

# Using species distribution models to gauge the completeness of the bat checklist of Eswatini

Ara Monadjem<sup>1,2,\*</sup>, Felicity Simelane<sup>1</sup>, Julie Teresa Shapiro<sup>3,4,5</sup>, Bonginkosi C. Gumbi<sup>1</sup>, Mngqobi L. Mamba<sup>1</sup>, Muzi D. Sibiyi<sup>1</sup>, Sifiso M. Lukhele<sup>1</sup>, Themb'alihlwa A.M. Mahlaba<sup>1</sup>

<sup>1</sup> Department of Biological Sciences, University of Eswatini, Private Bag 4, Kwaluseni, Eswatini

<sup>2</sup> Mammal Research Institute, Department of Zoology & Entomology, University of Pretoria, Private Bag 20, Hatfield 0028, Pretoria, Republic of South Africa

<sup>3</sup> School of Natural Resources and Environment, University of Florida, Gainesville FL USA

<sup>4</sup> Department of Wildlife Ecology and Conservation, University of Florida, Gainesville FL USA

<sup>5</sup> University of Lyon, CIRI INSERM U1111 - CNRS UMR5308 - ENS Lyon, Lyon, France, 46 Allée d'Italie, 69364 Lyon, France

\*Corresponding author:

Email address: ara@uniswa.sz

## Abstract

National species checklists are important for a variety of reasons, including biodiversity conservation. However, these national checklists are rarely complete, and it is not easy to gauge how many species have been overlooked or what the taxonomic identities of overlooked species would be. This is particularly the case for small, elusive, or nocturnal species such as bats. Despite their diversity and importance as ecosystem service providers, bat distributions are poorly known throughout much of Africa. We present a national checklist of bats for a small African country, Eswatini, by compiling species from museum specimens and literature records. A total of 32 species of bats have been recorded from the country. Since 1995, new species have continued to be

recorded in the country, with five additional species added since the last published checklist in 2016, suggesting that some species may have still been overlooked. In order to determine what species these may be, we used species distribution models based on the occurrence records of bats from southern Africa to predict what species would occur in Eswatini, which was then compared with what has been collected and deposited in museums. Our models predicted that a total of 47 species are likely to occur in Eswatini compared with 32 species collected to date. Our data suggest that the national checklist of bats of Eswatini is not yet complete and that further species are expected to be recorded for the country. We suggest that species distribution models can be useful in gauging the completeness of national checklists and in predicting which species may have been overlooked.

#### Keywords

Chiroptera; Maxent; species area curve; species richness

#### Introduction

Country checklists of species may serve several important functions, one of which is to inform conservation decisions. However, checklists are rarely complete as new species are discovered or described within the boundaries of even the most well surveyed countries. While African countries support a high known diversity of mammal species (Kingdon et al. 2013), this is the continent predicted to have the greatest number of undescribed mammal taxa in the world (Fisher et al. 2018), and vast regions of the continent have not been surveyed at all for small mammals (Monadjem et al. 2010a, 2015). Even in well surveyed parts of the continent, such as South Africa, making sense of species distributions is often difficult because of sampling bias; with accessibility being a critical factor in where past surveys have been conducted (Reddy and Davalos 2003). In other words, remote or inaccessible areas are typically under-represented in surveys.

Bats (order Chiroptera) are the second most diverse order of mammals after rodents (Simmons, 2005), with over 1400 species currently reported (Burgin et al. 2018; Simmons and Cirranello 2018) of which around 314 species (22% of global total) occur in Africa (ACR 2019). Bats are frequently used in conservation planning exercises and are specifically targeted in many biodiversity surveys (Decher et al. 2001, Fahr and Ebigbo 2003, Monadjem and Fahr 2007, Monadjem et al. 2016). Their importance for providing ecosystem services are also now well documented (Boyles et al. 2011; Kunz et al. 2011; Taylor et al. 2018a). Yet knowledge of bat distributions remains relatively poor compared with many other mammalian groups (Herkt et al. 2016), even in well-surveyed regions. For example, *Myotis alcaethoe* was added to the United Kingdom's national checklist only in 2010 (Jan et al. 2010) increasing the total number of resident species to 17 (Dietz and von Helversen 2004) and *Pipistrellus pygmaeus* was added in the decade before that (Barratt et al. 1997, Mayer and von Helversen 2001). This illustrates that even in a country like the United Kingdom with dozens of bat biologists and thousands of dedicated volunteers undertaking annual bat surveys (Mitchell-Jones et al. 1993) (also see <https://www.bats.org.uk/our-work/national-bat-monitoring-programme/reports/nbmp-annual-report>), new country records can still be made. In contrast, many tropical countries have a severe shortage of bat biologists or bat volunteers (Taylor 1999), with many African countries having no more than one or two dedicated professional bat biologists (A. Monadjem, personal observation).

In Africa, only a few countries have recent national checklists (Monadjem and Fahr 2007, Monadjem et al. 2010b, Bates et al. 2013, Amori et al. 2016, Child et al. 2016, Musila et al. 2019). For many countries, national checklists are not available, fragmentary in nature (having been published in numerous unrelated papers) or decades old (Kock 1969, Ansell 1978, Schlitter et al. 1982, Happold et al. 1987, Crawford-Cabral 1989, Yalden et al. 1996). While tools like distribution maps from the IUCN Red List (<https://www.iucnredlist.org/>) are available for Africa and used for ecological studies, they tend to underestimate species' ranges and the biodiversity of any given geographical area (Herkt et al. 2017).

In Eswatini (formerly Swaziland), bat surveys can be categorized as “historical” (pre-1995) or recent (since 1995) (see Methods). Prior to 1995, 12 species had been collected from the country and deposited in museums in South Africa and the United Kingdom (Monadjem 1998). However, the first checklist of bats in Eswatini was only published in 1997, and mostly based on surveys conducted from 1995 onward; it listed 16 species (Monadjem 1997) (Table 1). A year later, the total number of species recorded in Eswatini increased to 19 species (Monadjem 1998). By 2005, one additional species had been added to the national list (Monadjem 2005) and by this date, all the species collected prior to 1995 had been captured in recent surveys (Monadjem et al. 2005). The next published update affecting the bat checklist of the country was in 2008, when five new species were added (Monadjem and Reside 2008), raising the national total to 25 species. The last published checklist of Eswatini was in 2016 and listed 26 species (Shapiro and Monadjem 2016), but this paper erroneously omitted *Tadarida aegyptiaca*, which had been recorded previously (Monadjem 1998), and hence should have listed 27 species.

Table 1 – Bioclimatic and other environmental variables with Variance Inflation Factor (VIF) < 10, which were used in the Maxent models for bat distributions in this study

Variable	Description	VIF
Alt	Altitude	5.10
Alt_rough	Altitudinal roughness	1.40
Ecoregions	Ecoregions	1.53
Bio_2	Mean diurnal range	2.39
Bio_3	Isothermality	2.95
Bio_8	Mean temperature of wettest quarter	2.26
Bio_9	Mean temperature of driest quarter	7.72
Bio_13	Precipitation of wettest month	5.09
Bio_14	Precipitation of driest month	3.32
Bio_15	Precipitation seasonality	2.99
Bio_18	Precipitation of warmest quarter	2.89
Bio_19	Precipitation of coldest quarter	1.90

Clearly, the number of bat species recorded in Eswatini has risen significantly through time, raising doubts as to the completeness of this national checklist. This paper aims to assess how complete the current checklist is and to predict which species may have been overlooked. This is achieved by

comparing the actual number of species recorded in the country with species predicted based on species distribution models.

## Materials and Methods

### Study area

Eswatini is a small country situated in southern Africa covering an area of 17,360 km<sup>2</sup>. The country is topographically varied, with the Drakensberg mountain range in the west and the Lubombo mountain range on the eastern border with Mozambique. In between these two mountain ranges is a lowland region (Fig. 1). The western highlands comprise montane grassland with patches of forest, whereas the rest of the country is mostly covered in savanna (Monadjem et al. 2003).

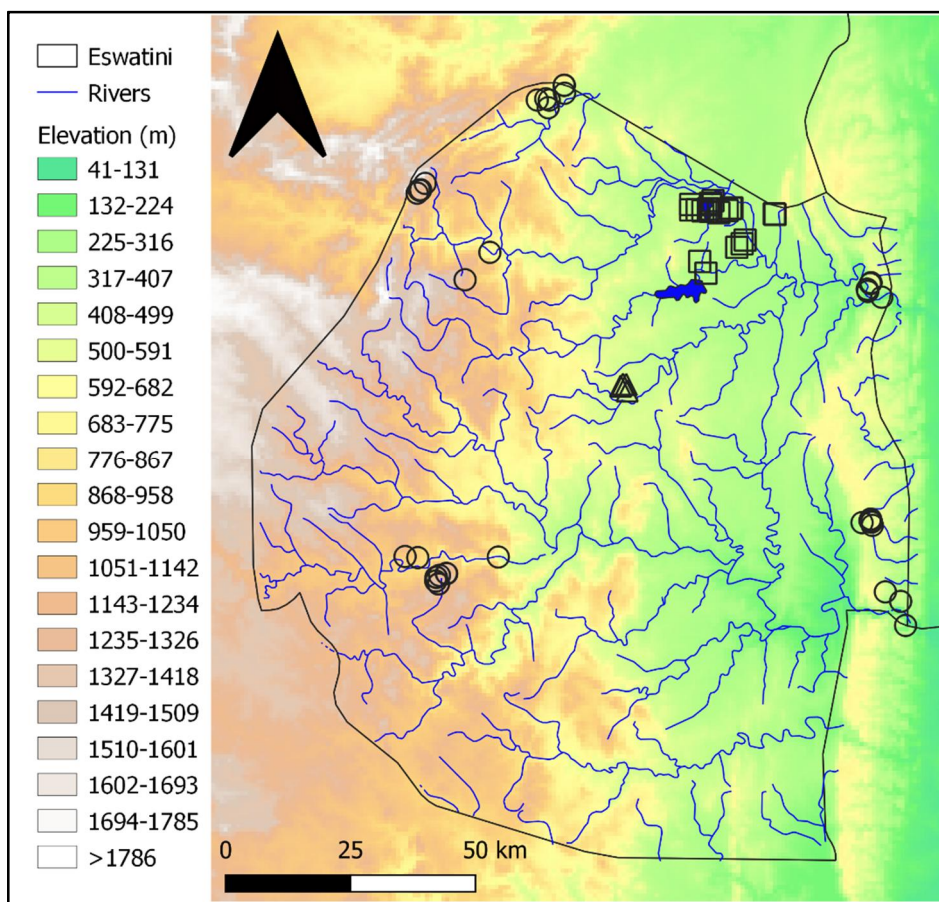


Fig. 1 – A digital elevation map of Eswatini showing the relief of the country including the major rivers flowing through. Also shown are the study sites in Eswatini at which new bat species not mentioned in Shapiro and Monadjem (2016) have been recorded. Squares = Inyoni Yami Swaziland Irrigation Scheme (YSIS) survey;

circles = Strengthening the National Protected Areas System of Swaziland (SNAP) survey; triangles = Dombeya Game Reserve survey (see Methods for further details)

## Species data

We compiled a dataset of all the bats collected in Eswatini based on historical pre-1995 collections and recent post-1995 records. All records were collected by A. Monadjem and various colleagues and students (for references to the various publications see the Introduction). For the post-1995 dataset, we compiled the year in which each species was first recorded in the country, and a cumulative total number of species for the period 1995-2018.

New bats recorded in Eswatini since 2015 have not yet been published and are presented here based on extensive trapping surveys conducted at: 1) Inyoni Yami Swaziland Irrigation Scheme cattle ranch (IYSIS, September 2015 to April 2016) near Tshaneni in northern Eswatini; 2) nine sites across the central and northern parts of the country under the Strengthening National Protected Areas project (SNPAS, December 2016 to February 2017); and 3) Dombeya Game Reserve (January 2018) (Fig. 1). Bats were captured using standard methods including setting up of mist nets and harp traps as described in Monadjem and Reside (2008). Voucher specimens of each species were collected and deposited in the Eswatini National Museum of Natural History, which were subsequently identified based on Monadjem et al. (2020a). Taxonomy follows Monadjem et al. (2020a) except for recent changes to the pipistrelle-like bat species (Monadjem et al. 2020b).

Beyond Eswatini, bat specimen records were obtained from Monadjem et al. (2020a) which included 125 species and 6,344 unique locality records from southern Africa (Fig. 2). We reduced this database in size by removing all species with less than six unique locality records in the region ( $n = 32$  species). We further reduced the dataset by removing duplicate occurrence records for the same species within a pixel (2.5 arc min, see below); see Table S1 for the number of occurrence points per

species used in this study. This dataset was then used to model distributions of bat species occurring in the region. Our choice of this region was based on three considerations: 1) this is a vast region comprising diverse landscapes to the south of the main rainforest bloc of the Congo basin where the taxonomy of bats is relatively well known and stable; 2) many of the bats occurring in southern Africa are endemic or near-endemic to this region, and hence the entire distribution of nearly all bat species that occur in Eswatini is encompassed by this region; and 3) this region is relatively well surveyed compared with other parts of Africa and distributional records have been published (Monadjem et al. 2020a).

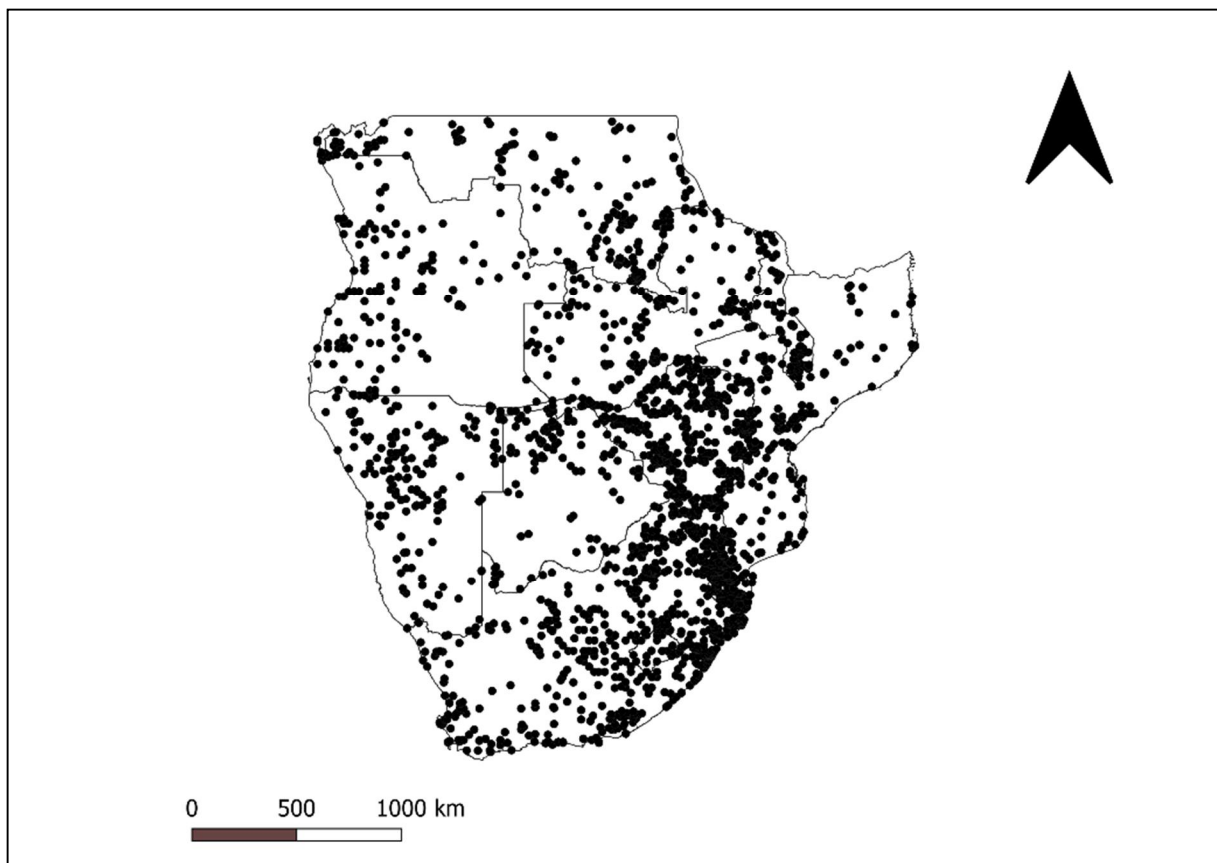


Fig. 2 – Map of the southern African region showing all the bat specimens with georeferenced localities used in this study

#### Species distribution modeling and statistical analysis

We modelled the predicted suitable environmental space of species using Maxent version 4.1.1 (Phillips et al. 2006, Phillips et al. 2020). Models were run at a resolution of approximately 5 km (2.5

arc min) using BIOCLIM variables from the WorldClim database (Hijmans et al. 2005), as well as altitude (Hijmans *et al.*, 2005), altitudinal roughness extracted from altitude using the program DIVA-GIS (available at [www.divagis.org](http://www.divagis.org)), and ecoregions as classified by Olson et al. (2001). Since BIOCLIM variables are frequently strongly correlated, we assessed the correlation between these variables in the R package 'usdm' (Naimi et al. 2014). We did this in two ways: 1) excluding one variable in every pair of variables with  $r \geq 0.8$  by removing the one with the higher Variance Inflation Factor (VIF) using the function 'vifcor'; and 2) removing variables with  $VIF > 10$  using the function 'vifstep' (Soultan et al. 2019). Both methods resulted in the inclusion of the same 12 variables, which are presented in Table 1.

We ran Maxent models in R version 3.6.2 (R Core Team 2019) using the package 'dismo' (Hijmans et al. 2013). We used hinged and categorical variables that smooth variable responses and generally improve model performance (Phillips and Dudik 2008, Merow et al. 2013). We divided bat species occurrence records from southern Africa into training (75%) and test (25%) datasets. The selection of the geographical background has important implications for the results of species distribution models (Acevedo et al. 2012; Phillips et al. 2009); a suitable background reflects the geographical space available to the species by dispersal (Zhu et al. 2014). Therefore, for each species we randomly sampled 10,000 background points from 100 km circular buffers around all occurrence points for that species. We used the value of 100 km because this is the distance that *Nycteris thebaica* (a particularly sedentary, clutter-foraging bat species) is able to cover during dispersal (Monadjem 2006), and therefore this buffered range would represent the minimum area available to any of the bat species we included in our analyses (Merow et al. 2013). We tested each model with the area under the receiver operating characteristic curve (AUC) statistic, which ranges from 0 to 1 with higher values signifying a better fit (Merow et al. 2013); values equal to or less than 0.5 indicate models no better than random, while values greater than 0.75 represent good model fit (Elith et al. 2006). We used the same 12 environmental variables (Table 1) and Maxent parameters for all



species (Cooper-Bohannon et al. 2016). We converted the predicted model outputs from Maxent (probabilities of suitability) into “presence-absence” maps using species-specific thresholds that maximized the sum of sensitivity and specificity, which is appropriate for presence-only data (Liu et al. 2013). We summed the modelled distributions of all the bats to quantify species richness using the “Raster Calculator” in QGIS (QGIS Development Team 2020).

We extracted all unique locality records for bats in Eswatini (n = 231 locality records) and prepared an actual species richness map for the country using the ‘Point to Grid’ tool in DIVA-GIS based on a “quarter-degree” grid size that is actually  $0.25^\circ \times 0.25^\circ$  in extent (approximately 24 km in length) (Hijmans et al. 2012). To test for a relationship between sampling effort (number of specimens captured) and species richness, we ran a linear regression using the “Analysis” function in DIVA-GIS. A regression of the bat species richness against area of southern African countries (south of the Zambezi-Kunene rivers) was conducted in the program R version 3.6.2 (R Core Development Team 2019).

## Results

The checklist of bat species in Eswatini has risen steadily over time from 12 species pre-1995 to 32 species at present (Table 2) (Fig. 3). This increase, however, has not been at a constant pace, with two short periods of stasis in the late 1990s and early 2000s and one longer period of stasis from 2007 to 2013 (Fig. 3).

Table 2 – The national checklist of bats of Eswatini listing all 32 species that have been collected in the country with confirmed identifications, including their global conservation status (IUCN 2019): LC – Least Concern; NE – Not Evaluated. Also shown are the dates of first mention of each species in the literature: “1997” = Monadjem (1997); “1998” = Monadjem (1998); “2005a” = Monadjem et al (2005); “2005b” = (Monadjem, 2005); “2008” = Monadjem and Reside (2008); “2016” = Shapiro and Monadjem (2016); “2020” = this study

Family	Genus	Species	IUCN status	Pre-1995	1997	1998	2005a	2005b	2008	2016	2020
Pteropodidae	<i>Epomophorus</i>	<i>crypturus</i>	LC		1						
Pteropodidae	<i>Epomophorus</i>	<i>wahlbergi</i>	LC	1	1						
Hipposideridae	<i>Hipposideros</i>	<i>caffer</i>	LC	1	1						
Rhinonycteridae	<i>Cloetis</i>	<i>percivali</i>	LC	1			1				
Rhinolophidae	<i>Rhinolophus</i>	<i>blasii</i>	LC					1			
Rhinolophidae	<i>Rhinolophus</i>	<i>clivusus</i>	LC	1	1						
Rhinolophidae	<i>Rhinolophus</i>	<i>darlingi</i>	LC		1						
Rhinolophidae	<i>Rhinolophus</i>	<i>rhodesiae</i>	NE								1
Rhinolophidae	<i>Rhinolophus</i>	<i>simulator</i>	LC	1		1					
Emballonuridae	<i>Taphozous</i>	<i>mauritanus</i>	LC			1					
Nycteridae	<i>Nycteris</i>	<i>thebaica</i>	LC	1	1						
Molossidae	<i>Chaerephon</i>	<i>pumilus</i>	LC	1	1						
Molossidae	<i>Mops</i>	<i>condylurus</i>	LC	1	1						
Molossidae	<i>Mops</i>	<i>midas</i>	LC							1	
Molossidae	<i>Tadarida</i>	<i>aegyptiaca</i>	LC	1		1					
Miniopteridae	<i>Miniopterus</i>	<i>fraterculus</i>	LC			1					
Miniopteridae	<i>Miniopterus</i>	<i>natalensis</i>	LC		1						
Vespertilionidae	<i>Afronycteris</i>	<i>nana</i>	LC	1	1						
Vespertilionidae	<i>Eptesicus</i>	<i>hottentotus</i>	LC								1
Vespertilionidae	<i>Kerivoula</i>	<i>argentata</i>	LC								1
Vespertilionidae	<i>Kerivoula</i>	<i>lanosa</i>	LC						1		
Vespertilionidae	<i>Laephotis</i>	<i>capensis</i>	LC	1	1						

Vespertilionidae	<i>Myotis</i>	<i>bocagii</i>	LC				1
Vespertilionidae	<i>Myotis</i>	<i>tricolor</i>	LC			1	
Vespertilionidae	<i>Myotis</i>	<i>welwitschii</i>	LC				1
Vespertilionidae	<i>Neoromicia</i>	<i>anchietae</i>	LC			1	
Vespertilionidae	<i>Neoromicia</i>	<i>zuluensis</i>	LC			1	
Vespertilionidae	<i>Nycticeinops</i>	<i>schlieffeni</i>	LC	1			
Vespertilionidae	<i>Pipistrellus</i>	<i>hesperidus</i>	LC		1		
Vespertilionidae	<i>Pipistrellus</i>	<i>rusticus</i>	LC				1
Vespertilionidae	<i>Scotophilus</i>	<i>dinganii</i>	LC	1	1		
Vespertilionidae	<i>Scotophilus</i>	<i>viridis</i>	LC			1	

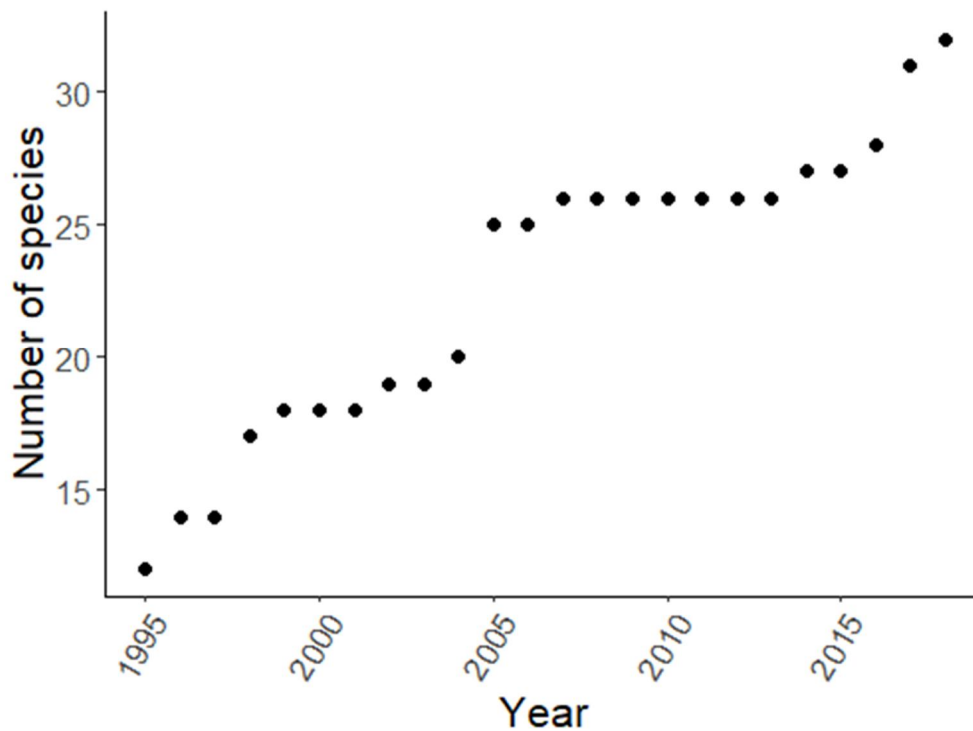


Fig. 3 – Species accumulation curve showing the increase in species richness over time from 1995 to 2018 in Eswatini

Although bats have been surveyed relatively widely in Eswatini, collecting effort has been skewed to just a few areas, particularly the northeast and northwest regions, while surveys have been more limited in the southern half of the country (Fig. 4). At a quarter-degree scale, only one complete grid had not been surveyed by 2018 (located in the centre of the country), as well as four partial grids on the borders of South Africa and Mozambique (two of which fall mostly outside of Eswatini) (Fig. 4). Based on this species richness map (Fig. 4), richness seems to vary considerably from grid to grid, but is much lower in the southern half of the country compared to the north. The number of species per grid is also low, ranging from 1-3 species in much of the south, to a maximum of 20 species in the north-east (Fig. 4). There was a strong correlation between species richness and sampling effort ( $F = 17900$ ,  $DF = 1022$ ,  $R^2 = 0.946$ ).

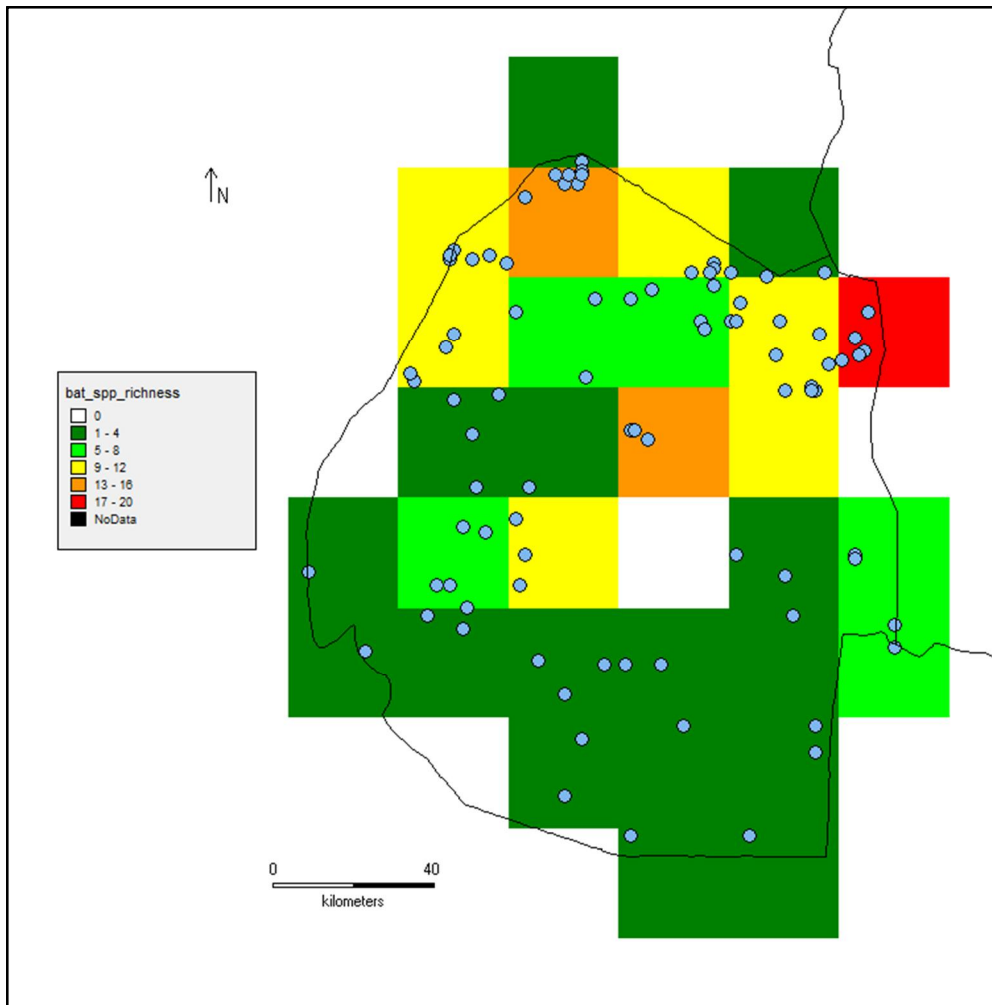


Fig. 4 – The distribution of all bat specimens collected in Eswatini (dots) laid over a “quarter-degree” grid (0.25° × 0.25°) showing the number of bat species recorded within each grid

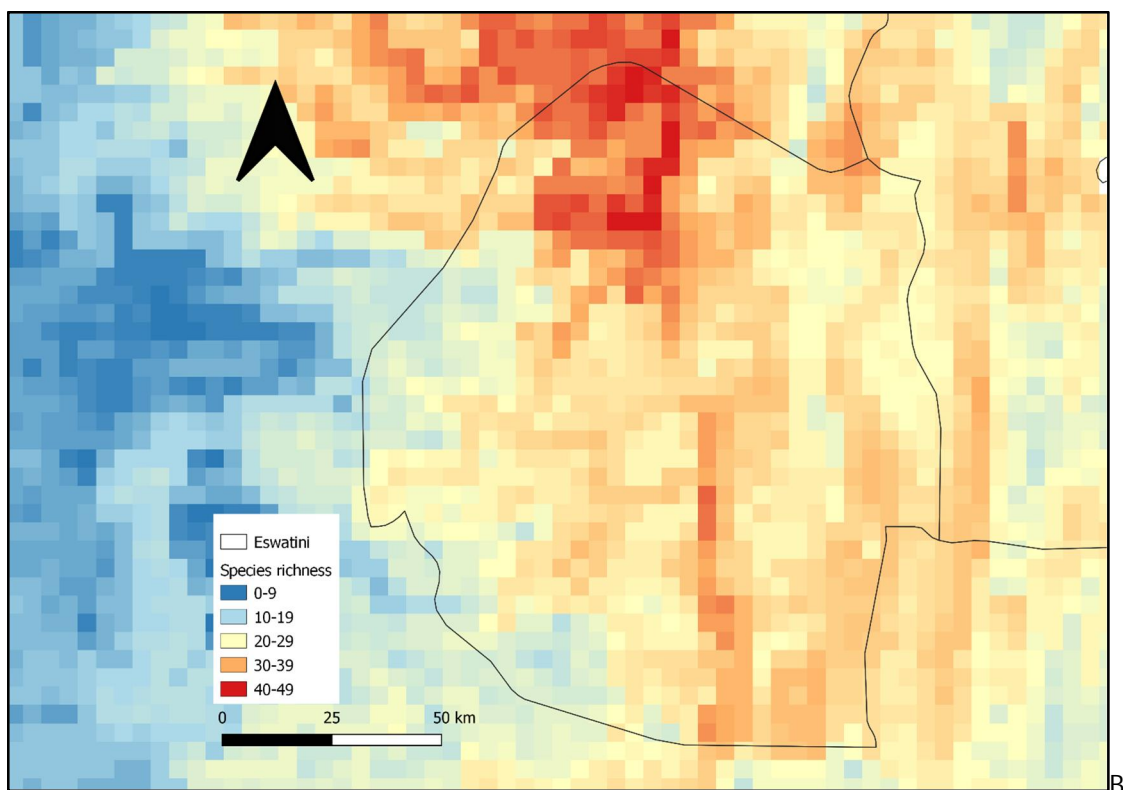
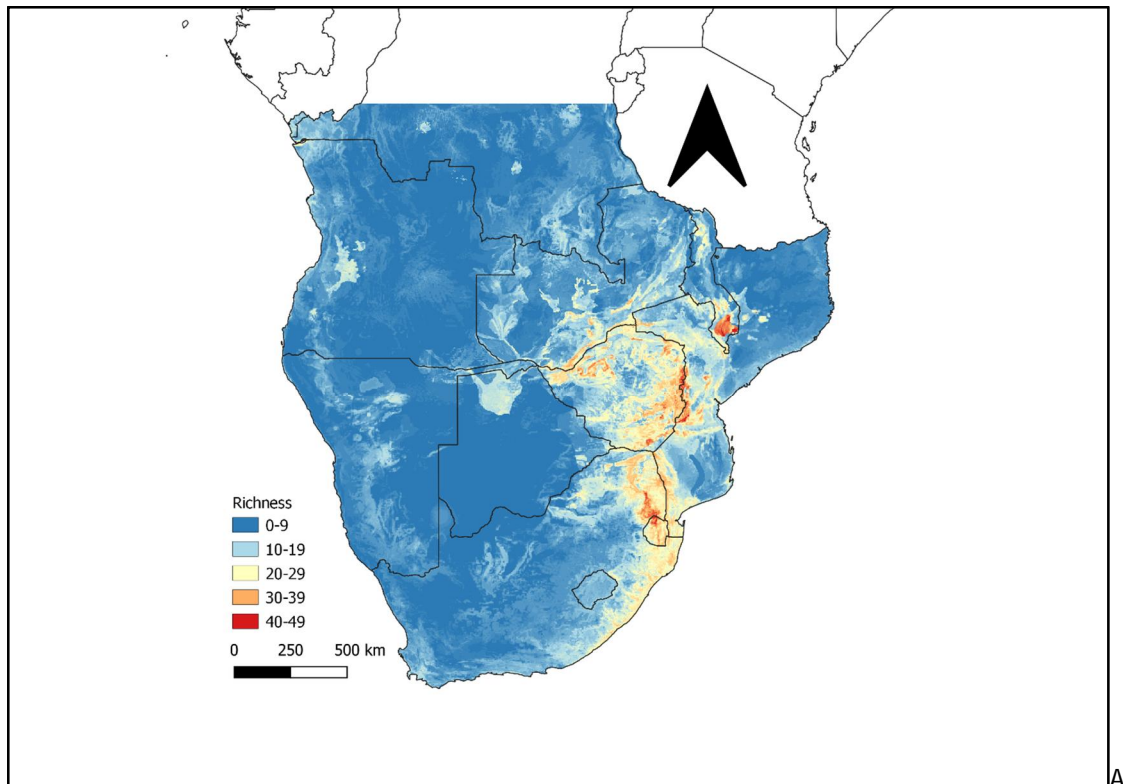


Fig. 5 – Maps showing predicted species richness of bats based on Maxent models: a) for southern Africa as defined in this study (following Monadjem et al. 2020a); and b) for Eswatini. Note that the map presented in (b) is simply a zoomed in section of (a)

However, the modelled species richness presents a different pattern (Fig. 5); the performance of each species model is presented in Table S1. On a regional scale, species richness is highest in the southern seaboard of South Africa, north through Eswatini into Zimbabwe, central Mozambique, and southern Malawi (Fig. 5a). Across the southern African region, species richness ranged from 1 to 43 species per pixel. When focusing on Eswatini, it is apparent that species richness is not uniformly high across the country (Fig. 5b). Modelled species richness was highest in the north-central parts of the country (reaching a maximum of 41 species per pixel), with a spur of high richness extending south along the boundary zone between high and low-lying regions of the country (see Fig. 1 for a digital elevation map of the country). Compared with the northern half of the country, species richness was generally lower in southern Eswatini (maximum of 35 species per pixel), especially in the southwest where richness was mostly between 11 and 15 species per pixel; the extreme west of the country also had low species richness (Fig. 5b). The median number of species per pixel in Eswatini was 21, and just 17 pixels supported less than 15 species of bats while 29 pixels support more than 35 species (Fig. S1). The total number of bat species estimated to occur in Eswatini based on species distribution models was 47 species, compared with the 32 species that have been recorded to date (Table S2).

There was a positive relationship between the area of southern African countries and the number of bats species recorded within them (Fig. S2). Based on this regression, the number of species predicted to occur in Eswatini is 34-35 species, which is 2-3 species more than currently recorded, and about 10 species less than that predicted from modeling distributions (Table S2).

A total of five species are reported for the first time in Eswatini since the last published national checklist of bats (Shapiro and Monadjem 2016). Additional details of the collecting localities and number of individuals captured for each of these species are provided in Table S3.

## Discussion

In this paper we present an updated checklist of the bats of Eswatini, which includes 32 species, an increase of five species from the most recent checklist (Shapiro and Monadjem 2016). This is lower than the species richness of many of the countries in the region. For example, Angola has 73 species (Taylor et al. 2018b), Mozambique 67 species (Monadjem et al. 2010b, Neves et al. 2018), Zambia 65 species, South Africa 63 species (Child et al. 2016), and Zimbabwe and Malawi both 62 species (Monadjem et al. 2020a). However, these countries are far larger than Eswatini and this difference accounts for most of the disparity. Correcting for surface area, the number of species recorded from Eswatini to date is 2-3 species less than what is predicted from its area alone (Fig. S2).

The relatively rapid addition of new bat species to the Eswatini checklist, including the five species added since 2016 (Table 1, S3), suggests that this latest checklist (of 32 species) is also incomplete. It is important to note that none of the additional species added to the checklist since 1995 are due to taxonomic rearrangements or recent splitting of species complexes. In every case, the additions were due to the discovery of a previously unrecorded species for the country, demonstrating the importance of field surveys. Taxonomic instability is unlikely to affect the bat fauna of a country as small as Eswatini since most African bat species complexes constitute two or more populations of non-overlapping taxa (Taylor et al. 2012, Monadjem et al. 2013, 2019). Hence, based on the continuous accumulation of new species, we expect the country total to continue to rise as further field surveys are conducted.

Our species distribution models also point to likely overlooked bat species in Eswatini. Based on our Maxent models, some additional 15 species could perhaps occur in Eswatini since there seems to be suitable environmental conditions for them in the country. It is important to note that our predicted species richness map for southern Africa is very similar to those previously published for the region (Schoeman et al. 2013, Cooper-Bohannon et al. 2016, Herkt et al. 2016), giving us confidence in our species distribution models. The 15 yet-unrecorded bat species that our models predict to occur in



Table 3 – The 15 species of bats predicted to occur in Eswatini but not yet recorded by vouchered museum specimens. Included are other details that may have an impact on whether they will occur in the country. Closest record – distance from the record to the border of Eswatini (from Monadjem et al. 2020a); habitat available – whether suitable habitat is available in Eswatini; status – migratory or resident; roost – tree, cave, or crevice roosting; taxonomic uncertainty – yes indicates that the species or species group requires revision that may affect the naming of species in the region; chance of occurrence – scored as low, medium, or high based on this information

Family	Species	Closest record (km)	Habitat available	Status	Roost type	Taxonomic uncertainty	Chance of occurrence
Emballonuridae	<i>Taphozous perforatus</i>	1	Yes	Resident	Crevice	No	High
Molossidae	<i>Chaerephon ansorgei</i>	40	Yes	Resident	Crevice	No	High
Pteropididae	<i>Eidolon helvum</i>	64	Yes	Migratory	Tree	No	High
Molossidae	<i>Otomops martiensseni</i>	250	Yes	Resident	Crevice	No	Medium
Pteropididae	<i>Rousettus aegyptiacus</i>	55	Yes	Migratory	Cave	No	Medium
Rhinolophidae	<i>Rhinolophus cohenae</i>	35	Yes	Resident	Cave	No	Medium
Vespertilionidae	<i>Laephotis botswanae</i>	280	Yes	Resident	Tree	No	Medium
Vespertilionidae	<i>Pseudoromicia rendalli</i>	110	?	Resident	Tree	No	Medium
Miniopteridae	<i>Miniopterus mossambicus</i>	680	?	Resident	Cave	No	Low
Nycteridae	<i>Nycteris macrotis</i>	275	Yes	Resident	Cave	No	Low
Rhinolophidae	<i>Rhinolophus smithersi</i>	350	?	Resident	Cave	No	Low
Vespertilionidae	<i>Nycticeinops grandidieri</i>	800	No	Resident	Tree	Yes	Low
Vespertilionidae	<i>Scotoecus albofuscus</i>	70	No	Resident	Tree	No	Low
Vespertilionidae	<i>Scotoecus hindei</i>	130	?	Resident	Tree	Yes	Low
Miniopteridae	<i>Miniopterus inflatus</i>	30	?	Resident	Cave	Yes	Low

Eswatini have been recorded at distances ranging from 1 to 800 km (Table 3) from the Eswatini border. This suggests that their occurrences in the country are not equally probable; those species occurring closer to the border are more likely to occur than those that are only known to occur much farther away. Furthermore, taking into consideration the ecology of each species can help determine the likelihood that it occurs in Eswatini. For example, in addition to suitable climate and elevation, the availability of habitat and roosts would also affect the probability of occurrence in Eswatini. Migration is another useful factor to consider since migratory species are more likely to turn up at distant localities. Finally, taxonomic uncertainties may affect distribution models because the occurrence points used in making the predictions may in fact refer to more than one species, affecting the resulting predicted distributions.

Based on these factors (see Table 3), we predict that three of the 15 species (*Taphozous perforatus*, *Chaerephon ansorgei*, and *Eidolon helvum*) have a high chance of occurring in Eswatini because the nearest records are < 65 km, and suitable habitat and roosts are available in the country. Another five species (*Otomops martiensseni*, *Rousettus aegyptiacus*, *Rhinolophus cohenae*, *Laephotis botswanae*, and *Pseudoromicia rendalli*) have a medium likelihood of occurrence based on closest records either being > 100 km away, or if <100 km but lacking suitable roosting sites in Eswatini. For example, Eswatini does not have the geology for the creation of large caves (Monadjem et al. 2003) and thus cave-roosting species, such as *Rousettus aegyptiacus* or *Rhinolophus cohenae*, are unlikely to occur. The remaining seven species have not been recorded within close distance of Eswatini and/or suitable habitat does not appear to be present in the country, and we therefore suggest that the probability of finding these species in Eswatini is rather low.

It is interesting to note that four of the five newly added species to the Eswatini bat checklist belong to the diverse family Vespertilionidae, and fifth belonging to the Rhinolophidae (Table S3). Two of the five species, *Rhinolophus rhodesiae* and *Kerivoula argentata*, were recorded within 2 km of the

national border, and without any further data may be assumed to have distributions marginal within the country. However, the remaining three species (*Eptesicus hottentotus*, *Myotis welwitschii*, and *Pipistrellus rusticus*) were captured well away from any border, suggesting that they may occur more widely in the country.

In conclusion, our study presents a bat checklist that includes 32 species in Eswatini, with up to 15 additional species that may still be recorded in the country. We suggest that species distribution models are a useful tool in gauging how complete national checklists are and identifying specific taxa that may have been overlooked, providing important baseline information to guide future conservation, management, and research strategies at both the national and regional level (Bungartz et al. 2012, Amori et al. 2012).

### Acknowledgements

Special appreciation goes to the “Strengthening the National Protected Areas Systems of Swaziland” (SNPAS) project and the project manager, Mr T. Methula, for funding this study. We would also like to extend our gratitude to the various community leaders that permitted us to conduct our study in their areas. We are also grateful to the following for assistance in the field: Mr Khoza, Mr Mkhwanazi, Mr Nhlabatsi, Mr Manana, Mr Mncina, Mr Sacolo, Mr Magagula, Mr Sibandze, Mr Dlamini, and Mr Echecolonea.

Funding: This study is based on bat specimens previously collected by different projects and therefore has no funding itself. JTS was supported by the National Science Foundation Graduate Research Fellowship under Grant No. DGE-1315138, a Student Research Grant from Bat Conservation International, a National Geographic Young Explorer’s Grant 9635-14, and The Explorers Club Exploration Fund – Mamont Scholars Program

Conflict of Interest: The authors declare that they have no conflict of interest.

## References

ACR (2019) African Chiroptera Report. AfricanBats NPC, Pretoria.

Amori G, Masciola S, Saarto J, Gippoliti S, Rondinini C, Chiozza F, Luiselli I (2012) Spatial turnover and knowledge gap of African small mammals: using country checklists as a conservation tool. *Biodiv Conserv* 21:1755-1793

Amori G, Segniagbeto GH, Decher J, Assou D, Gippoliti S, Luiselli L (2016) Non-marine mammals of togo (West Africa): an annotated checklist. *Zoosystema* 38:201–244

Ansell WFH (1978) The mammals of Zambia. The National Parks & Wildlife Service, Chilanga

Barratt EM, Deaville R, Burland TM, Bruford MW, Jones G, Racey PA, Wayne RK (1997) DNA answers the call of pipistrelle bat species. *Nature* 387:138–139

Bates PJJ, Cameron K, Pearch MJ, Hayes B (2013) A Review of the bats (Chiroptera) Of the Republic of Congo, including eight species new to the country. *Acta Chiropterol* 15:313–340

Boyles JG, Cryan PM, McCracken GF, Kunz TK (2011) Economic importance of bats in agriculture. *Science* 332:41–42

Bungartz F, Ziemmeck F, Tirado N, Jaramillo P, Herrera H, Jimenez-Uzategui, G (2011) The neglected majority – biodiversity inventories as an integral part of conservation biology. In: Wolff M, Gardener M (eds) *Fifty years of conservation in the Galapagos: the role of science in developing a sustainable future*. Routledge, Oxon, pp 119-142

Burgin CJ, Colella JP, Kahn PL, Upham NS (2018) How many species of mammals are there? *J Mammal* 99:1–14

Child MF, Roxburgh L, Do Linh San E, Raimondo D, Davies-Mostert HT (2016) The 2016 Red List of

Mammals of South Africa, Swaziland and Lesotho. South African National Biodiversity Institute and Endangered Wildlife Trust, South Africa.

Cooper-Bohannon R, Rebelo H, Jones G, Cotterill F, Monadjem A, Schoeman MC, Taylor P, Park K (2016) Predicting bat distributions and diversity hotspots in southern Africa. *Hystrix* 27: 38-48

Crawford-Cabral J (1989) A list of Angolan Chiroptera with notes on their distribution. *Garcia de Orta, Ser Zool, Lisboa* 13:7–48

Decher J, Oppong J, Fahr J (2001) Rapid assessment of small mammals at Draw River, Boi-Tano, and Krokosua Hills. In: Mcculloch J, Decher J, Guba-Kpelle D (eds) A biological assessment of the terrestrial ecosystems of the Draw River, Boi-Tana, Tano Nimiri and Krokosua Hills forest reserves, southwestern Ghana Conservation. International, New York, USA, pp 59–68

Dietz C, von Helversen S (2004) Illustrated Identification key to the bats of Europe. Electronic Publication, Tuebingen, Germany

Elith J, Graham CH, Anderson RP, Dudík M, Ferrier S, Guisan A, Hijmans RJ, Huettmann F, Leathwick JR, Lehmann A, Li J, Lohmann LG, Loiselle BA, Manion G, Moritz C, Nakamura M, Nakazawa Y, Overton J McC, Peterson AT, Phillips SJ, Richardson KS, Scachetti-Pereira R, Schapire R E, Soberón J, Williams S, Wisz MS, Zimmermann NE (2006) Novel methods improve prediction of species' distributions from occurrence data. *Ecography* 29: 129–151

Fahr J, Ebigbo NM (2003) A conservation assessment of the bats of the Simandou Range, Guinea, with the first record of *Myotis welwitschii* (Gray, 1866) from West Africa. *Acta Chiropterol* 5:125–141

Fisher MA, Vinson JE, Gittleman JL, Drake JM (2018) The description and number of undiscovered mammal species *Ecol Evol* 8:3628–3635

Happold DCD, Happold M, Hill JE (1987) The bats of Malawi. *Mammalia* 51:337–414

Herkt KMB, Barnikel G, Skidmore AK, Fahr J (2016) A high-resolution model of bat diversity and

endemism for continental Africa. *Ecol Model* 320:9–28

Herkt KMB, Skidmore AK, Fahr J (2017) Macroecological conclusions based on IUCN expert maps: a call for caution. *Global Ecol Biogeogr* 26:930-941

Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A (2005) Very high resolution interpolated climate surfaces for global land areas. *Int J Climat* 25:1965–1978

Hijmans RJ, Guarino L, Mathur P (2012) DIVA-GIS, version 75 manual

Hijmans RJ, Phillips S, Leathwick J, Elith J (2013) dismo: species distribution modeling: R package version 1.1-4.

IUCN (2019) The IUCN Red List of Threatened Species Version 20181. <http://www.iucnredlist.org>. Accessed 27 October 2018

Jan CMI, Frith K, Glover AM, Butlin RK, Scott CD, Greenaway F, Ruedi M, Frantz AC, Dawson DA, Altringham JD (2010) *Myotis alcaethoe* confirmed in the UK from mitochondrial and microsatellite DNA. *Acta Chiropterol* 12:471–483

Kingdon J, Happold D, Butynski T, Hoffmann M, Happold M, Kalina J (eds)(2013) *Mammals of Africa* (6 vols). Bloomsbury Publishing, London

Kock D (1969) Die Fledermaus-Fauna des Sudan. *Abhandlungen der Senckenbergischen Naturforschenden Gesellschaft* 521:1–238

Kunz TH, de Torrez EB, Bauer D, Lobova T, Fleming TH (2011) Ecosystem services provided by bats. *Ann New York Acad Sci* 1223:1–38

Liu C, White M, Newell G (2013) Selecting thresholds for the prediction of species occurrence with presence-only data. *J Biogeogr* 40:778–789

Mayer F, von Helversen O (2001) Sympatric distribution of two cryptic bat species across Europe. *Biol J Linn Soc* 74:365–374

- Merow C, Smith MJ, Silander JA (2013) A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. *Ecography* 36: 1058–1069
- Mitchell-Jones AJ, Hutson AM, Racey PA (1993) The growth and development of bat conservation in Britain. *Mammal Rev* 23:139–148
- Monadjem A (1997) An annotated checklist of the mammals of Swaziland. Conservation Trust of Swaziland, Manzini
- Monadjem A (1998) Mammals of Swaziland. Conservation Trust of Swaziland & Big Game Parks, Mbabane
- Monadjem A (2005) Recording of the call of the peak-saddle horseshoe bat (*Rhinolophus blasii* Peters, 1867) from Swaziland. *Afr Bat Conserv News* 3:5–6
- Monadjem A (2006) Longevity and movement of the common slit-faced bat *Nycteris thebaica*. *African Bat Conservation News* 9: 7
- Monadjem A, Fahr J (2007) A rapid survey of bats from North Lorma, Gola and Grebo National Forests, Liberia, with notes on shrews and rodents. In: Hoke P, Demey R, Peal A (eds) A rapid biological assessment of North Lorma, Gola and Grebo National Forests, Liberia. Conservation International, RAP Bulletin of Biological Assessment, Washington, DC 44, pp 47–58
- Monadjem A, Reside A (2008) The influence of riparian vegetation on the distribution and abundance of bats in an African savanna. *Acta Chiropterol* 10:339–348
- Monadjem A, Boycott RC, Parker V, Culverwell J (2003) Threatened vertebrates of Swaziland. Swaziland Red Data Book: fishes, amphibians, reptiles, birds and mammals. Ministry of Tourism, Environment and Communication, Swaziland
- Monadjem A, Cohen L, De Wet K (2005) Rediscovery of the short-eared trident bat (*Cloeotis percivali* Thomas 1901) in Swaziland. *Afr Bat Conserv News* 6:2–3

- Monadjem A, Taylor PJ, Cotterill FPD, Schoeman MC (2010a) Bats of southern and central Africa: a biogeographic and taxonomic synthesis. University of the Witwatersrand Press, Johannesburg
- Monadjem A, Schoeman MC, Reside A, Pio DV, Stoffberg S, Bayliss J, Cotterill FPD, Curran M, Kopp M, Taylor PJ (2010b) A recent inventory of the bats of Mozambique with documentation of seven new species for the country. *Acta Chiropterol* 12:371–391
- Monadjem A, Goodman S, Stanley W, Appleton B (2013) A cryptic new species of *Miniopterus* from south-eastern Africa based on molecular and morphological characters. *Zootaxa* 3746:123–142
- Monadjem A, Taylor PJ, Denys C, Cotterill FPD (2015) Rodents of sub-Saharan Africa: a biogeographic and taxonomic synthesis De Gruyter, Berlin
- Monadjem A, Richards L, Denys C (2016) An African bat hotspot: the exceptional importance of Mount Nimba for bat diversity. *Acta Chiropterol* 18:359–375
- Monadjem A, Shapiro JT, Richards LR, Karabulut H, Crawley WY, Nielsen IB, Hansen AJ, Bohmann K, Mourier T (2019) Systematics of West African *Miniopterus* with the description of a new species. *Acta Chiropterol* 21:237–256
- Monadjem A, Taylor PJ, Cotterill FPD, Schoeman MC (2020a) Bats of southern and central Africa: a biogeographic and taxonomic synthesis, 2<sup>nd</sup> edn. Wits University Press, Johannesburg
- Monadjem A, Demos TC, Dalton DL, Webala PW, Musila S, Kerbis Peterhans, JC Patterson BD (2020b) A revision of pipistrelle-like bats (Mammalia: Chiroptera: Vespertilionidae) in East Africa with the description of new genera and species. *Zoological Journal of the Linnean Society* (early view)
- Musila S, Monadjem A, Webala PW, Patterson BD, Hutterer R, De Jong YA, Butynski TM, Mwangi G, Chen ZZ, Jiang XL (2019) An annotated checklist of mammals of Kenya. *Zool Res* 40:1–51
- Naimi B, Hamm NAS, Groen TA, Skidmore AK, Toxopeus AG (2014). Where is positional uncertainty a problem for species distribution modelling? *Ecography* 37, 191–203



Neves IQ, Mathias ML, Bastos-Silveira C (2018) The terrestrial mammals of Mozambique: integrating dispersed biodiversity data. *Bothalia* 48(1):a2330

Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GVN, Underwood EC, D'Amico J, Itoua I, Strand HE, Morrison JC, Loucks CJ, Allnutt TF, Ricketts TH, Kura Y, Lamoreux JF, Wettengel WW, Hedao P, Kassem KR (2001) Terrestrial ecoregions of the World: a new map of life on Earth *BioScience* 51:933

Phillips SJ, Dudík M (2008) Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. *Ecography* 31:161–175

Phillips SJ, Anderson RP, Schapire RE (2006) Maximum entropy modeling of species geographic distributions. *Ecol Model* 190:231–259

Phillips SJ, Dudík M, Schapire RE (2020) Maxent software for modeling species niches and distributions (version 3.4.1). Available from:

[url://biodiversityinformatics.amnh.org/open\\_source/maxent/](http://biodiversityinformatics.amnh.org/open_source/maxent/)

QGIS Development Team (2020) QGIS Geographic Information System Open Source Geospatial Foundation Project. <http://qgisosgeorg>

R Core Development Team (2019) A language and environment for statistical computing Austria, Vienna

Rautenbach IL (1982) Mammals of the Transvaal *Ecoplan Monograph* 1:1–211

Rautenbach IL, Whiting MJ, Fenton MB (1996) Bats in riverine forests and woodlands: a latitudinal transect in southern Africa. *Can J Zool* 74:312–322

Reddy S, Davalos LM (2003) Geographical sampling bias and its implications for conservation priorities in Africa. *J Biogeogr* 30:1719–1727

Schlitter DA, Robbins LW, Buchanan SA (1982) Bats of the Central African Republic (Mammalia:

Chiroptera). *Ann Carnegie Mus* 51:133–155

Schoeman MC, Cotterill FPD, Taylor PJ, Monadjem A (2013) Using potential distributions to explore environmental correlates of bat species richness in southern Africa: Effects of model selection and taxonomy. *Current Zool* 59: 279–293

Shapiro JT, Sovie AR, Faller CR, Monadjem A, Fletcher RJ, Mccleery RA (2020) Ebola spillover correlates with bat diversity. *Eur J Wildl Res* 66:12

Shapiro JT, Monadjem A (2016) Two new bat species for Swaziland and a revised list for the country. *Mammalia* 80:353–357

Simmons N (2005) Order Chiroptera. In: Wilson DE, Reeder DM (eds) *Mammal species of the World: a taxonomic and geographic reference*. John Hopkins University Press, Baltimore

Simmons NB, Cirranello AL (2018) *Bat Species of the World: A taxonomic and geographic database*. [www.batnames.org](http://www.batnames.org) 17 December 2020.

Soultan A, Wikelski M, Safi K (2019) Risk of biodiversity collapse under climate change in the Afro-Arabian region. *Scientific Reports* 9: 955

Taylor PJ (1999) The role of amateurs in the growth of bat conservation and research in South Africa. *S Afr J Zool* 34:19–25

Taylor PJ, Stoffberg S, Monadjem A, Schoeman MC, Bayliss J, Cotterill FPD (2012) Four new bat species (*Rhinolophus hildebrandtii* complex) reflect Plio-Pleistocene divergence of dwarfs and giants across an Afromontane archipelago. *PLoS ONE* 7(9):e41744

Taylor PJ, Grass I, Alberts AJ, Joubert E, Tschardt T (2018a) Economic value of bat predation services – A review and new estimates from macadamia orchards. *Ecosys Serv* 30:372–381

Taylor PJ, Neef G, Keith M, Weier S, Monadjem A, Parker DM (2018b) Tapping into technology and the biodiversity informatics revolution: updated terrestrial mammal list of Angola, with new records

from the Okavango Basin. ZooKeys 779:51–88

Yalden DW, Largen MJ, Kock D, Hillman JC (1996) Catalogue of the mammals of Ethiopia and Eritrea: 7, Revised checklist, zoogeography and conservation. Trop Zool 9:73–164

### Supplementary material

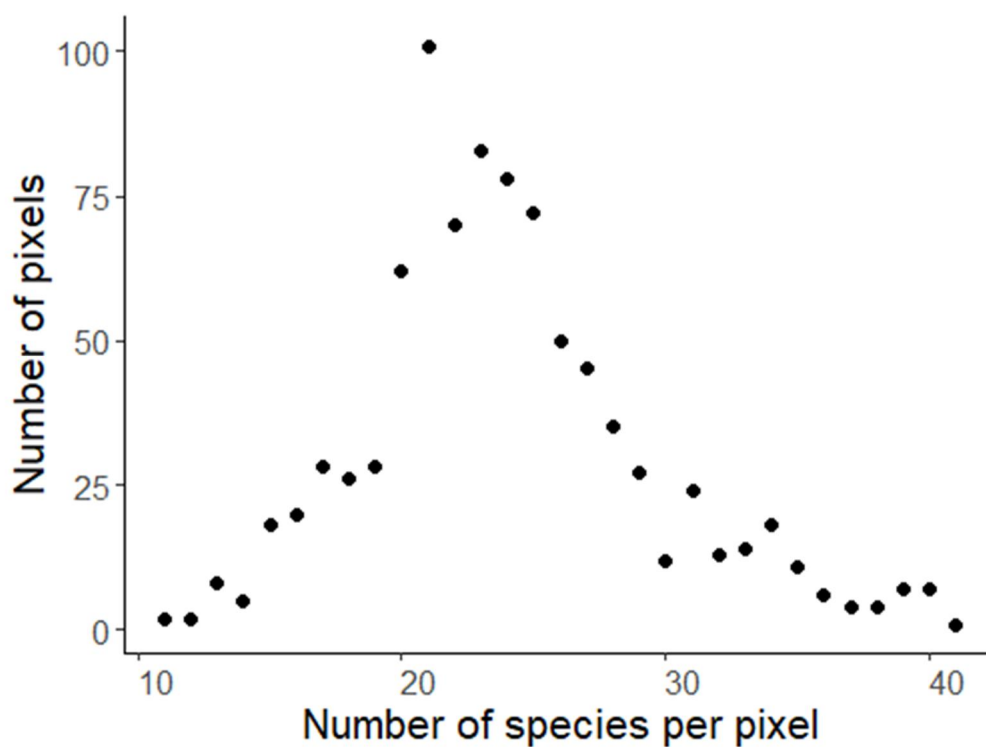


Fig. S1 – A count of the number of bat species per pixels in Eswatini based on Maxent species distribution models, ranging from a minimum of 11 bats per pixel up to a maximum of 41 bats per pixel

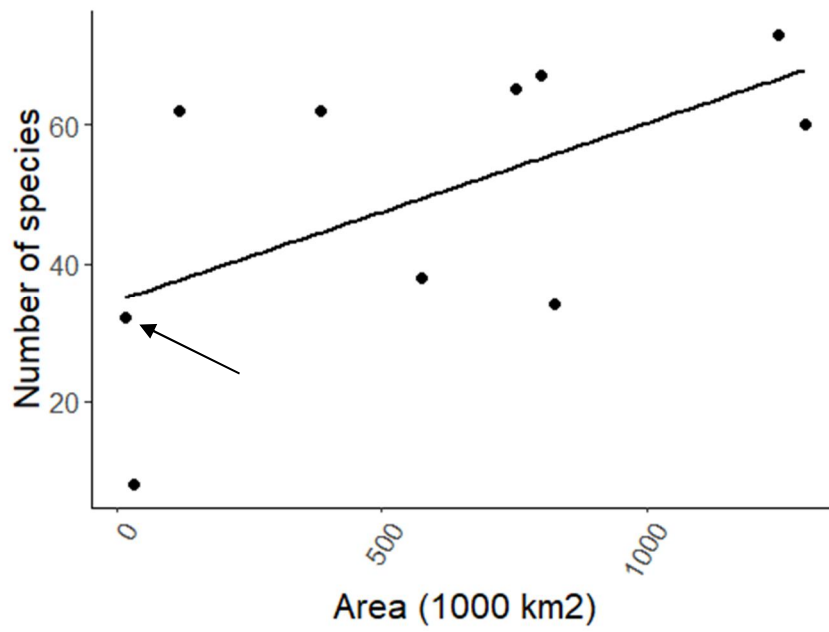


Fig. S2 – A linear regression of species richness against surface area for countries in southern Africa south of the Zambezi-Kunene rivers. The arrow points to Eswatini. Species richness =  $0.0000267 \cdot \text{area} + 34.25$  ( $F_{1,8} = 392$ ,  $P = 008318$ )

Table S1 – The 92 species used in the distribution models, number of occurrence points per species, model performance based on training and test AUC values and the individual contributions (%) of the 13 environmental variables (see Table 1 for a full description of each variable) used in this study. See the Methods for more details.

Species	Total no. of points	AUC (training)	AUC (test)	Altitude	Roughness	BIO2	BIO3	BIO8	BIO9	BIO13	BIO14	BIO15	BIO18	BIO19	Ecoregions
<i>Afronycteris nana</i>	237	0.8532	0.8354	1.2158	7.0918	16.4326	5.1115	1.6581	11.4016	12.8542	5.8932	1.4130	0.8772	0.3387	35.7124
<i>Chaerephon ansorgei</i>	33	0.9452	0.7636	3.8550	19.0007	3.8405	0.0000	0.1178	1.7195	0.2287	1.4951	0.6415	7.8641	3.4871	57.7500
<i>Chaerephon bivittatus</i>	14	0.9612	0.5445	0.9556	19.6017	0.0000	0.0000	0.0000	1.1705	0.0000	1.4648	12.9121	1.5171	11.3189	51.0593
<i>Chaerephon chapini</i>	17	0.9308	0.5131	0.9139	5.5956	1.8443	0.0000	1.5131	0.0000	3.9676	20.1952	0.0000	0.0000	0.6691	65.3012
<i>Chaerephon nigeriae</i>	43	0.9221	0.7796	0.3910	0.8433	5.6236	11.3447	0.0000	0.5892	6.6316	60.0022	0.0000	3.5109	1.1369	9.9266
<i>Chaerephon pumilus</i>	220	0.9303	0.8798	35.6290	2.3288	0.3260	0.6844	1.6550	9.1520	1.5603	3.9001	0.6091	7.9390	0.5834	35.6330
<i>Cistugo lesueuri</i>	19	0.9825	0.8068	28.9366	3.3900	0.0000	0.2051	0.0000	27.9977	0.0000	21.7848	0.0000	0.0000	0.0000	17.6858
<i>Cistugo seabrae</i>	13	0.9937	0.8445	1.4196	14.8602	0.0000	0.0148	1.1552	0.0000	14.9403	1.5762	0.0000	54.6127	4.4628	6.9585
<i>Cloeotis percivali</i>	35	0.9564	0.8870	1.7821	21.6107	6.2103	0.0000	0.0632	22.9375	1.4268	0.0000	0.0000	7.6078	8.0420	30.3197
<i>Eidolon helvum</i>	82	0.8624	0.7060	12.9345	18.2607	1.3297	0.0204	0.1540	2.4906	0.0000	3.6116	0.4624	0.1774	2.3003	58.2584
<i>Epomophorus angolensis</i>	20	0.9726	0.9852	4.5944	4.2278	0.5092	1.3528	0.0000	0.0000	6.1776	33.4143	0.0094	2.8973	2.4099	44.4073
<i>Epomophorus crypturus</i>	113	0.9008	0.8103	0.2702	1.3863	0.0000	12.6318	0.1877	3.6182	19.3685	0.0000	2.4149	2.1421	2.7560	55.2242
<i>Epomophorus dobsonii</i>	30	0.9393	0.8816	4.1088	2.9922	2.3809	1.2891	0.0000	4.0678	34.1131	22.4782	0.0000	0.0000	10.0407	18.5292
<i>Epomophorus labiatus</i>	25	0.9797	0.7978	1.0146	2.5009	14.2655	5.6922	0.0968	2.6638	22.1620	8.0059	0.0000	8.1289	0.4938	34.9757
<i>Epomophorus wahlbergi</i>	176	0.9084	0.8623	3.8243	16.9064	1.4962	1.2361	0.0282	6.4406	0.1478	24.3839	0.1153	14.0915	2.3204	29.0094
<i>Epomops franqueti</i>	12	0.9555	0.7098	0.0000	7.9718	0.2625	0.9954	4.8176	54.0177	0.0000	0.2743	0.0000	6.1448	2.4635	23.0524
<i>Eptesicus hottentotus</i>	50	0.9339	0.7693	4.0785	52.8493	2.5194	0.0000	0.6762	6.8676	0.1278	0.0000	6.2648	0.9092	0.7946	24.9126
<i>Glauconycteris argentata</i>	7	0.9928	0.9998	1.5726	4.9904	0.0000	0.0000	0.0000	19.8852	18.5073	0.0000	0.0000	0.0000	16.6869	38.3576
<i>Glauconycteris variegata</i>	42	0.9238	0.7142	27.4552	0.9681	0.0956	11.1731	0.2627	2.1851	16.2946	9.3796	0.0000	0.7953	2.1700	29.2206
<i>Hipposideros caffer</i>	263	0.8440	0.7949	7.8616	17.7340	0.2198	8.3113	0.7213	9.7156	14.2342	0.2389	0.2926	0.3677	0.0597	40.2433
<i>Hipposideros ruber</i>	37	0.9235	0.7582	0.0000	24.0274	7.3054	0.7577	0.0000	0.1329	51.1290	1.0131	0.0000	1.9979	7.3687	6.2678
<i>Hypsignathus monstrosus</i>	10	0.9829	0.9801	1.6269	19.9407	14.0432	0.0000	0.0000	27.4990	0.0000	0.0000	0.0000	13.3006	1.1572	22.4322
<i>Kerivoula argentata</i>	32	0.9493	0.6996	11.0618	6.6313	4.5121	3.6130	0.0000	0.0694	19.0043	11.5596	0.0626	3.0510	3.1440	37.2910
<i>Kerivoula lanosa</i>	31	0.9462	0.8545	0.2771	7.4098	0.0186	2.4834	0.9574	5.8713	1.3039	25.9882	0.0000	0.5044	0.4735	54.7124
<i>Laephotis angolensis</i>	6	0.9767	0.9375	0.0314	7.1930	0.9708	0.0000	0.0000	0.0000	16.8448	29.4669	0.0000	0.3131	6.6266	38.5534
<i>Laephotis botswanae</i>	37	0.9498	0.8231	0.8497	9.8305	0.6596	0.3023	0.0000	11.8609	7.6325	1.5413	0.7878	0.7207	0.8353	64.9794

<i>Laephotis capensis</i>	395	0.8415	0.8149	0.9225	5.0193	0.9398	3.5643	0.4605	28.1680	8.5279	10.8716	0.5138	3.0426	2.5063	35.4633
<i>Laephotis stanleyi</i>	14	0.9746	0.8354	0.0000	1.3885	2.1490	0.0000	2.2054	0.0000	0.0000	0.0000	0.6607	0.0000	15.9191	77.6774
<i>Lavia frons</i>	11	0.9777	0.8693	0.0000	2.1792	4.4226	0.1132	0.0000	32.1124	0.2672	9.2142	0.0000	0.0032	1.5024	50.1856
<i>Macronycteris vittatus</i>	84	0.8848	0.8469	0.1212	5.1241	1.6971	0.1409	0.0043	5.5559	22.4094	0.0000	26.2942	0.3016	2.3469	36.0045
<i>Micropteropus pusillus</i>	25	0.9576	0.8944	4.1369	6.0705	43.7518	0.5642	0.1036	11.8362	4.6980	0.2943	0.0000	7.8168	0.5598	20.1678
<i>Mimetillus thomasi</i>	13	0.8929	0.7340	0.2539	0.0674	17.0389	0.7319	0.0000	3.5222	44.7014	15.8144	0.0000	0.0000	4.3403	13.5296
<i>Miniopterus cf natalensis</i>	55	0.9372	0.8350	2.0445	5.4847	13.9829	10.3590	0.0000	2.0779	14.2389	0.0186	0.0000	5.5497	4.1043	42.1395
<i>Miniopterus fraterculus</i>	21	0.9953	0.9400	1.0821	17.3542	0.1048	0.0000	0.0000	6.9451	0.0000	40.9233	0.0000	11.1223	0.0000	22.4683
<i>Miniopterus inflatus</i>	20	0.9705	0.5760	1.5403	19.5136	0.7918	12.5234	0.0000	2.3230	4.6542	11.7527	0.0000	0.2823	8.2144	38.4042
<i>Miniopterus minor</i>	6	0.9845	0.7443	0.0000	26.7413	0.0000	0.0000	0.0000	0.0000	10.4897	34.5850	0.0000	1.3585	0.0000	26.8256
<i>Miniopterus mossambicus</i>	12	0.9945	0.8640	0.0000	28.0162	0.0000	0.0000	0.0790	1.8782	10.3093	29.8142	4.7041	13.3038	0.0000	11.8952
<i>Miniopterus natalensis</i>	182	0.9019	0.8784	3.4869	25.5127	0.2111	0.6353	0.0355	27.6593	8.1720	18.9091	0.0054	2.5438	0.7823	12.0466
<i>Mops condylurus</i>	112	0.9249	0.9327	36.5604	0.2296	1.6518	4.5063	2.8873	5.1039	8.3060	1.2671	0.1023	0.4459	0.0008	38.9387
<i>Mops midas</i>	36	0.9550	0.9261	2.9678	0.0498	2.7724	0.0108	16.1316	17.9252	0.1692	0.0518	0.3173	0.0013	0.3963	59.2064
<i>Mops niveiventer</i>	21	0.9703	0.7983	4.6054	5.2521	10.5126	1.3231	0.0000	0.1360	37.0715	5.4996	0.1160	0.5566	2.0858	32.8414
<i>Myonycteris angolensis</i>	17	0.9601	0.9708	0.0630	28.5964	0.4917	9.4142	0.0000	0.0000	0.0000	10.4808	1.8088	24.0523	4.7958	20.2969
<i>Myonycteris goliath</i>	7	0.9935	0.9819	0.0000	40.2764	0.0000	0.0243	0.0000	0.1658	0.0000	33.2158	8.0430	9.5458	0.0000	8.7290
<i>Myotis bocagii</i>	49	0.9032	0.7447	3.5968	23.7465	2.4614	0.0000	0.2623	3.4616	17.1170	0.0000	3.3546	2.8541	0.1632	42.9823
<i>Myotis tricolor</i>	71	0.9433	0.9562	1.6686	37.7972	0.8220	6.4332	0.0000	6.5687	0.0105	28.4150	0.2689	6.0518	2.2966	9.6677
<i>Myotis welwitschii</i>	35	0.9231	0.7658	0.0000	27.0073	1.6154	4.7562	0.0000	9.2968	10.5779	8.3692	7.2023	7.0558	11.7712	12.3480
<i>Neoromicia anchietae</i>	56	0.9462	0.8370	9.8689	3.9541	2.5035	0.0026	0.0000	19.6384	16.8214	0.0000	0.0000	0.2198	0.0988	46.8927
<i>Neoromicia zuluensis</i>	96	0.9034	0.8602	2.7323	7.6224	1.4583	1.6595	2.9758	16.8252	5.7586	0.0000	1.8101	2.1305	2.3172	54.7101
<i>Nycteris grandis</i>	15	0.9794	0.8348	29.6898	0.0089	0.0000	0.5515	1.0059	0.0043	5.5871	0.0000	0.0000	0.0706	9.2506	53.8313
<i>Nycteris hispida</i>	54	0.8918	0.8156	5.0432	3.6519	5.1881	0.1465	3.4084	2.9733	41.9111	7.0944	0.6973	0.2657	0.0962	29.5239
<i>Nycteris macrotis</i>	56	0.8901	0.7554	0.6626	3.4212	7.7364	0.0000	1.1729	0.0156	31.4818	0.0000	0.7033	3.6153	6.2222	44.9687
<i>Nycteris thebaica</i>	363	0.8097	0.8013	4.3870	12.9899	1.9347	27.4229	0.0814	6.7169	3.8348	0.0000	0.0589	0.6264	0.0000	41.9471
<i>Nycteris woodi</i>	25	0.9650	0.8781	2.0335	1.8798	0.0000	0.9470	0.4850	0.5055	1.0161	7.7741	0.3259	0.7296	4.0066	80.2969
<i>Nycticeinops grandidieri</i>	9	0.9758	0.7774	0.0000	8.2784	0.0000	0.5292	0.0000	0.7706	45.3365	11.6686	0.1254	0.0000	7.2432	26.0481
<i>Nycticeinops schlieffeni</i>	152	0.9255	0.8549	2.7654	4.9391	3.9315	7.0776	3.8613	7.2623	8.8766	0.0000	0.1786	1.2659	1.4342	58.4075
<i>Otomops martiensseni</i>	16	0.9950	0.7687	10.9056	15.5538	2.3339	1.5572	1.0921	3.9213	1.6908	0.0000	0.0000	0.0000	0.5571	62.3883
<i>Pipistrellus hesperidus</i>	74	0.9597	0.8685	2.3366	7.3454	0.0000	1.1846	0.2715	14.0020	10.9063	30.0142	0.0321	0.2449	3.9183	29.7442
<i>Pipistrellus rusticus</i>	58	0.9348	0.8794	0.0365	3.3261	3.4176	8.9899	3.9811	12.5936	13.9069	0.0000	1.8814	0.0000	0.3192	51.5478
<i>Plerotes anchietae</i>	10	0.9822	0.9927	26.5799	28.8237	0.0000	0.0000	0.3156	0.0176	19.1584	13.1813	0.0000	0.4905	3.0552	8.3778
<i>Pseudoromicia rendalli</i>	14	0.9606	0.7776	32.2637	0.0000	0.0000	0.0299	7.2714	0.0000	0.6096	0.0000	1.3704	0.0000	2.3077	56.1473
<i>Pseudoromicia tenuipinnis</i>	14	0.9730	0.8159	0.3173	0.1697	0.0000	2.7283	0.0000	41.0438	0.0000	0.6371	0.0035	5.3097	0.1805	49.6100
<i>Rhinolophus blasii</i>	46	0.9585	0.9394	0.0000	25.7082	0.4710	7.4229	0.0000	6.2197	17.4104	8.0682	3.4832	0.2859	3.1231	27.8074
<i>Rhinolophus capensis</i>	23	0.9964	0.9432	2.1479	1.1290	0.2134	0.3257	14.5860	0.0000	0.0047	0.1218	19.6917	1.5030	53.6176	6.6594
<i>Rhinolophus cf lobatus</i>	24	0.9728	0.8775	0.0000	12.4794	0.4596	0.0000	0.0000	1.2886	6.5792	0.0000	5.7057	0.9012	28.0060	44.5803

<i>Rhinolophus cf mossambicus</i>	19	0.9892	0.8490	1.2832	19.0425	0.0000	0.0000	0.0000	0.0000	0.0000	13.4920	0.4782	7.6000	1.6361	56.4680
<i>Rhinolophus clivosus</i>	197	0.9394	0.9091	0.9097	33.1864	1.0386	11.6198	8.3045	6.5032	3.2384	21.3214	0.1233	1.7769	0.1975	11.7805
<i>Rhinolophus cohenae</i>	6	0.9974	0.9946	0.0000	41.7889	0.0000	0.0000	0.0000	4.5746	0.0000	20.7921	0.0000	7.0924	0.2180	25.5340
<i>Rhinolophus damarensis</i>	39	0.9830	0.7128	10.0760	23.1949	0.5354	2.1802	0.0000	1.3450	21.2377	2.9865	1.6850	6.9147	3.7669	26.0777
<i>Rhinolophus darlingi</i>	103	0.9473	0.9318	0.8018	1.7941	4.4229	2.5362	0.0437	18.5562	0.4994	7.6533	0.0892	10.2271	0.8123	52.5640
<i>Rhinolophus denti</i>	19	0.9766	0.8949	0.0000	12.5557	9.7070	0.5099	0.0000	5.5230	2.3581	3.2699	0.0000	0.4380	0.2733	65.3652
<i>Rhinolophus fumigatus</i>	88	0.9373	0.7678	3.4506	27.5310	0.1970	3.0786	0.4679	5.3982	6.4739	5.7233	20.3085	0.5116	0.3033	26.5560
<i>Rhinolophus lobatus</i>	38	0.9588	0.8916	17.5468	0.0502	0.6370	1.6244	0.9505	0.7766	2.2627	0.0000	5.5399	0.0000	1.9773	68.6346
<i>Rhinolophus mossambicus</i>	119	0.9415	0.9433	0.9509	2.1741	3.0885	4.3495	0.0315	3.6046	14.5547	1.0594	13.1240	0.7391	0.2175	56.1062
<i>Rhinolophus rhodesiae</i>	43	0.9316	0.9169	0.0674	22.0597	4.3406	2.0565	0.4933	6.6218	15.3545	0.0000	1.8654	0.0000	0.7488	46.3920
<i>Rhinolophus simulator</i>	100	0.9414	0.9208	0.1590	18.2124	0.2207	16.6466	0.1365	16.8073	1.6791	5.2029	0.1603	20.2972	5.2596	15.2183
<i>Rhinolophus smithersi</i>	18	0.9929	0.9438	0.0000	15.5716	0.0000	0.0000	8.0517	10.3298	0.0000	9.4055	3.6980	0.7465	0.6702	51.5266
<i>Rhinolophus swinnyi</i>	8	0.9945	0.9971	0.2351	4.4765	2.6788	0.0000	0.0000	0.1818	0.0000	52.4311	0.0000	0.0000	0.0000	39.9968
<i>Rousettus aegyptiacus</i>	72	0.9138	0.8508	3.6178	31.1238	7.2709	7.4403	0.0000	6.1326	0.1772	22.2451	0.3421	3.0725	0.5288	18.0489
<i>Sauromys petrophilus</i>	63	0.9508	0.9248	0.4871	19.7270	3.7609	1.6278	0.5912	0.7925	33.7671	5.0069	5.3328	0.5012	3.1004	25.3050
<i>Scotoecus albofuscus</i>	7	0.9923	0.1066	57.7109	0.0204	0.0002	10.9834	0.0000	0.0000	0.0201	0.0000	0.0000	3.1984	3.7786	24.2881
<i>Scotoecus hindei</i>	23	0.9410	0.7052	5.1046	0.0500	0.0000	0.5918	0.0000	6.1618	20.2896	1.3124	0.0000	0.0573	0.7360	65.6965
<i>Scotophilus alvenslebeni</i>	6	0.9800	0.4529	44.3817	4.0470	0.0000	1.5088	0.0000	0.0000	0.0000	33.4946	0.0000	0.2059	0.0000	16.3620
<i>Scotophilus dinganii</i>	253	0.8667	0.8316	0.2909	4.9376	0.0464	2.9012	0.4727	20.1849	21.5715	7.1407	0.0143	0.1937	1.1306	41.1153
<i>Scotophilus leucogaster</i>	43	0.9444	0.8469	1.5794	13.5365	2.9127	0.1303	11.6024	7.9976	7.3935	12.5666	0.3096	1.8729	2.8650	37.2334
<i>Scotophilus viridis</i>	70	0.9432	0.8794	30.8308	2.5353	1.6851	9.8745	1.7311	4.6900	9.5083	0.0000	0.1188	0.0621	0.4414	38.5227
<i>Tadarida aegyptiaca</i>	175	0.8723	0.8092	0.8204	18.2536	2.5798	0.1574	2.9773	33.1609	3.8100	15.9902	0.1118	0.0958	0.0166	22.0263
<i>Tadarida fulminans</i>	19	0.9755	0.8211	0.1417	18.9670	0.0000	0.3375	0.0000	2.3951	2.1672	0.0000	16.3531	0.0000	16.0606	43.5778
<i>Tadarida ventralis</i>	9	0.9521	0.7656	0.0000	25.7713	0.0000	1.9103	0.0000	0.0000	6.2509	14.5104	4.7180	0.0000	0.0807	46.7584
<i>Taphozous mauritanus</i>	112	0.8415	0.7121	0.0234	1.5968	3.2110	0.7562	0.2448	0.8680	10.5668	0.0537	0.0000	4.6840	1.9270	76.0682
<i>Taphozous perforatus</i>	12	0.9498	0.4923	0.2108	1.9345	0.0000	0.0000	0.0000	4.8265	6.0352	9.5845	0.8747	1.7831	0.0000	74.7508
<i>Triaenops afer</i>	14	0.9736	0.8391	36.5579	5.2068	0.1397	0.0000	2.6036	0.0864	8.4952	0.0000	5.7682	0.3530	2.8360	37.9532
<i>Vansonia rueppellii</i>	42	0.8869	0.6934	0.0000	1.1738	0.0005	0.0000	2.7298	0.3157	0.2550	3.8797	13.7422	0.0035	4.5679	73.3319

Table S2 – Bat species predicted (n = 47) and recorded to date (n = 32) to occur in Eswatini based on species distribution models and museum specimens, respectively

Family	Genus	Species	Predicted	Recorded
Pteropodidae	<i>Eidolon</i>	<i>helvum</i>	1	
Pteropodidae	<i>Epomophorus</i>	<i>crypturus</i>	1	1
Pteropodidae	<i>Epomophorus</i>	<i>wahlbergi</i>	1	1
Pteropodidae	<i>Rousettus</i>	<i>aegyptiacus</i>	1	
Hipposideridae	<i>Hipposideros</i>	<i>caffer</i>	1	1
Rhinonycteridae	<i>Cloeotis</i>	<i>percivali</i>	1	1
Rhinolophidae	<i>Rhinolophus</i>	<i>blasii</i>	1	1
Rhinolophidae	<i>Rhinolophus</i>	<i>clivosus</i>	1	1
Rhinolophidae	<i>Rhinolophus</i>	<i>cohenae</i>	1	
Rhinolophidae	<i>Rhinolophus</i>	<i>darlingi</i>	1	1
Rhinolophidae	<i>Rhinolophus</i>	<i>rhodesiae</i>	1	1
Rhinolophidae	<i>Rhinolophus</i>	<i>simulator</i>	1	1
Rhinolophidae	<i>Rhinolophus</i>	<i>smithersi</i>	1	
Emballonuridae	<i>Taphozous</i>	<i>mauritanus</i>	1	1
Emballonuridae	<i>Taphozous</i>	<i>perforatus</i>	1	
Nycteridae	<i>Nycteris</i>	<i>macrotis</i>	1	
Nycteridae	<i>Nycteris</i>	<i>thebaica</i>	1	1
Molossidae	<i>Chaerephon</i>	<i>ansorgei</i>	1	
Molossidae	<i>Chaerephon</i>	<i>pumilus</i>	1	1
Molossidae	<i>Mops</i>	<i>condylurus</i>	1	1
Molossidae	<i>Mops</i>	<i>midas</i>	1	1
Molossidae	<i>Otomops</i>	<i>martiensseni</i>	1	
Molossidae	<i>Tadarida</i>	<i>aegyptiaca</i>	1	1
Miniopteridae	<i>Miniopterus</i>	<i>fraterculus</i>	1	1
Miniopteridae	<i>Miniopterus</i>	<i>inflatus</i>	1	



Miniopteridae	<i>Miniopterus</i>	<i>natalensis</i>	1	1
Miniopteridae	<i>Miniopterus</i>	<i>mossambicus</i>	1	
Vespertilionidae	<i>Afronycteris</i>	<i>nana</i>	1	1
Vespertilionidae	<i>Eptesicus</i>	<i>hottentotus</i>	1	1
Vespertilionidae	<i>Laephotis</i>	<i>botswanae</i>	1	
Vespertilionidae	<i>Laephotis</i>	<i>capensis</i>	1	1
Vespertilionidae	<i>Kerivoula</i>	<i>argentata</i>	1	1
Vespertilionidae	<i>Kerivoula</i>	<i>lanosa</i>	1	1
Vespertilionidae	<i>Myotis</i>	<i>bocagii</i>	1	1
Vespertilionidae	<i>Myotis</i>	<i>tricolor</i>	1	1
Vespertilionidae	<i>Myotis</i>	<i>welwitschii</i>	1	1
Vespertilionidae	<i>Neoromicia</i>	<i>anchietae</i>	1	1
Vespertilionidae	<i>Neoromicia</i>	<i>zuluensis</i>	1	1
Vespertilionidae	<i>Nycticeinops</i>	<i>grandidieri</i>	1	
Vespertilionidae	<i>Nycticeinops</i>	<i>schlieffeni</i>	1	1
Vespertilionidae	<i>Pipistrellus</i>	<i>hesperidus</i>	1	1
Vespertilionidae	<i>Pipistrellus</i>	<i>rusticus</i>	1	1
Vespertilionidae	<i>Pseudoromicia</i>	<i>rendalli</i>	1	
Vespertilionidae	<i>Scotoecus</i>	<i>albofuscus</i>	1	
Vespertilionidae	<i>Scotoecus</i>	<i>hindei</i>	1	
Vespertilionidae	<i>Scotophilus</i>	<i>dinganii</i>	1	1
Vespertilionidae	<i>Scotophilus</i>	<i>viridis</i>	1	1

Table S3 – The five new species recorded in Eswatini since the publication of last bat checklist for the country by Shapiro and Monadjem (2016), together with additional information on the collecting localities and number of individuals captured

Family, Species	Locality	Latitude	Longitude	Elevation (m)	Number of individuals captured
Rhinolophidae, <i>Rhinolophus</i> <i>rhodesiae</i>	Bulembu	-25.95977	31.11844	1210	2
Vespertilionidae, <i>Eptesicus</i> <i>hottentotus</i>	Ntfungula	-26.75476	31.16174	1200	1
Vespertilionidae, <i>Kerivoula</i> <i>argentata</i>	Mambane	-26.83912	32.13247	60	1
Vespertilionidae, <i>Myotis</i> <i>welwitschii</i>	Dombeya Game Reserve	-26.36068	31.5447	440	1
Vespertilionidae, <i>Pipistrellus</i> <i>rusticus</i>	Mvembili Sigcineni Velezizweni	-25.75719 -26.69995 -26.70120	31.41769 31.28313 31.11966	470 430 910	10 3 1