



Chemical Equilibrium at the Tick–Host Feeding Interface: A Critical Examination of Biological Relevance in Hematophagous Behavior

Ben J. Mans^{1,2,3*}

¹ Epidemiology, Parasites and Vectors, Agricultural Research Council-Onderstepoort Veterinary Research, Pretoria, South Africa, ² Department of Veterinary Tropical Diseases, University of Pretoria, Pretoria, South Africa, ³ Department of Life and Consumer Sciences, University of South Africa, Pretoria, South Africa

OPEN ACCESS

Edited by:

Abid Ali,
Abdul Wali Khan University Mardan,
Pakistan

Reviewed by:

Jose Ribeiro,
National Institute of Allergy
and Infectious Diseases (NIAID),
United States
Tae Kim,
Texas A&M University, United States
Jan Perner,
Institute of Parasitology (ASCR),
Czechia

*Correspondence:

Ben J. Mans
mansb@arc.agric.za

Specialty section:

This article was submitted to
Invertebrate Physiology,
a section of the journal
Frontiers in Physiology

Received: 12 March 2019

Accepted: 15 April 2019

Published: 01 May 2019

Citation:

Mans BJ (2019) Chemical
Equilibrium at the Tick–Host Feeding
Interface: A Critical Examination
of Biological Relevance in
Hematophagous Behavior.
Front. Physiol. 10:530.
doi: 10.3389/fphys.2019.00530

Ticks secrete hundreds to thousands of proteins into the feeding site, that presumably all play important functions in the modulation of host defense mechanisms. The current review considers the assumption that tick proteins have functional relevance during feeding. The feeding site may be described as a closed system and could be treated as an ideal equilibrium system, thereby allowing modeling of tick–host interactions in an equilibrium state. In this equilibrium state, the concentration of host and tick proteins and their affinities will determine functional relevance at the tick–host interface. Using this approach, many characterized tick proteins may have functional relevant concentrations and affinities at the feeding site. Conversely, the feeding site is not an ideal closed system, but is dynamic and changing, leading to possible overestimation of tick protein concentration at the feeding site and consequently an overestimation of functional relevance. Ticks have evolved different possible strategies to deal with this dynamic environment and overcome the barrier that equilibrium kinetics poses to tick feeding. Even so, cognisance of the limitations that equilibrium binding place on deductions of functional relevance should serve as an important incentive to determine both the concentration and affinity of tick proteins proposed to be functional at the feeding site.

Keywords: tick, host, feeding, affinity, equilibrium, function, relevance

BIOLOGICAL ACTIVITY AT THE TICK FEEDING INTERFACE

Biological function is central to the understanding of life and organismal biology. The context that function confers is exemplified in the frustration felt when a gene or protein of interest can only be annotated as a hypothetical protein with unknown function. Conversely, the ease of annotation by homology has made genome sequencing and assignment of function to orthologs an almost mundane task (Giles and Emes, 2017). Even so, biochemical characterization remains central to the elucidation and confirmation of new function and the understanding of functional mechanism. From a biochemical reductionist perspective, biological function may be explained within a structural mechanistic context as the action of a gene, surface, motif or residue that result

in a chemical reaction, activation or inhibition of an enzyme or receptor, or even a physiological process such as blood clotting or platelet aggregation. The current review considers biological function at the tick–host interface from this reductionist perspective (Elgin, 2010). Even so, it was recently suggested that before or during evolution of new function by gene duplicates, and before negative selection maintain the adaptive advantage conferred by such new functions, new duplicates may exist in a state of undefined or non-optimized function, where protein expression is maintained while functional space is explored by random mutation: the playground hypothesis of neutral evolution (Mans et al., 2017). There is also an increasing recognition that most proteins may exhibit “promiscuous activity,” i.e., irrelevant secondary activities (Copley, 2015, 2017). In addition, given the possibility that a biochemical assay may yield results for a broad class of proteins with diverse functions, or even results without biological meaning, it raises the question whether any particular measured function is relevant within a biological or physiological context. The current review therefore also considers functional relevance at the tick–host interface from this perspective.

Sabbatani (1899), after reading Haycraft’s work on the anti-clotting effects of the oral secretions from leeches (Haycraft, 1884), made the first prescient deduction that all hematophagous organisms must secrete components that interfere or regulate host defenses, such as the hemostatic system. He tested this by showing that crude whole body tick extracts led to *in vitro* inhibition of blood clotting and that injecting this extract into various animals led to prolongation of blood coagulation *in vivo*. He also injected what may very well be the first fractionation of tick proteins to show that this purified preparation retained its inhibitory properties. Sabbatani’s deduction has been confirmed by extensive research into tick–host interactions that established a veritable pharmacopeia of bioactive components secreted by ticks during feeding (reviewed in Mans and Neitz, 2004a; Francischetti et al., 2009; Mans et al., 2016). However, a question that must have plagued him and remains relevant today, is whether *in vitro* and even *in vivo* observations can be causally linked with biological relevant activity at the feeding site, i.e., do what we measure in a test tube really function as a modulator of host defenses during feeding? The observation that ticks can cause paralysis in a host (Hovell, 1824 cited in Scott, 1921) and the presence of salivary glands in ticks (von Siebold and Stannius, 1854; Leydig, 1855; Heller, 1858), must have suggested that ticks can secrete substances into the host. Phenotypic changes in the host such as itching or ecchymosis after tick bite also suggested that ticks secrete substances into the host (Nuttall et al., 1908). Secretion and therefore presence would imply activity at the feeding site. However, the presence of toxic and anti-hemostatic molecules in tick eggs, but not salivary glands or saliva, showed that measurement of biological activity in crude extracts does not necessarily imply function at the tick–host interface (Hoepli and Feng, 1933; de Meillon, 1942; Riek, 1957, 1959; Gregson, 1973; Neitz et al., 1981; Viljoen et al., 1985; Mans et al., 2004a). This implication was recognized soon after Sabbatani’s seminal study, when researchers extended his observations by proving that anti-hemostatic and toxic activities were present in salivary glands of

ticks (Nuttall and Strickland, 1908; Cornwall and Patton, 1914; Ross, 1926; Hoepli and Feng, 1933). It would take a number of years before anti-hemostatic and toxic activity could be showed to be secreted in tick saliva itself. This had to await chemical means, such as pilocarpine, or mechanical means, such as infrared light, to stimulate salivation in order to obtain adequate quantities of salivary secretion for demonstration of biological activity (Howell, 1966; Tatchell, 1967; Neitz et al., 1969, 1978; Dickinson et al., 1976; Ribeiro et al., 1985, 1987; Ribeiro and Spielman, 1986; Ribeiro et al., 1991). However, as Tatchell (1967) indicated: salivary secretions obtained with exogenous stimulants should be treated with caution, since it is unclear whether such secretions represent the total saliva complement or even represent saliva, since cement is not found in such secretions. This may be a pertinent observation since cement may readily form during feeding on artificial membranes (Kröber and Geurin, 2007), arguing that induced salivation is not entirely the same as salivation during actual feeding.

Confirmation of secretion during feeding remains a crucial component of validation of biological relevance (Law et al., 1992). This may be achieved to various extents, by direct determination of the presence of a specific activity or molecule in saliva, or detection of host-derived antibodies generated against components secreted during feeding (Ribeiro et al., 1991; Oleaga-Pérez et al., 1994; Mulenga et al., 2003). Detection in the salivary glands or salivary gland extract (SGE) may be used as an indication of secretion, especially if a secretory peptide signal is present in the immature protein sequence (Nielsen, 2017). The latter have been extensively used to identify potential secretory components during transcriptome analysis (reviewed in Mans et al., 2016). However, secretion of some proteins without canonical signal peptides and non-salivary gland derived proteins via apocrine or alternative secretion has complicated the distinction of true and false positive secretory components (Mulenga et al., 2003; Díaz-Martín et al., 2013b; Oliveira et al., 2013; Tirloni et al., 2014, 2015), thereby also obscuring deduction of biological relevance (Mans et al., 2016). Not all salivary gland proteins with signal peptides are necessarily secreted during feeding (Nielsen, 2017), nor are all secretory proteins secreted at the same time, such as the case for hard ticks, that show differential expression over the course of several days of feeding (McSwain et al., 1982; Paesen et al., 1999; Wang et al., 2001b; de Castro et al., 2016, 2017; Kim et al., 2017; Perner et al., 2018). Transcriptome and proteome data also shows a weak correlation (Schwarz et al., 2014). While this may be ascribed to technical limitations in the proteomic and transcriptomic analysis of complex samples from non-model organisms, it further complicates the assessment of the final relevant concentration of protein present during feeding.

The Equilibrium State and Functional Correlates

Le Chatelier’s principle states that an equilibrium system will tend to counteract changes to the system to maintain its equilibrium. In biological systems this is overcome by enzymes that create intermediate states to catalyze non-reversible chemical reactions.

In the case of receptor-ligand or protein-inhibitor binding, the affinity of the receptor for the ligand and the relative receptor and ligand concentrations determines the bound state at equilibrium (**Figure 1**). The affinity between molecules is expressed as the equilibrium dissociation constant (K_D) that range for most biological systems from the fM to mM range (Smith et al., 2012). The K_D is the ligand concentration at which 50% of the receptor is occupied at equilibrium (**Figure 1**). At a ligand concentration 10-fold the K_D , a receptor would be 91% saturated and above this concentration almost all of the receptor will be occupied. However, at a ligand concentration 10-fold less than the K_D , only 9% of the receptor would be occupied by the ligand and below this concentration essentially no binding occurs. At concentrations of the ligand that is equal to its K_D , only 50% of the receptor will be occupied if the stoichiometry of binding is 1:1. The K_D is therefore also considered to reflect a physiological relevant concentration (Sears et al., 2007). Concentrations of salivary proteins at the feeding site would therefore be physiologically relevant only at concentrations equal or exceeding their affinities for their targets by 10-fold or more if 50–100% receptor occupancy is necessary for effective inhibition. The K_i (inhibitor constant) is determined for inhibitors of enzyme active sites and reflect the concentration necessary to reduce enzyme activity by half. It is generally used in the characterization of inhibitors using enzyme kinetics and depends on both enzyme affinity for substrate and inhibitor affinity for enzyme. Even so, it is similar to the K_D and should produce similar values when enzyme activity is measured at physiological concentrations. Another measurement analogous to the K_D is the IC_{50} value of an inhibitor. The IC_{50} value (half-maximal inhibitory concentration) is the concentration at which 50% inhibition of function is observed. This is generally determined for complex processes such as inhibition of platelet

aggregation, blood clotting, or cell migration. This value would ultimately depend on the concentration of receptor or number of cells used in an assay, but would be close to the K_D if physiologically relevant receptor or cell concentrations were used. Another measure that may indicate physiological relevance is the stoichiometric inhibition ratio (SI). This ratio indicate physiological relevance when an inhibitor interact with an enzyme or receptor close to equimolar ratios, since this imply high affinity, so that for example an $SI = 2$ imply saturation of receptor at $2K_D$. This may be seen for suicide inhibitors that bind irreversible to the enzyme active site. In enzyme kinetics the K_m (Michaelis constant) is the substrate concentration that allows an enzyme to attain half V_{max} . V_{max} is the maximum reaction rate of the enzyme when saturated with substrate. While the K_m is dependent on rate constants rather than ligand concentration it is also an indication of affinity and provide an estimate of relevant concentrations of substrates where an enzyme will function. For example, if a given agonist functions at concentrations 10-fold lower than the K_m , the enzyme will not be able to effectively remove this agonist from the system and neutralize its effect. The K_D , K_i , IC_{50} , SI, and K_m are therefore all useful to assess the physiological relevance of any biological activity. The reader is referred to Kuriyan et al. (2013) for a general treatment of protein-ligand affinity.

Another measure of potential functional activity is the amount of salivary gland equivalents that may have a measurable impact or functionality. For example, ~ 0.125 salivary gland equivalents from the tick *Ornithodoros kalahariensis* (previously *Ornithodoros savignyi*, Bakkes et al., 2018), was able to increase clotting time for the APTT test by 400%, 0.4% of a salivary gland could inhibit fXa ~ 100 and 0.07% could inhibit thrombin by $\sim 100\%$ (Gaspar et al., 1995). As such, any function that may be measured from the equivalent of 2 salivary glands (~ 1 tick) or less that has an appreciable effect on some function may indicate the presence of relevant functionality. Even so, if characterizing crude extracts, it should be considered that the sum of multiple functions may be measured and that individual activities may be much less.

Whether a molecule will have relevant biological activity at the feeding site depends on their affinities for their respective ligands or receptors and whether they are secreted at relevant biological concentrations. In terms of chemical equilibrium, this implies that inhibitors or kratagonists (dealt with below) needs to be present at the feeding site at higher concentrations than the K_D for their respective receptors or ligands. At concentrations below the K_D little or no binding will occur and inhibitors may not be effective or physiologically relevant. In addition to satisfying concentration requirements, competition between host-derived substrates, ligands or receptors and tick-derived inhibitors, enzymes or scavengers for activating biomolecules will determine whether potential host-modulatory molecules may be biologically significant. This is essentially determined by the comparative affinities of host vs. tick-derived receptor-ligand interaction. Again, the biological relevance of host-derived agonists will be determined by their active concentrations at the feeding site. Effective concentration may also be determined by protein turnover or half-life at the feeding site. This will

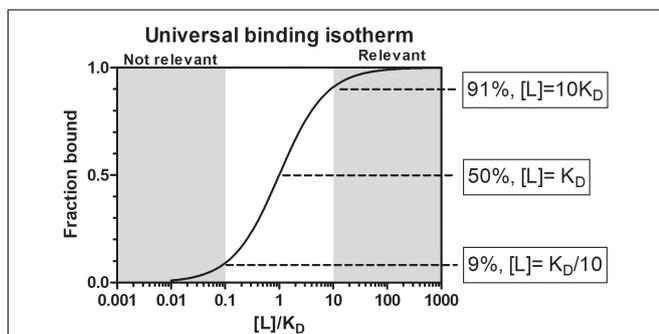


FIGURE 1 | The universal binding isotherm express the relationship between receptor occupation by a ligand (fraction bound) and the equilibrium dissociation constant (K_D) with regard to the concentration of ligand present for a 1:1 interaction at equilibrium. At a ligand concentration that equals the K_D ($1K_D$), 50% of the receptor will be occupied. At a ligand concentration 10-fold more than the K_D ($10K_D$), 91% of the receptor will be occupied. Above this concentration a given ligand should be physiologically significant. At a ligand concentration 10-fold less than the K_D ($K_D/10$) only 9% of the receptor will be occupied. Below this concentration physiological significance should be suspect. Between $10K_D$ and $K_D/10$ a gray area exist where physiological significance will depend on environmental factors.

depend on secretion into the feeding site, concentration and sequestration at the feeding site and protein stability. In the case of secretory proteins, most have disulphide bonds that increase their stability (Mans et al., 2016), contributing toward their effective concentrations. Modulation of host defenses at the tick–host interface therefore depends on interplay between these factors, with the implication that tick-derived antagonists need to be present at higher concentrations than host-derived agonists and have higher affinities for their shared targets (**Figure 2**).

The Feeding Site: Effective Volume Determines Functionality

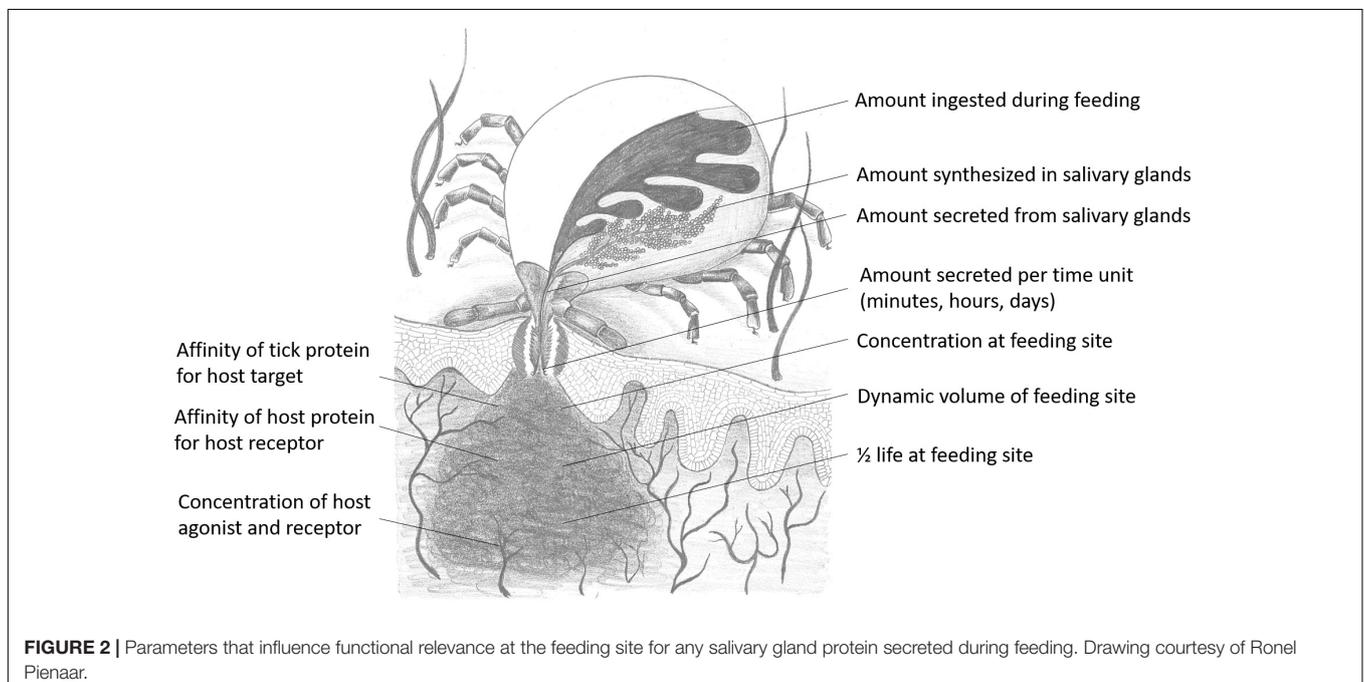
The equilibrium state is an ideal closed state and the feeding site may be considered as such. However, while the feeding site may have a defined volume approximating a closed state, the feeding site itself may be dynamic with constant changes in concentration of both host and tick proteins. Ticks ingest blood meal and salivary components, since ticks alternately salivate and ingest blood meal (Arthur, 1970; Kemp et al., 1982), thereby removing both volume and active components from the feeding site. These repetitive doses of saliva, alternating by sucking events may be considered to be independent equilibrium events and in such a case may suggest that estimates of concentration at the feeding site may be lower than described below. Tick-derived molecules may also be removed into the general systemic system of the host or the general area around the feeding site, as evidenced by the systemic effects seen during paralysis and tick toxicoses (Mans et al., 2004a). Influx of blood or lymph into the feeding site may also dilute tick-derived proteins as seen in the case of edema (Kemp et al., 1982). As such, while the feeding site itself has been described as a hematoma or cavity that seems self-contained,

the site itself may be much more dynamic than a defined cavity of blood.

A critical question remains as to what extent the feeding site may be reduced to a biochemical reaction in a test tube, i.e., can we measure the feeding site, calculate the feeding site volume and determine the amount of saliva secreted by the tick to derive estimations of biological concentrations at the feeding site? However, the feeding site is a complex and dynamic environment that changes constantly as determined by both the tick and the host (Nuttall, 2019). For example, ticks secrete enzymes that degrade the extracellular matrix over time, increasing the feeding site volume, while host wound healing responses will tend to counteract an increase in feeding site volume (Wikel, 2017). On the other hand, host inflammatory responses such as edema and cellular infiltration will tend to increase the feeding site volume, while hard ticks actively secrete excess blood-meal derived water back into the host, which will further contribute to feeding site volume, dilution and drainage of salivary proteins. As such, feeding site volumes have been conservatively estimated at 10–50 μl (Mans et al., 2008a,b). However, feeding site volume differs between larvae, nymphs and adults, whether natural or secondary hosts are parasitized and whether hosts are naive or immune (Tatchell and Moorhouse, 1968; Brown and Knapp, 1980a,b; Brown et al., 1983). Natural hosts tend to present smaller feeding lesions, while secondary hosts can produce large lesions due to inflammatory responses (Brown and Knapp, 1980a,b; Brown et al., 1983).

Protein Concentration at the Feeding Site and Functionally Suspect Proteins

It may be estimated what concentrations can be expected for the average secreted protein at the feeding site, by asking what



the highest protein concentration may be. In this regard, the lipocalins are known to be some of the most abundant proteins synthesized in tick salivary glands (Mans, 2011; Mans et al., 2017). The TSGP1-4 lipocalins from the soft tick *O. kalahariensis* has been characterized in detail. They each make up ~5% of the total soluble salivary gland protein, comprising ~20% of the total soluble protein content (Mans et al., 2001; Mans and Neitz, 2004b). In this case the total protein that may be secreted by a tick during a feeding event comprise ~6 μg protein with molecular masses of ~15 kDa each. Similarly, the savignygrins (Mr ~ 7 kDa) also make up ~3% of the total salivary gland protein with ~4 μg of protein secreted during a feeding event (Mans et al., 2002b). In the case of the lipocalins from the soft ticks *Argas monolakensis*, the histamine binding protein AM-10 (Mr ~ 16 kDa) comprise ~23% of the total soluble protein (Mans et al., 2008b,c), and in this case equates with 4.6 μg of protein secreted during feeding. These concentrations are likely to be the highest estimates, since it is known that hard ticks secrete ~30–300 times less protein than soft ticks in their saliva at any given feeding stage (Ribeiro, 1987; Dharampaul et al., 1993). The upper limit for any given protein in the adult tick salivary gland may thus be assumed to be ~10 μg with a dynamic protein concentration range of 10,000-fold (1 ng–10 μg). If it is also assumed that nymphs have 10-fold and larvae 100-fold lower salivary concentrations than adults some interesting observations may be made regarding feeding site concentrations. For the purpose of the current study, the feeding site volumes were estimated from the studies of Brown and Knapp (1980a,b) with feeding cavities for larvae from 0.1 to 50 nL, nymphs from 5 to 150 nL, and adults from 10 nL to 10 μL . Using these assumptions concentration fluctuation at the feeding site can be estimated at various salivary gland protein concentrations and molecular masses of proteins. Under these assumptions the concentrations at the feeding site do not differ extensively between larvae, nymphs or adults, even though feeding site volume and effective salivary gland concentrations may differ (Figure 3). Allowing for the concentration ranges estimated, maximum concentrations at the feeding site may range from nM to mM, and may even range from nM to μM for salivary gland concentrations such as 0.1, 0.01, or 0.001 μg for adults, nymphs, or larvae, respectively, that may be considered to be more representative of the average protein concentration. These concentrations may be an over estimation since the dynamic nature of the feeding site and the rate at which protein is secreted may never approximate these total estimated concentrations.

In ticks the IC_{50} , K_D , and K_i values range from pM–mM and falls within the concentration expected at the feeding site (Figures 3, 4). IC_{50} values generally tend to range from 10 nM to 10 μM (Figure 2). Conversely, K_D values range from 1 nM to 1 μM , while K_i values range from 10 pM to 10 nM. The higher IC_{50} values is probably due to the use of *in vitro* assays that measure complex reactions such as platelet aggregation, blood clotting, complement or cell migration. These systems do not necessarily represent true *in vivo* conditions and may be artificial to some extent, i.e., much higher concentrations of agonists or cell numbers are needed for observation than what is found

under physiological conditions. The K_D values may represent true affinities, while the K_i values are dependent on the enzyme and substrate concentrations used, and the lower pM values may be due to the availability of chromogenic and fluorogenic substrates that enable sensitive measurement of enzyme activity. In this case, some of the affinities may be overestimated.

A number of tick proteins characterized have affinities, inhibition constants or IC_{50} values in the nM or pM range, which would probably be functional at the feeding site (Figure 4 and Table 1). For some, concentrations at the feeding site may be estimated based on yields of purified inhibitor and these are generally in the same range as the affinities or higher, suggesting biological relevant concentrations at the feeding site (Table 1). For a number of proteins, estimates of concentrations at the feeding site is not available and the low nM to pM range of their K_D , K_i , or IC_{50} values is the only indication of functional relevance. A number of proteins also present extraordinary high concentration estimates at the feeding sites (>1 M), which may indicate that these proteins may not have been completely purified at the time of their characterization, or may reflect that they derived from whole body extracts and not tick salivary glands, which may suggest that these inhibitors originate from multiple organs (Ibrahim et al., 2001a,b). In many cases, the concentration of inhibitors that may occur at the feeding site has not been determined (Table 1). This will remain a major impediment in the assessment of functional relevance at the feeding site, since we are probably over rather than underestimating feeding site concentrations. To further consider functional relevance it is necessary to unpack the salivary gland repertoire into its functional modalities.

TICK FEEDING AND MODULATION OF HOST DEFENSES

Ticks need to modulate their vertebrate host's defense mechanisms to obtain a successful blood-meal (Francischetti et al., 2009). In this regard, ticks secrete numerous proteins during feeding that function as inhibitors, enzymes, or kratagonists that use a variety of mechanisms to overcome host defenses (Mans, 2011; Mans et al., 2016) (Figure 5).

Inhibitors That Target Enzyme Active Sites and/or Substrate-Binding Exosites

Various enzymes are involved in host hemostasis and many are part of enzyme cascades such as the blood clotting or complement cascade that enable efficient control as well as rapid response to injury or infection (Francischetti et al., 2009). Inhibitors for these cascades are found in all ticks. Inhibitors of blood-clotting enzymes generally target the active and exosites and inhibit primary enzymatic activity of thrombin, fXa, kallikrein, kallikrein-fXIIa-fXIa, fXa-TF-VIIa, plasmin, fV or carboxypeptidase B, thereby inhibiting formation or dissolution of the fibrin clot (Table 1 including references). Inhibitors may also target enzymes that may induce platelet aggregation such as cathepsin G, or enzymes involved in inflammation such as cathepsin B, C, H, L, S, and V, chymase,

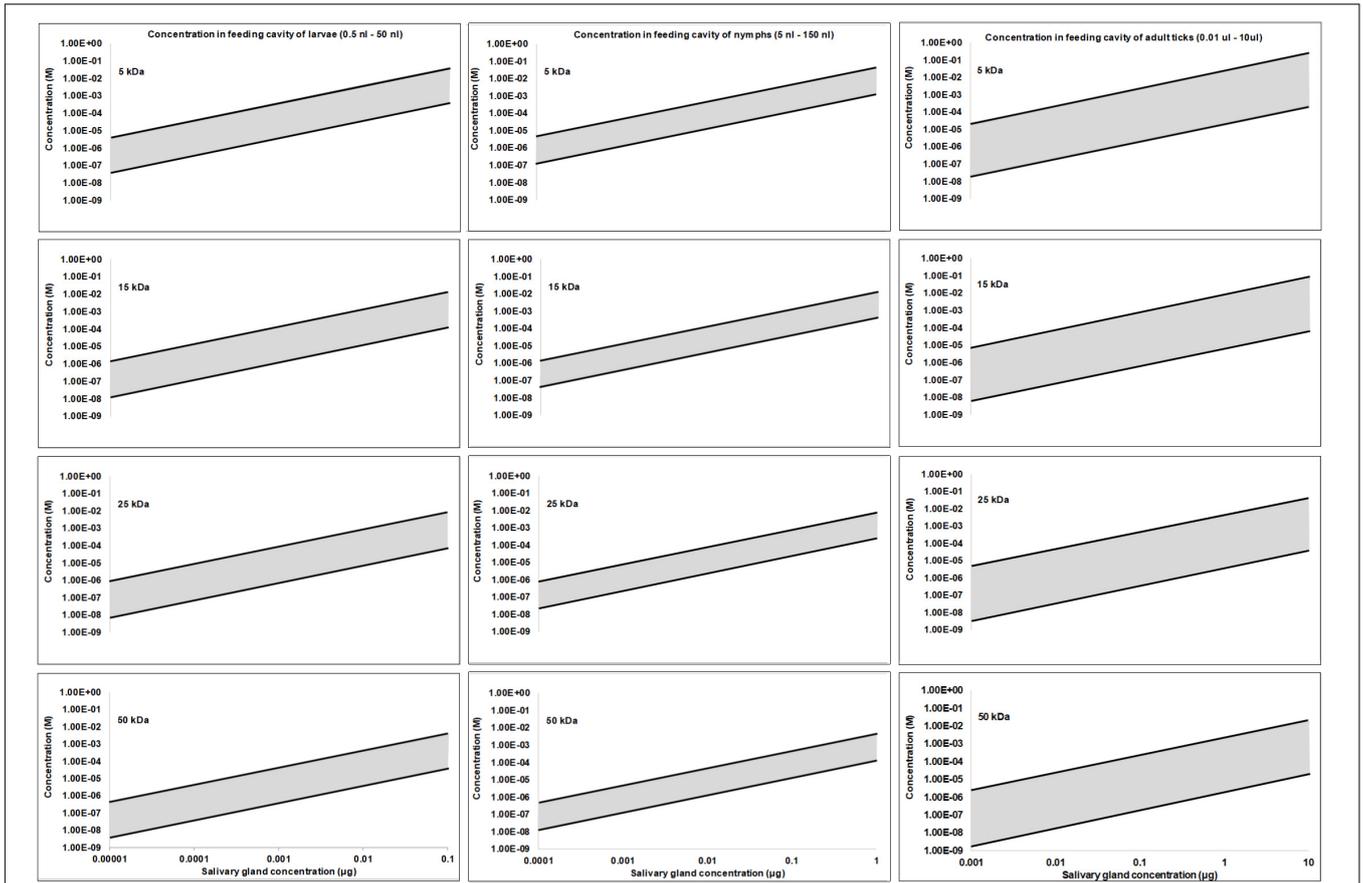


FIGURE 3 | Protein concentration estimates at the feeding site for larvae, nymphs and adults. Feeding sites volumes were estimated from Brown and Knapp (1980a; 1980b) and salivary concentrations were simulated over a 10,000-fold range assuming a maximum concentration of 10 µg in adults, 1 µg in nymphs and 0.1 µg in larvae.

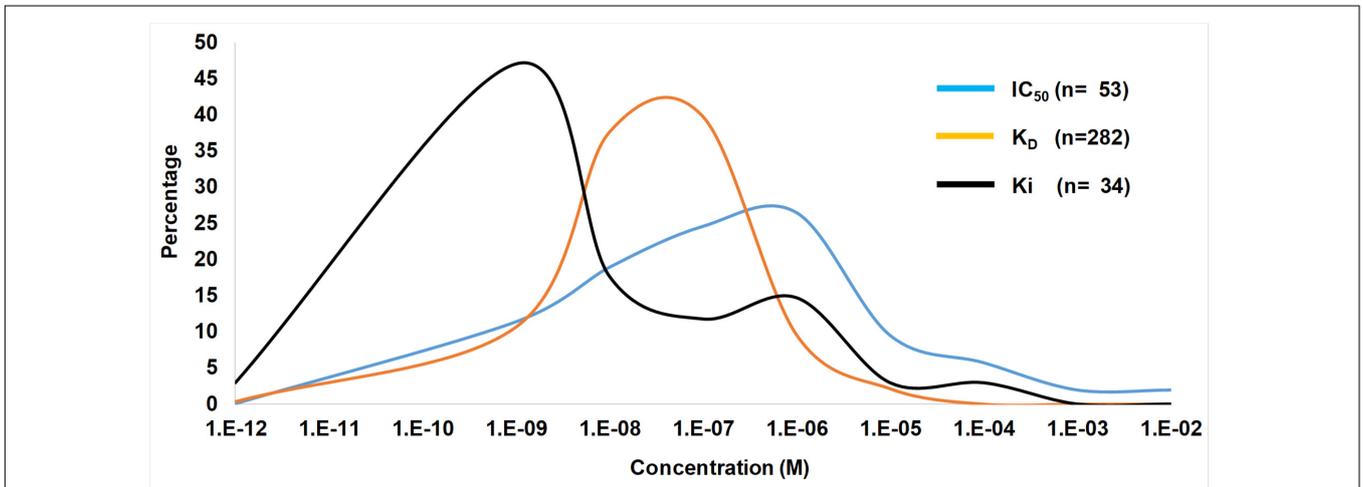


FIGURE 4 | Ranges of IC_{50} , K_D , and K_i values from the tick literature. Indicated are frequency distribution curves of values obtained from Table 1 and references therein.

TABLE 1 | Tick inhibitors of host defenses considered to have relevant physiological function.

| Protein | Tick | Target | Affinity (K_D , K_i , K_m , IC_{50}) | Concentration (Minimum–Maximum) | References |
|-------------|-------------------------------------|-------------------------------------|---|--------------------------------------|--|
| Apyrase | <i>Ixodes scapularis</i> | ATP/ADP | N/A | Activity in saliva | Ribeiro et al., 1985 |
| Apyrase | <i>Ornithodoros moubata</i> | ATP/ADP | N/A | Activity in saliva | Ribeiro et al., 1991 |
| Apyrase | <i>Ornithodoros kalahariensis</i> | ATP/ADP | $K_m \sim 1$ mM | Activity in saliva 1 μ M–1 mM | Mans, 1997; Mans et al., 1998b |
| Ir-CPI | <i>Ixodes ricinus</i> | fXIIa, fXIa, Plasmin | $K_D \sim 16$ –38 nM | Unknown | Decrem et al., 2009 |
| PAI | <i>Ixodes sinensis</i> | Platelets | $IC_{50} \sim 250$ nM | Unknown | Liu et al., 2005 |
| Savignygrin | <i>Ornithodoros kalahariensis</i> | $\alpha_{IIb}\beta_3$ | $IC_{50} \sim 130$ nM; $K_D \sim 60$ nM | 55 μ M–57 mM | Mans et al., 2002b; Mans and Neitz, 2004c |
| Monogrin | <i>Argas monolakensis</i> | $\alpha_{IIb}\beta_3$ | $IC_{50} \sim 150$ nM | 10 μ M–10 mM | Mans et al., 2008a |
| Variabilin | <i>Dermacentor variabilis</i> | $\alpha_{IIb}\beta_3$ | $IC_{50} \sim 150$ nM | 2.7 μ M–2.7 mM | Wang et al., 1996 |
| Disagregin | <i>Ornithodoros moubata</i> | $\alpha_{IIb}\beta_3$ | $IC_{50} \sim 104$ nM; $K_D \sim 40$ nM | 20 μ M–20 μ M | Karczewski et al., 1994 |
| YY-39 | <i>Ixodes scapularis</i> | $\alpha_{IIb}\beta_3$ | $IC_{50} \sim 4$ –20 μ M | Unknown | Tang et al., 2015 |
| TAI | <i>Ornithodoros moubata</i> | $\alpha_2\beta_1$ | $IC_{50} \sim 5$ –8 nM; $K_D \sim 40$ nM | 50 nM–50 μ M | Karczewski et al., 1995 |
| Moubatin | <i>Ornithodoros moubata</i> | TXA ₂ | $IC_{50} \sim 50$ nM $K_D \sim 24$ nM | 1.5 μ M–1.5 mM | Waxman and Connolly, 1993; Mans and Ribeiro, 2008b |
| TSGP2 | <i>Ornithodoros kalahariensis</i> | LTB ₄ | $K_D \sim 18$ nM | 38 μ M–38 mM | Mans et al., 2001; Mans and Ribeiro, 2008b |
| TSGP3 | <i>Ornithodoros kalahariensis</i> | TXA ₂ ; LTB ₄ | $K_D \sim 5$ –21 nM | 38 μ M–38 mM | Mans et al., 2001; Mans and Ribeiro, 2008b |
| Ir-LBP | <i>Ixodes ricinus</i> | LTB ₄ | $K_D \sim 0.5$ nM | Detected in saliva | Beaufays et al., 2008 |
| TSGP4 | <i>Ornithodoros kalahariensis</i> | LTC ₄ | $K_D < 2$ nM | 35 μ M–35 mM | Mans et al., 2001; Mans and Ribeiro, 2008a |
| AM-33 | <i>Argas monolakensis</i> | LTC ₄ | $K_D \sim 2$ nM | 3.5 μ M–3.5 mM | Mans et al., 2008c; Mans and Ribeiro, 2008a |
| TSGP1 | <i>Ornithodoros kalahariensis</i> | Histamine Serotonin | $K_D < 3$ nM; $K_D \sim 6$ nM | 32 μ M–32 mM | Mans et al., 2001, 2008b |
| OP-3 | <i>Ornithodoros parkeri</i> | Histamine Serotonin | $K_D \sim 106$ nM; $K_D \sim 116$ nM | Detected in SGE | Francischetti et al., 2008 Mans and Ribeiro, 2008b |
| OtLip | <i>Ornithodoros turicata</i> | Histamine | Unknown | Unknown | Neelakanta et al., 2018 |
| Monomine | <i>Argas monolakensis</i> | Histamine | $K_D \sim 7$ nM | 29 μ M–28 mM | Mans et al., 2008b,c |
| Monotonin | <i>Argas monolakensis</i> | Serotonin | $K_D < 2$ nM | 10 μ M–10 mM | Mans et al., 2008b,c |
| Is-14 | <i>Ixodes scapularis</i> | Histamine Serotonin | $K_D \sim 427$ nM; $K_D < 2$ | Unknown | Mans et al., 2008c |
| Is-15 | <i>Ixodes scapularis</i> | Histamine Serotonin | $K_D \sim 746$ nM; $K_D < 2$ | Unknown | Mans et al., 2008c |
| HBP1-3 | <i>Rhipicephalus appendiculatus</i> | Histamine | $K_D \sim 1$ –18 nM | Detected in SGE | Paesen et al., 1999 |
| SHBP | <i>Dermacentor reticulatus</i> | Histamine Serotonin | $K_D \sim 1$ –2 nM $K_D \sim 0.6$ nM | Unknown | Sangamnatdej et al., 2002 |
| Savignin | <i>Ornithodoros kalahariensis</i> | Thrombin | $K_i \sim 5$ pM | 850 nM–1 mM | Nienaber et al., 1999 |
| P5 | <i>Hyalomma dromedarii</i> | Thrombin | $K_i \sim 500$ nM | 30 μ M–30 mM | Ibrahim and Masoud, 2018 |
| Sculptin | <i>Amblyomma sculptum</i> | Thrombin | $IC_{50} \sim 2$ pM $K_i \sim 18$ pM; | Unknown | Iqbal et al., 2017 |
| Avathrin | <i>Amblyomma variegatum</i> | Thrombin | $IC_{50} \sim 7$ nM $K_i \sim 545$ pM | Unknown | Iyer et al., 2017 |
| Boophilin | <i>Rhipicephalus microplus</i> | Thrombin | $IC_{50} \sim 1$ μ M $K_i \sim 101$, 0.5–2 nM $K_D \sim 116$ nM | Unknown | Macedo-Ribeiro et al., 2008; Soares et al., 2012; Assumpção et al., 2016 |
| RmS-15 | <i>Rhipicephalus microplus</i> | Thrombin | SI ~ 2 | Unknown | Rodriguez-Valle et al., 2015; Xu T. et al., 2016 |
| Hyalomin-1 | <i>Hyalomma rufipes</i> | Thrombin | $K_i \sim 12$ nM $K_D \sim 19$ nM | Unknown | Jablonka et al., 2015 |

(Continued)

TABLE 1 | Continued

| Protein | Tick | Target | Affinity (K_D , K_i , K_m , IC_{50}) | Concentration (Minimum–Maximum) | References |
|---------------|---------------------------------------|---------------------------|--|---------------------------------|--|
| rixscS-1E1 | <i>Ixodes scapularis</i> | Thrombin | SI ~ 4 | Unknown | Ibelli et al., 2014 |
| Monobin | <i>Argas monolakensis</i> | Thrombin | K_i ~ 6 pM | 400 nM–400 μ M | Mans et al., 2008a |
| Chimadanin | <i>Haemaphysalis longicornis</i> | Thrombin | IC_{50} ~ 300 nM | Unknown | Nakajima et al., 2006 |
| Madanins | <i>Haemaphysalis longicornis</i> | Thrombin | K_D ~ 3–4 μ M; K_i ~ 31–55 nM | Unknown | Iwanaga et al., 2003; Figueiredo et al., 2013 |
| Varieggin | <i>Amblyomma variegatum</i> | Thrombin | IC_{50} ~ 1 nM; K_i ~ 10 pM | Unknown | Kazimirová et al., 2002; Koh et al., 2007 |
| NTI-2 | <i>Hyalomma dromedarii</i> | Thrombin | K_i ~ 211 nM | 3.8 mM–3.8 M | Ibrahim et al., 2001b |
| Americanin | <i>Amblyomma americanum</i> | Thrombin | K_i ~ 0.073 nM | 100 nM–100 μ M | Zhu et al., 1997a |
| Saliva | <i>Amblyomma americanum</i> | fXa; Thrombin | 100% I ~ 0.3 SU | Unknown | Zhu et al., 1997b |
| Ornithodorin | <i>Ornithodoros moubata</i> | Thrombin | K_i ~ 1 pM | Unknown | van de Locht et al., 1996 |
| Anticoagulant | <i>Ixodes holocyclus</i> | Clotting | IC_{50} ~ 0.25 SGU | Unknown | Anastopoulos et al., 1991 |
| Ixin | <i>Ixodes ricinus</i> | Thrombin | IC_{50} ~ 0.6 SGU | Unknown | Hoffmann et al., 1991 |
| Calcaratin | <i>Rhipicephalus calcaratus</i> | Thrombin | Unknown | Unknown | Motoyashiki et al., 2003 |
| Haemathrin | <i>Haemaphysalis bispinosa</i> | Thrombin | IC_{50} ~ 40 μ M | Unknown | Brahma et al., 2017 |
| Microphilin | <i>Rhipicephalus microplus</i> | Thrombin | IC_{50} ~ 6–42 μ M | 17 μ M–17 mM | Ciprandi et al., 2006 |
| BmAP | <i>Rhipicephalus microplus</i> | Thrombin | IC_{50} ~ 100 nM–1 μ M | 333 nM–33 μ M | Horn et al., 2000 |
| NTI-1 | <i>Hyalomma dromedarii</i> | Thrombin | K_i ~ 11.7 μ M | 650 μ M–6.5 M | Ibrahim et al., 2001b |
| HLS2 | <i>Haemaphysalis longicornis</i> | Thrombin | IC_{50} ~ μ M | Unknown | Imamura et al., 2005 |
| IRS-2 | <i>Ixodes ricinus</i> | Cathepsin G; Chymase | IC_{50} ~ 4–11 nM | Unknown | Chmelař et al., 2011 |
| AamS6 | <i>Amblyomma americanum</i> | Undefined | All tests μ M range; no parameters | Immunogenic | Chalaire et al., 2011; Mulenga et al., 2013 |
| Penthalaris | <i>Ixodes scapularis</i> | Tissue factor pathway | IC_{50} ~ 100 pM | Unknown | Francischetti et al., 2004 |
| Ixolaris | <i>Ixodes scapularis</i> | Tissue factor pathway fXa | IC_{50} ~ pM range | Unknown | Francischetti et al., 2002; Monteiro et al., 2005 |
| Hd_fXal | <i>Hyalomma dromedarii</i> | fXa | K_i ~ 134 nM | 0.3 mM–300 mM | Ibrahim et al., 2001a |
| Anticoagulant | <i>Rhipicephalus appendiculatus</i> | fXa | 450% clotting time increase for 2 SGU | 100 nM–125 μ M | Limo et al., 1991 |
| TAP | <i>Ornithodoros moubata</i> | fXa | K_i ~ 0.6 nM | 8 μ M–8 mM | Waxman et al., 1990 |
| fXal | <i>Ornithodoros kalahariensis</i> | fXa | K_i ~ 0.8 nM | 250 nM–250 μ M | Gaspar et al., 1996 |
| Salp14 | <i>Ixodes scapularis</i> | fXa | IC_{50} ~ 150 nM | Immunogenic | Narasimhan et al., 2002 |
| Amblyomin-X | <i>Amblyomma sculptum</i> | fXa | IC_{50} ~ 10 μ M; K_i ~ 4 μ M | Unknown | Batista et al., 2010; Branco et al., 2016 |
| TIX-5 | <i>Ixodes scapularis</i> | fV | IC_{50} ~ 3.2 μ M; all tests μ M range | Immunogenic | Schuijt et al., 2011, 2013 |
| Rhipilin-1 | <i>Rhipicephalus haemaphysaloides</i> | Elastase | SI ~ 6 | Unknown | Cao et al., 2013 |
| BmTI-A | <i>Rhipicephalus microplus</i> | Elastase; Kallikrein | K_i ~ 1.4 nM; K_i ~ 120 nM | Unknown | Tanaka et al., 1999 |
| Iris | <i>Ixodes ricinus</i> | Elastase TNF- α | IC_{50} ~ μ M IC_{50} ~ 50 nM | Unknown | Leboulle et al., 2002; Prevot et al., 2006, 2009 |
| RsTIQ 2, 5, 7 | <i>Rhipicephalus sanguineus</i> | Elastase; plasmin | K_i ~ 1–38 nM | Unknown | Sant'Anna Azzolini et al., 2003 |
| Haemaphysalin | <i>Haemaphysalis longicornis</i> | fXIIa | IC_{50} ~ 50–100 nM; K_D ~ 3 nM | Unknown | Kato et al., 2005 |
| AAS19 | <i>Amblyomma americanum</i> | Plasmin; Thrombin | SI ~ 3–9 | Unknown | Kim et al., 2015; Radulović and Mulenga, 2017 |

(Continued)

TABLE 1 | Continued

| Protein | Tick | Target | Affinity (K_D , K_i , K_m , IC_{50}) | Concentration (Minimum–Maximum) | References |
|--|--|--|--|---------------------------------|--|
| TdPI | <i>Rhipicephalus appendiculatus</i> | Tryptase | $K_i < 1.5$ nM | Unknown | Paesen et al., 2007 |
| Tryptogalinin | <i>Ixodes scapularis</i> | Plasmin Tryptase | $K_i \sim 5.83$ nM $K_i \sim 10$ pM | Unknown | Valdés et al., 2013 |
| Sialostatin L | <i>Ixodes scapularis</i> | Cathepsin L | $IC_{50} \sim 5$ nM; $K_i \sim 95$ pM | Detected in saliva | Kotsyfakis et al., 2006 |
| Sialostatin L2 | <i>Ixodes scapularis</i> | Cathepsin L | $IC_{50} \sim 70$ pM; $K_i \sim 65$ pM | Unknown | Kotsyfakis et al., 2007 |
| Iristatin | <i>Ixodes ricinus</i> | Cathepsin L and C | $IC_{50} \sim 500$ nM–3 μ M | Unknown | Kotál et al., 2019 |
| OmC2 | <i>Ornithodoros moubata</i> | Cathepsin L and S | $IC_{50} \sim 150$ pM; $K_i \sim 65$ pM | Detected in saliva | Grunclová et al., 2006; Salát et al., 2010 |
| BrBmcys2b | <i>Rhipicephalus microplus</i> | Cathepsin B and L | $K_i \sim 1$ –3 nM | Detected in saliva | Parizi et al., 2013 |
| RHcyst-2 | <i>Rhipicephalus haemaphysaloides</i> | Cathepsin S | $IC_{50} \sim 100$ pM | Detected in saliva | Wang et al., 2015 |
| TCI | <i>Rhipicephalus bursa</i> | Carboxy-peptidase | $K_i \sim 1$ –4 nM | 150 nM–150 μ M | Arolas et al., 2005 |
| HITCI | <i>Haemaphysalis longicornis</i> | Carboxy-peptidase | $IC_{50} \sim 10$ μ M | Unknown | Gong et al., 2007 |
| Evasin-1 | <i>Rhipicephalus sanguineus</i> | CCL3; CCL8; CCL18 | $K_D \sim 160$ pM; $K_D \sim 810$ pM; $K_D \sim 3$ nM | Detected in saliva | Frauschuh et al., 2007 |
| Evasin-3 | <i>Rhipicephalus sanguineus</i> | CXCL1; CXCL8 | $K_D \sim 340$ pM; $K_D \sim 700$ pM; | Detected in saliva | Déruaz et al., 2008 |
| Evasin-4 | <i>Rhipicephalus sanguineus</i> | CCL1–CCL27 | $K_D \sim 10$ –700 pM; | Detected in saliva | Déruaz et al., 2013 |
| P1243 (AAM-02) | <i>Amblyomma americanum</i> | CCL1–CCL27 | $K_D \sim 1$ –100 nM | Unknown | Alenazi et al., 2018 |
| P1156 | <i>Ixodes ricinus</i> | CXCL1–8 | $K_D \sim 3$ –70 nM | Unknown | Alenazi et al., 2018 |
| RPU-01, RPU-02, AAM-01, AAM-02, ACA-01, ACA-02, AMA-01, ATR-02, IRI-01 | <i>Rhipicephalus pulchellus</i> <i>Amblyomma americanum</i> <i>Amblyomma cajennense</i> <i>Ixodes ricinus</i> <i>Ixodes holocyclus</i> | Eotaxin-1; Eotaxin-2; Eotaxin-3; MCP-1; MCP-2; MCP-3 | $K_D \sim 5$ –1,000 nM | Unknown | Hayward et al., 2018 |
| P991, P985, P546, P974, P983, P1181, P1182, P1183, P1180, P467 | <i>Amblyomma maculatum</i> <i>Amblyomma triste</i> <i>Amblyomma parvum</i> <i>Rhipicephalus pulchellus</i> | CCL2, CCL3, CCL4, CCL7, CCL8, CCL11, CCL13, CCL14, CCL16, CL17, CCL18, CCL19 CCL20, CCL21, CCL22, CL23 CCL24, CCL27, CCL28, CCL25 | $K_D \sim 1$ pM–3 μ M | Unknown | Singh et al., 2017 |
| P672 | <i>R. pulchellus</i> | CCL3, CCL8, CCL11, CCL13, CCL14, CCL16, CCL18, CCL23 | $K_D \sim 1$ –10 nM; $IC_{50} \sim 2$ –6 nM | Unknown | Eaton et al., 2018 |
| OMCI | <i>Ornithodoros moubata</i> | C5 complement | $IC_{50} \sim 12$ –27 nM $K_D \sim 18$ nM $K_D < 100$ pM | 29 μ M–29 mM | Nunn et al., 2005; Hepburn et al., 2007; Macpherson et al., 2018 |
| TSGP2 | <i>Ornithodoros kalahariensis</i> | C5 complement | $K_D \sim 26$ nM | 38 μ M–38 mM | Mans et al., 2001; Mans and Ribeiro, 2008b |
| TSGP3 | <i>Ornithodoros kalahariensis</i> | C5 complement | $K_D \sim 14$ nM | 38 μ M–38 μ M | Mans et al., 2001; Mans and Ribeiro, 2008b |
| RaCI | <i>Rhipicephalus appendiculatus</i> | C5 complement | IC_{50} –6–21 nM | Unknown | Jore et al., 2016 |

(Continued)

TABLE 1 | Continued

| Protein | Tick | Target | Affinity (K_D , K_i , K_m , IC_{50}) | Concentration (Minimum–Maximum) | References |
|---------------------|-------------------------------------|--|--|---------------------------------|--|
| Isac; Salp9; Salp20 | <i>Ixodes scapularis</i> | C3 complement | $IC_{50} \sim 200$ nM | Detected in saliva | Valenzuela et al., 2000; Soares et al., 2005; Tyson et al., 2007, 2008 |
| IRAC; IxAC | <i>Ixodes ricinus</i> | C3 complement | $IC_{50} \sim 10$ –50 nM | Unknown | Daix et al., 2007; Schroeder et al., 2007; Couvreur et al., 2008 |
| HT1-12 | <i>Ixodes holocyclus</i> | Presynapse | $IC_{50} \sim 1$ –10 mM | Detected in saliva | Chand et al., 2016 |
| Salp15 | <i>Ixodes scapularis</i> | T cell proliferation IL2 production CD4 | $K_D \sim 47$ nM | Detected in saliva | Anguita et al., 2002; Garg et al., 2006 |
| Longistatin | <i>Haemaphysalis longicornis</i> | RAGE | $K_D \sim 72$ nM | Detected in saliva | Anisuzzaman et al., 2014 |
| Adrenomedullin | <i>Ornithodoros moubata</i> | Vasodilation | $IC_{50} \sim 7$ nM | Unknown | Iwanaga et al., 2014 |
| Ra-KLP | <i>Rhipicephalus appendiculatus</i> | maxiK channels | $AC_{50} \sim 1$ μ M | Unknown | Paesen et al., 2009 |
| Ixonnexin | <i>Ixodes scapularis</i> | Plasminogen-tPA | $K_m \sim 70$ –200 nM $K_D \sim 4$ –19 nM | Detected in saliva | Assumpção et al., 2018 |

Indicated are relative affinity reported as the dissociation equilibrium constant (K_D), inhibitory constant (K_i), Michaelis constant (K_m), stoichiometric inhibition ration (SI), or inhibitory concentration (IC_{50}). Salivary gland unit (SGU) refers to a fraction of a salivary gland. Minimum and maximum concentrations were calculated based on yields per tick reported and a feeding site volume of 10 nl–10 μ l. Were indicated that proteins were detected in saliva or salivary gland extract (SGE), this was done with either western blot, affinity pull-down or proteomics.

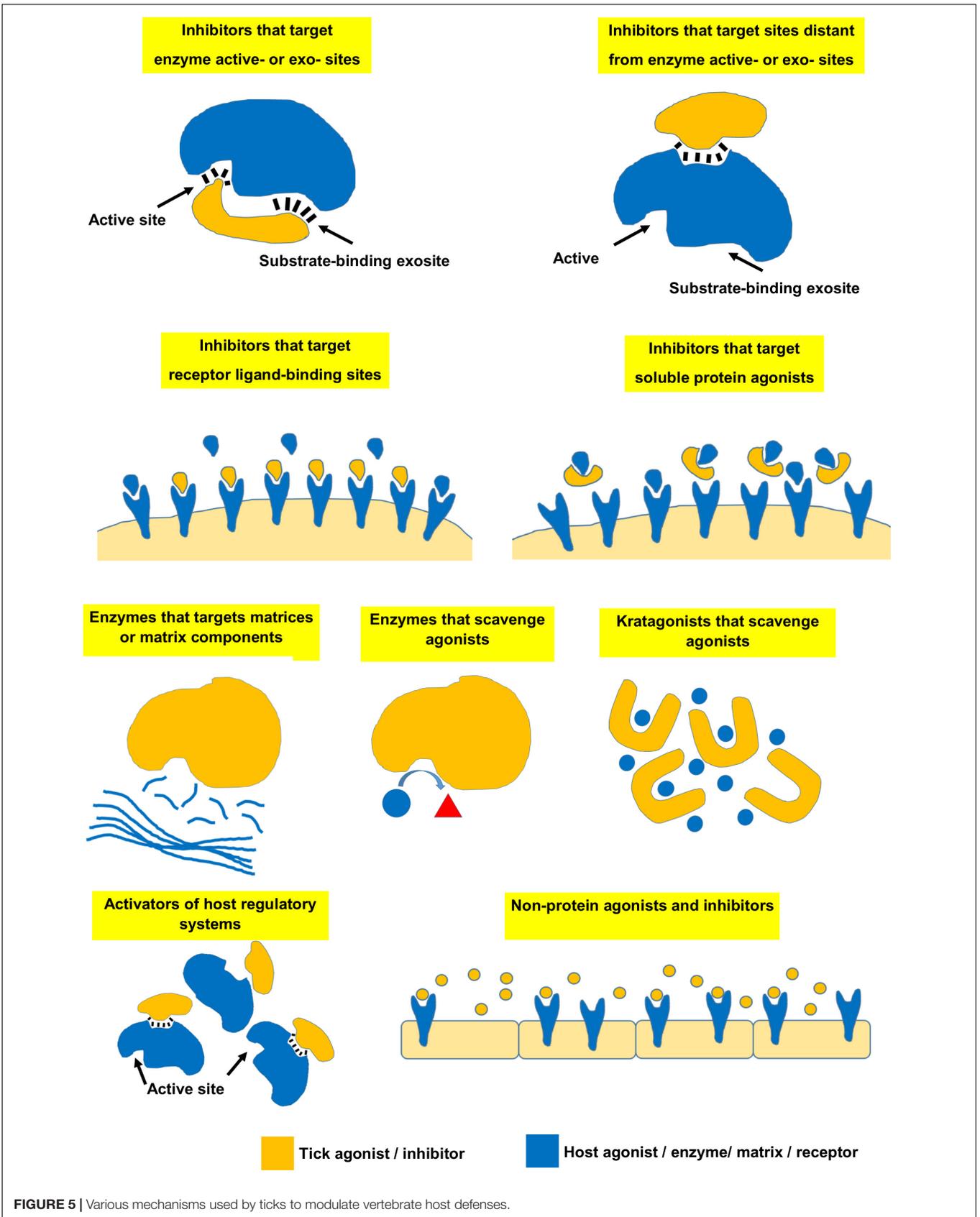
elastase, and trypsin (Table 1 and references therein). Targeting of enzyme active sites or substrate-binding exosites prevent binding of the substrate to the enzyme, thereby blocking its downstream effects. Most of these inhibitors have affinity measurements ranging from low pM to nM and for those whose concentrations could be estimated, would be present at relevant concentrations (Table 1).

With regard to host protein concentrations and affinities, prothrombin occurs at ~ 1.4 μ M in plasma, although only sub μ M quantities are generally converted to thrombin (Butenas and Mann, 2002). Fibrinogen and fibrin has a low affinity site for thrombin with $K_D \sim 2$ –5 μ M and a high affinity site with $K_D \sim 100$ nM, while occurring at a plasma concentration of ~ 7.6 μ M. In addition, the K_D for the thrombin platelet receptor GPIIb/IIIa is ~ 50 –200 nM (Adams and Huntington, 2006). It is therefore not unexpected that inhibitors would have K_D values in the low nM or pM range, while also occurring at low nM to μ M concentration ranges to allow efficient competition with fibrinogen. fX occur at a plasma concentration of ~ 170 nM, while fXa concentrations found in the clot range from 2 to 16 nM (Butenas and Mann, 2002). These concentrations also correlate with K_i values of tick inhibitors that are in the low nM range or below (Table 1). Other blood clotting factors have even lower concentrations ranging from low nM to pM concentrations in plasma (Butenas and Mann, 2002). Neutrophil derived proteases such as elastase or cathepsin G can reach concentrations of 100 nM at the site of release and is sufficient to induce platelet aggregation (Ferrer-Lopez et al., 1990). Again, the low nM IC_{50} or K_D values for the tick inhibitors correspond with the host target concentrations, supporting functional relevance at the tick feeding site (Table 1). Some clotting enzyme inhibitors have K_i or IC_{50} values in the μ M ranges (Table 1). While estimates indicate that tick proteins may achieve such high

concentrations at the feeding site, it remains crucial to confirm that they actually do.

Inhibitors That Target Sites Distant From Enzyme Active Sites

Inhibitors may target enzymes at sites distant from the active site, thereby disrupting complex assembly and downstream activation and include inhibitors of C5 complement such as *Ornithodoros moubata* complement inhibitor (OMCI) and *Rhipicephalus appendiculatus* complement inhibitor (RaCI) (Nunn et al., 2005; Jore et al., 2016). OMCI binds to the C5d, CUB, and C345c domains of C5, while RaCI binds to the MG1, MG2, and C5d domains (Jore et al., 2016). In both cases, these domains and the inhibitor binding sites are distant from C5a or the C5 convertase binding sites suggesting that these inhibitors do not directly inhibit interaction of convertases with C5, but probably inhibit rearrangement of domains within C5 that is necessary for activation to occur (Jore et al., 2016). Other complement inhibitors from soft ticks include the OMCI homologs TSGP2 and TSGP3 from *O. kalahariensis* (Mans and Ribeiro, 2008b). These inhibitors have K_D values in the low nM range (Table 1). A recent study that used site-specific immobilization of OMCI and multicycle kinetics indicated that the K_D may even be in the low pM ranges indicating that assay design can affect estimations of affinity (Macpherson et al., 2018). These inhibitors also show quite high expected concentrations at the feeding site, which is mostly due to their functioning as kratagonists (section below). Complement C5 occur at a concentration of ~ 370 nM in plasma and its activator (C3/C5 convertase) show K_m values of 5–16 nM for the classical and alternative complement pathways (Rawal and Pangburn, 2001, 2003). Therefore, even at low nM concentrations and the high concentrations of



inhibitor expected at the feeding site, the tick proteins should be functionally relevant. Hard ticks also possess complement inhibitors that belong to the Isac/IRAC family (Valenzuela et al., 2000). These inhibitors target the C3 convertase complex (C3bBbP) of the alternative pathway and dissociate this complex, preventing binding of the convertase to C3. IC₅₀ values that range from 10 to 200 nM has been reported (Table 1). Of these inhibitors, Salp20 specifically target properdin of the C3 convertase complex and bind with a $K_D \sim 0.6$ nM, which is much lower than the K_D of properdin (>85 nM) for C3b (Tyson et al., 2008). As such, these inhibitors should be functionally relevant at the feeding site although affinities for all homologs have not yet been determined. The blood-clotting inhibitor ixolaris targets the heparin-binding exosite of fXa, thereby disrupting formation of the prothrombinase complex with an IC₅₀ in the pM range (Francischetti et al., 2002; Monteiro et al., 2005). It would therefore be expected to be functionally relevant.

Inhibitors That Target Receptor Ligand-Binding Sites

Inhibitors that target cell receptors thereby blocking binding of natural ligands and receptor activation includes fibrinogen receptor (GPIIb/IIIa; $\alpha_{IIb}\beta_3$) antagonists that inhibit platelet aggregation induced by any agonist (Karczewski et al., 1994; Wang et al., 1996; Mans et al., 2002b, 2008a; Tang et al., 2015). It has been shown that these inhibitors can also compete with bound fibrinogen to disaggregate aggregated platelets (Mans et al., 2002a). Binding to the fibrinogen receptor do not seem to result in outside-in signaling by these agonists. Inhibitors that target fibrinogen receptors need to be present at high concentrations, since platelets possess high numbers of the $\alpha_{IIb}\beta_3$ integrin on their surface ($\sim 80,000$ receptors/platelet) (Wagner et al., 1996). This could result in concentrations of active receptor of ~ 39 nM at the feeding site. The affinity constant (K_D) for fibrinogen is ~ 1.4 μ M, while fibrinogen occur in 10-fold excess (Frojmovic et al., 1994). It is therefore not unexpected that inhibitors would have K_D values in the nM range to bind to the receptor, while also occurring at μ M concentrations to effectively compete with fibrinogen. Conversely, inhibitors with K_D or IC₅₀ values above 1 μ M may not be effective inhibitors.

Tick adhesion inhibitor (TAI) inhibits adhesion of platelets to collagen with an IC₅₀ ~ 8 nM (Karczewski et al., 1995). Competitive inhibition (IC₅₀ ~ 5 nM) of binding of the monoclonal antibody Gi9 to the platelet collagen receptor GPIIa-IIa ($\alpha_2\beta_1$) confirmed targeting of this receptor by TAI (Karczewski et al., 1995). The collagen receptor (GP1a/IIa) occur at low receptor numbers on platelets (800 receptors/platelet) resulting in a concentration of ~ 400 pM at the feeding site (Coller et al., 1989). The affinity of collagen for platelets is also ~ 35 – 90 nM (Jung and Moroi, 1998). It is therefore not surprising that the affinity for collagen receptor antagonists may be in the low nM range, with similar low nM concentrations at the feeding site.

Longistatin binds to the V domain of the receptor for advanced glycation end products (RAGE) with a $K_D \sim 72$ nM,

thereby inhibiting ligand-induced inflammation in tissues (Anisuzzaman et al., 2014). It has been detected at the feeding site using host antibodies and would therefore presumably be present at nM concentrations (Anisuzzaman et al., 2010). The concentration for RAGE is in the low pM ranges (Bopp et al., 2008) and longistatin should therefore be able to saturate the receptor at nM concentrations.

Inhibitors that target receptors and upon binding induce antagonistic responses do exist. Salp15 binds to the CD4 coreceptor on T cells with a $K_D \sim 47$ nM and inhibits T-cell receptor ligation induced activation resulting in immunosuppression (Anguita et al., 2002; Garg et al., 2006). Salp15 also interact with DC-SIGN on dendritic cells (DC) to activate the serine/threonine kinase Raf-1. This leads to modulation of Toll-like receptor induced DC activation (Hovius et al., 2008). However, in the latter case no affinity assessment was done, so it remains difficult to assess the biological relevance of this at the tick-feeding site. The CD4 coreceptor occurs at ~ 16 – 664 pM (Platt et al., 1998), and if Salp15 occur at concentrations equal or above its K_D for the CD4 coreceptor should saturate the receptor and will be biologically relevant. The neurotoxins from *Ixodes holocyclus* are presumed to target and inhibit presynaptic P/Q-type voltage gated calcium channels (Chand et al., 2016). Recombinant holocyclotoxins have IC₅₀ values ranging from 5 to 12 μ M, which seem to be orders of magnitude higher than the concentrations present in saliva (Chand et al., 2016), suggesting that a discrepancy still exist between the identified toxins and crude salivary composition (Pienaar et al., 2018).

Inhibitors That Target Soluble Protein Agonists

A number of soluble host protein agonists exist that play a role in inflammation and immunity. These generally bind to receptors on leukocytes to activate cellular responses and cellular migration and are collectively known as cytokines or chemokines (Sokol and Luster, 2015). Leukocytes include eosinophils, mast cells, monocytes, neutrophils and natural killer cells that migrate along chemokine concentration gradients caused by release of chemokines from sites of infection or inflammation (Moore et al., 2018). A large number of chemokines exist that have specificities for different cell types. Chemokines are classified based on their conserved disulphide bond patterns. C chemokines possess a single disulphide bond and consist of two chemokines (XCL1 and XCL2). CC chemokines (β -chemokines) possess two disulphide bonds, with adjacent cysteines near the N-terminal and consist of 28 chemokines (CCL1–CCL28), described thus far. CXC chemokines (α -chemokines) possess two disulphide bonds with the adjacent cysteines near the N-terminal separated by a single amino acid and consist of 17 chemokines (CXCL1–CXCL17), described to date. A number of inhibitors from ticks that interact directly with these soluble agonists and prevent binding to their receptors have been described and are known as the evasins (Frauensschuh et al., 2007; Déruaz et al., 2008). It has been shown that evasins can bind to a wide array of cytokines or chemokines (Table 1). Their K_D values range from low pM to nM.

In the case of chemokines where leukocytes respond to concentration gradients for directional chemotaxis, the measured gradients suggest that these may range across low pM to several hundred nM (Moore et al., 2018). Naturally formed gradients depend on various factors that will influence the concentration gradient, notably, the type of cell secreting the chemokine, the amount secreted that depend on environmental cues, the presence of flow-induced shear stress and interaction of the chemokines with the extracellular matrix (Moore et al., 2018). Depending on the K_D of the evasins (low nM), chemotaxis may only be efficiently inhibited somewhere along the concentration gradient and not across the whole range, especially since competition of evasins for different chemokines might occur.

Enzymes That Target Matrices or Matrix Components

Ticks feed from a feeding cavity where blood pools (Tatchell and Moorhouse, 1968). The feeding site needs to be remodeled to form this cavity (Wikel, 2017). Ticks may secrete a variety of enzymes that will enable such remodeling. This includes hyaluronidase that targets hyaluronic acid, a major component of the extracellular matrix (Neitz et al., 1978). Salivary transcriptomes also indicate that an abundant class of enzymes are the metalloproteases (Mans et al., 2016). While their role in feeding site remodeling has not been established beyond doubt, the general assumption is that these enzymes would play a role in remodeling (Mans, 2016; Wikel, 2017). Other metalloproteases that has been identified with a defined function include fibrin(ogen)ase activity that remove fibrinogen, both substrate for thrombin or platelet aggregation, or fibrin that forms the blood clot (Francischetti et al., 2003). Longistatin, a small EF-hand protein can hydrolyze α , β , and γ chains of fibrinogen, activates plasminogen to plasmin, degrade fibrin and dissolve fibrin clots (Anisuzzaman et al., 2011, 2012). Other enzymes that may target the fibrin clot without a direct interaction include plasminogen activators such as enolase that promote degradation of the fibrin clot via activation of the host enzyme plasminogen to plasmin (Díaz-Martín et al., 2013a; Xu X.L. et al., 2016). A serine protease that potentially activates protein C, a potent anticoagulant has also been identified in saliva of *Ixodes scapularis* (Pichu et al., 2014). For these “activating” enzymes the physiological effective concentrations in saliva may be low (pM–nM ranges) and detection of function in saliva (not salivary gland extract) may be enough to infer functional significance given their amplification/catalyzing nature.

Enzymes That Perform Scavenging Functions

Some enzymes may perform scavenging functions by targeting bioactive molecules and catalyzing chemical reactions that inactivate or remove these molecules. This may prevent activation of receptors, or induce antagonistic responses in receptors by removal of activating ligand. In ticks, the enzyme apyrase (ATP-diphosphohydrolase; EC 3.6.2.5) hydrolyse ATP that function in inflammation (Faas et al., 2017), as well as

ADP that induce platelet aggregation (Ribeiro et al., 1985, 1991; Mans et al., 1998a, 2008a). Apyrase was able to disaggregate platelets aggregated by ADP and caused platelet shape change from an activated spherical back to discoid form (Mans et al., 1998b, 2000), suggesting that bound ADP could be scavenged from its platelet receptor. Apyrase activity has been found in saliva or salivary glands of most ticks studied and have been assigned to the 5'-nucleotidase family (Stutzer et al., 2009). Family members have been found in all tick transcriptomes studied to date (Mans et al., 2016). Kinetic parameters from purified apyrase indicated a $K_m \sim 1$ mM for ATP and ADP and high turnover number (10^6 s⁻¹) and K_{cat}/K_m ratio (10^9 M⁻¹s⁻¹) (Mans et al., 1998b). These numbers indicate a highly efficient enzyme that would rapidly hydrolyze high local concentrations of ADP or ATP. Given the relatively high concentration of apyrase found in tick salivary glands (Mans et al., 1998b), it would probably also be functionally relevant. Another enzyme found in ticks that perform scavenging functions is a metallo dipeptidyl carboxypeptidase responsible for salivary kininase activity and breakdown of anaphylatoxin and bradykinin, involved in inflammation, pain and vasoconstriction (Ribeiro and Spielman, 1986; Ribeiro and Mather, 1998; Bastiani et al., 2002). In all cases of “scavenging” enzymes, the enzymes need to be able to rapidly remove host-derived agonist to levels below their functional ranges.

Kratagonists That Perform Scavenging Functions

Kratagonists are related to “scavenging” enzymatic functions, by scavenging or mopping up of bioactive molecules, but without chemically changing their structures. Kratagonists may have similar functional activity as “scavenging” enzymes, such as preventing activation of receptors by ligands, or competitive removal of ligand from receptors causing an antagonistic response (Ribeiro and Arcà, 2009; Andersen and Ribeiro, 2017). The term kratagonist was recently coined to describe the abundant proteins found in saliva of most hematophagous organisms that function in a scavenging capacity (Ribeiro and Arcà, 2009). The name derives from the Greek “to arrest or to seize” and was appropriately proposed independently by the Greek compatriots Babis Savakis and Michalis Kotsyfakis (Ribeiro and Arcà, 2009). Recently the etymology of the name was redefined to indicate “hold” or “grab/capture” and “agonist” (Andersen and Ribeiro, 2017; Arcà and Ribeiro, 2018). In ticks a large number of kratagonists have been described, that all belong to the lipocalin family. Lipocalin structure is composed of an eight stranded anti-parallel β -barrel closed off at one end by an N-terminal 3_{10} -helix, with a C-terminal α -helix anchored to the side of the barrel by disulphide bonds. This gives lipocalins the distinct appearance of a cup with an open end, where ligands can access the cup and bind in the cavity inside the barrel. Specificity is conferred by residues inside the barrel, as well as four loops that allow access to the barrel (Flower, 1996). The original name of lipocalin was assigned to “extracellular proteins capable of enclosing lipophiles within their structure to minimize solvent contact”

(Pervaiz and Brew, 1987). Subsequently, lipocalins were defined based on conserved sequence or structural motifs (Flower, 1996). Scavenging functions performed by tick lipocalins include scavenging of histamine and serotonin (Paesen et al., 1999; Sangamnatdej et al., 2002; Mans et al., 2008b; Neelakanta et al., 2018), leukotriene B₄ (Beaufays et al., 2008; Mans and Ribeiro, 2008b; Roversi et al., 2013), leukotriene C₄ (Mans and Ribeiro, 2008a), thromboxane A₂ (Mans and Ribeiro, 2008b), and cholesterol (Preston et al., 2013; Roversi et al., 2017).

In the case of small chemical agonists that are scavenged by either enzymes or kratagonists, the concentrations at which they activate their respective receptors are important. Platelets secrete ADP and ATP at 3–7 μM concentrations, which are also the concentration necessary for primary and secondary aggregation (Packham and Rand, 2011). Apyrase from ticks has been shown to effectively inhibit platelet aggregation at these activator concentrations, at enzyme concentrations well below the expected secretory levels (Mans et al., 1998b). Basophils and mast cells may secrete histamine to attain local concentrations of 20 μM in the skin that can lead to inflammatory responses when histamine binds to its receptors with $K_D \sim 10 \text{ nM}$ –30 μM (Petersen, 1997; MacGlashan, 2003). Serotonin is secreted by platelets at local concentrations of $\sim 5 \mu\text{M}$ where it can cause vasoconstriction and platelet aggregation by binding to various serotonin receptors with $K_i \sim 10 \text{ nM}$ –1 μM (Watts et al., 2012). The very high concentrations of biogenic amine binding kratagonists at the feeding site (μM – mM) and their low affinities (low nM) indicate that they will be biologically relevant at the feeding site (Table 1).

Leukotriene B₄ secreted by neutrophils may reach high concentrations at the site of neutrophil release ($\sim 950 \text{ nM}$) (Lewis et al., 1982). The affinity of LTB₄ for its neutrophil receptor BLT1 ranges from 0.1 to 2 nM (Yokomizo, 2015). Again, the low affinities observed for the LTB₄ scavengers and their high concentrations (μM) at the feeding sites would make them relevant competitors at the feeding site.

Cysteinyl leukotrienes can only be detected in plasma during inflammatory reactions such as asthma attacks and then occur at concentrations of ~ 100 –765 pM (Sasagawa et al., 1994). It has been shown to cause vasoconstriction and vasopermeability at a concentration of $\sim 100 \text{ nM}$, and bind with affinities from 5 to 35 nM to its receptors (Drazen et al., 1980; Krilis et al., 1983; Ghiglieri-Bertez et al., 1986; Prié et al., 1995). Scavengers of LTC₄ and LTD₄ have K_D values below 2 nM and also occur at μM concentrations (Table 1). It is therefore also expected that they would be physiologically relevant during feeding.

Thromboxane A₂ is released from platelets at concentrations of 11–35 nM, which is capable of inducing platelet aggregation and bind to platelet receptors with $K_D \sim 4 \text{ nM}$ (Hamberg et al., 1975; Dorn and DeJesus, 1991). The high concentrations of the TXA₂ kratagonists at the feeding site (μM – mM) and their low affinities (low nM) suggest that they would be able to neutralize TXA₂ binding to its receptor during feeding (Table 1).

Activators of Host Regulatory Systems

The majority of tick host modulatory mechanisms described so far comprise inhibitors. However, activators may also play important roles since these may target the natural regulatory feedback systems of host hemostasis. Plasminogen activators that result in degradation of the fibrin clot have already been discussed. Recently, a small peptide named ixonnexin that belongs to the basic tail family was described that act by promoting interaction of plasminogen and tissue plasminogen activator (tPA) by forming an enzymatically productive ternary complex that forms plasmin to promote fibrinolysis (Assumpção et al., 2018). Ixonnexin interacts with both plasminogen and tPA with similar K_D values (4–20 nM). Such an activator would need to be present at similar or higher concentrations than the target enzymes to be functionally relevant. In the case of ixonnexin, plasminogen and tPA occur at plasma concentrations of $\sim 2 \mu\text{M}$ and 100 pM, respectively (MacGregor and Prowse, 1983; Assumpção et al., 2018). While no exact concentration has been established for ixonnexin, it has been estimated to occur at high concentrations in saliva (Assumpção et al., 2018). It has also been suggested that all basic tail proteins may perform this function, including the fXa inhibitor Salp14, since this family in which the C-terminal is rich in basic amino acids such as lysine and arginine mimics the C-terminal lysine present in fibrin that serves as recognition site for plasminogen and tPA lysine binding sites (Assumpção et al., 2018).

The adrenomedullins are a special case of tick-derived inhibitors that bind to host calcitonin-receptor-like receptor and receptor activity-modifying protein receptor complexes to cause vasodilation (Iwanaga et al., 2014). These inhibitors have not converged to mimic host adrenomedullin, but have been acquired via horizontal gene transfer from a mammalian host (Iwanaga et al., 2014). At systemic concentrations of $\sim 7 \text{ nM}$ it reduced blood pressure by almost 50% (Table 1). It may therefore be assumed that secretion of such low quantities during feeding may result in local vasodilatory effects. However, its presence in saliva at functional concentrations still needs to be confirmed.

An activator of MaxiK channels have been described in *R. appendiculatus*, where it presumably play a role in regulation of blood vessel tonus and blood flow (Paesen et al., 2009). It invoked a half maximal response in MaxiK channels at 1 μM . Its functionality at the feeding site remains to be resolved.

Non-protein Agonists and Inhibitors

Ticks may also secrete non-protein agonists that could affect the host's defense mechanisms. As such, ixodid ticks secrete prostaglandin E₂ (PGE₂), a potent vasodilator, at ~ 40 –500 ng/ml saliva (Ribeiro et al., 1985, 1992; Inokuma et al., 1994). Concentrations of PGE₂ may range from 79 nM to 994 μM in adults at the feeding site, which would be pharmacologically active, since the K_D of the PGE₂ receptor is $\sim 0.7 \text{ nM}$ (Davis and Sharif, 2000). Another non-protein mechanism for modulating host inflammatory and pain

sensing responses was recently reported that involved secretion of saliva-specific microRNAs that was detected in salivary exosomes using next-generation sequencing (Hackenberg et al., 2017; Hackenberg and Kotsyfakis, 2018). The efficiency of exosomal miRNA will depend on the concentrations of exosomes in saliva, the specificity of the exosomes for specific cell types and the concentration of miRNA inside the exosomes (Hu et al., 2012). As yet, the functional relevance of saliva-derived exosomal miRNA still needs to be confirmed, with their kinetics of inhibition resolved, since the question remains whether a single exosome would only target a single lymphocyte or epithelial cell, in which case the effectiveness of inhibition would be determined by the number of cells that can be neutralized at the feeding site. It would, however, add another complex repertoire to the ticks expanding modulatory mechanisms.

STRATEGIES TO CIRCUMVENT THE AFFINITY/EQUILIBRIUM BARRIER

It is evident that the majority of tick proteins thus far characterized would have physiological functionality at the feeding site (Figure 3 and Table 1). However, expression and secretion of high concentrations of secretory proteins via the salivary secretory granules may be restricted to very few proteins given physical constraints of granule packaging (Mans and Neitz, 2004b). The dynamic nature of the feeding site may also make the actual concentrations of tick proteins present at any given moment much more haphazard than expected. The concentrations at the feeding site may very well be 100–10,000-fold lower than estimated, in which case concentrations may drop to pM–nM for most proteins, which may be below the K_D values for many proteins. To address this, ticks may employ various alternative strategies that allow adequate expression and optimal use of secreted proteins at the feeding site that allow ticks to circumvent or eliminate the problems posed by an equilibrium system (Figure 6). The next section discusses such strategies in more detail.

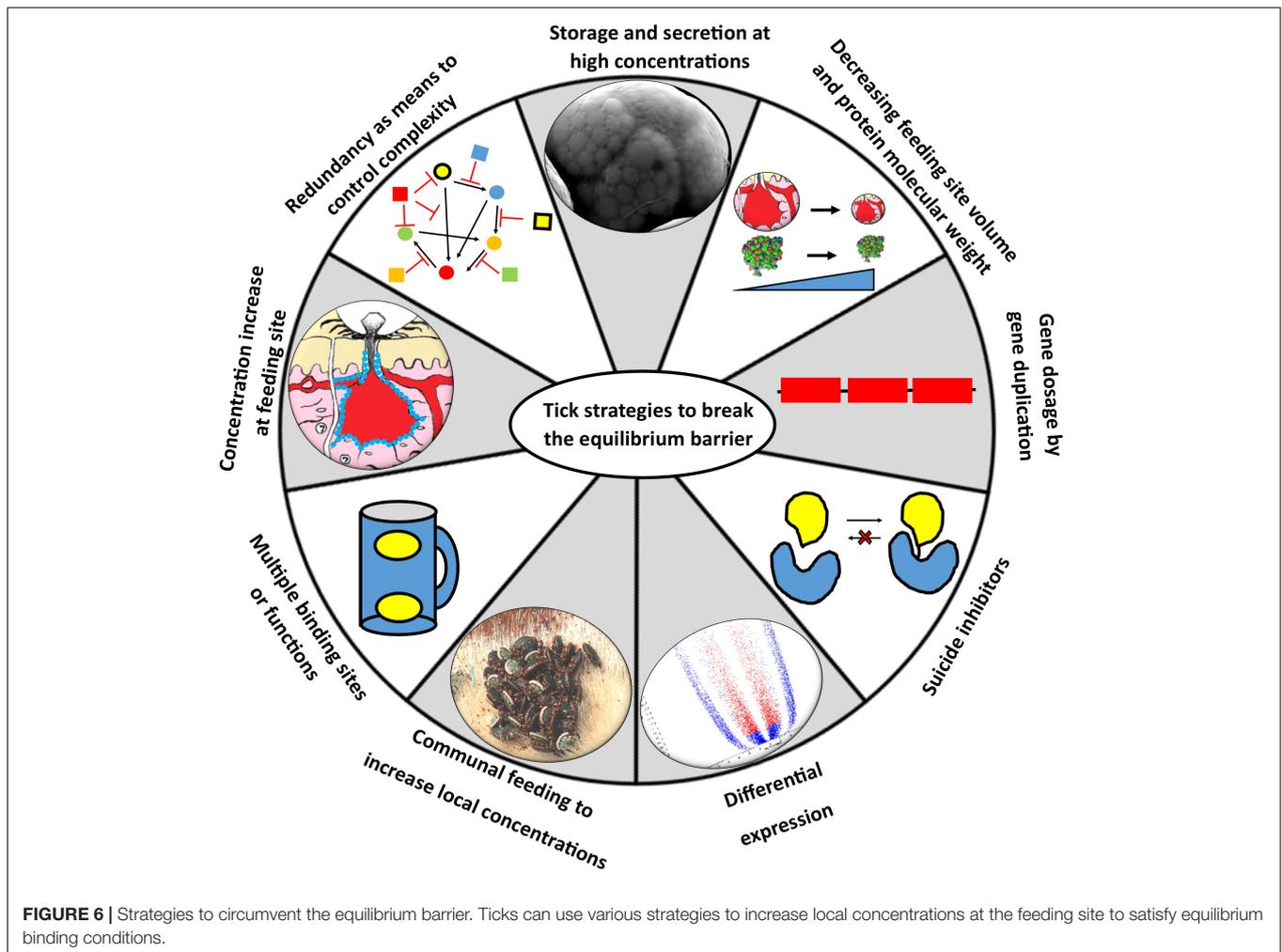
Redundancy as Means to Control Complexity and Chaos

The host's defense systems such as the clotting, platelet aggregation, complement and inflammatory cascades are part of one big redundant integrated feedback system that allows rapid response as well as control of the system (Delvaeye and Conway, 2009; Deppermann and Kubes, 2016; Wiegner et al., 2016). It has been argued that this redundancy is mimicked in the complexity of functions observed in the salivary glands of blood-feeding arthropods (Ribeiro, 1995). Conversely, it has been argued that the complexity observed in the functional salivary repertoire of ticks is due to a highly optimized system of defense shaped by evolution (Nuttall, 2019). However, redundancy may serve the purpose of dealing with a highly dynamic and chaotic system, as may be seen at the feeding site, which is in constant flux with ever changing concentrations of target and inhibitor molecules.

Targeting of many different host proteins at once, even if not at optimal concentrations, may disrupt a redundant system enough to allow successful feeding. In this environment no protein needs to be 100% effective, but only good enough to get the job done. This may allow non-optimal proteins to function at the feeding site, or be maintained while evolving more optimized functions as postulated in the playground hypothesis of neutral evolution (Mans et al., 2017). The question then becomes a matter of how much inhibition of the host's defense systems would be necessary to ensure successful feeding. In this regard, three observations could be made: firstly, that inefficient inhibition was surely a given during the early stages of adaptation to a blood-feeding lifestyle since protein functions were still optimized by natural selection. Secondly, since ticks had to switch hosts throughout their evolution, we have no evidence that ticks are not still adapting to new hosts, and functions that seem to be only partially effective may indeed still be optimized in future through natural selection. Thirdly, given the relatively high concentrations that may be obtained at the feeding site, even if proteins may only be partially effective, may allow for selection of these non-optimal functions to improve efficiency, again supporting a neutral evolution of function hypothesis (Mans et al., 2017). These arguments should, however, not serve as a *carte blanche* to support every claim of functional significance in blood-feeding, since the contribution of salivary derived proteins to species fitness has not been elucidated yet or even proven beyond doubt.

Fast Feeding, Storage and Secretion of a Large Bolus of Salivary Proteins

Soft ticks feed within minutes to hours to repletion, drop off, digest the blood meal slowly over the course of weeks to months, lay a small egg batch and can then feed several times more using the same pattern (Mans and Neitz, 2004a). During fast feeding most salivary proteins may be secreted in the course of 10–30 min and will be replenished within several days after feeding. Secretory proteins are stored in large granules up to 10 μM in diameter that effectively fill the salivary gland cells to their maximum extent (Mans and Neitz, 2004b; Mans et al., 2004b). The protein profiles of soft tick SGE attest to this, since protein spectra rarely show the presence of genomic DNA, in contrast to SGE from hard ticks that predominantly show genomic DNA/RNA, while the protein peak and concentration is obscured (Figure 7). This strategy from soft ticks allows concentrations of proteins that can overcome relatively high equilibrium dissociation constants by sheer concentration effects alone. Hard ticks utilize this strategy to some extent, since different salivary gland cells are filled with secretory granules over the course of the feeding period that can last several days to weeks (Binnington and Kemp, 1980). However, the amount of protein found in crude salivary gland extract rarely exceeds the concentrations of genomic DNA and is generally less than what would be observed for soft ticks (Figure 7), and may range from 5 to 60 μg total soluble protein over the course of feeding for a small tick such as *R. appendiculatus* (Wang and Nuttall, 1994). Similarly,



concentrations in pilocarpine-induced salivary secretions in hard ticks are generally lower than soft ticks, with soft ticks attaining ~20–40 mg/ml and hard ticks ranging from 700 to 60 $\mu\text{g/ml}$ over the course of feeding (Howell et al., 1975; Ribeiro, 1987; Ribeiro et al., 1991; Dharampaul et al., 1993; Chand et al., 2016).

Communal Feeding to Increase Local Concentrations at the Feeding Site

Hard ticks feed for several days to weeks and mating on the host (or off the host for some *Ixodes* species) during the slow pre-feeding phase is required before the rapid engorgement phase can occur (Kiszewski et al., 2001). To allow mate finding, males secrete attraction-aggregation-attachment pheromones that result in ticks clustering and co-feeding at the same feeding site (Sonenshine, 2004). In a similar manner, specific tick species generally have preference sites of attachment on the host, presumably due to host environmental cues. While males do not engorge or take a significant blood meal, they do attach and secrete salivary components into the communal feeding site that apparently

assist females in blood meal acquisition (Wang et al., 1998, 2001a). Creation of a communal locality where all ticks contribute to the localized but systemic feeding site may result in combined concentrations that overcome affinity restricted barriers.

Gene Dosage and Cumulative Contributions From Multigene Families

Proteins may be maintained as gene duplicates to allow high level expression from each family member (Mans et al., 2017). While each member may by itself completely inhibit host functions, even partial inhibition by each gene duplicate can result in complete inhibition. This is a relatively simple example of the gene dosage effect that has been observed for fibrinogen receptor antagonists (Mans et al., 2003a, 2008a) and LTB_4 scavengers (Mans et al., 2003b; Mans and Ribeiro, 2008b). It has been proposed that multigene families express different antigenic variable family members at low concentration levels to escape the immune system, the varying epitope hypothesis (Couvreux et al., 2008; Chmelař et al., 2016b). In this scenario, the low-level expressed proteins cumulatively target

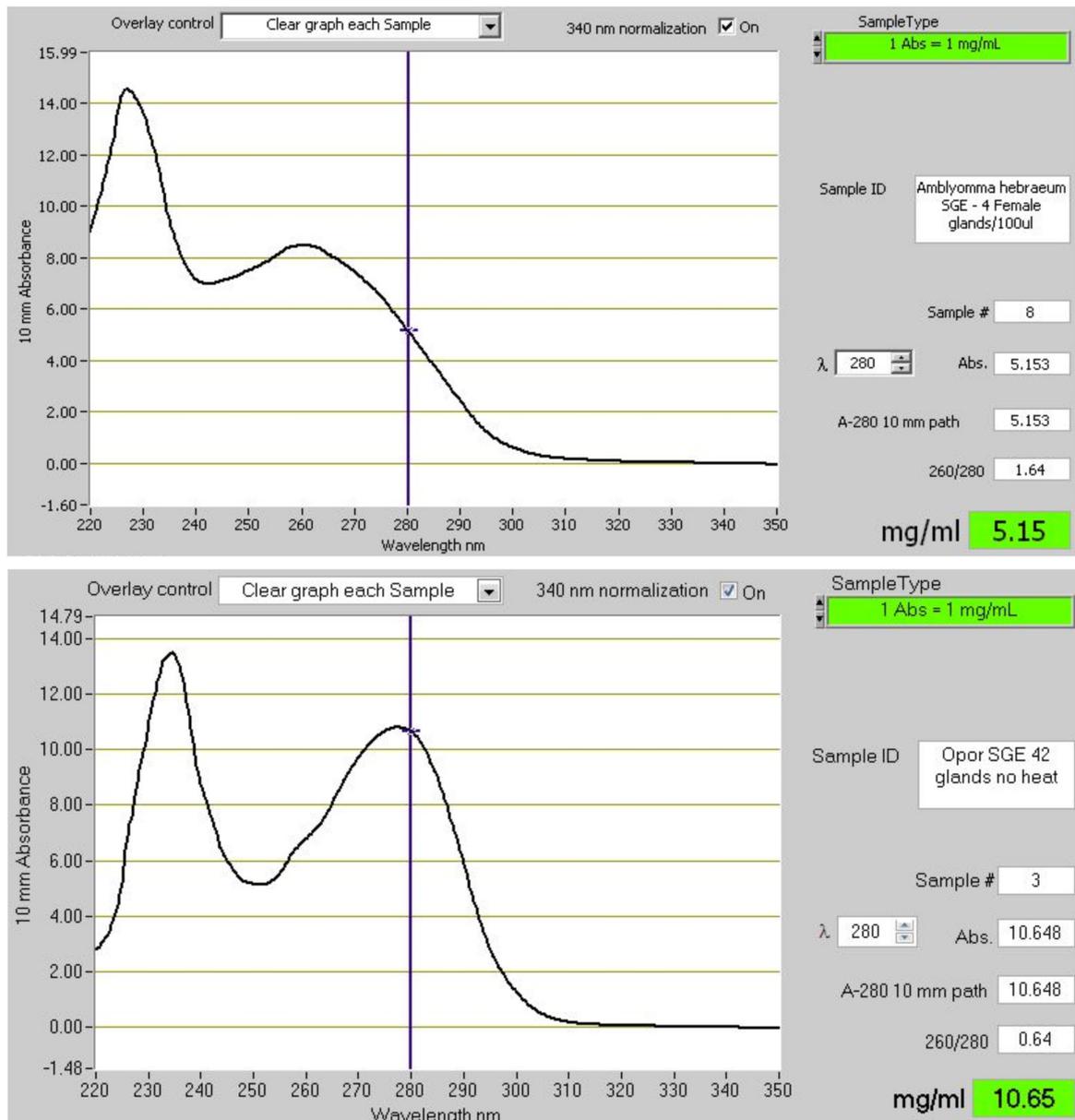


FIGURE 7 | The absorbance spectra of salivary gland extracts (SGE) from hard and soft ticks. The top graph shows the spectrum of SGE of 4 glands from *Amblyomma hebraeum* suspended in 100 μ l water. Based on the 280 nm absorbance a single gland would have a soluble protein concentration of \sim 128 μ g. Bradford determination indicated a concentration closer to \sim 60 μ g/gland. The bottom graph shows the spectrum of SGE of 42 glands from *Ornithodoros phacochoerus* suspended in 500 μ l water. Based on the 280 nm absorbance a single gland would have a soluble protein concentration of \sim 127 μ g. Bradford determination indicated a concentration of \sim 125 μ g/gland. These absorbance spectra are representative of hard and soft tick SGE in general.

the same receptor to attain a combined concentration that would allow receptor saturation and inhibition. This may be possible, even if expressed at concentrations well below the equilibrium dissociation constant, since the inhibitory effects may be summed if the inhibitors act in a mutually exclusive manner (1:1 receptor-ligand binding with no synergistic effects) (Chou and Talalay, 1977). A cumulative effect was recently shown for the holocyclotoxins from the paralysis tick, *Ixodes holocyclus*, where at least 19 holocyclotoxin genes were expressed

at low levels that showed a cumulative paralysis effect when combined (Rodriguez-Valle et al., 2017). It should be noted that while immune evasion was proposed as reason for the multiplicity of gene family members, low level expression of multiple genes to attain a cumulative threshold concentration that allows effective inhibition may also be possible and may fit with the overall neutral evolution of tick salivary gland proteins previously proposed (Mans et al., 2017). Low-level expression may in this scenario also be a function of the

constitutive differential expression observed in hard ticks, where the extreme high expression levels observed in soft ticks due to accumulation in granules may not be attainable. Even so, it remains to be quantitatively proven that cumulative contributions to attain a threshold concentration occur at the feeding site.

Increasing Concentration at the Feeding Site

The vertebrate host uses localized cues and responses to maintain and regulate haemostasis. As such, wounds or breakdown in system integrity are detected by exposure of localized collagen or other extracellular matrix components, activation of platelets and exposure of procoagulant platelet surfaces, initiation of blood-clotting and the complement cascade on activated platelet surfaces (Delvaeye and Conway, 2009; Deppermann and Kubes, 2016; Wiegner et al., 2016). In a similar manner, targeting of tick-derived bioactive molecules to activated surfaces, whether tick or host surfaces can increase local concentrations and prevent loss or dilution of components via blood meal ingestion or systemic diffusion. Salivary proteins have been found in tick cement where they presumably function as inhibitors of host defenses, or to prevent recognition of cement as a foreign or activation surface for host defenses (Bullard et al., 2016). Proteins such as apyrase have asymmetric charged surfaces (Stutzer et al., 2009), suggesting that these would be attracted to activated negatively charged platelet membranes. Other tick proteins has been shown to interact with membranes or have higher inhibitory activity in the presence of membranes, suggesting that these proteins may be concentrated on activated membrane surfaces at the feeding site (Ehebauer et al., 2002; Schuijt et al., 2013). Some Kunitz-domain inhibitors such as ixolaris have long carboxy-termini rich in serine and threonine residues that can be the site of glycosylation (Francischetti et al., 2002). These would create mucin tails that would be sticky and allow concentration on the walls of the feeding cavity preventing removal and increasing concentration (Francischetti et al., 2009). This may be a general strategy for glycosylated tick proteins (Uhlir et al., 1994).

Increasing Effective Concentration Using Multiple Binding Sites or Multiple Functions

In the case of some tick lipocalins their effective concentration is increased by possessing two independent binding sites for histamine or one for histamine and one for serotonin (Paesen et al., 2000; Sangamnatdej et al., 2002; Mans et al., 2008b). This increase their capacity for scavenging twofold. Another variation on this may be proteins that can potentially target many independent ligands or targets and may therefore have multiple functions. While this certainly expands the potential of proteins to work within a complex redundant environment, the most effective of these would be proteins that have independent function and mechanisms. For example, targeting of complement C5 and LTB₄ allows OMCI and its homologs to inhibit both complement and LTB₄ mediated inflammation at the same time (Nunn et al., 2005; Mans and Ribeiro, 2008b;

Roversi et al., 2013). In this case, both functions target the two key non-redundant mediators of neutrophil recruitment during inflammation that seems to be intricately linked (Sadik et al., 2018). This is a remarkable convergence of function in a single protein (Mans et al., 2017). On the other hand, if the functional mechanisms are not independent, i.e., use the same site for ligand binding or the same protein surface to target different proteins, competition between targets will effectively lower the concentration of available inhibitor, thereby impacting its functionality. This may be seen for moubatin and its homologs that bind both LTB₄ and TXA₂ within the same lipocalin cavity at similar affinities (Mans and Ribeiro, 2008b). If both LTB₄ and TXA₂ are present at similar concentrations, the effective scavenging capability may be halved. One way to compensate for this may be multiple proteins or very high concentrations at the feeding site as observed for moubatin, TSGP2 and TSGP3. The same problem would face many of the enzyme inhibitors that can inhibit two or more enzymes from a specific class such as the cystatins, where the presence of many different host enzymes at the feeding site would lead to competition with the tick inhibitors. Similarly, the evasins can bind to many chemokines with similar affinities that may lead to competition of evasins for different chemokines, thereby reducing their effectivity.

Suicide Inhibitors

Inhibitors that bind in a 1:1 manner to their target enzymes, and then serve as substrate, resulting in the formation of a covalent enzyme-inhibitor complex, circumvent the equilibrium dissociation problem completely. Their initial affinities need only to be high enough for the enzyme to catalyze its reaction and form the covalent complex to result in permanent inhibition. In ticks, such inhibitors are found in the serpins that target various enzymes of the clotting cascade (Chmelař et al., 2017). Serpins form covalent complexes with their respective serine proteases after cleavage (Whisstock et al., 2010).

Differential Expression

Ixodid ticks express different proteins at different periods during feeding (McSwain et al., 1982; Paesen et al., 1999; Wang et al., 2001b; Kim et al., 2017; Perner et al., 2018). Possible reasons proposed for this include antigenic variation to escape the immune system (Chmelař et al., 2016b), or responses to changes at the feeding site such as wound healing (Francischetti et al., 2005). Differential expression may also occur as salivary gland morphology changes during the course of feeding and different cell types play different roles in the feeding process (Binnington and Kemp, 1980). As such, differential expression may allow increased concentration spikes of proteins in specific feeding windows.

Minimizing Feeding Site Volume

Comparison of adult, nymphal, and larval concentrations at the feeding site indicates that the relative concentrations remain constant between the various life stages even though salivary gland concentrations may differ several fold (Figure 3). This is mostly due to the differences observed in feeding cavity

size that scale relative to the life stage and tick size. If this observation holds for all tick species it would imply that smaller ticks compensate by creating smaller feeding cavities. Feeding cavity size may then be related to protein concentration secreted during feeding.

Smaller Proteins Allow Higher Molar Concentrations

The majority of secretory salivary proteins in ticks have low molecular masses below 25 kDa. This include the majority of highly abundant protein families such as the basic tail secretory, Kunitz-BPTI and lipocalin families (Mans et al., 2008c, 2016; Mans, 2011). Lower molecular masses allow higher relative molar concentrations at the feeding site, which could result in an up to a 10-fold difference between a 5 and 50 kDa protein (Figure 3).

FUNCTIONAL RELEVANCE AND MECHANISM

Understanding the mechanism of action (how a given protein works) is important in accessing functional relevance. As example, the case of moubatin may be considered. Originally moubatin was identified as a specific inhibitor of collagen-induced platelet aggregation with an $IC_{50} \sim 50$ nM, that did not affect ADP, arachidonic acid, thrombin, ristocetin, or calcium ionophore A23187 induced platelet aggregation (Waxman and Connolly, 1993). Subsequently, recombinant moubatin was shown to inhibit collagen-induced platelet aggregation with $IC_{50} \sim 100$ nM but did not inhibit adhesion to collagen (Keller et al., 1993). At high concentrations (5.8 μ M) of moubatin and low concentrations of ADP (2 μ M) 40% inhibition of ADP-induced platelet aggregation was observed, suggesting that the cyclooxygenase pathway may be targeted. It was also shown that moubatin at these high concentrations could inhibit the TXA_2 mimetic U46619 and competed with the TXA_2 receptor antagonist SQ29548 for binding to platelet membranes with $IC_{50} \sim 10$ μ M (Keller et al., 1993). The leech inhibitor LAPP did not compete with SQ29548 binding to platelets, indicating that different receptors were targeted. Moubatin also did not inhibit binding of the monoclonal antibody Gi9 that inhibited adhesion to collagen and interacts with integrin $\alpha_2\beta_1$, the proposed receptor for adhesion to collagen. At the time, moubatin did not share sequence homology with any inhibitor of collagen-induced platelet aggregation or with collagen, and did not contain the RGD motif important in integrin recognition. From the complex data above it was suggested that moubatin might be a TXA_2 receptor antagonist (Keller et al., 1993). Subsequently, it was shown that moubatin's mechanism of collagen-induced platelet aggregation is exclusively via scavenging of TXA_2 with $K_D \sim 20$ nM, by gain and loss of function mutations in TSGP2 and TSGP3, respectively, two closely related homologs (Mans and Ribeiro, 2008b). In retrospect, it may be considered that the only inhibitory paradigms at the time were interaction with either collagen or specific platelet receptors and that the kratagonist paradigm as formulated recently did not exist. Interpreting the moubatin results from the previous paradigms may have been

logical, even if the high IC_{50} observed for SQ29548 should have raised flags. It may now be suggested that moubatin was scavenging SQ29548 (a TXA_2 mimetic), albeit with low affinity and did not compete for the receptor. Several lines of evidence converged on moubatin as scavenger of TXA_2 : the evidence that moubatin belonged to the lipocalin family (Paesen et al., 1999; Mans et al., 2003b); the fact that lipocalins are highly abundant in salivary glands and that abundant proteins generally act as scavengers, i.e., the kratagonist paradigm (Mans et al., 2001, 2003b; Mans and Neitz, 2004a,b; Calvo et al., 2006); the inhibitory effect of moubatin on the TXA_2 mimetic U46619 (Keller et al., 1993), and the observation that a closely related protein, OMCI bound ricinoleic acid, suggesting that moubatin may bind prostaglandins and thromboxanes (Roversi et al., 2007). The gain of function mutation in TSGP2 of R85G and a similar complete loss of function for TSGP3 with the mutation G85R, allowed unambiguous confirmation of functional relevance as TXA_2 scavengers (Mans and Ribeiro, 2008b). It not only highlighted the reductionist paradigm in elucidation of function, but also showed how elucidation of mechanism may inform on which function is considered relevant. As such, moubatin is an inhibitor of collagen-induced platelet aggregation, but perform this function by scavenging the secondary agonist TXA_2 . Its mechanism is primarily as kratagonist and not as receptor or ligand neutralizing inhibitor. Once mechanism is clarified the parameters necessary to assess functional significance can be better defined. In this case, that any homolog has to bind TXA_2 in the low nM range, be present at high concentrations and possess the R85G substitution to be functionally relevant.

SYSTEMS BIOLOGY, BIOINFORMATICS AND FUNCTIONAL RELEVANCE

While the majority of functions found in ticks may be assigned functional significance (Table 1), enough reports in the literature indicate that caution should be exercised when evaluating functional relevance. This is compounded by advances in technology that allow systems approaches to the analysis of salivary gland protein dynamics and bioinformatics that allows functional analysis *in silico*. As such, recent advances in technology, both in next-generation transcriptome sequencing and proteomics, has allowed an unprecedented view of salivary gland dynamics from a systems perspective (Schwarz et al., 2014; Chmelař et al., 2016a; Mans et al., 2016; Kim et al., 2017). This has indicated how ticks differentially up- or down-regulate proteins during various time intervals in feeding, which suggest that ticks actively respond to the feeding environment, reflecting a fine tuned adaptation to the hosts defense mechanisms. While the systems paradigm clearly show how dynamic expression may be in ticks, inferences regarding function rests on inference by homology or annotation. For example, a recent excellent proteomic study followed the expression profile of *I. scapularis* sampled every 24 h until detachment and indicated differential expression for a large number of lipocalins annotated as histamine-binding proteins (Kim et al., 2017). The discussion

focused on the functionality of lipocalins as histamine scavengers and how the data would support tick responses to host immunity and feeding. Interestingly, a functional study into biogenic amine binding lipocalins that specifically targeted *I. scapularis* lipocalins with biogenic amine binding motifs, failed to find any histamine-binding lipocalins, but only identified serotonin-binding lipocalins with a single binding site (Mans et al., 2008b). While this does not exclude the possibility that histamine-binding lipocalins exist in *Ixodes* ticks, the results thus far do not support it. Similarly, bioinformatic analysis predicted high affinity binding of histamine and serotonin in lipocalins from *Ixodes ricinus*, for which none of the critical residues involved in biogenic amine binding was conserved (Valdés et al., 2016). Molecular docking also recently predicted nM affinities for cystatins from *Ixodes persulcatus* without experimental verification of affinities or target enzymes (Rangel et al., 2017). While such bioinformatic and systems approaches can certainly direct research to proteins of interest, the data should be used with caution to infer functional relevance, especially if the possibility exists that some of these proteins may be only transiently expressed or at concentrations too low for functionality. For systems biology to come of age, we need accurate dissection of the feeding site in real-time to quantify fluxes in protein concentration while performing quality control with validated functions.

IS FUNCTIONAL RELEVANCE RELEVANT?

It may be considered whether it really matters whether a measured function is relevant during feeding, given the emerging recognition that all proteins may be moonlighting to some extent. From this perspective, any function present in tick saliva should be relevant at some level and our goal for the next few decades would be to assign functions to salivary proteins, whether relevant or not. A more comprehensive understanding would later emerge once we have gathered enough data to truly assess relevance. This position is appealing since it buys some time for dubious functions. It is, however, also a philosophical “everything goes” viewpoint (Feyerabend, 1975), that makes distinguishing important from trivial function very difficult. The same issue has been raised with regard to whether all ticks are venomous, or whether only some ticks secrete toxins that may cause the various well recognized forms of tick paralysis and toxicoses (Pienaar et al., 2018). By treating all ticks as venomous, the meaning of toxicity is obscured. Similarly, by treating all functions in saliva as relevant at the feeding site, even if their functional parameters suggest that they would not be relevant, may obscure those central in the feeding process from peripheral functions.

FUNCTIONAL RELEVANCE FROM A PRACTICAL PERSPECTIVE

The use of tick salivary proteins as therapeutic agents within a clinical or pharmaceutical setting remains an important

and promising goal (Mans, 2005). From this perspective any function determined for a protein need not be functionally relevant at the tick feeding site, as long as the specific parameters for use has been determined that would allow it to function under clinical or therapeutic controlled conditions. For example, the half-life of OMCI could be improved >50-fold by PASylation, making it more relevant for clinical use (Kuhn et al., 2016). In a similar vein, anti-tick vaccines may be developed against proteins with unknown functions or even irrelevant function, as long as the vaccine shows efficacy, as for example for hidden antigens (Nuttall et al., 2006). On the other hand, development of vaccines against exposed antigens may work better if antigens with real functional significance at the feeding site can be defined, their mechanism of action elucidated and this information used to rationally design target strategies that would neutralize function at the feeding site effectively.

CONCLUSION

Functional relevance is determined by the concentration of tick proteins at the feeding site as well as their affinity for their respective host targets. The current review showed that the majority of proteins found in tick saliva or salivary glands thus far characterized will be functional at the tick feeding site. It was also shown how ticks may circumvent the problems presented by an equilibrium system. Even so, inferring functional relevance without estimating concentration or affinity at the feeding site remains a risky endeavor. Future aims in salivary gland biology should focus on quantification of protein concentration secreted during feeding as well as in the actual feeding site. This should provide more accurate estimates of functional relevance.

AUTHOR CONTRIBUTIONS

BM conceptualized the study and wrote the manuscript.

FUNDING

This work was supported by the Economic Competitive Support Programme (30/01/V010), the National Research Foundation (NRF) Incentive Funding (IFR2011032400016) for Rated Researchers (NRF-Mans), and the National Research Foundation (NRF) Competitive Funding for Rated Researchers (CPRR180101296400). The funding bodies had no role in study design, data collection, analysis and interpretation, decision to publish, or preparation of the manuscript.

ACKNOWLEDGMENTS

Ronel Pienaar is thanked for assistance with the graphic figures in the study. The reviewers are thanked for constructive criticism and additional ideas that improved the study.

REFERENCES

- Adams, T. E., and Huntington, J. A. (2006). Thrombin–Cofactor interactions. Structural insights into regulatory mechanisms. *Arterioscler. Thromb. Vasc. Biol.* 26, 1738–1745. doi: 10.1161/01.atv.0000228844.65168.d1
- Alenazi, Y., Singh, K., Davies, G., Eaton, J. R. O., Elders, P., Kawamura, A., et al. (2018). Genetically engineered two-warhead evasins provide a method to achieve precision targeting of disease-relevant chemokine subsets. *Sci. Rep.* 8:6333. doi: 10.1038/s41598-018-24568-9
- Anastopoulos, P., Thurn, M. J., and Broady, K. W. (1991). Anticoagulant in the tick *Ixodes holocyclus*. *Aust. Vet. J.* 68, 366–367. doi: 10.1111/j.1751-0813.1991.tb00740.x
- Andersen, J. F., and Ribeiro, J. M. C. (2017). “Salivary kratagonists: scavengers of host physiological effectors during blood feeding,” in *Arthropod Vector: Controller of Disease Transmission*, Vol. 2, eds S. K. Wikel, S. Aksoy, and G. Dimopoulos (Amsterdam: Elsevier), 51–63. doi: 10.1016/b978-0-12-805360-7.00004-6
- Anguita, J., Ramamoorthi, N., Hovius, J. W., Das, S., Thomas, V., Persinski, R., et al. (2002). Salp15, an *Ixodes scapularis* salivary protein, inhibits CD4(+) T cell activation. *Immunity* 16, 849–859. doi: 10.1016/s1074-7613(02)00325-4
- Anisuzzaman, M., Hatta, T., Miyoshi, T., Matsubayashi, M., Islam, M. K., Alim, M. A., et al. (2014). Longistatin in tick saliva blocks advanced glycation end-product receptor activation. *J. Clin. Invest.* 124, 4429–4444. doi: 10.1172/jci74917
- Anisuzzaman, M., Islam, K., Alim, M. A., Miyoshi, T., Hatta, T., Yamaji, K., et al. (2011). Longistatin, a plasminogen activator, is key to the availability of blood-meals for ixodid ticks. *PLoS Pathog.* 7:e1001312. doi: 10.1371/journal.ppat.1001312
- Anisuzzaman, M., Islam, K., Alim, M. A., Miyoshi, T., Hatta, T., Yamaji, K., et al. (2012). Longistatin is an unconventional serine protease and induces protective immunity against tick infestation. *Mol. Biochem. Parasitol.* 182, 45–53. doi: 10.1016/j.molbiopara.2011.12.002
- Anisuzzaman, M., Islam, K., Miyoshi, T., Alim, M. A., Hatta, T., Yamaji, K., et al. (2010). Longistatin, a novel EF-hand protein from the ixodid tick *Haemaphysalis longicornis*, is required for acquisition of host blood-meals. *Int. J. Parasitol.* 40, 721–729. doi: 10.1016/j.ijpara.2009.11.004
- Arcà, B., and Ribeiro, J. M. C. (2018). Saliva of hematophagous insects: a multifaceted toolkit. *Curr. Opin. Insect Sci.* 29, 102–109. doi: 10.1016/j.cois.2018.07.012
- Arolas, J. L., Lorenzo, J., Rovira, A., Castellà, J., Aviles, F. X., and Sommerhoff, C. P. (2005). A carboxypeptidase inhibitor from the tick *Rhipicephalus bursa*: isolation, cDNA cloning, recombinant expression, and characterization. *J. Biol. Chem.* 280, 3441–3448. doi: 10.1074/jbc.m411086200
- Arthur, D. R. (1970). Tick feeding and its implications. *Adv. Parasitol.* 8, 275–292. doi: 10.1016/s0065-308x(08)60258-4
- Assumpção, T. C., Ma, D., Mizurini, D. M., Kini, R. M., Ribeiro, J. M., Kotsyfakis, M., et al. (2016). In vitro mode of action and anti-thrombotic activity of boophilin, a multifunctional Kunitz protease inhibitor from the midgut of a tick vector of babesiosis, *Rhipicephalus microplus*. *PLoS Negl. Trop. Dis.* 10:e0004298. doi: 10.1371/journal.pntd.0004298
- Assumpção, T. C., Mizurini, D. M., Ma, D., Monteiro, R. Q., Ahlstedt, S., Reyes, M., et al. (2018). Ixonnexin from tick saliva promotes fibrinolysis by interacting with plasminogen and tissue-type plasminogen activator, and prevents arterial thrombosis. *Sci. Rep.* 8:4806. doi: 10.1038/s41598-018-22780-1
- Bakkes, D. K., De Klerk, D., Latif, A. A., and Mans, B. J. (2018). Integrative taxonomy of Afrotropical *Ornithodoros* (*Ornithodoros*) (Acari: Ixodida: Argasidae). *Ticks Tick Borne Dis.* 9, 1006–1037. doi: 10.1016/j.ttbdis.2018.03.024
- Bastiani, M., Hillebrand, S., Horn, F., Kist, T. B., Guimarães, J. A., and Termignoni, C. (2002). Cattle tick *Boophilus microplus* salivary gland contains a thiol-activated metalloendopeptidase displaying kininase activity. *Insect Biochem. Mol. Biol.* 32, 1439–1446. doi: 10.1016/s0965-1748(02)00064-4
- Batista, I. F., Ramos, O. H., Ventura, J. S., Junqueira-de-Azevedo, I. L., Ho, P. L., and Chudzinski-Tavassi, A. M. (2010). A new factor Xa inhibitor from *Amblyomma cajennense* with a unique domain composition. *Arch. Biochem. Biophys.* 493, 151–156. doi: 10.1016/j.abb.2009.10.009
- Beaufays, J., Adam, B., Menten-Dedoyart, C., Fievez, L., Grosjean, A., Decrem, Y., et al. (2008). Ir-LBP, an *Ixodes ricinus* tick salivary LTB4-binding lipocalin, interferes with host neutrophil function. *PLoS One* 3:e3987. doi: 10.1371/journal.pone.0003987
- Binnington, K. C., and Kemp, D. H. (1980). Role of tick salivary glands in feeding and disease transmission. *Adv. Parasitol.* 18, 315–339. doi: 10.1016/s0065-308x(08)60403-0
- Bopp, C., Hofer, S., Weitz, J., Bierhaus, A., Nawroth, P. P., Martin, E., et al. (2008). sRAGE is elevated in septic patients and associated with patients outcome. *J. Surg. Res.* 147, 79–83. doi: 10.1016/j.jss.2007.07.014
- Brahma, R. K., Blanchet, G., Kaur, S., Kini, R. M., and Doley, R. (2017). Expression and characterization of haemathrins, madanin-like thrombin inhibitors, isolated from the salivary gland of tick *Haemaphysalis bispinosa* (Acari: Ixodidae). *Thromb. Res.* 152, 20–29. doi: 10.1016/j.thromres.2017.01.012
- Branco, V. G., Iqbal, A., Alvarez-Flores, M. P., Sciani, J. M., de Andrade, S. A., Iwai, L. K., et al. (2016). Amblyomin-X having a Kunitz-type homologous domain, is a noncompetitive inhibitor of FXa and induces anticoagulation in vitro and in vivo. *Biochim. Biophys. Acta* 1864, 1428–1435. doi: 10.1016/j.bbapap.2016.07.011
- Brown, S. J., and Knapp, F. W. (1980a). *Amblyomma americanum*: sequential histological analysis of larval and nymphal feeding sites on guinea pigs. *Exp. Parasitol.* 49, 188–205. doi: 10.1016/0014-4894(80)90116-2
- Brown, S. J., and Knapp, F. W. (1980b). *Amblyomma americanum*: sequential histological analysis of adult feeding sites on guinea pigs. *Exp. Parasitol.* 49, 303–318. doi: 10.1016/0014-4894(80)90067-3
- Brown, S. J., Worms, M. J., and Askenase, P. W. (1983). *Rhipicephalus appendiculatus*: larval feeding sites in guinea pigs actively sensitized and receiving immune serum. *Exp. Parasitol.* 55, 111–120. doi: 10.1016/0014-4894(83)90004-8
- Bullard, R., Allen, P., Chao, C. C., Douglas, J., Das, P., Morgan, S. E., et al. (2016). Structural characterization of tick cement cones collected from in vivo and artificial membrane blood-fed lone star ticks (*Amblyomma americanum*). *Ticks Tick Borne Dis.* 7, 880–892. doi: 10.1016/j.ttbdis.2016.04.006
- Butenas, S., and Mann, K. G. (2002). Blood coagulation. *Biochemistry* 67, 3–12.
- Calvo, E., Mans, B. J., Andersen, J. F., and Ribeiro, J. M. C. (2006). Function and evolution of a mosquito salivary protein family. *J. Biol. Chem.* 281, 1935–1942. doi: 10.1074/jbc.m510359200
- Cao, J., Shi, L., Zhou, Y., Gao, X., Zhang, H., Gong, H., et al. (2013). Characterization of a new Kunitz-type serine protease inhibitor from the hard tick *Rhipicephalus hemaphysaloides*. *Arch. Insect Biochem. Physiol.* 84, 104–113. doi: 10.1002/arch.21118
- Chalaise, K. C., Kim, T. K., Garcia-Rodriguez, H., and Mulenga, A. (2011). *Amblyomma americanum* (L.) (Acari: Ixodidae) tick salivary gland serine protease inhibitor (serpin) 6 is secreted into tick saliva during tick feeding. *J. Exp. Biol.* 214, 665–673. doi: 10.1242/jeb.052076
- Chand, K. K., Lee, K. M., Lavidis, N. A., Rodriguez-Valle, M., Ijaz, H., Koehbach, J., et al. (2016). Tick holocyclotoxins trigger host paralysis by presynaptic inhibition. *Sci. Rep.* 6:29446. doi: 10.1038/srep29446
- Chmelař, J., Kotál, J., Karim, S., Kopecký, P., Francischetti, I. M. B., Pedra, J. H. F., et al. (2016a). Sialomes and mialomes: a systems-biology view of tick tissues and tick-host interactions. *Trends Parasitol.* 32, 242–254. doi: 10.1016/j.pt.2015.10.002
- Chmelař, J., Kotál, J., Kopecký, J., Pedra, J. H., and Kotsyfakis, M. (2016b). All for one and one for all on the tick-host battlefield. *Trends Parasitol.* 32, 368–377. doi: 10.1016/j.pt.2016.01.004
- Chmelař, J., Kotál, J., Langhansová, H., and Kotsyfakis, M. (2017). Protease inhibitors in tick saliva: the role of serpins and cystatins in tick-host-pathogen interaction. *Front. Cell. Infect. Microbiol.* 7:216. doi: 10.3389/fcimb.2017.00216
- Chmelař, J., Oliveira, C. J., Rezacova, P., Francischetti, I. M., Kovarova, Z., Pejler, G., et al. (2011). A tick salivary protein targets cathepsin G and chymase and inhibits host inflammation and platelet aggregation. *Blood* 117, 736–744. doi: 10.1182/blood-2010-06-293241
- Chou, T.-C., and Talalay, P. (1977). A simple generalized equation for the analysis of multiple inhibition of Michaelis-Menten kinetics systems. *J. Biol. Chem.* 252, 6438–6442.
- Ciprandi, A., de Oliveira, S. K., Masuda, A., Horn, F., and Termignoni, C. (2006). *Boophilus microplus*: its saliva contains microphilin, a small thrombin inhibitor. *Exp. Parasitol.* 114, 40–46. doi: 10.1016/j.exppara.2006.02.010

- Coller, B. S., Beer, J. H., Scudder, L. E., and Steinberg, M. H. (1989). Collagen-platelet interactions: evidence for a direct interaction of collagen with platelet GPIa/IIa and an indirect interaction with platelet GPIIb/IIIa mediated by adhesive proteins. *Blood* 74, 182–192.
- Copley, S. D. (2015). An evolutionary biochemist's perspective on promiscuity. *Trends Biochem. Sci.* 40, 72–78. doi: 10.1016/j.tibs.2014.12.004
- Copley, S. D. (2017). Shining a light on enzyme promiscuity. *Curr. Opin. Struct. Biol.* 47, 167–175. doi: 10.1016/j.sbi.2017.11.001
- Cornwall, J. W., and Patton, W. S. (1914). Some observations on the salivary secretion of the commoner blood-sucking insects and ticks. *Indian J. Med. Res.* 2, 569–593.
- Couvreur, B., Beaufays, J., Charon, C., Lahaye, K., Gensale, F., Denis, V., et al. (2008). Variability and action mechanism of a family of anticomplement proteins in *Ixodes ricinus*. *PLoS One* 3:e1400. doi: 10.1371/journal.pone.0001400
- Daix, V., Schroeder, H., Praet, N., Georjgin, J. P., Chiappino, I., Gillet, L., et al. (2007). *Ixodes* ticks belonging to the *Ixodes ricinus* complex encode a family of anticomplement proteins. *Insect Mol. Biol.* 16, 155–166. doi: 10.1111/j.1365-2583.2006.00710.x
- Davis, T. L., and Sharif, N. A. (2000). Pharmacological characterization of [3H]-prostaglandin E2 binding to the cloned human EP4 prostanoid receptor. *Br. J. Pharmacol.* 130, 1919–1926. doi: 10.1038/sj.bjp.0703525
- de Castro, M. H., de Klerk, D., Pienaar, R., Latif, A. A., Rees, D. J., and Mans, B. J. (2016). De novo assembly and annotation of the salivary gland transcriptome of *Rhipicephalus appendiculatus* male and female ticks during blood feeding. *Ticks and Tick-borne Dis.* 7, 536–548. doi: 10.1016/j.ttbdis.2016.01.014
- de Castro, M. H., de Klerk, D., Pienaar, R., Rees, D. J. G. R., and Mans, B. J. (2017). Sialotranscriptomics of *Rhipicephalus zambeziensis* reveal intricate expression profiles of secretory proteins and suggest tight temporal transcriptional regulation during blood feeding. *Parasit. Vectors* 10:384.
- de Meillon, B. (1942). A toxin from the eggs of South African ticks. *S. Afr. J. Med. Sci.* 7, 226–235.
- Decrem, Y., Rath, G., Blasioli, V., Cauchie, P., Robert, S., Beaufays, J., et al. (2009). Ir-CPI, a coagulation contact phase inhibitor from the tick *Ixodes ricinus*, inhibits thrombus formation without impairing hemostasis. *J. Exp. Med.* 206, 2381–2395. doi: 10.1084/jem.20091007
- Delvaeye, M., and Conway, E. M. (2009). Coagulation and innate immune responses: can we view them separately? *Blood* 114, 2367–2374. doi: 10.1182/blood-2009-05-199208
- Deppermann, C., and Kubers, P. (2016). Platelets and infection. *Semin. Immunol.* 28, 536–545. doi: 10.1016/j.smim.2016.10.005
- Déruaz, M., Bonvin, P., Severin, I. C., Johnson, Z., Krohn, S., Power, C. A., et al. (2013). Evasin-4, a tick-derived chemokine-binding protein with broad selectivity can be modified for use in preclinical disease models. *FEBS J.* 280, 4876–4887. doi: 10.1111/febs.12463
- Déruaz, M., Frauenschuh, A., Alessandri, A. L., Dias, J. M., Coelho, F. M., Russo, R. C., et al. (2008). Ticks produce highly selective chemokine binding proteins with antiinflammatory activity. *J. Exp. Med.* 205, 2019–2031. doi: 10.1084/jem.20072689
- Dharampaul, S., Kaufman, W. R., and Belosevic, M. (1993). Differential recognition of saliva antigens from the ixodid tick *Amblyomma hebraeum* (Acari: Ixodidae) by sera from infested and immunized rabbits. *J. Med. Entomol.* 30, 262–266. doi: 10.1093/jmedent/30.1.262
- Díaz-Martín, V., Manzano-Román, R., Oleaga, A., Encinas-Grandes, A., and Pérez-Sánchez, R. (2013a). Cloning and characterization of a plasminogen-binding enolase from the saliva of the argasid tick *Ornithodoros moubata*. *Vet. Parasitol.* 191, 301–314. doi: 10.1016/j.vetpar.2012.09.019
- Díaz-Martín, V., Manzano-Román, R., Valero, L., Oleaga, A., Encinas-Grandes, A., and Pérez-Sánchez, R. (2013b). An insight into the proteome of the saliva of the argasid tick *Ornithodoros moubata* reveals important differences in saliva protein composition between the sexes. *J. Proteomics* 80, 216–235. doi: 10.1016/j.jprot.2013.01.015
- Dickinson, R. G., O'Hagan, J. E., Schotz, M., Binnington, K. C., and Hegarty, M. P. (1976). Prostaglandin in the saliva of the cattle tick *Boophilus microplus*. *Aust. J. Exp. Biol. Med. Sci.* 54, 475–486.
- Dorn, G. W. II, and DeJesus, A. (1991). Human platelet aggregation and shape change are coupled to separate thromboxane A2-prostaglandin H2 receptors. *Am. J. Physiol.* 260, H327–H334.
- Drazen, J. M., Austen, K. F., Lewis, R. A., Clark, D. A., Goto, G., Marfat, A., et al. (1980). Comparative airway and vascular activities of leukotrienes C-1 and D *in vivo* and *in vitro*. *Proc. Natl. Acad. Sci. U.S.A.* 77, 4354–4358.
- Eaton, J. R. O., Alenazi, Y., Singh, K., Davies, G., Geis-Asteggiante, L., Kessler, B., et al. (2018). The N-terminal domain of a tick evasin is critical for chemokine binding and neutralization and confers specific binding activity to other evasins. *J. Biol. Chem.* 293, 6134–6146. doi: 10.1074/jbc.RA117.000487
- Ehebauer, M. T., Mans, B. J., Gaspar, A. R., and Neitz, A. W. (2002). Identification of extrinsic blood coagulation pathway inhibitors from the tick *Ornithodoros savignyi* (Acari: Argasidae). *Exp. Parasitol.* 101, 138–148. doi: 10.1016/s0014-4894(02)00102-9
- Elgin, M. (2010). "Reductionism in biology: an example of biochemistry," in *The Present Situation in the Philosophy of Science. The Philosophy of Science in a European Perspective*, Vol. 1, ed. F. Stadler (Dordrecht: Springer).
- Faas, M. M., Sáez, T., and de Vos, P. (2017). Extracellular ATP and adenosine: the Yin and Yang in immune responses? *Mol. Aspects. Med.* 55, 9–19. doi: 10.1016/j.mam.2017.01.002
- Ferrer-Lopez, P., Renesto, P., Schattner, M., Bassot, S., Laurent, P., and Chignard, M. (1990). Activation of human platelets by C5a-stimulated neutrophils: a role for cathepsin G. *Am. J. Physiol.* 258, C1100–C1107.
- Feyerabend, P. (1975). *Against Method: Outline of an Anarchistic Theory of Knowledge*. Brooklyn, NY: Verso Books, 339.
- Figueiredo, A. C., de Sanctis, D., and Pereira, P. J. (2013). The tick-derived anticoagulant madanin is processed by thrombin and factor Xa. *PLoS One* 8:e71866. doi: 10.1371/journal.pone.0071866
- Flower, D. R. (1996). The lipocalin protein family: structure and function. *Biochem. J.* 318, 1–14. doi: 10.1042/bj3180001
- Francischetti, I. M., Mans, B. J., Meng, Z., Gudderra, N., Veenstra, T. D., Pham, V. M., et al. (2008). An insight into the salivome of the soft tick, *Ornithodoros parkeri*. *Insect Biochem. Mol. Biol.* 38, 1–21. doi: 10.1016/j.ibmb.2007.09.009
- Francischetti, I. M., Mather, T. N., and Ribeiro, J. M. (2003). Cloning of a salivary gland metalloprotease and characterization of gelatinase and fibrin(ogen)lytic activities in the saliva of the Lyme disease tick vector *Ixodes scapularis*. *Biochem. Biophys. Res. Commun.* 305, 869–875. doi: 10.1016/s0006-291x(03)00857-x
- Francischetti, I. M., Mather, T. N., and Ribeiro, J. M. (2004). Penthalaris, a novel recombinant five-Kunitz tissue factor pathway inhibitor (TFPI) from the salivary gland of the tick vector of Lyme disease, *Ixodes scapularis*. *Thromb. Haemost.* 91, 886–898. doi: 10.1160/th03-11-0715
- Francischetti, I. M., Mather, T. N., and Ribeiro, J. M. (2005). Tick saliva is a potent inhibitor of endothelial cell proliferation and angiogenesis. *Thromb. Haemost.* 94, 167–174. doi: 10.1160/th04-09-0566
- Francischetti, I. M., Sa-Nunes, A., Mans, B. J., Santos, I. M., and Ribeiro, J. M. (2009). The role of saliva in tick feeding. *Front. Biosci.* 14:2051–2088.
- Francischetti, I. M., Valenzuela, J. G., Andersen, J. F., Mather, T. N., and Ribeiro, J. M. (2002). Ixolaris, a novel recombinant tissue factor pathway inhibitor (TFPI) from the salivary gland of the tick, *Ixodes scapularis*: identification of factor X and factor Xa as scaffolds for the inhibition of factor VIIa/tissue factor complex. *Blood* 99, 3602–3612. doi: 10.1182/blood-2001-12-0237
- Frauenschuh, A., Power, C. A., Déruaz, M., Ferreira, B. R., Silva, J. S., Teixeira, M. M., et al. (2007). Molecular cloning and characterization of a highly selective chemokine-binding protein from the tick *Rhipicephalus sanguineus*. *J. Biol. Chem.* 282, 27250–27258. doi: 10.1074/jbc.m704706200
- Frojmovic, M. M., Mooney, R. F., and Wong, T. (1994). Dynamics of platelet glycoprotein IIb-IIIa receptor expression and fibrinogen binding. I. Quantal activation of platelet subpopulations varies with adenosine diphosphate concentration. *Biophys. J.* 67, 2060–2068. doi: 10.1016/s0006-3495(94)80689-7
- Garg, R., Juncadella, I. J., Ramamoorthi, N., Ashish Ananthanarayanan, S. K., Thomas, V., Rincón, M., et al. (2006). Cutting edge: CD4 is the receptor for the tick saliva immunosuppressor, Salp15. *J. Immunol.* 177, 6579–6583. doi: 10.4049/jimmunol.177.10.6579
- Gaspar, A. R., Crause, J. C., and Neitz, A. W. (1995). Identification of anticoagulant activities in the salivary glands of the soft tick, *Ornithodoros savignyi*. *Exp. Appl. Acarol.* 19, 117–127. doi: 10.1007/bf00052551
- Gaspar, A. R., Joubert, A. M., Crause, J. C., and Neitz, A. W. (1996). Isolation and characterization of an anticoagulant from the salivary glands of the tick, *Ornithodoros savignyi* (Acari: Argasidae). *Exp. Appl. Acarol.* 20, 583–598. doi: 10.1007/bf00052809

- Ghiglieri-Bertez, C., Cristol, J. P., and Bonne, C. (1986). High-affinity binding site for leukotriene C4 in human erythrocytes. *Biochim. Biophys. Acta* 879, 97–102. doi: 10.1016/0005-2760(86)90271-7
- Giles, T. C., and Emes, R. D. (2017). Inferring function from homology. *Meth. Mol. Biol.* 1526, 23–40. doi: 10.1007/978-1-4939-6613-4_2
- Gong, H., Zhou, J., Liao, M., Hatta, T., Harnnoi, T., Umemiya, R., et al. (2007). Characterization of a carboxypeptidase inhibitor from the tick *Haemaphysalis longicornis*. *J. Insect Physiol.* 53, 1079–1087. doi: 10.1016/j.jinsphys.2007.06.008
- Gregson, J. D. (1973). *Tick Paralysis: An Appraisal of Natural and Experimental Data*. Ottawa, ON: Information Division, Canada Department of Agriculture. doi: 10.1016/j.jinsphys.2007.06.008
- Grunclová, L., Horn, M., Vancová, M., Sojka, D., Franta, Z., Mares, M., et al. (2006). Two secreted cystatins of the soft tick *Ornithodoros moubata*: differential expression pattern and inhibitory specificity. *Biol. Chem.* 387, 1635–1644.
- Hackenberg, M., and Kotsyfakis, M. (2018). Exosome-mediated pathogen transmission by arthropod vectors. *Trends Parasitol.* 34, 549–552. doi: 10.1016/j.pt.2018.04.001
- Hackenberg, M., Langenberger, D., Schwarz, A., Erhart, J., and Kotsyfakis, M. (2017). *In silico* target network analysis of *de novo*-discovered, tick saliva-specific microRNAs reveals important combinatorial effects in their interference with vertebrate host physiology. *RNA* 23, 1259–1269. doi: 10.1261/rna.061168.117
- Hamberg, M., Svensson, J., and Samuelsson, B. (1975). Thromboxanes: a new group of biologically active compounds derived from prostaglandin endoperoxides. *Proc. Natl. Acad. Sci. U.S.A.* 72, 2994–2998. doi: 10.1073/pnas.72.8.2994
- Haycraft, J. B. (1884). Ueber die Einwirkung eines Sekretes des officinellen Blutegels auf die Gerinnbarkeit des Blutes. *Arch. für exp. Pathol. u. Pharmacol.* 18:209. doi: 10.1007/bf01833843
- Hayward, J., Sanchez, J., Perry, A., Huang, C., Rodriguez Valle, M., Canals, M., et al. (2018). Ticks from diverse genera encode chemokine-inhibitory evasin proteins. *J. Biol. Chem.* 292, 15670–15680. doi: 10.1074/jbc.m117.807255
- Heller, C. (1858). Zur anatomie von *Argas persicus*. *Sitzungsberichte d. Kaiserl. Akad. Wien* 30, 297–326.
- Hepburn, N. J., Williams, A. S., Nunn, M. A., Chamberlain-Banoub, J. C., Hamer, J., Morgan, B. P., et al. (2007). In vivo characterization and therapeutic efficacy of a C5-specific inhibitor from the soft tick *Ornithodoros moubata*. *J. Biol. Chem.* 282, 8292–8299. doi: 10.1074/jbc.m609858200
- Hoepli, R., and Feng, L. C. (1933). Experimental studies on ticks. *Chin. Med. J.* 47, 29–43.
- Hoffmann, A., Walsmann, P., Riesener, G., Paintz, M., and Markwardt, F. (1991). Isolation and characterization of a thrombin inhibitor from the tick *Ixodes ricinus*. *Pharmazie* 46, 209–212.
- Horn, F., dos Santos, P. C., and Termignoni, C. (2000). *Boophilus microplus* anticoagulant protein: an antithrombin inhibitor isolated from the cattle tick saliva. *Arch. Biochem. Biophys.* 384, 68–73. doi: 10.1006/abbi.2000.2076
- Hovius, J. W., de Jong, M. A., den Dunnen, J., Litjens, M., Fikrig, E., van der Poll, T., et al. (2008). Salp15 binding to DC-SIGN inhibits cytokine expression by impairing both nucleosome remodeling and mRNA stabilization. *PLoS Pathog.* 4:e31. doi: 10.1371/journal.ppat.0040031
- Howell, C. J. (1966). Collection of salivary gland secretion from the argasid *Ornithodoros savignyi* (Audouin) (1827) by the use of a pharmacological stimulant. *J. S. Afr. Vet. Med. Assoc.* 37, 236–239.
- Howell, C. J., Neitz, A. W. H., and Potgieter, D. J. J. (1975). Some toxic and chemical properties of the oral secretion of the sand tampan, *Ornithodoros savignyi* (Audouin) (1827). *Onderstepoort J. Vet. Res.* 43, 99–102.
- Hu, G., Drescher, K. M., and Chen, X. M. (2012). Exosomal miRNAs: biological properties and therapeutic potential. *Front. Genet.* 3:56. doi: 10.3389/fgene.2012.00056
- Ibelli, A. M., Kim, T. K., Hill, C. C., Lewis, L. A., Bakshi, M., Miller, S., et al. (2014). A blood meal-induced *Ixodes scapularis* tick saliva serpin inhibits trypsin and thrombin, and interferes with platelet aggregation and blood clotting. *Int. J. Parasitol.* 44, 369–379. doi: 10.1016/j.ijpara.2014.01.010
- Ibrahim, M. A., Ghazy, A. H., Maharem, T. M., and Khalil, M. I. (2001a). Factor Xa (FXa) inhibitor from the nymphs of the camel tick *Hyalomma dromedarii*. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* 130, 501–512. doi: 10.1016/s1096-4959(01)00459-6
- Ibrahim, M. A., Ghazy, A. H., Maharem, T., and Khalil, M. (2001b). Isolation and properties of two forms of thrombin inhibitor from the nymphs of the camel tick *Hyalomma dromedarii* (Acari: Ixodidae). *Exp. Appl. Acarol.* 25, 675–698.
- Ibrahim, M. A., and Masoud, H. M. M. (2018). Thrombin inhibitor from the salivary gland of the camel tick *Hyalomma dromedarii*. *Exp. Appl. Acarol.* 74, 85–97. doi: 10.1007/s10493-017-0196-9
- Imamura, S., da Silva Vaz Junior, I., Sugino, M., Ohashi, K., and Onuma, M. (2005). A serine protease inhibitor (serpin) from *Haemaphysalis longicornis* as an anti-tick vaccine. *Vaccine* 23, 1301–1311. doi: 10.1016/j.vaccine.2004.08.041
- Inokuma, H., Kemp, D. H., and Willadsen, P. (1994). Comparison of prostaglandin E2 (PGE2) in salivary gland of *Boophilus microplus*, *Haemaphysalis longicornis* and *Ixodes holocyclus*, and quantification of PGE2 in saliva, hemolymph, ovary and gut of *B. microplus*. *J. Vet. Med. Sci.* 56, 1217–1218. doi: 10.1292/jvms.56.1217
- Iqbal, A., Goldfeder, M. B., Marques-Porto, R., Asif, H., Souza, J. G., Faria, F., et al. (2017). Revisiting antithrombotic therapeutics; sculptin, a novel specific, competitive, reversible, scissile and tight binding inhibitor of thrombin. *Sci. Rep.* 7:1431. doi: 10.1038/s41598-017-01486-w
- Iwanaga, S., Isawa, H., and Yuda, M. (2014). Horizontal gene transfer of a vertebrate vasodilatory hormone into ticks. *Nature Comm.* 5:3373. doi: 10.1038/ncomms4373
- Iwanaga, S., Okada, M., Isawa, H., Morita, A., Yuda, M., and Chinzei, Y. (2003). Identification and characterization of novel salivary thrombin inhibitors from the ixodidae tick, *Haemaphysalis longicornis*. *Eur. J. Biochem.* 270, 1926–1934. doi: 10.1046/j.1432-1033.2003.03560.x
- Iyer, J. K., Koh, C. Y., Kazimirova, M., Roller, L., Jobichen, C., Swaminathan, K., et al. (2017). Avathrin: a novel thrombin inhibitor derived from a multicopy precursor in the salivary glands of the ixodid tick, *Amblyomma variegatum*. *FASEB J.* 31, 2981–2995. doi: 10.1096/fj.201601216R
- Jablonka, W., Kotsyfakis, M., Mizurini, D. M., Monteiro, R. Q., Lukszo, J., Drake, S. K., et al. (2015). Identification and mechanistic analysis of a novel tick-derived inhibitor of Thrombin. *PLoS One* 10:e0133991. doi: 10.1371/journal.pone.0133991
- Jore, M. M., Johnson, S., Sheppard, D., Barber, N. M., Li, Y. I., Nunn, M. A., et al. (2016). Structural basis for therapeutic inhibition of complement C5. *Nat. Struct. Mol. Biol.* 23, 378–386. doi: 10.1038/nsmb.3196
- Jung, S. M., and Moroi, M. (1998). Platelets interact with soluble and insoluble collagens through characteristically different reactions. *J. Biol. Chem.* 273, 14827–14837. doi: 10.1074/jbc.273.24.14827
- Karczewski, J., Endris, R., and Connolly, T. M. (1994). Disagregin is a fibrinogen receptor antagonist lacking the Arg-Gly-Asp sequence from the tick, *Ornithodoros moubata*. *J. Biol. Chem.* 269, 6702–6708.
- Karczewski, J., Waxman, L., Endris, R. G., and Connolly, T. M. (1995). An inhibitor from the argasid tick *Ornithodoros moubata* of cell adhesion to collagen. *Biochem. Biophys. Res. Commun.* 208, 532–541. doi: 10.1006/bbr.1995.1371
- Kato, N., Iwanaga, S., Okayama, T., Isawa, H., Yuda, M., and Chinzei, Y. (2005). Identification and characterization of the plasma kallikrein-kinin system inhibitor, haemaphysalin, from hard tick, *Haemaphysalis longicornis*. *Thromb. Haemost.* 93, 359–367. doi: 10.1160/th04-05-0319
- Kazimirová, M., Jancinová, V., Petriková, M., Takác, P., Labuda, M., and Nosál, R. (2002). An inhibitor of thrombin-stimulated blood platelet aggregation from the salivary glands of the hard tick *Amblyomma variegatum* (Acari: Ixodidae). *Exp. Appl. Acarol.* 28, 97–105. doi: 10.1007/978-94-017-3526-1_7
- Keller, P. M., Waxman, L., Arnold, B. A., Schultz, L. D., Condra, C., and Connolly, T. M. (1993). Cloning of the cDNA and expression of moubatin, an inhibitor of platelet aggregation. *J. Biol. Chem.* 268, 5450–5456.
- Kemp, D. H., Stone, B. F., and Binnington, K. C. (1982). “Tick attachment and feeding: role of the mouthparts, feeding apparatus, salivary gland secretions and the host response,” in *Physiology of Ticks*, eds F. D. Obenchain and R. Galun (Oxford: Pergamon Press).
- Kim, T. K., Tirloni, L., Pinto, A. F., Moresco, J., Yates, J. R. III, da Silva Vaz, I. Jr., et al. (2017). *Ixodes scapularis* tick saliva proteins sequentially secreted every 24 h during blood feeding. *PLoS Negl. Trop. Dis.* 10:e0004323. doi: 10.1371/journal.pntd.0004323
- Kim, T. K., Tirloni, L., Radulovic, Z., Lewis, L., Bakshi, M., Hill, C., et al. (2015). Conserved *Amblyomma americanum* tick serpin19, an inhibitor of blood clotting factors Xa and XIa, trypsin and plasmin, has anti-haemostatic functions. *Int. J. Parasitol.* 45, 613–627. doi: 10.1016/j.ijpara.2015.03.009

- Kiszewski, A. E., Matuschka, F. R., and Spielman, A. (2001). Mating strategies and spermiogenesis in ixodid ticks. *Annu. Rev. Entomol.* 46, 167–182.
- Koh, C. Y., Kazimirova, M., Trimnell, A., Takac, P., Labuda, M., Nuttall, P. A., et al. (2007). Variegatin, a novel fast and tight binding thrombin inhibitor from the tropical bont tick. *J. Biol. Chem.* 282, 29101–29113. doi: 10.1074/jbc.m705600200
- Kotál, J., Stergiou, N., Buša, M., Chlastáková, A., Beránková, Z., Řezáčová, P., et al. (2019). The structure and function of Iristatin, a novel immunosuppressive tick salivary cystatin. *Cell Mol. Life Sci.*
- Kotsyfakis, M., Karim, S., Andersen, J. F., Mather, T. N., and Ribeiro, J. M. (2007). Selective cysteine protease inhibition contributes to blood-feeding success of the tick *Ixodes scapularis*. *J. Biol. Chem.* 282, 29256–29263. doi: 10.1074/jbc.m703143200
- Kotsyfakis, M., Sá-Nunes, A., Francischetti, I. M., Mather, T. N., Andersen, J. F., and Ribeiro, J. M. (2006). Antiinflammatory and immunosuppressive activity of sialostatin L, a salivary cystatin from the tick *Ixodes scapularis*. *J. Biol. Chem.* 281, 26298–26307. doi: 10.1074/jbc.m513010200
- Krilis, S., Lewis, R. A., Corey, E. J., and Austen, K. F. (1983). Specific receptors for leukotriene C4 on a smooth muscle cell line. *J. Clin. Invest.* 72, 1516–1519. doi: 10.1172/jci111109
- Kröber, T., and Geurin, P. M. (2007). In vitro feeding assays for hard ticks. *Trends Parasitol.* 23, 445–449. doi: 10.1016/j.pt.2007.07.010
- Kuhn, N., Schmidt, C. Q., Schlapsch, M., and Skerra, A. (2016). PASylated Coversin, a C5-specific complement inhibitor with extended pharmacokinetics, shows enhanced anti-hemolytic activity *in vitro*. *Bioconjug. Chem.* 27, 2359–2371. doi: 10.1021/acs.bioconjchem.6b00369
- Kuriyan, J., Konforti, B., and Wemmer, D. (2013). *The Molecules of Life: Physical and Chemical Principles*. Routledge: Garland Science, Taylor & Francis Group, 1008.
- Law, J. H., Ribeiro, J. M. C., and Wells, M. A. (1992). Biochemical insights derived from insect diversity. *Ann. Rev. Biochem.* 64, 87–111. doi: 10.1146/annurev.biochem.61.1.87
- Leboulle, G., Crippa, M., Decrem, Y., Mejri, N., Brossard, M., Bollen, A., et al. (2002). Characterization of a novel salivary immunosuppressive protein from *Ixodes ricinus* ticks. *J. Biol. Chem.* 277, 10083–10089.
- Lewis, R. A., Mencia-Huerta, J. M., Soberman, R. J., Hoover, D., Marfat, A., Corey, E. J., et al. (1982). Radioimmunoassay for leukotriene B4. *Proc. Natl. Acad. Sci. U.S.A.* 79, 7904–7908.
- Leydig, F. (1855). Zum feineren Bau der Arthropoden. *Müller's Archiv. f. Anat. Physiol. u. wiss. Med.* 1855, 376–480.
- Limo, M. K., Voigt, W. P., Tumbo-Oeri, A. G., Njogu, R. M., and ole-MoiYoi, O. K. (1991). Purification and characterization of an anticoagulant from the salivary glands of the ixodid tick *Rhipicephalus appendiculatus*. *Exp. Parasitol.* 72, 418–429. doi: 10.1016/0014-4894(91)90088-e
- Liu, Y. H., Xu, C. H., Liu, Z. G., Liang, J. G., and Lai, R. (2005). Isolation and purification of an inhibitor on platelet aggregation from *Ixodes sinensis*. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 23, 424–427.
- Macedo-Ribeiro, S., Almeida, C., Calisto, B. M., Friedrich, T., Mentele, R., Stürzebecher, J., et al. (2008). Isolation, cloning and structural characterisation of boophilin, a multifunctional Kunitz-type proteinase inhibitor from the cattle tick. *PLoS One* 3:e1624. doi: 10.1371/journal.pone.0001624
- MacGlashan, D. Jr. (2003). Histamine: a mediator of inflammation. *J. Allergy Clin. Immunol.* 112, S53–S59.
- MacGregor, I. R., and Prowse, C. V. (1983). Tissue plasminogen activator in human plasma measured by radioimmunoassay. *Thromb. Res.* 31, 461–474. doi: 10.1016/0049-3848(83)90410-3
- Macpherson, A., Liu, X., Dedi, N., Kennedy, J., Carrington, B., Durrant, O., et al. (2018). The rational design of affinity-attenuated OmCI for the purification of complement C5. *J. Biol. Chem.* 293, 14112–14121. doi: 10.1074/jbc.RA118.004043
- Mans, B. J. (1997). *Biochemical Properties of a Platelet Aggregation Inhibitor of the tick, Ornithodoros savignyi*. . doi: 10.1074/jbc.ra118.004043
- Mans, B. J. (2005). Tick histamine-binding proteins and related lipocalins: potential as therapeutic agents. *Curr. Opin. Investig. Drugs* 6, 1131–1135.
- Mans, B. J. (2011). Evolution of vertebrate hemostatic and inflammatory control mechanisms in blood-feeding arthropods. *J. Innate Immun.* 3, 41–51. doi: 10.1159/000321599
- Mans, B. J. (2016). “Glandular matrices and secretions: blood-feeding arthropods,” in *Extracellular Composite Matrices in Arthropods*, eds E. Cohen and B. Moussian (Switzerland: Springer), 625–688. doi: 10.1007/978-3-319-40740-1_17
- Mans, B. J., Andersen, J. F., Schwan, T. G., and Ribeiro, J. M. (2008a). Characterization of anti-hemostatic factors in the argasid, *Argas monolakensis*: implications for the evolution of blood-feeding in the soft tick family. *Insect Biochem. Mol. Biol.* 38, 22–41. doi: 10.1016/j.ibmb.2007.09.002
- Mans, B. J., Ribeiro, J. M., and Andersen, J. F. (2008b). Structure, function, and evolution of biogenic amine-binding proteins in soft ticks. *J. Biol. Chem.* 283, 18721–18733. doi: 10.1074/jbc.M800188200
- Mans, B. J., Andersen, J. F., Francischetti, I. M., Valenzuela, J. G., Schwan, T. G., Pham, V. M., et al. (2008c). Comparative sialomics between hard and soft ticks: implications for the evolution of blood-feeding behavior. *Insect Biochem. Mol. Biol.* 38, 42–58. doi: 10.1016/j.ibmb.2007.09.003
- Mans, B. J., Coetzee, J., Louw, A. I., Gaspar, A. R., and Neitz, A. W. (2000). Disaggregation of aggregated platelets by apyrase from the tick, *Ornithodoros savignyi* (Acari: Argasidae). *Exp. Appl. Acarol.* 24, 271–282.
- Mans, B. J., de Castro, M. H., Pienaar, R., de Klerk, D., Gaven, P., Genu, S., et al. (2016). Ancestral reconstruction of tick lineages. *Ticks Tick Borne Dis.* 7, 509–535. doi: 10.1016/j.ttbdis.2016.02.002
- Mans, B. J., Featherston, J., de Castro, M. H., and Pienaar, R. (2017). Gene duplication and protein evolution in tick-host interactions. *Front. Cell Infect. Microbiol.* 7:413. doi: 10.3389/fcimb.2017.00413
- Mans, B. J., Gaspar, A. R., Louw, A. I., and Neitz, A. W. (1998a). Apyrase activity and platelet aggregation inhibitors in the tick *Ornithodoros savignyi* (Acari: Argasidae). *Exp. Appl. Acarol.* 22, 353–366.
- Mans, B. J., Gaspar, A. R., Louw, A. I., and Neitz, A. W. (1998b). Purification and characterization of apyrase from the tick, *Ornithodoros savignyi*. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* 120, 617–624. doi: 10.1016/s0305-0491(98)10061-5
- Mans, B. J., Venter, J. D., Vrey, P. J., Louw, A. I., and Neitz, A. W. (2001). Identification of putative proteins involved in granule biogenesis of tick salivary glands. *Electrophoresis* 22, 1739–1746. doi: 10.1002/1522-2683(200105)22%3A9%3C1739%3A%3Aaid-elps1739%3E3.0.co%3B2-7
- Mans, B. J., Louw, A. I., and Neitz, A. W. (2002a). Disaggregation of aggregated platelets by savignigrin, a α IIb β 3 antagonist from *Ornithodoros savignyi*. *Exp. Appl. Acarol.* 27, 231–239.
- Mans, B. J., Louw, A. I., and Neitz, A. W. (2002b). Savignigrin, a platelet aggregation inhibitor from the soft tick *Ornithodoros savignyi*, presents the RGD integrin recognition motif on the Kunitz-BPTI fold. *J. Biol. Chem.* 277, 21371–22378.
- Mans, B. J., Louw, A. I., and Neitz, A. W. (2003a). The influence of tick behavior, biotope and host specificity on concerted evolution of the platelet aggregation inhibitor savignigrin, from the soft tick *Ornithodoros savignyi*. *Insect Biochem. Mol. Biol.* 33, 623–629. doi: 10.1016/s0965-1748(03)00047-x
- Mans, B. J., Louw, A. I., and Neitz, A. W. (2003b). The major tick salivary gland proteins and toxins from the soft tick, *Ornithodoros savignyi*, are part of the tick Lipocalin family: implications for the origins of tick toxicoses. *Mol. Biol. Evol.* 20, 1158–1167. doi: 10.1093/molbev/msg126
- Mans, B. J., and Neitz, A. W. (2004a). Adaptation of ticks to a blood-feeding environment: evolution from a functional perspective. *Insect Biochem. Mol. Biol.* 34, 1–17. doi: 10.1016/j.ibmb.2003.09.002
- Mans, B. J., and Neitz, A. W. (2004b). Molecular crowding as a mechanism for tick secretory granule biogenesis. *Insect Biochem. Mol. Biol.* 34, 1187–1193. doi: 10.1016/j.ibmb.2004.07.007
- Mans, B. J., and Neitz, A. W. (2004c). The mechanism of α IIb β 3 antagonism by savignigrin and its implications for the evolution of anti-hemostatic strategies in soft ticks. *Insect Biochem. Mol. Biol.* 34, 573–584. doi: 10.1016/s0965-1748(04)00038-4
- Mans, B. J., Gothe, R., and Neitz, A. W. (2004a). Biochemical perspectives on paralysis and other forms of toxicoses caused by ticks. *Parasitology* 129, S95–S111.
- Mans, B. J., Venter, J. D., Coons, L. B., Louw, A. I., and Neitz, A. W. H. (2004b). A reassessment of argasid tick salivary gland ultrastructure from an immunocytochemical perspective. *Exp. Appl. Acarol.* 33, 119–129. doi: 10.1023/b%3Aaappa.0000030012.47964.b3

- Mans, B. J., and Ribeiro, J. M. (2008a). A novel clade of cysteinyl leukotriene scavengers in soft ticks. *Insect Biochem. Mol. Biol.* 38, 862–870. doi: 10.1016/j.ibmb.2008.06.002
- Mans, B. J., and Ribeiro, J. M. (2008b). Function, mechanism and evolution of the moubatin-clade of soft tick lipocalins. *Insect Biochem. Mol. Biol.* 38, 841–852. doi: 10.1016/j.ibmb.2008.06.007
- McSwain, J. L., Essenberg, R. C., and Sauer, J. R. (1982). Protein changes in the salivary glands of the female lone star tick, *Amblyomma americanum*, during feeding. *J. Parasitol.* 68, 100–106.
- Monteiro, R. Q., Rezaie, A. R., Ribeiro, J. M., and Francischetti, I. M. (2005). Ixolaris: a factor Xa heparin-binding exosite inhibitor. *Biochem. J.* 387, 871–877. doi: 10.1042/bj20041738
- Moore, J. E. Jr., Brook, B. S., and Nibbs, R. J. B. (2018). Chemokine transport dynamics and emerging recognition of their role in immune function. *Curr. Opin. Biomed. Eng.* 5, 90–95. doi: 10.1016/j.cobme.2018.03.001
- Motoyashiki, T., Tu, A. T., Azimov, D. A., and Ibragim, K. (2003). Isolation of anticoagulant from the venom of tick, *Boophilus calcaratus*, from Uzbekistan. *Thromb. Res.* 110, 235–241. doi: 10.1016/s0049-3848(03)00409-2
- Mulenga, A., Kim, T., and Ibelli, A. M. (2013). *Amblyomma americanum* tick saliva serine protease inhibitor 6 is a cross-class inhibitor of serine proteases and papain-like cysteine proteases that delays plasma clotting and inhibits platelet aggregation. *Insect Mol. Biol.* 22, 306–319. doi: 10.1111/imb.12024
- Mulenga, A., Macaluso, K. R., Simser, J. A., and Azad, A. F. (2003). The American dog tick, *Dermacentor variabilis*, encodes a functional histamine release factor homolog. *Insect Biochem. Mol. Biol.* 33, 911–919. doi: 10.1016/s0965-1748(03)00097-3
- Nakajima, C., Imamura, S., Konnai, S., Yamada, S., Nishikado, H., Ohashi, K., et al. (2006). A novel gene encoding a thrombin inhibitory protein in a cDNA library from *Haemaphysalis longicornis* salivary gland. *J. Vet. Med. Sci.* 68, 447–452. doi: 10.1292/jvms.68.447
- Narasimhan, S., Koski, R. A., Beaulieu, B., Anderson, J. F., Ramamoorthi, N., Kantor, F., et al. (2002). A novel family of anticoagulants from the saliva of *Ixodes scapularis*. *Insect Mol. Biol.* 11, 641–650. doi: 10.1046/j.1365-2583.2002.00375.x
- Neelakanta, G., Sultana, H., Sonenshine, D. E., and Andersen, J. F. (2018). Identification and characterization of a histamine-binding lipocalin-like molecule from the relapsing fever tick *Ornithodoros turicata*. *Insect Mol. Biol.* 27, 177–187. doi: 10.1111/imb.12362
- Neitz, A. W., Howell, C. J., Potgieter, D. J., and Bezuidenhout, J. D. (1978). Proteins and free amino acids in the salivary secretion and haemolymph of the tick *Amblyomma hebraeum*. *Onderstepoort J. Vet. Res.* 45, 235–240.
- Neitz, A. W., Prozesky, L., Bezuidenhout, J. D., Putterill, J. F., and Potgieter, D. J. (1981). An investigation into the toxic principle in eggs of the tick *Amblyomma hebraeum*. *Onderstepoort J. Vet. Res.* 48, 109–117.
- Neitz, A. W. H., Howell, C. J., and Potgieter, D. J. J. (1969). Purification of the toxic component in the oral secretion of the sand tamarin *Ornithodoros savignyi* (Audouin) (1827). *J. S. Afr. Chem. Ind.* 22, 142–149.
- Nielsen, H. (2017). Predicting secretory proteins with SignalP. *Methods Mol. Biol.* 1611, 59–73. doi: 10.1007/978-1-4939-7015-5_6
- Nienaber, J., Gaspar, A. R., and Neitz, A. W. (1999). Savignin, a potent thrombin inhibitor isolated from the salivary glands of the tick *Ornithodoros savignyi* (Acari: Argasidae). *Exp. Parasitol.* 93, 82–91. doi: 10.1006/expr.1999.4448
- Nunn, M. A., Sharma, A., Paesen, G. C., Adamson, S., Lissina, O., Willis, A. C., et al. (2005). Complement inhibitor of C5 activation from the soft tick *Ornithodoros moubata*. *J. Immunol.* 174, 2084–2091. doi: 10.4049/jimmunol.174.4.2084
- Nuttall, G. H. F., and Strickland, C. (1908). On the presence of an anticoagulin in the salivary glands and intestines of *Argas persicus*. *Parasitology* 1, 302–310. doi: 10.1017/s0031182000003590
- Nuttall, G. H. F., Warburton, C., Cooper, W. F., and Robinson, L. E. (1908). *Ticks. A Monograph of the Ixodidae. Part I: The Argasidae*. Cambridge: Cambridge University Press. doi: 10.1017/s0031182000003590
- Nuttall, P. A. (2019). Wonders of tick saliva. *Ticks Tick Borne Dis.* 10, 470–481. doi: 10.1016/j.ttbdis.2018.11.005
- Nuttall, P. A., Trimmell, A. R., Kazimirova, M., and Labuda, M. (2006). Exposed and concealed antigens as vaccine targets for controlling ticks and tick-borne diseases. *Parasite Immunol.* 28, 155–163. doi: 10.1111/j.1365-3024.2006.00806.x
- Oleaga-Pérez, A., Pérez-Sánchez, R., Astigarraga, A., and Encinas-Grandes, A. (1994). Detection of pig farms with *Ornithodoros erraticus* by pig serology. Elimination of non-specific reactions by carbohydrate epitopes of salivary antigens. *Vet. Parasitol.* 52, 97–111. doi: 10.1016/0304-4017(94)90040-x
- Oliveira, C. J., Anatriello, E., de Miranda-Santos, I. K., Francischetti, I. M., Sá-Nunes, A., Ferreira, B. R., et al. (2013). Proteome of *Rhipicephalus sanguineus* tick saliva induced by the secretagogues pilocarpine and dopamine. *Ticks Tick Borne Dis.* 4, 469–477. doi: 10.1016/j.ttbdis.2013.05.001
- Packham, M. A., and Rand, M. L. (2011). Historical perspective on ADP-induced platelet activation. *Purinergic. Signal.* 7, 283–292. doi: 10.1007/s11302-011-9227-x
- Paesen, G. C., Adams, P. L., Harlos, K., Nuttall, P. A., and Stuart, D. I. (1999). Tick histamine-binding proteins: isolation, cloning, and three-dimensional structure. *Mol. Cell* 3, 661–671. doi: 10.1016/s1097-2765(00)80359-7
- Paesen, G. C., Adams, P. L., Nuttall, P. A., and Stuart, D. I. (2000). Tick histamine-binding proteins: lipocalins with a second binding cavity. *Biochim. Biophys. Acta* 1482, 92–101. doi: 10.1016/s0167-4838(00)00168-0
- Paesen, G. C., Siebold, C., Dallas, M. L., Peers, C., Harlos, C., Nuttall, P. A., et al. (2009). An ion-channel modulator from the saliva of the brown ear tick has a highly modified Kunitz/BPTI structure. *J. Mol. Biol.* 389, 734–747. doi: 10.1016/j.jmb.2009.04.045
- Paesen, G. C., Siebold, C., Harlos, K., Peacey, M. F., Nuttall, P. A., and Stuart, D. I. (2007). A tick protein with a modified Kunitz fold inhibits human trypsin. *J. Mol. Biol.* 368, 1172–1186. doi: 10.1016/j.jmb.2007.03.011
- Parizi, L. F., Githaka, N. W., Acevedo, C., Benavides, U., Seixas, A., Logullo, C., et al. (2013). Sequence characterization and immunogenicity of cystatins from the cattle tick *Rhipicephalus (Boophilus) microplus*. *Ticks Tick Borne Dis.* 4, 492–499. doi: 10.1016/j.ttbdis.2013.06.005
- Perner, J., Kropáčková, S., Kopáček, P., and Ribeiro, J. M. C. (2018). Sialome diversity of ticks revealed by RNAseq of single tick salivary glands. *PLoS Negl. Trop. Dis.* 12:e0006410. doi: 10.1371/journal.pntd.0006410
- Pervaiz, S., and Brew, K. (1987). Homology and structure-function correlations between alpha 1-acid glycoprotein and serum retinol-binding protein and its relatives. *FASEB J.* 1, 209–214. doi: 10.1096/phaseb.1.3.3622999
- Petersen, L. J. (1997). Quantitative measurement of extracellular histamine concentrations in intact human skin in vivo by the microdialysis technique: methodological aspects. *Allergy* 52, 547–555. doi: 10.1111/j.1398-9995.1997.tb02598.x
- Pichu, S., Ribeiro, J. M., Mather, T. N., and Francischetti, I. M. (2014). Purification of a serine protease and evidence for a protein C activator from the saliva of the tick, *Ixodes scapularis*. *Toxicon* 77, 32–39. doi: 10.1016/j.toxicon.2013.10.025
- Pienaar, R., Neitz, A. W. H., and Mans, B. J. (2018). Tick paralysis: solving an Enigma. *Vet. Sci.* 5:53. doi: 10.3390/vetsci5020053
- Platt, E. J., Wehrly, K., Kuhmann, S. E., Chesebro, B., and Kabat, D. (1998). Effects of CCR5 and CD4 cell surface concentrations on infections by macrophagetropic isolates of human immunodeficiency virus type 1. *J. Virol.* 72, 2855–2864.
- Preston, S. G., Majtán, J., Kouremenou, C., Rysnik, O., Burger, L. F., Cabezas Cruz, A., et al. (2013). Novel immunomodulators from hard ticks selectively reprogramme human dendritic cell responses. *PLoS Pathog.* 9:e1003450. doi: 10.1371/journal.ppat.1003450
- Prevot, P. P., Adam, B., Boudjeltia, K. Z., Brossard, M., Lins, L., Cauchie, P., et al. (2006). Anti-hemostatic effects of a serpin from the saliva of the tick *Ixodes ricinus*. *J. Biol. Chem.* 281, 26361–26369. doi: 10.1074/jbc.m601497200
- Prevot, P. P., Beschin, A., Lins, L., Beaufays, J., Grosjean, A., Bruys, L., et al. (2009). Exosites mediate the anti-inflammatory effects of a multifunctional serpin from the saliva of the tick *Ixodes ricinus*. *FEBS J.* 276, 3235–3246. doi: 10.1111/j.1742-4658.2009.07038.x
- Prié, S., Guillemette, G., Boulay, G., Borgeat, P., and Sirois, P. (1995). Leukotriene C4 receptors on guinea pig tracheocytes. *J. Pharmacol. Exp. Ther.* 275, 312–318.
- Radulović, Ž.M., and Mulenga, A. (2017). Heparan sulfate/heparin glycosaminoglycan binding alters inhibitory profile and enhances anticoagulant function of conserved *Amblyomma americanum* tick saliva serpin 19. *Insect Biochem. Mol. Biol.* 80, 1–10. doi: 10.1016/j.ibmb.2016.11.002
- Rangel, C. K., Parizi, L. F., Sabadin, G. A., Costa, E. P., Romeiro, N. C., Isezaki, M., et al. (2017). Molecular and structural characterization of novel cystatins

- from the taiga tick *Ixodes persulcatus*. *Ticks Tick Borne Dis.* 8, 432–441. doi: 10.1016/j.ttbdis.2017.01.007
- Rawal, N., and Pangburn, M. (2001). Formation of high-affinity C5 convertases of the alternative pathway of complement. *J. Immunol.* 166, 2635–2642. doi: 10.4049/jimmunol.166.4.2635
- Rawal, N., and Pangburn, M. K. (2003). Formation of high affinity C5 convertase of the classical pathway of complement. *J. Biol. Chem.* 278, 38476–38483. doi: 10.1074/jbc.m307017200
- Ribeiro, J. M. (1987). *Ixodes dammini*: salivary anti-complement activity. *Exp. Parasitol.* 64, 347–353. doi: 10.1016/0014-4894(87)90046-4
- Ribeiro, J. M., and Arcá, B. (2009). From sialomes to the sialoverse: an insight into salivary portion of blood-feeding insects. *Adv. In. Insect Physiol.* 37, 59–118. doi: 10.1371/journal.pone.0044612
- Ribeiro, J. M., Evans, P. M., MacSwain, J. L., and Sauer, J. (1992). *Amblyomma americanum*: characterization of salivary prostaglandins E2 and F2 alpha by RP-HPLC/bioassay and gas chromatography-mass spectrometry. *Exp. Parasitol.* 74, 112–116. doi: 10.1016/0014-4894(92)90145-z
- Ribeiro, J. M., and Mather, T. N. (1998). *Ixodes scapularis*: salivary kininase activity is a metallo dipeptidyl carboxypeptidase. *Exp. Parasitol.* 89, 213–221. doi: 10.1006/expr.1998.4296
- Ribeiro, J. M. C. (1995). How ticks make a living. *Parasitol. Today* 11, 91–93. doi: 10.1016/0169-4758(95)80162-6
- Ribeiro, J. M. C., Endris, T. M., and Endris, R. (1991). Saliva of the soft tick *Ornithodoros moubata*, contains anti-platelet and apyrase activity. *Comp. Biochem. Physiol.* 100, 109–112. doi: 10.1016/0300-9629(91)90190-n
- Ribeiro, J. M. C., Makoul, G., Levine, J., Robinson, D., and Spielman, A. (1985). Antihemostatic, antiinflammatory, and immunosuppressive properties of the saliva of a tick *Ixodes dammini*. *J. Exp. Med.* 161, 332–344. doi: 10.1084/jem.161.2.332
- Ribeiro, J. M. C., and Spielman, A. (1986). *Ixodes dammini*: salivary anaphylatoxin inactivating activity. *Exp. Parasitol.* 62, 292–297. doi: 10.1016/0014-4894(86)90034-2
- Riek, R. F. (1957). Studies on the reactions of animals to infestation with ticks. *Aust. J. Agric. Res.* 8, 215–223.
- Riek, R. F. (1959). Studies on the reactions of animals to infestation with ticks. *Aust. J. Agric. Res.* 10, 604–613.
- Rodriguez-Valle, M., Moolhuijzen, P., Barrero, R. A., Ong, C. T., Busch, G., Karbanowicz, T., et al. (2017). Transcriptome and toxin family analysis of the paralysis tick, *Ixodes holocyclus*. *Int. J. Parasitol.* 48, 71–82. doi: 10.1016/j.ijpara.2017.07.007
- Rodriguez-Valle, M., Xu, T., Kurscheid, S., and Lew-Tabor, A. E. (2015). *Rhipicephalus microplus* serine protease inhibitor family: annotation, expression and functional characterisation assessment. *Parasit. Vectors* 8:7. doi: 10.1186/s13071-014-0605-4
- Ross, I. C. (1926). An experimental study of tick paralysis in Australia. *Parasitology* 18, 410–429. doi: 10.1186/s13071-018-3061-8
- Roversi, P., Johnson, S., Preston, S. G., Nunn, M. A., Paesen, G. C., Austyn, J. M., et al. (2017). Structural basis of cholesterol binding by a novel clade of dendritic cell modulators from ticks. *Sci. Rep.* 7:16057. doi: 10.1038/s41598-017-16413-2
- Roversi, P., Lissina, O., Johnson, S., Ahmat, N., Paesen, G. C., Ploss, K., et al. (2007). The structure of OMCI, a novel lipocalin inhibitor of the complement system. *J. Mol. Biol.* 369, 784–793. doi: 10.1016/j.jmb.2007.03.064
- Roversi, P., Ryffel, B., Togbe, D., Maillet, I., Teixeira, M., Ahmat, N., et al. (2013). Bifunctional lipocalin ameliorates murine immune complex-induced acute lung injury. *J. Biol. Chem.* 288, 18789–18802. doi: 10.1074/jbc.M112.420331
- Sabbatani, L. (1899). Fermenito anticoagulante del *Ixodes ricinus*. *Arch. Ital. Biol.* 31, 37–53.
- Sadik, C. D., Miyabe, Y., Sezin, T., and Luster, A. D. (2018). The critical role of C5a as an initiator of neutrophil-mediated autoimmune inflammation of the joint and skin. *Semin. Immunol.* 37, 21–29. doi: 10.1016/j.smim.2018.03.002
- Salát, J., Paesen, G. C., Rezáčová, P., Kotsyfakis, M., Kovárová, Z., Sanda, M., et al. (2010). Crystal structure and functional characterization of an immunomodulatory salivary cystatin from the soft tick *Ornithodoros moubata*. *Biochem. J.* 429, 103–112. doi: 10.1042/BJ20100280
- Sangamnatdej, S., Paesen, G. C., Slovak, M., and Nuttall, P. A. (2002). A high affinity serotonin- and histamine-binding lipocalin from tick saliva. *Insect Mol. Biol.* 11, 79–86. doi: 10.1046/j.0962-1075.2001.00311.x
- Sant'Anna Azzolini, S., Sasaki, S. D., Torquato, R. J., Andreotti, R., Andreotti, E., and Tanaka, A. S. (2003). *Rhipicephalus sanguineus* trypsin inhibitors present in the tick larvae: isolation, characterization, and partial primary structure determination. *Arch. Biochem. Biophys.* 417, 176–182. doi: 10.1016/s0003-9861(03)00344-8
- Sasagawa, M., Satoh, T., Takemoto, A., Hasegawa, T., Suzuki, E., and Arakawa, M. (1994). Blood levels of leukotrienes (LTC4, D4, E4, B4) in asthmatic patients during attack and remission. *Arerugi* 43, 28–36.
- Schroeder, H., Daix, V., Gillet, L., Renaud, J. C., and Vanderplasschen, A. (2007). The paralogous salivary anti-complement proteins IRAC I and IRAC II encoded by *Ixodes ricinus* ticks have broad and complementary inhibitory activities against the complement of different host species. *Microbes Infect.* 9, 247–250. doi: 10.1016/j.micinf.2006.10.020
- Schuijt, T. J., Bakhtiari, K., Daffre, S., Deponte, K., Wielders, S. J., Marquart, J. A., et al. (2013). Factor Xa activation of factor V is of paramount importance in initiating the coagulation system: lessons from a tick salivary protein. *Circulation* 128, 254–266. doi: 10.1161/CIRCULATIONAHA.113.003191
- Schuijt, T. J., Narasimhan, S., Daffre, S., DePonte, K., Hovius, J. W., Van't Veer, C., et al. (2011). Identification and characterization of *Ixodes scapularis* antigens that elicit tick immunity using yeast surface display. *PLoS One* 6:e15926. doi: 10.1371/journal.pone.0015926
- Schwarz, A., Tenzer, S., Hackenberg, M., Erhart, J., Gerhold-Ay, A., Mazur, J., et al. (2014). A systems level analysis reveals transcriptomic and proteomic complexity in *Ixodes ricinus* midgut and salivary glands during early attachment and feeding. *Mol. Cell. Proteomics* 13, 2725–2735. doi: 10.1074/mcp.M114.039289
- Scott, E. (1921). Hume and Hovell's journey to Port Phillip. *Roy. Aust. Hist. Soc.* 7, 289–380.
- Sears, D. W., Thompson, S. E., and Saxon, S. R. (2007). Reversible ligand binding reactions: why do biochemistry students have trouble connecting the dots? *Biochem. Mol. Biol. Educ.* 35, 105–118. doi: 10.1002/bmb.29
- Singh, K., Davies, G., Alenazi, Y., Eaton, J. R. O., Kawamura, A., and Bhattacharya, S. (2017). Yeast surface display identifies a family of evasins from ticks with novel polyvalent CC chemokine-binding activities. *Sci. Rep.* 7:4267. doi: 10.1038/s41598-017-04378-1
- Smith, R. D., Engdahl, A. L., Dunbar, J. B. Jr., and Carlson, H. A. (2012). Biophysical limits of protein-ligand binding. *J. Chem. Inf. Model.* 52, 2098–2106. doi: 10.1021/ci200612f
- Soares, C. A., Lima, C. M., Dolan, M. C., Piesman, J., Beard, C. B., and Zeidner, N. S. (2005). Capillary feeding of specific dsRNA induces silencing of the Isac gene in nymphal *Ixodes scapularis* ticks. *Insect Mol. Biol.* 14, 443–452. doi: 10.1111/j.1365-2583.2005.00575.x
- Soares, T. S., Watanabe, R. M., Tanaka-Azevedo, A. M., Torquato, R. J., Lu, S., Figueiredo, A. C., et al. (2012). Expression and functional characterization of Boophilin, a thrombin inhibitor from *Rhipicephalus* (Boophilus) microplus midgut. *Vet. Parasitol.* 187, 521–528. doi: 10.1016/j.vetpar.2012.01.027
- Sokol, C. L., and Luster, A. D. (2015). The chemokine system in innate immunity. *Cold Spring Harb. Perspect. Biol.* 7:a016303. doi: 10.1101/cshperspect.a016303
- Sonenshine, D. E. (2004). Pheromones and other semiochemicals of ticks and their use in tick control. *Parasitology* 129, S405–S425.
- Stutzer, C., Mans, B. J., Gaspar, A. R., Neitz, A. W., and Maritz-Olivier, C. (2009). *Ornithodoros savignyi*: soft tick apyrase belongs to the 5'-nucleotidase family. *Exp. Parasitol.* 122, 318–327. doi: 10.1016/j.exppara.2009.04.007
- Tanaka, A. S., Andreotti, R., Gomes, A., Torquato, R. J., Sampaio, M. U., and Sampaio, C. A. (1999). A double headed serine proteinase inhibitor-human plasma kallikrein and elastase inhibitor—from *Boophilus microplus* larvae. *Immunopharmacology* 45, 171–177. doi: 10.1016/s0162-3109(99)00074-0
- Tang, J., Fang, Y., Han, Y., Bai, X., Yan, X., Zhang, Y., et al. (2015). YY-39, a tick anti-thrombosis peptide containing RGD domain. *Peptides* 68, 99–104. doi: 10.1016/j.peptides.2014.08.008
- Tatchell, R. J. (1967). A modified method for obtaining tick oral secretion. *J. Parasitol.* 53, 1106–1107.
- Tatchell, R. J., and Moorhouse, D. E. (1968). The feeding processes of the cattle tick *Boophilus microplus* (Canestrini). II. The sequence of host-tissue changes. *Parasitology* 58, 441–459.

- Tirloni, L., Islam, M. S., Kim, T. K., Diedrich, J. K., Yates, J. R. III, Pinto, A. F., et al. (2015). Saliva from nymph and adult females of *Haemaphysalis longicornis*: a proteomic study. *Parasit. Vectors* 8:338. doi: 10.1186/s13071-015-0918-y
- Tirloni, L., Reck, J., Terra, R. M., Martins, J. R., Mulenga, A., Sherman, N. E., et al. (2014). Proteomic analysis of cattle tick *Rhipicephalus* (*Boophilus*) *microplus* saliva: a comparison between partially and fully engorged females. *PLoS One* 9:e94831. doi: 10.1371/journal.pone.0094831
- Tyson, K., Elkins, C., Patterson, H., Fikrig, E., and de Silva, A. (2007). Biochemical and functional characterization of Salp20, an *Ixodes scapularis* tick salivary protein that inhibits the complement pathway. *Insect Mol. Biol.* 16, 469–479. doi: 10.1111/j.1365-2583.2007.00742.x
- Tyson, K. R., Elkins, C., and de Silva, A. M. (2008). A novel mechanism of complement inhibition unmasked by a tick salivary protein that binds to properdin. *J. Immunol.* 180, 3964–3968. doi: 10.4049/jimmunol.180.6.3964
- Uhlir, J., Grubhoffer, L., Borski, L., and Dusbábek, F. (1994). Antigens and glycoproteins of larvae, nymphs and adults of the tick *Ixodes ricinus*. *Med. Vet. Entomol.* 8, 141–150. doi: 10.1111/j.1365-2915.1994.tb00154.x
- Valdés, J. J., Cabezas-Cruz, A., Sima, R., Butterill, P. T., Růžek, D., and Nuttall, P. A. (2016). Substrate prediction of *Ixodes ricinus* salivary lipocalins differentially expressed during *Borrelia afzelii* infection. *Sci. Rep.* 6:32372. doi: 10.1038/srep32372
- Valdés, J. J., Schwarz, A., Cabeza de Vaca, I., Calvo, E., Pedra, J. H., Guallar, V., et al. (2013). Tryptogalinin is a tick Kunitz serine protease inhibitor with a unique intrinsic disorder. *PLoS One* 8:e62562. doi: 10.1371/journal.pone.0062562
- Valenzuela, J. G., Charlab, R., Mather, T. N., and Ribeiro, J. M. (2000). Purification, cloning, and expression of a novel salivary anticomplement protein from the tick, *Ixodes scapularis*. *J. Biol. Chem.* 275, 18717–18723. doi: 10.1074/jbc.m001486200
- van de Locht, A., Stubbs, M. T., Bode, W., Friedrich, T., Bollschweiler, C., Höffken, W., et al. (1996). The ornithodorin-thrombin crystal structure, a key to the TAP enigma? *EMBO J.* 15, 6011–6017. doi: 10.1002/j.1460-2075.1996.tb00989.x
- Viljoen, G. J., Neitz, A. W. H., Prozesky, L., Bezuidenhout, J. D., and Vermeulen, N. M. J. (1985). Purification and properties of tick egg toxic proteins. *Insect Biochem.* 15, 475–482. doi: 10.1016/0020-1790(85)90060-5
- von Siebold, C. T. H., and Stannius, H. (1854). *Comparative Anatomy*. London: Trubner and Company.
- Wagner, C. L., Mascelli, M. A., Neblock, D. S., Weisman, H. F., Collier, B. S., and Jordan, R. E. (1996). Analysis of GPIIb/IIIa receptor number by quantification of 7E3 binding to human platelets. *Blood* 88, 907–914.
- Wang, H., Kaufman, W. R., Cui, W. W., and Nuttall, P. A. (2001b). Molecular individuality and adaptation of the tick *Rhipicephalus appendiculatus* in changed feeding environments. *Med. Vet. Entomol.* 15, 403–412. doi: 10.1046/j.0269-283x.2001.00328.x
- Wang, H., Hails, R. S., Cui, W. W., and Nuttall, P. A. (2001a). Feeding aggregation of the tick *Rhipicephalus appendiculatus* (Ixodidae): benefits and costs in the contest with host responses. *Parasitology* 123, 447–453.
- Wang, H., and Nuttall, P. A. (1994). Comparison of the proteins in salivary glands, saliva and haemolymph of *Rhipicephalus appendiculatus* female ticks during feeding. *Parasitology* 109, 517–523.
- Wang, H., Paesen, G. C., Nuttall, P. A., and Barbour, A. G. (1998). Male ticks help their mates to feed. *Nature* 391, 753–754. doi: 10.1038/35773
- Wang, X., Coons, L. B., Taylor, D. B., Stevens, S. E. Jr., and Gartner, T. K. (1996). Variabilin, a novel RGD-containing antagonist of glycoprotein IIb-IIIa and platelet aggregation inhibitor from the hard tick *Dermacentor variabilis*. *J. Biol. Chem.* 271, 17785–17790. doi: 10.1074/jbc.271.30.17785
- Wang, Y., Yu, X., Cao, J., Zhou, Y., Gong, H., Zhang, H., et al. (2015). Characterization of a secreted cystatin from the tick *Rhipicephalus haemaphysaloides*. *Exp. Appl. Acarol.* 67, 289–298. doi: 10.1007/s10493-015-9946-8
- Watts, S. W., Morrison, S. F., Davis, R. P., and Barman, S. M. (2012). Serotonin and blood pressure regulation. *Pharmacol. Rev.* 64, 359–388. doi: 10.1124/pr.111.004697
- Waxman, L., and Connolly, T. M. (1993). Isolation of an inhibitor selective for collagen-stimulated platelet aggregation from the soft tick *Ornithodoros moubata*. *J. Biol. Chem.* 268, 5445–5449.
- Waxman, L., Smith, D. E., Arcuri, K. E., and Vlasuk, G. P. (1990). Tick anticoagulant peptide (TAP) is a novel inhibitor of blood coagulation factor Xa. *Science* 248, 593–596. doi: 10.1126/science.2333510
- Whisstock, J. C., Silverman, G. A., Bird, P. I., Bottomley, S. P., Kaiserman, D., Luke, C. J., et al. (2010). Serpins flex their muscle: II. Structural insights into target peptidase recognition, polymerization, and transport functions. *J. Biol. Chem.* 285, 24307–24312. doi: 10.1074/jbc.R110.141408
- Wiegner, R., Chakraborty, S., and Huber-Lang, M. (2016). Complement-coagulation crosstalk on cellular and artificial surfaces. *Immunobiology* 221, 1073–1079. doi: 10.1016/j.imbio.2016.06.005
- Wikel, S. (2017). “Arthropod modulation of wound healing. in arthropod vector,” in *Controller of Disease Transmission*, Vol. 2, eds S. K. Wikel, S. Aksoy, and G. Dimopoulos (Amsterdam: Elsevier), 31–50. doi: 10.1016/b978-0-12-805360-7.00003-4
- Xu, T., Lew-Tabor, A., and Rodriguez-Valle, M. (2016). Effective inhibition of thrombin by *Rhipicephalus microplus* serpin-15 (RmS-15) obtained in the yeast *Pichia pastoris*. *Ticks Tick Borne Dis.* 7, 180–187. doi: 10.1016/j.ttbdis.2015.09.007
- Xu, X. L., Cheng, T. Y., and Yang, H. (2016). Enolase, a plasminogen receptor isolated from salivary gland transcriptome of the ixodid tick *Haemaphysalis flava*. *Parasitol. Res.* 115, 1955–1964. doi: 10.1007/s00436-016-4938-0
- Yokomizo, T. (2015). Two distinct leukotriene B4 receptors, BLT1 and BLT2. *J. Biochem.* 157, 65–71. doi: 10.1093/jb/mvu078
- Zhu, K., Bowman, A. S., Brigham, D. L., Essenberg, R. C., Dillwith, J. W., and Sauer, J. R. (1997a). Isolation and characterization of Americanin, a specific inhibitor of thrombin, from the salivary glands of the lone star tick *Amblyomma americanum* (L.). *Exp. Parasitol.* 87, 30–38. doi: 10.1006/exp.1997.4175
- Zhu, K., Sauer, J. R., Bowman, A. S., and Dillwith, J. W. (1997b). Identification and characterization of anticoagulant activities in the saliva of the lone star tick, *Amblyomma americanum* (L.). *J. Parasitol.* 83, 38–43.

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Mans. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.