Potential candidate genes:
EgrIAA (55)

Potential candidate genes:
EgrIAA (105)

## Manual curation:

1) Protein motif scan
2) Complete partial sequence by FgeneSH


Supplementary Fig. S1 Procedure used for identifying Aux/IAA genes in Eucalyptus grandis. Arabidopsis Aux/IAA protein sequences were used to search for related proteins in the predicted E. grandis proteome by using BLASTP. Fifty-five E. grandis proteins identified in this initial search were further examined by manual curation using protein motif scanning and the FgeneSH (Solovyev 2007) program to complete partial sequences. Redundant and invalid genes were eliminated based on gene structure, the integrity of conserved motifs and EST support. Manual curation resulted in 26 EgrIAA protein sequences. We then used these 26 protein sequences in two subsequent additional searches: first, a BLASTP search against the $E$. grandis proteome to identify exhaustively all divergent E. grandis Aux/IAA gene family members and, second, tBLASTn searches against the E. grandis genome for any possible unpredicted genes. To confirm our findings, we also used poplar Aux/IAA proteins to repeat the complete search procedure described above with identical results obtained.


Supplementary Fig. S2 Phylogenetic analysis of E. grandis and Arabidopsis IAA proteins and the exon-intron organization of corresponding genes. (A) Neighbor-joining phylogenetic tree of E. grandis and Arabidopsis IAA proteins. Full-length predicted IAA protein sequences were aligned using Clustal_X2 program and phylogenetic tree was constructed by using the MEGA5 program and neighbor-joining method with 1,000 bootstrap replicates. Bootstrap values are indicated at each node. (B) Exon-intron organization of the EgrIAA (in red) and AtIAA (in green) genes corresponding to the proteins in the phylogenetic tree. The sizes of exons and introns are indicated by the scale at the top. The information on exon-intron structure was extracted from the Phytozome database and visualized by using the FancyGene software (http://bio.ieo.eu/fancygene/).
$\square$ Consensus
 HDIQLGLALPIHDHPAKGSEPKDHEGSYKKHFHKKRCHGEFFGEHSSGKSQLLAHSSGPNEEDDPHARRGKFSCSIHQFRYSEGRDDHDRYYGHPPIKSHRKKFHHGQRPPRRDLHENYRQA MDLQLGLALPISDHSAKGSEPKDHEGLYKKHAHHKRCHGERFGQYSSGKSQSLAMSGRPHEEDDPSARRSKFYCSLAKFRYSEGADDHDQYYGHPPIQTHRKKFFHGQRPPRRDLYKIYRQA
 $\begin{array}{llllllllrr}131 & 140 & 150 & 160 & 170 & 180 & 190 & 200 & 210 & 217\end{array}$ HNHFYKYKMEGYAIYRKINLRTYRSYNSLKGHLIAMFSRYKRDDFKDHASYTLTYQDKEGDHLLAGDLPHLHFYESYHRLQIQRSRD

EgrIfR29 Egrifhe9n EgrIAR29B Consensus

Supplementary Fig. S3 Protein sequences alignment of EgrIAA29 and two truncated genes, EgrIAA29A and 29B.


Supplementary Fig. S4 Multiple sequence alignment of predicted amino acid sequences of EgrIAA proteins. The multiple sequence alignment was obtained with Clustal_x2 and manual correction. The four highly conserved domains of Aux/IAA proteins were noted on the top of the alignment. Nuclear localization signals (NLSs) were indicated by dots. The amino acid position was given on the right of each sequence.


Supplementary Fig. S5 Chromosomal positions of the EgrIAA genes. The tandem duplications (TD) and segmental duplication (SD) are written on the right side of the corresponding genes. Chromosome scaffolds (1-11) are indicated at the top end of each chromosome.


Supplementary Fig. S6 Predicted proteins of all the transcripts of EgrIAA and their conserved domains. Conserved domains were predicted using the MEME web server (Bailey et al. 2009) with the following parameters: distribution of motif occurrences, zero or one per sequence; motif width ranges from 10 to 50 residues; maximum number of motifs to find, 4. The size and location of motifs can be estimated by the scale at bottom.

Low expression in phloem
Responsiveness to bending
$\square$ Preferential expression in mature versus juvenile xylem


Supplementary Fig. S7 Venn diagram of overall strategy to identify the best potential candidate(s) involved in wood formation. Best potential candidate(s) should be i) highly expressed in vascular tissues; ii) preferentially expressed in xylem/cambium compare to phloem; iii) showing a response to bending; iv) preferential expression in mature versus juvenile xylem. Only EgrIAA4 is fulfilling these conditions.

MAAQGEDLNLEATELRLGLPGTVEPEKQQAPLSGRSMKRNLIDVNNEYGSNEEESNGSSA QKCDKQDVHRPSKAQVVGWPPVRSYRKNCFQKKAEGESTGVFIKVSMDGAPYLRKIDLKP YKGYSDLLKDLQGMFKFKVGEYCEREGYNGSEFVPTYEDKDGDWMLVGDVPWNMFITSCK RLRIMKGSSEV


Supplementary Fig. S8 Protein sequence of EgrIAA4 and the phenotype of EgrIAA4m transgenic Arabidopsis. (A) Amino acid sequence of mutated EgrIAA4m protein. In the mutated EgrIAA4m cDNA, the 80th codon encoding the conserved Pro residue (highlighted in red) was changed to a codon encoding a Ser residue. (B) Transcript levels of EgrIAA4m in transgenic Arabidopsis under the control of the 35 SCaMV promoter. Three technical replicates were used for each qRT-PCR experiment. Bars shows averages ratio of the EgrIAA4m relative to that of $A t U B Q 10$. Error bars represent SE. (C) EgrIAA4m transgenic and wild type Arabidopsis plants grown for 40 days. Scale bar: 1 cm .

