

Additional Methods

DNA extraction

Mycelia were scraped off the surface of the plate in a 1.5 mL Eppendorf tube. Three glass beads (0.5 mg) and 600 µl of Cetyl trimethylammonium bromide (CTAB) extraction buffer (2 % CTAB, 100 mM Tris-HCl, pH 8.0; 1.4 M NaCl; 20 mM EDTA, 0.2 % mercaptoethanol) were added and the tissue was disrupted by vortex. After incubation for 3 min at 100 °C, 500 µl chloroform: isoamylalcohol (24:1) was added and mixed by inversion. The supernatant was recovered after 30 minutes centrifugation at maximum speed and was mixed with ammonium acetate (2.5 M final concentration) and ice-cold isopropanol. The DNA was precipitated by incubation for 60 min at -20 °C and pelleted by centrifugation for 30 min at 12 000 rpm. DNA pellets were air-dried and suspended in 100 µl double distilled water mixed with 200 µl Tris-EDTA buffer (Thermo Fisher Scientific, Wellington, USA). DNA was quantified using a Nanodrop nd1000 instrument (Thermo Fisher Scientific, Wilmington, USA) and visualized by electrophoresis with ethidium bromide on a 1% agarose borax anhydrous gel (Sigma-Aldrich, USA).

PCR specifications

ITS region was targeted using the primer set ITS1F (5' CTTGGTCTTTAGAGGAAGTAA 3') (Gardes & Bruns, 1993) and ITS4 (5' TCCTCCGCTTAGATATGC 3') (White *et al.*, 1990). Additionally, *S. minor* and *S. sclerotiorum* specific primers were used that could distinguish between these two species. For *S. minor*, the primer set targeted the laccase 2 (*lac*) gene (SMLcc2F 5' CCCTCCTATCTCTCTTCCAAACA 3' and SMLcc2R 5' TGACCAATACCAATGAGGAGAG 3'; (Abd-Elmagid *et al.*, 2013), while for *S. sclerotiorum* the aspartyl protease (*aspr*) gene was targeted (SSasprF 5' CATTGGAAGTCTCGTCGTCA 3' and SSasprR 5' TCAAACGCCAAAGCTGTATG 3'; (Abd-Elmagid *et al.*, 2013)

All PCR reactions were carried out in 25 µl reactions that included 1U KapaTaq DNA polymerase (Kapa Biosystems, USA), 1× KapaTaq Buffer A, 0.25mM of each dNTP and 0.4mM of each primer. PCR reactions for all three regions were carried out on a GeneAmp 9700 thermocycler (Applied Biosystems,

Foster City, California, USA) following the protocol developed by Abd-Elmagid *et al.*, (2013), but with the annealing temperature adjusted to 57°C. All PCR products were visualized by electrophoresis on a 2% (w/v) borax anhydrous agarose gel stained with GelRed Nucleic Acid Gel stain (Biotium, Hayward, USA). All PCR reactions included a no template negative control, as well as two positive controls consisting of DNA from a known *S. sclerotiorum* isolate 1980 (Denton-Giles *et al.*, 2018) and a *S. minor* isolate CBS 339.39 (Wingfield *et al.*, 2022).