

Supplementary figures for: Buttstedt A. 2022: The role of 10-hydroxy- Δ^2 -decenoic acid in the formation of fibrils of the major royal jelly protein 1/apisimin/24-methylenecholesterol complex isolated from honey bee (*Apis mellifera*) royal jelly. — *Eur. J. Entomol.* **119**: 448–453.

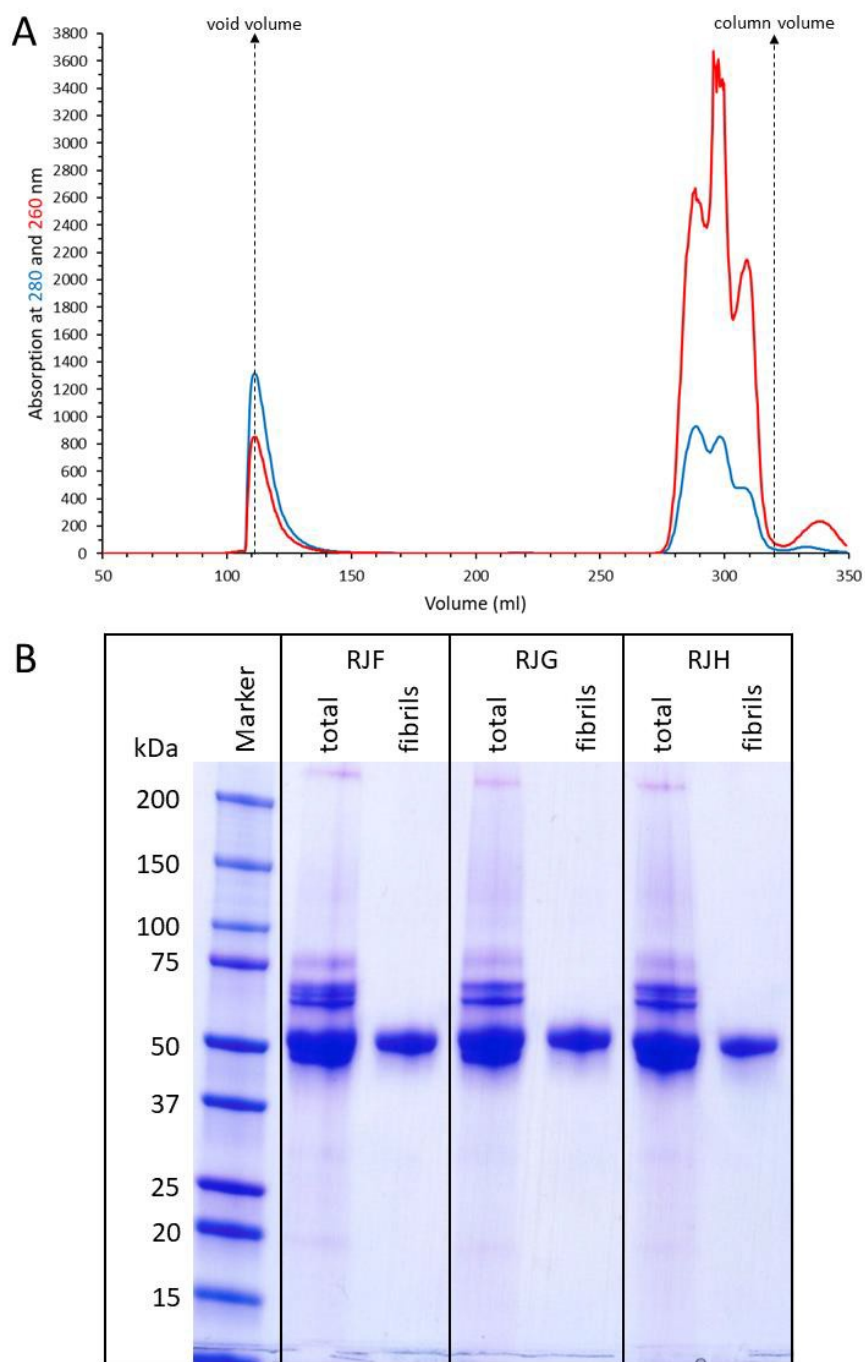


Fig. S1. Purification of fibrillar MRJP1/apisimin/24MC. **A)** Gel filtration exemplarily shown for the protein extract from RJF (column: HiLoad 26/600 Superdex 200 pg; buffer: 50 mM sodium citrate/citric acid, 150 mM NaCl, pH 4.0; flow rate: 2.0 ml/min; injection volume: 10 ml). Fibrillar MRJP1₄/apisimin₄/24MC₈ elutes within the void volume ($V_0 = 112$ ml) of the column and has, typical for a protein, a higher absorption at 280 (blue line) than at 260 nm (red line). The gel filtration step was necessary to separate the fibrillar protein from non-proteinaceous small molecular weight substances eluting shortly before the column volume ($V_t = 320$ ml) which had a higher absorption at 260 nm than at 280 nm and could not be removed using dialysis. **B)** SDS polyacrylamide gel (4-20% MiniProtein TGX stain-free (BioRad), separated at 175 V for 1 h, stained with Coomassie R-250) showing for all three RJ samples the protein components before (total) and after purification of the fibrils. The fibrillar fraction shows at around 50 kDa only MRJP1. Apisimin with 5 kDa is too small to be resolved within this gel.

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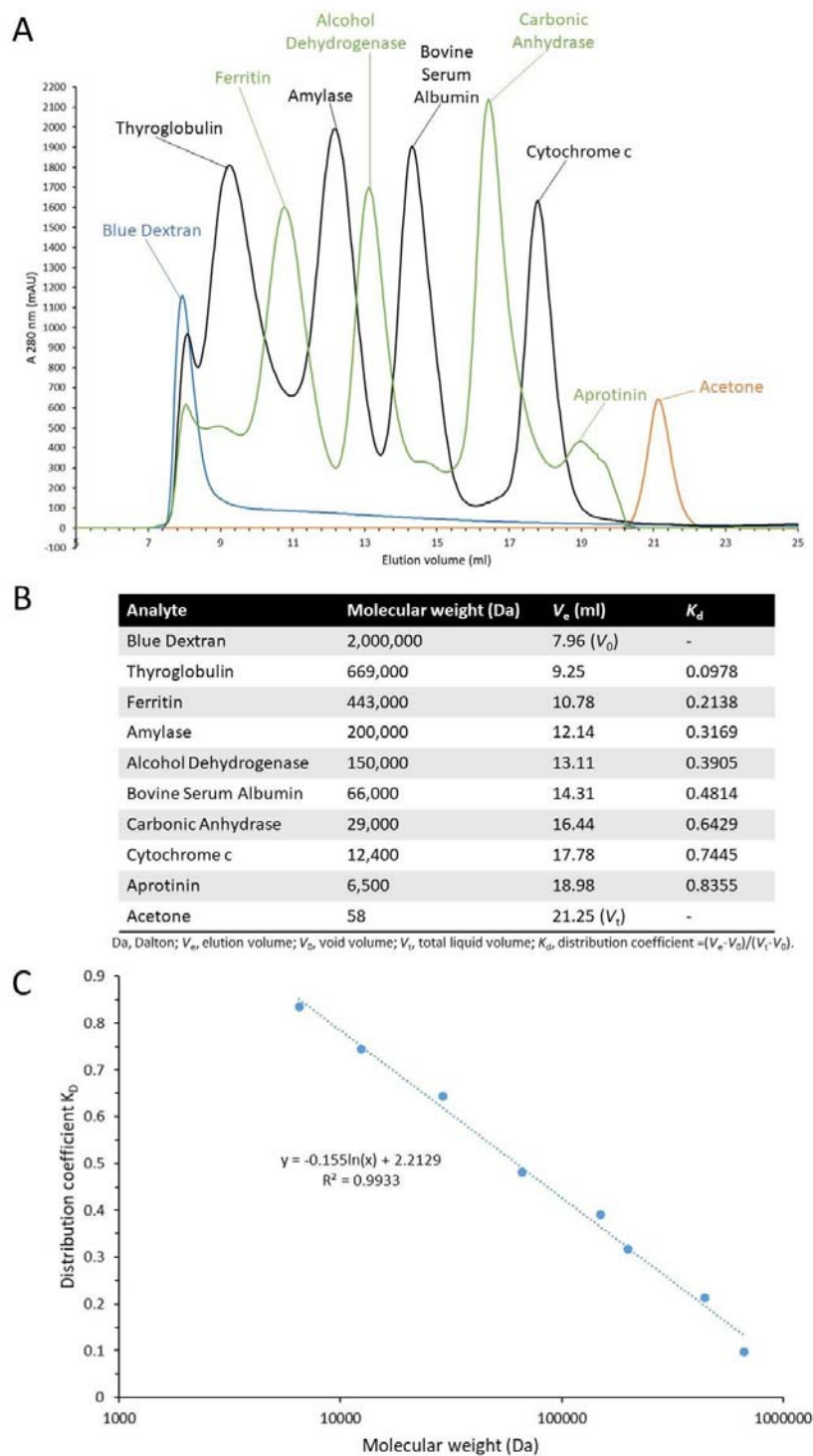


Fig. S2. Calibration of the Superdex 200 Increase 10/300 GL gel filtration column. **A**) Four separate gel filtration runs (buffer: 50 mM Tris/HCl, 150 mM NaCl, pH 7.5; flow rate: 0.7 ml/min; injection volume: 500 μ l) using Blue Dextran to determine the void volume (V_0) (blue line), acetone to determine the total liquid volume (V_t) (orange line) of the column and two runs using different marker proteins (green and black lines). **B**) Elution volumes (V_e) of Blue Dextran, acetone and the different marker proteins as well as calculated distribution coefficients (K_d) for the marker proteins. **C**) Plot showing the calculated distribution coefficients depending on molecular weight. The fitted logarithmic curve was used to calculate distribution coefficients and molecular weights during the subsequent analytical runs.

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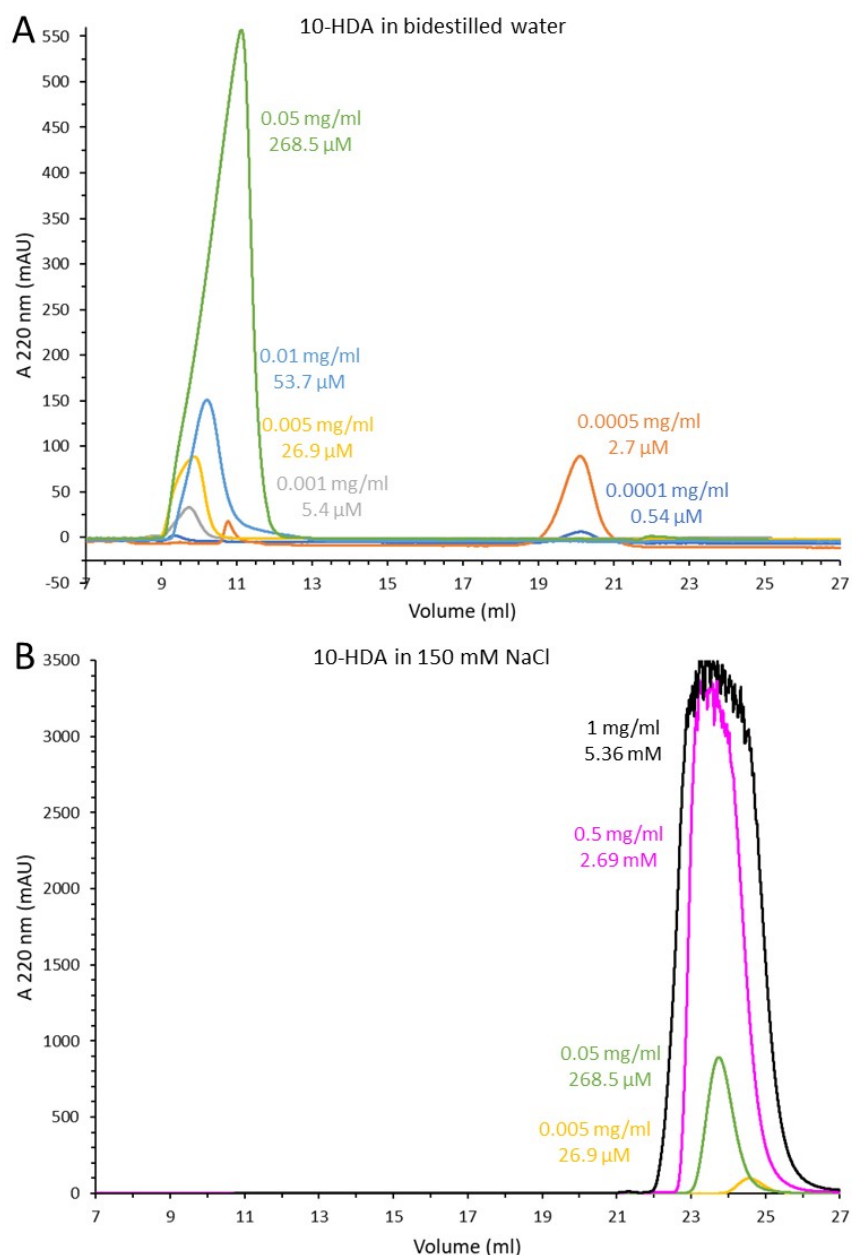


Fig. S3. Prevention of 10-HDA micelle formation by NaCl (column: Superdex 200 Increase 10/300 GL; flow rate: 0.7 ml/min; injection volume: 500 μ l). 10-HDA concentrations: Dark blue, 0.54 μ M; orange, 2.7 μ M; grey, 5.4 μ M; yellow, 26.9 μ M; light blue, 53.7 μ M; green, 268.5 μ M; magenta, 2.69 mM; black, 5.36 mM. Without addition of NaCl, micelles were formed from a concentration of 5.4 μ M 10-HDA. **A**) 10-HDA in double distilled water. **B**) 10-HDA in 150 mM NaCl.

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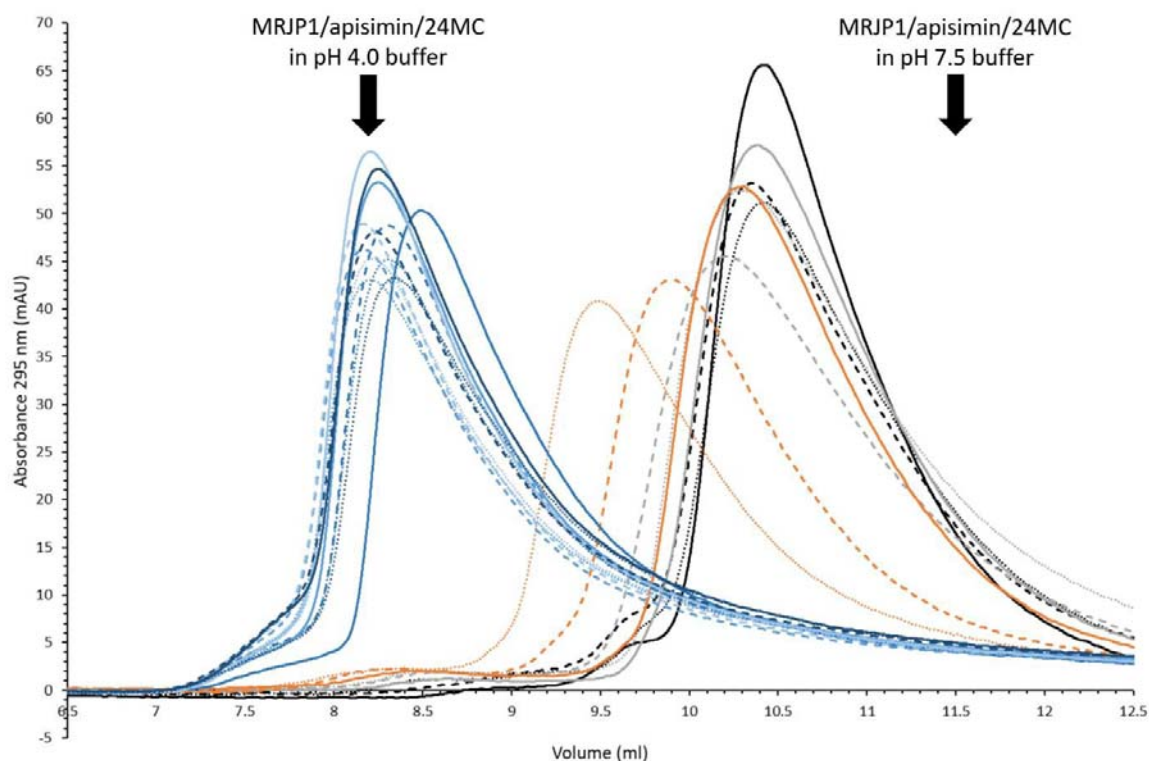


Fig. S4. Gel filtration analysis of 500 $\mu\text{g/ml}$ MRJP1/apisimin/24MC in 150 mM NaCl supplemented with different concentrations of 10-HDA (0 – 805.2 μM) (column: Superdex 200 Increase 10/300 GL; flow rate: 0.7 ml/min; injection volume: 500 μl). 10-HDA concentrations: Black, 0 μM ; grey, 26.8 μM ; orange 134.2 μM ; dark blue to light blue, 201.3 – 805.2 μM . RJP, solid line; RJG, dashed line; RJH dotted line.