

Figure S1. Physical map of a chromosome 10 region in maize line B73 corresponding to QTL10 from CML444. The B73 physical map (v4) positions of four SSR markers used for marker assisted backcross breeding to introgress the region from CML444 into B73 to produce B7-QTL are shown. The genetic map size of the QTL10 region in cM is indicated (Berger et al. 2014).



Figure S2a. Primer positions relative to the maize B73 L-RLK gDNA exon4-exon7 region. The primers L-RLK-qFor e4 and L-RLK-qRev e7 were used in RT-PCR to determine splice variants. The primers L-RLK e7 F1 and L-RLK e7 R1 were used for expression analysis by RT-qPCR. L-RLK e4e5 F1 (cml) and L-RLK e5 R1 (cml) were used for RT-qPCR expression analysis of the CML444 copy of the L-RLK gene. L-RLK e4e5 F2 (b73) and L-RLK e5 R2 (b73) were used for RT-qPCR expression analysis of the B73 copy of the L-RLK gene. Sequences are shown in Supplementary Figure S2b. The gDNA sequence shown (1427 bp) is identical in both versions of the B73 genome (Zm00001d026382 [v4]; Zm00001e041728 [v5]).

				1 Exon 4 L-RLK-qFor e4 primer 5' TGCCGAAGAGTTCAGGAATG 3' 99
CML I	L-RLK	cDNA	(1)	GAATGTTGGAAGGAACCAAAGAAGTAGCCATCAAGAGACTGAGCAAGAATTCTGGGCAAGGTGCCGAAGAGTTCAGGAATGAAGTAGTTTTGATTGCAA
в73 І	L-RLK	т2	(1)	GAATGTTGGAAGGAACCAAAGAAGTAGCCATCAAGAGACTGAGCAAGAGTTCTGGGCAAGGTGCCGAAGAGTTCAGGAATGAAGTA <mark>ATC</mark> TTGATTGCAA
в73 І	L-RLK	gDNA	(1)	GAATGTTGGAAGGAACCAAAGAAGTAGCCATCAAGAGACTGAGCAAGAGTTCTGGGCAAGGTGCCGAAGAGTTCAGGAATGAAGTA <mark>ATC</mark> TTGATTGCAA
				100 198
CML I	L-RLK	cDNA	(100)	${\tt AGTTGCAGCACAAGAACCTAGTTAAGCTTCTTGGTTTCTGTGTTCATGAAGATGAGAAGATGTTAGTTTATGAGTACTTGCCGAACAAAAGCTTAGATTAGATTAGAGTACTTAGGTTCATGAGATGAGAAGATGTTAGGTTTATGAGTACTTGCCGAACAAAAGCTTAGATTAGAGTAGTTAGATTAGAGTACTTGCCGAACAAAAGCTTAGATTAGAGATGAGAAGATGTTAGTTTATGAGTACTTGCCGAACAAAAGCTTAGATTAGAGTAGTTAGAGTAGTTAGAGTAGTTGAGATGAGAAGA$
B73 I	L-RLK	Т2	(100)	AGTTGCAGCACAAGAACCTAGTTAAGCTTCTTGGTT <mark>G</mark> CTGTGTTCATGAAGATGAGAAG <mark>C</mark> TGTTAGTTTATGAGTACTTGCCGAACAAAAGCTTAGATT
B73 I	L-RLK	gDNA	(100)	AGTTGCAGCACAAGAACCTAGTTAAGCTTCTTGGTT <mark>G</mark> CTGTGTTCATGAAGATGAGAAG <mark>C</mark> TGTTAGTTTATGAGTACTTGCCGAACAAAAGCTTAGATT
				199 297
				5' <mark>CTTCCTCTTTG</mark> ACTCTTC 3' L-RLK e4e5 F1 (cml) primer
CML I	L-RLK	cDNA	(199)	A <mark>CTTCCTCTTTG</mark>
				'CTTCCTCTTTGATTCTGC 3' L-RLK e4e5 F2 (b73) primer
B73 I	L-RLK	Т2	(199)	ACTTCCTCTTTG
B73 I	L-RLK	gDNA	(199)	ACTTCCTCTTTG <mark>GTACATTGAGTTTTAAAAACTTTCCATTTAGTTAACAAAATCAACAAATATACATGCATATGCCTTCTGATATGTATG</mark>
				298 Exon 5 396
		-	(011)	5'CTTCCTCTTTGACTCTTC 3' L-RLK e4e5 Fi (cml) primer
CML I	-RTK	CDNA	(211)	
				5/ CHARGE MARCH CARGE 2/ T. DIV AASE D2 (1772) Primer
ד כדם	שום	шЭ	(211)	
ן כום ד כדם	-KLK		(211)	
в/з I	-ктк	GUNA	(∠98)	TTTTATTCATATTAGA <mark>H</mark> TUT <mark>G</mark> CAAGAAAGTCAACGCTTCAGTGGCCCAACAGGTTCAAGATAATCCACGGGGTAGCTAGAGGAATTATGTATCTTCATC

49 CML L-RLK cDNA (295) AGGACTCGAGATTAACAATAATCCATAGAGATCTCAAAGCAAGC
B73 L-RLK T2 (295) AAGACTCGAGATTAACAATAATCCATAG <mark>C</mark> GATCTCAAAGCAAGCAAGCAACATCTTGTTAGATAAGGACATGAGCCCAAAAATATCAGATTTTGGCATGGCT 3' GTTCTGAGCTCTAATTG 5' L-RLK e4e5 R2 (b73) primer B73 L-RLK qDNA (397) AAGACTCGAGATTAACAATAATCCATAG <mark>C</mark> GATCTCAAAGCAAGCAAGCAACATCTTGTTAGATAAGGACATGAGCCCAAAAATATCAGATTTTGGCATGGC
496 CML L-RLK cDNA (394) GAATATTCAGTAGCGACCAGCTCCATGCAAATACTAACCGAGTTGTAGGGACATAIntron 5 B73 L-RLK T2 (394) GAATATTCAGTAGCGACCAGCTCCATGCAAATACTAACCGAGTTGTAGGGACATAIntron 5 B73 L-RLK gDNA (496) GAATATTCAGTAGCGACCAGCTCCATGCAAATACTAACCGAGTTGTAGGCACATAIntron 5
595Exon 669CML L-RLK cDNA(449)
79 CML L-RLK cDNA (491) AGTCAAGTCTGACACTTACAGCTTTGGTGTTTTAATGCTGGAGATTGTAAGTGGTTTGAAGATCAGCTCACCACATCTTCACATGGATTTTCCTAATC B73 L-RLK T2 (491) AGTCAAGTCTGACACTTACAGCTTTGGTGTTTTAATGCTGGAGATTGTAAGTGGTTTGAAGATCAGCTCACCACATCTTCACATGGATTTTCCTAATC B73 L-RLK gDNA (694) AGTCAAGTCTGACACTTACAGCTTTGGTGTTTTAATGCTGGAGATTGTAAGTGGTTTGAAGATCAGCTCACCACATCTTCACATGGATTTTCCTAATC
Exon 793 CML L-RLK CDNA (590) TAGAGCATATGCAT B73 L-RLK T2 (590) TAGAGCATATGIntron 6CAT B73 L-RLK gDNA (793) TAGAGCATATGTAAGGACTGGCACACTCTCATAATAATTCAGAACTGTCCACATGATAAAAACGATCCTGATATGAGATTGTGTCATGTTTCAGGCAT
99 CML L-RLK cDNA (605) GAACATGTGGAAAGAAGAAGAAGAAGAAGAATTGGTGGACTCATCAGTCATGGAGAATTGCTCCCTTGATGAAGTTTCACAATGTGTCCATATAGGAC B73 L-RLK 72 (605) GAACATGTGGAAAGAAGGCAAAATAGAAGATTTGGTGGACTCATCAGTCATGGAGAATTGCTCCCTTGATGAAGTTTCACAATGTGTACATATAGGAC B73 L-RLK gDNA (892) GAACATGTGGAAAGAAGGCAAAATAGAAGATTTGGTGGACTCATCAGTCATGGAGAATTGCTCCCTTGATGAAGTTTCACAATGTGTACATATAGGAC L-RLK-qFor e4 primer 3' CTAAACCACCTGAGTAGTCAGT 5'
991 L-RLK e7 F1 5 AACGACGACACTTCCAACTC 3 108 CML L-RLK cDNA (704) CTTGTGTGTTCAAGACAGTCCGAGCTTCAGGCCACTCATGTCGGCGGTTGTGTCCATGCTGGAGAACAAAACGACGACAACTTCCAACTCCAAGTCAAG B73 L-RLK T2 (704) CTTGTGTGTTCAAGACAGTCCGAGCTTCAGGCCACTCATGTCGGCGGTTGTGTCCATGCTGGAGAACAAAACGACGACAACTTCCAACTCCAAGTCAAG B73 L-RLK gDNA (991) CTTGTGTGTTCAAGACAGTCCGAGCTTCAGGCCACTCATGTCGGCGGTTGTGTCCATGCTGGAGAACAAAACGACGACAACTCCAACTCCAAGTCAAG
1090 118 CML L-RLK cDNA (803) TGTATATTTTGCAGTTAGGGACCCCTACCAACCTGGAAAGGCAGTTGGCAACAAGGAGTTGTCTATATAGAGTCTCACTGTGCCAGAAGGTC B73 L-RLK T2 (803) TGTATATTTTGCAGTTAGGGACCCCTACCAACCTGGAAAGGCAGTTGGCAACAAGGAGTTGTCTATATATGATATGAGTCTCACTGTGCCAGAAGGTC B73 L-RLK gDNA (1090) TGTATATTTTGCAGTTAGGGACCCCTACCAACCTGGAAAGGCAGTTGGCAACAAGGAGTTGTCTATATATGATATGAGTCTCACTGTGCCAGAAGGTC L-RLK gDNA (1090) TGTATATTTTGCAGTTAGGGACCCCTACCAACCTGGAAAGGCAGTTGGCAACAAGGAGTTGTCTATATATGATATGAGTCTCACTGTGCCAGAAGGTC L-RLK e7 R1 3 CTATACTCAGAGTGACACGGTC 5'

CML L-RLK CDNA B73 L-RLK T2 B73 L-RLK gDNA	(902) (902) (1189)	1287 TRAAATGCTTTATTTTTATAACTGATTGGGAGGATTAATTAA
CML L-RLK CDNA B73 L-RLK T2 B73 L-RLK gDNA	(1001) (1001) (1288)	288 TATGTGAATGGTCGGTAATTTGCATTTTGCAACAGTTTGGTAAGCCTGCTTATATAATTGTATAAAGACAGGTGATTTCCTTCTCTCAGCTCATGTGGT TATGTGAATGGTCGGTAATTTGCATTTTGCAACAGTTTGGTAAGCCTGCTTATATAATTGTATAAAGACAGGTGATTTCCTTCTCTCAGCTCATGTGGT TATGTGAATGGTCGGTAATTTGCATTTTGCAACAGTTTGGTAAGCCTGCTTATATAATTGTATAAAGACAGGTGATTTCCTTCTCTCAGCTCATGTGGT
CML L-RLK CDNA B73 L-RLK T2 B73 L-RLK gDNA	(1100) (1089) (1387)	1387 1427 TGTACTGTTTATGTGCTTATTATGTGGATCCAATTTGACCG TGTACTGTTTATGTGCCTTATTATGTGGATCCAATTTGACCG

Figure S2b. Nucleotide alignments to illustrate maize L-RLK gene-specific primer design. The maize B73 L-RLK gDNA exon 4–exon 7 region (B73 L-RLK gDNA) is aligned with corresponding regions of the fully spliced B73 L-RLK T2 transcript (XM_008663415, Zm00001e041728_T002) and the CML L-RLK cDNA (de novo assembled transcript in this study; MT108451). Introns are labeled in yellow. Nucleotides in B73 that are different from CML444 are marked blue. SNPs between B73 and CML at the start of exon 5 were used to design gene copy-specific exon-exon primers with differences in their 3' ends [L-RLK e4e5 F1 (cml) and L-RLK e4e5 F2 (b73)]. There was also one nucleotide difference in the 3' end of the reverse primers [L-RLK e5 R1 (cml) and L-RLK e5 R2 (b73)]. The primers specific to the CML copy are marked in green, and the primers specific to the B73 copy are marked in purple. The positions of the RT-PCR primers (L-RLK e7 R1) are also shown. The stop codon is marked in red.

		1 100
CML444 L-RLK-CML (817aa)	(1)	MMYVPIFILLVLLSSLCQSDDQLTHTKPL <mark>SPKD</mark> TLISAGGDFALGFFSPTSSSNKLYIGIWYNNVPERERTVVWIANRDSPVTTPATSVKLAVTNDSELV
B73 v5 L-RLK P2 (818aa)	(1)	$\verb MMYVPIFILLVVLSSLCOSDDQLTHTKPLFPKETLISAGGDFALGFFSPTSSSNKLYIGIWYNNVPERERTVVWIANRDSPVTTPATSVKLAVTNDSELV$
B73 v5 L-RLK P1 (666aa)	(1)	$\verb MMYVPIFILLVVLSSLCOSDDOLTHTKPLFPKETLISAGGDFALGFFSPTSSSNKLYIGIWYNNVPERERTVVWIANRDSPVTTPATSVKLAVTNDSELV$
B73 v4 L-RLK P1 (698aa)	(1)	MMYVPIFILLWVLSSLCOSDOOLTHTKPLFPKETLISAGGDFALGFFSPTSSSNKLYIGIWYNNVPERETVWWIANRDSPVTTPATSVKLAVTNDSELV
	(-)	101 200
CMI444 L-RIK-CMI (817aa)	(101)	LSDSOGNTFWMATSSTSGGAGAFAVILNSGNFVLRSPNGTDVWOSFDHPTDTLLPTMRFGTSYRSOPATRLFAWKGPDDPSTGNTSFSLDPASSNLOMFT
B73 v5 L-RLK P2 (818aa)	(101)	LSDSOGHTEWMATSSTSGGAGAFAVLLNSGNFVLRSPNGTDVWOSFDHPTDTVLPTMRSMTRYRSOPATGLFAWKSPDDPSTGDTSSGMDPASSNVOVFT
B73 v5 L-RLK P1 (666aa)	(101)	LSDSOGHTEWMATSSTSGGAGAFAVLLNSGNFVLRSPNGTDVWOSFDHPTDTVLPTMRSMTRYRSOPATGLFAWKSPDDPSTGDTSSGMDPASSNVOVFT
B73 v4 L-BLK P1 (698aa)	(101)	LSDSOCHTEWMATSSTSCCACAFAVLINSCNFVLRSPNGTDVWOSEDHPTDTVLPTMRSMTRYRSOPATCLEAWKSPDDPSTCDTSSCMDPASSNVOVET
	(101)	
CMI.444 L-RIK-CMI. (817aa)	(201)	WIGTLEY FRSSVVNDVSVSGGTYOANNTSVTYOGTLUDTGDE I YYTETVLAGS PYMHESLHYTGKMR I LAWENSTSSWVVT FEGESVGCDLYASCOPOO
B73 v5 L-RLK P2 (818aa)	(201)	WNGTLPYYRSSFFNDVPASGGTYQANNTSVTYQWMWVDTGDEIYYTFTVLAGSPYMHFSLHHTGKMRILIWENSTSSWUTSFFPSVGCDLYASCGPFGY
B73 v5 L-RLK P1 (666aa)	(201)	WNGTL PYYR SSFENDVPA SGGTYOANNTSVTYOWMVDTGDE I YYTFTVL AGSPYMH FSLHHTGKMRLI JWENSTSSWULTSE PSVGCDL YASCGPEGY
B73 v4 L-RLK P1 (698aa)	(201)	WNGTLPYYRSSFFNDVPASGGTYQANNTSVTYOWMWDTGDEIYYTFTVLAGSPYMHFSLHHTGKMRLLVWENSTSSWVLTSEEPSVGCDLYASCGPFGY
	(201)	301
CMT.444 IRI.K-CMI. (817aa)	(301)	CDLTTAMPTCOCPDHEELVDSLNESEGCORKEELKCGTDSYEMTMESMKTPDKELHTENRTEDOCAAECARNCTCTAYAYANLSTAGATGDTSRCLVMIO
B73 v5 L-BLK P2 (818aa)	(301)	CDLTKIAMPTCOCPDHEELVDSLNFSRCORKEELKCGTESYEMTMPNMKTPDKELHTRNRTFDCCAAFCARNCTCMAYTYATLSTASTTGDASRCLVMTO
B73 v5 L-BLK P1 (666aa)	(301)	CDITKIAMPTCOCPDHEELVDSLNFSRCORKEELKCGTESYEMTMPNMKTPDKELHTRNRTFDCCAAFCARNCTCMAYTYATLSTASTGDASRCLVMTO
B73 v4 L-BLK P1 (698aa)	(301)	CDITKIAMPTCOCPDHEELVDSLNFSRCORKEELKCGTESYEMTMPNMKTPDKELHTRNRTFDCCAAFCARNCTCMAYTYATLSTASTGDASRCLVMTO
	(001)	401
CML444 L-RLK-CML (817aa)	(401)	DLIDMEKASHLLENLYIRLGESPDQKS-NFQKILLSTIACLLLLTIGALGWACKIRGKWHKKKIQTRRMLEYLSSTDEAGGKNIEFSFIAFENIVTATDN
B73 v5 L-RLK P2 (818aa)	(401)	DLIDMEKASVFLENLYIRLGESPDQKKRNFQKILLSAIACLLLLTIGALGWACKIRGKWHKKKIQTRMMLEYLSSTDEAGGKNIEFPFITFENIVTATDN
B73 v5 L-RLK P1 (666aa)	(401)	DLIDMEKASVFLENLYIRLGESPDQKKRNFQKILLSAIACLLLLTIGALGWACKIRGKWHKKKIQTRMMLEYLSSTDEAGGKNIEFPFITFENIVTATDN
B73 v4 L-RLK P1 (698aa)	(401)	DLIDMEKASVFLENLYIRLGESPDQKKRNFQKILLSAIACLLLLTIGALGWACKIRGKWHKKKIQTRMMLEYLSSTDEAGGKNIEFPFITFENIVTATDNCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC
		501 600
CML444 L-RLK-CML (817aa)	(500)	FSDCNILGKGGFGKVYKGMLEGTKEVAIKRLSKNSGQGAEEFRNEVVLIAKLQHKNLVKLLGFCVHEDEKMLVYEYLPNKSLDYFLFDSSRKSTLQWPTR
B73 v5 L-RLK P2 (818aa)	(501)	FSDCNILGKGGFGKVYKGMLEGTKEVAIKRLSKSSGQGAEEFRNEVILIAKLQHKNLVKLLGCCVHEDEKLLVYEYLPNKSLDYFLFDSARKSTLQWPTR
B73 v5 L-RLK P1 (666aa)	(501)	FSDCNILGKGGFGKVYKGMLEGTKEVAIKRLSKSSGQGAEEFRNEVILIAKLQHKNLVKLLGCCVHEDEKLLVYEYLPNKSLDYFLFDSARKSTLQWPTR
B73 v4 L-RLK P1 (698aa)	(501)	FSDCNILGKGGFGKVYKGMLEGTKEVAIKRLSKSSGQGAEEFRNEVILIAKLQHKNLVKLLGCCVHEDEKLLVYEYLPNKSLDYFLFGTLS
		601 Kinase active site 700
CML444 L-RLK-CML (817aa)	(600)	FK II H GVARGIMYLHQDSRLT <mark>IIHRDLKASNILL</mark> DKDMSPKISDFGMARIFSSDQLHANTNRVVGTYGYMSPEYAMEGAFSVKSDTYSFGVLMLEIVSGL
B73 v5 L-RLK P2 (818aa)	(601)	FK II H GVARGIMYLHQDSRLT <mark>IIHRDLKASNILL</mark> DKDMSPKISDFGMARIFSSDQLHANTNRVVGTYGYMSPEYAMEGAFSVKSDTYSFGVLMLEIVSGL
B73 v5 L-RLK P1 (666aa)	(601)	FKIIHGVARGIMYLHQDSRLT <mark>IIHRDLKASNILL</mark> DKDMSPKISDFGMARIFSSDQLHANTNRVVGT*
B73 v4 L-RLK P1 (698aa)	(592)	FKTFH
	. ,	701 800
CML444 L-RLK-CML (817aa)	(700)	KISSPHLHMDFPNLRAYAWNMWKEGKIEDLVDSSVMENCSLDEVSOCVHIGLLCVODSPSFRPLMSAVVSMLENKTTTLPTPSOPVYFAVRDPYOPGKAV
B73 v5 L-RLK P2 (818aa)	(701)	KISSPHLHMDFPNLRAYAWNMWKEGKIEDLVDSSVMENCSLDEVSQCVHIGLLCVQDSPSFRPLMSAVVSMLENKTTTLPTPSQPVYFAVRDPYQPGKAV
B73 v5 L-RLK P1 (666aa)	(667)	
B73 v4 L-RLK P1 (698aa)	(597)	LAWNMWKEGKIEDLVDSSVMENCSLDEVSQCVHIGLLCVQDSPSFRPLMSAVVSMLENKTTTLPTPSQPVYFAVRDPYQPGKAV
		801 818
CML444 L-RLK-CML (817aa)	(800)	GNKELSIYDMSLTVPEGR*
B73 v5 L-RLK P2 (818aa)	(801)	GNKELSIYDMSLTVPEGR*
B73 v5 L-RLK P1 (666aa)	(667)	
B73 v4 L-RLK P1 (698aa)	(681)	GNKELSIYDMSLTVPEGR*
	,	

Figure S3a. Amino acid alignment of predicted G-lectin S-receptor-like kinase proteins from maize B73 [B73 v5 L-RLK P2 (818aa) annotated in both the v4 genome (XP_008661637) and v5 genome (Zm00001e041728_P001), B73 v5 L-RLK P1 (666aa) (Zm00001e041728_P001), B73 v4 L-RLK P1 (698aa)(AQK46433)], and the protein predicted from the de novo assembled transcript (MT108451) from B73-QTL (CML444 L-RLK-CML; QIH29483).



S3b. Figure structures predicted G-lectin S-receptor-like kinases from B73 Domain of maize genome annotations: B73 v4 L-RLK P1 (AQK46433), B73 v5 L-RLK P1 (Zm00001e041728_P001), B73 v5 L-RLK P2 (Zm00001e041728_P002, XP_008661637), and the de novo assembled transcript (MT108451) from B73-QTL (L-RLK-CML; QIH29483). Domains: SP = Signal peptide (purple); G-lectin (brown); S-locus glycoprotein (blue), PAN-like domain (green), trans-membrane (TM), kinase (red); kinase active site (dark purple). Drawn to scale. AQK464433 was removed from the v5 annotation of the B73 genome.



Figure S3c. Predicted transcripts of the G-lectin S-receptor-like kinase from the v4 and v5 annotations of the B73 genome. The 3582 genomic DNA fragment is identical between the v4 gene model (Zm00001d026382) and the v5 gene model (Zm00001e041728). Zm00001d026382_T001 encoding AQK46433 was removed from the v5 annotation. Zm00001e041728_T001 retains intron 5 resulting in a stop codon directly after exon 5 resulting in a truncated protein of 666 AA. Zm00001e041728_T002 (XM_008663415 in v4) encodes the full-length 818 AA L-RLK with all the functional domains.



Figure S3d. B73-QTL RNAseq read coverage mapped against the B73 genomic DNA sequence on chromosome 10 corresponding to the lectin receptor kinase Zm00001d026382.



Figure S3e. CML444 RNAseq read coverage mapped against the 2811 nt *l-rlk-cml* transcript assembled from the B73-QTL reads (MT108451). RNA was extracted from a pool of leaves from six CML444 plants at 103 days after planting at Baynesfield, South Africa, and challenged with *C.zeina*. This was the same field experiment reported in Christie et al (2017) The Plant Journal 89 (4):746-763.



Figure S4a. BamHI and HphI restriction map of the lectin receptor kinase gene from B73 and CML444 used for the PCR-RFLP. Panel A: A 430 bp fragment of Exon 1 from B73 gene Zm00001d026382 gDNA between Lectin primers F and R, showing the position of the single HphI site. Panel B: The corresponding 430 bp fragment predicted from the de novo assembled transcript encoding L-RLK-CML from B73-QTL and CML444 (MT108451), showing BamHI and HphI sites. The products of digestion with either BamHI or HphI are shown with blue dotted lines next to their sizes.



Figure S4b. Sequences of parts of the 430 bp PCR products from Exon 1 from B73 gene Zm00001d026382 and CML444 to show the polymorphisms at the BamHI and HphI restriction enzyme sites, and presence of both alleles in CML444.



Figure S5. Reverse transcriptase-qPCR of lectin receptor-like kinase gene expression in B73 and B73-QTL maize plants inoculated with *C. zeina*. Maize leaves were either inoculated with *C. zeina* CMW25467 or mock-inoculated, and, at 43 days post inoculation, samples were collected for RNA extraction and RT-qPCR analysis. **Panel A**: Expression of the lectin receptor-like kinase with primers in exon 7 (L-RLK e7 F1 & R1 primers) designed to amplify both the B73 and CML444 copy of the gene. **Panel B**: Expression of the lectin receptor-like kinase with the L-RLK e4e5 F1 (cml) and L-RLK e5 R1 (cml) primer pair, designed to amplify the CML444 copy of the gene. **Panel C**: Expression of the lectin receptor-like kinase with the L-RLK e4e5 F2 (b73) and L-RLK e5 R2 (b73) primer pair, designed to amplify the B73 copy of the gene. Average expression of the L-RLK transcripts are expressed relative to the RPOL reference gene in all panels. Maize genotype treatments are labeled on the x-axis. Expression values that were not significantly different from one another for each primer pair, based on a one-way ANOVA analysis with the Tukey-Kramer post hoc test (p < 0.05), are labeled with the same letter above each column. The 95% confidence intervals are shown by error bars for each column. There were no detectable transcripts for the B73 samples in Panel B. The maximum 95% confidence interval for B73-QTL in Panel C was 15.8. Panel C and the same letter above each column.



Figure S6. Synteny plot on region of maize chromosome 10 to compare gene organization around the B73 Zm00001d026382 (lectin receptor like kinase) gene in maize, teosinte and sorghum. Percentage amino acid identity to the predicted 817 AA L-RLK-CML protein from CML444 (QIH29483) is shown in red boxes.



Figure S7. Phylogram of G-Lectin-RLKs from diverse monocots and *Arabidpsis thaliana*. Protein BLAST was performed within the PLAZA database using the Zm00001d026382 amino acid sequence (XP_008661637, B73 v5 L-RLK P2) as the query.

The CML444 L-RLK-CML (QIH29483) and the best Zm00001d026382 BLAST hits (max. 3 from each species) were aligned using MAFFT. A phylogenetic tree was constructed using the Maximum-Likelihood method implemented in FastTree. Bootstrapping values (1000 replicates) are shown at the nodes.