

**BRCA1 R1699Q variant displaying ambiguous functional abrogation
confers intermediate breast and ovarian cancer risk**

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Abstract

Background: Clinical classification of rare sequence changes identified in the breast cancer susceptibility genes *BRCA1* and *BRCA2* is essential for appropriate genetic counseling of individuals carrying these variants. We previously showed that variant *BRCA1* c.5096G>A p.Arg1699Gln in the *BRCA1* transcriptional transactivation domain demonstrated equivocal results from a series of functional assays, and proposed that this

variant may confer low to moderate risk of cancer. **Methods:** Measures of genetic risk (report of family history, segregation) were assessed for 68 *BRCA1* c.5096G>A p.Arg1699Gln (R1699Q) families recruited through family cancer clinics, comparing results to 34 families carrying the previously classified pathogenic *BRCA1* c.5095C>T p.Arg1699Trp (R1699W) mutation at the same residue and to 243 breast cancer families with no *BRCA1* pathogenic mutation (BRCA-X). **Results:** Comparison of *BRCA1* carrier prediction scores of probands using the BOADICEA risk prediction tool revealed that *BRCA1* c.5096G>A p.Arg1699Gln variant carriers had family histories that were less “*BRCA1*-like” than *BRCA1* c.5095C>T p.Arg1699Trp mutation carriers ($p < 0.00001$), but more “*BRCA1*-like” than BRCA-X families ($p = 0.0004$). Further, modified segregation analysis of the subset of 30 families with additional genotyping showed that *BRCA1* c.5096G>A p.Arg1699Gln had reduced penetrance compared to the average truncating *BRCA1* mutation penetrance ($p = 0.0002$), with estimated cumulative risks to age 70 of breast or ovarian cancer of 24%. **Conclusion:** Our results provide substantial evidence that the *BRCA1* c.5096G>A p.Arg1699Gln (R1699Q) variant, demonstrating ambiguous functional deficiency across multiple assays, is associated with intermediate risk of breast and ovarian cancer, highlighting challenges for risk modeling and clinical management of patients of this and other potential moderate-risk variants.

INTRODUCTION

The clinical classification of rare sequence changes identified in the high risk breast cancer susceptibility genes *BRCA1* and *BRCA2* is essential for appropriate genetic counseling of individuals carrying these variants. Classification of *BRCA1* and *BRCA2*

variants was facilitated by the development of a multifactorial likelihood model[1], which provides a quantitative estimate of pathogenicity by assessing measures of genetic and other features of variant carriers relative to characteristics observed for classical high-risk mutations. Moreover, this quantitative assessment of risk has been linked to clinical management guidelines to provide a basis for standardized variant reporting, variant classification and management of families with such variants[2]. The multifactorial likelihood methodology has been applied in multiple studies[1, 3-15], with more than 200 *BRCA1* or *BRCA2* variants now classified using this approach[16]. However, the multifactorial approach is designed to distinguish high-risk mutations from variants with no or little clinical significance, and it is likely that additional methods are required to detect and validate *BRCA1* or *BRCA2* rare variants associated with more modest risks than the average penetrance reported for classical mutations in these genes i.e. 65% risk of breast cancer and 39% risk of ovarian cancer to age 70 years for *BRCA1* mutations, and 45% risk of breast cancer and 11% risk of ovarian cancer to age 70 years for *BRCA2* mutations [17].

We previously showed that the variant *BRCA1* R1699Q (c.5096G>A p.Arg1699Gln) located in the *BRCA1* carboxyl terminal (BRCT) regions of the transcriptional transactivation domain (TAD) demonstrated equivocal results from a series of functional assays, when compared to wildtype control and known pathogenic missense mutation *BRCA1* A1708E (c.5123C>A p.Ala1708Glu) which was null in all assays[8]. In particular, this variant displayed intermediate transcriptional transactivation activity in human 293T and T47D cell lines and wild type centrosome amplification function, but

behaved as a deleterious mutation when assayed for formation of nuclear foci and trypsin sensitivity. There is also inconsistency in assay results from other functional studies, including discrepancies between yeast and mammalian transcriptional transactivation assay results in a single report[18], and categorization of R1699Q as a variant with strong functional effect due to compromised peptide binding activity and specificity, and compromised transcriptional activity in yet another study[19]. Most recently, Chang et al.[20] performed an extensive study of the R1699Q substitution using mouse embryonic stem (ES) cell-based functional assays, and demonstrated that this variant affected mouse ES cell survival and differentiation and was unable to rescue embryonic lethality of *Brcal*-null mice. However, this study *also* demonstrated that the variant did not cause significant cell cycle defects and had no effect on genomic stability, but it was suggested that abrogated repression of oncomir miR-155 was the underlying mechanism for BRCA1-mediated tumor suppression. The equivocal behavior of this variant can be explained at a protein level, as demonstrated by protein modeling predictions shown in Supplementary Figure 1. R1699 is located in the linker connecting the BRCT repeat domain, and participates in a salt bridge between the BRCT repeats[21]. The loss of salt-bridging interactions and steric strain associated with accommodating a tryptophan substitution contributes to conformational instability of the R1699W (c.5095C>T p.Arg1699Trp) pathogenic mutation and subsequently disrupts transcriptional transactivation function. In contrast, substitutions with no/little effect on structure such as R1699Q may be fully or partially active in these assays. Moreover, R1699 lies in a conserved phosphopeptide binding groove of the BRCA1 repeat and plays an important role in phosphopeptide recognition through its interaction. Specifically, our protein

modeling results directly comparing R1699Q and R1699W show that the volume of R1699W is likely to cause steric clashes with the phosphopeptide, whereas the smaller surface and volume presentation of R1699Q will not cause steric clashes but may modestly alter phosphopeptide recognition (Supplementary Figure 1). These modeling predictions explain the experimental results from biophysical assays of BACH1 binding affinity which demonstrated that R1699W leads to a significant 160-fold reduction in affinity compared to wild type, whereas the reduction is only 24-fold for R1699Q [22].

We previously proposed that the R1699Q variant has partial abrogation of BRCA1 functions and may confer low to moderate risk of cancer that would be better measured using pooled family studies[8]. In a study assessing pathogenicity of 1433 variants based on family history, co-occurrence and co-segregation data from a large dataset derived from clinical testing at Myriad Genetic Laboratories, the combined odds that *BRCA1* R1699Q was a pathogenic variant compared to neutral/no clinical significance was 2.5:1, based on a sample of 16 family histories with co-segregation data on only 3 of these[11]. By contrast, *BRCA1* R1699W at the same residue was classified as pathogenic with odds in favor of pathogenicity of 39,978:1[11]. Bioinformatic analysis shows that the arginine at position 1699 is conserved through tunicate, but the severity of the amino acid substitution is much less marked for glutamine (Grantham deviation 43) compared to tryptophan (Grantham deviation 101). Accordingly, the Align-GVGD algorithm (<http://brca.iarc.fr>) classifies R1699Q as a C35, while R1699W falls in the most severe C65 category for missense alterations. Based on an analysis of the same Myriad data set,

C35 variants were estimated to have a prior probability of pathogenicity of 0.66, while C65 were associated with a prior probability of 0.81[11, 23].

Mohammadi et al. assessed the likelihood of causality by co-segregation analysis of a single family, and reported a likelihood ratio of 1.4 for R1699Q[24]. In another genetic study of several *BRCA1/2* sequence variants, Gomez-Garcia et al.[25] examined the R1699Q and R1699W variants as part of a model-building exercise that incorporated family history, and estimated the probability of pathogenicity to be 0.87 for R1699Q and >0.99 for R1699W. Although this model classified both variants as pathogenic mutations, the authors noted that R1699Q did not co-segregate completely with disease in one of three of the families in which such data were available.

In summary, a number of different studies to date indicate that the R1699Q variant demonstrates inconsistent or inconclusive results at the functional and genetic level. In an extension of our previous study[8], we confirmed the intermediate transcriptional transactivation activity of BRCA1 R1699Q in the 293T cell line relative to pathogenic variant R1699W at the same residue, and then initiated large-scale genetic studies to assess if this intermediate function might translate to the lower risk of breast and ovarian cancer in families for R1699Q compared to R1699W.

METHODS

Confirmation of Transcriptional Transactivation Activity

Using methods previously described[8], we first compared transcriptional transactivation activity of BRCA1 R1699Q in the 293T cell line to that of pathogenic variant R1699W at the same residue, and also to pathogenic control A1708E, and confirmed our original findings that this variant displayed intermediate function compared to wildtype sequence and known pathogenic TAD variants (Supplementary Figure 2).

Genetic Analyses

With ethical approval from the relevant institutional review boards, we then initiated large-scale genetic studies to assess if this intermediate function might translate to the risk of breast and ovarian cancer in families. Informed consent was obtained from all participants. Through collaboration facilitated in part by the ENIGMA consortium[26], we ascertained sufficient information from multiple clinical cancer genetics centers around the world (Table 1) to compare family history and risk profiles of families in which the R1699Q variant had been identified, to families with the known pathogenic mutation R1699W at the same residue. For an additional reference group, we also collected a set of pedigrees that had been tested clinically for *BRCA1* and *BRCA2* mutations from the same centers within the same time frame as the R1699Q and R1699W families but for which no pathogenic mutation or any other unclassified variant had been found (BRCA-X). The time-frame was determined by center to ensure that a similar

criterion for testing was used. The proband in each instance was defined as the individual initially screened for *BRCA1/2* mutations.

Table 1. Number of families included in the family history and penetrance analyses.

Group	# sites*	BRCA-X	R1699Q		R1699W	
		Family History	Family History	Penetrance	Family History	Penetrance
Netherlands & Belgium	8	42	15	5	11	4
Germany	13	39	5	1	13	8
France	6	4	5	3	2	1
Spain	1	28	0	0	1	0
Denmark	2	30	10	4	0	0
Mayo Clinic, USA	1	8	7	5	0	0
Sweden	3	14	14	5	5	5
Ohio, USA	1	8	1	1	0	0
Australia	3	44	6	2	0	0
Pennsylvania, USA	1	2	1	1	0	0
United Kingdom	4	24	4	2	2	1
South Africa	1	0	1	1	0	0
TOTAL		243	68	30	34	19

* Number of individual clinic sites for each contributing country or region.

Family history analysis: As a measure of how each family fit the characteristics of a *BRCA1* mutation-positive family, we used the BOADICEA risk prediction algorithm [27-30] to calculate the probability that the proband from each family was a carrier of a *BRCA1* mutation based on the pedigree structure and the phenotypes of individuals in the pedigree. BOADICEA uses a full pedigree likelihood approach and incorporates ages at diagnosis of breast and ovarian cancer, presence of pancreatic and prostate cancer, the age at last follow-up for unaffected individuals, and the year of birth to account for cohort effects in penetrance. The model estimates the simultaneous effects of the high risk genes *BRCA1* and *BRCA2* using the age-specific penetrance estimates derived from 22 population-based studies [17], while allowing for unknown genetic effects that explain

the residual familial clustering of breast cancer. The residual familial clustering is explained by a polygenic component with variance that decreases linearly with age.

The estimated probabilities of the proband carrying a pathogenic *BRCA1* mutation based on the BOADICEA prediction model, B_i , were then transformed in order to better fit a Gaussian distribution using a logit transformation $b_i = \text{logit}(B_i) = \ln(B_i/(1-B_i))$, so that standard statistical methods could be used. For each center that contributed R1699Q/W families, we calculated the mean and standard deviation of the probabilities calculated for the BRCA-X families from this center. This distribution was used to create z-scores as $Z_{ij} = (b_{ij} - X_j)/S_j$, where b_{ij} is the logit of the BOADICEA predicted probability of a *BRCA1* mutation in the i th BRCA-X family in the j th Center, X_j and S_j are the sample mean and standard deviation of the logit-transformed B_i from the j th center. For families with the sequence variants of interest, R1699Q and R1699W, these Z_{ij} thus represent the position of family histories of probands carrying an R1699Q or R1699W variant within the distribution of families tested negative for *BRCA1/2* mutations in the same centers and time frame. Letting Z_{Qi} be the standardized logit score of the i th R1699Q family and Z_{Wi} represent the corresponding score for the i th R1699W family. Assuming further that the Z_{Qi} and Z_{Wi} are Normally distributed with means μ_Q and μ_W and variances σ_Q^2 and σ_W^2 respectively, these scores can then be used to test the following two hypotheses:

- (1) The family histories of R1699Q probands are more *BRCA1*-like than those of matched BRCA-X. That is, we test the null hypothesis $\mu_Q = 0$ versus the alternative $\mu_Q > 0$ with a one-sample t-test. Rejection of the null hypothesis indicates that the

R1699Q families have proband/family histories more compatible with a pathogenic *BRCA1* mutation than the center matched BRCA-X families.

- (2) The family histories of R1699Q are less “*BRCA1*-like” than those of R1699W mutations. This is tested by a two sample t-test of the null hypothesis $\mu_Q = \mu_W$ against the one-sided alternative $\mu_Q < \mu_W$.

If both these null hypotheses are rejected, this indicates that R1699Q variants are in some sense intermediate in terms of their *BRCA1* family history profile compared to BRCA-X and *BRCA1* R1699W families.

Segregation Analyses: Risk was analyzed more directly through analysis of co-segregation of the R1699Q/W genotypes in the relatives of probands presenting with R1699Q/W variants [31]. Analyses included 30 R1699Q informative families with 111 total tested individuals and 19 R1699W families with 80 tested individuals. Risks were estimated by examining the likelihood of the genotypes of the family members (both women affected with breast or ovarian cancer, and healthy women) as a function of *BRCA1* penetrance, conditional on the proband’s genotype and all pedigree phenotypes. The conditioning is needed to account for the fact that families were ascertained on the basis of the cancer phenotypes in the entire family, and the fact that the proband carried the variant. In this situation, most information about penetrance derives from the distribution of variant genotypes among unaffected women. Because there was insufficient additional genotyping in these families to reliably estimate age-specific risk ratios for each age group, we examined the risk associated with the R1699Q/W variants relative to those associated with the “average pathogenic *BRCA1* mutation” as found in

much larger studies of predominantly truncating mutations[17]. In these analyses the age-specific hazard ratio (by decade) was assumed to be a constant multiple of the estimate of Antoniou et al.[17], with cumulative penetrances re-estimated at each trial value of the multiplier. This allowed for a similar pattern of age-specific effects as in *BRCA1*, but only required estimation of a single parameter. We also repeated the analyses allowing for separate penetrance multipliers for breast cancer and ovarian cancer to allow for the possibility that the functional effects of R1699Q or R1699W might be more relevant to cancer risk for one but not both of these cancers. We varied the multiplier of the assumed standard penetrance of *BRCA1* from 0.05 to 2, in increments of 0.01 in order to find the value that maximized the likelihood of the observed data (and to obtain confidence intervals). If under a particular model, a given value of the penetrance implied risks of cancer in carriers lower in a given age group than in non-carriers, these were constrained to be the same as the non-carrier rates.

The analysis of penetrance was done using the LINKAGE package of programs [32] to calculate pedigree likelihoods and the other statistical analyses were performed using STATA v11.0 (StatCorp, College Station TX).

RESULTS

Table 2 shows the results of the analyses comparing family history scores of probands from R1699Q families, R1699W families, and families with no *BRCA1* pathogenic mutation. Of note is the ordered progression of the BOADICEA raw scores showing

clear differences between all three groups of families, and the tests of significance between groups. The Z-scores for R1699Q (adjusted for the mean and standard deviation of the BRCA-X families from the same centers) were significantly greater than 0 ($p=0.0004$), indicating that carriers of the R1699Q variant have more “*BRCA1*-like” family histories than families that test negative for both genes, and that they have some of the characteristics of family history (e.g, ovarian cancer) of *BRCA1*. However, they are also clearly less “*BRCA1*-like” than family histories of probands carrying the previously classified pathogenic R1699W mutation ($p<0.00001$).

Table 2. Analysis of family history scores.

Group	Number of Families ^a	Mean BOADICEA Score ^b	Z-Scores ^c	
			Mean	Standard Error
BRCA-X	243	0.085	0.0	0.06
R1699Q	67	0.12	0.48	0.14
R1699W	34	0.36	1.81	0.20
<i>H0: Q=0</i> $t= 3.51, 66 df, p=0.0004$				
<i>H0: Q=W</i> $t= -5.5, 99 df, p=1.3 \times 10^{-7}$				

^a Analysis includes 67 families in which the proband carried a R1699Q variant and for which there was at least 1 matching BRCA-X family, and the 34 matching families in which the proband had a R1699W variant. One R1699Q family was excluded because there was no BRCA-X family available for comparison. For another R1699Q family with only a single matched BRCA-X family, the standard deviation could not be determined so the average standard deviation across all centers was used for analysis. Families were matched by country, as shown in Table 1.

^b All BOADICEA scores represent “pre-test” calculations based on family history information only. Scores ranges were as follows:- Q: 0.0006 to 0.88; W: 0.0026 to 0.97; X: 0.0002 to 0.92.

^c See text for definition of Z-scores

Although the above analyses indicate that families carrying R1699Q are different in terms of their personal and family history from both BRCA-X families and families carrying the R1699W variant, these analyses do not directly address the question of cancer risks conferred by these mutations. They also do not provide a level of evidence that the variant is pathogenic as in the typical assessment of co-segregation within the framework of the multifactorial model [1, 33]. Segregation analyses were thus undertaken. For R1699W, the maximum likelihood estimate of the relative proportion of the standard *BRCA1* penetrance was 0.24 (95% CI=0.06-1.10), which was not significantly different from 1.0 (Likelihood Ratio (LR) $X_1^2=3.44$; $p=0.06$). The odds in favor of pathogenicity at this value of the penetrance multiplier were 314,100:1. When we allowed the possibility that there were different multipliers for breast and ovarian cancer, the estimates were 0.11 for breast cancer and 2.35 for ovarian cancer, with corresponding odds of 2,420,000:1 in favor of pathogenicity. The LR test provided some evidence for difference from a single value ($X_1^2 = 4.08$; $p=0.043$) and for a difference from standard penetrance ($X_2^2 = 7.53$; $p=0.023$).

For R1699Q, the maximum likelihood estimate of the penetrance multiplier parameter was 0.20 (95% CI=0.09-0.45), significantly reduced compared to the standard model ($X_1^2 = 14.2$; $p=0.0002$). The odds in favor of pathogenicity were 6226:1 for R1699Q at this value of the multiplier, whereas they were only 5:1 under the standard model. In contrast to R1699W, allowing separate multipliers for breast and ovarian cancer did not result in a big difference in likelihood, with estimated parameters of 0.18 for breast cancer and 0.30

for ovarian cancer (odds of 6787:1), which was not significantly different from a single value of 0.20 ($p=0.7$) for breast and ovarian cancer.

Clearly there is a reduced penetrance for this variant, compared to the standard penetrance of *BRCA1* as estimated by Antoniou et al.[17]. To represent these estimated parameters in terms of absolute risks which are perhaps more clinically relevant, we can translate the penetrance multipliers into age-specific relative risks of breast and ovarian cancer and use these to obtain cumulative risks of breast and/or ovarian cancer by age, based on the age-specific relative risks in Antoniou et al.[17]. Figure 1 shows the predicted cumulative risks of developing either breast or ovarian cancer based on the maximum likelihood parameter estimates of the breast and ovarian relative risk multiplier parameters for R1699Q and R1699W compared to the standard model and population rates. Similar figures for breast cancer risk and ovarian cancer risk individually are provided in the supplementary Figure 3. If our model is correct, the risk of breast or ovarian cancer to age 70 is 24% (95% CI 10%-40%) for carriers of *BRCA1* R1699Q, and 58% (95% CI 7%-72%) for carriers of *BRCA1* R1699W assuming the best fitting model of separate risk multipliers for breast and ovarian. This compares to 4.6% for women in the general population, and 68% for the carriers of an average pathogenic mutation. The risks for R1699Q are higher than that conferred by family history alone, but still lower than that conferred by *BRCA2* and *PALB2* mutations.

DISCUSSION

Although we have presented results of analyses examining risk, our goal was not to estimate penetrance *per se*, but rather to compare these two specific variants to the penetrance of the “average” *BRCA1* mutation (the vast majority of which are truncating), both in terms of family histories of probands carrying these variants and in terms of co-segregation of the variants within families. Here we provide, for the first time, significant evidence that a *BRCA1* variant can be associated with reduced risks of breast cancer compared to the “average” pathogenic mutation. It is of particular relevance and consequence for future studies, since the variant R1699Q was selected for study due to its behavior in a variety of functional assays. Depending on the assay, this missense variant has demonstrated either wildtype function, abrogated function akin to known pathogenic mutations, or functional activity intermediate between that observed for wild type *BRCA1* and known truncating pathogenic and missense pathogenic mutations.

Interestingly there was also evidence that the R1699W variant was associated with significantly lower breast cancer risk and a markedly increased risk of ovarian cancer. We recognize that the estimation of breast and ovarian parameters separately is somewhat difficult given the necessity of conditioning the data on all pedigree phenotypes, but the results nevertheless raise the question that differences in risk of breast versus ovarian cancer may be a characteristic of some missense mutations in the BRCT repeat domains. In this regard, we note that in the 34 R1699W families there were an average of 2.24 breast cancers and 1.48 ovarian cancers, while in the 68 R1699Q families there were 2.35

breast cancers and 0.85 ovarian cancers per family, consistent with the higher estimated risk of ovarian cancer in these families. Further study of a large number of such variants will be necessary to address such an intriguing possibility that would have clear clinical implications.

Using the standard multifactorial model, the posterior probability for R1699Q is calculated to be 0.79 from the available data, namely: prior probability of pathogenicity of 0.66 based on the A-GVGD class C35 [23]; segregation odds of 5:1 in favor of pathogenicity from *this* study of 30 families; likelihood ratios from Easton et al.[11] of 8:1 against pathogenicity for family history, and 3:1 in favor of pathogenicity for co-occurrence data. That is, using the model developed based on the characteristics of *BRCA1* pathogenic mutations of “average” penetrance, R1699Q would be classified as IARC Class 3 “uncertain”.

Our conclusive finding that *BRCA1* c.5096G>A R1699Q can be shown to both have intermediate functional deficiency in several assays, and is associated with breast and ovarian cancer risk at significantly lower levels than truncating *BRCA1* mutations, has a number of consequences. Our findings suggest that results from a battery of functional assays may highlight other variants with intermediate or equivocal results for investigation as potential moderate risk variants. Indeed, the variant *BRCA1* A1708V showed abrogated centrosome amplification but normal nuclear foci formation and trypsin sensitivity equivocal results from a series of functional assays in our original report[8], and is a candidate for further investigation as a potential moderate risk variant.

If this observation of intermediate function translating to intermediate risk is a general finding, it is likely that there will be a subset of variants that are difficult to classify using the standard multifactorial likelihood approaches that are based on comparing data for a particular variant under the hypothesis that it is a fully penetrant pathogenic *BRCA1* mutation against the hypothesis that it is neutral or of no clinical significance with respect to risk. As shown for the R1699Q variant with more families available for analysis than will likely be achieved for most other rare variants, the standard co-segregation analysis yielded odds of only 5:1 in favor of the variant being pathogenic compared to the >6000:1 odds when a lower penetrance was allowed. Further, and more importantly, we must now face the question of how these women should be counseled in terms of cancer risk and the management of that risk. We do not propose that counseling be any different for R1699W, although results from the two parameter analyses suggest that particular attention should perhaps be paid to ovarian cancer for this known pathogenic variant. We emphasize however that the confidence intervals are wide, particularly for cancer site-specific risks and future studies are necessary to confirm the markedly increased ovarian cancer risk observed in our dataset. While there is certainly significant evidence that R1699Q carriers are at increased risk over population rates, this risk is markedly lower than that observed for the average *BRCA1* mutation. The findings presented here are likely to provide impetus for research studies considering approaches to clinical management of patients with cancer risks intermediate to those conferred by *BRCA1/2* mutations and those from family history alone. In the case of R1699Q, counseling could be similar to that for other moderate-penetrance genes such as *PALB2*, *CHEK2* and

RAD51C, although that may change if ovarian cancer screening improves given the increased rate of ovarian cancer over the general population. In all these cases, the incorporation of the now 30+ common breast cancer susceptibility alleles into comprehensive risk prediction models will be of great value in allowing women and their providers to make informed management decisions. In addition, it would be interesting to specifically explore if *BRCA1* haplotypes altering promoter activity [34, 35] or potentially altering 3'UTR microRNA binding [36] influence the level of function of R1699Q *in vivo*, and explain in part the variable presentation of families.

In summary, we provide evidence that a *BRCA1* variant demonstrating equivocal functional deficiency across multiple assays is associated with intermediate risk of breast and ovarian cancer, highlighting challenges for risk modeling and clinical management of patients of this and other potential moderate-risk variants.

Contributor Statement

All authors made a significant contribution to data collection, data analysis, writing, and critical assessment of this study. Specifically: DEG and ABS were responsible for study concept and design. DEG performed the statistical analyses. DEG, ABS and MPG V wrote the manuscript. PJW, SH, and BT were responsible for data collection and management. CJVA, AAKM, MGEMA, RKS, CE, AM, SC, OS, RL, FJC, LG, TVOH, MT, DME, KT, JB, SMD, AET, EJVT, BW, AB, MPG V provided the family data analyzed in this manuscript; MAB and CP performed functional analyses, and SC performed structural protein modeling. All authors have approved the final draft submitted.

Acknowledgements:

We thank the many families who participated in this study. This work is supported by the efforts of laboratory and clinical staff from many centers around the world. In particular, we would like to acknowledge the efforts of the individuals named in Appendix A for their contribution to this specific study. kConFaB thanks Heather Thorne, Eveline Niedermayr, kConFab research nurses and staff, heads and staff of the Family Cancer Clinics, and the Clinical Follow Up Study for their contributions to kConFab, and the many families who contribute to kConFab.

Funding:

This work was supported in part by project grants from The National Health and Medical Research Council (NHMRC) to ABS. ABS is supported by an NHMRC Senior Research Fellowship. kConFab is supported by grants from the National Breast Cancer Foundation, the NHMRC and by the Queensland Cancer Fund, the Cancer Councils of New South Wales, Victoria, Tasmania and South Australia, and the Cancer Foundation of Western Australia. The kConFab Clinical Follow Up Study was funded by NHMRC grants [145684 and 288704]. BJF is supported by the Canadian Institutes of Health Research Team Grant in Familial Risks of Breast Cancer CRN-87521. AL thanks the Swedish Cancer Society for support. The work of the German Consortium GC-HBOC is supported by a grant of the German Cancer Aid (grant 107364, RKS) and by the Centre for Molecular Medicine Cologne, Cologne, Germany (RKS, BW). The French Consortium thanks the Association d'Aide à la Recherche Cancérologique de Saint Cloud (ARCs)

and the Ligue 92 contre le Cancer for their financial support. FJC and DEG are supported by NIH Grant CA116167, an NIH Recovery Act supplement (CA116167Z) and an NIH Specialized Program of Research Excellence (SPORE) in Breast Cancer (CA116201). LG is supported by a Komen Race for the Cure Fellowship. Research by TvOH was supported by the NEYE Foundation. SMD is supported by funding from the Komen Foundation for the Cure. Ohio State University CCG is supported by the OSU Comprehensive Cancer Center (AET). EJVR is funded by grants from the Cancer Association of South Africa. The research coordinated by MPG was supported by Dutch Cancer Society grants 2001-2471 and 2006-3677. DEG is supported by NIH Grant CA116167. Co-ordination of ENIGMA is funded by The National Institutes of Health Recovery Act supplement award (CA116167Z).

References

1. Goldgar DE, Easton DF, Deffenbaugh AM, Monteiro AN, Tavtigian SV, Couch FJ. Integrated evaluation of DNA sequence variants of unknown clinical significance: application to BRCA1 and BRCA2. *Am J Hum Genet.* 2004;**75**:535-44.
2. Plon SE, Eccles DM, Easton D, Foulkes WD, Genuardi M, Greenblatt MS, Hogervorst FB, Hoogerbrugge N, Spurdle AB, Tavtigian SV. Sequence variant classification and reporting: recommendations for improving the interpretation of cancer susceptibility genetic test results. *Hum Mutat.* 2008;**29**:1282-91.
3. Whiley PJ, Guidugli L, Walker LC, Healey S, Thompson BA, Lakhani SR, Da Silva LM, Tavtigian SV, Goldgar DE, Brown MA, Couch FJ, Spurdle AB. Splicing and multifactorial analysis of intronic BRCA1 and BRCA2 sequence variants identifies clinically significant splicing aberrations up to 12 nucleotides from the intron/exon boundary. *Hum Mutat.* 2011;**32**:678-87.
4. Walker LC, Whiley PJ, Couch FJ, Farrugia DJ, Healey S, Eccles DM, Lin F, Butler SA, Goff SA, Thompson BA, Lakhani SR, Da Silva LM, Tavtigian SV, Goldgar DE, Brown MA, Spurdle AB. Detection of splicing aberrations caused by BRCA1 and BRCA2 sequence variants encoding missense substitutions: implications for prediction of pathogenicity. *Hum Mutat.* 2010;**31**:E1484-505.
5. Thomassen M, Blanco A, Montagna M, Hansen TV, Pedersen IS, Gutierrez-Enriquez S, Menendez M, Fachal L, Santamarina M, Steffensen AY, Jonson L, Agata S, Whiley P, Tognazzo S, Tornero E, Jensen UB, Balmana J, Kruse TA, Goldgar DE,

- Lazaro C, Diez O, Spurdle AB, Vega A. Characterization of BRCA1 and BRCA2 splicing variants: a collaborative report by ENIGMA consortium members. *Breast Cancer Res Treat.* 2011.
6. Spurdle AB, Lakhani SR, Healey S, Parry S, Da Silva LM, Brinkworth R, Hopper JL, Brown MA, Babikyan D, Chenevix-Trench G, Tavtigian SV, Goldgar DE. Clinical classification of BRCA1 and BRCA2 DNA sequence variants: the value of cytokeratin profiles and evolutionary analysis--a report from the kConFab Investigators. *J Clin Oncol.* 2008;**26**:1657-63.
 7. Spurdle AB, Lakhani SR, Da Silva LM, Balleine RL, Goldgar DE. Bayes analysis provides evidence of pathogenicity for the BRCA1 c.135-1G>T (IVS3-1) and BRCA2 c.7977-1G>C (IVS17-1) variants displaying in vitro splicing results of equivocal clinical significance. *Hum Mutat.* 2010;**31**:E1141-5.
 8. Lovelock PK, Spurdle AB, Mok MT, Farrugia DJ, Lakhani SR, Healey S, Arnold S, Buchanan D, Couch FJ, Henderson BR, Goldgar DE, Tavtigian SV, Chenevix-Trench G, Brown MA. Identification of BRCA1 missense substitutions that confer partial functional activity: potential moderate risk variants? *Breast Cancer Res.* 2007;**9**:R82.
 9. Lovelock PK, Healey S, Au W, Sum EY, Tesoriero A, Wong EM, Hinson S, Brinkworth R, Bekessy A, Diez O, Izatt L, Solomon E, Jenkins M, Renard H, Hopper J, Waring P, Tavtigian SV, Goldgar D, Lindeman GJ, Visvader JE, Couch FJ, Henderson BR, Southey M, Chenevix-Trench G, Spurdle AB, Brown MA. Genetic, functional, and histopathological evaluation of two C-terminal BRCA1 missense variants. *J Med Genet.* 2006;**43**:74-83.
 10. Chenevix-Trench G, Healey S, Lakhani S, Waring P, Cummings M, Brinkworth R, Deffenbaugh AM, Burbidge LA, Pruss D, Judkins T, Scholl T, Bekessy A, Marsh A, Lovelock P, Wong M, Tesoriero A, Renard H, Southey M, Hopper JL, Yannoukakos K, Brown M, Easton D, Tavtigian SV, Goldgar D, Spurdle AB. Genetic and histopathologic evaluation of BRCA1 and BRCA2 DNA sequence variants of unknown clinical significance. *Cancer Res.* 2006;**66**:2019-27.
 11. Easton DF, Deffenbaugh AM, Pruss D, Frye C, Wenstrup RJ, Allen-Brady K, Tavtigian SV, Monteiro AN, Iversen ES, Couch FJ, Goldgar DE. A systematic genetic assessment of 1,433 sequence variants of unknown clinical significance in the BRCA1 and BRCA2 breast cancer-predisposition genes. *Am J Hum Genet.* 2007;**81**:873-83.
 12. Sweet K, Senter L, Pilarski R, Wei L, Toland AE. Characterization of BRCA1 ring finger variants of uncertain significance. *Breast Cancer Res Treat.* 2010;**119**:737-43.
 13. Spearman AD, Sweet K, Zhou XP, McLennan J, Couch FJ, Toland AE. Clinically applicable models to characterize BRCA1 and BRCA2 variants of uncertain significance. *J Clin Oncol.* 2008;**26**:5393-400.
 14. Tavtigian SV, Deffenbaugh AM, Yin L, Judkins T, Scholl T, Samollow PB, de Silva D, Zharkikh A, Thomas A. Comprehensive statistical study of 452 BRCA1 missense substitutions with classification of eight recurrent substitutions as neutral. *J Med Genet.* 2006;**43**:295-305.
 15. Farrugia DJ, Agarwal MK, Pankratz VS, Deffenbaugh AM, Pruss D, Frye C, Wadum L, Johnson K, Mentlick J, Tavtigian SV, Goldgar DE, Couch FJ. Functional assays for classification of BRCA2 variants of uncertain significance. *Cancer Res.* 2008;**68**:3523-31.

16. Vallee MP, Francy TC, Judkins MK, Babikyan D, Lesueur F, Gammon A, Goldgar DE, Couch FJ, Tavtigian SV. Classification of missense substitutions in the BRCA genes: A database dedicated to Ex-UVs. *Hum Mutat.* 2011.
17. Antoniou A, Pharoah PD, Narod S, Risch HA, Eyfjord JE, Hopper JL, Loman N, Olsson H, Johannsson O, Borg A, Pasini B, Radice P, Manoukian S, Eccles DM, Tang N, Olah E, Anton-Culver H, Warner E, Lubinski J, Gronwald J, Gorski B, Tulinius H, Thorlacius S, Eerola H, Nevanlinna H, Syrjakoski K, Kallioniemi OP, Thompson D, Evans C, Peto J, Lalloo F, Evans DG, Easton DF. Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case Series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet.* 2003;**72**:1117-30.
18. Vallon-Christersson J, Cayan C, Haraldsson K, Loman N, Bergthorsson JT, Brondum-Nielsen K, Gerdes AM, Moller P, Kristoffersson U, Olsson H, Borg A, Monteiro AN. Functional analysis of BRCA1 C-terminal missense mutations identified in breast and ovarian cancer families. *Hum Mol Genet.* 2001;**10**:353-60.
19. Lee MS, Green R, Marsillac SM, Coquelle N, Williams RS, Yeung T, Foo D, Hau DD, Hui B, Monteiro AN, Glover JN. Comprehensive analysis of missense variations in the BRCT domain of BRCA1 by structural and functional assays. *Cancer Res.* 2010;**70**:4880-90.
20. Chang S, Wang RH, Akagi K, Kim KA, Martin BK, Cavallone L, Haines DC, Basik M, Mai P, Poggi E, Isaacs C, Looi LM, Mun KS, Greene MH, Byers SW, Teo SH, Deng CX, Sharan SK. Tumor suppressor BRCA1 epigenetically controls oncogenic microRNA-155. *Nat Med.* 2011;**17**:1275-82.
21. Williams RS, Green R, Glover JN. Crystal structure of the BRCT repeat region from the breast cancer-associated protein BRCA1. *Nat Struct Biol.* 2001;**8**:838-42.
22. Coquelle N, Green R, Glover JN. Impact of BRCA1 BRCT domain missense substitutions on phosphopeptide recognition. *Biochemistry.* 2011;**50**:4579-89.
23. Tavtigian SV, Byrnes GB, Goldgar DE, Thomas A. Classification of rare missense substitutions, using risk surfaces, with genetic- and molecular-epidemiology applications. *Hum Mutat.* 2008;**29**:1342-54.
24. Mohammadi L, Vreeswijk MP, Oldenburg R, van den Ouweland A, Oosterwijk JC, van der Hout AH, Hoogerbrugge N, Ligtenberg M, Ausems MG, van der Luijt RB, Dommering CJ, Gille JJ, Verhoef S, Hogervorst FB, van Os TA, Gomez Garcia E, Blok MJ, Wijnen JT, Helmer Q, Devilee P, van Asperen CJ, van Houwelingen HC. A simple method for co-segregation analysis to evaluate the pathogenicity of unclassified variants; BRCA1 and BRCA2 as an example. *BMC Cancer.* 2009;**9**:211.
25. Gomez Garcia EB, Oosterwijk JC, Timmermans M, van Asperen CJ, Hogervorst FB, Hoogerbrugge N, Oldenburg R, Verhoef S, Dommering CJ, Ausems MG, van Os TA, van der Hout AH, Ligtenberg M, van den Ouweland A, van der Luijt RB, Wijnen JT, Gille JJ, Lindsey PJ, Devilee P, Blok MJ, Vreeswijk MP. A method to assess the clinical significance of unclassified variants in the BRCA1 and BRCA2 genes based on cancer family history. *Breast Cancer Res.* 2009;**11**:R8.
26. Spurdle AB, Healey S, Devereau A, Hogervorst FB, Monteiro AN, Nathanson KL, Radice P, Stoppa-Lyonnet D, Tavtigian S, Wappenschmidt B, Couch FJ, Goldgar DE. ENIGMA-Evidence-based network for the interpretation of germline mutant alleles:

An international initiative to evaluate risk and clinical significance associated with sequence variation in BRCA1 and BRCA2 genes. *Hum Mutat.* 2011.

27. Antoniou AC, Cunningham AP, Peto J, Evans DG, Lalloo F, Narod SA, Risch HA, Eyfjord JE, Hopper JL, Southey MC, Olsson H, Johannsson O, Borg A, Pasini B, Radice P, Manoukian S, Eccles DM, Tang N, Olah E, Anton-Culver H, Warner E, Lubinski J, Gronwald J, Gorski B, Tryggvadottir L, Syrjakoski K, Kallioniemi OP, Eerola H, Nevanlinna H, Pharoah PD, Easton DF. The BOADICEA model of genetic susceptibility to breast and ovarian cancers: updates and extensions. *Br J Cancer.* 2008;**98**:1457-66.
28. Mavaddat N, Rebbeck TR, Lakhani SR, Easton DF, Antoniou AC. Incorporating tumour pathology information into breast cancer risk prediction algorithms. *Breast Cancer Res.* 2010;**12**:R28.
29. Antoniou AC, Pharoah PP, Smith P, Easton DF. The BOADICEA model of genetic susceptibility to breast and ovarian cancer. *Br J Cancer.* 2004;**91**:1580-90.
30. Antoniou AC, Hardy R, Walker L, Evans DG, Shenton A, Eeles R, Shanley S, Pichert G, Izatt L, Rose S, Douglas F, Eccles D, Morrison PJ, Scott J, Zimmern RL, Easton DF, Pharoah PD. Predicting the likelihood of carrying a BRCA1 or BRCA2 mutation: validation of BOADICEA, BRCAPRO, IBIS, Myriad and the Manchester scoring system using data from UK genetics clinics. *J Med Genet.* 2008;**45**:425-31.
31. Goldgar DE, Healey S, Dowty JG, Da Silva L, Chen X, Spurdle AB, Terry MB, Daly MJ, Buys SM, Southey MC, Andrulis I, John EM, Khanna KK, Hopper JL, Oefner PJ, Lakhani S, Chenevix-Trench G. Rare variants in the ATM gene and risk of breast cancer. *Breast Cancer Res.* 2011;**13**:R73.
32. Lathrop GM, Lalouel JM, Julier C, Ott J. Strategies for multilocus linkage analysis in humans. *Proc Natl Acad Sci U S A.* 1984;**81**:3443-6.
33. Goldgar DE, Easton DF, Byrnes GB, Spurdle AB, Iversen ES, Greenblatt MS. Genetic evidence and integration of various data sources for classifying uncertain variants into a single model. *Hum Mutat.* 2008;**29**:1265-72.
34. Chan KY, Liu W, Long JR, Yip SP, Chan SY, Shu XO, Chua DT, Cheung AN, Ching JC, Cai H, Au GK, Chan M, Foo W, Ngan HY, Gao YT, Ngan ES, Garcia-Barcelo MM, Zheng W, Khoo US. Functional polymorphisms in the BRCA1 promoter influence transcription and are associated with decreased risk for breast cancer in Chinese women. *J Med Genet.* 2009;**46**:32-9.
35. Cox DG, Simard J, Sinnott D, Hamdi Y, Soucy P, Ouimet M, Barjhoux L, Verny-Pierre C, McGuffog L, Healey S, Szabo C, Greene MH, Mai PL, Andrulis IL, Thomassen M, Gerdes AM, Caligo MA, Friedman E, Laitman Y, Kaufman B, Paluch SS, Borg A, Karlsson P, Askmalms MS, Bustinza GB, Nathanson KL, Domchek SM, Rebbeck TR, Benitez J, Hamann U, Rookus MA, van den Ouweland AM, Ausems MG, Aalfs CM, van Asperen CJ, Devilee P, Gille HJ, Peock S, Frost D, Evans DG, Eeles R, Izatt L, Adlard J, Paterson J, Eason J, Godwin AK, Remon MA, Moncoutier V, Gauthier-Villars M, Lasset C, Giraud S, Hardouin A, Berthet P, Sobol H, Eisinger F, Bressac de Paillerets B, Caron O, Delnatte C, Goldgar D, Miron A, Ozcelik H, Buys S, Southey MC, Terry MB, Singer CF, Dressler AC, Tea MK, Hansen TV, Johannsson O, Piedmonte M, Rodriguez GC, Basil JB, Blank S, Toland AE, Montagna M, Isaacs C, Blanco I, Gayther SA, Moysich KB, Schmutzler RK, Wappenschmidt B, Engel C, Meindl A, Ditsch N, Arnold N, Niederacher D, Sutter C, Gadzicki D, Fiebig B, Caldes T, Laframboise R, Nevanlinna H,

Chen X, Beesley J, Spurdle AB, Neuhausen SL, Ding YC, Couch FJ, Wang X, Peterlongo P, Manoukian S, Bernard L, Radice P, Easton DF, Chenevix-Trench G, Antoniou AC, Stoppa-Lyonnet D, Mazoyer S, Sinilnikova OM. Common variants of the BRCA1 wild-type allele modify the risk of breast cancer in BRCA1 mutation carriers. *Hum Mol Genet.* 2011;**20**:4732-47.

36. Pelletier C, Speed WC, Paranjape T, Keane K, Blitzblau R, Hollestelle A, Safavi K, van den Ouweland A, Zelterman D, Slack FJ, Kidd KK, Weidhaas JB. Rare BRCA1 haplotypes including 3'UTR SNPs associated with breast cancer risk. *Cell Cycle.* 2011;**10**:90-9.

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Competing Interests:

The authors declare there are no conflicts of interest.

Figure Legends

Figure 1: Cumulative risk of breast or ovarian cancer by age, assuming the best fitting models of penetrance for R1699Q (dotted line) and R1699W (dashed line). The corresponding curves for the standard penetrance (solid line) and for the general population (dash-dotted line) are also shown.

