

## Review Article

# African Herbal Remedies with Antioxidant Activity: A Potential Resource Base for Wound Treatment

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The use of traditional herbal remedies as alternative medicine plays an important role in Africa since it forms part of primary health care for treatment of various medical conditions, including wounds. Although physiological levels of free radicals are essential to the healing process, they are known to partly contribute to wound chronicity when in excess. Consequently, antioxidant therapy has been shown to facilitate healing of such wounds. Also, a growing body of evidence suggests that, at least, part of the therapeutic value of herbals may be explained by their antioxidant activity. This paper reviews African herbal remedies with antioxidant activity with the aim of indicating potential resources for wound treatment. Firstly, herbals with identified antioxidant compounds and, secondly, herbals with proven antioxidant activity, but where the compound(s) responsible for the activity has not yet been identified, are listed. In the latter case it has been attempted to ascribe the activity to a compound known to be present in the plant family and/or species, where related activity has previously been documented for another genus of the species. Also, the tests employed to assess antioxidant activity and the potential caveats thereof during assessment are briefly commented on.

## 1. Introduction

Human cells are continuously exposed to exogenous oxidants as well as to those produced endogenously during normal physiological processes. Antioxidants form part of protective mechanisms that exist in human cells to scavenge and neutralize these oxidants. Oxidants such as the reactive oxygen species (ROS) and reactive nitrogen species (RNS) are involved in several diseases [1, 2]. Antioxidant defenses are defective in these diseases and therefore it is possible to limit oxidative damage and ameliorate disease progression with antioxidant supplementation [3].

With reference to wounds, antioxidants play pivotal roles that consequently restore normalcy to injured skin. Basal levels of ROS and other free radicals are essential in almost all phases of the wound healing process (Figure 1) [4]. During haemostasis, ROS regulates the constriction of blood vessels to limit loss of blood. Furthermore, ROS facilitates the migration of neutrophils and monocytes from surrounding

blood vessels towards the injury site. The presence of ROS and other free radicals in the wound vicinity during the inflammatory phase of the healing process is also required for infection control and general maintenance of sterility. Finally, ROS promotes the proliferation of keratinocytes, endothelial cells, and fibroblasts, thereby enhancing angiogenesis and collagen deposition. However, uncontrolled release of ROS could cause oxidative stress, resulting in cellular and tissue damage, thereby causing delayed healing [1].

To keep ROS within physiological levels, antioxidants serve as electron donors, thereby preventing them from capturing electrons from other molecules which ultimately leads to their destruction [4]. Both nonenzymatic antioxidants such as glutathione, ascorbic acid, and  $\alpha$ -tocopherol, as well as enzymatic antioxidants like catalase and peroxiredoxin, have shown potential to normalize high ROS levels and thus stimulate healing [4]. By normalizing ROS, antioxidants can enhance their physiological roles and thereby accelerate the wound healing process. Naturally occurring antioxidants are

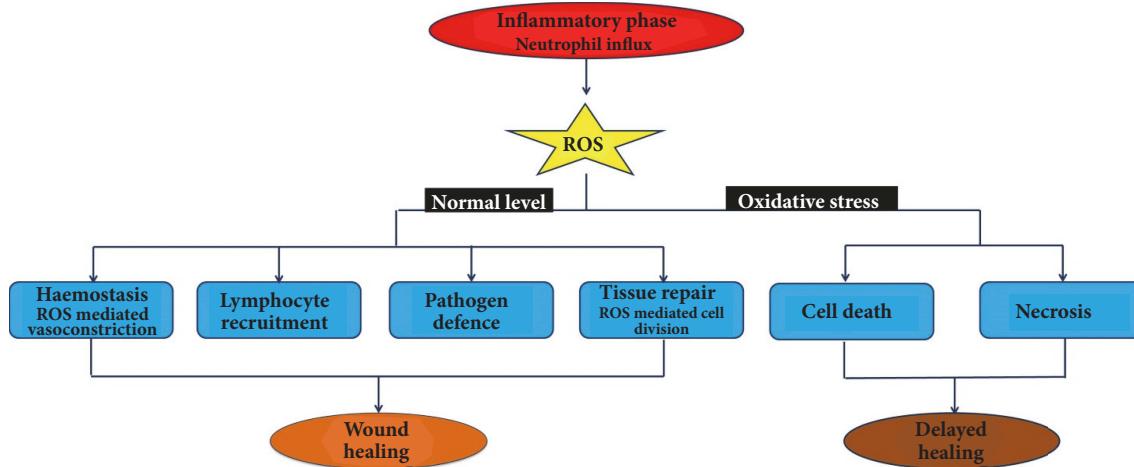


FIGURE 1: Role of reactive oxygen species (ROS) in the wound healing process.

generally favoured over their synthetic counterparts, as the latter are suspected to cause or promote negative health effects [5]. This has resulted in the restricted use of synthetic antioxidants in several countries [6].

This review provides a comprehensive list of African medicinal plants and isolated compounds with antioxidant activities, with the aim of highlighting the continent's rich herbal resource base for possible management of wounds and allied conditions. Previous reviews have listed a number of these African medicinal plants with antioxidant properties [7–9]. The present work has therefore aimed to expand the list to include medicinal plant species with antioxidant properties that are used in different African countries including those from Madagascar and Mauritius. For the sake of inclusivity, plants that have been shown to contain compounds that hold the potential of being novel antioxidants are also considered. In addition, those with anti-inflammatory properties were also included due to an earlier observation that the anti-inflammatory activities of the same extracts could be explained, at least in part, by their antioxidant properties [10]. Additional efforts were also made to include information, where available, on their vernacular names, their regional distribution, and medicinal use and plant parts used for these preparations or for the isolation of the antioxidant ingredient(s). Table 1 lists medicinal plants that have been investigated and have confirmed antioxidant and/or anti-inflammatory activity and that contain compounds which are known to have such activities. Table 2 on the other hand lists medicinal plants that have confirmed antioxidant activity but the compounds responsible for their antioxidant property have not yet been identified.

Many edible and culinary herbs and condiments were also included in these two tables as they were used in certain instances as medicinal herbs to treat diseases. These included fruits and seeds of *Balanites aegyptiaca*, leaves of *Boscia senegalensis*, leaves of *Entada africana* and seeds of *Parkia biglobosa*, from Niger [11], also leaves, seeds, and stem-bark of *Mangifera indica* from Benin and Burkina Faso [12, 13], leaves of *Cynara scolymus* from Ethiopia [14,

15], leaves of *Aspalathus linearis* from South Africa [16–21], leaves of *Cinnamomum zeylanicum* from Madagascar and Ethiopia [22–24], essential oils from the bark and leaves of *Ravensara aromatica* from Madagascar [23, 25], buds of *Syzygium aromaticum* from Madagascar [23], seeds of *Trigonella foenumgraecum* from Ethiopia and Morocco [26–28], and oils in seeds of *Nigella sativa* from African countries of the Mediterranean region [29–31].

## 2. Tests Used to Assess Antioxidant Activities of African Medicinal Plant Extracts

A variety of test systems were employed to assess the antioxidant properties of the medicinal plant extracts and compounds listed in Tables 1 and 2. A comprehensive list of the methods used in antioxidant activity determination, as well as their merits and demerits, has already been published [343–346]. The methods used in the determination of antioxidant activity of natural products and isolated compounds result in varied outcomes when the same samples are tested in different laboratories and by other researchers [347]. Furthermore, results of different methods cannot be correlated, as contradictory results are usually obtained. Hence, although several assays are available, none of them is capable of accurately and completely determining the antioxidant activity of a test substance because of the complex nature of the redox-antioxidant system *in vivo* (Figure 2). Based on this complexity, antioxidants are broadly classified as (i) inhibitors of free radical formation, (ii) free radical scavengers, (iii) cellular and tissue damage repairers, and (iv) signalling messengers [347].

The inhibition of free radical formation could protect against oxidative damage by suppressing the formation of active ROS/RNS. This typically involves reduction or inhibition of substrates required for free radical formation such as metal ions like iron (Fe) and copper (Cu). The sequestration of these metal ions by antioxidant compounds like ellagic acid and glutathione is known to suppress formation of

TABLE 1: Medicinal plants with confirmed antioxidant activity, shown to contain compounds that are known to have such activity.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Aloaceae</b>						
<i>Aloe barbadensis</i> Mill.	Burn plant, siber, sbar'essoutkouri /mar, shar	Leaf exudate	Algeria, Morocco, Tunisia	Antioxidant activity. Used as laxative, purgative, diuretic, asthma, baldness, cuts, burns, skin rash.	Flavonoids, two dihydrocoumarin derivatives and two flavone glycosides	[32-34]
<i>Aloe claviflora</i> Burch.	Kraal aloe	Leaf exudate	South Africa	Radical scavenging activity and moderate activity in the lipid peroxidation assay	Chromone glycoside	[35, 36]
<i>A. saponaria</i> (Ait.) Haw.	Mpelu Mnemvu Soap aloe, African aloe	Leaf exudate	South Africa	Radical scavenging activity and moderate activity in the lipid peroxidation assay	Chromone glycoside	[35, 37]
<i>A. thraskii</i> Baker	Dune aloe, ikhala, umhlaba	Leaf exudate	South Africa	Radical scavenging activity and moderate activity in the lipid peroxidation assay	Chromone glycoside	[35, 36]
<b>Amaranthaceae</b>						
<i>Amaranthus caudatus</i> L.	Tassel flower	Seed; Young shoots	Ethiopia	Antioxidant properties	Tocopherols, phenolic acids	[38-40]
<b>Anacardiaceae</b>						
<i>Anacardium occidentale</i> L.	Not signalized	Stern-bark	Nigeria	Anti-inflammatory properties.	Agathisflavone, querctein 3-O-rutinoside, querctein 3-O-rhamnoside	[41, 42]
<i>Lannea edulis</i> Engl.	Wild Grape	Root-bark	Zimbabwe	Semipolar extracts high activity both as radical scavengers and lipoxygenase inhibitors. Lipophilic extracts inhibitor of 15-lipoxygenase.		
<i>Lannea velutina</i> A. Rich	Bemmbeiyi Raisnier velu, Lannéa velouté	Leaves, bark, root	Mali	Used for painful menstruation, urogenital infection, sexually transmitted diseases.	Proanthocyaniidins	[46, 47]
<i>Mangifera indica</i> L.	Mango Mangoro	Leaves, seeds, stem-bark	Benin Burkina Faso	Anti-inflammatory, analgesic, and hypoglycemic effects. Used to treat urogenital infection, tonic, diarrhoea, tooth ache, gingivitis, liver disease, diabetes.	Polyphenolics, flavonoids	[12, 13, 46, 47]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Apiaceae</b>						
<i>Centella asiatica</i> (L.) Urb.	Gotu kola	Leaves	South Africa	Antioxidant and anti-inflammatory activities. Used for wound healing. Protection against radiation-induced injury. Cardio protective effect. Oral treatment increased antioxidant enzymes.	Rutin, Quercetin robinobioside, Kaempferol-3-O-rutinoside, Kaempferol-3-O-robinobioside	[48–55]
<b>Apocynaceae</b>						
<i>Alstonia boonei</i> De Wild.	Awin, Egbi	Stem-bark Root-bark	Nigeria Ghana	Anti-inflammatory activity. Used for its analgesic and anti-inflammatory properties.		[56–59]
<i>Catharanthus roseus</i> (L.) G. Don	Madagascar periwinkle kaka poul, karakté dezom blan, zèb sosoé	Whole plant	Madagascar	Antioxidant activity and ability to increase antioxidant enzymes. Used for conjunctivitis.	Phenols	[60]
<b>Arecaceae</b>						
<i>Elaeis guineensis</i> Jacq.	Ori	Nuts	Ghana Nigeria	Anti-inflammatory activity. Used to treat rheumatoïd arthritis.	3,4 hydroxybenzaldehyde, p-hydroxybenzoic acid, vanillic acid, syringic acid, ferulic acid, carotenoids, $\alpha$ -tocopherol	[12, 61]
<b>Asclepiadaceae</b>						
<i>Secamone afzelii</i> Rhoem.	Ahaban Kroratima	Stem	Central Africa	Antioxidant and anti-inflammatory properties. Used for wound healing.	Flavonoids, caffeic acid derivatives and $\alpha$ -tocopherol.	[62–64]
<b>Asphodelaceae</b>						
<i>Bulbine capitata</i> Poelln.	Scented grass bulbine	Roots Aerial parts	South Africa	Anti-inflammatory and weak antioxidant and free radical scavenging and lipid Peroxidation inhibition activities. Kniphofolone as a selective inhibitor of leukotriene metabolism. Used as a mild purgative and to cure gonorrhoeal infections.	Anthraquinone Kniphofolone	[65–73]
<i>Bulbine frutescens</i> Willd.	Snake flower, cat's tail, burn jelly plant	Leaf juice Roots	South Africa	Anti-inflammatory and weak antioxidant and free radical scavenging and lipid Peroxidation inhibition activities. Kniphofolone is a selective inhibitor of leukotriene metabolism. Used to treat burns, rashes, blisters, insect bites, cracked lips, acne, cold sores, mouth ulcers and areas of cracked skin.	Phenylanthraquinones, Isouranopaphthoquinones, Gaboroquinones A and B and $4'$ -O-demethylkniphofolone- $4'$ -O-beta-D-anthroquinone)	[65, 67, 70, 74, 75]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Kniphofia foliosa</i> Hochst. Red-not-peker			Kenya	Anti-inflammatory and weak antioxidant and free radical scavenging and lipid peroxidation inhibition activities. Knipholone as a selective inhibitor of leukotriene metabolism. Used for abdominal cramps, wound healing	Anthraquinone: Knipholone	[65, 76–78]
<b>Asteraceae</b>						
<i>Artemisia abyssinica</i> Sch.Bip.	Chikugn (Amharic) <i>Arrta bera (Or)</i>	Whole plant	Ethiopia	Radical scavenging and antioxidant activities. Used for stomach pain and wound healing.	Essential oils and flavonoids	[79–82]
<i>A. afra</i> Jacq. ex Willd.	African wormwood Wild wormwood	Roots, stems and leaves	Ethiopia South Africa	Radical scavenging and antioxidant activities. Used for stomach pain, coughs, colds, fever, loss of appetite, colic, headache, earache, intestinal worms to malaria.	Essential oils and flavonoids	[79, 82–84]
<i>A. arvensis</i> L.	Mugwort Wormwood	Whole plant	Algeria	Radical scavenging and antioxidant activities.	Phenolic compounds and flavonoids.	[85]
<i>A. campestris</i> L.	Field sagewort Field wormwood	Whole plant	Algeria	Radical scavenging and antioxidant activities. Used to treat insomnia	Phenolic compounds and flavonoids.	[85–87]
<i>Bidens pilosa</i> L.	Black jack	Leaves Roots	South Africa	Antioxidant and anti-inflammatory; antibacterial, antihypertensive activities. Used to treat diabetes and backache.	Phenolic compounds: quercetin 3-O-rabinobioside, quercetin 3-O-rutinoside. Two novel methoxylated flavone glycosides: quercetin 3,3'-dimethyl ether 7-O- $\beta$ -D-glucopyranosyl-(1 $\sim$ 6)- $\beta$ -D-glucopyranoside and the known quercetin 3,3'-dimethyl ether 7-O- $\beta$ -D-glucopyranoside	[19, 88–91]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Cynara scolymus</i> L.	Globe artichoke	Leaves	Ethiopia	Antioxidative and lipid-lowering properties and eNOS up-regulating ability. Used to treat chronic liver and gall bladder diseases, jaundice, hepatitis and atherosclerosis.	Polyphenolic flavonoid compounds	[14, 15, 92, 93]
<i>Helichrysum dasycanthum</i> Sweet	common name of kooigoed (bedding material)	Leaves	South Africa	Antioxidant, radical scavenging and anti-inflammatory activities. Used to treat wounds, infections, respiratory conditions.	Essential oils	[94–96]
<i>H. petiolare</i> Hilliard & B.L. Burtt.	Everlasting, Imphepho	Leaves	South Africa	Antioxidant, radical scavenging and anti-inflammatory activities. Used to treat wounds, infections, respiratory conditions, asthma, chest problems and high blood pressure	Essential oils	[94–96]
<b>Balanophoraceae</b>		Knaki bush stinking roger muster John Henry wild marigold	Leaves	Madagascar	Antimicrobial and antioxidant activity. Used as anthelmintic, antispasmodic, purgative and for the treatment of gastritis, indigestion and internal worms.	Essential oils.
<i>Thonningia sanguinea</i> Vahl.	Nkomango	Roots	Ghana	Antioxidative and radical scavenging activities and lipid peroxidation inhibitory activity. Used for bronchial asthma, rheumatoïd arthritis, atherosclerosis and diabetes.	Ellagittannins: Thonningianin A and B	[98–103]
<b>Balanitaceae</b>				Antioxidant properties <i>in vitro</i> confirmed. The bark and roots are used as laxatives, and for colic. The bark is used for sore throats, and as a remedy for sterility, mental diseases, epilepsy, yellow fever, syphilis, and tooth aches.	Coumarins, flavonoids, saponins (Balanin 1 ( $\beta,12\beta,14\beta,16\beta$ )-cholest-5-ene-3,16-diybis ( $\beta$ -d-glucopyranoside)-12-sulphate, a new sterol sulfonated and Balanin 2 ( $3\beta,20S,22R,25R$ )-26-hydroxy-22-acetoxysterost-5-en-3-yl-rhamnopyranosyl-(1 $\rightarrow$ 2)-glucopyranoside, a novel furostanol saponin)	[11, 104–106]
<b>Bignoniaceae</b>						
<i>Jacaranda mimosifolia</i> D.Don.	Sharpleaf Jacaranda	Leaves Stem-bark	Nigeria	Shown to have antimicrobial activity and used to treat infections	Phenylethanoid glucoside, jacaranone	[107–109]
<i>Spathodea campanulata</i> P.Beauv.	African tulip	Stem-bark	Nigeria, Ghana, Cameroon (Yaounde region)	Anti-inflammatory, antioxidant, hypoglycemic, anticomplement and anti-HIV activities. Used to treat itching, arthritis, and diabetes.	Flavonoids and caffeic acid derivatives	[63, 110]

TABLE I. Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Tecoma stans</i> (L.) H.B. & K.	Yellow trumpet bush	Leaves Stem-bark	Nigeria	Anti-diabetic activity is shown.	4-O-E-caffeoxy]-alpha-L-rhamnopyranosyl-[1' → 3)-alpha/beta-D-glucopyranose, E/Z-acetoside, isoacetoside	[107, 111]
<b>Capparaceae</b>						
<i>Cleome arabica</i> L.	Cleome eféina	Leaves	Egypt	Antioxidant activity, inhibited lipoxygenase activity and calcium ionophore-stimulated LTB4 synthesis in human neutrophils. Used to treat wounds and prevent inflammation	Rutin and quercetin.	[112, 113]
<b>Clusiaceae</b>						
<i>Garcinia kola</i> Heckel	Bitter cola/aku ilu, agbu ilu. Nigeria Hausa: Góóró p/ gwârrâá or góorrârkâí	Seeds	Nigeria	Inhibit lipid peroxidation and protective against H <sub>2</sub> O <sub>2</sub> -induced DNA strand breaks and oxidized bases. Used for laryngitis, coughs, liver disease, bronchitis and throat infections. Inhibits Aflatoxin B <sub>1</sub> induced genotoxicity.	Biflavanoid: kolaviron	[114–120]
<i>Harungana madagascariensis</i> Poir.	Otori	Stem-bark	Eastern Nigeria	Significant antioxidant activity. Used to treat skin diseases.	Prenylated Anthronoids: harunnumadagascarin A [8,9-dihydroxy-4,4-bis-(3,3-dimethylallyl)-6-methyl-2,3-(2,2-dimethylpyran)oanthrone], harunganol B	[121–123]
<i>Hypericum carinatum</i> Griseb.	Not signalized	Leaves	Egypt	Antioxidant and radical scavenging activities.	Benzophenones: cariphenone A (6-benzoyl-5,7-dihydroxy-2,2,8-trimethyl-2H-chromene) and cariphenone B (8-benzoyl-5,7-dihydroxy-2,2,6-trimethyl-2H-chromene).	[124, 125]
<i>H. perforatum</i> L.	Common St. John's wort	Whole plant	Egypt	Anti-inflammatory and anti-oxidant activities. Free radical scavenging, metal-chelation, and reactive oxygen quenching activities. Protective against scopolamine-induced altered brain oxidative stress status and amnesia in rats. Ability to suppress the activities of 5-lipoxygenase (5-LO) and cyclooxygenase-2 (COX-2), key enzymes in the formation of proinflammatory eicosanoids from arachidonic acid (AA). Analgesic, antiseptic, antispasmodic, digestive, diuretic and sedative.	Flavonoids: Rutin, hyperoside, isoquercitrin, avicularin, quercitrin, and quercetin.	[124, 126–131]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Cochlospermaceae</b>						
<i>Cochlospermum tinctorium</i> A.Rich.	N'iribara	Roots	Sudan, Uganda West Africa	Antioxidant activity. Used for malaria, jaundice.	Polyphenols: gallotannins and ferulic acids	[35]
<b>Combretaceae</b>						
<i>Combretum woodii</i> Drum.	Large-leaved forest bushwillow	Leaf	South Africa	Antioxidant and antibacterial activities. Also tannins showed inhibitory effect on Fe <sup>2+</sup> -induced lipid peroxidation and radical scavenger activity. Used for pneumonia, syphilis, abdominal pain and conjunctivitis.	Polyphenols: Combretastatin B5 (2',3'-trihydroxy-3,5,4'-trimethoxybibenzyl). Tannins.	[132-137]
<i>Combretum imberbe</i>	Not signified		South Africa	Combretum species are widely used for treating abdominal disorders (e.g. abdominal pains, diarrhea) backache, bilharziasis, chest coughs, colds, conjunctivitis, dysmenorrhoea, earache, fattening babies, fever, headache	$\alpha,3\beta$ -dihydroxy-12-oleanen-29-oic, 1-hydroxy-12-olean-30-oic acid, 3,30-dihydroxy-12-oleanen-22-one, and 1,3,24-trihydroxy-12-olean-29-oic acid, a new pentacyclic triterpenoid ( $\alpha,23$ -dihydroxy-12-oleanen-29-oic acid- $3\beta$ -O-2,4-di-acetyl-1-rhamnopyranoside)	[138]
<i>Guiera senegalensis</i> J.F.Gmel.	N'kundjé	Leaf	Western Africa	Antioxidant and radical scavenging activities. Used to treat dysentery, diarrhoea, gastro-intestinal pains and disorders, rheumatism, diabetes and fever.	Flavonol aglycones, flavonol glycosides and flavonoids (catechin, myricitrin, rutin and quercetin) as well as tannins (galloylquinic acids (hydrolysable tannins).	[139-143]
<i>Terminalia sericea</i> Burch. ex DC.	Silver cluster-leaf	Bark	South Africa	Radical scavenging and antioxidant activities. Used to treat diabetes and pneumonia and to relieve colic	Pentacyclic triterpenoids Anolignan B	[21, 136, 144]
<b>Commelinaceae</b>						
<i>Commelina diffusa</i> Burn.f.	Wandering Jew Climbing day flower	Leaves	Ghana	Anti-inflammatory and antioxidant properties. Used to treat fever and is diuretic	Flavonoids	[63, 145]
<i>Palisota hirsuta</i> K.Schum.,	Not signified	Aqueous leaf extracts	Nigeria	Anti-inflammatory effects against carageenan induced hind paw oedema	Not identified	[146, 147]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Crassulaceae</b>						
<i>Bryophyllum pinnatum</i> (Lam.) Oken Synonym: <i>Kalanchoe pinnata</i> (Lam.) Peters.	Ufu ivo	Leaves	Nigeria, South Africa	Anti-inflammatory properties. Used for earache.	Flavonoids, polyphenols, triterpenoids	[12, 148, 149]
<b>Cupressaceae</b>						
<i>Juniperus procera</i> Hochst ex. Endl.	African Juniper	Young twigs and buds	Ethiopia	Antioxidant and free radical scavenging activities. Used to relieve stomach pain.	Essential oils	[79, 150, 151]
<b>Dioscoreaceae</b>						
<i>Dioscorea dumetorum</i> (Kunth) Pax	Yam	Tubers	Nigeria	Antioxidant activity to modify serum lipid and anti-inflammatory activity. Used to treat diabetes.	Dioscorea and Dioscoretine	[152-154]
<i>Drosera madagascariensis</i> (DC.) D. ramentacea Burchell	Sundew	Roots and flowers	Madagascar	Anti-inflammatory effects. Used to treat coughs and asthma	Flavonoids: hyperoside, quercetin and isoquercitrin	[155, 156]
<i>Drosera rotundifolia</i> L.	Round-leaf Sundew	Roots and flowers	Madagascar	Anti-inflammatory effects. Used to treat coughs and asthma	Flavonoids: hyperoside, quercetin and isoquercitrin	[155, 157]
<b>Euphorbiaceae</b>						
<i>Alchornea laxiflora</i> (Benth) Pax & K. Hoffm.	Wild banana	Leaf and root	Nigeria	Antioxidant and anti-microbial activity. Used to treat jaundice and liver disorders. Also used in food preservation.	Quercetin-7,4'-disulphate, quercetin, quercetin-3',4'-disulphate, quercetin-3,4'-diacetate, rutin and querctein	[158-161]
<i>Bridelia ferruginea</i> Benth.	Ora	Leaves, stem and bark	West Africa Democratic republic of Congo, Nigeria	Anti-inflammatory. Used to treat diarrhea, dysentery, gastro-intestinal disorders, gynecological disorders (including sterility), and rheumatic pains.	A bioflavonoid: Gallocatechin-(4' → O →)7-Epigallocatechin.	[12, 57, 162-166]
<i>Mallotus oppositifolius</i> (Geiseler) Muell. Arg.	Jororo	Káfar mütuwàà Senampendi Mvundzajembe	Leaves, roots	West Africa Nigeria	Antioxidant, anti-inflammatory and antimicrobial activities. Used for abortion.	[167-172]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Fabaceae</b>						
<i>Aspalathus linearis</i> (Brum. F.) R. Dahgr.	Rooibos	Leaves	South Africa	Radical Scavenging Capacity Used to treat stomach cramps, insomnia, and to reduce stress.	Phenolic Fractions, Tannins and monomeric flavonoids aspalathin, nothofagin, quercetin, rutin, isoquercitrin, orientin, isorientin, luteolin, vitexin, isovitexin, and chrysoceriol.	[16-21, 173, 174]
<i>Burkea africana</i> Hook	Wild Syringa	Bark	Mali and Sub Saharan Africa	Antioxidant and radical scavenging activity. Used to treat coughs, colds, stomach obstruction, infusions against gonorrhoea and syphilis.	Proanthocyanidins; fisetinidol-(4alpha->8)-catechin 3-gallate and bis-fisetinidol-(4alpha->6, 4alpha->8)-catechin 3-gallate, with smaller amounts of flavan-3-ols (catechin, epicatechin and fisetinidol)	[175, 176]
<i>Crotalaria podocarpa</i> DC.	Crotalaria	Roots	South Africa	Anti-inflammatory activity. Used for the treatment of sore-eyes and boils. Expectorant.	Flavonoids	[67, 177]
<i>Cyclopia intermedia</i> E. Mey. and C. subternata Vog.	Honeybush	Leaves and stem	South Africa	Antioxidant activity. Used as tonic for colds, catarrh and tuberculosis.	Pinitol, shikimic acid, p-coumaric acid, 4-glucosyltyrosol, epigallocatechin gallate, the isoflavone orobol, the flavanones hesperedin, narirutin and eriocitrin, a glycosylated flavan, the flavones luteolin, 5-deoxyluteolin and scolymoside, the xanthone mangiferin and the flavonol C-6-glucosylkaempferol. Phenolic content: tyrosol and a methoxy analogue, 2-[4-([O-alpha-apiofuranosyl-(1'>6')-beta-d-glucopyranosyloxy]phenyl)ethanol, 4-[O-alpha-apiofuranosyl-(1'>2')-beta-d-glucopyranosyloxy]benzaldehyde, five glycosylated flavonols, two isoflavones, four flavanones, two isoflavones, and two flavones	[19, 21, 178-181]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area validation	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Eriosema robustum</i>	Twigs	Burundi, Ethiopia, Kenya, Rwanda, Tanzania, Uganda, Democratic Republic of Congo and Cameroon	Used traditionally for the treatment of coughs in East Africa and skin diseases in Central Africa	$2',3',5',5,7\text{-pentahydroxy-3,40-dimethoxyflavone}$ , $2',3,5',5,7\text{-pentahydroxy-4',methoxyflavone}$	[182, 183]	
<i>Erythrina latissima</i> E. Mey.	Broad-leaved coral tree	Stem Wood Root wood Seeds	South Africa Botswana	Antimicrobial activity and weak radical scavenging properties. Purgative.	Flavonoids and isoflavonoids. Isoflavones: erytissin A and B. Flavanone: erytissin C and flavonoids and Isoflavone glycosides: $4'\text{-hydroxyisoflavanone-7-O-beta-D-glucopyranoside}$ (compound 1); $4'\text{-hydroxyisoflavanone-7-O-alpha-L-rhamnose}$ ( $1\rightarrow6$ )-beta-D-glucopyranoside (compound 2); and a new compound $4',8\text{-dimethoxyisoflavanone-7-O-alpha-L-rhamnose}$ ( $1\rightarrow6$ ) glucopyranoside ( $8\text{-O-methylretusin-7-O-alpha-L-rhamnose}$ ( $1\text{-}6$ )-beta-D-glucopyranoside) (compound 3)	[67, 184-186]
<i>E. lysistemon</i> Hutch.	Common coral tree; lucky bean tree	Bark	South Africa	Mild antioxidant activity. Used to treat sores, wounds, abscesses and arthritis.	Isoflavonoids: 5,7-dihydroxy-2',4',5'-trimethoxyisoflavanone. Three prenylated flavonoid derivatives: $5,7,4'\text{-trihydroxy-8-(3''\text{-methylbut-2''-enyl})-6-(2''\text{-hydroxy-3''\text{-methylbut-3''-enyl}) isoflavanone}$ (isoerysenegalesein E), $5,7,2'\text{-trihydroxy-4',methoxy-5'-}(3''\text{-methylbut-2''-enyl})$ isoflavanone (lysiseisoflavanone), $5,4'\text{-dihydroxy-6-(3'''-methylbut-2'''-enyl)-2',}-hydroxyisopropyl dilydrofuran-4'',5'',8,7] isoflavanone (isosenelegaleinsin), together with the four known flavonoids abyssinone V-4'-methylether, alpinumisoflavanone, wighteone and burttinone$	[187-190]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
Anti-inflammatory properties.						
<i>Melilotus elegans</i> Salzm. ex Ser. (syn. M. abyssinica Baker)	Egug, Gugi, Yemen berri Elegant sweet clover	Leaves	Ethiopia	Used for asthma, haemorrhoid, wounds, excavated sore, piles, ulcers mouth infection, lacerated wounds, haemorrhoids, bronchial asthma (personal communication)	Flavonoids: kaempferol	[191–194]
<i>Milletia griffoniana</i> Baill.	Not signalized	Root-bark and seeds	Cameroon	Anti-inflammatory activity. Used as an antimalarial.	Coumarin: 4-hydroxy-3-(3',4'-methylenedioxyphenyl)-5,6,7-trimethoxycoumarin, durmilloine, odorantin, 7-methoxybenosin, calopogonium isoflavone B and 7,2'-dimethoxy-4',5'-methylenedioxy isoflavone maxima isoflavone G (5) and 7-hydroxy-6-methoxy-3',4'-methylenedioxyisoflavone and new prenylated isoflavonoids griffonianones A, B, C, D and E. Griffonianone D ((7E)-6'',7''-dihydroxy-3'',7''-dimethyl-oct-2''-enyl)oxy-4'-methoxyisoflavone), an isoflavone.	[195–202]
<i>Parkia biglobosa</i> (Jacq.) Benth	African Locust Bean Néré Ojinyi	Bark Seeds	Mali Sudan Ivory Coast	Anti-inflammatory activity. Used as antiseptic and to treat coughs, chest pain, and wound healing	Tocopherol, ascorbic acid (Seeds)	[12, 33, 34, 36–39, 43–53, 55, 64, 66–72, 118, 119, 121, 138, 159, 182, 195, 203–235]
<i>Peltophorum africanum</i> Sond.	Weeping wattle	Root and bark	South Africa	Antioxidant and antibacterial activities Used to treat diarrhoea, dysentery, sore throat, wounds, back and joint pains, HIV-AIDS, venereal diseases and infertility.	Flavonol glycosides and flavonol glucoside gallates	[236–238]
<i>Pliostigma thomningii</i> (Schum.) Milne- Redh	Camel's foot tree, Monkey Bread Niam (Mali). Abefe Kalgo Okpoatu Omepa		Nigeria, Ethiopia Botswana, Kenya, Namibia, Senegal, South Africa, Sudan, Tanzania, Uganda, Zambia	Anti-oxidant and anti-inflammatory properties. Used to treat wounds, chronic ulcers, cough, respiratory disorders and toothache, gum inflammation, arthritis, headache, backache, and infertility.	Proanthocyanidins epicatechin, catechin trimers and oligomers, flavonoids, polyphenolics, C-methylflavonols (in the leaf extract)	[12, 58, 239–245]
<i>Sutherlandia frutescens</i> R Br.	Cancerbush Phetola	Leaves	South Africa	Superoxide and hydrogen peroxide scavenging activities. Used as tonic to boost the immune system.	Canavanine, pinitol	[246–248]
<i>Trigonella foenumgraecum</i> L.	Fenugreek	Seeds	Ethiopia, Morocco	Protective effect against Oxidative stress during ischaemia-reperfusion. It is hypolipidemic, and is also used to treat boils and to improve appetite.	Free phenolics and Vit C.	[26–28, 249, 250]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Humiriaceae</b>						
<i>Sacoglottis gabonensis</i> Urb.	Cherry tree, ozouga	Stem-bark	West Africa	Antioxidant activity.	Bergenin	[251-254]
<b>Hypooxidaceae</b>						
<i>Hypoxis hemerocallidea</i> Fisch. & C.A. Mey.	African potato	Corms	South Africa	Antioxidant activity. Used to treat tuberculosis, cancer, bladder Rooperol disorders, benign prostatic hyperplasia.		[188, 255-257]
<b>Lamiaceae</b>						
<i>Ocimum basilicum</i> L.	Mitkandu Basil	Leaves	Burkina Faso Ethiopia	Intermediate antioxidant activity and high antibacterial activity. Used in Ethiopia to treat Conjunctivitis and in Kenya to treat colds and stomachache.	Linalool basil oil Methyl chavicol, eugenol, (E)-methyl cinnamate, thymol, linalool	[23, 258]
<i>Ocimum gratissimum</i> L.	Tea bush, Scent leaf/Nchuanwu. Ujuju okpevu Basil	Leaves	Popular republic of Congo (ex Brazzaville Congo) Eastern Nigeria	Antioxidant activity Popular republic of Congo it is used as a laxative, purgative, and to treat snakebite, diabetes, tooth ache, gingivitis.	Xanthomicrol, cirsimarin, rutin, kaempferol 3-O-rutinoside and vicenin-2 were identified as the major flavonoids, whereas luteolin 5-O-glucoside, luteolin 7-O glucoside, apigenin 7-O-glucoside, viexin, isovitexin, quercetin 3-O-glucoside and isothymusin were detected as minor constituents.	[12, 58, 258-262]
<b>Lauraceae</b>						
<i>Cinnamomum zeylanicum</i> Breyne	Cinnamon leaf	Leaves	Madagascar Ethiopia	Very high antioxidant and high antimicrobial activities. Used to treat diarrhoea, rheumatism, colds and hypertension	Cinnamaldehyde, eugenol and eugenyl acetate to be the main constituents of cinnamon oil.	[22-24, 263]
<i>Ocotea bullata</i> (Burch.) Baill.	Black stinkwood Unukane (Zulu)	Bark	South Africa	Anti-inflammatory, cyclooxygenase inhibitory activity. Urinary disorders, headaches.	Monoterpenoids	[188, 264]
<i>Ravensara aromatica</i> Sonn.	Nutmeg havozo	Bark Leaf	Madagascar	Low antioxidant and antimicrobial activity. Useful for chronic respiratory conditions, barks, estragole (methyl chavicol) but leaves and sometimes helpful in cases of asthma.	Essential oils, principally composed of the monoterpenic hydrocarbons a-pinene, sabinene, myrcene, limonene, & the azulene: iso-lidene. In barks, estragole (methyl chavicol) but leaves contain b-myrcene, 1,8-cineole, linalool, and carotol.	[23, 25, 265]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Malvaceae</b>						
<i>Hibiscus sabdariffa</i> L.	Red tea, sorelle Rosella	Flowers	Nigeria South Africa	Antimutagenic activity and free radical scavenging effects on active oxygen species Used against insomnia, colic.	Flavonol glucoside hibiscitrin Anthocyanins. Such as cyanidin 3-O-β-D-glucopyranoside, cyanidin 3-O-(2-O-β-D-xylopyranosyl)-β-D-glucopyranoside, delphinidin 3-O-β-D-glucopyranoside and delphinidin 3-O-(2-O-β-D-xylopyranosyl)-β-D-glucopyranoside.	[19, 21, 266-269]
<b>Meliaceae</b>						
<i>Trichilia roka</i> Chiiov.	Soulafinzan	Root	Tropical Africa Mali	Significantly protective against CCl <sub>4</sub> -induced liver damage and prevented perisinusoidal fibrosis. Used to treat malaria, abdominal pain and dermatitis.	Polyphenols	[270, 271]
<b>Menispermaceae</b>						
<i>Sphenocentrum jollyanum</i> Pierre	Akerejupon ajo	Fruit Root	West Africa	Anti-inflammatory activity. Used to treat inflammatory-based diseases	Furanoditerpenes: columbin, isocolumbin. Flavonoids-rich fraction.	[272-274]
<i>Timospora bakis</i>	Whole plant	Sudan		Anti-inflammatory activity. To treat headache and rheumatism	A diterpenoid furanolactone, columbin	[275]
<b>Moraceae</b>						
<i>Dorstenia barteri</i> var. <i>subtriangularis</i> (Engler)	Contrayerva	Twigs/leaves	Cameroon	Antioxidant properties account for the anti-inflammatory action of these extracts chalcones: 3,5'-di-(2-hydroxy-3-methylbut-3-enyl)-4,2',4'-trihydroxy-3',5'-prenylchalcone and bartericins A (-)-3-(3,3-dimethylallyl)-5'-(2-hydroxy-3-methylbut-3-enyl)-4,2',4'-trihydroxychalcone, bartericins B (+)-3-(3,3-dimethylallyl)-4',5'-[2'''-(1-hydroxy-1-methylethyl)-dihydrofuranol]-4,2' -dihydroxychalcone and bartericins C 3,4-(6'',6''-dimethyl)dihydropyrano)-4',5'-[2'''-(1-hydroxy-1-methylethyl)-dihydrofuranol]-4,2' -hydroxychalcone and also two novel diprenylated chalcones: 3,5'-di-(2-hydroxy-3-methylbut-3-enyl)-4,2',4'-trihydroxy-3',5'-prenylchalcone and 4,2',4'-trihydroxy-3',5'-diprenylchalcone; and 5,7,4'-trihydroxy-8-prenyflavone. Other known compounds such as stipulin, 4-hydroxylichocarpin, kanzanol B, 3'-(2-Hydroxy-3-methylbut-3-enyl)-5'-(3,3-dimethylallyl)-4,2',4'-trihydroxychalcone, and dorstenone.	[67, 276-281]	

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>D. ciliata</i> Engl.	Contrayerva	Aerial parts	Cameroon Central Africa	Antiradical and antioxidant activities. Used as food additive.	phenolic compound (6-prenylapigenin) Flavones: (ciliatin A) 5,4'-Dihydroxy-5''-isopropenylidihyfuranol[2'',3'',7;6]flavone (ciliatin B) 7,4'-Dihydroxy-3'-methoxy-6'',6''-dimethylidihydropyranol[2',3';5,6].	[282-284]
<i>D. convexa</i> De Wild.	Contrayerva	Twigs and leaves	Democratic Republic of the Congo	Antioxidant properties account for the anti-inflammatory action of these extracts. Used to treat arthritis, rheumatism, gout, headache and other forms of body pains.	Prenylated flavonoids	[67, 276, 280]
<i>D. manni</i> Hook.f.	Contrayerva	Twigs/leaves Aerial parts	Central Africa	Antioxidant action against copper-induced LDL oxidation, this activity is like the non-prenylated flavonoid quercetin. Also, inhibition of platelet aggregation and influence of cyclooxygenase and lipoxygenase activity. Used to treat rheumatism, stomach disorders. Anti-trichomonial activity.	Grenylated and prenylated flavonoids and flavonones: Flavonones: 6,8-diprenyl-5,7,3',4'-tetrahydroxyflavanone, 4-hydroxylonchocarpin, 4-methoxylonchocarpin, 6-prenylchrysoeriol, 6,8-diprenyleriodictyol, ganaconin P and Prenylated flavonoids: 6,8-diprenyleriodictyol, dorsmanin C 7,8-(2,2-Dimethylchromeno)-6-geranyl-3,5,3',4'-tetrahydroxyflavone and dorsmanin D 6,8-Diprenyl-3,5,7,4'-tetrahydroxy-3'-methoxyflavone, dorsmanins I, J and 2''-epimers of dorsmanins F [67, 187, 207, (6,7-(2,2-dimethylpyrano)-8-prenyl-5,3',4'-trihydroxyflavanone, G (6,7-(2,2-dimethylidihydro-pyrano)-8-prenyl-5,3',4'-trihydroxflavanone). Also, dorsmanins F and G. Four new prenylated flavanones, named dorsamine F (7,8-[2''-(1-hydroxy-1-methylethyl)]-dihyfurano]-6-prenyl-5,3',4'-trihydroxyflavanone), dorsamine G (6,7-[2''-(1-hydroxy-1-methylethyl)dihyfurano]-8-prenyl-5,3',4'-trihydroxyflavanone) and dorsamine H (6-prenyl-8-(2-hydroxy-3-methylbut-3-enyl)-5,7,3',4'-tetrahydroxyflavanone).	[285-287]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>D. poinsettifolia</i> var. <i>angusta</i> Engl.	Dingetenga	Whole plant	Cameroun	Antiradical and antioxidant activities. Used to treat infected wounds.	Germylated and prenylated flavonoids. The unusual 4-phenyl-substituted dihydrocoumarin and the rare geranyl-and prenyl-substituted Chalcone.	[207, 288, 289]
<i>D. psilurus</i> Welw.	Dingetenga	Roots	Cameroon Central Africa	Antiradical and antioxidant activities. Used against snakebite and to treat rheumatism, headache and stomach disorders.	Germylated and prenylated flavonoids. Three phenolic compounds: 6,8-diprenyl-3'-[O],4'-(2,2-dimethylpyrano)-3,5,7-trihydroxyflavone, 3,6-diprenyl-8-(2-hydroxy-3-methylbut-3-enyl)-5,7,2',4'-tetrahydroxyflavone and an unusual B/C ring modified flavonoid derivative for which the names dorsilurins C, D and E, respectively, are proposed. Two new flavones, dorsilurins A and B, and a new benzofuran derivative have been isolated from Dorstenia psilurus, together with three known phenylpropanoid derivatives, stearyl-p-coumarate [octadecanyl 3-(4-hydroxyphenyl)prop-2-enate], stearyl ferulate [octadecanyl 3-(4-hydroxy-3-methoxyphenyl)prop-2-enate] and psoralen.	[206, 282, 290-292]
<b>Myrtaceae</b>						
<i>Eugenia elliptica</i> Sm. <i>Syzygium smithii</i> (Poir.) Nied.	Lily Pilly	Leaves	Mauritius	Modulate the expression of the antioxidant enzyme genes.	Quercetin-3-O-galactoside (hyperoside), kaempferol-3-glucoside (astragalin), queretin-3-O-glucoside (isoquercitrin), (+)-catech.	[293, 294]
<i>E. fasciculata</i> Wall.	Not signalized	Leaves	Mauritius	Modulate the expression of the antioxidant enzyme genes.	Quercetin-3-O-galactoside (hyperoside), kaempferol-3-glucoside (astragalin), queretin-3-O-glucoside (isoquercitrin), (+)-catech.	[293]
<i>E. orbiculata</i> Lam.	Not signalized	Leaves	Mauritius	Modulate the expression of the antioxidant enzyme genes.	Quercetin-3-O-galactoside (hyperoside), kaempferol-3-glucoside (astragalin), queretin-3-O-glucoside (isoquercitrin), (+)-catech.	[293, 295]
					queretin-3-O-rutinoside (rutin),	

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>E. pollicina</i> J.Guelho & A.J.Scott	Not signalized	Leaves	Mauritius	Modulate the expression of the antioxidant enzyme genes.	Quercetin-3-O-galactoside (hyperoside), kaempferol-3-glucoside (astragalin), queretin-3-O-glucoside (isoquercitrin), (+)-catech. (-)-epicatechin gallate	[293, 296]
<i>Monnieriastrum acutispalum</i> J.Guelho & A.J.Scott.	Not signalized	Leaves	Mauritius	Modulate the expression of the antioxidant enzyme genes.	Quercetin-3-O-galactoside (hyperoside), kaempferol-3-glucoside (astragalin), and queretin-3-O-glucoside (isoquercitrin). (+)-catechin	[293-295]
<i>M. globosum</i> J.Guelho & A.J.Scott	Not signalized	Leaves	Mauritius	Modulate the expression of the antioxidant enzyme genes.	Quercetin-3-O-galactoside (hyperoside), kaempferol-3-glucoside (astragalin), and queretin-3-O-glucoside (isoquercitrin). (-)-catechin	[293]
<i>Syzygium aromaticum</i> (L.) Merr. & L.M.Perry	Clove bud	Dried flowers Buds	Madagascar Sudan	Antioxidant and antimicrobial activities. Used to treat tooth ache and throat inflammation.	Eugenol Methyleugenol	[23, 297, 298]
<i>S. coriaceum</i> J.Bosser & J.Guého	Bois de pomme		Mauritius	Abilities to modulate the expression of the antioxidant enzyme genes.	Phenols and flavonoids: Quercetin-3-O-rutinoside, kaempferol-3-glucoside (astragalin) and queretin-3-O-glucoside (isoquercitrin), (+)-catechin, procyanidin B1 dimer, (-)-epicatechin gallate	[293]
<i>S. glomeratum</i> DC.	Bois de pomme	Leaves	Mauritius	Abilities to modulate the expression of the antioxidant enzyme genes. Used to treat boils, abscesses, fever and wounds and as expectorant.	Phenols and flavonoids: kaempferol-3-glucoside (astragalin) and queretin-3-O-glucoside (isoquercitrin), procyanidin B1 dimer, (-)-epicatechin, chlorogenic acid, (-)-epicatechin	[293]
<i>S. guehoui</i>	Not signalized		Mauritius	Abilities to modulate the expression of the antioxidant enzyme genes.	Phenols and flavonoids: quercetin-3-O-rutinoside (rutin), kaempferol-3-glucoside (astragalin) and queretin-3-O-glucoside (isoquercitrin), (+)-catechin, chlorogenic acid, procyanidin B2 dimer	[293]
<i>S. mauritianum</i> J.Guelho & A.J.Scott	Not signalized	Leaves	Mauritius	Abilities to modulate the expression of the antioxidant enzyme genes.	Phenols and flavonoids: quercetin-3-O-rutinoside (rutin), kaempferol-3-glucoside (astragalin) and queretin-3-O-glucoside (isoquercitrin), (+)-catechin, chlorogenic acid	[293]
<i>S. petrinense</i> J.Bosser & J.Guého	Not signalized		Mauritius	Abilities to modulate the expression of the antioxidant enzyme genes.	Phenols and flavonoids: quercetin-3-O-rutinoside (rutin), kaempferol-3-glucoside (astragalin) and queretin-3-O-glucoside (isoquercitrin), procyanidin B1 dimer, chlorogenic acid	[293]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area validation	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>S. venosum</i> (Lam.) J.Guelo & A.J.Scott	Not signalized		Mauritius	Abilities to modulate the expression of the antioxidant enzyme genes.	Phenols and flavonoids: quercetin-3-O-rutinoside (rutin), kaempferol-3-glucoside (astragalin) and quercentin-3-O-glucoside (isoquercitrin), (+)-catechin, procyanidin B2 dimer	[293, 295, 299]
<b>Oleaceae</b>				Potent antioxidant activity. Used as eye lotions and tonics, lower blood pressure, improve kidney function and deal with sore throats. The early Cape settlers used the fruits to treat diarrhoea	Oleafricein (mixture of oleanolic acid and ursolic acids), Triterpenoids and oleoprotein.	[84, 300, 301]
<i>Olea europaea</i> subsp <i>africana</i> (Mill.)P.S. Green	African wild olive	Leaves	South Africa			
<b>Pedaliaceae</b>						
<i>Harpagophytum procumbens</i> D.C. ex Meissner	Devil's claw	Root	South Africa Native to the Kalahari Desert of southern Africa, Namibia and Botswana.	Anti-inflammatory and ability to inhibit the expression of cyclooxygenase-2 and inducible nitric oxide by suppression of NF-kappaB activation. Used for pain, muscular tension, osteoarthritis, degenerative rheumatism or painful arthrosis and tendonitis as well as tonic for loss of appetite and dyspeptic complaints.	Roots contain iridoid glycosides mainly harpagoside. Other constituents are flavones and flavonols kaempferol, and tuteolin.	[302–312]
<b>Piperaceae</b>						
<i>Piper guineense</i> Schum. & Thonn.	West African black pepper Bush pepper Ikom, Amana kakwale iyeyeh ashoesie taquale Meshoro	Fruit, seed and leaf	Ghana, West Africa Nigeria Cameroon	Antioxidant activity.	Volatile oil components-monoterpenes, sesquiterpenes, terpenoids, lignans and sterols.	[313–316]
<b>Podocarpaceae</b>						
Podocarpus species						
<i>Podocarpus elongatus</i>						
<i>Podocarpus falcatus</i> ,						
<i>Podocarpus henkelii</i>						
and <i>Podocarpus latifolius</i>						
					These species are used to treat fevers, asthma, coughs, cholera, chest complaints, arthritis, rheumatism, painful joints and venereal diseases	[317]
					Eastern and Southern Africa	Diterpenoids, bioflavonoids and Totarol

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Ranunculaceae</b>						
<i>Nigella sativa</i> L.	Black cumin	Seed	African countries in the Mediterranean region	Antioxidant potentials through scavenging ability of different free radicals including the superoxide anion radical, inhibition of lipid peroxidation, and protection of liver against carbon tetrachloride (CCl <sub>4</sub> ) -induced liver fibrosis in rabbits Used to treat diarrhoea, asthma, and as a gastroprotective agent.	Oil: Thymoquinone	[29–31, 318, 319]
<b>Rosaceae</b>						
<i>Crataegus monogyna</i> Jacq.	Hawthorn, May Blossom, May Day Flower, White Thorn.	Fresh vegetative and reproductive organs	Mauritius, Northern Africa	Antioxidant activities. Used for its neuro- and cardiosedative actions.	Polyphenols: (proanthocyanidin, flavonoid, anthocyanin, (-)-epicatechin, procyanidin B2, chlorogenic acid). Flavonoids: quercetin and quercentrin, glycosides, proanthocyanidins, anthocynaidins, saponins, tannins, and catechins Also, Vitamin C.	[320–323]
<i>Leucoidea sericea</i>		Leaf, bark and roots	Southern Africa	Antimicrobial and anti-inflammatory properties	Phenolics, alkaloids and saponins	[210]
<i>Pygeum africanum</i> Hook. f.	African plum tree Red Stinkwood	Bark	South Africa	Anti-inflammatory. Used to treat against benign prostatic hyperplasia, prostatitis	14% triterpenes (urolic acids, oleanolic acid, crataegolic acid), 0.5% n-docosanol Phytosterol ( $\beta$ -sitosterol, $\beta$ -sitostерон, Campesterol	[188, 324–327]
<b>Rubiaceae</b>						
<i>Crossopteryx febrifuga</i> Benth.	Roger Blench “rima” jogoo-hi/je”	Seeds Leaf and roots	Mali Nigeria	Radical scavenging and lipoxygenase inhibition activities. Used to treat fever and various respiratory diseases	Flavonoids	[328–330]
<b>Rutaceae</b>						
<i>Agathosma betulina</i> (Berg.) Pillans.	Round-leaf buchu	Leaves, stems	South Africa	Hydroxyl radical ion scavenging ability. Used for stomach problems, kidney and urinary tract diseases.	Anti-inflammatory activity. Used to treat benign prostatic hyperplasia, prostatitis, diabetes, inflammation of the colon, gums, and mucous membranes. Leaves chewed to relieve stomach complaints.	[188, 331, 332]
<i>A. crenulata</i> (L.) Pillans	Oval-leaf buchu	Leaves, stems	South Africa		Essential oils and flavonoids	[84, 188, 331, 332]

TABLE I: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Fagara zanthoxyloides</i> Lam.	xeti, xe Wô	Roots, root-bark	Cameroon, Uganda	Antioxidant activity. Used to treat gingivitis, toothache, urinary and venereal diseases, rheumatism and lumbago, malaria and other infections.	Phenylethanoid derivative, lignans and fagaronine	[333–336]
<b>Sapindaceae</b>						
<i>Dodonaea viscosa</i> Jacq.				Anti-inflammatory activity by inhibiting the synthesis of prostaglandin (PG) E(2). Used to treat rheumatism, skin infections, diarrhea, stomachaches, pains of hepatic and splenic origin, uterine colic. It is also used as an antipruritic in skin rashes and for the treatment of some throat, dermatitis and hemorrhoids.	Quercetin, isorhamnetin	[337–341]
Synonyms: <i>Dodonaea angustifolia</i> L. f.; <i>Ptelea viscosa</i> L.	Umusasa	Leaves	Rwanda			
<b>Xanthorrhoeaceae</b>						
<i>Aloe ferox</i> Mill.	Bitter aloe or Cape aloe	Leaves	South Africa, Lesotho	<i>A. ferox</i> gel contains at least 130 medicinal Chromones, anthraquinones, anthrone, agents with anti-inflammatory, analgesic, anthrone-C-glycosides, and other phenolic calming, antiseptic, antiviral, antiparasitic compounds and anticancer effects	Barbolin	[9]
<b>Zingiberaceae</b>				Anti-inflammatory activity through cyclooxygenase inhibitory (prostaglandin-synthetase inhibition), activity.	Sesquiterpenoid	[188, 264, 342]
<i>Siphonochilus aethiopicus</i> (Schweinf.) B.L. Burtt.	Wild ginger Natal ginger African Ginger	Rhizome	South Africa	Used to treat Coughs, colds, asthma.		

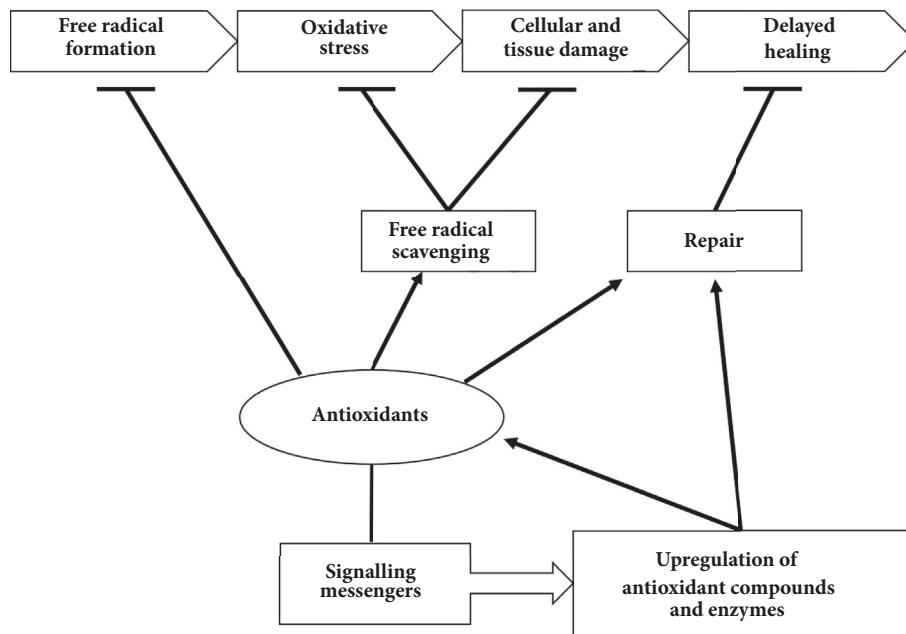


FIGURE 2: Mechanism of antioxidant action in wounds.

hydrogen peroxide ( $H_2O_2$ ) and other free radicals [348, 349]. Furthermore, increasing evidence suggests a relationship between metal overload and several chronic diseases through the induction of oxidative stress [350]. Therefore, inhibition of free radical formation using metal ions as targets could be useful therapeutically. Antioxidant assays designed for this purpose include the cupric and ferric reducing antioxidant power (CUPRAC/FRAP). These methods measure the ability of antioxidants to reduce cupric ( $Cu^{2+}$ ) and ferric ( $Fe^{3+}$ ) ions, respectively.

Another mechanism by which antioxidants act is through the suppression of oxidative stress by directly scavenging active free radicals. Most commonly reported antioxidant assays such as 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS), 2,2'-diphenyl-p-picrylhydrazyl radical (DPPH), oxygen radical absorbance capacity (ORAC), Trolox equivalent antioxidant capacity (TEAC), total oxyradical scavenging capacity (TOSC), and total radical antioxidant parameter (TRAP) are focused on testing the ability to scavenge free radicals. Furthermore, there are diverse cellular antioxidant assays that assess the ability of antioxidant compounds and substances to protect cells against excessive free radical generation. Such assays involve the use of a fluorescent compound such as 2,7-dichlorofluoroscein to determine the ability of test samples to quench intracellularly generated free radicals and inhibit radical formation and lipid peroxidation [345].

There are also numerous reports of the ability of antioxidants to repair damaged tissues and improve healing. Topical application of kojic acid and deferiprone, two compounds with the ability to scavenge free radicals, enhanced healing of wounds in rats [351]. Also, the mitochondria-targeted antioxidant, 10-(6'-plastoquinonyl) decyltriphenylphosphonium,

accelerated wound closure, stimulated epithelialization, granulation tissue formation, and vascularization, and lowered lipid peroxidation in mice [352]. Moreover, an antioxidant peptide (cathelicidin-OA1) promoted wound healing in a mouse model with full-thickness skin wounds, accelerated reepithelialization and granulation tissue formation by enhancing the recruitment of macrophages to the wound site, and induced cell proliferation and migration [353]. Some antioxidants have also been reported to contribute to healing by enhancing the activity of endogenous antioxidant compounds and enzymes. The induction of the nuclear factor E2-related factor 2-(Nrf2) mediated antioxidative pathway by a rhomboid family protein (RHBDL2) promoted healing of injured tissues, suggesting a relationship between antioxidant gene induction and healing [354]. Niconyl-peptide enhanced wound healing and protected against hydrogen peroxide-induced cell death by increasing the expression of Nrf2 expression in human keratinocytes [355].

The most common tests used to determine the antioxidant activity of samples included the assessment of the ability to scavenge free radicals such as DPPH, ABTS<sup>+</sup> [16, 19, 35, 62, 85, 94, 98, 99, 139, 158, 175, 184, 187, 266, 282, 302, 356–364], or the hydroxyl radicals [79, 188, 267, 365, 366], as well as the hydroperoxyl radicals by the Briggs-Rauscher reaction [104]. The ability of the extracts to chelate metal ions was also determined as further indication of their ability to contribute in the reduction of free radicals such as the hydroxyl radical [114]. In addition, assessment of the ability of these medicinal plant extracts to protect against lipid peroxidation was also included, which in turn was measured by the malondialdehyde-thiobarbituric acid (MDA) test [320, 367], the modified thiobarbituric acid reactive species (TBARS) assay [18, 22], or conjugated diene

formation [367]. Moreover, lipid peroxidation was assessed using the fluorescent probe, diphenyl-1-pyrenylphosphine (DPPP) [188], or using the inhibition of Cu(2+)-mediated oxidation of human low-density lipoprotein (LDL) [187, 367]. The ability of extracts to protect against damage to DNA using the Comet assay was also employed [114, 188].

The antioxidant capacity of the medicinal plant extracts was determined using either the TEAC or FRAP assays [11, 85, 302, 313, 321, 368]. The ability of extracts to modulate the gene expression of the antioxidant enzymes, such as Cu, Zn-superoxide dismutase (Cu, Zn-SOD), Mn-superoxide dismutase (Mn-SOD), catalase, and glutathione peroxidase (GPx), was also used as a measure of their antioxidant properties [293]. The photochemiluminescence (PLC) assay is a more recent antioxidant capacity assessment method and was employed for the evaluation of antioxidant capacity of baobab fruit pulp extracts [369].

Anti-inflammatory properties of these extracts were assessed by their ability to inhibit 5-lipoxygenases [94, 370, 371] or cyclooxygenase (COX-1 and COX-2) activities [65, 275, 317, 372, 373]. Using the former [374] and the latter [264, 331] methodologies, respectively, a great number of South African medicinal plant extracts were screened for their anti-inflammatory properties. The effect of medicinal extracts on the biosynthesis of different prostaglandins was assessed as a measure of their anti-inflammatory effect [239, 337, 375]. Extracts of *Podocarpus* species were shown to inhibit the activities of the COX enzymes [317]. Once again, using this test, the anti-inflammatory properties of the aqueous and ethanolic extracts of 39 plants used in traditional Zulu medicine were screened [376]. The Hen's Egg Test-Chorioallantoic Membrane (HET-CAM) assay which utilizes the CAM's capillary system in bred hen eggs was also used to assess the anti-inflammatory activity through antiangiogenic effects of the ethanol and aqueous extracts of *Drosera rotundifolia* and *D. madagascariensis* [155].

The antioxidant and anti-inflammatory abilities of the herbal extracts were further assessed by evaluating their ability to control the production of ROS produced by oxidative burst in neutrophils stimulated with L-formyl-L-methionyl-L-leucyl-L-phenylalanine (FMLP) [21, 246]. The inhibition of neutrophils elastase was used as a measure of anti-inflammatory property and it was proposed that the presence of flavonoids such as hyperoside, quercetin, and isoquercitrin in *D. rotundifolia* [377] and five flavonoid compounds in two *Polyodium* species (*P. decumanum* and *P. triseriale*) [378] were thought to contribute to this anti-inflammatory activity. These and other *in vitro* tests were used to assess the antioxidant properties of three Ghanaian species: *Spathodea campanulata*, *Commelina diffusa*, and *Secamone afzelii* [63].

Inflammation is a complex mechanism with many pathways. Several extracts derived from medicinal plants have been shown to modulate or inhibit the activities of mediators of inflammation. For instance, kolaviron, a bioflavonoid compound isolated from the seeds of *Garcinia kola*, has been reported to possess anti-inflammatory and antioxidant activities via its effects on COX-2 and inducible nitric oxide synthase (iNOS) by inhibiting the expression of nuclear factor kappa B (NF- $\kappa$ B) [115]. Quercetin is a flavonoid molecule

ubiquitous in nature and functions as an antioxidant and anti-inflammatory agent. Dose- and time-dependent effects of quercetin have been investigated on proinflammatory cytokine expression and iNOS, focusing on its effects on NF- $\kappa$ B signal transduction pathways in lipopolysaccharide-stimulated RAW 264.7 cells by using real time polymerase chain reaction (RT-PCR) and immunoblotting. Curcumin, a yellow pigment of turmeric, has been shown to exhibit anti-inflammatory activity. Curcumin has been found effective in the treatment or control of chronic inflammatory conditions such as rheumatism, atherosclerosis, type II diabetes, and cancer [203]. Calixto et al. reported that the anti-inflammatory action of active spice-derived components results from the disruption of the production of various inflammatory proteins (e.g., cytokines such as tumour necrosis factor-alpha (TNF- $\alpha$ ), iNOS, and COX-2) [379].

Animal studies were also conducted to assess the antioxidant properties of several medicinal extracts. The antioxidant potential of *Hypericum perforatum*, containing many polyphenolic compounds, was evaluated on splanchnic artery occlusion (SAO) shock-mediated injury [477] and also against elevated brain oxidative status induced by amnestic dose of scopolamine in rats [126]. Some medicinal plant extracts were tested for their ability to protect against carbon tetrachloride-, 2-acetylaminofluorene- (2-AAF-), and galactosamine-induced liver as well as aflatoxin B1-(AFB1)-induced genotoxicity. Using this test, it was found that an extract of *Garcinia kola* seeds [116, 478, 479], a decoction of *Trichilia roka* root [270], extracts of *Entada africana* [442], and *Thonningia sanguinea* [98, 480] possessed protective abilities. The antioxidant properties of plant extracts against potassium bromate (KBrO(3))-induced kidney damage showed the ability of *G. kola* seed extract to protect the kidneys [481].

Animal studies were also used to assess the anti-inflammatory ability of a great number of medicinal plant extracts using the carrageenan-induced rat paw oedema model. Plants investigated include seed extracts of *Picralima nitida* [399], crude methanol extract of the root of *Moringa oleifera* [469], powdered leaves and root of *Mallotus oppositifolium* [167], methanolic extract of *Picralima nitida* fruit [400], hot water extract of *Alstonia boonei* root-bark, *Rauvolfia vomitoria* root-bark, and *Elaeis guineensis* nuts [56], secondary root aqueous extract of *Harpagophytum procumbens* [303], crude extracts of *Sphenocentrum jollyanum* [272], aqueous and methanolic extracts of *Hypoxis hemerocallidea* corm [482], aqueous and methanolic extracts of *Sclerocarya birrea* stem-bark [483], aqueous extract of *Mangifera indica* stem-bark [13], aqueous extracts of *Leonotis leonurus* leaves [484], leaf extracts of *Bryophyllum pinnatum* [148], methanol extracts of the stem-bark of *Alstonia boonei* [485], aerial parts of *Amaranthus caudatus* [486], methanolic extracts of *Kigelia pinnata* flower [415], and leaf and twig extracts of *Dorstenia barteri* [276]. In all of these studies, the anti-inflammatory effect against carrageenan-induced rat paw oedema was attributed to flavonoids and other polyphenolic compounds. Animal tests also employed to assess the anti-inflammatory effects of the medicinal plant extracts included inflammatory cell response such as neutrophil chemotaxis

TABLE 2: Medicinal plants with confirmed antioxidant activity or medicinal plants that contain compounds that are not known to have antioxidant activity.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Acanthaceae</b>						
<i>Barleria species</i>		Leaves, twigs and roots	South Africa	Anti-inflammatory and antioxidant activities	Not identified	[212, 213]
<i>B. albostellata</i> , <i>B. greenii</i> , <i>B. prioritis</i>						
<b>Aizoaceae</b>						
<i>Hypoestes rosea</i> Deone.	Not signalized	Leaf extract	Nigeria	Anti-inflammatory activity due in part to its ability to inhibit NF-kappaB activation through direct inhibition of I kappaB kinase (IKK).	Diterpene: Hypoestoxide (a bicyclo [9,3,1] pentadecane)	[380, 381]
<i>Glinus lotoides</i> L.	“Mettete” Hairy carpet -weed	Seeds	Cameroon Ethiopia, Sudan, Uganda, Egypt.	Used to treat cardiovascular and gastrointestinal system.	Three flavonoids: apigenin-7-O-glucoside, isovitexin, and luteolin-7-O-glucoside Three isoflavonoids: 5,7,2',4'-tetrahydroxy-6-(3,3-dimethylallyl)isoflavone, 5,7,4'-trihydroxy-6,3'-di-(3,3-dimethylallyl)isoflavone, and 5,7,2',4'-tetrahydroxy-6,3'-di-(3,3-dimethylallyl)isoflavone.	[290, 382-386]
<i>G. oppositifolius</i> (L.) Aug. DC.	Balasa	Whole plant	Mali	Antioxidant and radical scavenging abilities.	kaempferol 3-O-galactopyranoside	[387, 388]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Aloaceae</b>						
<i>Aloe daviflora</i> Burch.	Kraal aloe		South Africa	Free radical scavenging and moderate inhibition in lipid peroxidation. Used as a purgative.	Not identified	[35]
<i>A. maculata</i> Forssk. (=A. saponaria)	“Yellow Form” Tiger Aloe, Soap Aloe		South Africa	Free radical scavenging and moderate inhibition in lipid peroxidation. Used as a purgative.	Not identified	[35]
<i>A. thraskii</i> Baker	Dune aloe		South Africa	Free radical scavenging and moderate inhibition in lipid peroxidation. Used as a purgative.	Not identified	[35]
<b>Anacardiaceae</b>						
<i>Sclerocarya birrea</i> (A.Rich.) Hochst	Marula	Stem-bark		Anti-inflammatory activity. Used to treat diabetes, tonsillitis, snake bite and also diarrhoea.	Not identified	[389]
<b>Annonaceae</b>						
<i>Eenantha chlorantha</i> Oliver	Erenbabogo, Mfisi Muamba	Root, stem-bark	Nigeria	Anti-inflammatory activity. Used to treat ulcers and leprosy spots wounds. Bark sap is taken as decoction against diarrhoea.	Not identified	[390–393]
<i>Uvaria ejzelli Sc. Elliot</i>	Pareho-houon, Bahie oulin	Leaves, roots and stem-bark	Ivory Coast	Used as for its antiparasitic activity	Anthocyanins and other flavonoids	[394–396]
<i>U. chamae</i> P.Beauv.	Okandii Anweda tsoga	Stem, bark Leaves, root	Ivory Coast Nigeria	Used for its antiplasmoidal activity.	Polyphenols	[12, 397, 398]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Apocynaceae</b>						
<i>Picralima nitida</i> Th. & H. Dur.	Ghana: Kpetepeteiso, Kanwini, Kanwini Cameroun: <i>motoko-toko</i>	Seeds Stem-bark	Ghana	Anti-inflammatory activity. Used for its analgesic and anti-inflammatory properties.	Not identified	[168, 399–402]
<i>Rauvolfia vomitoria</i> Afzel.	Asofeyeje, adapopo Mwanje	Root-bark	Ghana	Anti-inflammatory activity. Used for its analgesic, antipyretic and anti-inflammatory activities. Also to treat scabies, high blood pressure, fever and snakebites.	Not identified	[56]
<b>Araliaceae</b>						
<i>Cussonia barteri</i> Seem.	Cabbage tree	Leaves Roots	Nigeria, Mali	Antioxidant and radical scavenging abilities. Inhibitory activity on 5-lipoxygenase and cyclooxygenase-1.	Not identified	[357, 403]
<b>Arecaceae</b>						
<i>Hyphaene thebaica</i> Mart.	Not signalized	Shell	Niger	Antioxidant activity	Not identified	[11]
<b>Asclepiadaceae</b>						
<i>Calotropis procera</i> (Aiton) W.T.Aiton	African milk weed Sodom apple/Giant milkweed/ Swallow-wort/Auricula tree.	Latex	Ethiopia Sudan	Anti-inflammatory and antioxidant activities. Used to control dermal fungal infections and for pain relief. Latex used against scorpion stings and roots for jaundice.	Not identified	[404]
<i>Gongronema latifolium</i> Benth.	Not signalized	Leaves	Nigeria	Antioxidant activity	Not identified	[405–407]
<i>Leptadenia hastata</i> Deone.	Not signalized	Leaves	Niger	Antioxidant activity	Not identified	[11]
<i>Pachycarpus rigidus</i> E. Mey.	Not signalized	Bark	South Africa	Antioxidant activity. Used to treat pain in the joints	Not identified	[188]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Asparagaceae</b>						
<i>Asparagus virgatus</i> Baker Refug. Bot. (Saunders)	Broom asparagus	Bark	South Africa	Antioxidant activity Used to treat syphilis, antihelminthic	Not identified	[35]
<b>Asteraceae</b>						
<i>Ageratum conyzoides</i> L.	Inkuruba Herbe à bouc	Whole plant	Central Africa, Rwanda Ethiopia	Properties. Used to treat mastitis and urogenital infections and to dress wounds. Also as a gastroprotective.	Not identified	[12, 408, 409]
<i>Artemisia herba-alba</i>	Desert wormwood, shih Herbe à alouette	Aerial parts	Algeria, Tunisia, Israel, Morocco	Herbal tea from <i>A. herba-alba</i> has been used as analgesic, antibacterial, antispasmodic, and hemostatic agents in folk medicines	Camphor (17–33%), $\alpha$ -thujone (7–28%), and chrysanthene (4–19%)	[9]
<i>Artemisia judaica</i> L.	Wormwood	Leaves	Egypt	Used for gastrointestinal disorders	Flavonoids with antioxidant activities.	[410]
<i>Callilepis laureola</i> DC.	Ox-eye daisy, Impila	Tuber	South Africa	Antioxidant and radical scavenging activities. Used to induce fertility, impotence, tapeworm infestations but induces hepatic and renal tubular necrosis.	Not identified	[188, 411, 412]
<i>Psiadia punctulata</i> (DC) Vatke	Mwendathigo	Leaf exudate	Kenya, East Africa	Used to treat colds, fevers and abdominal pains.	Flavones: 5,7-dihydroxy-2',3',4',5'-tetramethoxyflavone, 5,4'-dihydroxy-7,2',3',5'-tetramethoxyflavone, 5,7,4'-trihydroxy-2',3',5'-trimethoxyflavone, 5-hydroxy-7,2',3',4',5'-pentamethoxyflavone and 5,7,3'-trihydroxy-2',4',5'-trimethoxyflavone.	[359, 413]
<i>Vernonia kotschyana</i> Sch. Bip. ex Walp.	Buaye	Leaves, roots	Mali	Anti-inflammatory activity. Used to treat gastritis, gastro duodenal ulcers, as an aid to ameliorate digestion and as a wound healing remedy. Immunomodulating activities.	Not identified	[187, 414]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Bignoniaceae</b>						
<i>Kigelia pinnata</i> DC.	Sausage tree, Cucumber tree	Root fruit	Egypt	Used as dressing for ulcers and used to treat rheumatism Anti-inflammatory activity	Naphthoquinones: kigelinone, isopinnatal, dehydro-alpha-lapachone, and lapachol and the phenylpropanoids: p-coumaric acid, ferulic acid (root), kigelinone and cafféic acid (fruits).	[415, 416]
<i>Tabebuia rosea</i> (Bertol.) DC.	Pink tecoma Pink trumpet tree	Leaves Stem-bark	Nigeria	Used to treat arthritis.	Tannins, flavonoids, alkaloids, quinones and traces of saponins	[107]
<i>Crescentia cujete</i> L.	Calabash Gourd tree	Leaves Stem-bark	Nigeria	Used as purgative and to treat coughs.	Tannins, flavonoids, alkaloids, quinones and traces of saponins	[107]
<b>Bombacaceae</b>						
<i>Bombax costatum</i> Pellegrin & Vuillet	Not signalized	Fruit	Niger	Antioxidant activity	Not identified	[11]
<b>Boraginaceae</b>						
<i>Heliotropium indicum</i> L.	Nonsikou	Leaves	Mali	Moderate antioxidant activity. Used for wound healing and for ocular infection.	Not identified	[417-419]
<b>Buddlejaceae</b>						
<i>Buddleja madagascariensis</i> Lam.	Butterfly-bush	Leaves	Egypt	Used to treat coughs, asthma, and bronchitis.	Flavonoids trglycosides: hesperetin and diosmetin 7-O (2'',6'' - di-O-alpha-L-rhamnopyranosyl)-beta-D-glucopyranosides	[420]
<b>Caesalpiniaceae</b>						
<i>Cassia fistula</i> L.	Golden shower tree	Fruit	Mauritius	Laxative.	Phenolics and flavonoids	[368]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Canellaceae</b>						
<i>Warburgia salutaris</i> (Bertol F.) Chiiov.	Pepper-bark tree Isibaha	Bark	South Africa	Antioxidant and radical scavenging activities. Used to treat coughs, stomach ulcers, malaria, rheumatism, liver and venereal diseases	Not identified	[188]
<i>W. ugandensis</i> Sprague	Fever tree	Stem-bark Leaves	Kenya Ethiopia	Used to treat stomach ache, chest pains, malaria, toothache and coughs.	Flavonol glycoside Kaempferol, kaempferol 3-rhamnoside, kaempferol 3-Rhamnoside-7,4'-digalactoside and Quercetin: 3-Rhamnosyl[1→6]glucosyl(1→2)glucoside]-7-rhamnoside, kaempferide 3-O-beta-xylosyl(1→2)-beta-glucoside, kaempferol 3-O-alpha-rhamnoside-7,4'-di-O-beta-galactoside, kaempferol 3,7,4'-tri-O-beta-glucoside, kaempferol 3-rutinoside, myricetin, quercetin 3-rhamnoside, kaempferol 3-arabinoside, quercetin 3'-glucoside, quercetin, kaempferol 3-rhamnoside-4'-galactoside, myricetin 3'-galactoside and kaempferol 3-glucoside.	[421-424]
<b>Capparaceae</b>						
<i>Boscia senegalensis</i> (Pers.) Lam. ex Poiret	Senegal Boscia	Fruit hull Roots and leaf	Mali Niger	Antioxidant activity. Used to treat diarrhoea, cholera, tachycardia, pectoral pain.	Not identified	[12]
<i>Gynandropsis gymnantha</i> Merr.	Not signalized	Leaves	Niger	Antioxidant activity	Not identified	[11]
<b>Celastraceae</b>						
<i>Solacia leptoclada</i> Tul.	Lemon rope	Root	South Africa	Antioxidant activity. Used as an aphrodisiac.	Not identified	[188]
<b>Chenopodiaceae</b>						
<i>Salsola somalensis</i> N.E.Br.	Dingetegna	Roots	Ethiopia	Used as taenicide.	Nine new isoflavones, 5,3'-dihydroxy-6,7,2'-trimethoxy isoflavone, 5,8,3'-trihydroxy-6,7,2'-dimethoxyisoflavone, 8,3'-dihydroxy-5,7,2'-trimethoxyisoflavone, 5,6,3'-trihydroxy-7,2'-dimethoxyisoflavone, 6,7,3'-trihydroxy-5,2'-dimethoxyisoflavone, 5,8,3'-trihydroxy-2'-methylenedioxyisoflavone, or 5,6,3'-trihydroxy-6,7-methylenedioxyisoflavone, 3'-hydroxy-5,6,7,2'-tetramethoxyisoflavone, 7,3'-dihydroxy-5,6,2'-trimethoxyisoflavone and 6,3'-dihydroxy-5,7,2'-trimethoxyisoflavone.	[425]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Clusiaceae</b>						
<i>Psotospurum guineense</i> Hochr.	Kardjakouma	Leaves	Mali	Antioxidant activity. Used as diuretic and febrifuge.	Not identified	
<b>Combretaceae</b>						
<i>Pteleopsis suberosa</i> Engl. & Diels.	Girga	Stem-bark	Mali	Antioxidant properties. Used to treat gastric and duodenal ulcers.	Not identified	[329, 426]
<b>Dioscoreaceae</b>						
<i>Dioscorea dumetorum</i> Th. Duret Schinz	Cluster yam African bitter yam Trifoliate yam	Tubers	Nigeria Tropical West Africa	Antioxidant and hypolipidemic activities. Used to treat diabetes.	Not identified	[152, 153, 427]
<b>Ebenaceae</b>						
<i>Diospyros abyssinica</i> (Hiern) F. White	Giant diospyros	Leaves, roots Root-bark	Mali	Radical scavengers and lipooxygenase inhibitors.	Not identified	[357]
<b>Euclea divinorum</b> Hiern						
	Diamond-leaved euclea Magic guarri	Roots	Ethiopia	Used to treat venereal diseases, chest pains, pneumonia, internal body pains, stomach-ache and diarrhea. Chewed roots ease toothache.	Flavonoids	[428]
<b>Euphorbiaceae</b>						
<i>Acalypha hispida</i> Burm. f.	Chenille plant Red-hot cattail	Leaves Flowers	Nigeria	Used as anti-bacterial agent.	Gallic acid and Quercetin 3-O-rutinoside and kaempferol 3-O-rutinoside	[228, 429]
<i>A. wilkesiana</i> Müll. Arg.	Copper leaf	Leaves	Nigeria	Used to treat ailments of microbial origin	The main anthocyanin is the known cyanidin 3-O-(2-O-galloyl)galactose, but a minor pigment (5%) is the new cyanidin Cy 3-O-(2-O-galloyl)-6-O-rhamnosylgalactoside	[430]
<i>Croton gratissimus</i> Burch.	Lavender fever-berry	Bark	South Africa	Used as purgative for abdominal disorders, fever. The charred and powdered bark is used to treat bleeding gums	Gallic acid and Quercetin 3-O-rutinoside and kaempferol 3-O-rutinoside	
<i>Euphorbia hirta</i> L.	Kasandasanda Ufu idire	Whole plant Leaves	Ethiopia	Used to treat diarrhoea and asthma.	Flavonoid: quercitrin (1→3)galactoside)	[12, 431–433]
<b>Fabaceae</b>						
<i>Acacia caffra</i> (Thunb.) Wild.	Hook-thorn Cat-thorn	Bark	South Africa	Used to treat diarrhoea and as emetics.	Proanthocyanidins: oritin-(4alpha→5)-epioritin-4beta-ol, ent-epioritin-(4alpha→5)-epioritin-4beta-ol and epioritin-(4beta→5)-epioritin-4alpha-ol and ent-ortin-(4beta→5)-epioritin-4alpha-ol.	[434–436]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>A. galpinii</i> Burtt Davy.	Monkey-thorn	Bark	South Africa	Used to treat diarrhoea.	Proanthocyanidins: oritin-(4alpha—>5)-epioritin-4beta-ol, ent-epioritin-(4alpha—>5)-epioritin-4beta-ol and epioritin-(4beta—>5)-epioritin-4alpha-ol and ent-oritin-(4beta—>5)-epioritin-4alpha-ol.	[434, 435]
<i>Afzelia bella</i> Harms	Pretty Afzelia	Stem-bark	Ivory Coast	Used to treat skin diseases and cough.	An acylated dihydroflavonol glycoside identified as 2R,3R-trans-aramadendrin 7-O-beta-D-glucopyranoside-6''-(4''-hydroxy-2''-m ethylene flavonoids:butanoate), along with five known flavonoids and the lignan glycoside (+)-isolariciresinol 9-O-xylloside.	[437]
<i>Bolusanthus speciosus</i> Harms	Tre Wisteria	Root Stem-bark	South Africa, Botswana, Mozambique, Zimbabwe, Zambia.	Used to treat abdominal pains, emetism and tuberculosis.	Three new flavonoids from the root: 5,7,4'-trihydroxy-6-[1-hydroxy-2-methylbuten-2-y]isoflavone (isogancanon C), 7,2'-dihydroxy-4'-methoxyisoflav-3-ene (bolusanthin III), 6,6'-dihydroxy-4'-methoxy-2-arylbenzofuran (bolusanthin IV) in addition to eight known derrone, medicarpin, genistein, wighteone, lupiwighteone, gancaonin C, 7-hydroxy-4'-methoxyisoflavone and 7,3'-dihydroxy-4'-methoxyisoflavone flavonoids 2R,3R-Aromadendrin 7-(6-[4-hydroxy-2-methylenebutanyl]glucoside). Two new isoflavonoids from the combined ethyl acetate/methanolic extracts of the stem bark of Bolusanthus speciosus have been established as 4,7,2'-trihydroxy-4', methoxyisoflavanol (1) and 5,7,3',4'-tetrahydroxy-5'-(2-epoxy-3-methylbutyl)isoflavone (2). Five other known isoflavonoids, 5,7,3',-trihydroxy-4'-methoxy-5', $\gamma$ , $\gamma$ -dimethylallylisoflavone, 5,7,2',-trihydroxy-4'-methoxy-5',-di(y, $\gamma$ -dimethylallylisoflavone, 5,7,2',4'-tetrahydroxy-8,5',-di(y, $\gamma$ -dimethylallylisoflavone, 5,7,2',4'-tetrahydroxy-8,3',-di(y, $\gamma$ -dimethylallylisoflavone), and derrone.	[67, 358, 438]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Crotalaria lanceolata</i> E. Mey.	Lanceleaf rattlebox	Root	South Africa	Antioxidant activity. Used to treat coughs.	Not identified	[188]
<i>Derris trifoliata</i> Lour.	Common derris	Root-bark. Stem-bark. Seeds.	Kenya	Used for prevention of cancer. Entire plant is used as stimulant, antispasmodic. Bark is used as an alternative in rheumatism.	An isoflavanoid derivative, named 7a-O-methyldeguelol, a modified rotenoid with an open ring-C, representing a new sub-class of isoflavonoids (the sub-class is here named as rotenoloid). In addition, the known rotenoids, rotenone, deguelin and alpha-toxicarol. In addition, two unusual rotenoid derivatives, a rotenoloid (named 7a-O-methyl-12a-hydroxydeguelol) and a spirohomooxarotenoid (named spiro-13-homo-13-oxaelliptone).	[438-441]
<i>Entada africana</i> Guill. & Perr.	Samanere	Leaves	Mali Niger	Protective against carbon tetrachloride-induced liver damage. Used to treat fever and various respiratory diseases.	Not identified	[329, 357, 442, 443]
<i>Erythrina abyssinica</i> Lam.	Red hot poker tree	Stem bark Root bark	Kenya	Used to treat malaria.	New isoflav-3-ene [7,4'-dihydroxy-2',5'-dimethoxyisoflav-3-ene] in addition to the known compounds erycristagallin, licoagrochalcone A, octacosyl ferulate and triacontyl 4-hydroxycinnamate were identified. A new chalcone, 2',3,4,4'-tetrahydroxy-5-prenylchalcone (trivial name 5-prenylbutein) and a new flavanone, 4',7-dihydroxy-3'-methoxy-5'-prenyflavanone (trivial name, 5-deoxyabysinii II) along with known flavonoids	[444, 445]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>E. burttii</i> Baker f.	Not signalized	Stem-bark Root-bark	Kenya	Used as antifungal and antibacterial agent.	Two new flavanones: 5,7-dihydroxy-4'-methoxy-3',5'-di-(3-methylbut-2-enyl)flavanone (trivial name, abyssinone V-4'-methyl ether) and 5,7-dihydroxy-4'-methoxy-3',-(3-hydroxy-3-methylbut-1-enyl)-5'-(3-methylbut-2-enyl)favanone (trivial name, burttinone). A new isoflavone, 5,2',4'-trihydroxy-7-methoxy-6-(3-methylbut-2-enyl)isoflavone (trivial name, 7-O-methylfoluteone) and a new flavanone, 5,7-dihydroxy-4'-methoxy-3',-(3-methylbutadienyl)-5'-(3-methylbut-2-enyl)favanone, 3 isoflavanoids (8-prenylluteone, 3-O-methylcalopocarpin and genistein)	[446-449]
<i>E. eriotticha</i> Harms.	Not signalized	Root-bark	Cameroon	Anti-microbial activity	Three isoflav-3-enes, 7,4'-dihydroxy-2',-methoxy-6-(1',1'-dimethylallyl)isoflav-3-ene (trivial name, burttinol-A), 4',-hydroxy-2',-methoxy-2'',2''-dimethylpyrano[5',6'':8,7]isoflav-3-ene (trivial name, burttinol-B), 7,4'-dihydroxy-2',-methoxy-8-(3'',3'')-dimethylallyl)isoflav-3-ene (trivial name, burttinol-C), and 2-arylbenzofuran, 6,4'-dihydroxy-2',-methoxy-5-(1'',1''-dimethylallyl)-2-arylbenzofuran (trivial name, burttinol-D).	[450, 451]
<i>E. saculeuxii</i> Hua	Kinyarwanda	Bark	Kenya	Used to treat fever, malaria and leprosy.	Two new isoflavonones (R)-5,7-dihydroxy-2',4',5'-trimethoxysoflavone (trivial name, (R)-2,3-dihydro-7-demethylrobustigenin) and (R)-5-hydroxy-2',4',5'-trimethoxy-2'',2''-dimethylpyrano[5',6'':6,7]isoflavan one (trivial name, (R)-saccone)	[452, 453]
<i>Millettia ferruginea</i> (Hochst.) Baker	Birbira Sotallo Sari	Bark	Ethiopia	Used for skin disorders.	O-Geranylated and O-prenylated flavonoids, C-prenylated isoflavones Geranylated and prenylated flavonoids	[199]
<i>M. dura</i> Dunn.	Runyankore Ummuyogoro	Stem-bark	Rwanda Uganda	Used for blood parasitism	Flavonoids: A new isoflavone (7,3',-dimethoxy-4',5',-methylenedioxyisoflavone) and three known isoflavones [isoerythrin A 4-(3-methylbut-2-enyl) ether, isojamaicin and nordurlettone].	[454, 455]
<i>Ostryoderris stuhlmannii</i> (Taub.) Dunn ex Harms	Mnyinga	Leaves	Mali	Antioxidant activity. Used to treat painful menstruation, peritonitis, gastritis, colitis and gingivitis.	Not identified	[357]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Pliostigma reticulatum</i> (DC.) Hochst	Kalga	Leaves Bark	Nigeria	High antioxidant activity. Used to treat wounds, bronchitis, malaria, sterility (leaves) and diarrhoea and dysentery (bark).	Not identified	[240]
<i>Sesbania pachycarpa</i> DC.	Not signalized	Leaves	Niger	Antioxidant activity	Not identified	[11]
<i>Tephrosia polystyphlla</i> (Chiiov.) J.B. Gillett	Hoary pea	Aerial part	Kenya		Flavonoids	[456]
<i>T. deflexa</i> Baker	Hoary pea	Aerial part	Senegal		Flavonoids: Rutin 1 - quercetine 3-O- $\alpha$ -L-rhamnopyranosyl (1-6) glucopyranose - and morin 2 - 3,5,7,2',4'-pentahydroxyflavone.	[457]
<i>T. albifoliolis</i> A.Nongonierna & T.Sarr	Hoary pea	Aerial part	Senegal		Flavonoids: Rutin 1 - quercetine 3-O- $\alpha$ -L-rhamnopyranosyl (1-6) glucopyranose - and morin 2 - 3,5,7,2',4'-pentahydroxyflavone.	[457]
<i>Taverniera abyssinica</i> A. Rich.	Dingetegna	Root	Ethiopia	Used to treat fever, discomfort and pain, stomach ache.	Four isoflavonoids	[290, 458, 459]
<i>Flacourtiaceae</i>						
<i>Flacouritia flavescentis</i> Willd.	Not signalized	Leaves	Mali	Antioxidant activity.	Not identified	[357]
<i>Geraniaceae</i>						
<i>Pelargonium reniforme</i> Spreng.	Xhosa (Umckaloabo)	Root	Southern Africa	Used to treat liver disorders, laxative, purgative, cancer, and pulmonary disorders	Polyphenols: catechol (3/4' dihydroxy) element in the B-ring, which possesses higher antioxidant activity than ascorbic acid.	[362, 460, 461]
<i>Gunneraceae</i>						
<i>Gunnera perpensa</i> L.	River pumpkin Ugbollo	Leaves and stem.	South Africa	Decreased lucigenin enhanced chemiluminescence. Used to treat wounds and psoriasis.	Not identified	[21, 462]
<i>Irvingiaceae</i>						
<i>Irvingia gabonensis</i> (Aubry-Lecomte ex O'Rorke) Baill.	Bush mango Ono	Seeds	Nigeria Cameroon	Antioxidant activity. Used as laxative and for stomach and kidney pain. Shown to lower total cholesterol.	Not identified	[12, 313, 463]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Lamiaceae</b>						
<i>Leonotis leonurus</i> (L.)R.Br.	Wild dagga	Leaves	South Africa	Anti-inflammatory properties. Used to treat headaches, dysentery, coughs and colds.	Not identified	[13]
<i>Salvia stenophylla</i> Burch. ex Benth.	Sage	Leaves	South Africa	Solvent extracts: antioxidant activity but poor anti-inflammatory properties. Essential oils: anti-inflammatory activity but poor anti-oxidant activity. Used against fever and digestive disorders.	Not identified	[360]
<i>S. repens</i> Burch. ex Benth.	Not signalized	Leaves	South Africa	Solvent extracts: antioxidant activity but poor anti-inflammatory properties. Essential oils: anti-inflammatory activity but poor anti-oxidant activity. Used for fevers and digestive disorders.	Not identified	[360]
<i>S. runcinata</i> L.f.	Not signalized	Leaves	South Africa	Solvent extracts: antioxidant activity but poor anti-inflammatory properties. Essential oils: anti-inflammatory activity but poor anti-oxidant activity. Used against fever and digestive disorders.	Not identified	[360]
<b>Loranthaceae</b>						
<i>Tapinanthus globiferus</i> Tiegh.	Not signalized	Leaves	Niger	Antioxidant activity	Not identified	[11]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Malvaceae</b>						
	English: baobab, Afrikaans: kremetart, Hausa: kuka, Sotho: seboi, Tswana: mowana, Tsonga: shimuwu, Venda: muvhuyu, Arabic: tabladi	Leaves, root, bark and fruits	All over Africa, but limited trees in Central Africa	Antioxidant, analgesic and anti-inflammatory properties of extracts	L-ascorbic acid	[36, 464]
<b>Mimosaceae</b>						
	Albizia lebbeck (L.) Benth.	East Indian walnut, frywood, koko, lebbek, lebbek tree, rain tree, raom tree, silver raintree, siris rain tree, siris tree, soros-tree, woman's tongue.	Leaves and bark	Egypt	Used to treat asthma and skin disorders (bark) and eye diseases and dysentery (leaves)	Two new tri-O-glycoside flavonols: kaempferol and quercetin 3-O-alpha-rhamnopyranosyl(1→6)-beta- glucopyranosyl(1→6)-beta- galactopyranosides [465]
<b>Moraceae</b>						
<i>Dorstenia angusticornis</i> Engl.	Not signalized	Twigs	Cameroon	Used for snakebite and to treat infection, rheumatism, headache, cough and stomach pain.	Two novel diprenylated chalcones: 3,5'-di-(2-hydroxy-3-methylbut-3-enyl)-4,2',4'- trihydroxychalcone, 3, 4-(2,2-dimethylpyrano)-3'-(2-hydroxy-3- methylbut-3-enyl)-2',4'-dihydroxychalcone and the known stipulin. 3-(2-Hydroxy-3-methylbut-3-enyl)-5'-(3,3- dimethylallyl)-4,2',4'-trihydroxy chalcone and the known compounds: gancanin Q, paratocarpins C, F, and lupeol	[67, 278]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>D. dinklagei</i> Engl.	Not signalized	Twigs	Cameroon	Used for snakebite and to treat infection, rheumatism, headache, cough and stomach pain.	Three prenylated flavonoids, dinklagins A, B and C identified, respectively, as (dinklagin B): (+)-5,4',5'- $\xi$ -Trihydroxy-6'',6''-dimethylidihydropyranol[2'',3'':7,6]flavone. (dinklagin C): (+)-6-(2 $\xi$ -Hydroxy-3-methyl-3-buteny)-5,74'-trihydroxyflavone (-)-6-(3,3-dimethylallyl)-7-hydroxy-6''',6'''-dimethylchromeno(4',3',2'',3'')-flavanone, (+)-5,4',5'- $\xi$ -trihydroxy-6'',6''-dimethylchromano-(7,6,2'',3'')-flavone and (+)-6-(2 $\xi$ -hydroxy-3-methyl-3-buteny)-5,74'-trihydroxyflavone. 6-prenyliapigenin, 4-hydroxylonchocarpin, stipulin and 5,4'-dihydroxy-6'',6''-dimethylchromano-(7,6,2'',3'')-flavone.	[67, 226]
<i>D. elliptica</i> Bur.	Not signalized	Twigs	Botswana	Used to treat eye infection.	Monoprenylated flavan	[466]
<i>D. Kameruniana</i> . Engl.	Not signalized	Leaves	Botswana	Used for snakebite and to treat infection, rheumatism, headache, cough and stomach pain.	Two novel flavonoids: 6,7-(2,2-dimethylchromano)-5,4'-dihydroxyflavone and 3,4,4',5'-bis-(2,2-dimethylchromano)-2'-hydroxychalcone together with the known 6-(3-methylbut-2-enyl)apigenin and two chalcones (E)-1-[2,4-dihydroxy-3-[3-methylbut-2-enyl]phenyl]-3-[4-hydroxyphenyl]-prop-2-en-1-one and (E)-[2,4-dihydroxy-5-[3-methylbut-2-enyl]phenyl]-3-[4-hydroxy-3-[3-methylbut-2-enyl]phenyl]-prop-2-en-1-one.	[467]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>D. prorepens</i> Engl.	Not signalized	Twigs	Botswana	Used for snakebite and to treat infection, rheumatism, headache, cough and stomach pain.	Digeranylated chalcone, 5,3'-(3,7-dimethyl-2,6-octadienyl)-3,4,2',4'-tetrahydroxychalcone. 4-Hydroxylonchocarpin Chalcone: 3,4,2',4'-Tetrahydroxy-5,3'-digeranylchalcone	[67, 468]
<i>D. poinsettifolia</i> Engl.	Not signalized	Twigs	Botswana	Used for snakebite and to treat infection, rheumatism, headache, cough and stomach pain.	Grenylated and prenylated flavonoids. In addition, the flavone 5,7,4-trihydroxy-8-prenylflavone (licoflavone C), the chalcones 4,2',4'-trihydroxy-3'-prenylchalcone (isobavachromene) and isobavachromene, the triterpene butyrospermol, and the carotenoid lutein.	[67, 206, 289]
<i>D. zenkeri</i> Engl.	Not signalized	Twigs	Botswana	Used for snakebite and to treat infection, rheumatism, headache, cough and stomach pain.	3',4'-(3-hydroxy-2,2-dimethylidihydropyrano)-4,2'-dihydroxychalcone and a bichalcone. 4-Hydroxylonchocarpin. P-hydroxybenzaldehyde, dorsmanin A, 4,2',4'-trihydroxychalcone and 4,2',4'-trihydroxy-3'-prenylchalcone Chalcones: 4,2',5''-Trihydroxy-6'',6''-dimethylidihydropyranol[2'',3'':4',3']chalcone	[67, 468]
<b>Moringaceae</b>						
<i>Moringa oleifera</i> Lam.	Horse-radish tree Drumstick Moringo Zakalanda	Root	West Africa Zimbabwe	Anti-inflammatory activity. Used as aphrodisiac and to treat asthma, gout and rheumatism.	Not identified	[469]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<i>Myriaceae</i>						
<i>Eucalyptus camaldulensis</i> Dehnh.	Not signalized	Leaves	Egypt	Antioxidant activity	Not identified	[470]
<i>Polygonaceae</i>						
<i>Polygonum senegalense</i> Meisn.	Fotsimbarin'akoholahy	Leaves	Madagascar	Flavonoids: quercetin, kaempferol and luteolin and their glycosides such as dihydrochalcone glucoside and quercetin glycosides.	[413, 471]	
<i>Rumex abyssinicus</i> Jacq.	Mekmeko	Leaves	N. Africa - Ethiopia	Anti-inflammatory properties Used to treat itching, skin eczema and leprosy.	Flavonoids.	[337, 472]
<i>R. nervosus</i> Vahl, Dengogo	Alcgango Dengogo	Leaves	Ethiopia	Anti-inflammatory properties Used to treat acne, wounds, eczema, typhus and as an ophthalmic antiseptic.	Not identified	[337]
<i>Rubiaceae</i>						
<i>Nauclea latifolia</i> Smith	Pin Cushion Tree Iguyaà	Leaves and root	Nigeria	Used as anthelmintic and to treat malaria, fever, stomachache and liver diseases.	Proanthocyanidins.	[12, 58, 473-475]
<i>Solanaceae</i>						
<i>Datura stramonium</i> L.	Thorn-apple rwiziringa	Seeds	South Africa	Antioxidant activity. Used to treat asthma, headaches and wounds.	Not identified	[188]
<i>Tiliaceae</i>						
<i>Grewia occidentalis</i> L.	Cross-berry Four-corner	Bark	South Africa	Antioxidant activity. Used to treat bladder ailments, wounds, impotence and sterility, and to help in childbirth.	Not identified	[188]

TABLE 2: Continued.

Family and plant name	Vernacular name	Plant part	Country/area	Medicinal use and/or experimental validation	Compounds isolated	Reference
<b>Vahliaeae</b>						
<i>Vahlia capensis</i> (L.f.) Thunb.	Vahlia of the Cape	Zimbabwe		Used to treat bacterial infections.	Kaempferol, quercetin, afzelin, astragalin, queritrin, isoquercitrin, rutin, gallic acid, chiro-inositol, dulcitol, and a novel biflavonoid, VC-15B (vahlia biflavone)	[475]
<b>Vitaceae</b>						
<i>Cyphostemma natalitium</i> (Syzs.) J.v. d. Merwe	Tick-berry bush	Root	South Africa	Anti-inflammatory and anti-microbial agents with significant inhibition of COX-1	Not identified	[374]
<i>Rhoisssus digitata</i> Gilg. & Brandt						
	Wilde patataf	Roots, stems and leaves	South Africa	At high concentrations possessed some prooxidative properties. Anti-inflammatory and anti-microbial agents with significant inhibition of COX-1. Used to facilitate delivery.	Not identified	[364, 374]
<i>R. rhomboidea</i> (E. Meyer ex Harvey) Planchon						
	Glossy forest grape	Roots, stems and leaves	South Africa Mozambique	Radical scavenging activity, inhibitory effect on xanthine oxidase activity, prevention of lipid peroxidation and damage to DNA and ability to chelate iron. Anti-inflammatory through inhibition of COX-1.	Not identified	[364, 374]
<i>R. tomentosa</i> (Lam.) Wild & R.B.Drum.						
	Wild grape Forest Grape, Monkey rope,	Roots, stems and leaves	South Africa	Antioxidant and anti-inflammatory activities. Anti-inflammatory through inhibition of COX-1. Used to facilitate delivery.	Not identified	[364, 374]
<i>R. tridentata</i> (L.f.) Wild & Drum.						
	Bitter grape Bushmans grape Isinwazi	Roots, stems and leaves	South Africa : Venda	Radical scavenging activity, inhibitory effect on xanthine oxidase activity, prevention of lipid peroxidation and damage to DNA and ability to chelate iron. Anti-inflammatory through inhibition of COX-1. Used to treat colds, infertility and stomach ailments.	Not identified	[364, 374, 476]

and degranulation [112, 487], antiatherosclerosis effects [486], and pain assessment in experimental animals [117].

The effect of the medicinal plants on the induction or inhibition of drug metabolizing enzymes was also studied in animals. The effect of the aqueous extract of *Thonningia sanguinea* on 7-ethoxyresorufin O-deethylase (EROD, CYP1A1), 7-pentoxyresorufin O-dealkylase (PROD, CYP2B1/2), 7-methoxyresorufin O-demethylase (MROD, CYP1A2), aniline hydroxylase (aniline, CYP2E1), *p*-nitrophenol hydroxylase (PNPH, CYP2E1), and erythromycin N-demethylase (ERDM, CYP3A1) in rat liver was found to selectively modulate CYP isoenzymes [100] and suppress CYP3A2 and CYP1A2 gene expression [101].

### **3. Compounds Isolated from African Medicinal Plant Extracts with Confirmed Antioxidant Activities**

Several medicinal plant extracts were studied at research centres in African countries for their antioxidant properties. The major findings of these investigations have indicated that, in addition to known antioxidant compounds such as ascorbic acid in the seeds of *Parkia biglobosa* [204] and fruits/pulp of *Adansonia digitata* [369], alpha-tocopherol in methanol extracts of the stems of *Secamone afzelii* [62] or from the seeds [38] and methanol extracts of leaves of *Amaranthus caudatus* [39], and apigenin and luteolin in aerial parts of *Bulbine capitata* [66], several other antioxidant compounds were identified. Although known antioxidant compounds such as ascorbic acid have been confirmed to promote wound healing, not all the newly identified compounds have been tested for such activity [488–491].

The identified compounds included mainly flavonoids such as flavones and flavonols, flavone and flavonol glycosides, chalcones and dihydrochalcones, and flavonones, although some anthocyanins, proanthocyanidins, and anthrones were also isolated with antioxidant properties. A wide range of plant extracts investigated have been shown to contain flavonoids. *Dorstenia* species are rich in flavonoids some of which are unique to this genus [67, 205], namely, prenylated flavonoids as found in *Dorstenia kameruniana* and twigs of *D. mannii* [206, 207]. Earlier studies have shown that prenylated flavonoids had antioxidant properties, which protected human LDL from oxidation [208]. Those isolated from African medicinal plant extracts were also tested and their antioxidant properties confirmed. The antioxidant activities of three prenylated flavonoids from *D. mannii* (6,8-diprenyleriodictyol, dorsmanin C, 7,8-(2,2-dimethylchromeno)-6-geranyl-3,5,3',4'-tetrahydroxyflavonol and dorsmanin F, (+)-7,8-[2''-(1-hydroxy-1-methylethyl)-dihydrofuran]-6-prenyl-5,3',4'-trihydroxyflavanone) against LDL oxidation and also their free radical scavenging activity have been indicated [187]. Similarly, a diprenylated chalcone, Bartericin A, present in *D. barteri* leaf and twig extracts was shown to have potent antioxidant properties. It was found that this and other prenylated and geranylated chalcones were as active as the prenylated flavones and

may account for the anti-inflammatory action of these extracts [276]. Free radical scavenging activity was also confirmed for prenylated anthronoids isolated from the stem-bark of *Harungana madagascariensis* [121] and for proanthocyanidins isolated from the bark of *Burkea africana* [175]. The anti-inflammatory and antioxidant activities of kolaviron, a biflavonoid isolated from a *Garcinia kola* seed extract to scavenge free radicals, which protect against lipid peroxidation and H<sub>2</sub>O<sub>2</sub>-induced DNA strand breaks and oxidized bases, were also reported [114, 116–119, 209]. In addition, the ability of free radical scavenging activity and ability to inhibit lipid peroxidation of Thonningianin A and Thonningianin B, ellagitannins, isolated from *Thonningia sanguinea* have been shown [99, 366]. The anti-inflammatory ability of Griffonianone D ((7E)-(6'',7''-dihydroxy-3'',7''-dimethyoct-2''-enyl)oxy-4'-methoxyisoflavone), an isoflavone present in *Millettia griffoniana*, has been established [195]. Prenylated anthronoids, harunmadagascarins A (8,9-dihydroxy-4,4-bis-(3,3-dimethylallyl)-6-methyl-2,3-(2,2-dimethylpyrano)anthrone and B (8,9-dihydroxy-4,4,5-tris-(3,3-dimethylallyl)-6-methyl-2,3-(2,2-dimethylpyrano)anthrone), harunganol B, and harungin anthrone from the stem-bark of *Harungana madagascariensis* have exhibited significant antioxidant activity [121]. Saponins and isofuranonaphthoquinones isolated from different medicinal plant extracts showed antioxidant properties and include the saponin, Balanin 1 (3β,12β,14β,16β) cholest-5-ene-3,16-diyl bis (β-d-glucopyranoside)-12-sulphate, sterol sulfonated, Balanin 2 (3β,20S,22R,25R)-26-hydroxy-22-acetoxyfurost-5-en-3-yl-rhamnopyranosyl-(1→2)-glucopyranoside, and a furostanol saponin isolated from *Balanites aegyptiaca* [104]. Isofuranonaphthoquinones isolated from the roots of *Bulbine capitata*, 5,8-dihydroxy-1-tigloylmethylnaphtho[2,3-c]furan-4,9-dione, 1-acetoxymethyl-8-hydroxynaphtho[2,3-c]furan-4,9-dione, and 1-acetoxymethyl-5,8-dihydroxynaphtho[2,3-c]furan-4,9-dione possess antioxidant activities [68]. Though none of these antioxidant compounds has been directly assessed for wound healing potential, the enhanced wound closure observed with treatment of prenylated flavonoids such as genistein [492] and the demonstrated effect of chalcones on the inflammation process [493] attest to the potential of isolated antioxidants in wound management.

### **4. Crude Extracts of African Medicinal Plants with Confirmed Antioxidant Activities**

The antioxidant properties of a larger proportion of African medicinal plants listed in Tables 1 and 2 were tested using either aqueous or organic plant extracts. After confirming antioxidant properties, a correlation was proposed between this property and the general groups of antioxidant compounds that are present in these extracts. No further attempts were made to isolate the specific compounds that may have contributed towards this property. Flavonoids in *Aloe barbadensis* [32], chromone glycosides in *A. claviflora* [35], essential oils in *Artemisia abyssinica*, and *Juniperus procera* [79] as well as *Helichrysum dasyanthum*, *H. felinum*, *H.*

*excisum*, and *H. petiolare* [94], proanthocyanidins in *Burkea africana* bark [175], polyphenols in extracts of *Crataegus monogyna* [321], saponins, and alkaloids in extracts of *Leucosidea sericea* [210, 211] are all considered as major compounds that have contributed to the antioxidant properties of these plants. Reports on a number of *Barleria* species, which includes *B. albostellata*, *B. greenii*, and *B. prionitis*, have indicated their anti-inflammatory [212] and antioxidant capacities [213]. Unlike the isolated compounds, most of the plants listed for possessing antioxidant activity, including extracts of *Agerantum conyzoides*, *Euphorbia hirta*, *Kigelia africana*, and *Nauclea latifolia*, have been shown to possess wound healing ability [494–496].

Furthermore, studies have focused on screening a vast number of plants, used in a specific region, so as to determine their antioxidant properties, Mali [357], South Africa [19, 188, 267, 364], Cameroon [182, 313], Algeria [85], Ghana [98], Burkina Faso [266], Madagascar [23], and Mauritius [293], and anti-inflammatory properties, South Africa [168, 264, 374, 376] and West Africa [400].

## 5. Discussion and Conclusion

The use of traditional herbal remedies as alternative medicine plays a significant role in Africa since it features extensively in primary health care. The search for natural antioxidants, especially from plant sources, as a potential intervention for treatment of free radical mediated diseases is an important research field, especially for those in developing countries. Many polyphenols, including phenolic acids, flavonoids (anthocyanins and anthoxanthins), tannins, and lignans, are known to act as antioxidants and protect against various pathological conditions such as coronary artery disease and wounds, in addition to their anti-inflammatory, antimicrobial, and anticancer activities [214–216].

Flavonoids are a large group of compounds containing several hydroxyl groups on their ring structures and include isoflavonoids and isoflavanoid glycosides, flavones, and flavone glycosides, flavonols and flavonol glycosides, anthocyanins, chalcones and dihydrochalcones, aurones, flavonones and dihydroflavonols, and flavans and biflavonyls. To date, approximately 9000 different flavonoids have been identified from plant sources [217]. Great interest has been dedicated to the antioxidant properties of flavonoids that may function as potent free radical scavengers, reducing agents, and protectors against peroxidation of lipids [208, 218]. Reviews have been published documenting numerous studies on antioxidant efficacy of flavonoids and phenolic compounds as well as on the relationship between their antioxidant activities, as hydrogen donating free radical scavengers, in relation to their chemical structures. The importance of the unsaturation in the C ring of quercetin compared to catechin in the increased antioxidant activity of the former has been presented [216, 219–223]. Also, the importance of the position and number of hydroxyl groups on the phenolic rings in increasing or decreasing the antioxidant properties of these compounds has been emphasized [216, 219–223].

Although many flavonoids have been isolated from different African medicinal plant extracts, the

structure-activity relationship of these compounds has not yet been investigated. Recent studies have also shown that some flavonoids are modulators of proinflammatory gene expression, thus leading to the attenuation of the inflammatory response [224]. Examples of these include the lipophilic flavones and flavonols 5,7-dihydroxy-2',3',4',5'-tetramethoxyflavone, 5,4'-dihydroxy-7,2',3',5'-tetramethoxyflavone, and 5,7,4'-trihydroxy-2',3',5'-trimethoxyflavone isolated from *Psiadia punctulata* [225] and Dinklagin B and C isolated from *Dorstenia dinklagei* [226]. Isolated flavone and flavonol glycosides include kaempferide 3-O-beta-xylosyl (1→2)-beta-glucoside, kaempferol 3-O-alpha-rhamnoside-7,4'-di-O-beta-galactoside, kaempferol 3,7,4'-tri-O-beta-glucoside and quercetin 3-O-[alpha-rhamnosyl (1→6)] [beta-glucosyl (1→2)]-beta-glucoside-7-O-alpha-rhamnoside from *Warburgia ugandensis*, and quercetin-7,4'-disulphate from *Alchornea laxiflora* [159]. Flavanones and dihydroflavonols include dorsmanin I and J and epidorsmanin F and G isolated from *Dorstenia mannii* [227] and Dinklagins A, isolated from the twigs of *Dorstenia dinklagei* [226] and two flavones isolated from the twigs of *Eriosema robustum* [182] and 1 $\alpha$ ,3 $\beta$ -dihydroxy-12-oleanen-29-oic (1), 1-hydroxy-12-olean-30-oic acid (2), 3,30-dihydroxyl-12-oleanen-22-one (3), and 1,3,24-trihydroxyl-12-oleanen-29-oic acid (4), a new pentacyclic triterpenoid (1 $\alpha$ , 23-dihydroxy-12-oleanen-29-oic acid-3 $\beta$ -O-2,4-di-acetyl-1-rhamnopyranoside) (5) from *Combretum imberbe* [138]. Anthocyanins isolated include the cyanidins 3-O-(2''-galloyl- $\beta$ -galactopyranoside) and 3-O-(2''-galloyl-6''-O- $\alpha$ -rhamnopyranosyl- $\beta$ -galactopyranoside) from *Acalypha hispida* [228] and cyanidin 3-O- $\beta$ -D-glucopyranoside and cyanidin 3-O-(2-O- $\beta$ -D-xylopyranosyl)- $\beta$ -D-glucopyranoside from *Hibiscus sabdariffa* [266]. When revising the literature, it became apparent that even though most of these medicinal plants and compounds have confirmed antioxidant activity, not many of them have been screened for wound healing potential. As there is an association between antioxidative therapy and wound healing, research in this direction is as imminent as it is important. Furthermore, structure-activity studies on the isolated compounds from African medicinal extracts will be of great interest.

Antioxidants may exert their protective effects via different mechanisms at different stages of the oxidation process. There are those that are able to inhibit the production of free radicals via their ability to chelate transition metal ions and those that are able to quench and stabilise free radicals [229, 230]. Additionally, they are further subdivided into categories according to their functions [230]. Such classification of the newly isolated antioxidant compounds from African medicinal plant extracts is warranted to better understand their antioxidant properties.

It should be noted that the antioxidant activity of the extracts and compounds listed in this review was mostly determined using either single assays or *in vitro* analysis. It is therefore possible that some of these extracts and compounds may not show antioxidant activity when alternative testing methods are used. Furthermore, although *in vivo* studies are encouraged, most studies cited used *in vitro* assays. As

antioxidant activity *in vitro* does not necessarily translate to activity *in vivo*, due to pharmacokinetic and pharmacodynamic processes that occurs *in vivo*, it is possible that samples may not be active when tested in animals. Activity of such samples should therefore be confirmed using animal models.

Additionally, attempts should be made to identify the compounds responsible for the proven antioxidant properties where not yet done, and in cases where they have been isolated, their wound healing properties should be investigated. If the activity of the compounds and plants identified in this review is confirmed *in vivo*, they could serve as viable sources for the treatment of wounds in future.

## Conflicts of Interest

The authors declare that they have no conflicts of interest.

## References

- [1] C. Dunnill, T. Patton, J. Brennan et al., "Reactive oxygen species (ROS) and wound healing: the functional role of ROS and emerging ROS-modulating technologies for augmentation of the healing process," *International Wound Journal*, vol. 12, no. 6, pp. 1–8, 2015.
- [2] E. Moasser, N. Azarpira, A. Ghorbani dalini, and B. Shirazi, "Paraoxonase 1 (PON1) gene polymorphism and haplotype analysis in type 2 diabetes mellitus: a case-control study in the south Iranian population," *International Journal of Diabetes in Developing Countries*, vol. 38, no. 1, pp. 62–68, 2018.
- [3] A. Benabbou, M. B. Khaled, and A. S. Alchalabi, "Evaluation of the Efficiency of Combined and Separated Antioxidant Supplementation of Vitamin C and E on Semen Parameters in Strepto-zotocin-Induced Diabetic Male Wistar Rats," *South Asian Journal of Experimental Biology*, vol. 7, no. 4, pp. 166–72, 2018.
- [4] T. Kurahashi and J. Fujii, "Roles of Antioxidative Enzymes in Wound Healing," *Journal of Developmental Biology*, vol. 3, no. 2, pp. 57–70, 2015.
- [5] G. Calviello, G. M. Filippi, A. Toesca et al., "Repeated exposure to pyrrolidine-dithiocarbamate induces peripheral nerve alterations in rats," *Toxicology Letters*, vol. 158, no. 1, pp. 61–71, 2005.
- [6] B. Poljsak, D. Šuput, and I. Milisav, "Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants," *Oxidative Medicine and Cellular Longevity*, vol. 2013, Article ID 956792, 11 pages, 2013.
- [7] S. E. Atawodi, "Antioxidant potential of African medicinal plants," *African Journal of Biotechnology*, vol. 4, no. 2, pp. 128–133, 2005.
- [8] E. O. Iwalewa, L. J. McGaw, V. Naidoo, and J. N. Eloff, "Inflammation: the foundation of diseases and disorders. A review of phytomedicines of South African origin used to treat pain and inflammatory conditions," *African Journal of Biotechnology*, vol. 6, no. 25, pp. 2868–2885, 2007.
- [9] M. F. Mahomoodally, "Traditional medicines in Africa: an appraisal of ten potent African medicinal plants," *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, Article ID 617459, 14 pages, 2013.
- [10] G. R. Schinella, H. A. Tournier, J. M. Prieto, P. M. de Buschiazzo, and J. L. Ríos, "Antioxidant activity of anti-inflammatory plant extracts," *Life Sciences*, vol. 70, no. 9, pp. 1023–1033, 2002.
- [11] J. A. Cook, D. J. Vanderjagt, A. Dasgupta et al., "Use of the trolox assay to estimate the antioxidant content of seventeen edible wild plants of niger," *Life Sciences*, vol. 63, no. 2, pp. 105–110, 1998.
- [12] J. Igoli, O. Ogaji, T. Tor-Anyiin, and N. Igoli, "Traditional Medicine Practice amongst the Igede People of Nigeria. Part II," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 2, no. 2, 2005.
- [13] J. Ojewole, "Antiinflammatory, analgesic and hypoglycemic effects of *Mangifera indica* Linn. (Anacardiaceae) stem-bark aqueous extract," *Methods and Findings in Experimental and Clinical Pharmacology*, vol. 27, no. 8, pp. 547–554, 2005.
- [14] R. Gebhardt, "Antioxidative and protective properties of extracts from leaves of the artichoke (*Cynara scolymus* L.) against hydroperoxide-induced oxidative stress in cultured rat hepatocytes," *Toxicology and Applied Pharmacology*, vol. 144, no. 2, pp. 279–286, 1997.
- [15] H. Li, N. Xia, I. Brausch, Y. Yao, and U. Förstermann, "Flavonoids from artichoke (*Cynara scolymus* L.) up-regulate endothelial-type nitric-oxide synthase gene expression in human endothelial cells," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 310, no. 3, pp. 926–932, 2004.
- [16] L. Bramati, F. Aquilano, and P. Pietta, "Unfermented Rooibos Tea: Quantitative Characterization of Flavonoids by HPLC-UV and Determination of the Total Antioxidant Activity," *Journal of Agricultural and Food Chemistry*, vol. 51, no. 25, pp. 7472–7474, 2003.
- [17] L. Bramati, M. Minoggio, C. Gardana, P. Simonetti, P. Mauri, and P. Pietta, "Quantitative characterization of flavonoid compounds in Rooibos tea (*Aspalathus linearis*) by LC-UV/DAD," *Journal of Agricultural and Food Chemistry*, vol. 50, no. 20, pp. 5513–5519, 2002.
- [18] O. Inanami, T. Asanuma, N. Inukai et al., "The suppression of age-related accumulation of lipid peroxides in rat brain by administration of Rooibos tea (*Aspalathus linearis*)," *Neuroscience Letters*, vol. 196, no. 1-2, pp. 85–88, 1995.
- [19] K. L. Lindsey, M. L. Motsei, and A. K. Jäger, "Screening of South African food plants for antioxidant activity," *Journal of Food Science*, vol. 67, no. 6, pp. 2129–2131, 2002.
- [20] C. Rabe, J. A. Steenkamp, E. Joubert, J. F. W. Burger, and D. Ferreira, "Phenolic metabolites from rooibos tea (*Aspalathus linearis*)," *Phytochemistry*, vol. 35, no. 6, pp. 1559–1565, 1994.
- [21] V. Steenkamp, E. Mathivha, M. C. Gouws, and C. E. J. Van Rensburg, "Studies on antibacterial, antioxidant and fibroblast growth stimulation of wound healing remedies from South Africa," *Journal of Ethnopharmacology*, vol. 95, no. 2-3, pp. 353–357, 2004.
- [22] M. T. Baratta, H. J. D. Dorman, S. G. Deans, A. C. Figueiredo, J. G. Barroso, and G. Ruberto, "Antimicrobial and antioxidant properties of some commercial essential oils," *Flavour and Fragrance Journal*, vol. 13, no. 4, pp. 235–244, 1998.
- [23] R. Juliani Hector, J. E. Simon, M. M. Roland Ramboatiana, O. Behra, A. S. Garvey, and I. Raskin, "Malagasy aromatic plants: Essential oils, antioxidant and antimicrobial activities," *Acta Horticulturae*, vol. 629, pp. 77–81, 2004.
- [24] J. Mancini-Filho, A. Van-Koijij, D. A. P. Mancini, F. F. Cozzolino, and R. P. Torres, "Antioxidant activity of cinnamon (*Cinnamomum zeylanicum*, breyne) extracts," *Bollettino Chimico Farmaceutico*, vol. 137, no. 11, pp. 443–447, 1998.
- [25] S. Möllenbeck, T. König, P. Schreier, W. Schwab, J. Rajaonarivony, and L. Ranarivelo, "Chemical composition and analyses

- of enantiomers of essential oils from Madagascar," *Flavour and Fragrance Journal*, vol. 12, no. 2, pp. 63–69, 1997.
- [26] N. Dilsiz, A. Sahaboglu, M. Z. Yildiz, and A. Reichenbach, "Protective effects of various antioxidants during ischemia-reperfusion in the rat retina," *Graef's Archive for Clinical and Experimental Ophthalmology*, vol. 244, no. 5, pp. 627–633, 2006.
- [27] R. Randhir, Y.-T. Lin, and K. Shetty, "Phenolics, their antioxidant and antimicrobial activity in dark germinated fenugreek sprouts in response to peptide and phytochemical elicitors," *Asia Pacific Journal of Clinical Nutrition*, vol. 13, no. 3, pp. 295–307, 2004.
- [28] K. Srinivasan, K. Sambaiah, and N. Chandrasekhara, "Spices as beneficial hypolipidemic food adjuncts: A review," *Food Reviews International*, vol. 20, no. 2, pp. 187–220, 2004.
- [29] O. A. Badary, R. A. Taha, A. M. Gamal El-Din, and M. H. Abdel-Wahab, "Thymoquinone is a potent superoxide anion scavenger," *Drug and Chemical Toxicology*, vol. 26, no. 2, pp. 87–98, 2003.
- [30] N. Farah, H. Benguzzzi, M. Tucci, and Z. Cason, "The effects of isolated antioxidants from black seed on the cellular metabolism of A549 cells," *Biomedical Sciences Instrumentation*, vol. 41, pp. 211–216, 2005.
- [31] M. F. Ramadan, L. W. Kroh, and J.-T. Mörsel, "Radical scavenging activity of black cumin (*Nigella sativa* L.), coriander (*Coriandrum sativum* L.), and Niger (*Guizotia abyssinica* Cass.) crude seed oils and oil fractions," *Journal of Agricultural and Food Chemistry*, vol. 51, no. 24, pp. 6961–6969, 2003.
- [32] S. Lee, S. Do, S. Y. Kim, J. Kim, Y. Jin, and C. H. Lee, "Mass spectrometry-based metabolite profiling and antioxidant activity of *Aloe vera* (*Aloe barbadensis* Miller) in different growth stages," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 45, pp. 11222–11228, 2012.
- [33] X.-f. Zhang, H.-m. Wang, Y.-l. Song et al., "Isolation, structure elucidation, antioxidative and immunomodulatory properties of two novel dihydrocoumarins from *Aloe vera*," *Bioorganic & medicinal chemistry letters*, vol. 16, no. 4, pp. 949–953, 2006.
- [34] M. Moniruzzaman, B. Rokeya, S. Ahmed, A. Bhowmik, M. I. Khalil, and S. H. Gan, "In vitro antioxidant effects of *aloë barbadensis miller* extracts and the potential role of these extracts as antidiabetic and antilipidemic agents on streptozotocin-induced type 2 diabetic model rats," *Molecules*, vol. 17, no. 11, pp. 12851–12867, 2012.
- [35] K. L. Lindsey, A. M. Viljoen, and A. K. Jäger, "Screening of *Aloe* species for antioxidant activity," *South African Journal of Botany*, vol. 69, no. 4, pp. 599–602, 2003.
- [36] S. O. Amoo, A. O. Aremu, and J. Van Staden, "Unraveling the medicinal potential of South African *Aloe* species," *Journal of Ethnopharmacology*, vol. 153, no. 1, pp. 19–41, 2014.
- [37] P. J. Zapata, D. Navarro, F. Guillén et al., "Characterisation of gels from different *Aloe* spp. as antifungal treatment: Potential crops for industrial applications," *Industrial Crops and Products*, vol. 42, no. 1, pp. 223–230, 2013.
- [38] R. Bruni, A. Guerrini, S. Scalia, C. Romagnoli, and G. Sacchetti, "Rapid techniques for the extraction of vitamin E isomers from *Amaranthus caudatus* seeds: ultrasonic and supercritical fluid extraction," *Phytochemical Analysis*, vol. 13, no. 5, pp. 257–261, 2002.
- [39] P. Veeru, M. P. Kishor, and M. Meenakshi, "Screening of medicinal plant extracts for antioxidant activity," *Journal of Medicinal Plants Research*, vol. 3, no. 8, pp. 608–612, 2009.
- [40] D. M. Jiménez-Aguilar and M. A. Grusak, "Minerals, vitamin C, phenolics, flavonoids and antioxidant activity of Amaranthus leafy vegetables," *Journal of Food Composition and Analysis*, vol. 58, pp. 33–39, 2017.
- [41] O. O. Ajileye, E. M. Obuotor, E. O. Akinkunmi, and M. A. Aderogba, "Isolation and characterization of antioxidant and antimicrobial compounds from *Anacardium occidentale* L. (Anacardiaceae) leaf extract," *Journal of King Saud University - Science*, vol. 27, no. 3, pp. 244–252, 2015.
- [42] R. Velagapudi, O. O. Ajileye, U. Okorji, P. Jain, M. A. Aderogba, and O. A. Olajide, "Agathisflavone isolated from *Anacardium occidentale* suppresses SIRT1-mediated neuroinflammation in BV2 microglia and neurotoxicity in APPS we-transfected SH-SY5Y cells," *Phytotherapy Research*, vol. 32, no. 10, pp. 1957–1966, 2018.
- [43] A. Maroyi, "Traditional use of medicinal plants in south-central Zimbabwe: review and perspectives," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, article 31, 2011.
- [44] T. Munodawafa, L. S. Chagonda, and S. R. Moyo, "Antimicrobial and phytochemical screening of some Zimbabwean medicinal plants," *Journal of Biologically Active Products from Nature*, vol. 3, no. 5–6, pp. 323–330, 2013.
- [45] E. F. Queiroz, C. Kuhl, C. Terreaux, S. Mavi, and K. Hostettmann, "New dihydroalkylhexenones from *Lannea edulis*," *Journal of Natural Products*, vol. 66, no. 4, pp. 578–580, 2003.
- [46] A. Maiga, K. E. Malterud, D. Diallo, and B. S. Paulsen, "Antioxidant and 15-lipoxygenase inhibitory activities of the Malian medicinal plants *Diospyros abyssinica* (Hiern) F. White (Ebenaceae), *Lannea velutina* A. Rich (Anacardiaceae) and *Crossopteryx febrifuga* (Afzel) Benth. (Rubiaceae)," *Journal of Ethnopharmacology*, vol. 104, no. 1–2, pp. 132–137, 2006.
- [47] L. Ouattara, J. Koudou, C. Zongo et al., "Antioxidant and antibacterial activities of three species of *Lannea* from Burkina Faso," *Journal of Applied Sciences*, vol. 11, no. 1, pp. 157–162, 2011.
- [48] R. Arora, D. Gupta, R. Chawla et al., "Radioprotection by plant products: present status and future prospects," *Phytotherapy Research*, vol. 19, no. 1, pp. 1–22, 2005.
- [49] Y.-J. Chen, Y.-S. Dai, B.-F. Chen et al., "The effect of tetrrandrine and extracts of *centella asiatica* on acute radiation dermatitis in rats," *Biological & Pharmaceutical Bulletin*, vol. 22, no. 7, pp. 703–706, 1999.
- [50] G. Jayashree, G. Kurup Muraleedhara, S. Sudarslal, and V. B. Jacob, "Anti-oxidant activity of *Centella asiatica* on lymphoma-bearing mice," *Fitoterapia*, vol. 74, no. 5, pp. 431–434, 2003.
- [51] D. MacKay and A. L. Miller, "Nutritional support for wound healing," *Alternative Medicine Review*, vol. 8, no. 4, pp. 359–377, 2003.
- [52] F. Pittella, R. C. Dutra, D. D. Junior, M. T. P. Lopes, and N. R. Barbosa, "Antioxidant and cytotoxic activities of *Centella asiatica* (L) Urb.," *International Journal of Molecular Sciences*, vol. 10, no. 9, pp. 3713–3721, 2009.
- [53] J. Sharma and R. Sharma, "Radioprotection of Swiss Albino Mouse by *Centella asiatica* Extract," *Phytotherapy Research*, vol. 16, no. 8, pp. 785–786, 2002.
- [54] R. Sharma and J. Sharma, "Modification of gamma ray induced changes in the mouse hepatocytes by *Centella asiatica* extract: In vivo studies," *Phytotherapy Research*, vol. 19, no. 7, pp. 605–611, 2005.
- [55] A. Shukla, A. M. Rasik, and B. N. Dhawan, "Asiaticoside-induced elevation of antioxidant levels in healing wounds," *Phytotherapy Research*, vol. 13, no. 1, pp. 50–54, 1999.

- [56] G. Kweifio-Okai, "Antiinflammatory activity of a Ghanaian antiarthritic herbal preparation: I," *Journal of Ethnopharmacology*, vol. 33, no. 3, pp. 263–267, 1991.
- [57] O. A. Olajide, J. M. Makinde, D. T. Okpako, and S. O. Awe, "Studies on the anti-inflammatory and related pharmacological properties of the aqueous extract of Bridelia ferruginea stem bark," *Journal of Ethnopharmacology*, vol. 71, no. 1-2, pp. 153–160, 2000.
- [58] B. B. Fakae, A. M. Campbell, J. Barrett et al., "Inhibition of glutathione S-transferases (GSTs) from parasitic nematodes by extracts from traditional Nigerian medicinal plants," *Phytotherapy Research*, vol. 14, no. 8, pp. 630–634, 2000.
- [59] N. Okoye and C. Okoye, "Anti-oxidant and Antimicrobial Flavonoid Glycosides from Alstonia boonei De Wild Leaves," *British Journal of Pharmaceutical Research*, vol. 10, no. 6, pp. 1–9, 2016.
- [60] W. Zheng and S. Y. Wang, "Antioxidant activity and phenolic compounds in selected herbs," *Journal of Agricultural and Food Chemistry*, vol. 49, no. 11, pp. 5165–5170, 2001.
- [61] S. E. Atawodi, L. M. Yusufu, J. C. Atawodi, O. Asuku, and O. E. Yakubu, "Phenolic Compounds and Antioxidant Potential of Nigerian Red Palm Oil (*Elaeis Guineensis*)," *International Journal of Biology*, vol. 3, no. 2, 2011.
- [62] A. Y. Mensah, P. J. Houghton, G. N. A. Akyirem et al., "Evaluation of the antioxidant and free radical scavenging properties of Secamone afzelii Rhoem," *Phytotherapy Research*, vol. 18, no. 12, pp. 1031–1032, 2004.
- [63] P. J. Houghton, P. J. Hylands, A. Y. Mensah, A. Hensel, and A. M. Deters, "In vitro tests and ethnopharmacological investigations: wound healing as an example," *Journal of Ethnopharmacology*, vol. 100, no. 1-2, pp. 100–107, 2005.
- [64] H. Zabri, C. Kodjo, A. Benie, J. M. Bekro, and Y. A. Bekro, "Phytochemical screening and determination of flavonoids in Secamone afzelii (Asclepiadaceae) extracts," *African Journal of Pure and Applied Chemistry*, vol. 2, no. 8, pp. 80–82, 2008.
- [65] A. A. Wube, F. Bucar, K. Asres et al., "Knipholone, a selective inhibitor of leukotriene metabolism," *Phytomedicine*, vol. 13, no. 6, pp. 452–456, 2006.
- [66] M. Bezabhi and B. M. Abegaz, "4'-Demethylknipholone from aerial parts of Bulbine capitata," *Phytochemistry*, vol. 48, no. 6, pp. 1071–1073, 1998.
- [67] R. R. T. Majinda, B. M. Abegaz, M. Bezabih et al., "Recent results from natural product research at the University of Botswana," *Pure and Applied Chemistry*, vol. 73, no. 7, pp. 1197–1208, 2001.
- [68] M. Bezabih, B. M. Abegaz, K. Dufall, K. Croft, T. Skinner-Adams, and T. M. E. Davis, "Antiplasmodial and antioxidant isofuranonaphthoquinones from the roots of Bulbine capitata," *Planta Medica*, vol. 67, no. 4, pp. 340–344, 2001.
- [69] B. M. Abegaz, "Novel phenylanthraquinones, isofuranonaphthoquinones, homoisoflavanoids, and biflavonoids from African plants in the genera Bulbine, Scilla, Ledebouria, and Rhus," *Phytochemistry Reviews*, vol. 1, no. 3, pp. 299–310, 2002.
- [70] B. M. Abegaz, M. Bezabih, T. Msuta et al., "Gaboroquinones A and B and 4 $\beta$ -O-demethylknipholone-4 $\beta$ -O- $\beta$ -D-glucopyranoside, phenylanthraquinones from the roots of Bulbine frutescens," *Journal of Natural Products*, vol. 65, no. 8, pp. 1117–1121, 2002.
- [71] M. Bezabih, S. Motlhagodi, and B. M. Abegaz, "Isofuranonaphthoquinones and phenolic and knipholone derivatives from the roots of Bulbine capitata," *Phytochemistry*, vol. 46, no. 6, pp. 1063–1067, 1997.
- [72] G. J. Grubben, *Plant Resources of Tropical Africa (PROTA)*, Prota, 2008.
- [73] A. Mats' eliso and P. Karuso, "Secondary Metabolites from Basotho Medicinal Plants. II. Bulbine capitata," *Australian Journal of Chemistry*, vol. 54, no. 7, pp. 427–430, 2001.
- [74] J. Mutanyatta, M. Bezabih, B. M. Abegaz et al., "The first 6 $\beta$ -O-sulfated phenylanthraquinones: Isolation from Bulbine frutescens, structural elucidation, enantiomeric purity, and partial synthesis," *Tetrahedron*, vol. 61, no. 35, pp. 8475–8484, 2005.
- [75] P. Tambama, B. Abegaz, and S. Mukanganyama, "Antiproliferative activity of the isofuranonaphthoquinone isolated from *Bulbine frutescens* against jurkat T cells," *BioMed Research International*, vol. 2014, Article ID 752941, 14 pages, 2014.
- [76] M. Adams and R. Bauer, "Inhibition of leukotriene biosynthesis by secondary plant metabolites," *Current Organic Chemistry*, vol. 12, no. 8, pp. 602–618, 2008.
- [77] S. Habtemariam, "Knipholone anthrone from Kniphofia foliosa induces a rapid onset of necrotic cell death in cancer cells," *Fitoterapia*, vol. 81, no. 8, pp. 1013–1019, 2010.
- [78] S. Habtemariam, "Antioxidant activity of Knipholone anthrone," *Food Chemistry*, vol. 102, no. 4, pp. 1042–1047, 2007.
- [79] M. Burits, K. Asres, and F. Bucar, "The antioxidant activity of the essential oils of *Artemisia afra*, *Artemisia abyssinica* and *Juniperus procera*," *Phytotherapy Research*, vol. 15, no. 2, pp. 103–108, 2001.
- [80] S. A. Emami, J. Asili, Z. Mohagheghi, and M. K. Hassanzadeh, "Antioxidant activity of leaves and fruits of Iranian conifers," *Evidence-Based Complementary and Alternative Medicine*, vol. 4, no. 3, pp. 313–319, 2007.
- [81] M. Esteban, L. G. Collado, F. A. Macías, G. M. Massanet, and F. R. Luis, "Flavonoids from *Artemisia lanata*," *Phytochemistry*, vol. 25, no. 6, pp. 1502–1504, 1986.
- [82] V. Naidoo, L. J. McGaw, S. P. R. Bisschop, N. Duncan, and J. N. Elloff, "The value of plant extracts with antioxidant activity in attenuating coccidiosis in broiler chickens," *Veterinary Parasitology*, vol. 153, no. 3-4, pp. 214–219, 2008.
- [83] L. V. Buwa and A. J. Afolayan, "Antimicrobial activity of some medicinal plants used for the treatment of tuberculosis in the Eastern Cape Province, South Africa," *African Journal of Biotechnology*, vol. 8, no. 23, pp. 6683–6687, 2009.
- [84] B.-E. Van Wyk, B. v. Oudtshoorn, and N. Gericke, *Medicinal Plants of South Africa*, Briza, 1997.
- [85] A. Djeridane, M. Yousfi, B. Nadjemli, D. Boutassouna, P. Stocker, and N. Vidal, "Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds," *Food Chemistry*, vol. 97, no. 4, pp. 654–660, 2006.
- [86] A. Akroud, L. A. Gonzalez, H. El Jani, and P. C. Madrid, "Antioxidant and antitumor activities of *Artemisia campestris* and *Thymelaea hirsuta* from southern Tunisia," *Food and Chemical Toxicology*, vol. 49, no. 2, pp. 342–347, 2011.
- [87] M. B. Naili, R. O. Alghazeer, N. A. Saleh, and A. Y. Al-Najjar, "Evaluation of antibacterial and antioxidant activities of *Artemisia campestris* (Astraceae) and *Ziziphus lotus* (Rhamnaceae)," *Arabian Journal of Chemistry*, vol. 3, no. 2, pp. 79–84, 2010.
- [88] M. G. L. Brandão, C. G. C. Nery, M. A. S. Mamão, and A. U. Krettli, "Two methoxylated flavone glycosides from *Bidens pilosa*," *Phytochemistry*, vol. 48, no. 2, pp. 397–399, 1998.

- [89] Y.-M. Chiang, D.-Y. Chuang, S.-Y. Wang, Y.-H. Kuo, P.-W. Tsai, and L.-F. Shyur, "Metabolite profiling and chemopreventive bioactivity of plant extracts from *Bidens pilosa*," *Journal of Ethnopharmacology*, vol. 95, no. 2-3, pp. 409–419, 2004.
- [90] L.-P. Yuan, F.-H. Chen, L. Ling et al., "Protective effects of total flavonoids of *Bidens pilosa* L. (TFB) on animal liver injury and liver fibrosis," *Journal of Ethnopharmacology*, vol. 116, no. 3, pp. 539–546, 2008.
- [91] F. Deba, T. D. Xuan, M. Yasuda, and S. Tawata, "Chemical composition and antioxidant, antibacterial and antifungal activities of the essential oils from *Bidens pilosa* Linn. var. *Radiata*," *Food Control*, vol. 19, no. 4, pp. 346–352, 2008.
- [92] F. Fratianni, M. Tucci, M. D. Palma, R. Pepe, and F. Nazzaro, "Polyphenolic composition in different parts of some cultivars of globe artichoke (*Cynara cardunculus* L. var. *scolymus* (L.) Fiori)," *Food Chemistry*, vol. 104, no. 3, pp. 1282–1286, 2007.
- [93] E. Speroni, R. Cervellati, P. Govoni, S. Guizzardi, C. Renzulli, and M. C. Guerra, "Efficacy of different *Cynara scolymus* preparations on liver complaints," *Journal of Ethnopharmacology*, vol. 86, no. 2-3, pp. 203–211, 2003.
- [94] A. C. U. Lourens, D. Reddy, K. H. C. Bašer, A. M. Viljoen, and S. F. van Vuuren, "In vitro biological activity and essential oil composition of four indigenous South African *Helichrysum* species," *Journal of Ethnopharmacology*, vol. 95, no. 2-3, pp. 253–258, 2004.
- [95] S. Albayrak, A. Aksoy, O. Sagdic, and E. Hamzaoglu, "Compositions, antioxidant and antimicrobial activities of *Helichrysum* (Asteraceae) species collected from Turkey," *Food Chemistry*, vol. 119, no. 1, pp. 114–122, 2010.
- [96] A. C. U. Lourens, A. M. Viljoen, and F. R. van Heerden, "South African *Helichrysum* species: a review of the traditional uses, biological activity and phytochemistry," *Journal of Ethnopharmacology*, vol. 119, no. 3, pp. 630–652, 2008.
- [97] L. G. Ranilla, Y.-I. Kwon, E. Apostolidis, and K. Shetty, "Phenolic compounds, antioxidant activity and *in vitro* inhibitory potential against key enzymes relevant for hyperglycemia and hypertension of commonly used medicinal plants, herbs and spices in Latin America," *Bioresource Technology*, vol. 101, no. 12, pp. 4676–4689, 2010.
- [98] M. A. Gyamfi, M. Yonamine, and Y. Aniya, "Free-radical scavenging action of medicinal herbs from GhanaThonningia sanguinea on experimentally-induced liver injuries," *General Pharmacology: The Vascular System*, vol. 32, no. 6, pp. 661–667, 1999.
- [99] I. I. Ohtani, N. Gotoh, J. Tanaka, T. Higa, M. A. Gyamfi, and Y. Aniya, "Thonningianins A and B, new antioxidants from the African medicinal herb *Thonningia sanguinea*," *Journal of Natural Products*, vol. 63, no. 5, pp. 676–679, 2000.
- [100] M. A. Gyamfi, N. Hokama, K. Oppong-Boachie, and Y. Aniya, "Inhibitory effects of the medicinal herb, *Thonningia sanguinea*, on liver drug metabolizing enzymes of rats," *Human & Experimental Toxicology*, vol. 19, no. 11, pp. 623–631, 2000.
- [101] M. A. Gyamfi, T. Tanaka, and Y. Aniya, "Selective suppression of cytochrome P450 gene expression by the medicinal herb, *Thonningia sanguinea* in rat liver," *Life Sciences*, vol. 74, no. 14, pp. 1723–1737, 2004.
- [102] M. A. Gyamfi, I. I. Ohtani, E. Shinno, and Y. Aniya, "Inhibition of glutathione S-transferases by thonningianin A, isolated from the African medicinal herb, *Thonningia sanguinea*, *in vitro*," *Food and Chemical Toxicology*, vol. 42, no. 9, pp. 1401–1408, 2004.
- [103] J. D. N'Guessan, A. P. Bidié, B. N. Lenta, B. Weniger, P. André, and F. Guédé-Guina, "*In vitro* assays for bioactivity-guided isolation of anti salmonella and antioxidant compounds in *Thonningia sanguinea* flowers," *African Journal of Biotechnology*, vol. 6, no. 14, pp. 1685–1689, 2007.
- [104] E. Speroni, R. Cervellati, G. Innocenti et al., "Anti-inflammatory, anti-nociceptive and antioxidant activities of *Balanites aegyptiaca* (L.) Delile," *Journal of Ethnopharmacology*, vol. 98, no. 1-2, pp. 117–125, 2005.
- [105] D. L. Chothani and H. U. Vaghasiya, "A review on *Balanites aegyptiaca* Del (desert date): phytochemical constituents, traditional uses, and pharmacological activity," *Pharmacognosy Reviews*, vol. 5, no. 9, pp. 55–62, 2011.
- [106] A. El Tahir, A. M. Ibrahim, G. M. H. Satti, T. G. Theander, A. Kharazmi, and S. A. Khalid, "The potential antileishmanial activity of some Sudanese medicinal plants," *Phytotherapy Research*, vol. 12, no. 8, pp. 576–579, 1998.
- [107] O. A. Binutu and B. A. Lajubutu, "Antimicrobial potentials of some plant species of the Bignoniaceae family," *African Journal of Medicine and Medical Sciences*, vol. 23, no. 3, pp. 269–273, 1994.
- [108] J. J. Rojas, V. J. Ochoa, S. A. Ocampo, and J. F. Muñoz, "Screening for antimicrobial activity of ten medicinal plants used in Colombian folkloric medicine: a possible alternative in the treatment of non-nosocomial infections," *BMC Complementary and Alternative Medicine*, vol. 6, article 2, 2006.
- [109] A. Rana, S. Bhangalia, and H. P. Singh, "A new phenylethanoid glucoside from *Jacaranda mimosifolia*," *Natural Product Research (Formerly Natural Product Letters)*, vol. 27, no. 13, pp. 1167–1173, 2013.
- [110] K. Ofori-Kwakye, A. A. Kwapon, and F. Adu, "Antimicrobial activity of extracts and topical products of the stem bark of *Spathodea campanulata* for wound healing," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 6, no. 2, pp. 168–174, 2009.
- [111] M. Marzouk, A. Gamal-Eldeen, M. Mohamed, and M. El-Sayed, "Anti-proliferative and antioxidant constituents from *Tecoma stans*," *Zeitschrift fur Naturforschung - Section C Journal of Biosciences*, vol. 61, no. 11-12, pp. 783–791, 2006.
- [112] L. Selloum, L. Sebilhi, A. Mekhalfia, R. Mahdadi, and A. Senator, "Antioxidant activity of *Cleome arabica* leaves extract," *Biochemical Society Transactions*, vol. 25, no. 4, p. S608, 1997.
- [113] U. S. Akula and B. Odhay, "In vitro 5-lipoxygenase inhibition of polyphenolic antioxidants from undomesticated plants of South Africa," *Journal of Medicinal Plants Research*, vol. 2, no. 9, pp. 207–212, 2008.
- [114] E. O. Farombi, P. Møller, and L. O. Dragsted, "Ex-vivo and *in vitro* protective effects of kolaviron against oxygen-derived radical-induced DNA damage and oxidative stress in human lymphocytes and rat liver cells," *Cell Biology and Toxicology*, vol. 20, no. 2, pp. 71–82, 2004.
- [115] E. O. Farombi, S. Shrotriya, and Y.-J. Surh, "Kolaviron inhibits dimethyl nitrosamine-induced liver injury by suppressing COX-2 and iNOS expression via NF-κB and AP-1," *Life Sciences*, vol. 84, no. 5-6, pp. 149–155, 2009.
- [116] E. O. Farombi, J. G. Tahnteng, A. O. Agboola, J. O. Nwankwo, and G. O. Emerole, "Chemoprevention of 2-acetylaminofluorene-induced hepatotoxicity and lipid peroxidation in rats by kolaviron—a *Garcinia kola* seed extract," *Food and Chemical Toxicology*, vol. 38, no. 6, pp. 535–541, 2000.

- [117] S. Olaleye, E. Farombi, E. Adewoye, B. Owoyele, S. Onasanwo, and R. Elegbe, "Analgesic and anti-inflammatory effects of kaviiron (a *Garcinia kola* seed extract)," *African journal of biomedical research*, vol. 3, no. 3, pp. 171–174, 2000.
- [118] O. A. Adaramoye and O. Akinloye, "Possible protective effect of kolaviron on CCl<sub>4</sub>-induced erythrocyte damage in rats," *Bioscience Reports*, vol. 20, no. 4, pp. 259–264, 2000.
- [119] O. A. Adaramoye, V. O. Nwaneri, K. C. Anyanwo, E. O. Farombi, and G. O. Emerole, "Possible anti-atherogenic effect of kolaviron (a *Garcinia kola* seed extract) in hypercholesterolaemic rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 32, no. 1-2, pp. 40–46, 2005.
- [120] J. O. Nwankwo, J. G. Tahnteng, and G. O. Emerole, "Inhibition of aflatoxin B1 genotoxicity in human liver-derived HepG2 cells by kolaviron biflavonoids and molecular mechanisms of action," *European Journal of Cancer Prevention*, vol. 9, no. 5, pp. 351–361, 2000.
- [121] S. F. Kouam, B. T. Ngadjui, K. Krohn, P. Wafo, A. Ajaz, and M. I. Choudhary, "Prenylated anthronoid antioxidants from the stem bark of Harungana madagascariensis," *Phytochemistry*, vol. 66, no. 10, pp. 1174–1179, 2005.
- [122] P.-C. N. Biapa, G. A. Agbor, J. E. Oben, and J. Y. Ngogang, "Phytochemical studies and antioxidant properties of four medicinal plants used in Cameroon," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 4, no. 4, pp. 495–500, 2007.
- [123] E. O. Iwalewa, I. O. Adewale, B. J. Taiwo et al., "Effects of Harungana madagascariensis stem bark extract on the antioxidant markers in alloxan induced diabetic and carrageenan induced inflammatory disorders in rats," *Journal of Complementary and Integrative Medicine*, vol. 5, no. 1, 2008.
- [124] A. P. M. Bernardi, A. B. F. Ferraz, D. V. Albring et al., "Benzophenones from *Hypericum carinatum*," *Journal of Natural Products*, vol. 68, no. 5, pp. 784–786, 2005.
- [125] J. D. M. Nunes, P. S. Pinto, S. A. D. L. Bordignon, S. B. Rech, and G. L. von Poser, "Phenolic compounds in *Hypericum* species from the Trigynobrathys section," *Biochemical Systematics and Ecology*, vol. 38, no. 2, pp. 224–228, 2010.
- [126] D. A. El-Sherbiny, A. E. Khalifa, A. S. Attia, and E. D. Eldershary, "*Hypericum perforatum* extract demonstrates antioxidant properties against elevated rat brain oxidative status induced by amnestic dose of scopolamine," *Pharmacology Biochemistry & Behavior*, vol. 76, no. 3-4, pp. 525–533, 2003.
- [127] A. Herold, L. Cremer, A. Calugaru et al., "Antioxidant properties of some hydroalcoholic plant extracts with antiinflammatory activity," *Romanian Archives of Microbiology and Immunology*, vol. 62, no. 3-4, pp. 217–227, 2003.
- [128] A. Herold, L. Cremer, A. Calugaru et al., "Hydroalcoholic plant extracts with anti-inflammatory activity," *Romanian Archives of Microbiology and Immunology*, vol. 62, no. 1-2, pp. 117–129, 2003.
- [129] H. Hosseinzadeh, G.-R. Karimi, and M. Rakhshanizadeh, "Anticonvulsant effect of *Hypericum perforatum*: Role of nitric oxide," *Journal of Ethnopharmacology*, vol. 98, no. 1-2, pp. 207–208, 2005.
- [130] D. Z. Orčić, N. M. Mimica-Dukić, M. M. Francišković, S. S. Petrović, and E. T. Jovin, "Antioxidant activity relationship of phenolic compounds in *Hypericum perforatum* L," *Chemistry Central Journal*, vol. 5, no. 1, p. 34, 2011.
- [131] Y. Zou, Y. Lu, and D. Wei, "Antioxidant activity of a flavonoid-rich extract of *Hypericum perforatum* L. in vitro," *Journal of Agricultural and Food Chemistry*, vol. 52, no. 16, pp. 5032–5039, 2004.
- [132] M. A. Aderogba, D. T. Kgatle, L. J. McGaw, and J. N. Eloff, "Isolation of antioxidant constituents from *Combretum apiculatum* subsp. *apiculatum*," *South African Journal of Botany*, vol. 79, pp. 125–131, 2012.
- [133] P. H. Coombes and C. B. Rogers, "Methyl gardenolate A, a novel cycloartenoid ester from the leaves of *Combretum woodii* (Combretaceae)," *Natural Product Research (Formerly Natural Product Letters)*, vol. 16, no. 5, pp. 301–304, 2002.
- [134] J. N. Eloff, J. O. Famakin, and D. R. P. Katerere, "Combretum woodii (Combretaceae) leaf extracts have high activity against Gram-negative and Gram-positive bacteria," *African Journal of Biotechnology*, vol. 4, no. 10, pp. 1161–1166, 2005.
- [135] J. N. Eloff, J. O. Famakin, and D. R. P. Katerere, "Isolation of an antibacterial stilbene from *Combretum woodii* (Combretaceae) leaves," *African Journal of Biotechnology*, vol. 4, no. 10, pp. 1167–1171, 2005.
- [136] P. Masoko and J. N. Eloff, "Screening of twenty-four South African *Combretum* and six *Terminalia* species (Combretaceae) for antioxidant activities," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 4, no. 2, pp. 231–239, 2007.
- [137] V. K. Zishiri, *Potentising and application of a Combretum woodii leaf extract with high antibacterial and antioxidant activity*, University of Pretoria, 2005.
- [138] J. E. Angeh, X. Huang, I. Sattler et al., "Antimicrobial and anti-inflammatory activity of four known and one new triterpenoid from *Combretum imberbe* (Combretaceae)," *Journal of Ethnopharmacology*, vol. 110, no. 1, pp. 56–60, 2007.
- [139] N. Bouchet, L. Barrier, and B. Fauconneau, "Radical scavenging activity and antioxidant properties of tannins from *Guiera senegalensis* (Combretaceae)," *Phytotherapy Research*, vol. 12, no. 3, pp. 159–162, 1998.
- [140] S. Amos, E. Kolawole, P. Akah, C. Wambebe, and K. Gamaniel, "Behavioral effects of the aqueous extract of *Guiera senegalensis* in mice and rats," *Phytomedicine*, vol. 8, no. 5, pp. 356–361, 2001.
- [141] R. Ficarra, P. Ficarra, S. Tommasini et al., "Isolation and characterization of *Guiera senegalensis* J.F.Gmel. active principles," *Bollettino Chimico Farmaceutico*, vol. 136, no. 5, pp. 454–459, 1997.
- [142] Ž. Maleš, M. Medić-Šarić, and F. Bucar, "Flavonoids of *Guiera senegalensis* J. F. GMEL. -Thin-layer Chromatography and Numerical Methods," *Croatica Chemica Acta*, vol. 71, no. 1, pp. 69–79, 1998.
- [143] P. A. E. D. Sombié, A. Hilou, C. Mounier et al., "Antioxidant and anti-inflammatory activities from galls of *guiera senegalensis* J.F. Gmel (Combretaceae)," *Research Journal of Medicinal Plant*, vol. 5, no. 4, pp. 448–461, 2011.
- [144] I. M. S. Eldeen, E. E. Elgorashi, D. A. Mulholland, and J. Van Staden, "Anolignan B: A bioactive compound from the roots of *Terminalia sericea*," *Journal of Ethnopharmacology*, vol. 103, no. 1, pp. 135–138, 2006.
- [145] J. Ezea, T. Iwuji, and M. Ogwike, "Growth responses of pregnant rabbits and their litters fed Spreading day flower (*Commelina diffusa* Burm. F.) and rock fig (*Ficus ingens* Miquel) leaves," *Journal of Global Biosciences*, vol. 3, no. 2, pp. 619–625, 2014.
- [146] P. A. Akah and A. I. Nwambie, "Evaluation of Nigerian traditional medicines: 1. Plants used for rheumatic (inflammatory) disorders," *Journal of Ethnopharmacology*, vol. 42, no. 3, pp. 179–182, 1994.
- [147] E. Boakye-Gyasi, G. K. Ainooson, and W. K. Abotsi, "Anti-inflammatory, antipyretic and antioxidant properties of a

- hydroalcoholic leaf extract of *Palisota hirsuta* K. Schum. (Commelinaceae)," *West African Journal of Pharmacy*, vol. 22, no. 1, 2011.
- [148] J. A. O. Ojewole, "Antinociceptive, anti-inflammatory and antidiabetic effects of *Bryophyllum pinnatum* (Crassulaceae) leaf aqueous extract," *Journal of Ethnopharmacology*, vol. 99, no. 1, pp. 13–19, 2005.
- [149] S. J. N. Tatsumi, J. D. D. Tamokou, L. Havyarimana et al., "Antimicrobial and antioxidant activity of kaempferol rhamnoside derivatives from *Bryophyllum pinnatum*," *BMC Research Notes*, vol. 5, article 158, 2012.
- [150] S. I. Alqasoumi and M. S. Abdel-Kader, "Terpenoids from *Juniperus procera* with hepatoprotective activity," *Pakistan Journal of Pharmaceutical Sciences*, vol. 25, no. 2, pp. 315–322, 2012.
- [151] N. Orhan, I. E. Orhan, and F. Ergun, "Insights into cholinesterase inhibitory and antioxidant activities of five *Juniperus* species," *Food and Chemical Toxicology*, vol. 49, no. 9, pp. 2305–2312, 2011.
- [152] M. Araghniknam, S. Chung, T. Nelson-White, C. Eskelson, and R. R. Watson, "Antioxidant activity of disoscorea and dehydroepiandrosterone (DHEA) in older humans," *Life Sciences*, vol. 59, no. 11, pp. PL147–PL157, 1996.
- [153] M. M. Iwu, C. O. Okunji, G. O. Ohiaeri, P. Akah, D. Corley, and M. S. Tempesta, "Hypoglycaemic activity of dioscoretine from tubers of *Dioscorea dumetorum* in normal and alloxan diabetic rabbits," *Planta Medica*, vol. 56, no. 3, pp. 264–267, 1990.
- [154] M. A. Sonibare and R. B. Abegunde, "In vitro antimicrobial and antioxidant analysis of *Dioscorea dumetorum* (Kunth) Pax and *Dioscorea hirtiflora* (Linn.) and their bioactive metabolites from Nigeria," *Journal of Applied Biosciences*, vol. 51, pp. 3583–3590, 2012.
- [155] D. H. Paper, E. Karall, M. Kremser, and L. Krenn, "Comparison of the antiinflammatory effects of *Drosera rotundifolia* and *Drosera madagascariensis* in the HEX-CAM assay," *Phytotherapy Research*, vol. 19, no. 4, pp. 323–326, 2005.
- [156] P. A. Egan and F. Van Der Kooy, "Phytochemistry of the carnivorous sundew genus *Drosera* (Droseraceae) - Future perspectives and ethnopharmacological relevance," *Chemistry & Biodiversity*, vol. 10, no. 10, pp. 1774–1790, 2013.
- [157] M. T. Giardi, G. Rea, and B. Berra, *Bio-Farms for Nutraceuticals: Functional Food and Safety Control by Biosensors*, Springer Science & Business Media, 2011.
- [158] E. O. Farombi, O. O. Ogundipe, E. S. Uhunwangho, M. A. Adeyanju, and J. O. Moody, "Antioxidant properties of extracts from *Alchornea laxiflora* (Benth) Pax and Hoffman," *Phytotherapy Research*, vol. 17, no. 7, pp. 713–716, 2003.
- [159] O. O. Ogundipe, J. O. Moody, P. J. Houghton, and H. A. Ode-lola, "Bioactive chemical constituents from *Alchornea laxiflora* (benth) pax and hoffman," *Journal of Ethnopharmacology*, vol. 74, no. 3, pp. 275–280, 2001.
- [160] R. N. Okigbo, C. L. Anuagasi, and J. E. Amadi, "Advances in selected medicinal and aromatic plants indigenous to Africa," *Journal of Medicinal Plants Research*, vol. 3, no. 2, pp. 86–95, 2009.
- [161] G. K. Oloyede, P. A. Onocha, J. Soyinka, O. Oguntokun, and E. Thonda, "Phytochemical screening, antimicrobial and antioxidant activities of four Nigerian medicinal plants," *Annals of Biological Research*, vol. 1, no. 2, pp. 114–120, 2010.
- [162] A. Adetutu, W. A. Morgan, and O. Corcoran, "Antibacterial, antioxidant and fibroblast growth stimulation activity of crude extracts of *Bridelia ferruginea* leaf, a wound-healing plant of Nigeria," *Journal of Ethnopharmacology*, vol. 133, no. 1, pp. 116–119, 2011.
- [163] B. Bakoma, B. Berké, K. Eklu-Gadegbeku et al., "Total phenolic content, antioxidant activity and In vitro inhibitory potential against key enzymes relevant for hyperglycemia of *Bridelia ferruginea* extracts," *Research Journal of Phytochemistry*, vol. 6, no. 4, pp. 120–126, 2012.
- [164] T. De Bruyne, K. Cimanga, L. Pieters, M. Claeys, R. Domisse, and A. Vlietinck, "Gallocatechin - (4'—O—>7) - epigallocatechin, a new biflavonoid isolated from *Bridelia ferruginea*," *Natural Product Research (Formerly Natural Product Letters)*, vol. 11, no. 1, pp. 47–52, 1998.
- [165] K. Cimanga, T. de Bruyne, S. Apers et al., "Complement-inhibiting constituents of *Bridelia ferruginea* stem bark," *Planta Medica*, vol. 65, no. 3, pp. 213–217, 1999.
- [166] O. A. Fabiyi, A. Olubunmi, O. S. Adeyemi, and G. A. Olatunji, "Antioxidant and Cytotoxicity of β-Amyrin acetate fraction from *Bridelia ferruginea* leaves," *Asian Pacific Journal of Tropical Biomedicine*, vol. 2, no. 2, pp. S981–S984, 2012.
- [167] E. O. Farombi, O. Ogundipe, and J. O. Moody, "Antioxidant and anti-inflammatory activities of *Mallotus oppositifolium* in model systems," *African Journal of Medicine and Medical Sciences*, vol. 30, no. 3, pp. 213–215, 2001.
- [168] J. C. Chukwujekwu, J. Van Staden, and P. Smith, "Antibacterial, anti-inflammatory and antimalarial activities of some Nigerian medicinal plants," *South African Journal of Botany*, vol. 71, no. 3-4, pp. 316–325, 2005.
- [169] V. Barku, Y. Opoku-Boahen, E. Owusu-Ansah, N. Dayie, and F. Mensah, "In-vitro assessment of antioxidant and antimicrobial activities of methanol extracts of six wound healing medicinal plants," *In-Vitro*, vol. 3, no. 1, 2013.
- [170] E. O. Farombi, "African indigenous plants with chemotherapeutic potentials and biotechnological approach to the production of bioactive prophylactic agents," *African Journal of Biotechnology*, vol. 2, no. 12, pp. 662–671, 2003.
- [171] R. Kamgang, E. Vidal Pouokam Kamgne, M. C. Fonkoua, V. Penlap N Beng, and M. Biwolé Sida, "Activities of aqueous extracts of *Mallotus oppositifolium* on *Shigella dysenteriae* A1-induced diarrhoea in rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 33, no. 1-2, pp. 89–94, 2006.
- [172] C. O. Nwaehujor, M. I. Ezeja, N. E. Udeh, D. N. Okoye, and R. I. Udegbunam, "Anti-inflammatory and anti-oxidant activities of *Mallotus oppositifolius* (Geisel) methanol leaf extracts," *Arabian Journal of Chemistry*, vol. 7, no. 5, pp. 805–810, 2014.
- [173] P. W. Sinjman, E. Joubert, D. Ferreira et al., "Antioxidant activity of the dihydrochalcones aspalathin and nothofagin and their corresponding flavones in relation to other rooibos (*Aspalathus linearis*) flavonoids, epigallocatechin gallate, and Trolox," *Journal of Agricultural and Food Chemistry*, vol. 57, no. 15, pp. 6678–6684, 2009.
- [174] R. Johnson, D. D. Beer, P. V. Sludla, D. Ferreira, C. J. F. Muller, and E. Joubert, "Aspalathin from Rooibos (*Aspalathus linearis*): A Bioactive C -glucosyl Dihydrochalcone with Potential to Target the Metabolic Syndrome," *Planta Medica*, 2018.
- [175] E. Mathisen, D. Diallo, Ø. M. Andersen, and K. E. Malterud, "Antioxidants from the bark of *Burkea africana*, an African medicinal plant," *Phytotherapy Research*, vol. 16, no. 2, pp. 148–153, 2002.
- [176] R. Dave, "In vitro models for antioxidant activity evaluation and some medicinal plants possessing antioxidant properties: an overview," *African Journal of Microbiology Research*, vol. 3, no. 13, pp. 981–996, 2009.

- [177] F. Stoddard, "Novel feed and non-food uses of legumes," *Legume Futures Report*, vol. 1, 2013.
- [178] E. Joubert, E. S. Richards, J. D. Van Der Merwe, D. De Beer, M. Manley, and W. C. A. Gelderblom, "Effect of species variation and processing on phenolic composition and in vitro antioxidant activity of aqueous extracts of cyclopia spp. (Honeybush tea)," *Journal of Agricultural and Food Chemistry*, vol. 56, no. 3, pp. 954–963, 2008.
- [179] B. I. Kamara, D. J. Brand, E. V. Brandt, and E. Joubert, "Phenolic metabolites from honeybush tea (*Cyclopia subternata*)," *Journal of Agricultural and Food Chemistry*, vol. 52, no. 17, pp. 5391–5395, 2004.
- [180] B. I. Kamara, E. V. Brandt, D. Ferreira, and E. Joubert, "Polyphenols from honeybush tea (*Cyclopia intermedia*)," *Journal of Agricultural and Food Chemistry*, vol. 51, no. 13, pp. 3874–3879, 2003.
- [181] D. L. McKay and J. B. Blumberg, "A review of the bioactivity of South African herbal teas: Rooibos (*Aspalathus linearis*) and honeybush (*Cyclopia intermedia*)," *Phytotherapy Research*, vol. 21, no. 1, pp. 1–16, 2007.
- [182] M. D. Awouafack, P. Tane, and J. N. Eloff, "Two new antioxidant flavones from the twigs of *Eriosema robustum* (Fabaceae)," *Phytochemistry Letters*, vol. 6, no. 1, pp. 62–66, 2013.
- [183] A. Yenesew, S. Derese, B. Irungu et al., "Flavonoids and isoflavonoids with antiplasmoidal activities from the root bark of *Erythrina abyssinica*," *Planta Medica*, vol. 69, no. 7, pp. 658–661, 2003.
- [184] M. Chacha, G. Bojase-Moleta, and R. R. T. Majinda, "Antimicrobial and radical scavenging flavonoids from the stem wood of *Erythrina latissima*," *Phytochemistry*, vol. 66, no. 1, pp. 99–104, 2005.
- [185] C. C. W. Wanjala, B. F. Juma, G. Bojase, B. A. Gashe, and R. R. T. Majinda, "Erythraline alkaloids and antimicrobial flavonoids from *Erythrina latissima*," *Planta Medica*, vol. 68, no. 7, pp. 640–642, 2002.
- [186] C. C. W. Wanjala and R. R. T. Majinda, "Isoflavone glycosides from the root wood of *Erythrina latissima*," *Journal of AOAC International*, vol. 84, no. 2, pp. 451–453, 2001.
- [187] K. G. Dufall, B. T. Ngadjui, K. F. Simeon, B. M. Abegaz, and K. D. Croft, "Antioxidant activity of prenylated flavonoids from the West African medicinal plant *Dorstenia mannii*," *Journal of Ethnopharmacology*, vol. 87, no. 1, pp. 67–72, 2003.
- [188] V. Steenkamp, H. Grimmer, M. Semano, and M. Gulumian, "Antioxidant and genotoxic properties of South African herbal extracts," *Mutation Research - Genetic Toxicology and Environmental Mutagenesis*, vol. 581, no. 1-2, pp. 35–42, 2005.
- [189] S. El-Masry, M. E. Amer, M. S. Abdel-Kader, and H. H. Zaatout, "Prenylated flavonoids of *Erythrina lysistemon* grown in Egypt," *Phytochemistry*, vol. 60, no. 8, pp. 783–787, 2002.
- [190] U. Mabona and S. F. Van Vuuren, "Southern African medicinal plants used to treat skin diseases," *South African Journal of Botany*, vol. 87, pp. 175–193, 2013.
- [191] K. Asres, S. Gibbons, and V. Nachname, "Anti-inflammatory activity of extracts and a saponin isolated from *Melilotus elegans*," *Die Pharmazie-An International Journal of Pharmaceutical Sciences*, vol. 60, no. 4, pp. 310–312, 2005.
- [192] S. Chorespsima, K. Tentolouris, D. Dimitroulis, and N. Tentolouris, "Melilotus: Contribution to wound healing in the diabetic foot," *Journal of Herbal Medicine*, vol. 3, no. 3, pp. 81–86, 2013.
- [193] T. Gebre-Mariam, K. Asres, M. Getie, A. Endale, R. Neubert, and P. C. Schmidt, "In vitro availability of kaempferol glycosides from cream formulations of methanolic extract of the leaves of *Melilotus elegans*," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 60, no. 1, pp. 31–38, 2005.
- [194] T. Gebre-Mariam, R. Neubert, P. C. Schmidt, P. Wutzler, and M. Schmidtke, "Antiviral activities of some Ethiopian medicinal plants used for the treatment of dermatological disorders," *Journal of Ethnopharmacology*, vol. 104, no. 1-2, pp. 182–187, 2006.
- [195] E. Yankep, D. Njamen, M. T. Fotsing et al., "Griffonianone D, an isoflavone with anti-inflammatory activity from the root bark of *Millettia griffoniana*," *Journal of Natural Products*, vol. 66, no. 9, pp. 1288–1290, 2003.
- [196] S. Combes, J.-P. Finet, and D. Siri, "On the optical activity of the 3-aryl-4-hydroxycoumarin isolated from *Millettia griffoniana*: Molecular modelling and total synthesis," *Journal of the Chemical Society, Perkin Transactions 1*, vol. 2, no. 1, pp. 38–44, 2002.
- [197] D. Ngamga, E. Yankep, P. Tane et al., "Antiparasitic prenylated isoflavonoids from seeds of *Millettia griffoniana*," *Bulletin of the Chemical Society of Ethiopia*, vol. 19, no. 1, pp. 75–80, 2005.
- [198] D. Ngamga, E. Yankep, P. Tane et al., "Isoflavonoids from seeds of *Millettia griffoniana* (Bail), 15," *Zeitschrift fur Naturforschung - Section B Journal of Chemical Sciences*, vol. 60, no. 9, pp. 973–977, 2005.
- [199] E. Yankep, Z. T. Fomum, D. Bisrat, E. Dagne, V. Hellwig, and W. Steglich, "O-geranylated isoflavones and a 3-phenylcoumarin from *Millettia griffoniana*," *Phytochemistry*, vol. 49, no. 8, pp. 2521–2523, 1998.
- [200] E. Yankep, Z. T. Fomum, and E. Dagne, "An O-geranylated isoflavone from *Millettia griffoniana*," *Phytochemistry*, vol. 46, no. 3, pp. 591–593, 1997.
- [201] E. Yankep, J. T. Mbafor, Z. T. Fomum et al., "Further isoflavonoid metabolites from *Millettia griffoniana* (Bail)," *Phytochemistry*, vol. 56, no. 4, pp. 363–368, 2001.
- [202] S. Zingue, D. Njamen, J. Tchoumtchoua et al., "Effects of *Millettia macrophylla* (Fabaceae) extracts on estrogen target organs of female Wistar rat," *Journal of Pharmacological Sciences*, vol. 123, no. 2, pp. 120–131, 2013.
- [203] R. A. Sharma, A. J. Gescher, and W. P. Steward, "Curcumin: the story so far," *European Journal of Cancer*, vol. 41, no. 13, pp. 1955–1968, 2005.
- [204] D. A. Alabi, O. R. Akinsulire, and M. A. Sanyaolu, "Qualitative determination of chemical and nutritional composition of *Parkia biglobosa* (Jacq.) Benth," *African Journal of Biotechnology*, vol. 4, no. 8, pp. 812–815, 2005.
- [205] B. M. Abegaz, B. T. Ngadjui, E. Dongo, and M.-T. Bezabih, "Chemistry of the genus *Dorstenia*," *Current Organic Chemistry*, vol. 4, no. 10, pp. 1079–1090, 2000.
- [206] B. T. Ngadjui, E. Dongo, E. N. Happi, M.-T. Bezabih, and B. M. Abegaz, "Prenylated flavones and phenylpropanoid derivatives from roots of *Dorstenia psilurus*," *Phytochemistry*, vol. 48, no. 4, pp. 733–737, 1998.
- [207] B. T. Ngadjui, G. W. F. Kapche, H. Tamboue, B. M. Abegaz, and J. D. Connolly, "Prenylated flavonoids and a dihydro-4-phenylcoumarin from *Dorstenia poinsettifolia*," *Phytochemistry*, vol. 51, no. 1, pp. 119–123, 1999.
- [208] C. L. Miranda, J. F. Stevens, V. Ivanov et al., "Antioxidant and prooxidant actions of prenylated and nonprenylated chalcones and flavanones in vitro," *Journal of Agricultural and Food Chemistry*, vol. 48, no. 9, pp. 3876–3884, 2000.

- [209] G. O. Adegoke, M. Vijay Kumar, K. Sambaiah, and B. R. Lokesh, "Inhibitory effect of *Garcinia kola* on lipid peroxidation in rat liver homogenate," *Indian Journal of Experimental Biology (IJEB)*, vol. 36, no. 9, pp. 907–910, 1998.
- [210] A. O. Aremu, O. A. Fawole, J. C. Chukwujekwu, M. E. Light, J. F. Finnie, and J. Van Staden, "In vitro antimicrobial, antihelminthic and cyclooxygenase-inhibitory activities and phytochemical analysis of *Leucosidea sericea*," *Journal of Ethnopharmacology*, vol. 131, no. 1, pp. 22–27, 2010.
- [211] A. O. Aremu, S. O. Amoo, A. R. Ndhlala, J. F. Finnie, and J. Van Staden, "Antioxidant activity, acetylcholinesterase inhibition, iridoid content and mutagenic evaluation of *Leucosidea sericea*," *Food and Chemical Toxicology*, vol. 49, no. 5, pp. 1122–1128, 2011.
- [212] S. O. Amoo, J. F. Finnie, and J. Van Staden, "In vitro pharmacological evaluation of three *Barleria* species," *Journal of Ethnopharmacology*, vol. 121, no. 2, pp. 274–277, 2009.
- [213] S. O. Amoo, A. R. Ndhlala, J. F. Finnie, and J. Van Staden, "Antifungal, acetylcholinesterase inhibition, antioxidant and phytochemical properties of three *Barleria* species," *South African Journal of Botany*, vol. 77, no. 2, pp. 435–445, 2011.
- [214] D. Barron, A. Di Pietro, C. Dumontet, and D. B. McIntosh, "Isoprenoid flavonoids are new leads in the modulation of chemoresistance," *Phytochemistry Reviews*, vol. 1, no. 3, pp. 325–332, 2002.
- [215] M. G. L. Hertog, D. Kromhout, C. Aravanis et al., "Flavonoid intake and long-term risk of coronary heart disease and cancer in the Seven Countries Study," *JAMA Internal Medicine*, vol. 155, no. 4, pp. 381–386, 1995.
- [216] E. Middleton Jr., C. Kandaswami, and T. C. Theoharides, "The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer," *Pharmacological Reviews*, vol. 52, no. 4, pp. 673–751, 2000.
- [217] C. A. Williams and R. J. Grayer, "Anthocyanins and other flavonoids," *Natural Product Reports*, vol. 21, no. 4, pp. 539–573, 2004.
- [218] Y. Hanasaki, S. Ogawa, and S. Fukui, "The correlation between active oxygens scavenging and antioxidative effects of flavonoids," *Free Radical Biology & Medicine*, vol. 16, no. 6, pp. 845–850, 1994.
- [219] G. Cao, E. Sofic, and R. L. Prior, "Antioxidant and prooxidant behavior of flavonoids: structure-activity relationships," *Free Radical Biology & Medicine*, vol. 22, no. 5, pp. 749–760, 1997.
- [220] Z. Y. Chen, P. T. Chan, K. Y. Ho, K. P. Fung, and J. Wang, "Antioxidant activity of natural flavonoids is governed by number and location of their aromatic hydroxyl groups," *Chemistry and Physics of Lipids*, vol. 79, no. 2, pp. 157–163, 1996.
- [221] N. Cottelle, J.-L. Bernier, J.-P. Catteau, J. Pommery, J.-C. Waller, and E. M. Gaydou, "Antioxidant properties of hydroxyflavones," *Free Radical Biology & Medicine*, vol. 20, no. 1, pp. 35–43, 1996.
- [222] P. G. Pietta, "Flavonoids as antioxidants," *Journal of Natural Products*, vol. 63, no. 7, pp. 1035–1042, 2000.
- [223] C. A. Rice-Evans, N. J. Miller, and G. Paganga, "Structure-antioxidant activity relationships of flavonoids and phenolic acids," *Free Radical Biology & Medicine*, vol. 20, no. 7, pp. 933–956, 1996.
- [224] A. García-Lafuente, E. Guillamón, A. Villares, M. A. Rostagno, and J. A. Martínez, "Flavonoids as anti-inflammatory agents: implications in cancer and cardiovascular disease," *Inflammation Research*, vol. 58, no. 9, pp. 537–552, 2009.
- [225] B. F. Juma, A. Yenesew, J. O. Midiwo, and P. G. Waterman, "Flavones and phenylpropenoids in the surface exudate of *Psiadia punctulata*," *Phytochemistry*, vol. 57, no. 4, pp. 571–574, 2001.
- [226] B. T. Ngadjui, E. Dongo, B. M. Abegaz, S. Fotso, and H. Tamboue, "Dinklagins A, B and C: Three prenylated flavonoids and other constituents from the twigs of *Dorstenia dinklagei*," *Phytochemistry*, vol. 61, no. 1, pp. 99–104, 2002.
- [227] B. T. Ngadjui, E. Dongo, H. Tamboue, K. Fogue, and B. M. Abegaz, "Prenylated flavanones from the twigs of *Dorstenia mannii*," *Phytochemistry*, vol. 50, no. 8, pp. 1401–1406, 1999.
- [228] B. Reiersen, B. T. Kiremire, R. Byamukama, and Ø. M. Andersen, "Anthocyanins acylated with gallic acid from chenille plant, *Acalypha hispida*," *Phytochemistry*, vol. 64, no. 4, pp. 867–871, 2003.
- [229] M. Gordon, *The mechanism of antioxidant action in vitro*, Springer, Food antioxidants, 1990.
- [230] I. Pinchuk and D. Lichtenberg, "The mechanism of action of antioxidants against lipoprotein peroxidation, evaluation based on kinetic experiments," *Progress in Lipid Research*, vol. 41, no. 4, pp. 279–314, 2002.
- [231] N. Noguchi, A. Watanabe, and H. Shi, "Diverse functions of antioxidants," *Free Radical Research*, vol. 33, no. 6, pp. 809–817, 2000.
- [232] H. Kim, J. Y. Moon, H. Kim et al., "Antioxidant and antiproliferative activities of mango (*Mangifera indica* L.) flesh and peel," *Food Chemistry*, vol. 121, no. 2, pp. 429–436, 2010.
- [233] L. T. Ling, S.-A. Yap, A. K. Radhakrishnan, T. Subramaniam, H. M. Cheng, and U. D. Palanisamy, "Standardised *Mangifera indica* extract is an ideal antioxidant," *Food Chemistry*, vol. 113, no. 4, pp. 1154–1159, 2009.
- [234] A. Lamien-Meda, C. Lamien, M. Compaoré et al., "Polyphenol content and antioxidant activity of fourteen wild edible fruits from Burkina Faso," *Molecules*, vol. 13, no. 3, pp. 581–594, 2008.
- [235] S. E. Bizimenyera, G. E. Swan, H. Chikoto, and J. N. Eloff, "Rationale for using *Peltophorum africanum* (Fabaceae) extracts in veterinary medicine," *Journal of the South African Veterinary Association*, vol. 76, no. 2, pp. 54–58, 2005.
- [236] N. Mongalo, "Peltophorum africanum Sond [Mosetlha]: A review of its ethnomedicinal uses, toxicology, phytochemistry and pharmacological activities," *Journal of Medicinal Plants Research*, vol. 7, no. 48, pp. 3484–3491, 2013.
- [237] L. J. Shai, S. R. Magano, S. L. Lebelo, and A. M. Mogale, "Inhibitory effects of five medicinal plants on rat alpha-glucosidase: comparison with their effects on yeast alpha-glucosidase," *Journal of Medicinal Plants Research*, vol. 5, no. 13, pp. 2863–2867, 2011.
- [238] O. Mazimba, "Pharmacology and phytochemistry studies in *Peltophorum africanum*," *Bulletin of Faculty of Pharmacy, Cairo University*, vol. 52, no. 1, pp. 145–153, 2014.
- [239] J. C. Ibewuike, F. O. Ogungbamila, A. O. Ogundaini, I. N. Okeke, and L. Bohlin, "Antiinflammatory and antibacterial activities of C-methylflavonols from *piliostigma thonningii*," *Phytotherapy Research*, vol. 11, no. 4, pp. 281–284, 1997.
- [240] M. Aderogba, E. Okoh, T. Adelanwa, and O. AwolowoUniv, "Antioxidant properties of the Nigerian *Piliostigma* species," *Journal of Biological Sciences*, 2004.
- [241] T. Ajiboye, A. Salau, M. Yakubu, A. Oladiji, M. Akanji, and J. Okogun, "Aqueous extract of *Securidaca longepedunculata* root induce redox imbalance in male rat liver and kidney," *Human & Experimental Toxicology*, vol. 29, no. 8, pp. 679–688, 2010.

- [242] E. Bombardelli, A. Cristoni, A. Lolla et al., "Chemical and biological characterisation of *Piliostigma thonningii* polyphenols," *Fitoterapia*, vol. 65, no. 6, pp. 493–501, 1994.
- [243] J. C. Ibewuike, A. O. Ogundaini, F. O. Ogungbamila et al., "Piliostigmin, a 2-phenoxychromone, and C-methylflavonol from *Piliostigma thonningii*," *Phytochemistry*, vol. 43, no. 3, pp. 687–690, 1996.
- [244] O. M. Ighodaro, S. O. Agunbiade, J. O. Omole, and O. A. Kuti, "Evaluation of the chemical, nutritional, antimicrobial and antioxidant-vitamin profiles of *Piliostigma thonningii* leaves (Nigerian species)," *Research Journal of Medicinal Plant*, vol. 6, no. 7, pp. 537–543, 2012.
- [245] F. O. Jimoh and A. T. Oladiji, "Preliminary Studies on *Piliostigma thonningii* seeds: Proximate analysis, mineral composition and phytochemical screening," *African Journal of Biotechnology*, vol. 4, no. 12, pp. 1439–1442, 2005.
- [246] A. C. Fernandes, A. D. Cromarty, C. Albrecht, and C. E. Jansen Van Rensburg, "The antioxidant potential of *Sutherlandia frutescens*," *Journal of Ethnopharmacology*, vol. 95, no. 1, pp. 1–5, 2004.
- [247] J. Tai, S. Cheung, E. Chan, and D. Hasman, "In vitro culture studies of *Sutherlandia frutescens* on human tumor cell lines," *Journal of Ethnopharmacology*, vol. 93, no. 1, pp. 9–19, 2004.
- [248] B.-E. van Wyk and C. Albrecht, "A review of the taxonomy, ethnobotany, chemistry and pharmacology of *Sutherlandia frutescens* (Fabaceae)," *Journal of Ethnopharmacology*, vol. 119, no. 3, pp. 620–629, 2008.
- [249] S. Kaviarasan, G. H. Naik, R. Gangabhirathi, C. V. Anuradha, and K. I. Priyadarsini, "In vitro studies on antiradical and antioxidant activities of fenugreek (*Trigonella foenum graecum*) seeds," *Food Chemistry*, vol. 103, no. 1, pp. 31–37, 2007.
- [250] A. Wojdylo, J. Oszmiański, and R. Czemerys, "Antioxidant activity and phenolic compounds in 32 selected herbs," *Food Chemistry*, vol. 105, no. 3, pp. 940–949, 2007.
- [251] H. C. C. Maduka and Z. S. C. Okoye, "The effect of *Sacoglottis gabonensis* stem bark extract, a Nigerian alcoholic beverage additive, on the natural antioxidant defences during 2,4-dinitrophenyl hydrazine-induced membrane peroxidation in vivo," *Vascular Pharmacology*, vol. 39, no. 1-2, pp. 21–31, 2002.
- [252] H. C. C. Maduka, Z. S. C. Okoye, and A. Eje, "The influence of *Sacoglottis gabonensis* stem bark extract and its isolate bergenin, Nigerian alcoholic beverage additives, on the metabolic and haematological side effects of 2,4-dinitrophenyl hydrazine-induced tissue damage," *Vascular Pharmacology*, vol. 39, no. 6, pp. 317–324, 2002.
- [253] H. Maduka and Z. Okoye, "The Effect of *Sacoglottis gabonensis* and its Isolate Bergenin on Doxorubicin - Ferric Ions (Fe<sup>3+</sup>) - Induced Degradation of Deoxyribose," *Journal of Medical Sciences (Faisalabad)*, vol. 1, no. 5, pp. 316–319, 2001.
- [254] D. K. Patel, K. Patel, R. Kumar, M. Gadewar, and V. Tahilyani, "Pharmacological and analytical aspects of bergenin: A concise report," *Asian Pacific Journal of Tropical Disease*, vol. 2, no. 2, pp. 163–167, 2012.
- [255] V. D. P. Nair, A. Dairam, A. Agbonon, J. T. Arnason, B. C. Foster, and I. Kanfer, "Investigation of the antioxidant activity of African potato (*Hypoxis hemerocallidea*)," *Journal of Agricultural and Food Chemistry*, vol. 55, no. 5, pp. 1707–1711, 2007.
- [256] P. M. O. Owira and J. A. O. Ojewole, "'African potato' (*Hypoxis hemerocallidea* corm): A plant-medicine for modern and 21st century diseases of mankind? - A review," *Phytotherapy Research*, vol. 23, no. 2, pp. 147–152, 2009.
- [257] M. J. Van Der Merwe, K. Jenkins, E. Theron, and B. J. Van Der Walt, "Interaction of the di-catechols rooperol and nordihydroguaiaretic acid with oxidative systems in the human blood: a structure-activity relationship," *Biochemical Pharmacology*, vol. 45, no. 2, pp. 303–311, 1993.
- [258] A. C. Akinmoladun, E. O. Ibukun, E. Afor, E. M. Obuotor, and E. O. Farombi, "Phytochemical constituent and antioxidant activity of extract from the leaves of *Ocimum gratissimum*," *Scientific Research and Essays*, vol. 2, no. 5, pp. 163–166, 2007.
- [259] R. J. Grayer, G. C. Kite, M. Abou-Zaid, and L. J. Archer, "The application of atmospheric pressure chemical ionisation liquid chromatography-mass spectrometry in the chemotaxonomic study of flavonoids: Characterisation of flavonoids from *Ocimum gratissimum* var. *gratissimum*," *Phytochemical Analysis*, vol. 11, no. 4, pp. 257–267, 2000.
- [260] O. A. Odukoya, O. O. Ilori, M. O. Sofidiya, O. A. Aniunoh, B. M. Lawal, and I. O. Tade, "Antioxidant activity of Nigerian dietary spices," *Electronic Journal of Environmental, Agricultural and Food Chemistry*, vol. 4, pp. 1086–1093, 2005.
- [261] B. Prakash, R. Shukla, P. Singh, P. K. Mishra, N. K. Dubey, and R. N. Kharwar, "Efficacy of chemically characterized *Ocimum gratissimum* L. essential oil as an antioxidant and a safe plant based antimicrobial against fungal and aflatoxin B1 contamination of spices," *Food Research International*, vol. 44, no. 1, pp. 385–390, 2011.
- [262] R. F. Vieira, R. J. Grayer, A. Paton, and J. E. Simon, "Genetic diversity of *Ocimum gratissimum* L. based on volatile oil constituents, flavonoids and RAPD markers," *Biochemical Systematics and Ecology*, vol. 29, no. 3, pp. 287–304, 2001.
- [263] G. K. Jayaprakasha, P. S. Negi, B. S. Jena, and L. J. M. Rao, "Antioxidant and antimutagenic activities of *Cinnamomum zeylanicum* fruit extracts," *Journal of Food Composition and Analysis*, vol. 20, no. 3-4, pp. 330–336, 2007.
- [264] A. K. Jäger and J. Van Staden, "Cyclooxygenase inhibitory activity of South African plants used against inflammation," *Phytochemistry Reviews*, vol. 4, no. 1, pp. 39–46, 2005.
- [265] M. Zabka, R. Pavela, and L. Slezakova, "Antifungal effect of *Pimenta dioica* essential oil against dangerous pathogenic and toxinogenic fungi," *Industrial Crops and Products*, vol. 30, no. 2, pp. 250–253, 2009.
- [266] É. Palé, M. Kouda-Bonafos, and M. Nacro, "Caractérisation et mesure des activités anti-radicalaires d'anthocyanes de plantes du Burkina Faso," *Comptes Rendus Chimie*, vol. 7, no. 10-11, pp. 973–980, 2004.
- [267] E. O. Farombi and A. Fakoya, "Free radical scavenging and antigenotoxic activities of natural phenolic compounds in dried flowers of *Hibiscus sabdariffa* L," *Molecular Nutrition & Food Research*, vol. 49, no. 12, pp. 1120–1128, 2005.
- [268] E. Prenesti, S. Berto, P. G. Daniele, and S. Toso, "Antioxidant power quantification of decoction and cold infusions of *Hibiscus sabdariffa* flowers," *Food Chemistry*, vol. 100, no. 2, pp. 433–438, 2007.
- [269] C.-J. Wang, J.-M. Wang, W.-L. Lin, C.-Y. Chu, F.-P. Chou, and T.-H. Tseng, "Protective effect of *Hibiscus* anthocyanins against tert-butyl hydroperoxide-induced hepatic toxicity in rats," *Food and Chemical Toxicology*, vol. 38, no. 5, pp. 411–416, 2000.
- [270] M. P. Germanò, V. D'Angelo, R. Sanogo, A. Morabito, S. Pergolizzi, and R. De Pasquale, "Hepatoprotective activity of *Trichilia roka* on carbon tetrachloride-induced liver damage in rats," *Journal of Pharmacy and Pharmacology*, vol. 53, no. 11, pp. 1569–1574, 2001.

- [271] O. Nana, J. Momeni Nzangué, R. Tepongning, M. B. Ngassoum, and J. Momeni Nzangué, "Phytochemical screening, antioxidant and antiplasmodial activities of extracts from *Trichilia roka* and *Sapium ellipticum*," *The Journal of Phytopharmacology*, vol. 2, pp. 22–29, 2013.
- [272] J. O. Moody, V. A. Robert, J. D. Connolly, and P. J. Houghton, "Anti-inflammatory activities of the methanol extracts and an isolated furanoditerpene constituent of *Sphenocentrum jollyanum* Pierre (Menispermaceae)," *Journal of Ethnopharmacology*, vol. 104, no. 1-2, pp. 87–91, 2006.
- [273] O. S. Olorunnisola, A. O. Akintola, and A. J. Afolayan, "Hepatoprotective and antioxidant effect of *Sphenocentrum jollyanum* (Menispermaceae) stem bark extract against CCl<sub>4</sub>-induced oxidative stress in rats," *African Journal of Pharmacy and Pharmacology*, vol. 5, no. 9, pp. 1241–1246, 2011.
- [274] O. S. Olorunnisola and A. J. Afolayan, "In vivo antioxidant and biochemical evaluation of *Sphenocentrum jollyanum* leaf extract in *P. berghei* infected mice," *Pakistan Journal of Pharmaceutical Sciences*, vol. 26, no. 3, pp. 445–450, 2013.
- [275] S. I. Abdelwahab, W. S. Koko, M. M. E. Taha et al., "In vitro and in vivo anti-inflammatory activities of columbin through the inhibition of cyclooxygenase-2 and nitric oxide but not the suppression of NF-κB translocation," *European Journal of Pharmacology*, vol. 678, no. 1–3, pp. 61–70, 2012.
- [276] N. O. A. Omisore, C. O. Adewunmi, E. O. Iwalewa et al., "Antitrichomonad and antioxidant activities of *Dorstenia barteri* and *Dorstenia convexa*," *Brazilian Journal of Medical and Biological Research*, vol. 38, no. 7, pp. 1087–1094, 2005.
- [277] V. Kuete, B. Ngameni, A. T. Mbaveng, B. Ngadjui, J. J. M. Meyer, and N. Lall, "Evaluation of flavonoids from *Dorstenia barteri* for their antimycobacterial, antigonorrhreal and anti-reverse transcriptase activities," *Acta Tropica*, vol. 116, no. 1, pp. 100–104, 2010.
- [278] B. T. Ngadjui, J. Watchueng, F. Keumedjio, B. Ngameni, I. K. Simo, and B. M. Abegaz, "Prenylated chalcones, flavone and other constituents of the twigs of *Dorstenia angusticornis* and *Dorstenia barteri* var. *subtriangularis*," *Phytochemistry*, vol. 66, no. 6, pp. 687–692, 2005.
- [279] B. Ngameni, B. T. Ngadjui, G. N. Folefoc, J. Watchueng, and B. M. Abegaz, "Diprenylated chalcones and other constituents from the twigs of *Dorstenia barteri* var. *subtriangularis*," *Phytochemistry*, vol. 65, no. 4, pp. 427–432, 2004.
- [280] N. O. A. Omisore, C. O. Adewunmi, E. O. Iwalewa et al., "Antinociceptive and anti-inflammatory effects of *Dorstenia barteri* (Moraceae) leaf and twig extracts in mice," *Journal of Ethnopharmacology*, vol. 95, no. 1, pp. 7–12, 2004.
- [281] A. Tsopmo, M. Tene, P. Kamnaing, J. F. Ayafor, and O. Sterner, "A new Dieis-Alder-type adduct flavonoid from *Dorstenia barteri*," *Journal of Natural Products*, vol. 62, no. 10, pp. 1432–1434, 1999.
- [282] G. Kansci, E. Dongo, and C. Genot, "2,2-Diphenyl-1-picrylhydrazyl (DPPH•) test demonstrates antiradical activity of *Dorstenia psilurus* and *Dorstenia ciliata* plant extracts," *Molecular Nutrition & Food Research*, vol. 47, no. 6, pp. 434–437, 2003.
- [283] N. A. Al-Jaber, A. S. Awaad, and J. E. Moses, "Review on some antioxidant plants growing in Arab world," *Journal of Saudi Chemical Society*, vol. 15, no. 4, pp. 293–307, 2011.
- [284] B. T. Ngadjui, B. Ngameni, E. Dongo, S. F. Kouam, and B. M. Abegaz, "Prenylated and geranylated chalcones and flavones from the aerial parts of *Dorstenia ciliata*," *Bulletin of the Chemical Society of Ethiopia*, vol. 16, no. 2, pp. 157–163, 2002.
- [285] A. T. Mbaveng, V. Kuete, B. Ngameni et al., "Antimicrobial activities of the methanol extract and compounds from the twigs of *Dorstenia mannii* (Moraceae)," *BMC Complementary and Alternative Medicine*, vol. 12, no. 1, p. 83, 2012.
- [286] B. T. Ngadjui, B. M. Abegaz, E. Dongo, H. Tamboue, and F. Kouam, "Geranylated and prenylated flavonoids from the twigs of *Dorstenia mannii*," *Phytochemistry*, vol. 48, no. 2, pp. 349–354, 1998.
- [287] B. T. Ngadjui, S. F. Kouam, E. Dongo, G. W. F. Kapche, and B. M. Abegaz, "Prenylated flavonoids from the aerial parts of *Dorstenia mannii*," *Phytochemistry*, vol. 55, no. 8, pp. 915–919, 2000.
- [288] V. Kuete and L. P. Sandjo, "Isobavachalcone: An overview," *Chinese Journal of Integrative Medicine*, vol. 18, no. 7, pp. 543–547, 2012.
- [289] A. Tsopmo, M. Tene, P. Kamnaing, D. Ngnokam, J. F. Ayafor, and O. Sterner, "Geranylated flavonoids from *Dorstenia poinsettifolia*," *Phytochemistry*, vol. 48, no. 2, pp. 345–348, 1998.
- [290] B. M. Abegaz, "Novel natural products from marketed plants of eastern and southern Africa," *Pure and Applied Chemistry*, vol. 71, no. 6, pp. 919–926, 1999.
- [291] C. Etoundi, D. Kuaté, J. Ngondi, and J. Oben, "Anti-amylase, anti-lipase and antioxidant effects of aqueous extracts of some Cameroonian spices," *Journal of Natural Products*, vol. 3, no. 2010, pp. 165–171, 2010.
- [292] B. T. Ngadjui, T. K. Tabopda, E. Dongo, G. W. F. Kapche, P. Sandor, and B. M. Abegaz, "Dorsilurins C, D and E, three prenylated flavonoids from the roots of *Dorstenia psilurus*," *Phytochemistry*, vol. 52, no. 4, pp. 731–735, 1999.
- [293] S. Toyokuni, T. Tanaka, W. Kawaguchi et al., "Effects of the phenolic contents of Mauritian endemic plant extracts on promoter activities of antioxidant enzymes," *Free Radical Research*, vol. 37, no. 11, pp. 1215–1224, 2003.
- [294] S. A. Angaji, S. F. Mousavi, and E. Babapour, "Antioxidants: A few key points," *Annals of Biological Research*, vol. 3, no. 8, pp. 3968–3977, 2012.
- [295] V. S. Neergheen, T. Bahorun, L.-S. Jen, and O. I. Aruoma, "Bioefficacy of mauritian endemic medicinal plants: Assessment of their phenolic contents and antioxidant potential," *Pharmaceutical Biology*, vol. 45, no. 1, pp. 9–17, 2007.
- [296] D. Ramful, B. Aumjaud, V. S. Neergheen et al., "Polyphenolic content and antioxidant activity of *Eugenia pollicina* leaf extract in vitro and in model emulsion systems," *Food Research International*, vol. 44, no. 5, pp. 1190–1196, 2011.
- [297] H. Barakat, "Composition, antioxidant, antibacterial activities and mode of action of clove (*Syzygium aromaticum* L.) buds essential oil," *British Journal of Applied Science & Technology*, vol. 4, no. 13, p. 1934, 2014.
- [298] M. I. Nassar, A. H. Gaara, A. H. El-Ghorab et al., "Chemical constituents of clove (*Syzygium aromaticum*, Fam. Myrtaceae) and their antioxidant activity," *Revista Latinoamericana de Química*, vol. 35, no. 3, p. 47, 2007.
- [299] M. A. Abbasi, D. Shahwar, M. Wahab, and M. F. Saddiqui, "Antibacterial and antioxidant activities of an ethnobotanically important plant *Sauromatum venosum* (Ait.) Schott. of District Kotli, Azad Jammu & Kashmir," *Pakistan Journal of Botany*, vol. 43, no. 1, pp. 579–585, 2011.
- [300] O.-H. Lee and B.-Y. Lee, "Antioxidant and antimicrobial activities of individual and combined phenolics in *Olea europaea* leaf extract," *Bioresource Technology*, vol. 101, no. 10, pp. 3751–3754, 2010.

- [301] L. I. Somova, F. O. Shode, P. Ramnanan, and A. Nadar, "Antihypertensive, antiatherosclerotic and antioxidant activity of triterpenoids isolated from *Olea europaea*, subspecies *africana* leaves," *Journal of Ethnopharmacology*, vol. 84, no. 2-3, pp. 299–305, 2003.
- [302] A. Betancor-Fernández, A. Pérez-Gálvez, H. Sies, and W. Stahl, "Screening pharmaceutical preparations containing extracts of turmeric rhizome, artichoke leaf, devil's claw root and garlic or salmon oil for antioxidant capacity," *Journal of Pharmacy and Pharmacology*, vol. 55, no. 7, pp. 981–986, 2003.
- [303] I. M. Mahomed and J. A. O. Ojewole, "Analgesic, antiinflammatory and antidiabetic properties of *Harpagophytum procumbens* DC (Pedaliaceae) secondary root aqueous extract," *Phytotherapy Research*, vol. 18, no. 12, pp. 982–989, 2004.
- [304] J. J. Gagnier, S. Chrubasik, and E. Manheimer, "Harpagophytum procumbens for osteoarthritis and low back pain: a systematic review," *BMC Complementary and Alternative Medicine*, vol. 4, article 13, 2004.
- [305] H. Göbel, A. Heinze, M. Ingwersen, U. Niederberger, and D. Gerber, "Effects of Harpagophytum procumbens LI 174 (devil's claw) on sensory, motor and vascular muscle reactivity in the treatment of unspecific back pain," *Der Schmerz*, vol. 15, no. 1, pp. 10–18, 2001.
- [306] L. Grant, D. E. McBean, L. Fyfe, and A. M. Warnock, "A review of the biological and potential therapeutic actions of *Harpagophytum procumbens*," *Phytotherapy Research*, vol. 21, no. 3, pp. 199–209, 2007.
- [307] T. H.-W. Huang, V. H. Tran, R. K. Duke et al., "Harpagoside suppresses lipopolysaccharide-induced iNOS and COX-2 expression through inhibition of NF- $\kappa$ B activation," *Journal of Ethnopharmacology*, vol. 104, no. 1-2, pp. 149–155, 2006.
- [308] M. Kaszkin, K. F. Beck, E. Koch et al., "Downregulation of inos expression in rat mesangial cells by special extracts of *Harpagophytum procumbens* derives from harpagoside-dependent and independent effects," *Phytomedicine*, vol. 11, no. 7-8, pp. 585–595, 2004.
- [309] A. A. Elujoba, O. M. Odeleye, and C. M. Ogunnyemi, "Traditional Medical Development for medical and dental primary health care delivery system in Africa," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 2, no. 1, pp. 46–61, 2004.
- [310] I. M. Mahomed and J. A. O. Ojewole, "Anticonvulsant activity of *Harpagophytum procumbens* DC [Pedaliaceae] secondary root aqueous extract in mice," *Brain Research Bulletin*, vol. 69, no. 1, pp. 57–62, 2006.
- [311] I. M. Mahomed and J. A. O. Ojewole, "Oxytocin-like effect of *Harpagophytum procumbens* DC [Pedaliaceae] secondary root aqueous extract on rat isolated uterus," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 3, no. 1, pp. 82–89, 2006.
- [312] G. McGregor, B. Fiebich, A. Wartenberg, S. Brien, G. Lewith, and T. Wegener, "Devil's claw (*Harpagophytum procumbens*): an anti-inflammatory herb with therapeutic potential," *Phytochemistry Reviews*, vol. 4, no. 1, pp. 47–53, 2005.
- [313] G. A. Agbor, J. E. Oben, J. Y. Ngogang, G. Xinxing, and J. A. Vinson, "Antioxidant capacity of some herbs/spices from Cameroon: a comparative study of two methods," *Journal of Agricultural and Food Chemistry*, vol. 53, no. 17, pp. 6819–6824, 2005.
- [314] G. A. Agbor, J. A. Vinson, J. E. Oben, and J. Y. Ngogang, "In vitro antioxidant activity of three piper species," *Journal of Herbal Pharmacotherapy*, vol. 7, no. 2, pp. 49–64, 2007.
- [315] E. U. Isong and I. B. Essien, "Nutrient and antinutrient composition of three varieties of *Piper* species," *Plant Foods for Human Nutrition*, vol. 49, no. 2, pp. 133–137, 1996.
- [316] K. S. Natarajan, M. Narasimhan, K. R. Shanmugasundaram, and E. R. B. Shanmugasundaram, "Antioxidant activity of a salt-spice-herbal mixture against free radical induction," *Journal of Ethnopharmacology*, vol. 105, no. 1-2, pp. 76–83, 2006.
- [317] H. S. Abdillahi, J. F. Finnie, and J. Van Staden, "Anti-inflammatory, antioxidant, anti-tyrosinase and phenolic contents of four *Podocarpus* species used in traditional medicine in South Africa," *Journal of Ethnopharmacology*, vol. 136, no. 3, pp. 496–503, 2011.
- [318] N. Erkan, G. Ayrancı, and E. Ayrancı, "Antioxidant activities of rosemary (*Rosmarinus Officinalis* L.) extract, blackseed (*Nigella sativa* L.) essential oil, carnosic acid, rosmarinic acid and sesamol," *Food Chemistry*, vol. 110, no. 1, pp. 76–82, 2008.
- [319] I. Meral, Z. Yener, T. Kahraman, and N. Mert, "Effect of *Nigella sativa* on Glucose Concentration, Lipid Peroxidation, Anti-Oxidant Defence System and Liver Damage in Experimentally-Induced Diabetic Rabbits," *Journal of Veterinary Medicine Series A*, vol. 48, no. 10, pp. 593–599, 2001.
- [320] T. Bahorun, F. Trotin, J. Pommery, J. Vasseur, and M. Pinkas, "Antioxidant activities of *Crataegus monogyna* extracts," *Planta Medica*, vol. 60, no. 4, pp. 323–328, 1994.
- [321] T. Bahorun, E. Aumjaud, H. Ramphul et al., "Phenolic constituents and antioxidant capacities of *Crataegus monogyna* (Hawthorn) callus extracts," *Molecular Nutrition & Food Research*, vol. 47, no. 3, pp. 191–198, 2003.
- [322] T. Bahorun, B. Gressier, F. Trotin et al., "Oxygen species scavenging activity of phenolic extracts from hawthorn fresh plant organs and pharmaceutical preparations," *Arzneimittel-Forschung/Drug Research*, vol. 46, no. 11, pp. 1086–1089, 1996.
- [323] J. Bernatoniene, R. Masteikova, D. Majiene et al., "Free radical-scavenging activities of crataegus monogyna extracts," *Medicina*, vol. 44, no. 9, pp. 706–712, 2008.
- [324] J. Breza, O. Dzurny, A. Borowka et al., "Efficacy and acceptability of Tadenan® (*Pygeum africanum* extract) in the treatment of benign prostatic hyperplasia (BPH): A multicentre trial in central Europe," *Current Medical Research and Opinion*, vol. 14, no. 3, pp. 127–139, 1998.
- [325] A. Ishani, R. MacDonald, D. Nelson, I. Rutks, and T. J. Wilt, "Pygeum africanum for the treatment of patients with benign prostatic hyperplasia: A systematic review and quantitative meta-analysis," *American Journal of Medicine*, vol. 109, no. 8, pp. 654–664, 2000.
- [326] M. Paubert-Braquet, A. Cave, R. Hocquemiller et al., "Effect of Pygeum africanum extract on A23187-stimulated production of lipoxygenase metabolites from human polymorphonuclear cells," *Journal of Lipid Mediators and Cell Signalling*, vol. 9, no. 3, pp. 285–290, 1994.
- [327] D. Wang, Y. Li, G. Hou et al., "Pygeum africanum: Effect on oxidative stress in early diabetes-induced bladder," *International Urology and Nephrology*, vol. 42, no. 2, pp. 401–408, 2010.
- [328] S. O. Adeola, T. A. Yahaya, B. Hafsatu et al., "Gastro-protective effect of crossopteryx febrifuga in wistar rats," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 8, no. 3, pp. 300–306, 2011.
- [329] F. Occhiuto, R. Sanogo, M. P. Germano, A. Keita, V. D'Angelo, and R. De Pasquale, "Effects of some Malian medicinal plants on the respiratory tract of guinea-pigs," *Journal of Pharmacy and Pharmacology*, vol. 51, no. 11, pp. 1299–1303, 1999.

- [330] F. A. Tomas-Barberan and K. Hostettmann, "A cytotoxic triterpenoid and flavonoids from *Crossopteryx febrifuga*," *Planta Medica*, vol. 54, no. 3, pp. 266–267, 1988.
- [331] V. Steenkamp, M. C. Gouws, M. Gulumian, E. E. Elgorashi, and J. Van Staden, "Studies on antibacterial, anti-inflammatory and antioxidant activity of herbal remedies used in the treatment of benign prostatic hyperplasia and prostatitis," *Journal of Ethnopharmacology*, vol. 103, no. 1, pp. 71–75, 2006.
- [332] M. Lis-Balchin, S. Hart, and E. Simpson, "Buchu (*Agathosma betulina* and *A. crenulata*, Rutaceae) essential oils: Their pharmacological action on guinea-pig ileum and antimicrobial activity on microorganisms," *Journal of Pharmacy and Pharmacology*, vol. 53, no. 4, pp. 579–582, 2001.
- [333] F. Chaaib, E. F. Queiroz, K. Ndjoko, D. Diallo, and K. Hostettmann, "Antifungal and antioxidant compounds from the root bark of *Fagara zanthoxyloides*," *Planta Medica*, vol. 69, no. 4, pp. 316–320, 2003.
- [334] O. Ngozi, O. Samson, and S. K. Akindele, "In vitro biochemical investigations of the effects of *Carica papaya* and *Fagara zanthoxyloides* on antioxidant status and sickle erythrocytes," *African Journal of Biochemistry Research*, vol. 5, no. 8, pp. 226–236, 2011.
- [335] W. M. Messmer, M. Tin-wa, H. H. S. Fong et al., "Fagaronine, a new tumor inhibitor isolated from *Fagara zanthoxyloides* lam. (Rutaceae)," *Journal of Pharmaceutical Sciences*, vol. 61, no. 11, pp. 1858–1859, 1972.
- [336] J. W. Ogwal-Okeng, C. Obua, and W. W. W. Anokbonggo, "Acute toxicity effects of the methanolic extract of *Fagara zanthoxyloides* (Lam.) root-bark," *African Health Sciences*, vol. 3, no. 3, pp. 124–126, 2003.
- [337] M. Getie, T. Gebre-Mariam, R. Rietz et al., "Evaluation of the anti-microbial and anti-inflammatory activities of the medicinal plants *Dodonaea viscosa*, *Rumex nervosus* and *Rumex abyssinicus*," *Fitoterapia*, vol. 74, no. 1, pp. 139–143, 2003.
- [338] M. Getie, T. Gebre-Mariam, R. Rietz, and R. H. H. Neubert, "Evaluation of the release profiles of flavonoids from topical formulations of the crude extract of the leaves of *Dodonea viscosa* (Sapindaceae)," *Die Pharmazie*, vol. 57, no. 5, pp. 320–322, 2002.
- [339] R. A. A. Mothana, S. A. A. Abdo, S. Hasson, F. M. N. Althawab, S. A. Z. Alaghbari, and U. Lindequist, "Antimicrobial, antioxidant and cytotoxic activities and phytochemical screening of some yemeni medicinal plants," *Evidence-Based Complementary and Alternative Medicine*, vol. 7, no. 3, pp. 323–330, 2010.
- [340] A. Rojas, S. Cruz, H. Ponce-Monter, and R. Mata, "Smooth muscle relaxing compounds from *Dodonaea viscosa*," *Planta Medica*, vol. 62, no. 2, pp. 154–159, 1996.
- [341] L. S. Teffo, M. A. Aderogba, and J. N. Eloff, "Antibacterial and antioxidant activities of four kaempferol methyl ethers isolated from *Dodonaea viscosa* Jacq. var. *angustifolia* leaf extracts," *South African Journal of Botany*, vol. 76, no. 1, pp. 25–29, 2010.
- [342] A. Opoku, M. Nethengwe, P. Dladla et al., "Larvicidal and antimalarial activity of some Zulu medicinal plants," *Planta Medica*, vol. 77, no. 12, 2011.
- [343] G. Beretta and R. M. Facino, "Recent advances in the assessment of the antioxidant capacity of pharmaceutical drugs: From in vitro to in vivo evidence," *Analytical and Bioanalytical Chemistry*, vol. 398, no. 1, pp. 67–75, 2010.
- [344] M. Carocho and I. C. F. R. Ferreira, "A review on antioxidants, prooxidants and related controversy: natural and synthetic compounds, screening and analysis methodologies and future perspectives," *Food and Chemical Toxicology*, vol. 51, no. 1, pp. 15–25, 2013.
- [345] M. E. Kellett, P. Greenspan, and R. B. Pegg, "Modification of the cellular antioxidant activity (CAA) assay to study phenolic antioxidants in a Caco-2 cell line," *Food Chemistry*, vol. 244, pp. 359–363, 2018.
- [346] K. M. Schaich, X. Tian, and J. Xie, "Reprint of "Hurdles and pitfalls in measuring antioxidant efficacy: A critical evaluation of ABTS, DPPH, and ORAC assays"," *Journal of Functional Foods*, vol. 18, pp. 782–796, 2015.
- [347] E. Niki, "Assessment of antioxidant capacity in vitro and in vivo," *Free Radical Biology & Medicine*, vol. 49, no. 4, pp. 503–515, 2010.
- [348] L. T. Dalvi, D. C. Moreira, R. Andrade, J. Ginani, A. Alonso, and M. Hermes-Lima, "Ellagic acid inhibits iron-mediated free radical formation," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 173, pp. 910–917, 2017.
- [349] P. M. Hanna and R. P. Mason, "Direct evidence for inhibition of free radical formation from Cu(I) and hydrogen peroxide by glutathione and other potential ligands using the EPR spin-trapping technique," *Archives of Biochemistry and Biophysics*, vol. 295, no. 1, pp. 205–213, 1992.
- [350] M. Valko, K. Jomova, C. J. Rhodes, K. Kuča, and K. Musilek, "Redox- and non-redox-metal-induced formation of free radicals and their role in human disease," *Archives of Toxicology*, vol. 90, no. 1, pp. 1–37, 2016.
- [351] M. Mohammadpour, M. Behjati, A. Sadeghi, and A. Fassihi, "Wound healing by topical application of antioxidant iron chelators: Kojic acid and deferiprone," *International Wound Journal*, vol. 10, no. 3, pp. 260–264, 2013.
- [352] I. A. Demyanenko, V. V. Zakharova, O. P. Ilyinskaya et al., "Mitochondria-Targeted Antioxidant SkQ1 Improves Dermal Wound Healing in Genetically Diabetic Mice," *Oxidative Medicine and Cellular Longevity*, vol. 2017, 2017.
- [353] X. Cao, Y. Wang, C. Wu et al., "Cathelicidin-OA1, a novel antioxidant peptide identified from an amphibian, accelerates skin wound healing," *Scientific Reports*, vol. 8, no. 1, 2018.
- [354] V. Hosur, L. M. Burzenski, T. M. Stearns et al., "Early induction of NRF2 antioxidant pathway by RHBD2 mediates rapid cutaneous wound healing," *Experimental and Molecular Pathology*, vol. 102, no. 2, pp. 337–346, 2017.
- [355] D. Son, D. Yang, J. Sun et al., "A Novel Peptide, Nicotinyl-Isoleucine–Valine–Histidine (NA-IVH), Promotes Antioxidant Gene Expression and Wound Healing in HaCaT Cells," *Marine Drugs*, vol. 16, no. 8, p. 262, 2018.
- [356] J. Benedí, R. Arroyo, C. Romero, S. Martín-Aragón, and A. M. Villar, "Antioxidant properties and protective effects of a standardized extract of *Hypericum perforatum* on hydrogen peroxide-induced oxidative damage in PC12 cells," *Life Sciences*, vol. 75, no. 10, pp. 1263–1276, 2004.
- [357] D. Diallo, A. Marston, C. Terreaux, Y. Toure, B. S. Paulsen, and K. Hostettmann, "Screening of malian medicinal plants for antifungal, larvicidal, molluscicidal, antioxidant and radical scavenging activities," *Phytotherapy Research*, vol. 15, no. 5, pp. 401–406, 2001.
- [358] R. R. T. Majinda, P. Erasto, and G. Bojase-Moleta, "Antimicrobial and antioxidant flavonoids from the root wood of *Bolusanthus speciosus*," *Phytochemistry*, vol. 65, no. 7, pp. 875–880, 2004.
- [359] B. F. Juma and R. R. T. Majinda, "Erythrinoline alkaloids from the flowers and pods of *Erythrina lysistemon* and their DPPH radical scavenging properties," *Phytochemistry*, vol. 65, no. 10, pp. 1397–1404, 2004.

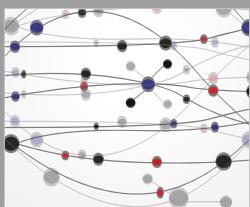
- [360] G. P. P. Kamatou, A. M. Viljoen, A. B. Gono-Bwalya et al., "The in vitro pharmacological activities and a chemical investigation of three South African *Salvia* species," *Journal of Ethnopharmacology*, vol. 102, no. 3, pp. 382–390, 2005.
- [361] D. R. Katerere and J. N. Eloff, "Variation in chemical composition, antibacterial and antioxidant activity of fresh and dried *Acacia* leaf extracts," *South African Journal of Botany*, vol. 70, no. 2, pp. 303–305, 2004.
- [362] K. P. Latté and H. Kolodziej, "Antioxidant properties of phenolic compounds from *Pelargonium reniforme*," *Journal of Agricultural and Food Chemistry*, vol. 52, no. 15, pp. 4899–4902, 2004.
- [363] C. S. Nergard, D. Diallo, K. Inngjerdingen et al., "Medicinal use of *Cochlospermum tinctorium* in Mali: Anti-ulcer-, radical scavenging- and immunomodulating activities of polymers in the aqueous extract of the roots," *Journal of Ethnopharmacology*, vol. 96, no. 1-2, pp. 255–269, 2005.
- [364] A. R. Opoku, N. F. Maseko, and S. E. Terblanche, "The in vitro antioxidative activity of some traditional Zulu medicinal plants," *Phytotherapy Research*, vol. 16, no. 1, pp. S51–S56, 2002.
- [365] M. Burits and F. Bucar, "Antioxidant activity of *Nigella sativa* essential oil," *Phytotherapy Research*, vol. 14, no. 5, pp. 323–328, 2000.
- [366] M. A. Gyamfi and Y. Aniya, "Antioxidant properties of Thonningianin A, isolated from the African medicinal herb, *Thonningia sanguinea*," *Biochemical Pharmacology*, vol. 63, no. 9, pp. 1725–1737, 2002.
- [367] E. O. Farombi and I. A. Nwaokeafor, "Anti-oxidant mechanisms of kolaviron: Studies on serum lipoprotein oxidation, metal chelation and oxidative membrane damage in rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 32, no. 8, pp. 667–674, 2005.
- [368] A. Luximon-Ramma, T. Bahorun, M. A. Soobrattee, and O. I. Aruoma, "Antioxidant activities of phenolic, proanthocyanidin, and flavonoid components in extracts of *Cassia fistula*," *Journal of Agricultural and Food Chemistry*, vol. 50, no. 18, pp. 5042–5047, 2002.
- [369] S. Singh, V. Parasharami, and S. Rai, "Medicinal uses of *adansonia digitata* L.: An endangered tree species," *Journal of Pharmaceutical and Scientific Innovation*, vol. 2, no. 3, pp. 14–16, 2013.
- [370] F. Bucar, M. Resch, R. Bauer, M. Burits, E. Knauder, and M. Schubert-Zsilavecz, "5-methylflavasperone and rhamnetin from *Guiera senegalensis* and their antioxidative and 5-lipoxygenase inhibitory activity," *Die Pharmazie*, vol. 30, no. 13, 1999.
- [371] F. Bucar, I. Schneider, H. Ögmundsdóttir, and K. Ingólfssdóttir, "Anti-proliferative lichen compounds with inhibitory activity on 12(S)-HETE production in human platelets," *Phytomedicine*, vol. 11, no. 7-8, pp. 602–606, 2004.
- [372] I. Schneider and F. Bucar, "Lipoxygenase inhibitors from natural plant sources, part 1: medicinal plants with inhibitory activity on arachidonate 5-lipoxygenase and 5-lipoxygenase/cyclooxygenase," *Phytotherapy Research*, vol. 19, no. 2, pp. 81–102, 2005.
- [373] A. A. Wube, B. Streit, S. Gibbons, K. Asres, and F. Bucar, "In vitro 12(S)-HETE inhibitory activities of naphthoquinones isolated from the root bark of *Euclea racemosa* ssp. *schimperi*," *Journal of Ethnopharmacology*, vol. 102, no. 2, pp. 191–196, 2005.
- [374] J. Lin, A. R. Opoku, M. Geheeb-Keller et al., "Preliminary screening of some traditional zulu medicinal plants for anti-inflammatory and anti-microbial activities," *Journal of Ethnopharmacology*, vol. 68, no. 1-3, pp. 267–274, 1999.
- [375] A. Hiermann and F. Bucar, "Influence of some traditional medicinal plants of senegal on prostaglandin biosynthesis," *Journal of Ethnopharmacology*, vol. 42, no. 2, pp. 111–116, 1994.
- [376] A. K. Jäger, A. Hutchings, and J. Van Staden, "Screening of Zulu medicinal plants for prostaglandin-synthesis inhibitors," *Journal of Ethnopharmacology*, vol. 52, no. 2, pp. 95–100, 1996.
- [377] L. Krenn, G. Beyer, H. H. Pertz et al., "In vitro antispasmodic and anti-inflammatory effects of *Drosera rotundifolia*," *Arzneimittel-Forschung/Drug Research*, vol. 54, no. 7, pp. 402–405, 2004.
- [378] M. Vasänge, B. Liu, C. J. Welch, W. Rolfsen, and L. Bohlin, "The flavonoid constituents of two *Polypodium* species (*Calaguala*) and their effect on the elastase release in human neutrophils," *Planta Medica*, vol. 63, no. 6, pp. 511–517, 1997.
- [379] J. B. Calixto, M. F. Otuki, and A. R. S. Santos, "Anti-inflammatory compounds of plant origin. part i. action on arachidonic acid pathway, nitric oxide and nuclear factor κB (NF-κB)," *Planta Medica*, vol. 69, no. 11, pp. 973–983, 2003.
- [380] E. A. Ojo-Amaize, P. Kapahi, V. N. Kakkanaiah et al., "Hypoestoxide, a novel anti-inflammatory natural diterpene, inhibits the activity of IκB kinase," *Cellular Immunology*, vol. 209, no. 2, pp. 149–157, 2001.
- [381] E. A. Ojo-Amaize, E. J. Nchekwube, H. B. Cottam et al., "Hypoestoxide, a natural nonmutagenic diterpenoid with antiangiogenic and antitumor activity: Possible mechanisms of action," *Cancer Research*, vol. 62, no. 14, pp. 4007–4014, 2002.
- [382] G. Chernishov, M. Arragie, and A. Etana, "Preliminary pharmacological studies on Mettere (*Glinus lotoides*). II. Effects upon the cardiovascular and gastrointestinal system," *Ethiopian Medical Journal*, vol. 16, no. 3, pp. 105–110, 1978.
- [383] A. E. Mengesha, *Isolation, Structural Elucidation, Quantification and Formulation of the Saponins and Flavonoids of the Seeds of Glinus Lotoides*, 2005.
- [384] A. Endale, B. Kammerer, T. Gebre-Mariam, and P. C. Schmidt, "Quantitative determination of the group of flavonoids and saponins from the extracts of the seeds of *Glinus lotoides* and tablet formulation thereof by high-performance liquid chromatography," *Journal of Chromatography A*, vol. 1083, no. 1-2, pp. 32–41, 2005.
- [385] A. Endale, P. C. Schmidt, and T. Gebre-Mariam, "Standardisation and physicochemical characterisation of the extracts of seeds of *Glinus lotoides*," *Die Pharmazie*, vol. 59, no. 1, pp. 34–38, 2004.
- [386] M. El-Sayed, "Phytochemical investigation of *Glinus lotoides* growing in Egypt," *Egyptian journal of pharmaceutical sciences*, vol. 38, no. 4-6, pp. 377–390, 1997.
- [387] D. Diallo, B. Hveem, M. Ag Mahmoud, G. Berge, B. S. Paulsen, and A. Maiga, "An ethnobotanical survey of herbal drugs of Gourma district, Mali," *Pharmaceutical Biology*, vol. 37, no. 1, pp. 80–91, 1999.
- [388] P. Sahakitpichan, W. Disadee, S. Ruchirawat, and T. Kanchanapoom, "L-(--)-(N-trans-Cinnamoyl)-arginine, an Acyl-amino Acid from *Glinus oppositifolius* (L.) Aug. DC," *Molecules*, vol. 15, no. 9, pp. 6186–6192, 2010.
- [389] J. A. O. Ojewole, "Evaluation of the analgesic, anti-inflammatory and anti-diabetic properties of *Sclerocarya birrea* (A. Rich.) Hochst. stem-bark aqueous extract in mice and rats," *Phytotherapy Research*, vol. 18, no. 8, pp. 601–608, 2004.
- [390] E. Agbaje, A. Tijani, and O. Braimoh, "Effects of Enantia chlorantha extracts in Laboratory-Induced Convulsion and

- Inflammation," *Orient Journal of Medicine*, vol. 15, no. 1, pp. 68–71, 2004.
- [391] R. F. Atata, S. Alhassan, and S. M. Ajewole, "Effect of stem bark extracts of *Enantia chlorantha* on some clinical isolates," *Bioekemistri*, vol. 15, no. 2, pp. 84–92, 2003.
- [392] J. O. Moody, S. F. Bloomfield, and P. J. Hylands, "In-vitro evaluation of the antimicrobial activities of *Enantia chlorantha* Oliv. extractives," *African Journal of Medicine and Medical Sciences*, vol. 24, no. 3, pp. 269–273, 1995.
- [393] P. V. Tan, B. Nyasse, T. Dimo, P. Wafo, and B. T. Akahkuh, "Synergistic and potentiating effects of ranitidine and two new anti-ulcer compounds from *Enantia chlorantha* and *Voacanga africana* in experimental animal models," *Die Pharmazie*, vol. 57, no. 6, pp. 409–412, 2002.
- [394] A. M. Koffi, C. Kanko, H. Ramiarantsoa et al., "Essentials oils phenolic and benzenic derivatives from three *Uvaria* (Annonaceae) of Ivory Coast: *Uvaria chamae* (P. Beauv), *Uvaria afzelii* (Sc. Elliot), and *Uvaria* sp. (Aké Assi)," *Comptes Rendus Chimie*, vol. 7, no. 10-11, pp. 997–1002, 2004.
- [395] H. Ménan, J.-T. Banzouzi, A. Hocquette et al., "Antiplasmodial activity and cytotoxicity of plants used in West African traditional medicine for the treatment of malaria," *Journal of Ethnopharmacology*, vol. 105, no. 1-2, pp. 131–136, 2006.
- [396] R. I. Uchegbu and D. E. Okwu, "An Evaluation of the Phytochemical and Nutrient Composition of the Seeds and Stem bark of *Detarium senegalense* Gmelin," *Journal of Natural Science Research*, vol. 2, no. 5, pp. 107–111, 2012.
- [397] D. Fall, C. Gleye, X. Franck, A. Laurens, and R. Hocquemiller, "Cis-bullatencin, a linear acetogenin from roots of *Uvaria chamae*," *Natural Product Research (Formerly Natural Product Letters)*, vol. 16, no. 5, pp. 315–321, 2002.
- [398] S. Philipov, N. Ivanovska, R. Istatkova, M. Velikova, and P. Tuleva, "Phytochemical study and cytotoxic activity of alkaloids from *Uvaria chamae* P. Beauv," *Die Pharmazie*, vol. 55, no. 9, pp. 688–689, 2000.
- [399] M. Duwiejua, E. Woode, and D. D. Obiri, "Pseudoakuammigine, an alkaloid from *Picralima nitida* seeds, has anti-inflammatory and analgesic actions in rats," *Journal of Ethnopharmacology*, vol. 81, no. 1, pp. 73–79, 2002.
- [400] I. C. Ezeamuzie, M. C. Ojinnaka, E. O. Uzogara, and S. E. Oji, "Anti-inflammatory, antipyretic and anti-malarial activities of a West African medicinal plant—*Picralima nitida*," *African Journal of Medicine and Medical Sciences*, vol. 23, no. 1, pp. 85–90, 1994.
- [401] J. Betti, *An ethnobotanical study of medicinal plants among the Baka pygmies in the Dja biosphere reserve, Cameroon*, 2004.
- [402] T. O. Fakaye, O. A. Itiola, and H. A. Odelola, "Evaluation of the antimicrobial property of the stem bark of *Picralima nitida* (Apocynaceae)," *Phytotherapy Research*, vol. 14, no. 5, pp. 368–370, 2000.
- [403] S. Papajewski, B. Vogler, J. Conrad et al., "Isolation from *Cussonia barteri* of 1 $\beta$ -O-chlorogenoylchlorogenic acid and 1 $\beta$ -o-chlorogenoylneochlorogenic acid, a new type of quinic acid esters," *Planta Medica*, vol. 67, no. 8, pp. 732–736, 2001.
- [404] S. Roy, R. Sehgal, B. M. Padhy, and V. L. Kumar, "Antioxidant and protective effect of latex of *Calotropis procera* against alloxan-induced diabetes in rats," *Journal of Ethnopharmacology*, vol. 102, no. 3, pp. 470–473, 2005.
- [405] N. H. Ugochukwu and N. E. Babady, "Antioxidant effects of *Gongronema latifolium* in hepatocytes of rat models of non-insulin dependent diabetes mellitus," *Fitoterapia*, vol. 73, no. 7-8, pp. 612–618, 2002.
- [406] N. H. Ugochukwu and N. E. Babady, "Antihyperglycemic effect of aqueous and ethanolic extracts of *Gongronema latifolium* leaves on glucose and glycogen metabolism in livers of normal and streptozotocin-induced diabetic rats," *Life Sciences*, vol. 73, no. 15, pp. 1925–1938, 2003.
- [407] N. H. Ugochukwu, N. E. Babady, M. Cobourne, and S. R. Gasset, "The effect of *Gongronema latifolium* extracts on serum lipid profile and oxidative stress in hepatocytes of diabetic rats," *Journal of Biosciences*, vol. 28, no. 1, pp. 1–5, 2003.
- [408] R. H. Nébié, R. T. Yaméogo, A. Bélanger, and F. S. Sib, "Composition chimique des huiles essentielles d'*Ageratum conyzoides* du Burkina Faso," *Comptes Rendus Chimie*, vol. 7, no. 10-11, pp. 1019–1022, 2004.
- [409] A. Shirwaikar, P. M. Bhilegaonkar, S. Malini, and J. Sharath Kumar, "The gastroprotective activity of the ethanol extract of *Ageratum conyzoides*," *Journal of Ethnopharmacology*, vol. 86, no. 1, pp. 117–121, 2003.
- [410] C. Z. Liu, S. J. Murch, M. El-Demerdash, and P. K. Saxena, "Artemisia judaica L.: Micropropagation and antioxidant activity," *Journal of Biotechnology*, vol. 110, no. 1, pp. 63–71, 2004.
- [411] A. Popat, N. H. Shear, I. Malkiewicz et al., "The toxicity of *Callilepis laureola*, a South African traditional herbal medicine," *Clinical Biochemistry*, vol. 34, no. 3, pp. 229–236, 2001.
- [412] V. Steenkamp, M. J. Stewart, and M. Zuckerman, "Detection of poisoning by impila (*Callilepis laureola*) in a mother and child," *Human & Experimental Toxicology*, vol. 18, no. 10, pp. 594–597, 1999.
- [413] J. O. Midiwo, A. Yenesew, B. F. Juma et al., "Bioactive compounds from some Kenyan ethnomedicinal plants: myrsinaceae, polygonaceae and psadia punctulata," *Phytochemistry Reviews*, vol. 1, no. 3, pp. 311–323, 2002.
- [414] C. S. Nergard, D. Diallo, T. E. Michaelsen et al., "Isolation, partial characterisation and immunomodulating activities of polysaccharides from *Vernonia kotschyana* Sch. Bip. ex Walp," *Journal of Ethnopharmacology*, vol. 91, no. 1, pp. 141–152, 2004.
- [415] M. William Carey, N. V. Rao, B. R. Kumar, and G. K. Mohan, "Anti-inflammatory and analgesic activities of methanolic extract of *Kigelia pinnata* DC flower," *Journal of Ethnopharmacology*, vol. 130, no. 1, pp. 179–182, 2010.
- [416] Y. G. Gouda, A. M. Abdel-baky, F. M. Darwish, K. M. Mohamed, R. Kasai, and K. Yamasaki, "Iridoids from *Kigelia pinnata* DC. fruits," *Phytochemistry*, vol. 63, no. 8, pp. 887–892, 2003.
- [417] J. S. Reddy, P. R. Rao, and M. S. Reddy, "Wound healing effects of *Heliotropium indicum*, *Plumbago zeylanicum* and *Acalypha indica* in rats," *Journal of Ethnopharmacology*, vol. 79, no. 2, pp. 249–251, 2002.
- [418] J. S. N. Souza, L. L. Machado, O. D. L. Pessoa et al., "Pyrrolizidine alkaloids from *Heliotropium indicum*," *Journal of the Brazilian Chemical Society*, vol. 16, no. 6 B, pp. 1410–1414, 2005.
- [419] A. Togola, D. Diallo, S. Dembélé, H. Barsett, and B. S. Paulsen, "Ethnopharmacological survey of different uses of seven medicinal plants from Mali, (West Africa) in the regions Doila, Kolokani and Siby," *Journal of Ethnobiology and Ethnomedicine*, vol. 1, article 7, 2005.
- [420] A. M. Emam, R. Elias, A. M. Moussa, R. Faure, L. Debrauwer, and G. Balansard, "Two flavonoid triglycosides from *Buddleja madagascariensis*," *Phytochemistry*, vol. 48, no. 4, pp. 739–742, 1998.
- [421] L. O. A. Manguro, I. Ugi, R. Hermann, and P. Lemmen, "Flavonol and drimane-type sesquiterpene glycosides of *Warburgia stuhlmannii* leaves," *Phytochemistry*, vol. 63, no. 4, pp. 497–502, 2003.

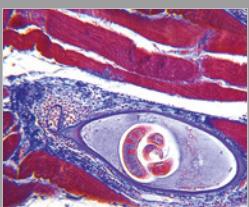
- [422] L. O. Arot Manguro, I. Ugi, P. Lemmen, and R. Hermann, "Flavonol glycosides of *Warburgia ugandensis* leaves," *Phytochemistry*, vol. 64, no. 4, pp. 891–896, 2003.
- [423] A. A. Wube, F. Bucar, K. Asres, S. Gibbons, L. Rattray, and S. L. Croft, "Antimalarial compounds from *Kniphofia foliosa* roots," *Phytotherapy Research*, vol. 19, no. 6, pp. 472–476, 2005.
- [424] A. A. Wube, F. Bucar, S. Gibbons, and K. Asres, "Sesquiterpenes from *Warburgia ugandensis* and their antimycobacterial activity," *Phytochemistry*, vol. 66, no. 19, pp. 2309–2315, 2005.
- [425] B. M. Abegaz and Y. Woldu, "Isoflavonoids from the roots of *Salsola somalensis*," *Phytochemistry*, vol. 30, no. 4, pp. 1281–1284, 1991.
- [426] M. P. Germanò, R. Sanogo, M. Guglielmo, R. De Pasquale, G. Crisafi, and G. Bisignano, "Effects of *Pteleopsis suberosa* extracts on experimental gastric ulcers and *Helicobacter pylori* growth," *Journal of Ethnopharmacology*, vol. 59, no. 3, pp. 167–172, 1998.
- [427] R. Nimenibo-Uadua, "Control of hyperlipidaemia, hypercholesterolaemia and hyperketonaemia by aqueous extract of *Dioscorea dumetorum* tuber," *Tropical Journal of Pharmaceutical Research*, vol. 2, no. 1, pp. 183–189, 2003.
- [428] E. Dagne, M. Alemua, and O. Sterner, "Flavonoids from *Euclea divinorum*," *Bulletin of the Chemical Society of Ethiopia*, vol. 7, no. 2, 1993.
- [429] S. K. Adesina, O. Idowu, A. O. Ogundaini et al., "Antimicrobial constituents of the leaves of *Acalypha wilkesiana* and *Acalypha hispida*," *Phytotherapy Research*, vol. 14, no. 5, pp. 371–374, 2000.
- [430] K. O. Akinyemi, O. Oladapo, C. E. Okwara, C. C. Ibe, and K. A. Fasure, "Screening of crude extracts of six medicinal plants used in South-West Nigerian unorthodox medicine for anti-methicillin resistant *Staphylococcus aureus* activity," *BMC Complementary and Alternative Medicine*, vol. 5, 2005.
- [431] J. Gálvez, M. E. Crespo, J. Jiménez, A. Suárez, and A. Zarzuelo, "Antidiarrhoeic activity of quercitrin in mice and rats," *Journal of Pharmacy and Pharmacology*, vol. 45, no. 2, pp. 157–159, 1993.
- [432] J. B. Harborne and C. A. Williams, "Anthocyanins and other flavonoids," *Natural Product Reports*, vol. 18, no. 3, pp. 310–333, 2001.
- [433] L. Bennie, J. Coetzee, E. Malan, and D. Ferreira, "(4→6)-coupled proteracacinidins and promelacacinidins from *Acacia galpinii* and *Acacia caffra*," *Phytochemistry*, vol. 60, no. 5, pp. 521–532, 2002.
- [434] L. Bennie, J. Coetzee, E. Malan, and D. Ferreira, "Structure and stereochemistry of dimeric proteracacinidins possessing the rare C-4(C) → C-5(D) interflavanyl linkage," *Phytochemistry*, vol. 59, no. 6, pp. 673–678, 2002.
- [435] L. Bennie, E. Malan, J. Coetzee, and D. Ferreira, "Structure and synthesis of ether-linked proteracacinidin and promelacacinidin proanthocyanidins from *Acacia caffra*," *Phytochemistry*, vol. 53, no. 7, pp. 785–793, 2000.
- [436] O. A. Binutu and G. A. Cordell, "Constituents of *Afzelia bella* stem bark," *Phytochemistry*, vol. 56, no. 8, pp. 827–830, 2001.
- [437] G. Bojase, C. C. W. Wanjala, and R. R. T. Majinda, "Flavonoids from the stem bark of *Bolusanthus speciosus*," *Phytochemistry*, vol. 56, no. 8, pp. 837–841, 2001.
- [438] C. Ito, M. Itoigawa, N. Kojima et al., "Cancer chemopreventive activity of rotenoids from *Derris trifoliata*," *Planta Medica*, vol. 70, no. 6, pp. 585–588, 2004.
- [439] A. Yenesew, J. T. Kiplagat, S. Derese et al., "Two unusual rotenoid derivatives, 7a-O-methyl-12a-hydroxydeguelol and spiro-13-homo-13-oxaelliptone, from the seeds of *Derris trifoliata*," *Phytochemistry*, vol. 67, no. 10, pp. 988–991, 2006.
- [440] A. Yenesew, E. K. Mushibe, M. Induli et al., "7a-O-methyldeguelol, a modified rotenoid with an open ring-C, from the roots of *Derris trifoliata*," *Phytochemistry*, vol. 66, no. 6, pp. 653–657, 2005.
- [441] S. Ragusa, R. De Pasquale, M. Flores, M. P. Germanò, R. Sanogo, and A. Rapisarda, "Micromorphological investigations on *Entada africana* Guill. et Perr. (Mimosaceae)," *Farmaco*, vol. 56, no. 5–7, pp. 361–363, 2001.
- [442] R. Sanogo, M. P. Germanò, V. D'Angelo, M. Guglielmo, and R. De Pasquale, "Antihepatotoxic properties of *Entada africana* (Mimosaceae)," *Phytotherapy Research*, vol. 12, no. 1, pp. S157–S159, 1998.
- [443] J. O. Kokwaro, *Medicinal plants of east Africa*, University of Nairobi press, 2009.
- [444] A. Yenesew, M. Induli, S. Derese et al., "Anti-plasmodial flavonoids from the stem bark of *Erythrina abyssinica*," *Phytochemistry*, vol. 65, no. 22, pp. 3029–3032, 2004.
- [445] A. Yenesew, S. Derese, J. O. Midiwo, C. C. Bii, M. Heydenreich, and M. G. Peter, "Antimicrobial flavonoids from the stem bark of *Erythrina burttii*," *Fitoterapia*, vol. 76, no. 5, pp. 469–472, 2005.
- [446] A. Yenesew, B. Irungu, S. Derese, J. O. Midiwo, M. Heydenreich, and M. G. Peter, "Two prenylated flavonoids from the stem bark of *Erythrina burttii*," *Phytochemistry*, vol. 63, no. 4, pp. 445–448, 2003.
- [447] A. Yenesew, J. O. Midiwo, S. M. Guchu, M. Heydenreich, and M. G. Peter, "Three isoflav-3-enes and a 2-arylbenzofuran from the root bark of *Erythrina burttii*," *Phytochemistry*, vol. 59, no. 3, pp. 337–341, 2002.
- [448] A. Yenesew, J. O. Midiwo, M. Miessner, M. Heydenreich, and M. G. Peter, "Two prenylated flavanones from stem bark of *Erythrina burttii*," *Phytochemistry*, vol. 48, no. 8, pp. 1439–1443, 1998.
- [449] A. E. Nkengfack, J. C. Vardamides, Z. T. Fomum, and M. Meyer, "Prenylated isoflavanone from *Erythrina eriota*," *Phytochemistry*, vol. 40, no. 6, pp. 1803–1808, 1995.
- [450] A. E. Nkengfack, T. W. Vouffo, J. C. Vardamides et al., "Phenolic metabolites from *Erythrina* species," *Phytochemistry*, vol. 46, no. 3, pp. 573–578, 1997.
- [451] A. W. Andayi, A. Yenesew, S. Derese et al., "Antiplasmodial flavonoids from *Erythrina sacleuxii*," *Planta Medica*, vol. 72, no. 2, pp. 187–189, 2006.
- [452] A. Yenesew, J. O. Midiwo, M. Heydenreich, and M. G. Peter, "Four isoflavones from the stem bark of *Erythrina sacleuxii*," *Phytochemistry*, vol. 49, no. 1, pp. 247–249, 1998.
- [453] A. Yenesew, J. O. Midiwo, M. Heydenreich, D. Schanzenbach, and M. G. Peter, "Two isoflavanones from the stem bark of *Erythrina sacleuxii*," *Phytochemistry*, vol. 55, no. 5, pp. 457–459, 2000.
- [454] S. Derese, A. Yenesew, J. O. Midiwo, M. Heydenreich, and M. G. Peter, "A new isoflavone from stem bark of *Millettia dura*," *Bulletin of the Chemical Society of Ethiopia*, vol. 17, no. 1, pp. 113–115, 2003.
- [455] E. Dagne, W. Mammo, and O. Sterner, "Flavonoids of *Tephrosia polypylla*," *Phytochemistry*, vol. 31, no. 10, pp. 3662–3663, 1992.
- [456] B. Niassy, B.-H. Um, A. Lobstein, B. Weniger, M. Koné, and R. Anton, "Flavonoids of *Tephrosia deflexa* and *Tephrosia albifoliolis*," *Comptes Rendus Chimie*, vol. 7, no. 10–11, pp. 993–996, 2004.
- [457] H. Duddeck, A. Yenesew, and E. Dagne, "Isoflavonoids from *Taverniera abyssinica*," *Bulletin of the Chemical Society of Ethiopia*, vol. 1, no. 1, p. pp, 1987.

- [458] B. K. Noamesi, M. Bogale, and E. Dagne, "Intestinal smooth muscle spasmolytic actions of the aqueous extract of the roots of *Taverniera abyssinica*," *Journal of Ethnopharmacology*, vol. 30, no. 1, pp. 107–113, 1990.
- [459] K. P. Latté, *Phytochemische und pharmakologische Untersuchungen an Pelargonium reniforme CURT*, 1999.
- [460] H. Wagner and S. Bladt, "Cumarine aus südafrikanischen Pelargonium-arten," *Phytochemistry*, vol. 14, no. 9, pp. 2061–2064, 1975.
- [461] S. E. Drewes, F. Khan, S. F. van Vuuren, and A. M. Viljoen, "Simple 1, 4-benzoquinones with antibacterial activity from stems and leaves of *Gunnera perpensa*," *Phytochemistry*, vol. 66, no. 15, pp. 1812–1816, 2005.
- [462] J. L. Ngondi, J. E. Oben, and S. R. Minka, "The effect of *Irvingia gabonensis* seeds on body weight and blood lipids of obese subjects in Cameroon," *Lipids in Health and Disease*, vol. 4, no. 1, 12 pages, 2005.
- [463] S. Vertuani, E. Braccioli, V. Buzzoni, and S. Manfredini, "Antioxidant capacity of *Adansonia digitata* fruit pulp and leaves," *Acta Phyttherapeutica*, vol. 2, no. 5, pp. 2–7, 2002.
- [464] A. M. D. El-Mousallamy, "Leaf flavonoids of *Albizia lebbeck*," *Phytochemistry*, vol. 48, no. 4, pp. 759–761, 1998.
- [465] B. M. Abegaz, B. T. Ngadjui, G. N. Folefoc et al., "Prenylated flavonoids, monoterpenoid furanocoumarins and other constituents from the twigs of *Dorstenia elliptica* (Moraceae)," *Phytochemistry*, vol. 65, no. 2, pp. 221–226, 2004.
- [466] B. M. Abegaz, B. T. Ngadjui, E. Dongo, and H. Tamboue, "Prenylated chalcones and flavones from the leaves of *Dorstenia kameruniana*," *Phytochemistry*, vol. 49, no. 4, pp. 1147–1150, 1998.
- [467] B. M. Abegaz, B. T. Ngadjui, E. Dongo, B. Ngameni, M. N. Nindi, and M. Bezabih, "Chalcones and other constituents of *Dorstenia prorprens* and *Dorstenia zenkeri*," *Phytochemistry*, vol. 59, no. 8, pp. 877–883, 2002.
- [468] A. H. El-Ghorab, K. F. El-Massry, F. Marx, and H. M. Fadel, "Antioxidant activity of Egyptian *Eucalyptus camaldulensis* var. *brevirostris* leaf extracts," *Molecular Nutrition Food Research*, vol. 47, no. 1, pp. 41–45, 2003.
- [469] I. C. Ezeamuzie, A. W. Ambakederemo, F. O. Shode, and S. C. Ekwебelem, "Antiinflammatory effects of *Moringa oleifera* root extract," *International Journal of Pharmacognosy*, vol. 34, no. 3, pp. 207–212, 1996.
- [470] J. O. Midiwo, A. Yenesew, B. Juma, K. L. Omosa, I. L. Omosa, and D. Mutisya, "Phytochemical evaluation of some Kenyan medicinal plants," in *Proceedings of the Phytochemical evaluation of some Kenyan medicinal plants. 11th NAPRECA Symposium Book of Proceedings*, Antananarivo, Madagascar, 2001.
- [471] J. O. Midiwo, N. Gikonyo, D. Wanjau, J. Matasi, and P. Waterman, "Flavonoids of *Polygonum senegalense* (Meisn) Part II: More surface and internal tissue flavonoid aglycones," *Bulletin of the Chemical Society of Ethiopia*, vol. 6, no. 2, 1992.
- [472] M. I. Akpanabiati, I. B. Umoh, E. O. Udosen, A. E. Udoeh, and E. E. Edet, "Rat serum electrolytes, lipid profile and cardiovascular activity on *Nauclea latifolia* leaf extract administration," *Indian Journal of Clinical Biochemistry*, vol. 20, no. 2, pp. 29–34, 2005.
- [473] A. Gidado, D. A. Ameh, and S. E. Atawodi, "Effect of *Nauclea latifolia* leaves aqueous extracts on blood glucose levels of normal and alloxan-induced diabetic rats," *African Journal of Biotechnology*, vol. 4, no. 1, pp. 91–93, 2005.
- [474] P. A. Onyeyili, C. O. Nwosu, J. D. Amin, and J. I. Jibike, "Anthelmintic activity of crude aqueous extract of *Nauclea latifolia* stem bark against ovine nematodes," *Fitoterapia*, vol. 72, no. 1, pp. 12–21, 2001.
- [475] R. R. T. Majinda, M. Motswaledi, R. D. Waigh, and P. G. Waterman, "Phenolic and antibacterial constituents of *Vahlia capensis*," *Planta Medica*, vol. 63, no. 3, pp. 268–270, 1997.
- [476] L. C. Katsoulis, D. J. H. Veale, and I. Havlik, "The pharmacological action of *Rhoicissus tridentata* on isolated rat uterus and ileum," *Phytotherapy Research*, vol. 14, no. 6, pp. 460–462, 2000.
- [477] R. De Paola, C. Muià, E. Mazzon et al., "Effects of *Hypericum perforatum* extract in a rat model of ischemia and reperfusion injury," *Shock*, vol. 24, no. 3, pp. 255–263, 2005.
- [478] E. O. Farombi, "Mechanisms for the hepatoprotective action of kolaviron: studies on hepatic enzymes, microsomal lipids and lipid peroxidation in carbontetrachloride-treated rats," *Pharmacological Research*, vol. 42, no. 1, pp. 75–80, 2000.
- [479] E. O. Farombi, B. F. Adepoju, O. E. Ola-Davies, and G. O. Eme-role, "Chemoprevention of aflatoxin B1-induced genotoxicity and hepatic oxidative damage in rats by kolaviron, a natural biflavonoid of *Garcinia kola* seeds," *European Journal of Cancer Prevention*, vol. 14, no. 3, pp. 207–214, 2005.
- [480] M. A. Gyamfi and Y. Aniya, "Medicinal herb, *Thonningia sanguinea* protects against aflatoxin B1 acute hepatotoxicity in Fischer 344 rats," *Human & Experimental Toxicology*, vol. 17, no. 8, pp. 418–423, 1998.
- [481] E. O. Farombi, M. C. Alabi, and T. O. Akuru, "Kolaviron modulates cellular redox status and impairment of membrane protein activities induced by potassium bromate (KBrO<sub>3</sub>) in rats," *Pharmacological Research*, vol. 45, no. 1, pp. 63–68, 2002.
- [482] J. A. O. Ojewole, "Antinociceptive, anti-inflammatory and antidiabetic properties of *Hypoxis hemerocallidea* Fisch. & C.A. Mey. (Hypoxidaceae) corm ['African Potato'] aqueous extract in mice and rats," *Journal of Ethnopharmacology*, vol. 103, no. 1, pp. 126–134, 2006.
- [483] J. A. O. Ojewole, "Evaluation of the anti-inflammatory properties of *Sclerocarya birrea* (A. Rich.) Hochst. (family: Anacardiaceae) stem-bark extracts in rats," *Journal of Ethnopharmacology*, vol. 85, no. 2–3, pp. 217–220, 2003.
- [484] J. A. O. Ojewole, "Antinociceptive, antiinflammatory and antidiabetic effects of *Leonotis leonurus* (L.) R. BR. (Lamiaceae) leaf aqueous extract in mice and rats," *Methods and Findings in Experimental and Clinical Pharmacology*, vol. 27, no. 4, pp. 257–264, 2005.
- [485] O. A. Olajide, S. O. Awe, J. M. Makinde et al., "Studies on the anti-inflammatory, antipyretic and analgesic properties of *Alstonia boonei* stem bark," *Journal of Ethnopharmacology*, vol. 71, no. 1–2, pp. 179–186, 2000.
- [486] N. Kabiri, S. Asgary, H. Madani, and P. Mahzouni, "Effects of *Amaranthus caudatus* l. extract and lovastatin on atherosclerosis in hypercholesterolemic rabbits," *Journal of Medicinal Plants Research*, vol. 4, no. 5, pp. 355–361, 2010.
- [487] L. Selloum, L. Arrar, B. Medani, A. Khenchouche, and H. Bisker, "Effect of *Cleome arabica* leaves extract on inflammatory cells response in rat," *Biochemical Society Transactions*, vol. 23, no. 4, p. 609, 1995.
- [488] B. M. Mohammed, B. J. Fisher, D. Kraskauskas et al., "Vitamin C promotes wound healing through novel pleiotropic mechanisms," *International Wound Journal*, vol. 13, no. 4, pp. 572–584, 2016.
- [489] J. M. Larrosa, V. Polo, T. Ramirez, I. Pinilla, L. E. Pablo, and F. M. Honrubia, "Alpha-tocopherol derivatives and wound healing in

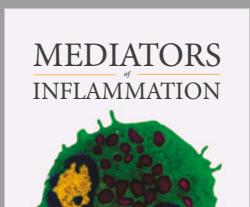
- an experimental model of filtering surgery," *Ophthalmic Surgery, Lasers & Imaging Retina*, vol. 31, no. 2, pp. 131–135, 2000.
- [490] I. Süntar, E. K. Akkol, H. Keles, E. Yesilada, and S. D. Sarker, "Exploration of the wound healing potential of *Helichrysum graveolens* (Bieb.) Sweet: isolation of apigenin as an active component," *Journal of Ethnopharmacology*, vol. 149, no. 1, pp. 103–110, 2013.
- [491] S. Lodhi and A. K. Singhai, "Wound healing effect of flavonoid rich fraction and luteolin isolated from *Martynia annua* Linn. on streptozotocin induced diabetic rats," *Asian Pacific Journal of Tropical Medicine*, vol. 6, no. 4, pp. 253–259, 2013.
- [492] E. Park, S. M. Lee, I.-K. Jung, Y. Lim, and J.-H. Kim, "Effects of genistein on early-stage cutaneous wound healing," *Biochemical and Biophysical Research Communications*, vol. 410, no. 3, pp. 514–519, 2011.
- [493] V. R. Yadav, S. Prasad, B. Sung, and B. B. Aggarwal, "The role of chalcones in suppression of NF- $\kappa$ B-mediated inflammation and cancer," *International Immunopharmacology*, vol. 11, no. 3, pp. 295–309, 2011.
- [494] E. I. O. Ajayi, G. Popoola, and E. Ojediran, "Wound healing potential of *Nauclea latifolia* and *Manihot esculenta* leaf extracts in type 1 diabetic rats," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 13, no. 1, pp. 1–5, 2016.
- [495] C. Agyare, Y. D. Boakye, E. O. Bekoe, A. Hensel, S. O. Dapaah, and T. Appiah, "Review: African medicinal plants with wound healing properties," *Journal of Ethnopharmacology*, vol. 177, pp. 85–100, 2016.
- [496] R. H. Tuhin, M. Begum, S. Rahman et al., "Wound healing effect of *Euphorbia hirta* linn. (Euphorbiaceae) in alloxan induced diabetic rats," *BMC Complementary and Alternative Medicine*, vol. 17, no. 1, article no. 423, 2017.



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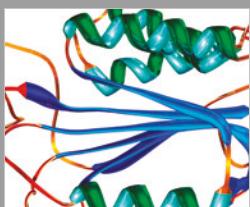
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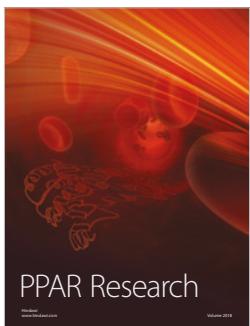
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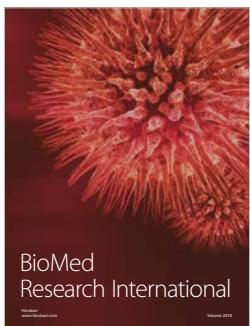


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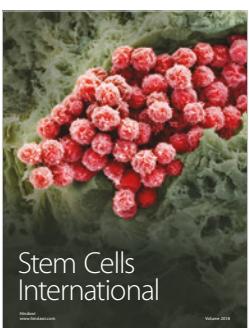
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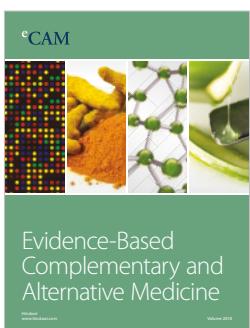
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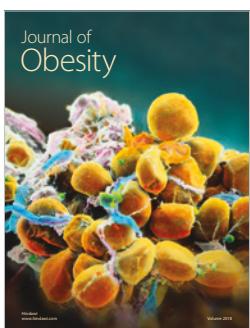
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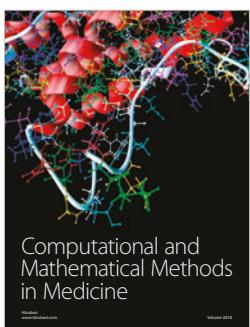
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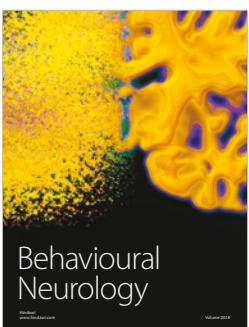
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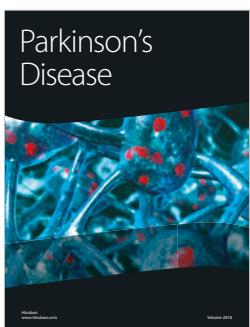
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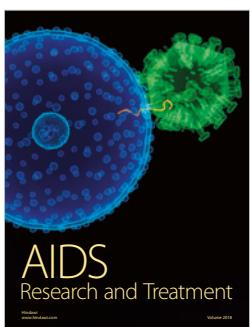
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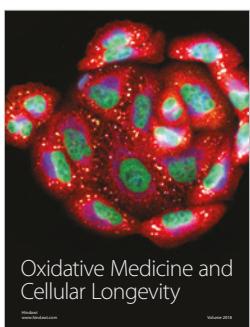
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