

CHAPTER 6

Electroanalysis of inorganic redox systems – potassium iron

(III) cyanide and cerium (III) sulphate

6.1 Introduction

Boron-doped diamond electrodes, according to literature [1,2], have several advantages over the conventional glassy carbon electrode :

- Wide potential window of water stability
- Low background current (low baseline current)
- Chemical and structural stability
- Resistance to fouling

Voltammetry studies were undertaken to investigate the advantages of diamond electrodes over glassy carbon electrodes and to compare the electrochemical behaviour of the De Beers diamond electrodes with that of other diamond electrodes reported in literature.

In voltammetric techniques, an electrical potential gradient is applied across an electrode-solution interface to oxidise or reduce species present in solution [3]. The potential of the working electrode is measured relative to the reference electrode, for example a Ag/AgCl

electrode. The current passed through the system is measured between the working electrode and a counter electrode, for example a platinum mesh electrode. The counter electrode is essential since current flow through a reference interface might perturb the reference potential. A total of three electrodes are therefore generally used: a working electrode at which the redox reaction of interest takes place, a reference electrode where no current flows, and the counter electrode at which the current is measured.

After a voltammetric analysis, a voltammogram (a current versus potential plot), is produced. A voltammogram reflects the chemical identity of the redox species (as indicated by its oxidation or reduction potential) and its concentration (by the magnitude of the current at a given potential). The current versus potential plot has three basic features that have useful thermodynamic, kinetic, and analytical information: the size of the current, the peak shape, and its position on the applied potential axis.

The voltammetric study entailed the use of three techniques: cyclic voltammetry, linear sweep voltammetry and chronoamperometry. Cyclic voltammetry is a popular technique that is particularly powerful for initial studies of a new system. Essentially, the potential applied to the working electrode is swept through the potential range where an electrochemical reaction occurs. The direction of the scan is then reversed in order to determine whether the product of electron transfer is stable or whether the reaction intermediates or the final product are electroactive [4]. Linear sweep voltammetry is similar to cyclic voltammetry except that the direction of scan is not reversed, i.e. the voltammetric scan is restricted to one direction only.

Chronoamperometry is a potential step experiment where the current versus time response is recorded. The potential of the electrode is changed almost instantaneously from a value where no current passes (i.e. no chemical reaction occurs at the electrode surface) to one where the electrode reaction of interest takes place [4]. This gives rise to a diffusion limited current whose value varies with time. The current-time relationship is described by the Cottrell equation [5].

In this study, diamond electrodes containing varying amounts of boron were characterised by observing the effect of boron concentration on the potential window in an aqueous solution, and on the size of the baseline current. The effect of the surface area of the diamond electrode in an ascorbic acid (AA) system was also investigated. The potassium iron (III) cyanide and cerium (III) sulphate redox systems were used to compare the electrochemical behaviour of boron-doped diamond electrodes with that of the glassy carbon electrode.

6.2 Experimental

An AutoLab PGStat 100 (**Figure 6.1**), connected to a computer with the GPES AutoLab software, was used for the voltammetric measurements. All measurements were carried out in a three-electrode electrochemical cell (**Figure 6.2**). The cell comprised a platinum mesh counter electrode, a boron-doped diamond working electrode (refer to **Section 5.3.2** for the construction of the diamond electrode) and a Ag/AgCl reference electrode with a Luggin capillary.

All electrochemical measurements were carried out at room temperature (25°C) in a stationary analyte solution. All solutions were deaerated using nitrogen prior to any electrochemical measurement. The chemicals used were analytical grade quality. Potassium iron (III) cyanide ($K_3[Fe(CN)_6]$), potassium nitrate (KNO_3) and sodium sulphate (Na_2SO_4) were obtained from Merck, cerium (III) sulphate ($Ce_2(SO_4)_3$) and the phosphate buffer were obtained from Sigma Aldrich and the sulphuric acid (H_2SO_4) was obtained from Saarchem.

The supporting electrolytes (KNO_3 , H_2SO_4 and Na_2SO_4) and the phosphate buffer were made up to the desired concentration using ultrapure water from a Millipore purification system, having a conductivity of $18 \mu\Omega.cm$. The $K_3[Fe(CN)_6]$ and the $Ce_2(SO_4)_3$ stock solutions were made up to the predetermined concentrations using the supporting electrolyte solutions. $Ce_2(SO_4)_3$, however, is known to polymerise in solution on heating. $Ce_2(SO_4)_3$ was therefore slowly added to a stirred solution of H_2SO_4 at a temperature of below $0^\circ C$, until the $Ce_2(SO_4)_3$ had completely dissolved.



Figure 6.1 : An AutoLab PGStat 100 connected to a PC carrying the GPES

AutoLab software

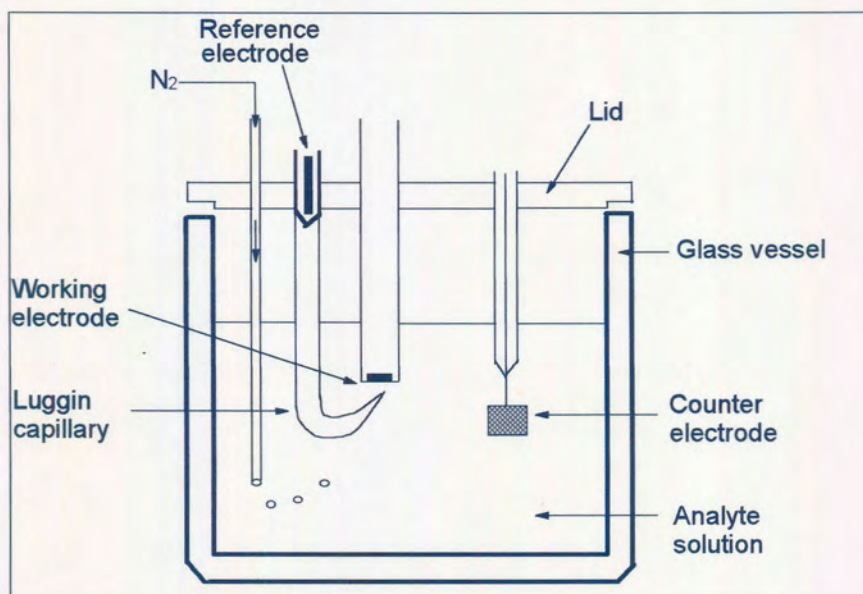


Figure 6.2 : Schematic diagram of a three electrode electrochemical cell

Voltammograms for the inorganic redox systems were then obtained at both glassy carbon and boron-doped diamond (BDD) electrodes. The surface areas of the glassy carbon and BDD electrodes were 0.071 cm^2 and 0.15 cm^2 respectively. The CVDBD4 electrodes having a boron concentration of 543 mg/L were used to investigate the $\text{K}_3[\text{Fe}(\text{CN})_6]$ and $\text{Ce}_2(\text{SO}_4)_3$ redox systems.

6.3 Results and discussion

6.3.1 Electrochemical characterisation of diamond electrodes of varying boron concentrations

The boron concentrations for the various samples of the BDD are given in **Table 4.4**. CVDBD6 is the highest boron-doped sample whereas CVDBD2 is the lowest boron-doped sample. **Figure 6.3** represents the electrochemical behaviour of diamond electrodes containing varying amounts of boron in 0.5 mol/L perchloric acid (HClO_4).

The observed onset of water electrolysis, i.e. water oxidation and water reduction for the various BDD electrodes as well as the glassy carbon electrode, is found in **Table 6.1**. It appears that, as the boron concentration of the BDD increases, the potential window decreases. The decrease in the potential window of the BDD electrodes is thought to be due to the small increase of sp^2 carbon (graphitic or amorphous carbon) in the BDD electrodes. The observation was made previously in **Table 4.5** that, as the boron concentration of the CVD samples increases, the sp^2/sp^3 ratio of the respective samples also increases. It is clear, however, that the potential window for the highest BDD

electrode is still wider than that of glassy carbon, thereby confirming the advantage that the BDD electrode has over glassy carbon when investigating aqueous systems.

Table 6.1 : Onset of water oxidation and water reduction for the various boron-doped CVD diamond electrodes as well as that of the glassy carbon electrode, in 0.5 mol/L HClO₄ solution

CVD sample	Hydrogen evolution / V	Oxygen evolution / V
CVDBD2	-1.34	1.80
CVDBD3	-1.14	1.77
CVDBD5	-1.06	1.77
CVDBD6	-0.95	1.77
Glassy carbon	-0.88	1.65

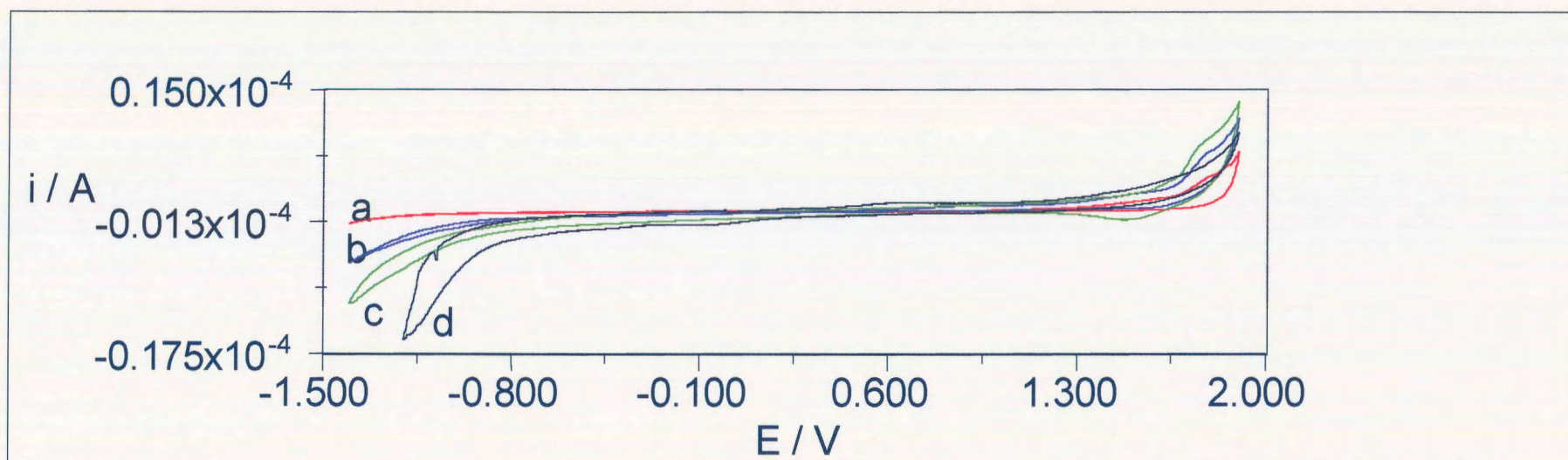


Figure 6.3 : Electrochemical behaviour of diamond electrodes containing varying amounts of boron in 0.5 mol/L HClO₄, (a)

CVDBD2, (b) CVDBD3, (c) CVDBD5, (d) CVDBD6 – scan rate is 50 mV/s. $E_i = -1$ V, scan direction = positive.

6.3.2 Surface area effects

Three different diamond electrode projected surface areas were investigated: 0.1590 cm^2 , 0.0707 cm^2 and 0.0078 cm^2 . The electrochemical behaviour of these diamond electrodes was determined using an ascorbic acid (AA) system. The structure of AA is found in **Figure 6.4**. **Figure 6.5** represents the cyclic voltammograms for 6 mmol/L AA in 0.1 mol/L HClO_4 at diamond electrodes having varied surface areas. An anodic peak ascribed to the oxidation of the ascorbic acid molecule is seen at approximately 1.3 V . The magnified electrochemical response of the 0.0078 cm^2 diamond electrode in AA is given in **Figure 6.6**.

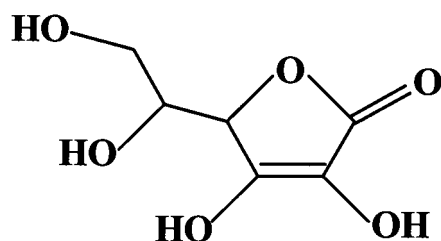


Figure 6.4 : Chemical structure of ascorbic acid [3]

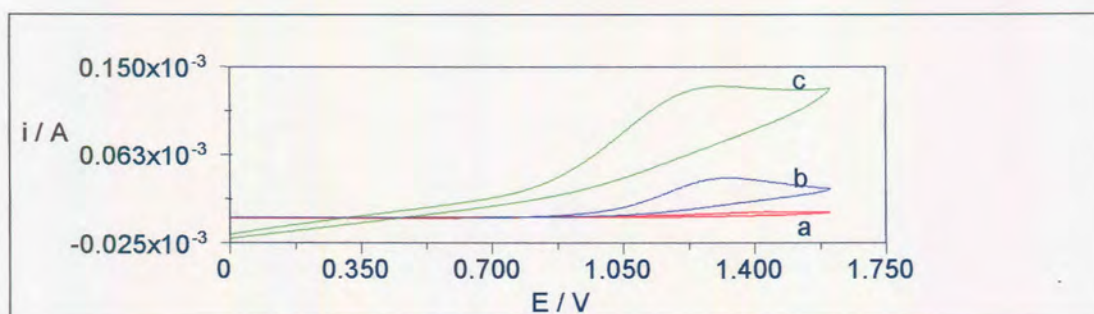


Figure 6.5 : Cyclic voltammograms for 6 mmol/L AA in 0.1 mol/L HClO₄ at CVDBD6 diamond electrodes with varying surface areas, (a) 0.0078 cm², (b) 0.0707 cm², (c) 0.1590 cm² - scan rate is 50 mV/s. E_i = 0 V, scan direction = positive.

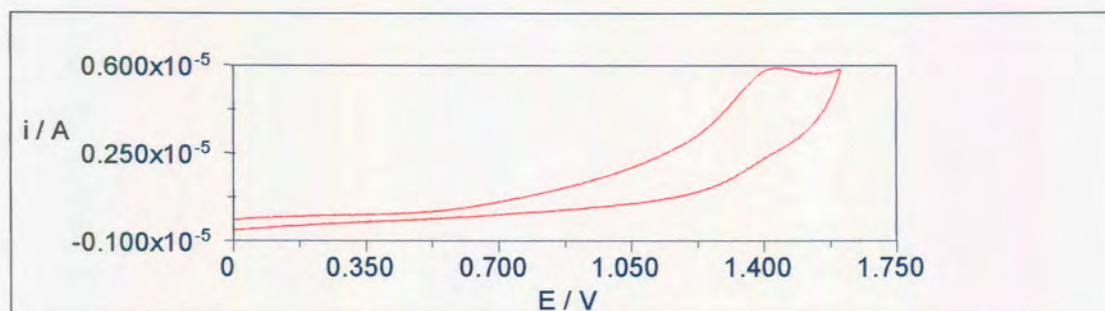


Figure 6.6 : Magnified electrochemical response of the 0.0078 cm² diamond electrode in AA. Scan rate is 50 mV/s.

From **Figure 6.5**, it can be seen that the largest diamond electrode surface area yields the greatest oxidation current. The oxidation current values are given in **Table 6.2**. This observation is in agreement with the fact that as the surface area of the electrode material increases, the corresponding electrochemical response also increases. The oxidation current values (**Table 6.2**) do not increase linearly with increasing surface area. This could be due to the difference between the true surface area and the projected surface area, brought about by the rough nature of the electrode surface. Polishing of the

electrode surface could rectify this, however, this would also alter the electrode surface state in terms of chemical functionalities. A method that could be employed to determine the true surface area of the BDD electrode is anodic stripping voltammetry.

Table 6.2 : Oxidation current values for boron-doped diamond (BDD) electrodes of varied surface areas - taken from Figure 6.5

Surface area of CVDBD6 / cm ²	Oxidation current values / μA
0.0078	5.90
0.0707	40
0.1590	132

6.3.3 Potassium iron (III) cyanide

The $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple has a complex chemistry associated with it. Despite this complexity, the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ couple has been used to evaluate the performance of diamond electrodes [6-8]. Although it is often presumed that this redox couple undergoes electron transfer via a simple outer-sphere mechanism, there are several pieces of evidence suggesting otherwise:

- (a) The electron-transfer rate constant of the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ system, k^0 , has been shown to depend on the nature and concentration of the supporting electrolyte cation [9].

- (b) A semipassivating layer of $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ (Prussian blue) can be formed on the electrode surface and this layer serves to decrease the rate of electron transfer [10,11].
- (c) Chen and McCreery [12-14] have reported a dependence of k^0 on the electrode surface structure. This work indicates that the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple reacts on carbon via specific surface interactions through an innersphere pathway.

It has been suggested that boron-doped diamond electrodes are nothing more than an ensemble of non-diamond impurity domains isolated within an inactive diamond matrix. The relative role of the non-diamond carbon in electron transfer reactions at diamond electrode surfaces has also been questioned [15].

The work undertaken by Swain et. al. [16] subsequently entailed a direct comparison of the electrochemical behaviour of a high-quality boron-doped diamond electrode (containing a lower volume fraction of non-diamond carbon impurity) with that of a low-quality electrode (higher volume fraction of non-diamond carbon impurity) in the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ system.

They found that the high quality electrode was more active for $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ than the low quality electrode. This implies that the electron transfer at diamond does not appear to occur through the non-diamond impurity sites but rather seems to occur through the doped diamond lattice.

Figure 6.7 shows the cyclic voltammetric i - E curve for 20 mmol/L $[\text{Fe}(\text{CN})_6]^{3-}$ in 1 mol/L potassium nitrate (KNO_3) and 0.1 mol/L phosphate buffer, pH 6.7, at a BDD electrode (CVDBD4). The i - E curve shows two well-defined waves with a peak separation (ΔE_p) of 690 mV. This peak separation should, in fact, be 59 mV for a reversible one-electron transfer reaction such as the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox reaction. A large peak separation implies a slow electron-transfer reaction occurring at the electrode surface between the analyte and the electrode. This means that the response time for the electrode is slow.

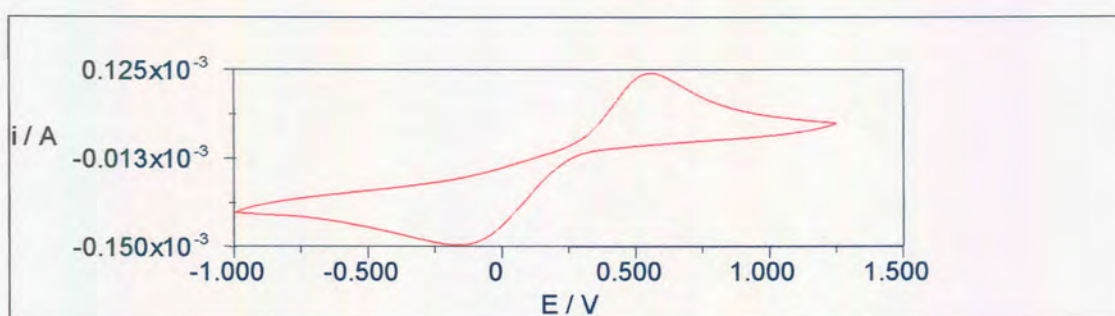


Figure 6.7 : Cyclic voltammetric curve for 20 mmol/L $[\text{Fe}(\text{CN})_6]^{3-}$ in 1 mol/L KNO_3 and 0.1 mol/L phosphate buffer, pH 6.7, at a BDD electrode (CVDBD4) - scan rate is 50 mV/s. $E_i = 1.25$ V, scan direction = negative

Figure 6.8 illustrates the cyclic voltammogram for 20 mmol/L $[\text{Fe}(\text{CN})_6]^{3-}$ in 1 mol/L KNO_3 at a BDD electrode as well as at a glassy carbon electrode. The cyclic voltammogram for $[\text{Fe}(\text{CN})_6]^{3-}$ at the glassy carbon electrode yielded a ΔE_p of 61 mV which is an order of magnitude lower than that of the diamond electrode.

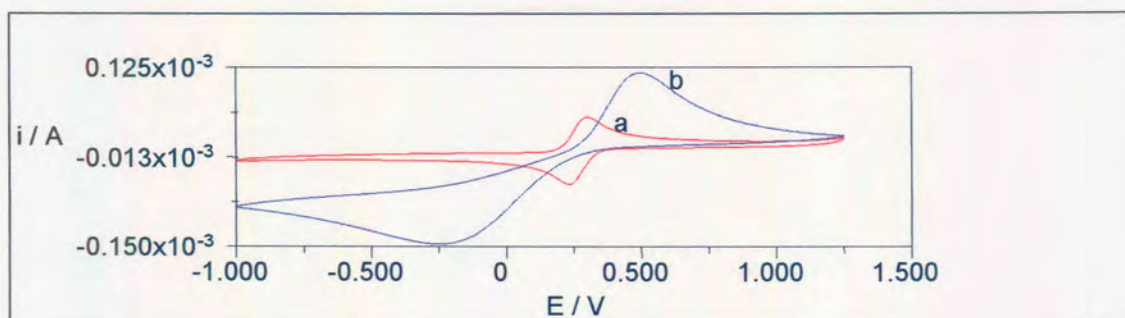


Figure 6.8 : Cyclic voltammograms for 20 mmol/L $[\text{Fe}(\text{CN})_6]^{3-}$ in 1 mol/L KNO_3 at a BDD diamond electrode (b) as well as at a glassy carbon electrode (a). Scan rate is 50 mV/s. $E_i = 1.25$ V, scan direction = negative

The contact angle measurement of the diamond electrode (CVDBD4) used in this experiment was between 72° - 88° (refer to **Section 4.6**). The XPS result (refer to **Section 4.5**) for the as-received sample showed the oxygen content to be 2.37×10^{15} atoms/cm², with an oxygen coverage of 1.26 monolayers. The contact angle measurement together with the XPS analysis implies that the surface of the diamond is partially hydrophobic. This therefore indicates that there are oxygen functionalities present on the diamond surface, i.e. the surface is partially oxygen terminated.

Many authors [17-19] report that the peak separation of the BDD electrode in the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ system is very sensitive to the surface termination of the electrode. The smallest ΔE_p is observed at the hydrogen terminated surface, whilst the largest ΔE_p is observed at the oxygen terminated surface. Ramesham and Rose [18] reported a ΔE_p of 192 mV in the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ system at a MA-CVD as-grown diamond electrode, whilst Yagi et. al. [20] reported a ΔE_p of 120 mV at a hydrogen

plasma treated diamond electrode and a ΔE_p of 1072 mV at an oxygen plasma treated electrode.

The evidence above suggests that oxygen termination inhibits $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ electron transfer, because of chemical effects. These chemical influences could involve electrostatic or site-blocking effects. In the light of the above literature findings, the large peak separation of 690 mV observed for the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple could be explained as follows :

- (a) Carbon-oxygen functionalities (for example carboxyl groups) on the surface of the diamond electrodes can be deprotonated, thereby producing a negative surface charge. This charge could repel the highly charged $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ species and in so doing, reduce the rate of electron transfer. Hence, a large ΔE_p would be observed.
- (b) Granger and Swain [17] believe that the hydrogen terminated surface appears to provide a specific site through which the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ couple can interact, which lowers the activation barrier for electron transfer. These surface interactions seem to be blocked on the oxygen terminated surface. This theory therefore explains the large ΔE_p observed since the surface of CVDBD4 is partially oxygen terminated.

It can therefore be assumed that in order to reduce the ΔE_p , the surface of the diamond electrode will have to be made hydrophobic.

The influence of the supporting electrolyte cation was also investigated. **Figure 6.9** represents the cyclic voltammogram for 20 mmol/L $K_3[Fe(CN)_6]$ in 1 mol/L KNO_3 and 0.1 mol/L sodium sulphate (Na_2SO_4) at a BDD electrode.

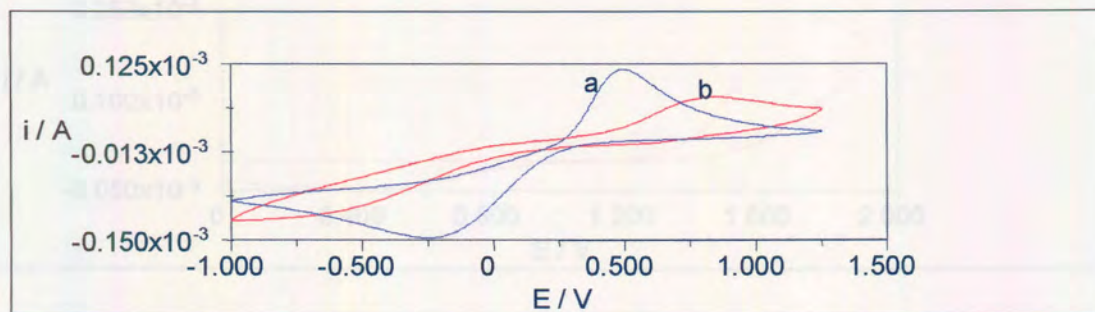


Figure 6.9 : Cyclic voltammogram for 20 mmol/L $K_3[Fe(CN)_6]$ in 1 mol/L KNO_3 and 0.1 mol/L Na_2SO_4 at a BDD electrode, (a) KNO_3 , (b) Na_2SO_4 - scan rate is 50 mV/s.

$E_i = 1.25$ V, scan direction = negative.

Using KNO_3 as a supporting electrolyte, a well-defined redox couple was obtained. However, when the supporting electrolyte was changed to Na_2SO_4 , the ΔE_p increased significantly, and the peak size decreased. This result indicates a slower electron transfer reaction when a lower concentration of supporting electrolyte and a larger, less mobile ion such as the SO_4^{2-} ion is used. This finding clearly shows that the rate of the electron transfer of $[Fe(CN)_6]^{3-}/[Fe(CN)_6]^{4-}$ at a BDD electrode, is dependent on the nature of the supporting electrolyte.

6.3.4 Cerium (III) sulphate

One of the major advantages of diamond electrodes over all other electrodes is its wide potential window and high overpotential for oxygen evolution by water oxidation in aqueous electrolyte solutions. The $\text{Ce}^{3+}/\text{Ce}^{4+}$ redox couple has a more positive standard potential than that of the $\text{H}_2\text{O}/\text{O}_2$ couple. It was for this reason that the $\text{Ce}^{3+}/\text{Ce}^{4+}$ redox couple was chosen in order to highlight the fact that boron-doped diamond has a wider potential window than most other electrodes, especially the glassy carbon electrode. The anodic oxidation of Ce^{3+} cannot be clearly observed at conventional electrodes such as glassy carbon, platinum, gold, and other electrodes because of the influence of oxygen evolution [21].

Figure 6.10 illustrates the cyclic voltammogram for 3 mmol/L $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L sulphuric acid (H_2SO_4) at a BDD electrode. A large, well-defined anodic peak is observed at approximately 1.7 V. This anodic peak represents the oxidation of Ce^{3+} to Ce^{4+} . A weak cathodic peak representing the reduction of Ce^{4+} can be seen at approximately 0.4 V on the reverse scan.

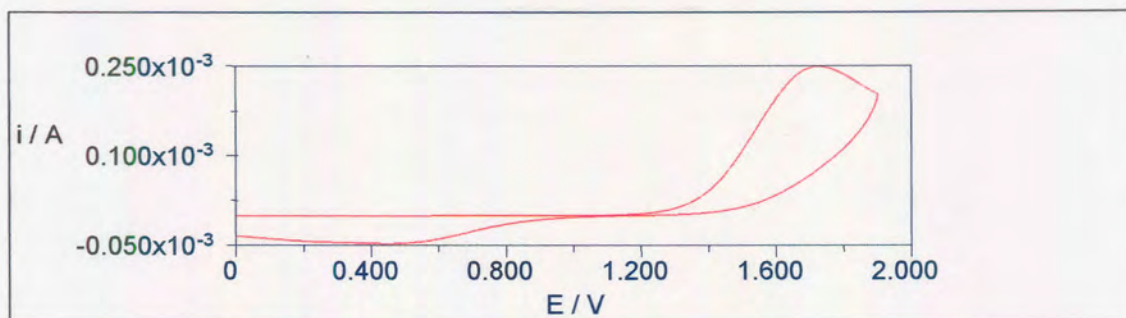


Figure 6.10 : Cyclic voltammogram for 3 mmol/L $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L H_2SO_4 at a BDD electrode - scan rate is 50 mV/s. $E_i = 0$ V, scan direction = positive.

Figure 6.11 shows the cyclic voltammograms for 3 mmol/L $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L H_2SO_4 , at a glassy carbon electrode as well as at a BDD electrode. For the glassy carbon electrode, water oxidation occurs at about 1.7 V. The electrolysis of water and oxygen evolution therefore negatively affects the resolution of the anodic peak. A weak cathodic peak occurring at 0.4 V is also observed using the glassy carbon electrode.

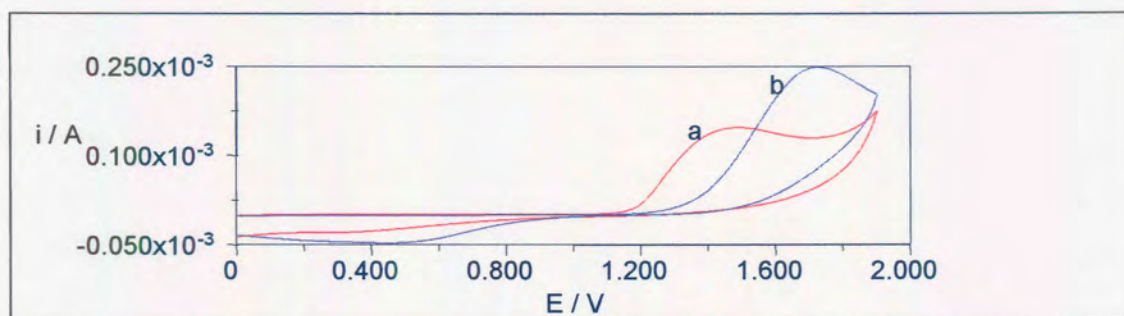


Figure 6.11: Cyclic voltammograms for 3 mmol/L $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L H_2SO_4 , at a glassy carbon electrode (a) as well as at a BDD electrode (b) - scan rate is 50 mV/s.

$E_i = 0$ V, scan direction = positive.

On comparison of the voltammograms representing the glassy carbon and diamond electrodes, it is quite clearly seen that a better anodic peak resolution independent of water electrolysis is obtained for the diamond electrode. This implies that the use of diamond electrodes in systems such as that of the $\text{Ce}^{3+}/\text{Ce}^{4+}$ redox couple, which require a large potential window, is favourable.

Figure 6.12 represents the effect of scan rate for 3 mmol/L $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L H_2SO_4 , at a BDD electrode. As the scan rate increases from 5 mV/s to 100 mV/s, the oxidation peak current also increases.

The cyclic voltammogram representing the reproducibility of the diamond electrode in cerium (III) sulphate is given in **Figure 6.13**. It can clearly be seen that as the number of scans increases, the oxidation peak potential shifts to more positive values, until it finally can no longer be resolved from the oxidation peak of water.

Initially, the surface of the diamond electrode [CVDBD4(3)] used in **Figure 6.13** was partially hydrophobic, i.e. contact angle measurements were between 72° - 88° (see **Section 4.6**). This indicates that the diamond electrode surface contained oxygen functionalities. Cerium is known to react with oxygen to form a cerium-oxygen polymer. Cerium could therefore have reacted with the oxygen functionalities on the surface of the diamond electrode to form a polymeric layer on the surface. This layer could then act as a passivating layer, hindering oxidation of Ce^{3+} ions at the electrode surface, and necessitating more and more positive potentials to achieve electron transfer, as seen in **Figure 6.13**.

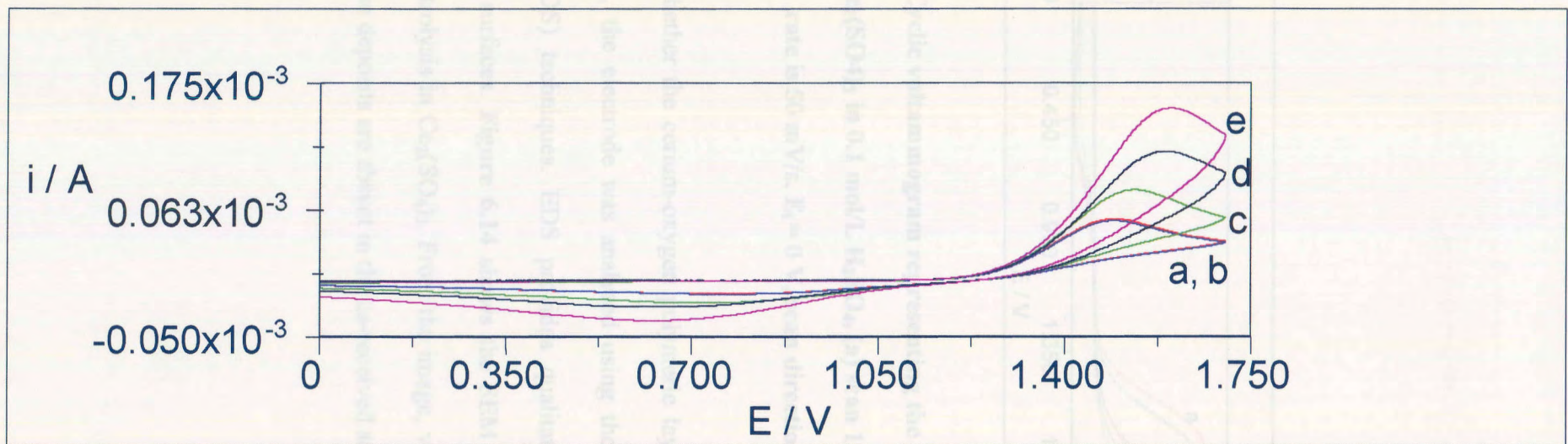


Figure 6.12 : Effect of scan rate for 3 mmol/L $Ce_2(SO_4)_3$ in 0.1 mol/L H_2SO_4 , at a BDD electrode, (a) 5 mV/s, (b) 10 mV/s, (c) 25 mV/s, (d) 50 mV/s, (e) 100 mV/s. $E_1 = 0$ V, scan direction = positive.

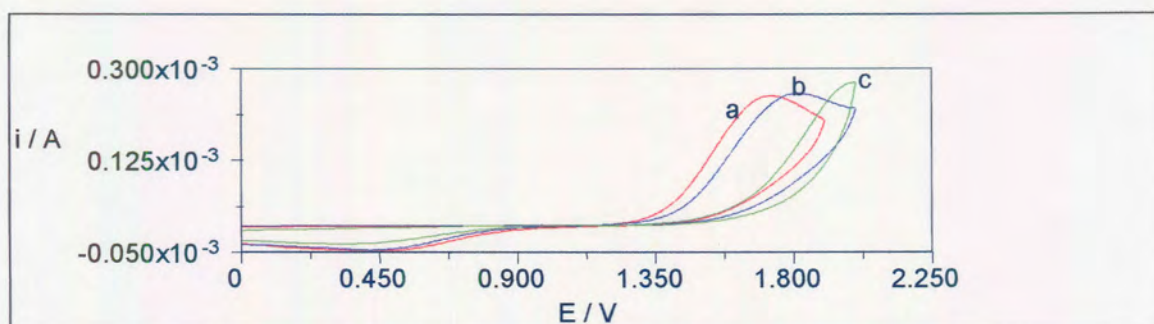


Figure 6.13 : Cyclic voltammogram representing the reproducibility of the BDD electrode for $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L H_2SO_4 , (a) scan 1, (b) scan 3, (c) scan 7 - scan rate is 50 mV/s. $E_1 = 0$ V, scan direction = positive

To investigate whether the cerium-oxygen polymeric layer had indeed formed on the electrode surface, the electrode was analysed using the SEM and energy dispersive spectroscopy (EDS) techniques. EDS provides qualitative information on elements present on solid surfaces. **Figure 6.14** shows the SEM image of the BDD electrode surface after electrolysis in $\text{Ce}_2(\text{SO}_4)_3$. From the image, various deposits or clusters can be observed. These deposits are absent in the as-received sample (**Figure 6.15**).

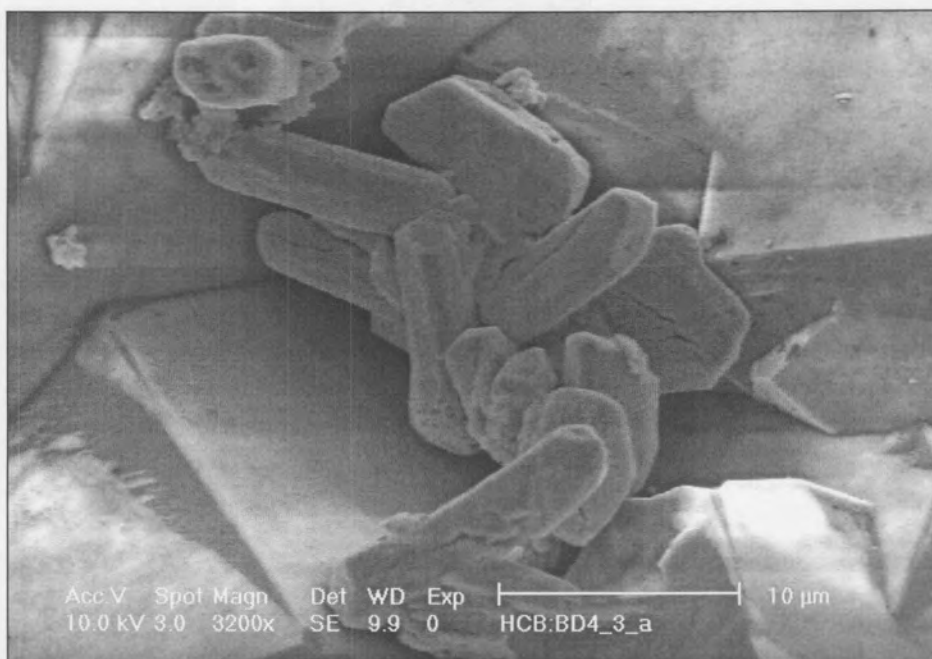


Figure 6.14 : SEM image of the BDD electrode surface after electroanalysis in $\text{Ce}_2(\text{SO}_4)_3$

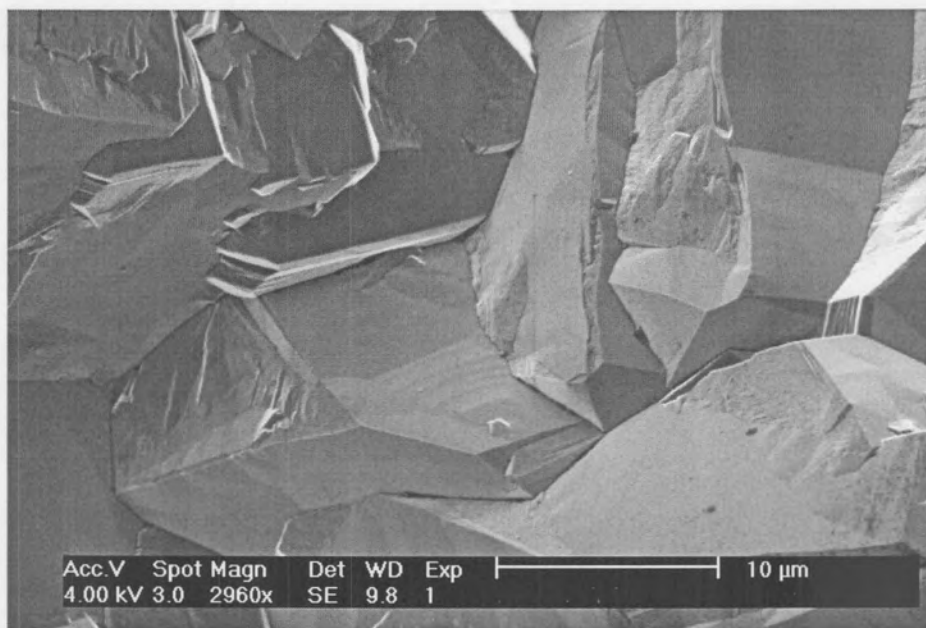


Figure 6.15 : SEM image of the as-received BDD electrode

EDS analysis identified these to be cerium deposits (**Figure 6.16**). The presence of cerium on the diamond electrode surface indicates that a cerium polymeric layer is likely to have formed on the electrode surface thereby hindering the further oxidation of Ce^{3+} to Ce^{4+} . In order to prevent the formation of the polymeric layer, the oxygen functionalities on the electrode surface would have to be removed via hydrogenation of the surface prior to analysis in $\text{Ce}_2(\text{SO}_4)_3$.

To test this theory, the diamond electrode (CVDBD4(3)) was acid cleaned in 20% H_2SO_4 and ultrasonically cleaned in hot water in order to remove the cerium deposits. The electrode was then hydrogenated in the TGA at 700°C under a flowing stream of 10% hydrogen in argon for 4 hours, in order to replace the oxygen atoms present on the surface with hydrogen atoms. A subsequent voltammetric scan for 3 mmol/L $\text{Ce}_2(\text{SO}_4)_3$ in 0.1 mol/L H_2SO_4 at the BDD electrode was carried out. A welcoming Ce^{3+} oxidation peak was evident at approximately 1.7 V. The peak potential and current remained constant on subsequent scans illustrating the reproducibility of the BDD electrode in $\text{Ce}_2(\text{SO}_4)_3$. However, the BDD electrode can only be reproducible in $\text{Ce}_2(\text{SO}_4)_3$ as long as the electrode surface is hydrophobic.

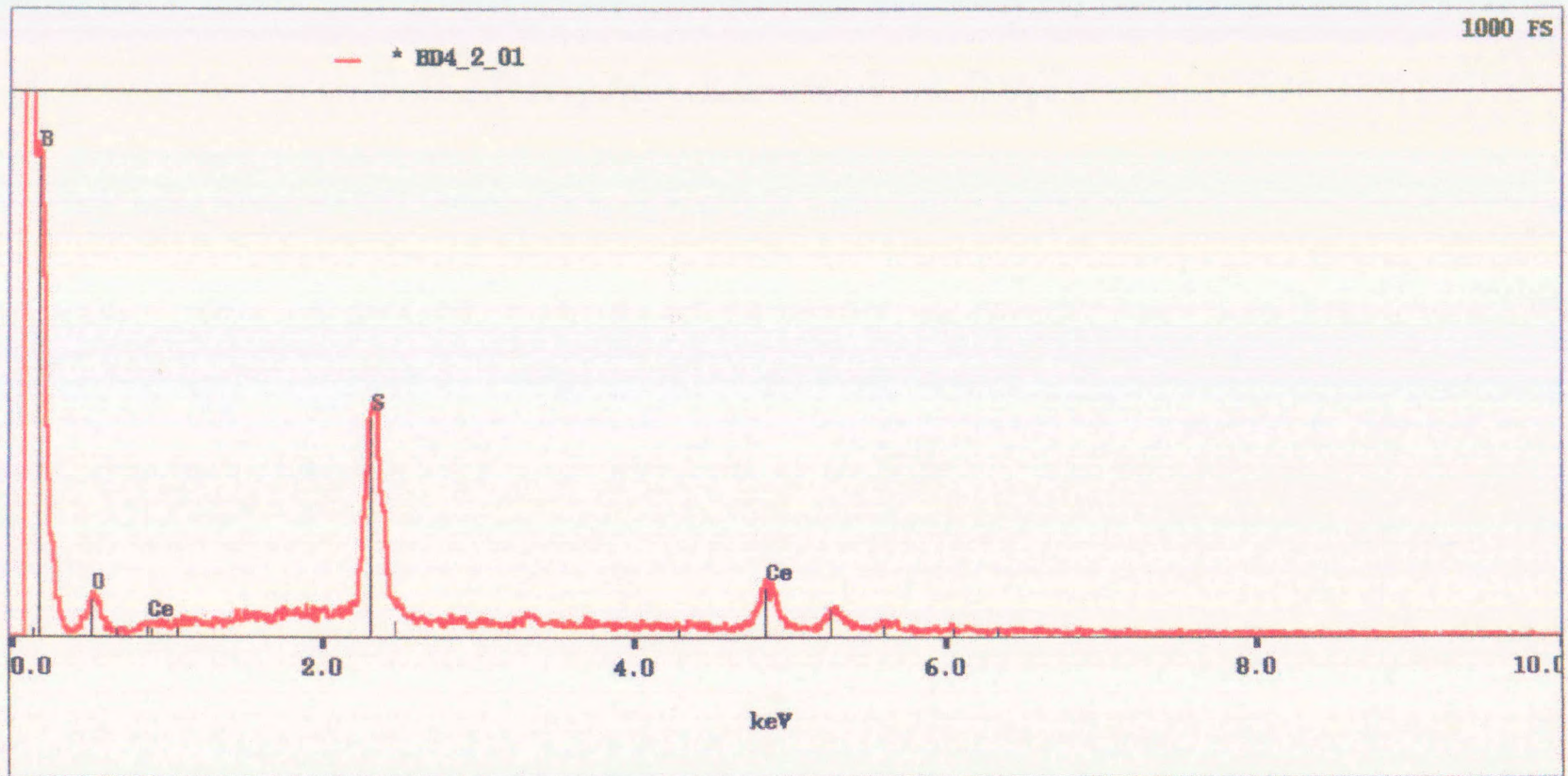


Figure 6.16 : EDS spectrum of CVDBD3 after electroanalysis in $\text{Ce}_2(\text{SO}_4)_3$

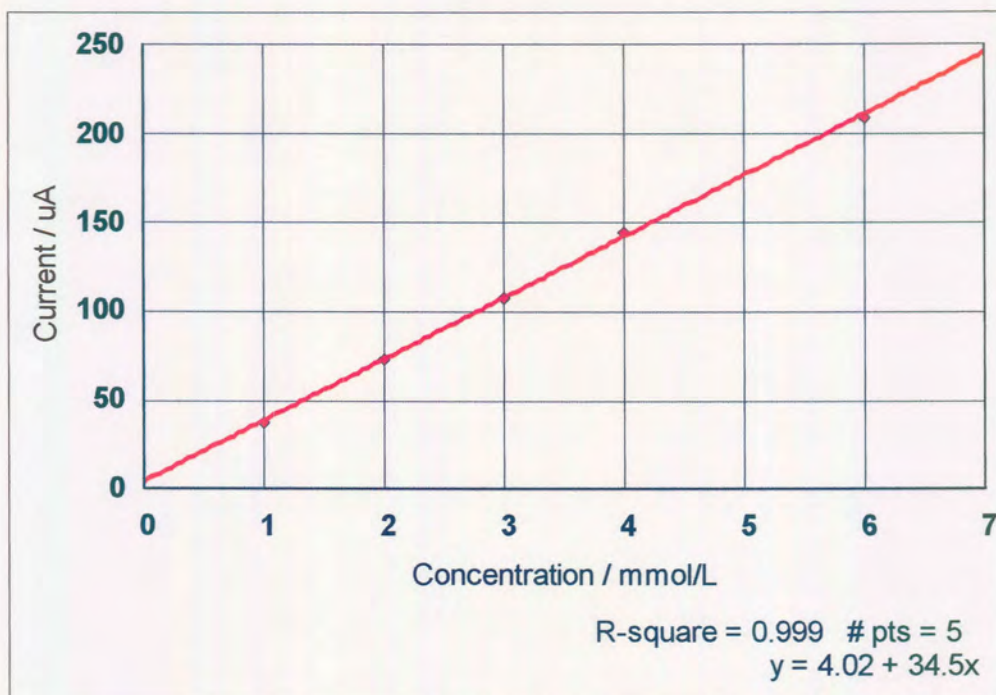


Figure 6.17 : Calibration plot for the BDD electrode

By varying the concentration of $\text{Ce}_2(\text{SO}_4)_3$ as a function of the respective oxidative peak current, a calibration plot was drawn for a partially hydrophobic BDD electrode (**Figure 6.17**) as well as the glassy carbon electrode (**Figure 6.18**). Percentage recoveries and the relative standard deviation of the samples measured are found in **Table 6.3**. The percentage recovery determines the ability of the electrode to analyse an unknown concentration. This value should be as close to 100% as possible in order to determine the reproducibility and stability of the electrode.

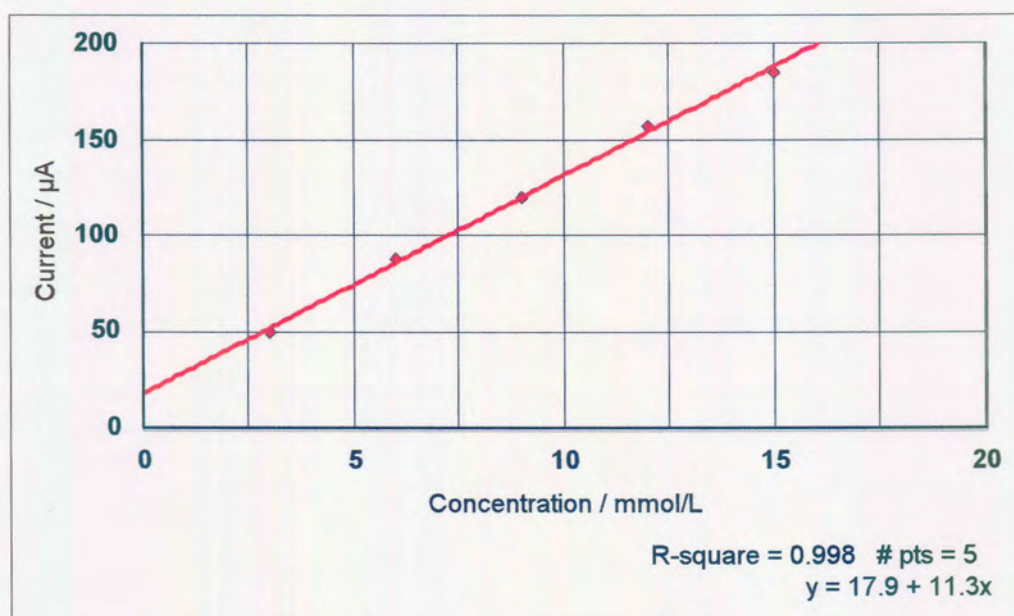


Figure 6.18 : Calibration plot for the glassy carbon electrode in $Ce_2(SO_4)_3$

Table 6.3 : Percentage recoveries and the relative standard deviation of the samples measured

Electrode	Sample	Percentage recovery	Average	Standard deviation
BDD	1	96.71	97.38	1.23
	2	98.80		
	3	96.63		
Glassy carbon	1	107.49	104.93	2.41
	2	102.70		
	3	104.59		

Both the BDD electrode and the glassy carbon electrode were linear in the concentration range measured. The linear regression (R-square) value for the diamond electrode (0.999) was found to be higher than that of the glassy carbon electrode (0.998). If all the points in a calibration graph lay in a straight line, then the R-square value would be 1. The R-square value is therefore regarded as a measure of the quality of the calibration plot. Thus, it can be clearly seen that the BDD electrode produces a better quality calibration plot than the glassy carbon electrode. Furthermore, the percentage recovery values (**Table 6.3**) for the BDD electrode is much closer to 100% than the glassy carbon electrode. This illustrates diamond to be preferable to glassy carbon for the quantitative analysis of $\text{Ce}_2(\text{SO}_4)_3$.

6.4 Conclusion

The electrochemical behaviour of diamond electrodes of varying boron concentrations was investigated. It was found that, as the boron concentration of the BDD increases, the potential window decreases. However, the potential window for the highest BDD electrode (CVDBD6) is still wider than that of the glassy carbon electrode, thereby confirming the advantage that the BDD electrode has over glassy carbon when investigating aqueous systems.

The peak separation of the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple (690 mV) was found to be an order of magnitude larger than that of the glassy carbon electrode (61 mV). It is suggested that oxygen termination of the BDD electrode surface inhibits the electron

transfer of the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple. It can thus be assumed that in order to reduce the peak separation, the surface of the electrode will have to be made hydrophobic.

The BDD electrode proved to be more effective (wider potential window, therefore better peak resolution) for the semi-quantitative analysis of cerium (III) sulphate, than the glassy carbon electrode. This is evidenced by the BDD electrode producing a better quality calibration plot and percentage recovery values than the glassy carbon electrode.

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CHAPTER 7

Electroanalysis of organic systems – dopamine in the presence of ascorbic acid

7.1 Introduction

Dopamine belongs to the family of catecholamines and is found in the extracellular fluid (ECF) in the brain. It is synthesised in the body by a two step process from the 1-amino acid tyrosine. The hydroxylation of tyrosine by tyrosine hydroxylase results in the formation of the catechol dihydroxyphenylalanine (DOPA). DOPA is then decarboxylated by the non-specific enzyme 1-amino acid decarboxylase to dopamine. **Figure 7.1** illustrates the reactions involved in the synthesis of dopamine.

Dopamine is a biologically important compound, which plays a key role in neurotransmission [2,3]. Neurotransmission is the conversion of an electrical impulse to a chemical event and subsequently to a second electrical event [4]. The mammalian brain contains approximately 10^{11} nerve cells or neurons, which assimilate and process information.

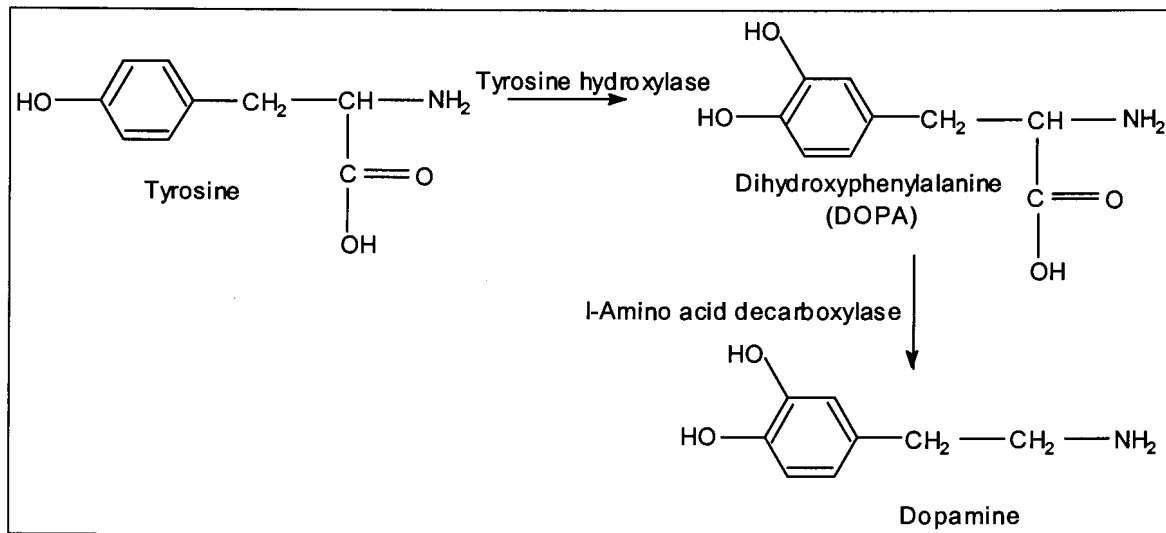


Figure 7.1 : Synthesis of Dopamine [1]

Figure 7.2 represents a schematic diagram of a neuron. Neurons receive information from cells via a treelike network called dendrites and then transfer this information in the form of electrical impulses along a cablelike axon to other nerves. These electrical impulses cause the release of a chemical messenger from a storage vesicle in the axon terminal. The chemical messenger, also called a neurotransmitter, travels across a synapse to bind to a postsynaptic receptor protein. The act of binding to the receptor sets in motion a series of events, which eventually brings about a change in the electrical state of the postsynaptic cell. As a chemical messenger, dopamine is similar to adrenaline. Dopamine affects brain processes that control movement, emotional response, and ability to experience pleasure and pain.

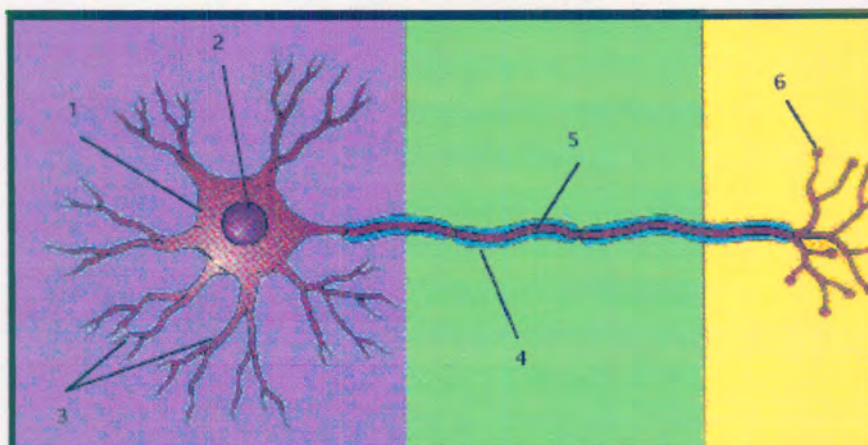
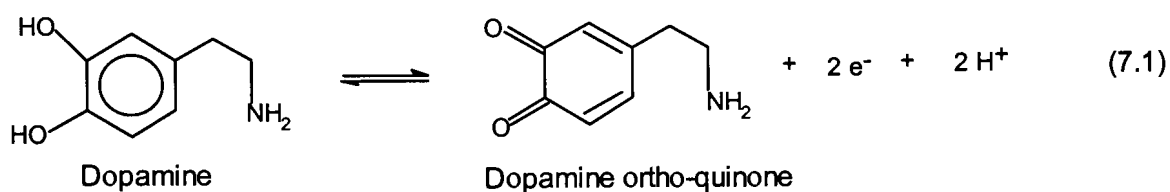


Figure 7.2 : Schematic diagram of a neuron - 1 = Cell Body, 2 = Nucleus, 3 = Dendrites, 4 = Axon, 5 = Myelin Sheath, 6 = Axon terminals [5]

Neurons containing the neurotransmitter dopamine are clustered in the midbrain in an area called the substantia nigra [6]. A deficiency of dopamine in the brain results in Parkinson's disease and an excess of dopamine results in schizophrenia. Dopamine is also one of the neurotransmitters that play a major role in the addiction to certain drugs such as cocaine. It is therefore vital that the presence of dopamine be efficiently detected in order to cure these dreaded diseases.

One of the methods used to detect for the presence of dopamine is voltammetry. Since dopamine, a neurotransmitter, is electroactive, it can be oxidised at an applied potential. The resulting current can then be measured and the concentration of analyte determined. The electro-oxidation of dopamine is illustrated in **Equation 7.1** [7].



The ideal technique for the detection of dopamine would be *in vivo* voltammetry. This technique involves the insertion of an electrode into the brain of the subject, which enables online monitoring of the dopamine levels. Ralph Adams and his colleagues [8] first attempted this technique by inserting a carbon paste electrode, having a diameter of approximately 250 μm , into the brain of a rat to detect electroactive materials *in vivo*. Although oxidation currents were obtained, this technique was non-specific, unreliable and irreproducible.

At the time that Adams was doing his pioneering work in the United States, Francois Gonon and his colleagues in France had been investigating electrodes made of other carbon-based materials for detecting catecholamines [4,7]. They used carbon fibre electrodes made from glass pipettes and these electrodes were vastly reduced in size (having a diameter of approximately 10 μm). The implantation of an electrode is traumatic to brain cells, and obviously the smaller the electrode the less damage occurs. The size of the electrode can also alter the species detected [4]. For example, larger electrodes may disrupt the blood-brain barrier in the brain and allow blood-borne compounds to be detected, or they can evoke a local gliosis, which is a tissue reaction to a foreign object.

The carbon fibre electrode gave a considerable improvement in terms of reproducibility and reliability over the carbon paste electrodes. This electrode, however, like the carbon paste electrode was not selective in detecting only the catecholamine of interest. Although oxidation currents were produced, several other oxidizable molecules, notably ascorbic acid which coexists with the catecholamines in the ECF in the brain [9], obscured the currents from catecholamines such as dopamine. Because the oxidation potential of ascorbic acid is similar to that of dopamine, selective detection of dopamine became a challenging task. It was subsequently found that, in order to detect catecholamines such as dopamine in the presence of ascorbic acid and other acidic metabolites, an anionic Nafion membrane could be used to repel these acids, as reported by Ralph Adams [10].

Since the carbon paste and carbon fibre electrodes were used to detect the presence of catecholamines in brain tissue, it can be assumed that boron-doped diamond (BDD) electrodes can do the same, if not better. BDD electrodes, however, have a significant advantage over the carbon paste and carbon fibre electrodes, in that they are more resistant to fouling and more biocompatible than these carbon electrodes. Another very powerful advantage of BDD electrodes is their low baseline current, which enables them to detect extremely low concentrations of catecholamines such as dopamine in the brain tissue. These advantages make BDD electrodes potentially very useful as biosensors for the *in vivo* or *in vitro* detection of analytes.

7.1.1 Biocompatibility studies of chemically vapour deposited diamond (CVDD)

The surface of an implant (such as a biosensor) should not only be inert in the body, but should also stimulate cell proliferation and differentiation. Two biocompatibility studies of CVD diamond are reported in the literature. In the first, an internal study was commissioned by De Beers and carried out by researchers at De Montfort University (Leicester, United Kingdom), which compared the performance of boron-doped CVD diamond with that of porous silicon [11].

Three indicators of biocompatibility were measured : the deposition of protein, the deposition of hydroxyapatite (the inorganic constituent of bone tissue), and cell proliferation and morphology. The deposition of protein is essential for cells to become firmly anchored at the implant surface. Deposition of hydroxyapatite is a standard indicator of the compatibility of material for bone implants such as hip replacements, and may negatively affect the adherence and function of cells. Lastly, cell proliferation at an implant surface should be normal or above average, and the cells should adhere well to the surface and have a normal morphology.

It was found that hydroxyapatite deposited more slowly on the diamond sample than on porous silicon, and that the hydroxyapatite deposited more slowly than the protein layer. This is an excellent indication that boron-doped diamond would be a suitable candidate for a biomedical implant. Furthermore, B50 cells (transformed rat neuronal tissue) were

found to grow vigorously on the diamond surface, with good adhesion and normal morphology.

In the second study [12], the performance of a CVD (undoped) diamond coating was compared with that of pure titanium (titanium alloys are commonly used in hip replacements). Cell proliferation at the diamond coating was found to be equal to that at a titanium surface. However, attachment of cells at the diamond surface was retarded in comparison with titanium. Two reasons are proposed : firstly, the diamond surface was considerably rougher than the titanium surface, and secondly, the diamond surface was hydrophobic, which is expected to seriously interfere with protein adsorption and therefore cell adhesion. This lack of adhesion was not encountered in the De Montfort study, and may be due to the presence of boron dopant affecting the surface properties.

In conclusion, indications are that boron-doped diamond is an excellent biomaterial. However, care should be taken that the surface state has the right degree of hydrophilic/hydrophobic nature to balance the requirements for electroactivity and biocompatibility.

Since diamond is biocompatible, it would be an ideal biosensor for the *in vivo* or *in vitro* detection of catecholamines such as dopamine. However, there are many requirements that an electrode material has to meet in order to become an effective biosensor [13-17].

7.1.2 Commercial requirements and benefits of biosensors

Commercial biosensors must be accurate and repeatable [18-21]. In order to accomplish this, the electrode surface must be resistant to fouling, since any adsorbed species (for example quinones) on the surface of the electrode will result in a diminished level of accuracy and repeatability of the electrode.

The sensitivity of a biosensor is very important [22]. Biosensors must have very low detection limits when used to measure compounds in the body fluid, as the concentration levels are extremely low. Biosensors are specific and sensitive in that they can detect low concentrations of analytes which are present in a complex matrix. Another necessary requirement for a biosensor is its speed of response. If a biosensor is to be used to measure neurotransmitters in the ECF, where there is a major concentration fluctuation due to the speed at which chemicals are transferred from synapse to synapse, the biosensor has to respond quickly in order to detect these changes.

Other requirements for biosensors include reliability, physical robustness, low running costs and long lifetime. If they are physically robust, they can very easily be used by unskilled personnel.

Biosensors are potentially more efficient and less time consuming. Using conventional methods, rapid diagnosis and treatment cannot be carried out immediately because there is a long time delay between when the sample is taken and when the result is obtained

from a central laboratory. Biosensors, because of their excellent response time, are able to produce immediate results.

A vital requirement for a biosensor is biocompatibility. A biosensor must not contaminate nor poison the sample host system when inserted into the body. For this reason, diamond, being a purely carbon-based material, is ideal for use as a biosensor.

The investigations detailed below were carried out in order to evaluate the potential use of De Beers BDD electrodes as a biosensor for the *in vivo* detection of dopamine in brain tissue. The electrochemical behaviour of De Beers BDD electrodes was compared to that of other boron-doped diamond electrodes used in the literature, and to that of the glassy carbon electrode, in the dopamine and ascorbic acid systems.

7.2 Experimental

A three electrode cell consisting of a BDD working electrode, a platinum counter electrode and a Ag/AgCl reference electrode were used for the voltammetry investigation. All the chemicals used were analytical grade. Ascorbic acid (AA) and perchloric acid (HClO₄) were obtained from Saarchem. Dopamine was obtained from Sigma Aldrich. The supporting electrolyte (HClO₄) was diluted to the desired concentration using ultrapure Millipore water (18 μΩ.cm). Stock solutions of dopamine and AA were prepared using the supporting electrolyte. Each solution was deaerated prior to the electrochemical analysis by purging the solution with helium for 5 minutes.

All the electrochemical measurements were performed at room temperature (25°C). The surface areas of the glassy carbon and BDD electrodes were 0.071 cm² and 0.15 cm² respectively. CVDBD4, having a boron concentration of 543 mg/L, was used as the electrode material to investigate the dopamine and AA systems.

Prior to each electrochemical measurement, the glassy carbon electrode was polished for approximately 20 minutes in order to clean the electrode surface and remove any adsorbed organic species. An electrode holder was made from Perspex, consisting of a flat plate with a hole drilled in it through which the glassy carbon electrode was inserted during polishing. This was done in order to ensure that the electrode surface remained flat on the polishing cloth so that no curvature of the surface was introduced during the polishing process. The glassy carbon electrode was polished with a Leco polisher, using 6 µm diamond suspension, followed by 1 µm diamond suspension. The electrode surface was cleaned thoroughly with deionised water after polishing.

7.3 Results and discussion

7.3.1 Dopamine

Figure 7.3 illustrates the cyclic voltammogram for the oxidation of 0.2 mmol/L dopamine in 0.1 mol/L HClO₄ at a BDD electrode. A well-defined redox couple is observed with the oxidation of dopamine occurring at approximately 0.8 V and the ensuing reduction of dopamine occurring at about 0.2 V. The voltammogram shown in **Figure 7.3** was compared to that of literature. Fujishima et. al. [9] reported an oxidation

peak current density of $40 \mu\text{A}/\text{cm}^2$ for 0.1 mmol/L dopamine in 0.1 mol/L HClO_4 . The corresponding oxidation peak potential was reported to be approximately 0.8 V . These values are similar to the experimental values obtained for 0.2 mmol/L dopamine in 0.1 mol/L HClO_4 , which gave an oxidation peak current density of $49 \mu\text{A}/\text{cm}^2$ and an oxidation peak potential of approximately 0.8 V . The observed current was greater than that reported in literature due to a higher dopamine concentration. The shape of the experimentally obtained cyclic voltammogram (**Figure 7.3**) was identical to that depicted in literature [23].

The effect of deaerating the solution before any electrochemical analysis was also investigated. **Figure 7.4** shows the cyclic voltammograms for 0.2 mmol/L dopamine in 0.1 mol/L HClO_4 in a deaerated as well as in an aerated solution. Both cyclic voltammograms yielded a dopamine oxidation peak current of approximately $5 \mu\text{A}$. The effect of purging the solution with helium was negligible.

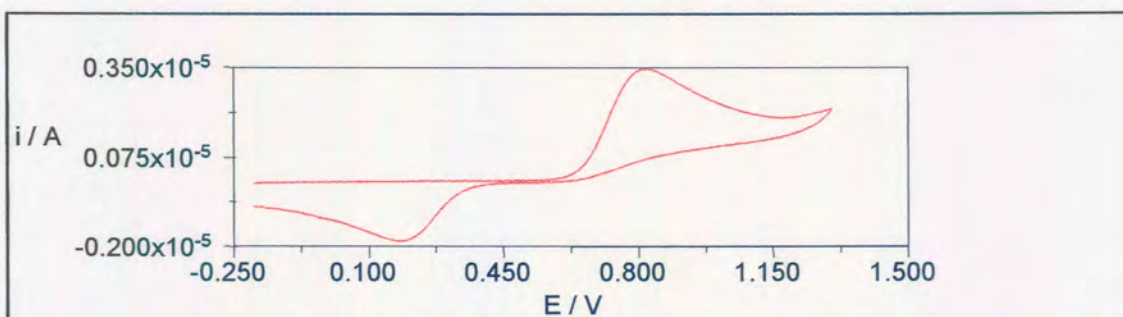


Figure 7.3 : Cyclic voltammogram for the oxidation of 0.2 mmol/L dopamine in 0.1 mol/L HClO_4 at a BDD electrode – scan rate is 50 mV/s . $E_i = -0.2 \text{ V}$, scan direction = positive

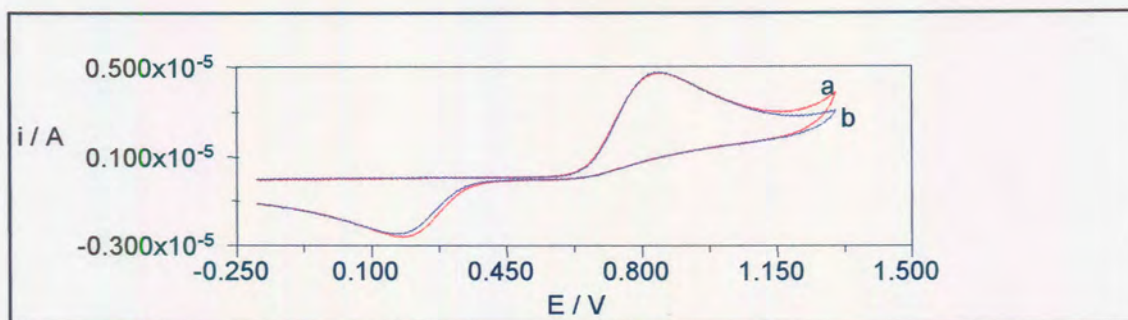


Figure 7.4 : Cyclic voltammograms for 0.2 mmol/L dopamine in 0.1 mol/L HClO₄ in an aerated (a), as well as in an deaerated (b) solution – scan rate is 100 mV/s.

$E_i = -0.2$ V, scan direction = positive.

Linear sweep voltammetry was employed to investigate the reproducibility of the BDD electrode for 0.4 mmol/L dopamine in 0.1 mol/L HClO₄ (Figure 7.5). The oxidation peak current for both the first and the second scan was found to be 9.8 μ A. This shows the reproducibility of the BDD electrode in the dopamine system to be quite acceptable.

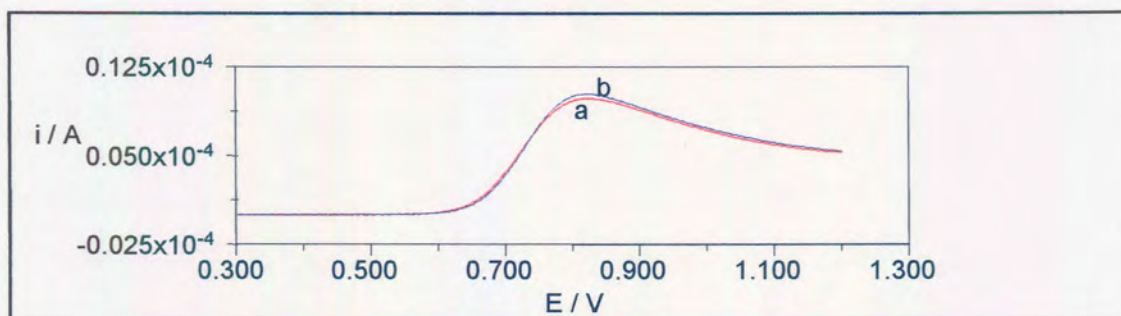


Figure 7.5 : Linear sweep voltammogram illustrating the reproducibility of the BDD electrode for 0.4 mmol/L dopamine in 0.1 mol/L HClO₄, (a) scan 1, (b) scan 2 – scan rate is 100 mV/s. $E_i = 0.3$ V, scan direction = positive.

Figure 7.6 represents the cyclic voltammograms for 0.2 mmol/L dopamine in 0.1 mol/L HClO₄ at a glassy carbon electrode and at a BDD electrode respectively. The separation between the oxidation peak and the reduction peak (peak separation, ΔE_p) for the BDD electrode was calculated to be 600 mV, whilst the ΔE_p for the glassy carbon electrode was calculated to be 56 mV. Granger et. al. [23] reported a ΔE_p of 480 mV for dopamine and a ΔE_p in the range of 125-175 mV at a polished glassy carbon electrode. Dopamine appears to therefore exhibit much more electrochemical irreversibility at the BDD electrode than at the glassy carbon electrode, as evidenced by the relatively large ΔE_p .

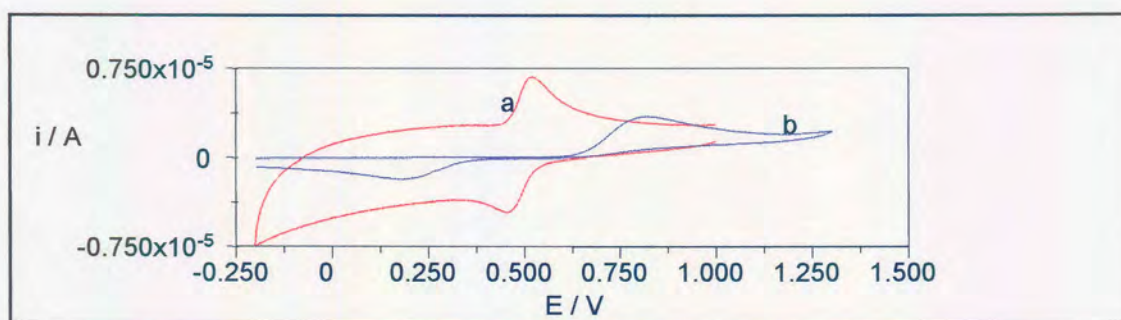


Figure 7.6 : Cyclic voltammograms for 0.2 mmol/L dopamine in 0.1 mol/L HClO₄ at a glassy carbon electrode (a), and BDD electrode (b) respectively – scan rate is 50 mV/s. $E_i = -0.2$ V and scan direction = positive.

The large ΔE_p for the BDD electrode can be ascribed to a slow electron transfer reaction occurring at the electrode surface. Swain et. al. [23] believe that a lack of adsorption of dopamine at the BDD electrode surface is one of the reasons for the relatively slow electrode kinetics. In addition, Hunt-Duvall and McCreery have presented a detailed

study showing that low ΔE_p values correlate with catechol and catecholamine adsorption on glassy carbon, and surface treatments that decreased adsorption also increased ΔE_p [24].

The apparent lack of adsorption of catecholamines such as dopamine on the BDD electrode surface demonstrates the high degree of resistance that BDD electrodes possess towards fouling. This feature is again one of the many advantages that BDD electrodes possess over most other electrode materials, for example glassy carbon. The surface of the glassy carbon electrode, like many solid electrodes, has a tendency to become deactivated over time when it is exposed to the atmosphere or working solution. The problem is more severe when biological sample solutions are used [25].

A further conspicuous feature in the above cyclic voltammogram (**Figure 7.6**), is the huge baseline current associated with the glassy carbon electrode. This observed baseline can be attributed to the sp^2 nature of the carbon in the glassy carbon electrode. BDD electrodes, however, possess a very low baseline current and this may be ascribed to several factors [26] :

- (a) Since the as-grown BDD electrodes are hydrophobic or at least partially hydrophobic in nature, there is a lack of electroactive carbon-oxygen functional groups on the electrode surface. The presence of these oxygen functional groups would increase the baseline current associated with the electrode.

(b) Diamond possesses a low density of surface states at the Fermi level as compared to that of a metal. Thus, the baseline current associated with the BDD electrode is expected to be much lower than that associated with a corresponding metal electrode.

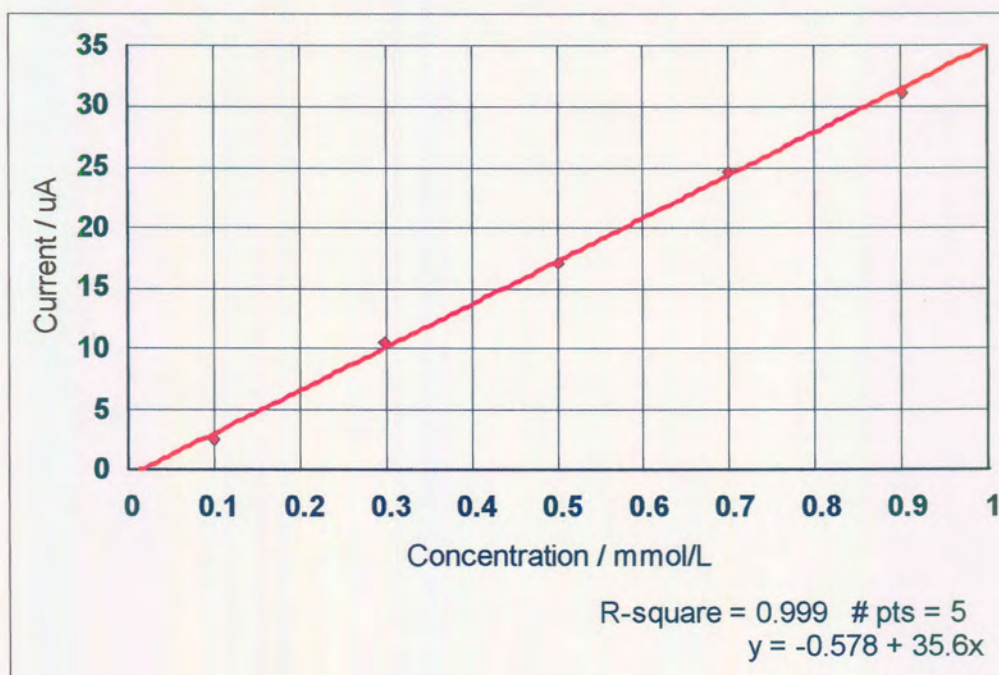


Figure 7.7 : Calibration plot for dopamine at a BDD electrode

Figure 7.7 and **Figure 7.8** illustrate the calibration plot for dopamine at a BDD electrode and at a glassy carbon electrode respectively. Percentage recoveries and the relative standard deviation of the samples measured are found in **Table 7.2**.

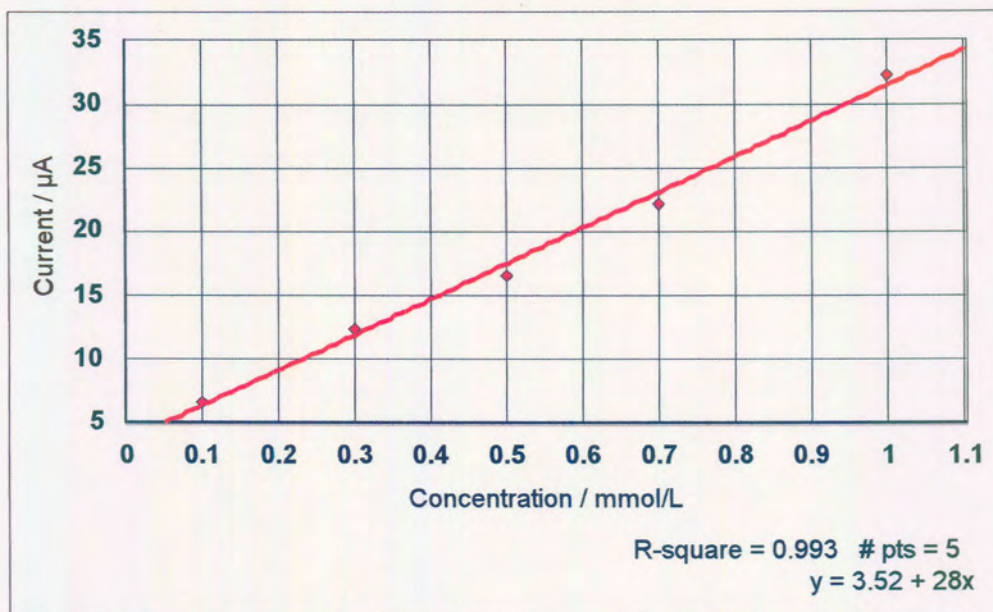


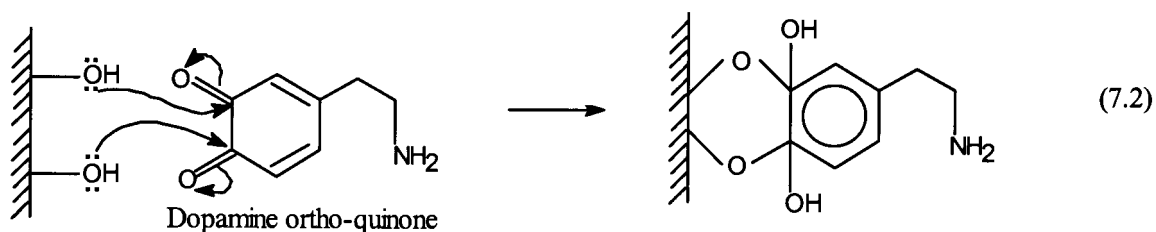
Figure 7.8 : Calibration plot for dopamine at a glassy carbon electrode

Table 7.2 : Percentage recoveries and the relative standard deviation of the samples measured

Electrode	Sample	Percentage recovery	Average	Standard deviation
BDD	1	101.10	98.81	2.21
	2	98.63		
	3	96.69		
Glassy carbon	1	86.52	102.96	14.72
	2	114.94		
	3	107.43		

Both the BDD electrode and the glassy carbon electrode responses were linear in the concentration range measured. The linear regression (R-square) value for the diamond electrode (0.999) was found to be higher than that of the glassy carbon electrode (0.993). It is evident from **Table 7.2** that the percentage recovery values for the BDD electrode is much closer to 100% than the glassy carbon electrode.

The low R-square and high variation in percentage recovery values for the glassy carbon electrode may be attributed to the fouling of the electrode surface by dopamine. According to **Equation 7.1**, the product from the electro-oxidation of dopamine is dopamine ortho-quinone. Quinones have a tendency to adsorb onto surfaces that have a high degree of oxygen-carbonyl functionalities, and these may be expected to form on glassy carbon during the anodic stage of the voltammetry cycle. **Equation 7.2** [27] illustrates a possible mechanism for dopamine ortho-quinone chemisorption at a glassy carbon electrode surface. This mechanism could explain why the glassy carbon electrode had to be polished after every dopamine determination during the dopamine calibration.



From the calibration graphs and the percentage recovery values, it can be deduced that BDD is a better electrode material for the determination of dopamine than the glassy

carbon electrode. This is accredited to the lack of adsorption of dopamine or its electro-oxidation product on the BDD electrode surface. It must be noted that, although BDD electrodes are preferred in the determination of dopamine, the rate of electron transfer at the electrode surface is slow.

In the determination of dopamine, the BDD electrode was found to be stable over a three month period. During this period, the electrode was stored in contact with air. Using chronoamperometry, the detection limit for dopamine at the BDD electrode was determined to be 0.56 $\mu\text{mol/L}$. Fujishima et. al. [9] reported a dopamine detection limit of 50 nmol/L, which is among the lowest values ever reported. The resistivity of the BDD electrode used by Fujishima et. al. was of the order of $10^{-3} \Omega\cdot\text{cm}$, whilst the resistivity of the CVDBD4 electrode employed for the detection of dopamine was calculated to be approximately 17 $\Omega\cdot\text{cm}$. This indicates that the BDD diamond electrode used by Fujishima et. al. must have been “heavily doped”. Therefore, in order to decrease the observed detection limit for dopamine, a higher level of boron dopant (probably in the order of 10^{21} boron atoms. cm^{-3}) is necessary.

Dopamine coexists in the ECF in the brain with ascorbic acid. The concentration of ascorbic acid and other acidic metabolites in the ECF is much greater than that of the catechols and catecholamines [4]. It is therefore essential that a biosensor should be capable of detecting low concentrations of dopamine in the presence of much larger concentrations of ascorbic acid.

7.3.2 Ascorbic acid

Ascorbic acid (AA) normally occurs in the extracellular fluid (ECF) in the brain. With its high concentration and ease of oxidation, AA is probably the simplest molecule to detect in the brain ECF. The structure of AA is found in **Figure 6.4**. Using voltammetry techniques, the oxidation of ascorbic acid was investigated at a glassy carbon electrode as well as at a boron-doped diamond electrode.

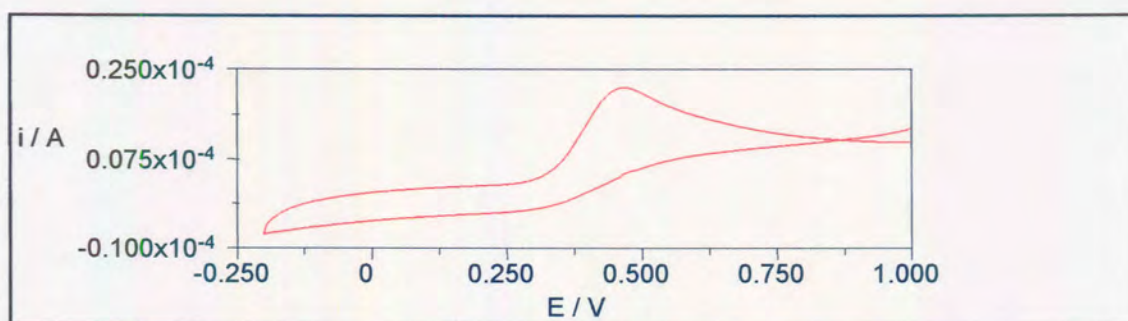


Figure 7.9 : Cyclic voltammogram for 2 mmol/L AA in 0.1 mol/L HClO₄ at a glassy carbon electrode - scan rate is 50 mV/s. E_i = -0.2 V, scan direction = positive

Figure 7.9 shows the cyclic voltammogram for 2 mmol/L AA in 0.1 mol/L HClO₄ obtained at a glassy carbon electrode. The oxidation peak of AA is seen at a potential of approximately 0.46 V. It must be noted that there is no reduction peak on the reverse scan, that is the cyclic voltammogram for AA is totally irreversible. This implies that the product of the electron transfer reaction for AA is stable, i.e. not electroactive.

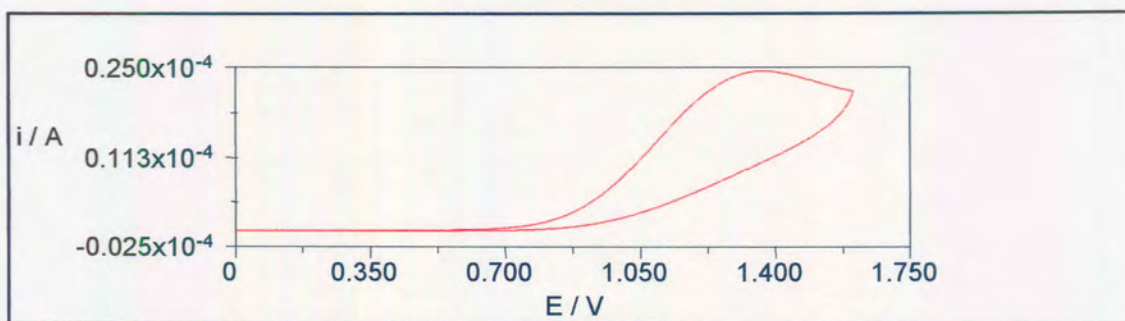


Figure 7.10 : Cyclic voltammogram for 2 mmol/L AA in 0.1 mol/L HClO₄ at a BDD electrode - scan rate is 50 mV/s. $E_i = 0$ V, scan direction = positive

The cyclic voltammogram representing the oxidation of 2 mmol/L AA in 0.1 mol/L HClO₄ at a BDD electrode is shown in **Figure 7.10**. If the surface of the BDD electrode was very hydrophobic, the oxidation peak potential of AA would have occurred at approximately 0.8 V, according to Fujishima et. al. [9]. However, since CVDBD4 is partially hydrophobic, a well-defined AA oxidation peak is evident at approximately 1.3 V. The cyclic voltammogram for AA at a BDD electrode is again entirely irreversible. The reproducibility of the BDD electrode in the AA system was investigated (**Figure 7.11**). It can be deduced from **Figure 7.11** that the BDD electrode is fairly reproducible in the AA system.

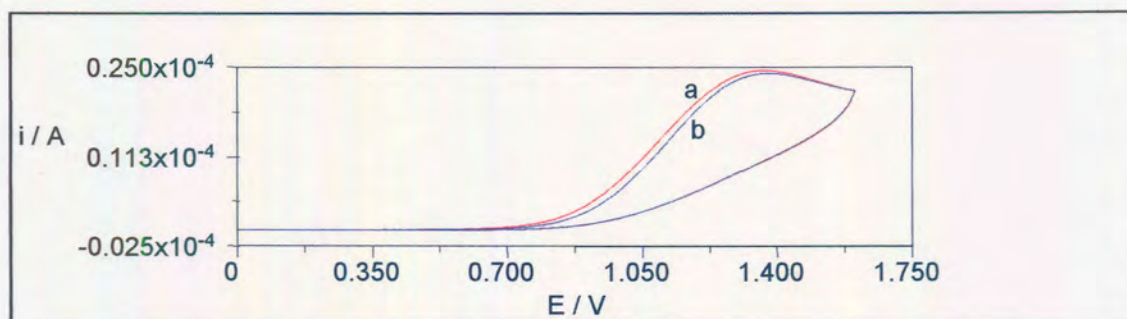


Figure 7.11 : Cyclic voltammogram illustrating the reproducibility of the BDD electrode for 2 mmol/L AA in 0.1 mol/L HClO₄; (a) first scan, (b) second scan - scan rate is 50 mV/s. $E_i = 0$ V, scan direction = positive

Figure 7.12 and **Figure 7.13** illustrate the calibration plot for AA at a BDD electrode and at a glassy carbon electrode respectively. Percentage recoveries and the relative standard deviation of the samples measured are found in **Table 7.3**. The glassy carbon electrode had to be polished between measurements during the calibration in order to remove any adsorbed species present on the electrode surface.

The BDD electrode and the glassy carbon electrode calibration curves were found to be linear in the concentration range measured. The R-square value for the diamond electrode (1) was again found to be higher than that of the glassy carbon electrode (0.998). Also, the percentage recovery values for AA at the BDD electrode were much closer to 100% than at the glassy carbon electrode. These positive results for the BDD electrode can be attributed to the lack of adsorption of AA on the electrode surface.

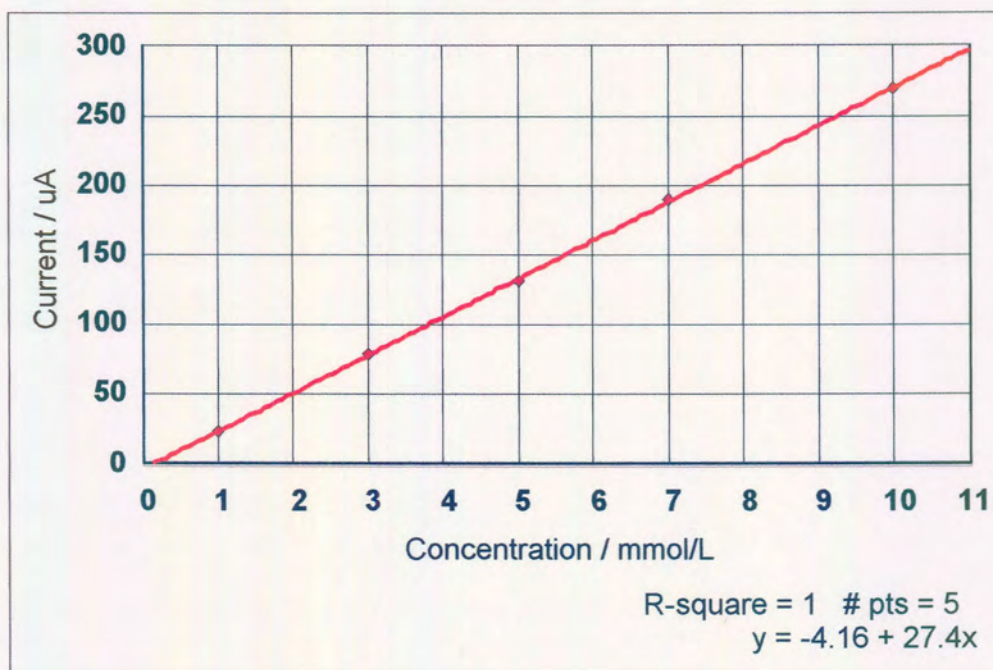


Figure 7.12 : Calibration plot for AA at a BDD electrode

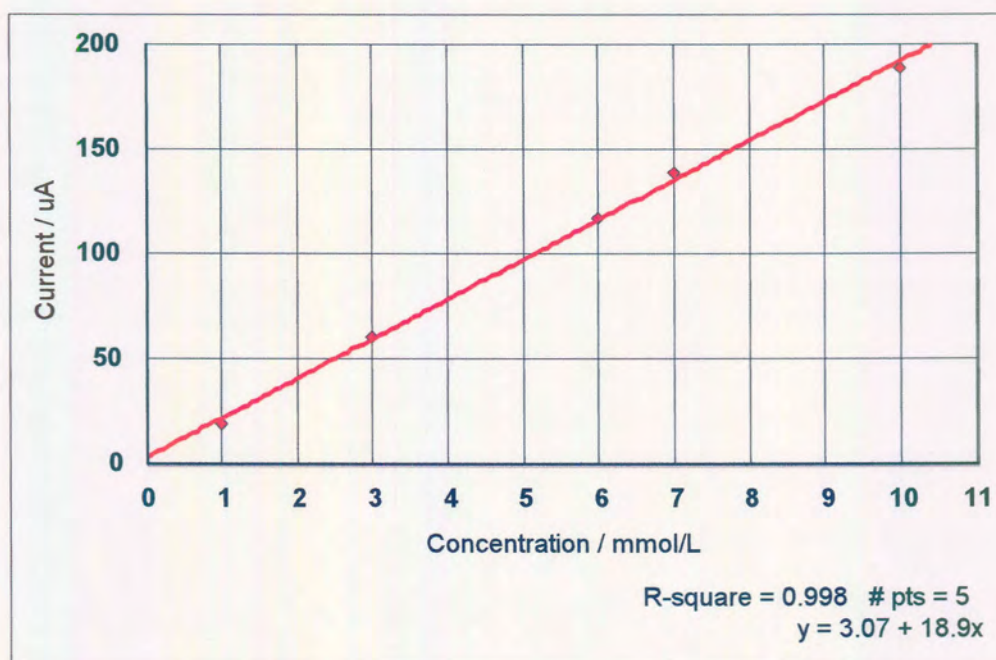


Figure 7.13 : Calibration plot for AA at a glassy carbon electrode

Table 7.3 : Percentage recoveries and the relative standard deviation of the samples measured

Electrode	Sample	Percentage recovery	Average	Standard deviation
BDD	1	100.53	100.36	0.49
	2	100.75		
	3	99.82		
Glassy carbon	1	111.27	107.14	3.88
	2	103.56		
	3	106.59		

The BDD electrode was also found to be stable over a three month period when used for the detection of AA. During this period, the electrode was exposed to a laboratory environment. The detection limit for AA at the BDD electrode was determined to be 20 nmol/L.

In the *in vivo* determination of dopamine, ascorbic acid is a major interferent. It is therefore essential to achieve good resolution between the responses for AA and dopamine. This may be accomplished using two well known techniques [4] :

- (a) By using an anionic Nafion membrane at the electrode surface, and a pH above that of the pKa for AA, the negatively charged ascorbic acid should be repelled from the surface while the neutral dopamine molecule passes through the membrane and is

detected at the electrode surface. The drawback of using such a membrane is a decrease in the response time of the electrode. This is a disadvantage if the electrode is to be used as an implantable biosensor giving fast feedback on fluctuating dopamine levels.

- (b) The surface of the electrode can be anodically pretreated such that the oxidation peak of the ascorbic acid is shifted towards more positive potentials, and away from the peak potential of dopamine. On anodisation of the BDD electrode, oxygen functionalities (which replace the surface hydrogen atoms) are adsorbed onto the electrode surface. Fujishima et. al. [9] believe that these oxygen functionalities are highly oriented. They proposed that this highly oriented oxygen layer exerts a dipolar field. This field might be effective in repelling a molecule such as AA, which has four OH groups and one $>C=O$ group on its periphery. Thus, the AA anodic peak position is expected to shift from its original position. The net effect is a dopamine and AA peak separation.

7.3.3 Dopamine in the presence of ascorbic acid

Figure 7.14 and **Figure 7.15** represents the cyclic voltammograms for a mixture of 0.2 mmol/L dopamine and 2 mmol/L ascorbic acid in 0.1 mol/L $HClO_4$ at a glassy carbon electrode and at a BDD electrode respectively. The concentration of the ascorbic acid was 10 times higher than that of dopamine.

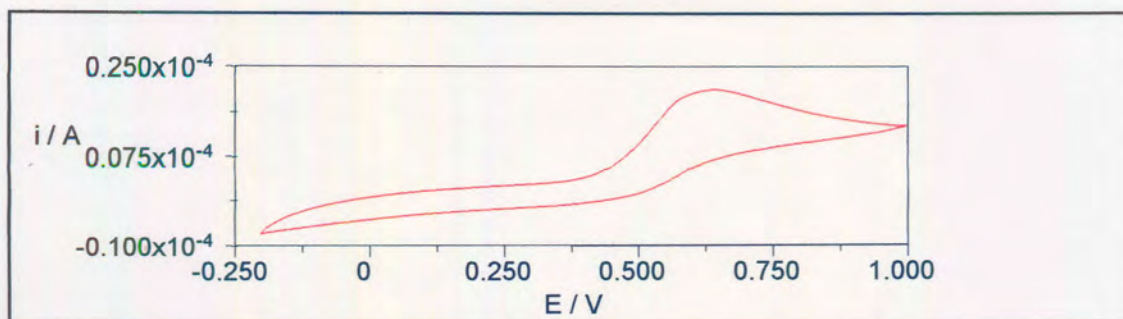


Figure 7.14 : Cyclic voltammogram for 0.2 mmol/L dopamine and 2 mmol/L ascorbic acid in 0.1 mol/L HClO₄ at a glassy carbon electrode - scan rate is 50 mV/s.

$E_i = -0.2$ V, scan direction = positive.

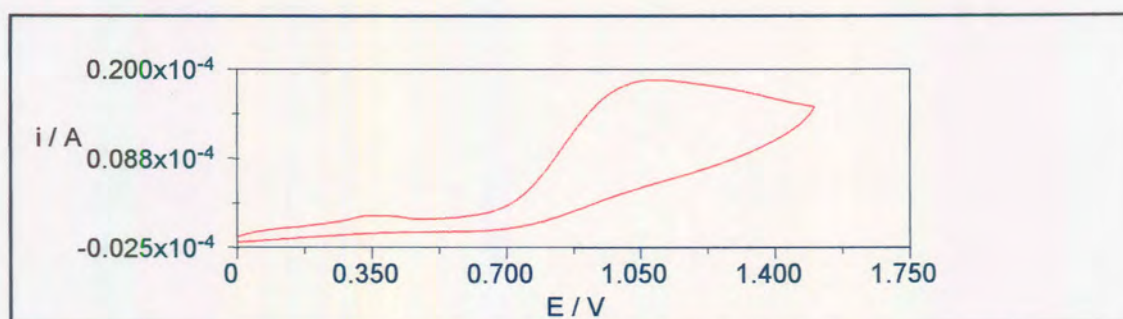


Figure 7.15 : Cyclic voltammogram for 0.2 mmol/L dopamine and 2 mmol/L ascorbic acid in 0.1 mol/L HClO₄ at a BDD electrode - scan rate is 50 mV/s. $E_i = 0$ V,

scan direction = positive.

It is evident from the voltammograms shown in **Figures 7.14** and **7.15** that no separation of the AA and dopamine peaks was achieved. Voltammograms with a single broad peak were obtained due to the overlap of the individual peaks. The accumulative AA and dopamine oxidation peak is seen at approximately 0.62 V for the glassy carbon electrode and at approximately 1 V for the BDD electrode.

To detect dopamine in the presence of ascorbic acid, the method of anodic pretreatment was adopted. The BDD electrode was anodically treated at 2.7 V for 90 minutes in a strong base (KOH). The method for anodisation is described in **Section 5.3.2**. This method attaches oxygen functionalities to the diamond surface, thereby converting the surface from hydrophobic or partially hydrophobic to hydrophilic. By employing this method, the AA peak potential is shifted more positive, whilst the shift in the dopamine peak potential is negligible.

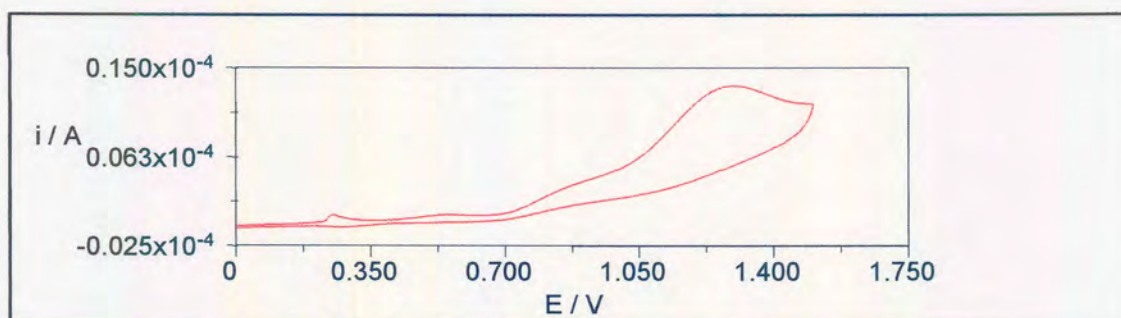


Figure 7.16 : Cyclic voltammogram for 0.2 mmol/L dopamine and 2 mmol/L AA in 0.1 mol/L HClO₄ at an anodised BDD electrode - scan rate is 50 mV/s. E_i = 0 V, scan direction = positive.

Figure 7.16 illustrates the cyclic voltammogram for 0.2 mmol/L dopamine and 2 mmol/L AA in 0.1 mol/L HClO₄ at an anodised BDD electrode. A distinct shoulder is observed at approximately 0.87 V. This shoulder can be attributed to the oxidation of dopamine. A well-defined oxidation peak occurs at approximately 1.3 V. This peak is most likely due to the oxidation of AA, and may be explained by the presence of highly oriented oxygen functionalities as proposed by Fujishima. et. al. [9] (see **Section 7.3.2**).

The boron concentration of the BDD electrode used in this investigation was 543 mg/L. According to the work undertaken by Fujishima et. al. [9], a BDD diamond electrode having a film resistivity in the order of $10^{-3} \Omega \cdot \text{cm}$, was used to obtain an effective dopamine and AA peak separation. This led to the belief that in order to convert the observed dopamine shoulder (**Figure 7.16**) to a more prominent peak, a BDD electrode having a higher boron concentration (probably in the order of 10^{21} boron atoms. cm^{-3}) is necessary. BDD electrodes containing this amount of boron are referred to as being “heavily doped”. A number of attempts were made to source “heavily doped” BDD electrodes from De Beers, but these are currently unavailable.

Unlike the glassy carbon electrode, the anodically pretreated BDD electrode is able to electrochemically distinguish between the dopamine and AA oxidation peaks without the use of a membrane, which is known to decrease the response time of an electrode. There is thus potential for a BDD electrode to be transformed into a microelectrode having a diameter similar to that of the common carbon fibre electrode (approximately $10 \mu\text{m}$) developed by Gonon and his colleagues [7] in 1978. This microelectrode, after the anodisation pretreatment, can be inserted into the ECF in the brain for online *in vivo* monitoring of dopamine.

A proposed method [28] for a microelectrode fabrication to be used for the *in vivo* monitoring of dopamine could be the chemical vapour deposition of boron-doped diamond onto the tip of a microscopic substrate that resembles a needle-like structure. This needle could contain a tiny reference electrode as well as a counter electrode.

Electrical contact can be made using silver wire and/or silver paste. Silver is chosen since it is less harmful to the body than most other common electrical contacts such as copper.

7.4 Conclusion

The BDD electrode was found to be superior to the glassy carbon electrode in the detection of dopamine and AA. This is evidenced by the excellent R-square and percentage recovery values obtained for the BDD electrode as compared to the values obtained for the glassy carbon electrode. However, in comparison to glassy carbon, the rate of electron transfer at the BDD electrode surface was found to be slow.

The BDD electrode also appeared to be reproducible in both the dopamine and the AA systems. On comparison of the electrochemical behaviour of the De Beers BDD electrode to that of literature in the dopamine as well as the AA system, no significant difference was observed. The BDD electrode was found to be stable over a three month period in both the dopamine and AA systems. The detection limits for dopamine and AA at a BDD electrode surface were found to be 0.56 $\mu\text{mol/L}$ and 20 nmol/L respectively. The expected concentrations of dopamine in tissue are 0.1 – 0.01 $\mu\text{mol/L}$ (10 – 100 nmol/L) [29]. In order to achieve these detection limits, it is suggested that “heavily boron-doped” (10^{21} boron atoms. cm^{-3}) diamond electrodes be used.

In order to detect dopamine in the presence of AA, the BDD electrode may be anodically pretreated in 0.1 mol/L KOH solution. This pretreatment shifts the AA peak position

more positive and in so doing, the dopamine and AA peaks may be resolved. It appears therefore that an anodically pretreated BDD electrode would make an ideal biosensor for the detection of dopamine in the presence of ascorbic acid, since it is accurate, reproducible, resistant to fouling, reliable and most importantly biocompatible. However, further work needs to be done to improve the sensitivity of the BDD electrode to be used for dopamine detection. One possible method is to use a “heavily boron-doped” diamond sample as the electrode material. This is expected to increase the size of the dopamine peak. A prolonged electrode lifetime and stability study on the anodically pretreated BDD electrode is also essential in order for the eventual design and fabrication of an implantable microelectrode.

7.5 References

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CHAPTER 8

Electroanalysis of organic systems via a bio-recognition element – thyroid hormones (L-T₃ and L-T₄)

8.1 Introduction

Thyroid hormones (iodinated amino acids), which are produced by the thyroid gland, are essential for adequate growth, development and energy metabolism [1]. In the foetus, they affect growth and differentiation, and in the mature human, they regulate metabolism. These hormones are secreted directly into the blood stream where they are transported to various cells and tissues in the body.

The two principal thyroid hormones are L-thyroxine (L-T₄) and L-triiodothyronine (L-T₃). The structures of these hormones are found in **Figures 8.1 and 8.2** respectively. L-T₄ has four iodine atoms attached to its aromatic rings, whereas L-T₃ contains three iodine atoms. These hormones are transported to various cells in the body via two plasma proteins, namely the thyroid hormone-binding globulin (TBG) and the thyroid hormone-binding prealbumin (TBPA). Only a small fraction (<0.3 %) of the total hormones in circulation is free. L-T₄, a prohormone, is converted into the more active L-T₃ by tissue deiodinases in the body. Approximately 90% of the L-T₃ in the body is formed by the

deiodination of L-T₄ [3]. The secretion of L-T₃ and L-T₄ by the thyroid gland is regulated by a complex system that originates in the central nervous system (CNS). Normal thyroid function yields about 28 – 50 µg of L-T₃ daily [3].

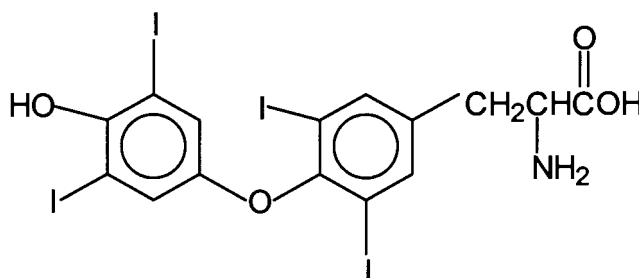


Figure 8.1 : Chemical structure of L-thyroxine (L-T₄) [2]

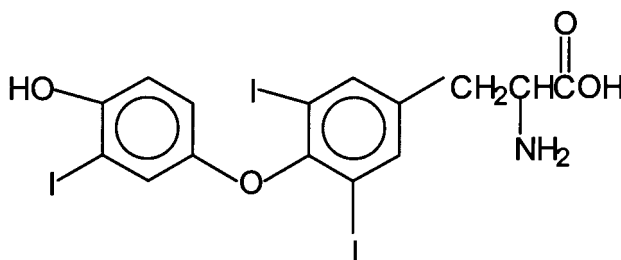


Figure 8.2 : Chemical structure of L-triiodothyronine (L-T₃) [2]

Since the L-T₃ and L-T₄ thyroid hormones affect growth in the foetus and regulate the metabolism of a mature human, the presence and equilibrium balance of these hormones are very important. It is common for humans in later life to develop thyroid hormone imbalances, and it is therefore necessary to be able to monitor these hormone levels in order to correct these imbalances by means of medication.

Classical methods of L-T₃ and L-T₄ analysis are very time consuming and more importantly, very expensive. An alternative technique that may be employed for the detection of L-T₃ and L-T₄, is voltammetry. In order to use this technique, the working electrode must be an immunosensor, which is very similar to a biosensor, except that it involves an antibody-antigen reaction [4].

Antibodies are proteins produced by B cells, which are designed to control the immune response in extracellular fluids. They are diverse, with more than 10¹⁰ possible variations, yet each antibody is designed to recognise only a specific antigen [5]. Antigens are foreign substances that enter the body, and are often proteins on the surface of bacteria and viruses [6]. Because of the ability of the antibody to recognise a specific antigen, an immunosensor is capable of selectively analysing for a biological compound in a complex matrix such as blood or ECF. An immunosensor is therefore said to function via a so-called bio-recognition system.

8.1.1 Bio-recognition system

A bio-recognition system (**Figure 8.3**) consists of bio-recognition elements (enzymes or antibodies) that are attached to an electrode surface [4,7-9]. Examples of potential electrode materials are diamond, glassy carbon, carbon paste, gold and platinum.

Once the bio-recognition element is immobilised on the electrode surface, the electrode is inserted into the sample matrix. The specific analyte of interest then combines with the

bio-recognition element in a “lock and key” [10] type mechanism. During this interaction, electrons are either released or consumed, depending on whether the reaction is an oxidation or reduction. The flow of electrons occurs via the electrode, and is recorded as a voltammogram.

For the thyroid hormone system, the mouse monoclonal anti-thyroxine (anti-T₄) combines with the L-T₄, thereby releasing electrons and hence producing an electrochemical signal. Likewise, the mouse monoclonal anti-triiodothyronine (anti-T₃) combines with the L-T₃ hormone to produce an electrochemical signal.

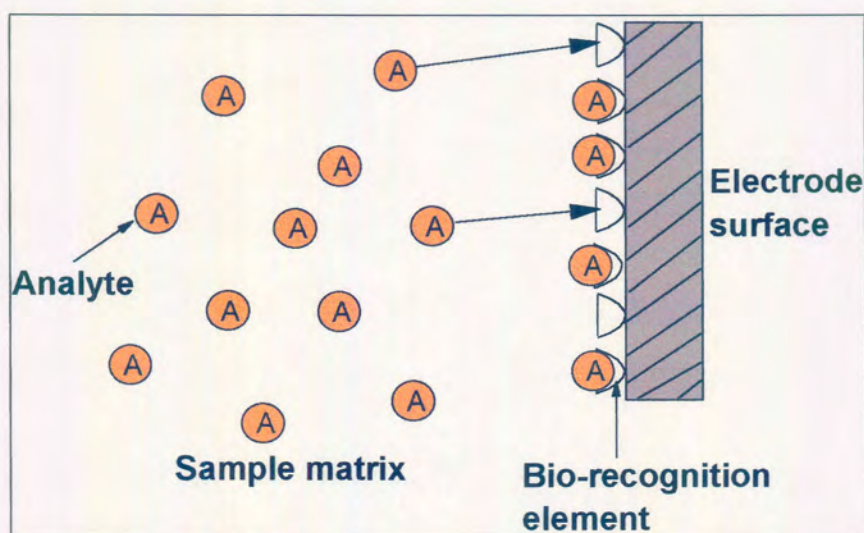


Figure 8.3 : Diagram illustrating a bio-recognition system

A bio-recognition element can be immobilised on an electrode surface using various techniques :

- (a) The element can be chemically or physically adsorbed onto the electrode surface

- (b) The electrode surface can be anodically treated such that the surface is terminated by oxygen functionalities. The bio-recognition element can then be chemically bound to these functionalities.
- (c) Chemical attachment and entrapment of bio-recognition elements in conductive polymer matrixes, which are in contact with the electrode surface, can also be used.
- (d) The bio-recognition element can be immobilised in a membrane, for example cellulose acetate. This membrane can then be attached to the electrode surface.

Bourdillon et. al. [11] attached a glucose oxidase conjugated antibody (antimouse IgG) to the surface of a glassy carbon electrode by soaking the electrode in the glucose oxidase solution overnight. Stefan et. al. [3] successfully impregnated graphite paste with mouse monoclonal anti-T₃. By using a chronoamperometric technique, the immunosensor was used for the L-T₃ assay at $\mu\text{g/L}$ - ng/L concentration levels. There are as yet no reports in the literature describing the detection of thyroid hormones at diamond electrodes.

A number of methods may be used to attach anti-T₃ and anti-T₄ to a BDD electrode surface. If the anti-T₃ and anti-T₄ are to be entrapped in a membrane or a polymer, the construction of the immunosensor has to be reproducible, and it is known that there are inherent problems with reproducibility in this approach [12]. For this reason, an alternative novel method for the attachment of the anti-T₃ and anti-T₄ to the electrode surface was developed. This technique involves the absorption of the anti-thyroid hormones into the pores of a boron-doped porous diamond electrode.

8.1.2 Porous diamond electrode

The manufacture of boron-doped porous diamond may be accomplished by the high temperature, high pressure sintering of crushed boron-doped CVD diamond. According to the shape and size of the crushed diamond particles, as well as the sintering conditions, various pore shapes are possible : cylindrical pores (circular in cross-section), ink-bottle pores having a narrow neck and wide body, and slit-shaped pores with parallel plates [13]. As this is a new electrode material, no data regarding the pore structure of porous diamond electrodes exists yet. The pore volume and pore size distribution may be measured using mercury porosimetry or low temperature gas adsorption–desorption (BET method) [14]. However, the research work detailed below was an initial feasibility study of the use of porous diamond electrodes as thyroid immunosensors, and detailed measurements of the pore structure lay outside the scope of this work.

For the electrochemical detection of the thyroid hormones (L-T₃ and L-T₄), the anti-thyroid hormones (anti-T₃ and anti-T₄) must be incorporated in the pores of the BDD electrode. One method of accomplishing this is to allow the anti-thyroid hormones to diffuse into the pores of the electrode by soaking the electrode in the anti-thyroid solution, and to rely on capillary forces to retain the solution within the pores. It is vital that the anti-thyroid hormone remains in the pores when the electrode is subsequently immersed in the thyroid hormone solution: if the anti-thyroid hormone diffuses out of the pores, no antibody-antigen reaction will be recorded, and the electrochemical signal will be lost.

Figure 8.4 illustrates a schematic diagram of a porous diamond substrate containing the anti-thyroid hormones and the formation of an antibody-antigen complex. When an antibody encounters an antigen, the two establish a bond which stays intact indefinitely or for a time of the order of hours [15]. The mean time until the molecules dissociate again depends on the antibody. This implies that the immunosensor (porous diamond electrode/antibody system) will become inactive after a measurement, and will have to be regenerated in some way, or be discarded, i.e. a disposable immunosensor.

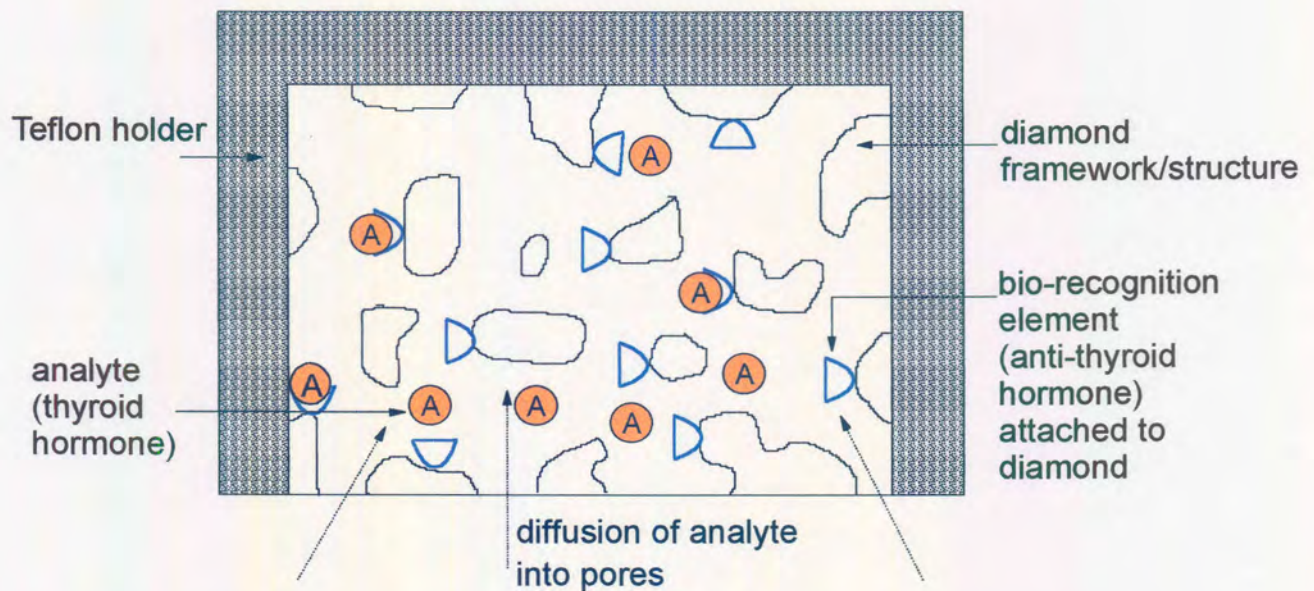


Figure 8.4 : Schematic diagram of a porous diamond substrate containing the anti-thyroid hormones and the formation of the antibody-antigen complex

8.2 Experimental

8.2.1 Crushing and sintering

Boron-doped CVD diamond webbing was crushed (to smaller size fractions) using iron rings in a ring crusher. Two size fractions were investigated: 49/57 US mesh (ca. 310 – 330 μm diameter) and 75/90 US mesh (ca. 180 – 200 μm diameter). During ring crushing of the CVD diamond, the diamond was inevitably contaminated with trace amounts of iron. Since iron is soluble in hydrochloric acid (HCl), the crushed CVD diamond was boiled in HCl to remove the iron impurity. The diamond particles were then compacted and sintered under high pressure and high temperature conditions. Afterwards, the sintered porous compact was lapped in order to produce discs of approximately 1 mm thickness and 20 mm diameter.

At this stage, the porosity of the two size fractions (49/57 US mesh and 75/90 US mesh) was determined by calculating the volume ($\pi \times \text{radius}^2 \times \text{height}$) of the circular lapped substrate. The theoretical mass of the substrate was then calculated (volume \times density of diamond (3.51 g/cm^3)). Thereafter, the substrate was weighed and the percentage ratio of the actual mass over the theoretical mass was taken as a measurement of the porosity of the porous diamond. The porosity of the two size fractions was calculated to be 14% in each case. The porous diamond compact was then laser cut to size and inserted into a Teflon electrode holder as described previously in **Section 5.3.2**, to give a projected surface area of 0.15 cm^2 .

8.2.2 Characterisation

Characterisation of the boron-doped porous diamond electrode was done using SEM, XPS and contact angle measurements.

One of the important milestones for the detection of the thyroid hormones using an immunosensor is the attachment of the antiserum in the pores of the BDD electrode. As mentioned previously, the antiserum may be absorbed into the pores by allowing the electrode to soak in the antiserum solution for a certain period of time. To determine the absorption kinetics, linear sweep voltammograms for the 75/90 US mesh porous BDD electrode were generated as a function of time in an ascorbic acid (AA) solution. Ascorbic acid was chosen since the solution is very stable, and the cyclic voltammograms that were generated thus far using the CVDBD electrodes were found to be very reproducible.

The surface chemistry of the porous BDD electrode was also manipulated in order to assess its effect on the electrochemical behaviour of the electrode.

8.2.3 Detection of thyroid hormones (L-T₃ and L-T₄)

The L-T₄ stock solution (12.51 mmol/L) was prepared by dissolving 0.064 g of solid L-T₄ in a mixture of 0.5886 g analytical grade sodium hydroxide (NaOH) and 4.738 g analytical grade ethanol. Similarly, L-T₃ stock solution (14.27 mmol/L) was prepared by

dissolving 0.0641 g of solid L-T₃ in a mixture of 0.6 g analytical grade NaOH and 4.8 g analytical grade ethanol. Both the thyroid hormones (L-T₃ and L-T₄) were obtained from Sigma Aldrich. The NaOH and ethanol were obtained from Saarchem.

The anti-T₃ was diluted to a working dilution of 1:70 in 0.01 mol/L phosphate buffered saline, pH = 7.4, containing 0.1% sodium azide, whilst the anti-T₄ was diluted to a working dilution of 1:40 in 0.01 mol/L phosphate buffered saline, pH = 7.4, containing 0.1% sodium azide. The 0.01 mol/L phosphate buffered saline solution was prepared using ultrapure water from a Millipore purification system (18 μΩ.cm). After preparation, both the antiserum solutions were stored at 0°C. The antisera (anti-T₃ and anti-T₄) were obtained from Sigma Adrich.

To speed up the process of impregnating the porous electrode with antiserum, a vacuum dessicator was used. Basically, the porous BDD was inserted into the antiserum solution and the solution together with the porous BDD was placed in a dessicator. A vacuum pump was connected to the dessicator and switched on. The vacuum thus created in the pores caused the liquid antiserum to be drawn quickly and efficiently into the pores of the electrode. The duration of the vacuum treatment was 10 minutes. After this period, the porous BDD electrode was removed from the antiserum solution and inserted into the Teflon electrode holder using carbon paste.

Electrochemical experiments were performed using an AutoLab PGStat 100 with the AutoLab GPES software. A porous BDD electrode, Ag/AgCl electrode and a platinum

electrode served as the working, reference and counter electrodes in the cell respectively. Cyclic voltammetry scans were initially performed in a phosphate buffered saline solution, pH = 7.4, containing 0.1% sodium azide, before and after the addition of L-T₃ and L-T₄, in order to optimise the working conditions for the chronoamperometry technique. Chronoamperometry techniques were used to obtain the detection limits of both L-T₃ and L-T₄.

After each voltammetric measurement, the surface of the porous BDD electrode was cleaned by boiling it in HCl. All measurements were carried out at room temperature (25°C) and at a pH = 7.4 in a deaerated, stationary analyte solution. The projected surface area of the 75/90 US mesh porous BDD electrode used in the experiments was 0.15 cm².

8.3 Results and discussion

8.3.1 Characterisation of the porous BDD electrode

8.3.1.1 Surface morphology

The morphology of the two porous BDD electrodes (made from 49/57 US mesh and 75/90 US mesh size fractions) was viewed under the SEM. **Figure 8.5** and **Figure 8.6** represents the SEM image of the 49/57 US mesh and 75/90 US mesh electrodes respectively. Randomly oriented facets can be seen in both the SEM images. The surface of the diamond is very rough and the presence of pores can be clearly seen. The pores appear to be randomly distributed, with a variation in the pore size.

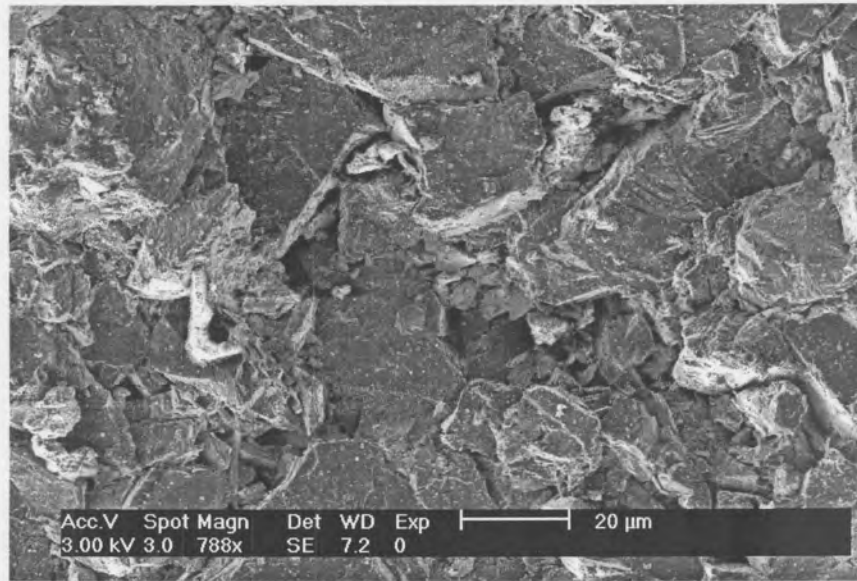


Figure 8.5 : SEM image depicting the surface of the 49/57 US mesh porous BDD size fraction

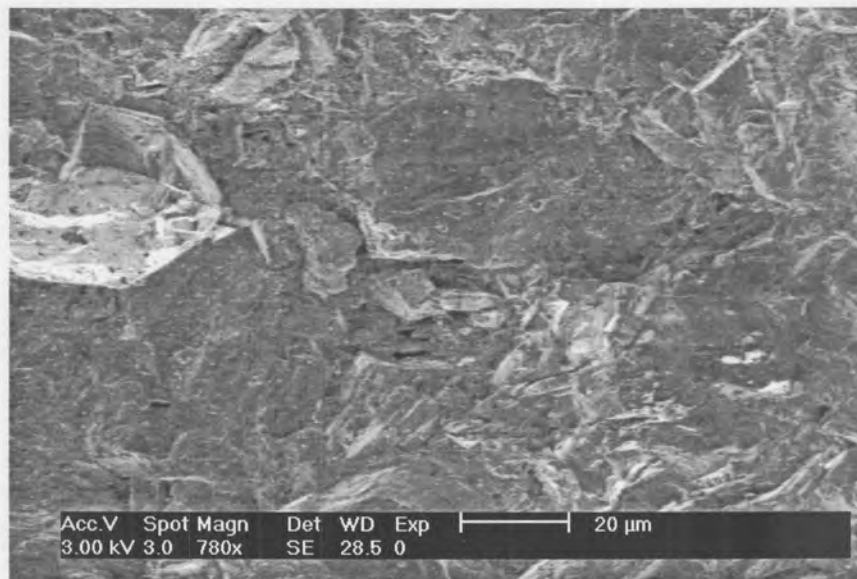


Figure 8.6 : SEM image depicting the surface of the 75/90 US mesh porous BDD size fraction

The surface of the 75/90 US mesh electrode was analysed using energy dispersive spectroscopy (EDS). The presence of trace amounts of iron was again detected in the EDS spectrum. It is assumed that this contaminant was introduced during ring crushing of the boron-doped CVD webbing using iron rings, and was not entirely removed by treating the crushed diamond powder with HCl, prior to sintering. Subsequent treatment of the sintered porous diamond with HCl effectively removed these traces of iron.

8.3.1.2 Surface chemistry

The contact angles (**Table 8.1**) of the two electrodes were measured to determine their surface state. The surfaces were found to be only partially hydrophobic, with the 49/57 US mesh electrode slightly more hydrophobic than the 75/90 US mesh electrode. This indicates that the surface of the diamond contains oxygen functionalities.

Table 8.1 : Contact angle measurements for the two porous BDD electrodes

Size fraction	Contact angle		
	Range / °	Average/ °	Std Deviation
49/57 US mesh	58 - 71	64.70	6.29
75/90 US mesh	43 - 57	52.55	5.02

The oxygen content of the 75/90 US mesh porous BDD electrode was determined using XPS (**Figure 8.7**). The oxygen content of the porous diamond was found to be 2.69×10^{15} atoms/cm² and the surface oxygen coverage was found to be 1.4 monolayers.

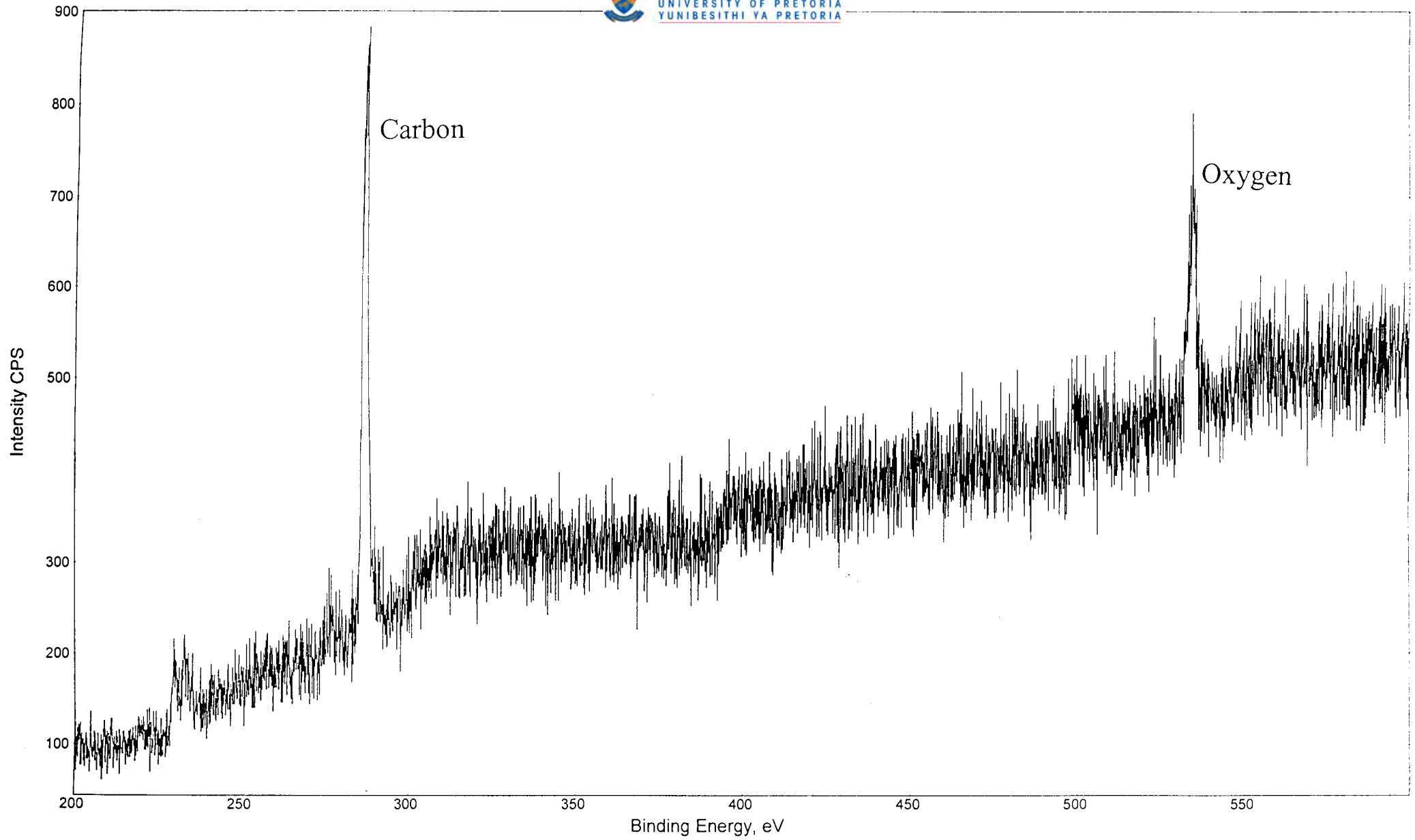


Figure 8.7 : XPS spectrum for the 75/90 US mesh porous BDD electrode

Contact angle measurements taken after treatment of the porous electrodes to remove trace iron impurities revealed that the hydrophilic/hydrophobic nature of the surface was not significantly changed.

8.3.1.3 Electrochemical behaviour of porous BDD electrodes

Figure 8.8 illustrates the cyclic voltammograms for 0.1 mol/L HClO₄ before and after HCl treatment at a 75/90 US mesh porous BDD electrode. A large baseline current is observed in the porous BDD electrode before treatment in HCl. This may be due to the presence of oxygen functionalities on the electrode surface and/or trace amounts of iron. A redox couple, probably the Fe²⁺/Fe³⁺ couple, is also evident. Subsequent treatment in HCl may have caused some of the surface functionalities to become hydrogen terminated, thus decreasing the observed baseline current. The redox couple was also absent.

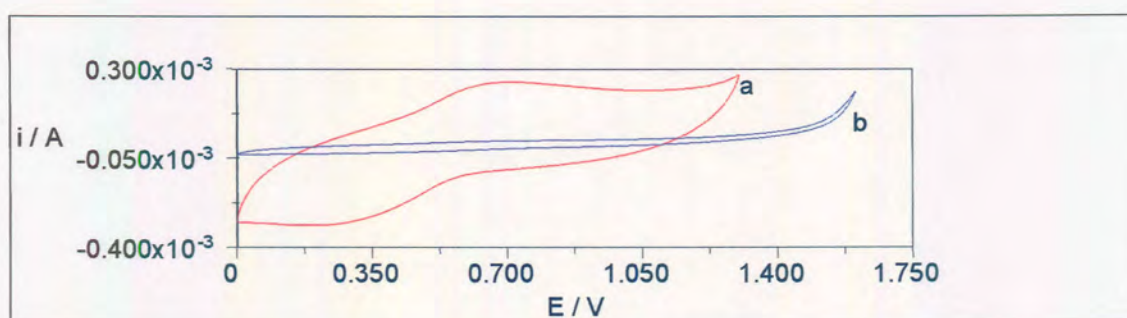


Figure 8.8 : Cyclic voltammograms for 0.1 mol/L HClO₄ at a 75/90 US mesh porous BDD electrode, (a) before HCl treatment, (b) after HCl treatment - scan rate is 50 mV/s. $E_i = 0$ V, scan direction = positive

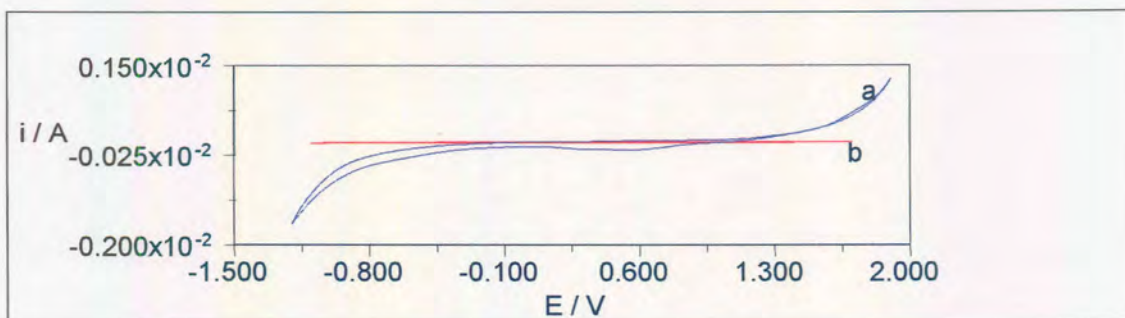


Figure 8.9 : Cyclic voltammograms for 0.5 mol/L HClO₄ at a 75/90 US mesh porous BDD electrode as well as at a CVDBD4 electrode : (a) 75/90 US mesh porous BDD electrode, (b) CVDBD4 electrode – scan rate is 50 mV/s.

Figure 8.9 represents the cyclic voltammograms for 0.5 mol/L HClO₄ at a 75/90 US mesh porous BDD electrode as well as at a CVDBD4 electrode. The baseline current for the 75/90 US mesh porous BDD electrode is much larger than that of the CVDBD4 electrode. This could be attributed to the increased surface area of the 75/90 US mesh diamond caused by an increase in the porosity, or the hydrophilic nature of the 75/90 US mesh diamond caused by oxygen functionalities on the electrode surface : the presence of these functionalities are known to make the diamond electrode more electroactive.

A further cause of the increased baseline current for the 75/90 US mesh porous BDD electrode could be the presence of non-diamond carbon on the electrode surface. This non-diamond carbon could have formed during the high pressure, high temperature sintering of the crushed CVD diamond. In order for diamond to convert to graphite, the presence of solvent metal catalysts (for example iron) is necessary. The EDS spectrum (discussed earlier) shows that trace amounts of iron were present on the electrode surface

after the sintering process. Iron could have therefore catalysed the conversion of diamond to graphite.

It is also evident from the cyclic voltammogram shown in **Figure 8.9**, that the potential window for water electrolysis for the 75/90 US mesh porous BDD electrode has decreased when compared with the CVDBD4 electrode. Oxygen evolution begins at approximately 1.4 V, whilst hydrogen evolution starts at about -0.8 V.

Figure 8.10 shows the cyclic voltammogram for 0.1 mol/L HClO_4 at a 75/90 US mesh and at a 49/57 US mesh porous BDD electrode respectively. It is apparent that the baseline current for the 49/57 US mesh electrode is greater than the baseline current for the 75/90 US mesh electrode. This could be due to surface area effects or the presence of non-diamond carbon on the surface of the 49/57 US mesh electrode.

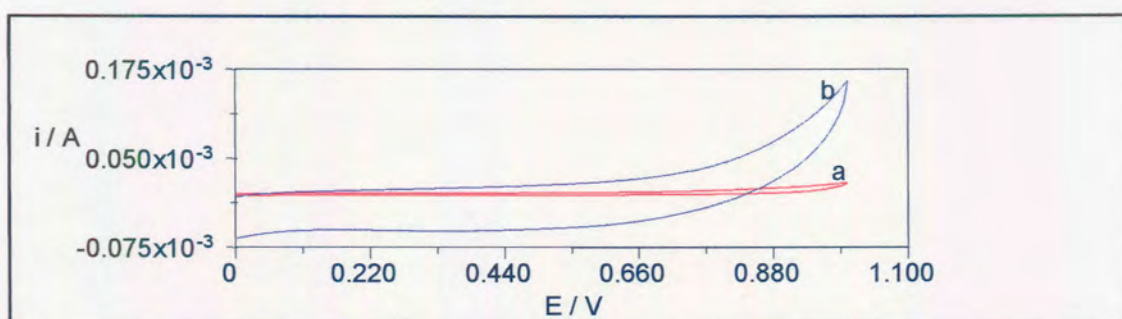
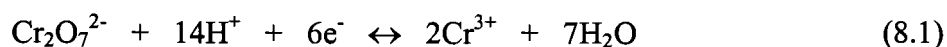


Figure 8.10 : Cyclic voltammogram for 0.1 mol/L HClO_4 at a 75/90 US mesh (a) and at a 49/57 US mesh (b) porous BDD electrode – scan rate is 50 mV/s. $E_i = 0$ V, scan direction = positive.

The surface chemistry of the 49/57 US mesh porous BDD electrode was modified by boiling the electrode in chromic acid (0.5 g potassium dichromate in 100 ml sulphuric acid) in order to oxidise the surface of the electrode. This surface manipulation was performed in order to observe the effect of the surface state of the electrode on its electrochemical behaviour.

Figure 8.11 illustrates the cyclic voltammogram for 0.1 mol/L HClO₄ at a 49/57 US mesh porous BDD electrode before and after treatment. After the chromic acid treatment, the baseline current had increased tremendously with the additional occurrence of a redox couple which could be due to the presence of chromium. The oxidation potential of the redox couple occurs approximately 0.5 V and the reduction potential occurs at about 0.3 V. **Equations 8.1** and **8.2** depict the reduction of two chromium species [16]. The relevant reduction potentials of the chromium species versus a hydrogen reference electrode as well as a Ag/AgCl reference electrode are shown in **Table 8.3** [16].



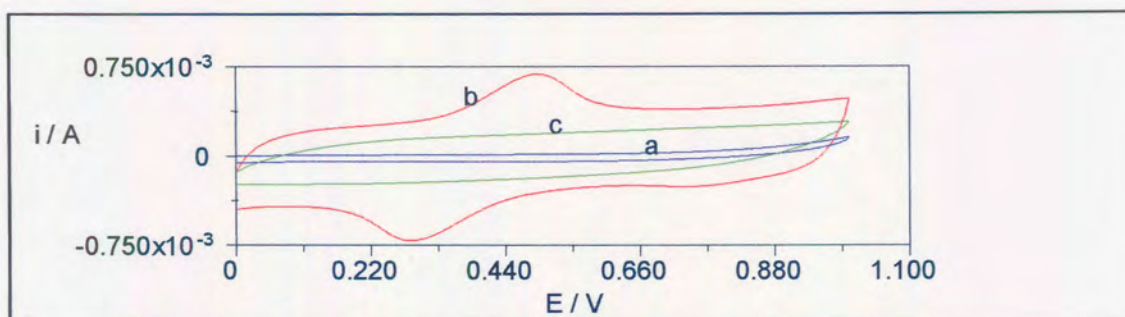


Figure 8.11 : Cyclic voltammogram for 0.1 mol/L HClO₄ at a 49/57 US mesh porous BDD electrode before and after treatment : (a) HCl treated 49/57 US mesh electrode, (b) 49/57 US mesh electrode after treatment in chromic acid, (c) 49/57 US mesh electrode after treatment in chromic acid, then boiled in HCl – scan rate is 50 mV/s. E_i = 0 V, scan direction = positive

It is apparent that the oxidation and reduction potentials of the redox couple shown in **Figure 8.11** are higher than the standard reduction potentials of the chromium species indicated in **Table 8.3**. However, this shift in peak potentials could be ascribed to the electrode material being used.

Table 8.3 : Reduction potentials of chromium species

Reduction reaction	Potential / V	
	Hydrogen reference electrode	Ag/AgCl reference electrode
$\text{Cr}_2\text{O}_7^{2-} + 14\text{H}^+ + 6\text{e}^- \leftrightarrow 2\text{Cr}^{3+} + 7\text{H}_2\text{O}$	1.33	1.11
$\text{Cr}^{6+} + 3\text{e}^- \leftrightarrow \text{Cr}^{3+}$	1.10	0.88

The contact angle of the 49/57 US mesh porous BDD electrode was measured after the chromic acid treatment and was found to be 0° . This implies the surface of the electrode to be extremely hydrophilic with numerous oxygen functionalities attached to the electrode surface. The hydrophilic nature of the electrode surface could explain the observed increase in the baseline current.

Interestingly, when the 49/57 US mesh porous BDD electrode was subsequently boiled in HCl, the assumed chromium redox couple was absent (**Figure 8.11**). In addition, the baseline current was also reduced in magnitude. It appears that the HCl removed the chromium from the surface of the electrode and partially terminated the electrode surface with hydrogen atoms, possibly making the electrode surface more hydrophobic. This would explain the observed decrease in the baseline current.

8.3.2 Absorption kinetics

Figure 8.12 illustrates the absorption curve for 2 mmol/L AA in 0.1 mol/L HClO₄ at a 75/90 US mesh porous BDD electrode. Linear sweep voltammograms (background corrected) were generated every hour for the period that the electrode was in contact with the stationary AA solution.

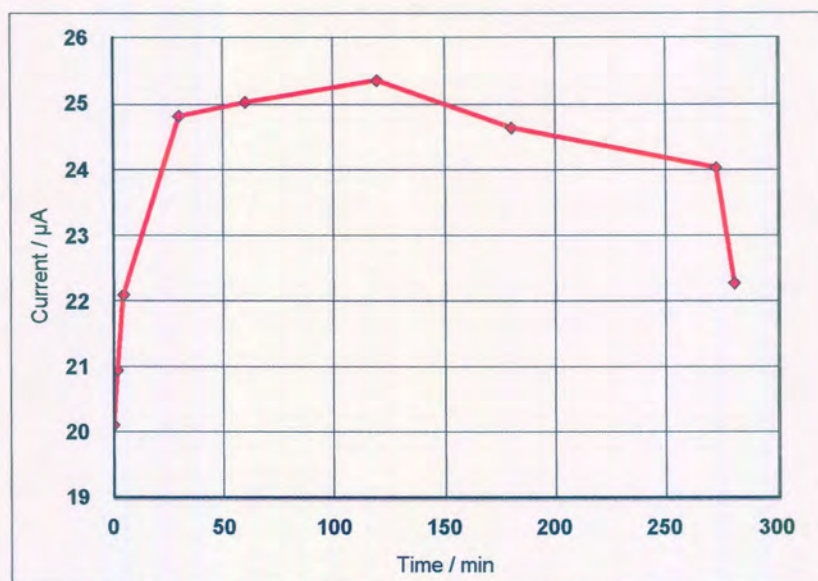


Figure 8.12 : Absorption curve for 0.6 mmol/L AA in 0.1 mol/L HClO₄ at a 75/90 US mesh porous BDD electrode

It can be seen from **Figure 8.12** that the current increased steeply for the first 30 minutes, then gradually increased for the next 100 minutes. Afterwards, the current began to slowly decrease. The increase in current is thought to be due to the diffusion of the AA into the pores of the BDD electrode, which effectively increased the electrode surface area in contact with the solution. The decrease in current after 120 minutes is ascribed to the depletion of AA in the pores during the periods when the potential was applied. Replenishment of AA in these pores, especially those deepest inside the electrode, would be retarded by diffusion, and by the fact that the capillary interaction which initially would have helped the pores to fill relatively quickly with AA, would no longer play a role once the pores were filled with liquid.

In all, it can be inferred from **Figure 8.12** that in order for the antiserum to absorb into the pores of the BDD electrode, the electrode will have to be soaked in the antiserum solution for a period of no less than two hours.

8.3.3 Detection of thyroid hormones (L-T₃ and L-T₄)

8.3.3.1 Detection of L-T₄

The antiserum (anti-T₃ or anti-T₄) was initially absorbed into the pores of the 75/90 US mesh BDD electrode by allowing the electrode to soak in the antiserum solution for a period of two hours. A linear sweep voltammogram (background current for 0.01 mol/L phosphate buffered saline, containing 0.1% sodium azide subtracted) for 0.44 mmol/L L-T₄ in 0.01 mol/L phosphate buffered saline was generated (**Figure 8.13**).

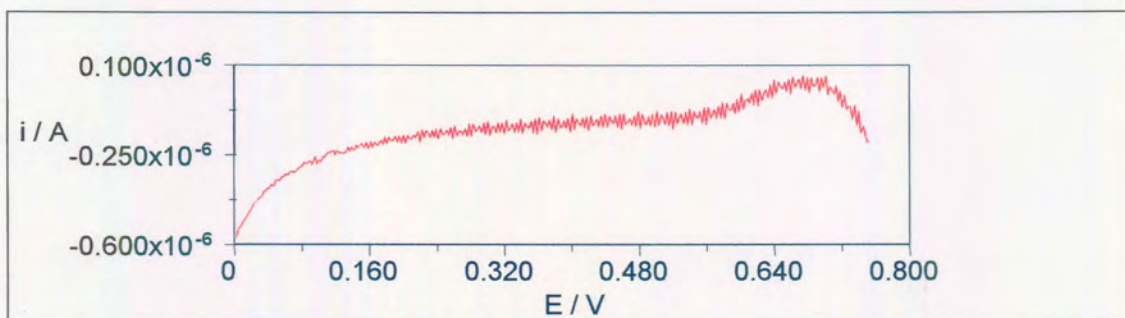


Figure 8.13 : Linear sweep voltammogram for 0.44 mmol/L L-T₄ in 0.01 mol/L phosphate buffered saline containing 0.1% sodium azide, pH = 7.4 – scan rate is 50 mV/s. E_i = 0 V, scan direction = positive.

An oxidation peak, which probably occurs as a result of the formation of the antigen-antibody complex, is seen at approximately 680 mV. Considering the potential at which this peak occurred, the chronoamperometric technique was performed at an optimum potential of 780 mV vs the Ag/AgCl electrode. The potential, however, was applied in two stages. The initial applied potential was 100 mV with a duration of 5 seconds. The potential was then stepped up to 780 mV.

During the course of the chronoamperometry measurements, it was found that the immunosensor was only able to detect the presence of L-T₄ on the first voltammetric scan. The subsequent scans did not yield an increase in the observed current. It was thought that the antiserum may be diffusing out of the diamond electrode pores, thereby resulting in a decrease in the observed signal.

The antiserum was thus drawn into the electrode pores using a vacuum dessicator technique. This technique proved to be quite effective in that the subsequent scans also showed an increase in the observed current. **Figure 8.14** shows a chronoamperometric scan for 0.01 mol/L phosphate buffered saline before and after the addition of 17.51 μmol/L L-T₄.

A definite increase in the current is observed on addition of 17.51 μmol/L L-T₄. This increase in the current was initially thought to be due to the presence of NaOH and ethanol in the solution. However, subsequent scans carried out in 0.01 mol/L phosphate buffered saline containing small quantities of NaOH and ethanol, failed to yield an

increase in the current. Thus, the increase in the observed current (**Figure 8.14**) is definitely due to the presence of L-T₄. According to **Figure 8.14**, a plateau was reached after approximately 16 seconds. The lower detection limit for L-T₄ was found to be 12.51 $\mu\text{mol/L}$. At the other end of the scale, L-T₄ concentrations above 37.53 $\mu\text{mol/L}$ could not be detected, due to the precipitation of L-T₄ in solution.

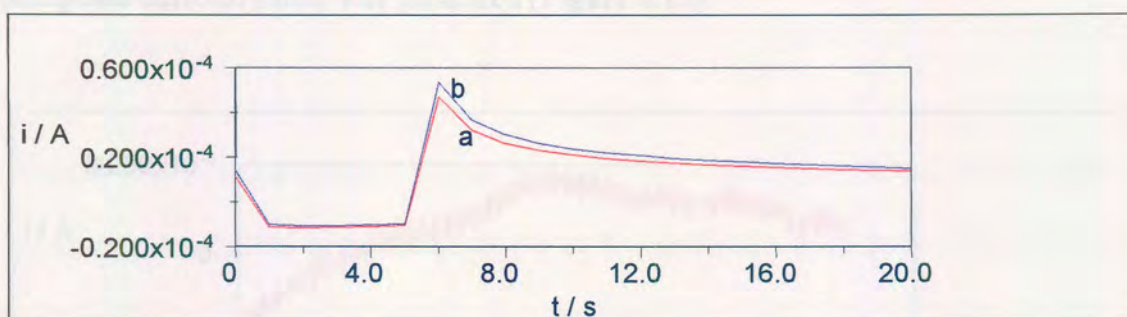


Figure 8.14 : Chronoamperometric scan for 0.01 mol/L phosphate buffered saline before (a) and after (b) the addition of 17.51 $\mu\text{mol/L}$ L-T₄

Although the porous BDD electrode was successful in the detection of L-T₄, the immunosensor response was found to be very irreproducible. Thus a calibration plot for the L-T₄ assay could not be generated. The irreproducibility of the porous BDD electrode could be due to the questionable reproducibility of the vacuum desiccator technique in impregnating the diamond pores with the antiserum or it may be due to differences in the diffusion rate of the phosphate buffer and L-T₄ solution into the electrode pores.

The immobilisation of the antiserum in the diamond electrode pores may be improved by blending the antiserum in a gel and then impregnating the pores with this gel containing antiserum. The gel should ideally not hinder the electrochemical detection of L-T₄.

Another method for the attachment of the antiserum onto the electrode surface is through the use of a membrane. However, both these methods are expected to further decrease the response time of the electrode.

The diffusion rate of the phosphate buffer and L-T₄ solution could be increased by oxygen terminating the surface of the porous BDD electrode, thereby making the surface hydrophilic. In this way, the electrolyte would wet the electrode surface more easily and diffuse into the electrode pores at a much faster rate. The response of the porous BDD electrode may be made more consistent by integrating the area under the chronoamperometric peak as a function of time, so that small differences in diffusion rates and response times would be eliminated. In this way, a calibration curve may be drawn.

A further improvement on the detection limit and sensitivity of the porous BDD electrode in the L-T₄ assay would be to decrease the baseline current obtained. This can be achieved by hydrogen terminating the electrode surface. However, a hydrogen terminated electrode surface may decrease the diffusion rate of the analyte into the electrode pores, due to poor wetting of the electrode surface by the solution. The baseline current can also be decreased by the removal of any non-diamond carbon present on the electrode surface. It is suspected that the surface of the 75/90 US mesh porous BDD electrode contained non-diamond carbon (as discussed previously under **Section 8.3.1.3**). The presence of this graphitic carbon may have resulted in the increased baseline current observed which would have negatively affected the detection limit of L-T₄.

8.3.3.2 Detection of L-T₃

For the detection of L-T₃, the corresponding antiserum was impregnated in the pores of the 75/90 US mesh BDD electrode using the vacuum dessicator technique. A background current corrected linear sweep voltammogram for 51 $\mu\text{mol/L}$ L-T₃ in 0.01 mol/L phosphate buffered saline was generated (**Figure 8.15**).

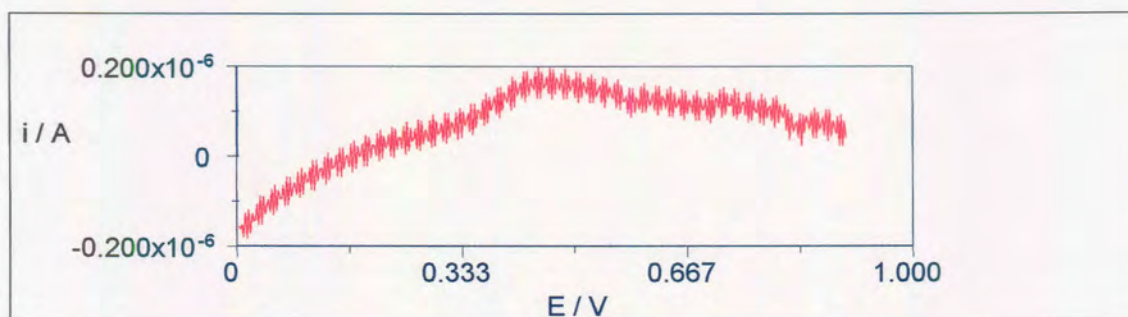


Figure 8.15 : A background current corrected linear sweep voltammogram for 51 $\mu\text{mol/L}$ L-T₃ in 0.01 mol/L phosphate buffered saline – scan rate is 50 mV/s. $E_i = 0$ V, scan direction = positive

An oxidation peak is evident at approximately 500 mV. This peak is most probably due to the formation of the antigen-antibody complex. In view of the oxidation peak potential, the chronoamperometric technique was performed at an optimum potential of 780 mV vs the Ag/AgCl electrode. The potential was again applied in two stages : the initial applied potential was 100 mV and the subsequent applied potential was increased to 780 mV.

Figure 8.16 illustrates a chronoamperometric scan for 0.01 mol/L phosphate buffered saline solution before and after the addition of 14.27 $\mu\text{mol/L}$ L-T₃ at a 75/90 US mesh

porous BDD electrode. A plateau was reached after 18 seconds. The lower limit of detection for L-T₃ was found to be 856 nmol/L.

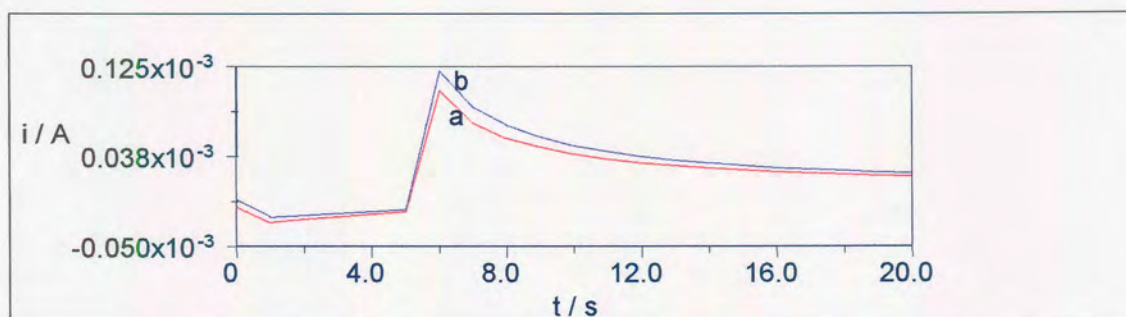


Figure 8.16 : Chronoamperometric scan for 0.01 mol/L phosphate buffered saline solution, pH = 7.4, (a) before addition of 14.27 $\mu\text{mol/L}$ T₃, (b) after addition of 14.27 $\mu\text{mol/L}$ L-T₃.

Due to the irreproducible nature of the 75/90 US mesh porous BDD electrode in the L-T₃ system (for the reasons mentioned previously), a calibration plot could not be generated.

8.4 Conclusion

Porous BDD electrodes were successfully used to analyse for L-T₃ and L-T₄ in synthetic samples, by impregnating the electrode with antiserum using a vacuum technique. The lower limit of detection for L-T₃ was found to be 856 nmol/L, while that of L-T₄ was 12.5 $\mu\text{mol/L}$. As the respective concentrations of free L-T₃ and L-T₄ in the blood are 0.89 pmol/L and 0.031 nmol/L (2.97 nmol/L and 0.103 $\mu\text{mol/L}$ respectively protein-bound)[1], further work is necessary to improve the sensitivity of this immunosensor.

The response of the porous BDD electrode was found to be highly dependent on its surface state, with a hydrophobic surface giving a lower baseline than that of a hydrophilic surface.

Problems were experienced with reproducibility in both the L-T₃ and L-T₄ cases, and were thought to be related to the electrode surface state, and/or the impregnation and retention of the antiserum in the pores.

In conclusion, the feasibility of using porous BDD electrodes for the detection of thyroid hormones has been demonstrated. In order to develop a commercial diamond biosensor for thyroid hormone detection based on this material, further work is required to improve the sensitivity and reproducibility of the electrode.

Generally, the porous BDD electrode offers a significant advantage over a CVD BDD electrode, as it is much cheaper to manufacture, and may be manufactured to have a vastly increased surface area, which opens up possibilities of future application in systems where the analyte is present in very low concentrations. These systems vary from biological or medical to wastewater applications.

8.5 References

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CHAPTER 9

Conclusions

The as-received samples were characterised using various techniques, namely SEM, LA-ICP-MS, Raman spectroscopy and XPS. Contact angle and resistance measurements were also performed. SEM analysis showed the surface of the boron-doped samples to have randomly oriented facets, whilst LA-ICP-MS measurements indicated that CVDBD2 contained the lowest boron concentration and CVDBD6 contained the highest boron concentration. It was inferred from the Raman spectroscopy analysis, that the presence of boron in the boron-doped CVD samples seems to suppress the formation of sp^2 carbon.

Using XPS, the oxygen content of the as-received CVDBD4 sample was found to be 2.37×10^{15} atoms/cm², with a surface oxygen coverage of 1.26 monolayers. The contact angle measurements revealed CVDBD1 and CVDBD2 to be hydrophobic, CVDBD3 and CVDBD4 to be partially hydrophobic and CVDBD5 and CVDBD6 to be more hydrophilic. The resistance measurements showed CVDBD2 and CVDBD3 to have a low conductivity, whilst CVDBD4 – CVDBD6 were found to have a high conductivity. Thus, the minimum level of boron required to make diamond a suitable electrode material was found to be 500 mg/L.

Various hydrogenation (tube furnace, TGA and hydrogen plasma) and oxygenation (tube furnace, TGA, chromic acid and anodisation) techniques were employed to modify the surface state of the boron-doped diamond electrodes. This investigation was deemed necessary since literature reports that the surface state of the electrode may significantly affect its electrochemical behaviour. The most effective hydrogenation technique was found to be the hydrogen plasma treatment (yielding a very hydrophobic surface), whilst all of the oxygenation techniques investigated were successful in producing a very hydrophilic electrode surface.

However, one of the drawbacks in producing an oxygen terminated electrode surface is the formation of boron oxide on the surface. Boron oxide could drastically affect the observed electrochemical signal. It is therefore proposed that in order to minimise the formation of boron oxide, mild oxygenation methods should be used, for example heat treating the boron-doped diamond sample in air at temperatures below 250°C for a short period of time or boiling the diamond sample in hydrogen peroxide for short intervals.

On investigating the electrochemical behaviour of various boron-doped CVD diamond electrodes, it was found that, as the boron concentration of the electrode increases, the potential window decreases. However, it was also found that the potential window for the highest boron-doped diamond electrode (CVDBD6) was still wider than that of the glassy carbon electrode, thereby confirming the advantage that the BDD electrode has over glassy carbon when investigating aqueous systems.

The electrochemical behaviour of two redox systems was investigated, namely potassium iron (III) cyanide and cerium (III) sulphate. It was found that the electrochemical behaviour of the potassium iron (III) cyanide redox couple was very complex when a boron-doped diamond electrode was used. The peak separation of the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple (690 mV) was found to be an order of magnitude larger than that of the glassy carbon electrode (61 mV). It is suggested that oxygen termination of the BDD electrode surface inhibits the electron transfer of the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple. It can thus be assumed that in order to reduce the peak separation, the surface of the electrode will have to be made hydrophobic.

The BDD electrode proved to be more effective (wider potential window, therefore better peak resolution) for the quantitative analysis of cerium (III) sulphate, than the glassy carbon electrode. This is evidenced by the BDD electrode producing a better quality calibration plot and percentage recovery values than the glassy carbon electrode.

The electrochemical behaviour of two biological systems, dopamine and ascorbic acid, were also investigated. Dopamine co-exists with ascorbic acid (a major electrochemical interferent) in the extracellular fluid in the brain. The boron-doped diamond electrode was found to be superior to the glassy carbon electrode in the detection of dopamine and ascorbic acid, as evidenced by the excellent R-square and percentage recovery values obtained for the boron-doped diamond electrode.

The detection limits for dopamine and ascorbic acid at a boron-doped diamond electrode surface were found to be 0.56 $\mu\text{mol/L}$ and 20 nmol/L respectively. The expected concentrations of dopamine in tissue are 0.1 – 0.01 $\mu\text{mol/L}$ (10 – 100 nmol/L). In order to achieve these detection limits, it is suggested that “heavily boron doped” (10^{21} boron atoms. cm^{-3}) diamond electrodes be used. Finally, the BDD electrode was found to be stable over a three month period in both the dopamine and ascorbic acid investigations.

In order to detect dopamine in the presence of ascorbic acid, the boron-doped diamond electrode may be anodically pretreated in potassium hydroxide. This pretreatment shifts the ascorbic acid peak position more positive and in so doing, the dopamine and ascorbic acid peaks may be resolved. It appears therefore that an anodically pretreated boron-doped diamond electrode would make an ideal biosensor for the detection of dopamine in the presence of ascorbic acid, since it is accurate, reproducible, resistant to fouling, reliable and most importantly biocompatible.

However, further work needs to be done to improve the sensitivity of the BDD electrode to be used for dopamine detection. One possible method is to use a “heavily” boron-doped diamond sample as the electrode material. This is expected to increase the size of the dopamine peak. A prolonged electrode lifetime and stability study on the anodically pretreated boron-doped diamond electrode is also essential in order for the eventual design and fabrication of an implantable microelectrode.

For the first time, the electrochemical detection of thyroid hormones was investigated at diamond electrodes, as detailed in this study. Also documented for the first time is the use of porous boron-doped diamond electrodes. This electrode material was successfully used to analyse for the thyroid hormones (L-T₃ and L-T₄) in synthetic samples, by impregnating the electrode with antiserum using a vacuum technique. Since the electrochemical determination of the thyroid hormones using porous boron-doped diamond electrodes is novel, the results obtained could not be compared to that of literature. The lower limit of detection for L-T₃ was found to be 856 nmol/L, while that of L-T₄ was 12.5 µmol/L. As the respective concentrations in blood are 0.89 pmol/L and 0.031 nmol/L (2.97 nmol/L and 0.103 µmol/L respectively protein-bound), further work is necessary to improve the sensitivity of this immunosensor.

The response of the porous boron-doped diamond electrode was found to be highly dependent on its surface state, with a hydrophobic surface giving a lower baseline than that of a hydrophilic surface. Problems were experienced with reproducibility in both the L-T₃ and L-T₄ cases, and were thought to be related to the electrode surface state, and/or the retention of the antiserum in the pores.

The feasibility of using porous boron-doped diamond electrodes for the detection of thyroid hormones has been demonstrated. In order to develop a commercial diamond biosensor for thyroid hormone detection based on this material, further work is required to improve the sensitivity and reproducibility of the electrode.

Generally, the porous boron-doped diamond electrode offers a significant advantage over a CVD boron-doped diamond electrode, as it is much cheaper to manufacture, and may be manufactured to have a vastly increased surface area, which opens up possibilities of future application in systems where the analyte is present in very low concentrations. These systems vary from biological or medical to wastewater applications.

In conclusion, the De Beers boron-doped diamond electrode has been shown to possess definite advantages over the conventional glassy carbon electrode, in that it has a wide potential window for water stability (as confirmed using the cerium (III) sulphate system), low baseline current, and is chemically inert and resistant to fouling (as evidenced by the results obtained for both the biological systems, dopamine and ascorbic acid). The electrochemical behaviour of the De Beers boron-doped diamond electrode was also found to be very similar to that reported in literature, for the various electrochemical systems investigated (except for the novel thyroid hormone system, where no literature is available).

In all, the boron-doped diamond electrode is an excellent candidate for the analysis of various electrochemical compounds or species and would therefore have wide commercial electrochemical applications, especially for the detection of low levels of dopamine in the presence of ascorbic acid, and the detection of the thyroid hormones. It is, however, recommended that further work be done in order to improve the electrochemical behaviour of the boron-doped diamond electrodes for use in these systems.