

A morphological analysis of the subtribe *Pegylina* Lacroix, 1989 (Scarabaeidae: Melolonthinae: Melolonthini) reconstitutes its generic composition

J. du G. Harrison^{1,2,3}

¹Department of Zoology and Entomology, Forestry and Agricultural Biotechnology Institute, University of Pretoria, Pretoria, 0002 South Africa

²School of Animal, Plant and Environmental Sciences, University of the Witwatersrand, Private Bag 3, Johannesburg, 2050 South Africa*

³Department of Invertebrates, Ditsong National Museum of Natural History (formerly Transvaal Museum), Pretoria, 0001 South Africa

Phylogenetic analysis using six outgroup and 26 ingroup taxa and 18 external adult morphological characters resulted in a paraphyletic *Pegylina* as composed by Lacroix (1989). To maintain the *Pegylina* as a monophyletic subtribe, the genus *Stenopegylis* Arrow, 1943 is removed from the *Pegylina* and placed in the Melolonthini *sensu lato*, pending further cladistic analyses of the Afrotropical melolonthines. Following the analyses presented here, eight character areas (antennomere number, labrum form, protarsal claw shape, protibial spur, pronotum, mesosternum and mesosternal process, body shape and sexual dimorphism) are discussed with reference to differentiating *Hypopholis* Erichson, 1847 syn. nov. from *Pegylis* Erichson, 1847. Consequently, the former is synonymized with the latter, resulting in the following new combinations: *Pegylis sommeri* (Burmeister, 1855) comb. nov.; *P. uelensis* (Burgeon, 1946) comb. nov. and *P. vittata* (Fåhraeus in Boheman, 1857) comb. nov. An appended checklist for the subtribe *Pegylina* as presently composed includes 37 species in two genera, *Eupegylis* Duvivier, 1892 (1 sp.) and *Pegylis* Erichson, 1847 (36 spp.). During outgroup selection for this analysis it became apparent that the monotypic *Wernerophylla nigra* Lacroix, 2001 syn. nov. is conspecific with *Stenopegylis cylindrica* Arrow, 1943 and consequently this synonymy is implemented here.

Key words: Melolonthinae, *Adoretopsis*, *Eupegylis*, *Hypopholis*, *Pegylis*, *Pegylidius*, *Wernerophylla*, *Stenopegylis*, southern Africa.

INTRODUCTION

Lacroix (1989) created the Melolonthinae tribe Pegylini to accommodate species from four genera: *Eupegylis* Duvivier, 1892 (1 sp.); *Hypopholis* Erichson, 1847 (3 spp.); *Pegylis* Erichson, 1847 (32 spp.); and *Stenopegylis* Arrow, 1943 (1 sp.) using the following morphological characters as support: (i) protibia without an apical spur; (ii) labrum elongate and bilobed; (iii) antennal club three-segmented (5–6 in *Eupegylis*); (iv) anterior margin of pronotum bordered by a membrane; (v) pronotum transverse; (vi) metepisternum wide (at the same level as the metepimeron); (vii) anterior femur non-subvertical; (viii) body slightly elongated, rounded or oval; (ix) sexual dimorphism limited, *i.e.* undeveloped.

Smith (2006), in his review of family-group names in the Scarabaeoidea, regarded the Pegylini as a subtribe (*Pegylina*) of the Melolonthini. Bouchard *et al.* (2011) followed Smith (2006), and this same rank is adopted here.

This study assesses the relationship of the genera *Eupegylis*, *Hypopholis*, *Pegylis*, *Stenopegylis* and

Wernerophylla based on a morphological phylogenetic analysis to determine or refute the monophyly of the *Pegylina*. It is the first of two companion papers, with the second (Harrison 2014) being a taxonomic review of the three South African species of *Pegylis*.

MATERIAL AND METHODS

Nomenclature and specimens used

The first mention of genera here includes authorship and date of publication, for example, *Pegylis* Erichson, 1847, while that for species is provided in Table 1 and Appendix 1.

Many of the specimens used in this analysis were primary or secondary museum type specimens. The advantage of this is that it allowed for wide taxon sampling within *Pegylis*, its related genera and synonyms, and scoring of characters from unequivocally authoritative specimens. The disadvantage is that disarticulation of these specimens to extend the character suite to include internal morphological characters is prohibited. Conse-

*Present address. E-mail: james.harrison@wits.ac.za

quently, extending the character suite to include internal morphological characters, *e.g.* mouthparts would undoubtedly result in the discovery of additional characters useful for testing the monophyly of the subtribe Pegylina. However, an overriding aim of this study was to examine the generic relationship between *Hypopholis* and *Pegylis* in order to compile the much needed review of the South African 'large wattle chafers' (Harrison 2014), which this study achieves.

Ingroup taxa

Currently there are 37 species from the genera placed by Lacroix (1989) in the Pegylina: *Eupegylis* (1); *Hypopholis* (3); *Pegylis* (32); and *Stenopegylis* (1). Twenty-six of these species (70 % of the known taxa) were examined for this study (Table 1). In order to enhance monophyly, *Pegylis* species previously placed in genera other than indicated above are also included. Thus the monotypic *Adoretopsis tenuitarsis* Fairmaire, 1887 and *Pegylidius mashunus* Péringuey, 1904 (now both placed in *Pegylis*) are included as ingroup taxa. Table 1 lists all ingroup species as originally placed at the generic level to provide an indication of their nomenclatural history. This accounts for the larger number of *Hypopholis* species in Table 1 than currently recognized. During on-going taxonomic work on African melolonthines it became apparent that *Wernerophylla nigra* Lacroix, 2001 described and placed by Lacroix (2001, 2007) in the Pachydemini, is actually a synonym (first proposed here) of *Stenopegylis cylindrica* Arrow, 1943. *Stenopegylis cylindrica* was separately placed by Lacroix (2010) in the Pegylina and thus by default, *W. nigra*, is included in the analysis as *S. cylindrica*.

Outgroup taxa

Outgroup taxa include: (i) *Asthenopholis adspersa*; (ii) *Brachylepis elephas*; (iii) *Eulepida lepidota*; (iv) *Psilonychus duponti*; (v) *Psilonychus eckloni*; and (vi) *Rhabdopholis albostrigata*. These genera were chosen because i–iii are included by Péringuey (1904) together with the following ingroup genera *Hypopholis*, *Pegylidius* and *Pegylis* in his group 'Leucopholides' (= Leucopholina), thus allowing for some degree of comparison between genera presently (2014) placed into two separate subtribes of the Melolonthini, *i.e.* Pegylina and Leucopholina. Additionally, *Rhabdopholis* Burmeister, 1855; *Asthenopholis* Brenske, 1898; and *Psilonychus* Burmeister, 1855 are morphologically similar to

both *Pegylis* and *Stenopegylis* Arrow, 1943 and were thus also included. *Rhabdopholis* shares a mesosternal process and similar body shape to *Hypopholis*, while *Asthenopholis* and *Psilonychus* share a propygidial groove with *Stenopegylis*.

Characters and coding

The most informative characters from recent phylogenetic studies (Jameson 1997; Ahrens 2005; Coca-Abia 2007; Katovich 2008) of other Melolonthinae taxa were examined. Characters reused here are indicated in Table 2, although in some instances their states were modified to make them applicable to this study. During character selection, characters used in taxonomic descriptions of genera or higher taxonomic categories were specifically included. However, some of these proved difficult to code, or were invariant across the taxa used in the analysis and were thus excluded from the final data matrix (*e.g.* the form of the labrum). The resulting 18 morphological characters chosen to score the taxa studied here are provided in Table 2, while the character matrix is presented in Table 1.

Cladistic analysis

The matrix was analysed using a heuristic search in the WinClada (Ver. 1.00.08) interface (Nixon 2002). From WinClada the data analyses were undertaken in Nona (Goloboff 1997) using these parameter settings: hold 10 000 (hold a maximum of 10 000 trees in memory); mult*100 (with 100 randomized replications); hold/10 (starting each replication of branch-swapping with 10 trees in memory); 0 = time ('pseudo-random number generator based on the time in order to select the sequence of addition of the replications' Hardy *et al.* (2003)). With the following search strategy mult*max* (multiple tree bisection-reconnection (TBR) + TBR), in an unconstrained search, as suggested by Hardy *et al.* (2003) for an initial search on data sets with less than 120 taxa. The characters were treated as non-additive and with equal weights. Characters were polarized by outgroups (Nixon & Carpenter 1993), represented herein by *Asthenopholis* (used for rooting), *Brachylepis*, *Eulepida*, *Rhabdopholis*, *Psilonychus* and including other genera of Pegylina as more closely-related outgroups, *i.e.* *Eupegylis* and *Stenopegylis*. WinClada was also used to view character states and to calculate a Nelsen strict consensus tree (SCT) and bootstrap and jack-knife support values for the nodes on the resulting SCT.

Table 1. Character states of the six outgroup and 26 ingroup taxa used in the analysis. Character descriptions provided in Table 2. '?' = unknown character state/s.

| Characters | | 1 | 1 | 1 |
|--------------------------------|--------------------------|------------|----------|---|
| | | 1234567890 | 12345678 | |
| Outgroup taxa | | | | |
| <i>Asthenopholis adspersa</i> | (Boheman, 1857) | 0000002102 | 01101201 | |
| <i>Brachylepis elephas</i> | (Gerstaecker, 1867) | 0000000122 | 01100202 | |
| <i>Eulepida lepidota</i> | (Klug, 1855) | 0200002120 | 01100201 | |
| <i>Psilonychus duponti</i> | Burmeister, 1855 | 2300002122 | 00001100 | |
| <i>Psilonychus eckloni</i> | Burmeister, 1855 | 2200002121 | 00001100 | |
| <i>Rhabdopholis albostrata</i> | Burmeister, 1855 | 0200002022 | 02000100 | |
| Ingroup taxa | | | | |
| <i>Adoretopsis tenuitarsis</i> | Fairmaire, 1887 | 1100011003 | 11110200 | |
| <i>Eupegylis confusa</i> | Duvivier, 1892 | 1100120100 | 11000303 | |
| <i>Hypopholis conspurcata</i> | Gerstaecker, 1867 | 1200011003 | 11110112 | |
| <i>Hypopholis sommeri</i> | Burmeister, 1855 | 1110011003 | 02010100 | |
| <i>Hypopholis uelensis</i> | Burgeon, 1946 | 1200011003 | 02010200 | |
| <i>Hypopholis vittata</i> | Fähræus in Boheman, 1857 | 1110010003 | 01010100 | |
| <i>Pegylidius mashunus</i> | Péringuey, 1904 | 1200010003 | 10110201 | |
| <i>Pegylis angolensis</i> | Moser, 1915 | 11?0010000 | 00110202 | |
| <i>Pegylis bennigseni</i> | Brenske, 1898 | 1200010003 | 10110201 | |
| <i>Pegylis burgeoni</i> | Decelle, 1968 | 1100110003 | 20110001 | |
| <i>Pegylis ertli</i> | Moser, 1919 | 1210010003 | 21110115 | |
| <i>Pegylis gestroi</i> | Brenske, 1895 | 1200010003 | 20110115 | |
| <i>Pegylis giraudetae</i> | Decelle, 1968 | 1100110013 | 11110200 | |
| <i>Pegylis hauseri</i> | Brenske, 1898 | 1000010003 | 20110100 | |
| <i>Pegylis kenyensis</i> | Decelle, 1968 | 1201010003 | 20110106 | |
| <i>Pegylis lindiana</i> | Moser, 1919 | 1110010003 | 11110200 | |
| <i>Pegylis maculipennis</i> | van Lansberge, 1889 | 1201010003 | 10110100 | |
| <i>Pegylis microchaeta</i> | Moser, 1919 | 1100011003 | 21110200 | |
| <i>Pegylis morio</i> | Blanchard, 1851 | 1201010000 | 00110106 | |
| <i>Pegylis neumanni</i> | Kolbe, 1894 | 1200010003 | 10110200 | |
| <i>Pegylis pilosa</i> | Lacroix, 2008b | 1000010010 | 00110101 | |
| <i>Pegylis pondoensis</i> | Arrow, 1943 | 1110011003 | 01010000 | |
| <i>Pegylis rufolineata</i> | Kolbe, 1894 | 1200010003 | 21110115 | |
| <i>Pegylis rufomaculatus</i> | Linell, 1896 | 1210010003 | 21110115 | |
| <i>Pegylis vestita</i> | Brenske, 1895 | 1201010013 | 20110200 | |
| <i>Stenopegylis cylindrica</i> | Arrow, 1943 | 1100003142 | 00001200 | |

RESULTS AND DISCUSSION

The parsimony analysis produced 46 equally most parsimonious (EMP) trees, with a length of 94 steps, and a CI = 0.39 and RI = 0.71. A Nelsen strict consensus tree (SCT) was calculated from these 46 trees. The SCT (Fig. 1) has a length of 144 steps, with a CI = 0.47 and RI = 0.47. Bootstrap (Fig. 2) and jack-knife (Fig. 3) support values were calculated for the nodes of the SCT. The number of steps, ci and ri for each character are

included in Table 2. The checklist in Appendix 1 lists all species of Pegylina including the generic changes resulting from this analysis.

Monophyly of the subtribe Pegylina Lacroix, 1989

Node: (((*Asthenopholis adspersa*) (*Brachylepis elephas* + *Eulepida lepidota* + *Rhabdopholis albostrata* + *Stenopegylis cylindrica*) (*Psilonychus eckloni* + *Psilonychus duponti*)) (((*Eupegylis confusa* + *Pegylis angolensis* + *Pegylis bennigseni* + *Hypopholis conspurcata* + *Pegylis giraudetae* + *Pegylis lindiana* + *Pegylidius mashunus* + *Pegylis*

Table 2. Adult morphological characters (scored for both sexes) used in the phylogenetic analysis of genera placed by Lacroix (1989) in the subtribe Pegylina. For each character, the number of steps in the tree (S), consistency index (ci) and the retention index (ri) are provided, based on the parsimony analysis performed in WinClada. References within the table indicate character sources, prior to these being modified and used in this analysis.

-
- 1) Protibial spur: (0) present; (1) absent; (2) residual. S = 4; ci = 50; ri = 50.
 - 2) Number of protibial teeth: (0) three; (1) two (third reduced); (2) two; (3) one (second reduced). S = 15; ci = 20; ri = 29.
 - 3) Protarsal claws (in males): (0) symmetrical; (1) asymmetrical. S = 5; ci = 20; ri = 20.
 - 4) Number of antennomeres: (0) nine or ten; (1) eight or less. S = 2; ci = 50; ri = 66. (Ahrens 2005: character 5).
 - 5) Eyes: (0) normally convex; (1) strongly convex; (2) with dorsal light patch. S = 3; ci = 33; ri = 0.
 - 6) Anterior margin of pronotum bordered by a membrane: (0) absent; (1) present; (2) present, but slight. S = 2; ci = 100; ri = 100.
 - 7) Anterior margin (dorsal section) of pronotum: (0) defined by a groove (marginal bead) continuous from side to side; (1) defined by a groove only on each side towards the angles; (2) groove absent; (3) notched on each side towards the angle. S = 11; ci = 27; ri = 50.
 - 8) Lateral margins of pronotum: (0) smooth; (1) finely serrated; (2) coarsely serrated. S = 3; ci = 33; ri = 66. (Coca-Abia 2007: character 16).
 - 9) Pronotum surface: (0) uniformly clothed with short fine setae; (1) uniformly clothed with long fine setae; (2) scaly; (3) no or few setae / scales; (4) uniformly clothed with short and long fine setae. S = 9; ci = 44; ri = 61.
 - 10) Colour of elytra: (0) brown; (1) russet; (2) black; (3) bi-coloured. S = 16; ci = 18; ri = 31.
 - 11) Mottling on elytra: (0) absent; (1) slightly mottled; (2) strongly mottled. S = 13; ci = 15; ri = 47.
 - 12) Mesosternum (as a mesosternal keel): (0) not produced; (1) weakly produced, not surpassing the base of mesocoxae; (2) produced anteriorly beyond base of the mesocoxae. S = 14; ci = 14; ri = 25. (Jameson 1997: character 110).
 - 13) Mesosternum: (0) without rim; (1) with rim. S = 8; ci = 12; ri = 12.
 - 14) Median depression on abdominal sternites (of male): (0) absent; (1) present. S = 2; ci = 50; ri = 85.
 - 15) Propygidial groove: (0) absent; (1) present. S = 0. Uninformative.
 - 16) Adeagus lateral profile: (0) straight; (1) smoothly curved; (2) sharply angulate; (3) laterally produced. S = 13; ci = 23; ri = 33.
 - 17) Adeagus symmetry: (0) symmetrical; (1) asymmetrical. S = 3; ci = 33; ri = 50.
 - 18) Adeagus symmetry and complexity: (0) symmetrical and simple (no protuberances); (1) symmetrical and complex (one set of protuberances); (2) symmetrical and complex (two or more sets of protuberances); (3) symmetrical and complex (including setae) (4) asymmetrical and simple (one set of protuberances); (5) asymmetrical and complex (two or more sets of protuberances); (6) symmetrical with a three-bladed internal section. S = 21; ci = 28; ri = 63.
-

microchaeta + *Pegylis neumanni* + *Pegylis pilosa* + *Pegylis pondoensis* + *Hypopholis sommeri* + *Adoretopsis tenuitarsis* + *Hypopholis uelensis* + *Hypopholis vittata* (*Pegylis maculipennis* + *Pegylis vestita*) (*Pegylis burgeoni* + *Pegylis hauseri*) (((*Pegylis gestroi* + *Pegylis rufolineata*) (*Pegylis ertli* + *Pegylis rufomaculatus*)) (*Pegylis kenyensis* + *Pegylis morio*))) = Fig. 1.

As currently composed, the Pegylina is paraphyletic (Fig. 1) due to *Stenopegyilis* (= *Wernerophylla*), a monotypic genus, occurring outside of the 'Pegylina clade'. However, the removal of *Stenopegyilis* from the Pegylina, based on the absence of a membrane on its anterior pronotal margin (an apparent synapomorphy for the Pegylina), returns

the Pegylina to a monophyletic group based on the following synapomorphies, with 'character: state' indicated in brackets: anterior margin of pronotum bordered by a *membrane* (C6:S1); *bi-coloured* elytra (C10:S3); *slight mottling* on elytra (C11:S1); and median depression on abdominal sternites (of male) *present* (C14:S1).

Support values for the Pegylina clade (((*Eupegyilis confusa* ... *Pegylis morio*))) are 65 % bootstrap support (Fig. 2) and 74 % jack-knife (Fig. 3).

Consequently, *Stenopegyilis cylindrica* Arrow, 1943 is transferred to the Melolonthini *sensu lato* pending further analyses of relationships among African

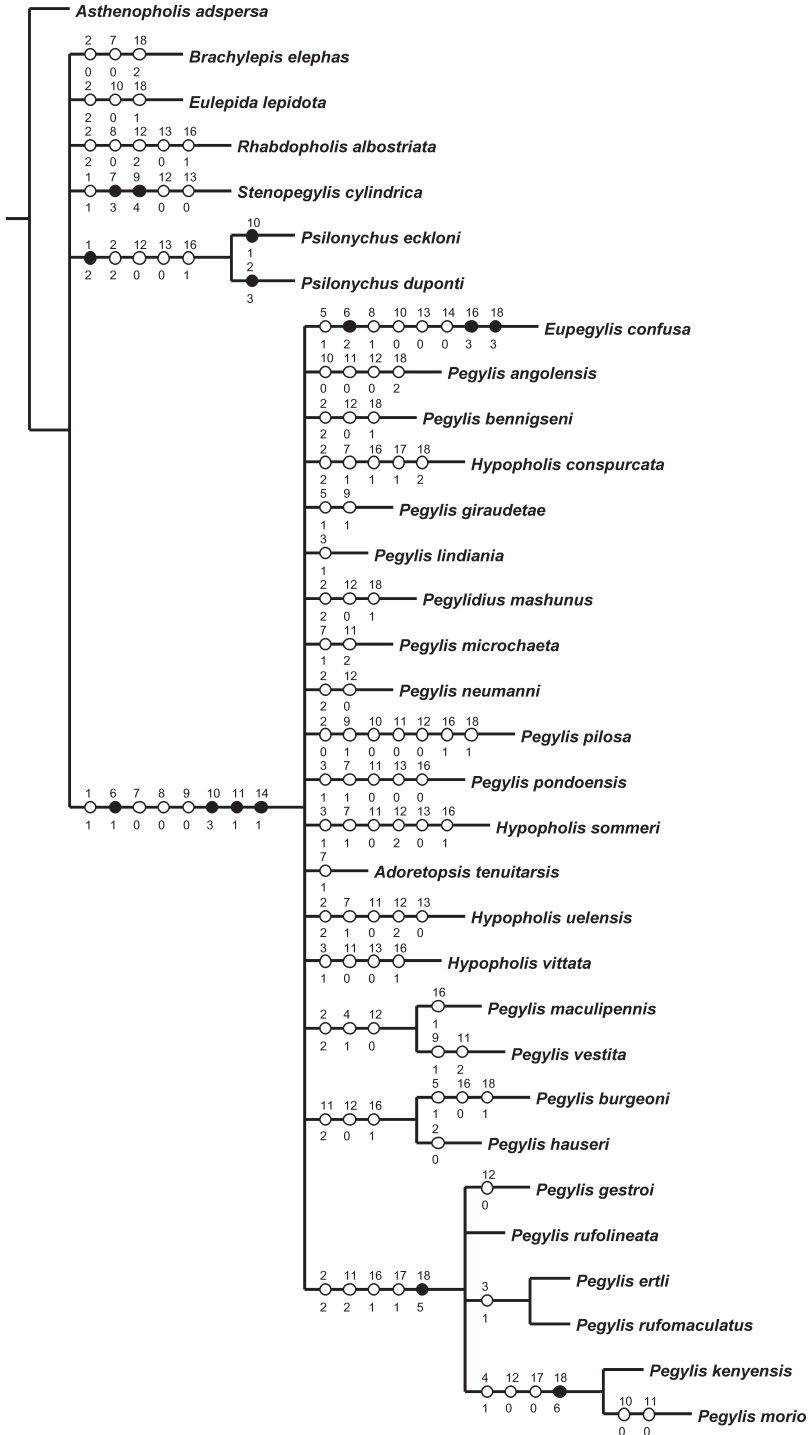


Fig. 1. Strict consensus tree for the subtribe Pegylina and outgroup taxa included in the analysis, using *Asthenopholis adspersa* to root the tree. Open circles (○) represent homoplasious character state changes. Filled circles (●) represent non-homoplasious character state changes. Character number above each circle; character state below each circle.

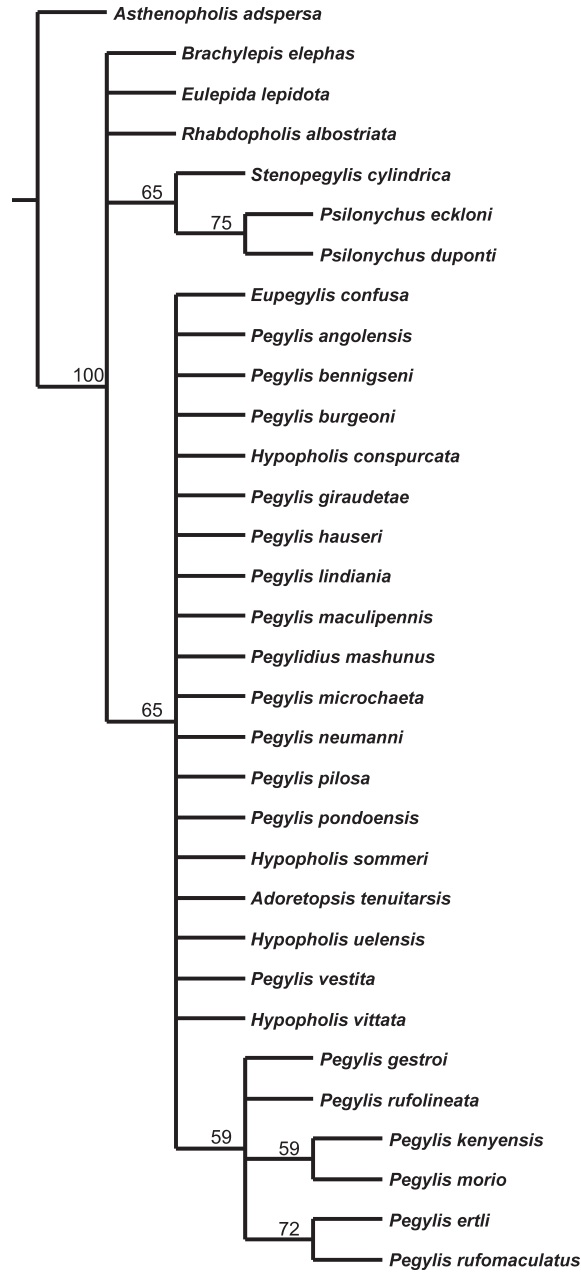


Fig. 2. SCT with bootstrap values indicated on supported nodes.

genera of Melolonthinae. Arrow (1943: 783–784) wrote the following after having described this monotypic genus:

Although differing very greatly from *Pegylis* in its external appearance, the insect for which this genus is proposed is nearly related to it and shares with it the peculiar sexual difference in claw-structure as

well as the no less peculiar absence of the normal tibial spur at the base of the front tarsus. It differs especially in its narrow cylindrical shape, instead of the broadly ovate form characteristic of *Pegylis*, the small head, with its prominent clypeus, the short diverging lobes of the labrum, and the existence of a longitudinal groove upon the propygidium, similar to that I described in 1902 [Arrow 1902] as distin-

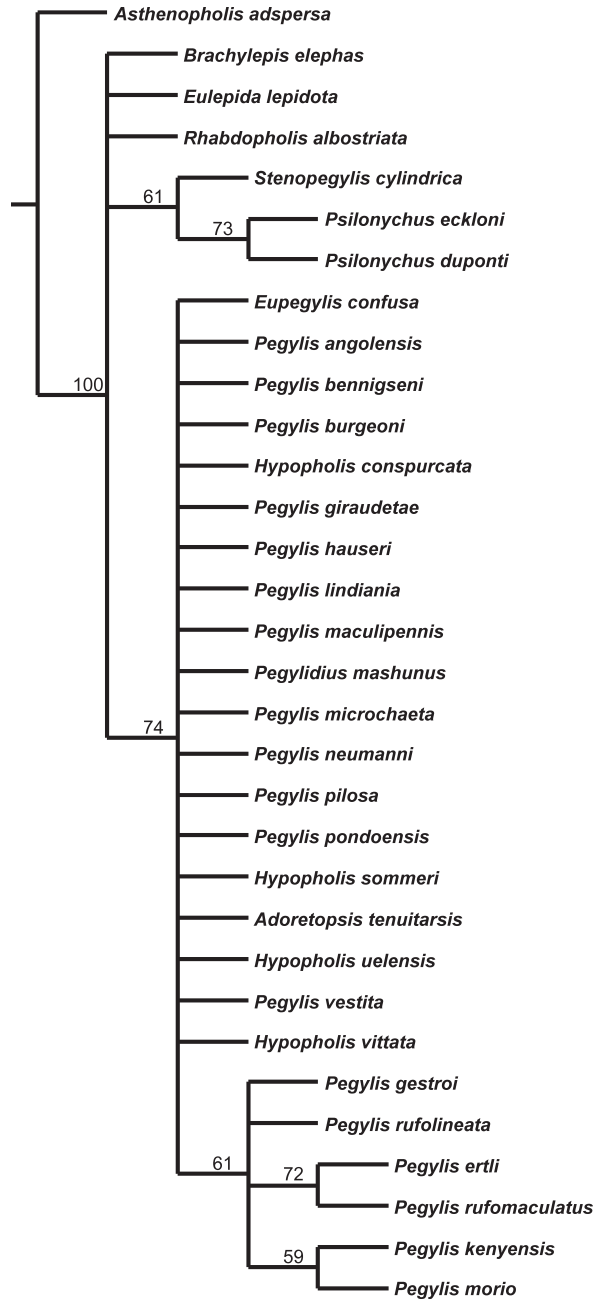


Fig. 3. SCT with jack-knife values on supported nodes.

guishing the genus *Asthenopholis*, to which *Stenopegylis* has some superficial resemblance, although clothed with hair instead of scales.

Harrison (2009) provided a revision of *Asthenopholis* including a discussion of the occurrence of a grooved propygidium within various scarab taxa.

Monophyly of *Adoretopsis*, *Hypopholis* and *Pegylidius*

Node: (((*Eupegylis confusa* + *Pegylis angolensis* + *Pegylis bennigseni* + *Hypopholis conspurcata* + *Pegylis giraudetae* + *Pegylis lindiania* + *Pegylidius mashunus* + *Pegylis microchaeta* + *Pegylis neumanni* + *Pegylis pilosa* + *Pegylis pondoensis* + *Hypopholis sommeri* + *Adoretopsis*

Table 3. Morphological features used by Erichson (1847) to differentiate *Hypopholis* from *Pegylis*.

| <i>Hypopholis</i> | <i>Pegylis</i> |
|---|---|
| (i) Antennae 10-segmented with 3-segmented club | Antennae 8-segmented with 3-segmented club |
| (ii) Labrum large, deeply split/incised* | Labrum as in <i>Hypopholis</i> * |
| (iii) Pronotum posterior closely adjacent to elytral base | Pronotum small and not closely adjacent to elytra (<i>i.e.</i> gap in between) |
| (iv) Mesosternum with a long process | Mesosternum simple |

*Character (ii) does not differentiate between *Hypopholis* and *Pegylis*, and it is thus an invalid character at the generic level. However, it is included here as done by Erichson (1847) for discussion purposes.

tenuitarsis + *Hypopholis uelensis* + *Hypopholis vittata* (*Pegylis maculipennis* + *Pegylis vestita*) (*Pegylis burgeoni* + *Pegylis hauseri*) (((*Pegylis gestroi* + *Pegylis rufolineata*) (*Pegylis ertli* + *Pegylis rufomaculatus*)) (*Pegylis kenyensis* + *Pegylis morio*))) = Pegylina clade

Four synapomorphies (as listed above and indicated (•) in Fig. 1) support the monophyly of the Pegylina clade. This analysis thus supports Arrow's (1943) previous synonymy of *Pegylidius* with *Pegylis*, the more recent synonymy of *Adoretopsis* with *Pegylis* (Lacroix 2010). Results provide evidence that *Hypopholis* is synonymous with *Pegylis*, and that *Pegylis* and *Eupegylis* are closely related (Fig. 1).

Validity of *Eupegylis* and *Pegylis* as genera

Node: ((((*Eupegylis confusa* ... *Pegylis morio*))) = Pegylina clade

Three synapomorphies support the validity of the genus *Eupegylis*, *i.e.* anterior margin of pronotum bordered by a slight membrane (C6:S2; S = 2; ci = 100; ri = 100); aedeagus laterally produced in profile (C16:S3; S = 13; ci = 23; ri = 33); aedeagus symmetrical and complex (including setae) (C18:S3; S = 21; ci = 28; ri = 63). Consequently, *Eupegylis* is retained as a valid genus due to the support of these synapomorphies (Fig. 1).

Four synapomorphies support the validity of the genus *Pegylis*, *i.e.* anterior margin of pronotum bordered by a membrane (C6:S1; S = 2; ci = 100; ri = 100); elytra bi-coloured (C10:S3; S = 16; ci = 18; ri = 31); elytra slightly mottled (C11:S1; S = 13; ci = 15; ri = 47); median depression on abdominal sternites (of male) present (C14:S1; S = 2; ci = 50; ri = 85). Of these characters, the anterior margin of the pronotum bordered by a membrane provides strong support (ci = 100; ri = 100) for the Pegylina clade and a synapomorphy for all species of *Pegylis*, with the other Pegylina genus, *Eupegylis*, also having this membrane (but being less developed).

An assessment of Erichson's (1847) characters used to differentiate *Hypopholis* from *Pegylis* and an evaluation of the characters used by Lacroix (1989) to define the subtribe Pegylina

Node: ((((*Eupegylis confusa* ... *Pegylis morio*))) = Pegylina clade

Erichson (1847) described both *Hypopholis* and *Pegylis* listing the characters provided in Table 3 to differentiate between them. To represent these genera he placed two South African species in *Hypopholis*, *i.e.* *Leucopholis sommeri* Dejean¹ and *Leucopholis vittata* Boheman², and one from 'Sennaar' in the Sudan in *Pegylis*, *i.e.* *Melolontha morio* Kolbe (in litt.)³.

Here follows a discussion of characters used by Erichson (1847) and previous authors to differentiate *Hypopholis* from *Pegylis* in order to reassess their validity as generic characters. Additionally, the characters used by Lacroix (1989) to define the subtribe Pegylina are included in the discussion.

(i) Antennae (character 4)

Erichson (1847) used 'antennae 10-segmented with a 3-segmented club' versus 'antennae 8-segmented with a 3-segmented club' to distinguish *Hypopholis* from *Pegylis*. In his generic description of *Pegylidius* Péringuey (1904) noted the antennal number as 'antennae 10-jointed'. In 1943, Arrow commented that:

In *P. mashunus* and *P. pondoensis* the antennae have all the joints of the footstalk separate, while in most of the species the males have them more or less fused together, but I consider this also useless for a generic separation, as applying to one sex only.

Consequently, Arrow (1943) synonymized *Pegylidius* with *Pegylis* mentioning that the num-

¹*Pegylis sommeri* (Burmeister, 1855) comb. n.

²*Pegylis vittata* (Fähræus in Boheman, 1857) comb. n.

³*Pegylis morio* Blanchard, 1850.

ber of antennal segments is an invalid generic character in this particular instance. In his characterization of the Pegylini Lacroix (1989) included a three-segmented antennal club for *Hypopholis*, *Pegylyis* and *Stenopegylyis*, but *Eupegylyis* with a 5–6 segmented antennal club.

The number of antennal segments has historically been used as a generic character in various phytophagous scarabs. However, a recent study (Ahrens & Vogler 2008) has shown that it is too variable to be a reliable generic character. In the analysis, the number of antennomeres (C4) was coded following Ahrens (2005) using only two states, *i.e.* (0) nine or 10 antennomeres and (1) eight or fewer antennomeres. Thus, character 4 is two steps in length on the SCT (Fig. 1), with a $ci = 50$ and $ri = 66$. The second state is found only supporting the following subclades (*Pegylyis maculipennis* + *Pegylyis vestita*) and (*Pegylyis kenyensis* + *Pegylyis morio*) on the SCT. Harrison (2014) provides clear illustrations in Figs 13–15 of the 10-segmented antennae in specimens of the following species (*P. sommeri*, *P. pondoensis* and *P. vittata*) examined here. Thus, although it is quite possible to have individuals or even populations with a reduced number of antennal segments, this character is not regarded here as valid to separate *Pegylyidius* from *Pegylyis*, nor *Hypopholis* from *Pegylyis*, at the generic level.

(ii) Labrum (uninformative character)

Erichson (1847) described the labrum of *Hypopholis* as ‘... large, deeply split/incised’, while that of *Pegylyis* as ‘labrum as in *Hypopholis*’, suggesting no morphological difference between these genera based on the labrum. Lacroix (1989) described the labrum of the subtribe Pegylyina taxa to be elongate and bilobed.

During selection of characters to code for the analysis the labrum of all available taxa (37 species; $n = 5$) were examined and other than morphological differences at species level, *i.e.* varying depths of the incision possibly related to intraspecific diets, no characters differentiating at the generic level were observed, and thus the labrum was excluded from the analysis when the data matrix was ‘mopped’ in WinClada to remove uninformative characters. Consequently, the shape of the labrum is also regarded here as an invalid generic character to differentiate *Hypopholis* from *Pegylyis*.

(iii) Protarsal claws (character 3)

Gerstaecker (1867) appears to be the earliest

author to comment on the protarsal claw morphology of *Hypopholis* and *Pegylyis* and noted the difference between *H. sommeri* and *H. vittata* as:

...the shape of the tarsal claws ... in *H. sommeri* are in the middle very strongly dentate, [while] in *H. vittata* the posterior tibial claws are simple; the protibial claws with a simple inner [claw] and split external [claw].

Gerstaecker (1873) added to this by noting the differing tarsal claw morphology between *H. conspurcata*, *H. sommeri* and *H. vittata* as:

It is intriguing that [of] the three species that are known to date, all show distinctly/unusually shaped claws: (i) all claws apically deeply split: *H. conspurcata*; (ii) all claws medially strongly dentate [toothed]: *H. sommeri*; (iii) meso- and metatarsal claws simple [unmodified]; protarsal inner claw simple, outer claw split: *H. vittata*...

Note that of the three species discussed above, one is subsequently transferred to *Pegylyis* and the other two retained in *Hypopholis* (see Appendix 1).

In the analysis, male protarsal claws were scored as symmetrical (0) or asymmetrical (1), and this character failed to contribute differentiation at the generic level among the taxa of Pegylyina included in the analysis (C3; $S = 5$; $ci = 20$; $ri = 20$). However, it did provide interspecific support for these species: *Pegylyis lindiana*, *P. pondoensis*, *Hypopholis sommeri*, and the sub-clade *P. ertli* + *P. rufomaculatus* (Fig. 1).

Arrow (1901) discussed the undesirability of using sexually dimorphic characters on which to base genera, and provided examples from the Scarabaeidae: Rutelinae to illustrate this. Péringuey (1904) described *Pegylyidius* as a new genus for a Zimbabwean species (*P. mashunus*) and remarked that, ‘plainly [it is] allied to *Hypopholis*, but differing mainly by the structure of the claws and simple mesosternum.’ Arrow (1943) in his discussion of *Pegylyis* and *Pegylyidius* concluded that, ‘It is evident for generic purposes the claws are useless and I therefore regard *Pegylyidius* as a synonym of *Pegylyis*’. Having examined most of the described species of *Pegylyis* (see * in Appendix 1), it is apparent that the protarsal claws are sexually dimorphic, inter- and intraspecifically variable and thus this character is also regarded here, for these taxa, as invalid on which to base genera.

(iv) Absence of a protibial spur (character 1)

Kolbe (1894) in his *Synopsis of the Genera of the Leucopholines from the Ethiopian Region* differentiated *Pegylyis* and *Hypopholis* from five other genera

which he described as new by 'protibia without spurs'. I am not aware of any earlier authors specifically noting this character in these genera.

The complete absence of a protibial spur in both sexes is a key character used by Lacroix (1989) to define the subtribe Pegylina. However, this character (C1:S1) is not unique to the Pegylina. Examples of other melolonthine genera that lack, or have reduced, protibial spurs include the following⁴: the monotypic *Oncerus floralis* LeConte, 1856, which is one of two monotypic genera in the tribe Oncerini LeConte (A.V. Evans, pers. comm.); the tribe Podolasiini Howden, 1997 comprising *Podolasia* Harold and *Podostena* Howden (A.V. Evans, pers. comm.) are both without a protarsal spur, as illustrated in Howden (1954: figs 1–5); *Chaunocolus* Saylor (monotypic) and *Chmaunanthus* Burmeister (comprising about three species) (formerly Chasmatopterini (Evans 2003)), recently placed by Evans & Smith (2009) in genera 'incertae sedis', also see Baraud & Branco (1990) and Branco (2004) for discussion on *Chasmatopterus* Dejean; *Ceratogonia* Kolbe (monotypic) from the tribe Diplotaxini Burmeister (Bezděk 2004a,b). In addition, many New World Macroductylini do not possess a protibial spur (Smith 2008), examples include, *Ampliodactylus* Smith, *Pusiiodactylus* Smith, *Macroductylus* Dejean and some *Pristerophora* Harold. This character is variable within the Macroductylini (A.B.T. Smith, pers. comm.) and Katovich (2008) provided a partial list of some taxa where this character is variably expressed (see Katovich's character 54 (pp. 10, 76)). Within the Melolonthini: Rhizotrogina, *Brachyllus* Brenske is a genus lacking a protibial spur (Keith 2003, 2005, 2007). Some *Eulasia* (*Rudeulasia*) Baraud (Glaphyridae) lack the spur at least in males, whereas most species have one, but D. Keith (pers. comm.) found that in these species it is more often vestigial rather than completely absent. Finally, members of the scarab subfamily Aclopininae Blanchard, have no protibial spurs (F.C. Ocampo, pers. comm.).

Character 1, state 1 (= protibial spur absent) occurs as a homoplasious character state change for *Stenopegyilis cylindrica* and for the Pegylina clade (Fig. 1), with character 1, state 2 (= protibial spur residual) as a non-homoplasious character state change (synapomorphy) for the genus *Psilonychus* (Fig. 1).

As indicated by the analysis and discussion

⁴These examples were provided by Scarab-L list server members, and consequently (pers. comm.) is used extensively here to acknowledge their source.

above, the complete absence of protibial spurs in both sexes of *Pegylis* is a homoplasious character state change for *Pegylis* and *Eupegyilis*, while being a synapomorphy for the two species of *Psilonychus* included in this analysis.

(v) Pronotum (characters 6, 7, 8 & 9)

Erichson (1847) used 'posterior pronotum closely adjacent to elytral base' for *Hypopholis versus* 'pronotum small and not closely adjacent to elytra' [separated by a gap] for *Pegylis*. The relative position of the posterior section of the pronotum in relation to the elytral base is entirely dependent on how the specimen being examined died, was mounted and dried. This is because in live specimens there is ample articulation between these two body sections. Thus, I do not regard this character as justifiable to separate *Hypopholis* from *Pegylis* at the generic level.

Character 6. Anterior margin of pronotum bordered by a membrane: (0) absent; (1) present; (2) present, but slight. S = 2; ci = 100; ri = 100. Character 6, state 2 represents a non-homoplasious character state change for *Eupegyilis confusa* (Fig. 1). Lacroix (1989) included the anterior margin of the pronotum bordered by a membrane as a defining feature for the Pegylina. Character 6, state 1 represents a non-homoplasious character state change (synapomorphy) for the Pegylina clade.

Character 7. Anterior margin (dorsal section) of pronotum: (0) defined by a groove continuous from side to side; (1) defined by a groove only on each side towards the angles; (2) groove absent; (3) notched on each side towards the angle. S = 11; ci = 27; ri = 50. Character 7, state 0 represents a homoplasious character state change for *Brachylepis elephas* and the Pegylina clade. Character 7, state 1 represents a homoplasious character state change for *Hypopholis conspurcata*, *Pegylis microchaeta*, *P. pondoensis*, *H. sommeri*, *Adoretopsis tenuitarsis* and *H. uelensis*. Character 7, state 3 represents a non-homoplasious character state change for *Stenopegyilis cylindrica*.

Character 8. Lateral margin of pronotum: (0) smooth; (1) finely serrated; (2) coarsely serrated. S = 3; ci = 33; ri = 66. Character 8, state 0 represent homoplasious character state changes for *Rhabdopholis albostrigata* and the Pegylina clade in the analysis (Fig. 1). Character 8, state 1 represent homoplasious character state changes for *Eupegyilis confusa*.

Character 9. Pronotum surface: (0) uniformly

clothed with short fine setae; (1) uniformly clothed with long fine setae; (2) scaliferous; (3) no or few setae/scales; (4) uniformly clothed with short and long fine setae. $S = 9$; $ci = 44$; $ri = 61$. Character 9, state 0 represents a homoplasious character state change for the *Pegylina* clade. Character 9, state 1 represents a homoplasious character state change for *Pegylis giraudetae*, *P. pilosa* and *P. vestita*. Character 9, state 4 represents a non-homoplasious character state change for *Stenopegylis cylindrica*.

(vi) Mesosternum process (character 12) and mesosternum (character 13)

Erichson (1847) indicated that *Hypopholis* has a mesosternum with a long process, as opposed to a simple mesosternum in *Pegylis*. The presence or absence of a mesosternal process appears to occur at specific rather than at generic level within the *Pegylina*, as species separated from *Hypopholis* based on this character alone, are equally placed into *Pegylis* using other characters. The presence of a mesosternal process (keel or peg-like projection) is an obvious character used to separate *Hypopholis* from *Pegylis*, and historically any species with such a process was placed in *Hypopholis* and those without it into *Pegylis*. A brief overview of the presence of a mesosternal process in some other scarab taxa is provided below.

Melolonthinae and Rutelinae: in combination with other characters, the presence/lack of mesosternal process was/is used to differentiate the following generic couples: *Melolontha* Fabricius from *Hoplosternus* Blanchard (now regarded as a synonym of *Melolontha* (see Bunalski 2002)), *Leucopholis* Blanchard from *Lepidiota* Hope (D. Keith, pers. comm.), and *Mimela* Kirby from *Anomala* Samouelle (Kim 1996). In the endemic Australian genus *Sciton* Blackburn (Melolonthinae: Liparetrini), almost all species belonging to this genus possess a mesosternal projection, ranging from a long setose peg to a short peg, others with a longitudinal keel of varying height to none at all (A. Szitó, pers. comm.). Within *Sciton*, A. Szitó (pers. comm.; presently revising this genus) does not consider it to be a generic character. A. Bezděk (pers. comm.) is not aware of a mesosternal keel in any diplotaxine species, thus it appears to be absent in the Diplotaxini. All known species of the following genera (*Proagosternus* Blanchard; *Pseudodicrania* Gutiérrez (Evans 2003); *Rhabdopholis* Burmeister (Harrison 2004) have some degree of a mesosternal keel.

Many rutelines have a well-developed meso-metasternal process which is suspected to be for flight muscle attachment, as strong fliers often have a well-developed process, but specimens would need to be dissected to show that this is the case (M.L. Jameson, pers. comm.). For example, a mesosternal process is observed in several species of *Parastasia* (Kuijten 1992). In *Parastasia*, the presence of the process is not a generic character, but does discriminate a few species from others within the genus (A. Bezděk, pers. comm.).

Cetoniinae: within the cetoniines many genera and species have a mesosternal process (see Holm & Marais 1992). Several Ethiopian Trichiinae (*sensu* Krikken 1984) genera bear a noticeable mesosternal projection (E. Ricchiardi, pers. comm.). Within a wide selection of Cremastocheilini examined by G. Mynhardt during research for Mynhardt & Wenzel (2010), the presence of a mesosternal projection was variably apparent, but always present (Mynhardt, pers. comm.).

Dynastinae: many genera (or parts of genera) of dynastines have a prosternal peg, the shape of which is used as a character in some of Endrödi's (1985) keys, for example *Peltonotus* Burmeister (Jameson & Wada 2004, 2009).

Scarabaeinae: the presence of a well-developed spiniform prosternal projection is one of the defining characters of the genus *Oxysternon* (Phanaeini) (Edmonds & Zidek 2004), whereas this same feature is less well developed in a related genus *Phanaeus* (Philips *et al.* 2004; Price 2007, 2009).

Phaenomeridinae: a mesosternal projection is found in the genus *Phaenomeris* Hope (Phaenomerinae) (A. Ballerio, pers. comm.).

B. Warner's (pers. comm.) response to the scarab list provides an informative summary of a pro/mesosternal projection, and thus I include it here.

Absent in many smaller scarabs. Actually, they are commonplace in many scarabaeoid groups, and fairly common in Coleoptera. No doubt there have been many independent evolutions of such pegs, as they are the simple answer to stiffen prothoracic/mesothoracic flexion, or head (mentum in cremastocheilines)/prothoracic flexion, and act like a 'door stop.' The standard locations are anterior to procoxae, posterior to procoxae, and protruding from the mesometasternal junction. As for use as a generic character, *Mimela* vs. *Anomala* (and relatives) is an example. However, in many genera pegs may range from absent to large. Such pegs are generally not used at higher taxonomic categories given frequent parallelisms and variable expression.

Character 12. Mesosternum (as a mesosternal keel): (0) not produced; (1) weakly produced, not surpassing the base of mesocoxae; (2) produced anteriorly beyond base of the mesocoxae. $S = 14$; $ci = 14$; $ri = 25$ (Jameson 1997: Char 110). Character 12, state 0 represents a homoplasious character state change for *Stenopegyilis cylindrica*, *Psilonychus eckloni* + *Psilonychus duponti*, *Pegylis angolensis*, *P. bennigseni*, *Pegylidius mashunus*, *P. neumanni*, *P. pilosa*, *P. maculipennis* + *P. vestita*, *P. burgeoni* + *P. hauseri*, *P. gestroi* and *P. kenyensis* + *P. morio*. Character 12, state 2 represents a homoplasious character state change for *Rhabdopholis albostrata*, *Hypopholis sommeri* and *H. uelensis*.

Character 13. Mesosternum: (0) without rim; (1) with rim. $S = 8$; $ci = 12$; $ri = 12$. Character 13, state 0 represents a homoplasious character state change for *Rhabdopholis albostrata*, *Stenopegyilis cylindrica*, *Psilonychus duponti* + *Psilonychus eckloni*, *Eupegyilis confusa*, *Pegylis pondoensis*, *Hypopholis sommeri*, *Hypopholis uelensis* and *Hypopholis vittata*.

Consequently, mesosternal characters and especially the presence of a mesosternal keel/process is not a homology (synapomorphy) for *Hypopholis* or any other genera included in this analysis. Therefore, the presence or absence of a mesosternal process is regarded as an invalid character to base genera on within the Pegylina.

(vii) Body shape (too subjective to score)

Lacroix (1989) includes the body shape (body slightly elongated, rounded or oval) in his description of the Pegylina. It is true that all genera of Pegylina share this 'streamlined' body shape, but so do many other genera of melolonthines, for example, *Eulepida*, *Lepidiota*, *Leucopholis* and *Rhabdopholis*. Consequently, it is regarded as an invalid character for tribal/subtribal level discrimination, but is obviously valid as part of a habitus description for taxa within a respective taxonomic group.

(viii) Sexual dimorphism (characters 3, 14)

For the Pegylini Lacroix (1989) defines sexual dimorphism as limited or undeveloped. Within the Scarabaeoidea sexual dimorphism varies from quite marked to almost indistinguishable. Large horned Cetoniinae, Dynastinae and Scarabaeinae males can differ so much from their conspecific females that they can and have been regarded as different species. However, once associated via locality data or if collected in copula, the sexes of

conspecifics are easily separated macroscopically. Minor males of the above taxa merge into the female body form, and are thus our exception to this rule-of-thumb. However, experienced taxonomists quickly learn to differentiate the sexes of the groups on which they work. Male Pegylina have a distinctive median-lateral groove on their ventral sternites, slightly longer antennal clubs, and different tarsal claws compared to females.

Apart from the form of the male genitalia two additional gender-related characters were included in the analysis, *i.e.* C3 and C14.

Character 3. Protarsal claws (in males): (0) symmetrical; (1) asymmetrical. $S = 5$; $ci = 20$; $ri = 20$. The morphology of the male protarsal claws is covered under point (iii) above and only provided interspecific support.

Character 14. Median depression on abdominal sternites (of male): (0) absent; (1) present. $S = 2$; $ci = 50$; $ri = 85$. Character 14, state 0 represents a homoplasious character state change for *Eupegyilis confusa*, and character 14, state 1 represents a non-homoplasious character state change (synapomorphy) for the subtribe Pegylina (Fig. 1).

Synonymy of *Wernerophylla* Lacroix, 2001 with *Stenopegyilis* Arrow, 1943

The variable placement of one species (*Stenopegyilis cylindrica* Arrow, 1943) in two different tribes/subtribes within the Melolonthinae indicates the value of phylogenetic analyses as a tool to understand relationships between taxa within taxonomic hierarchies. *A priori* taxa selection for this study warranted no rationale to include *Wernerophylla nigra* Lacroix, 2001 in the analysis or discussion. However, during examination of taxa for the study and wider literature on African melolonthines it became apparent that *W. nigra* (Pachydemini) is conspecific with *S. cylindrica* (Melolonthini). Therefore, their synonymy follows below.

Stenopegyilis Arrow, 1943

Stenopegyilis Arrow, 1943: 782–783. Lacroix 2010: 122–123.

cylindrica Arrow, 1943: 783–784. Lacroix 2010: 122–123.

Wernerophylla Lacroix, 2001

Wernerophylla Lacroix, 2001: 180–181. Lacroix 2007: 23–25, 211–212 (key), 223–225, 337, 397, 399 (Pachydemini catalogue). **Syn. n.**

nigra Lacroix, 2001: 181–183; Lacroix 2007: 224–225, 337. **Syn. n.**

Remarks. Arrow (1943) described *Stenopegylis cylindrica* from seven male and three female specimens from: Nyasaland, Mlanje, Dec. 1913, leg. Dr S.A. Neave. In a study of the Pachydemini of Kenya and Tanzania, Lacroix (2001) described *Wernerophylla nigra* as a monotypic genus using 11 male and two female specimens from 'Tanzania, Ruvuma Pr., near Songea, 9–13.XII.1997, leg. Werner & Lizler'. I have examined Arrow's type series of *S. cylindrica* and specimens of the same locality series collected by Werner and Lizler used by Lacroix to describe *W. nigra* and there is no doubt that these taxa are conspecific in all respects. Lacroix (2007) includes *W. nigra* in his first catalogue of the genera and species of African Pachydemini, while Arrow's *S. cylindrica* is included in his second catalogue covering the non-Pachydemini genera and species of African Melolonthinae.

Nomenclatural changes resulting from the analysis and character discussion

The phylogenetic analysis aimed to resolve the relationship of species in the following genera: *Adoretopsis*, *Hypopholis*, *Pegylis* and *Pegylidius* and further to investigate the validity of the subtribe Pegylina which, prior to this study, included the following genera: *Eupegylis*, *Hypopholis*, *Pegylis* and *Stenopegylis*. In the analysis, four synapomorphies support the monophyly of the subtribe Pegylina, but only if the monotypic *Stenopegylis* is transferred to the Melolonthini *sensu lato*. Thus, to maintain a monophyletic Pegylina the genus *Stenopegylis* is removed from the subtribe Pegylina and moved to the Melolonthini *sensu lato*.

Priority of *Pegylis* over *Hypopholis*

Erichson (1847: 657) first mentioned *Hypopholis* followed by *Pegylis* in a key to new Melolonthini, and then again in the works (Erichson 1847) index. One could argue that as *Hypopholis* appears just before *Pegylis* it has line-priority over *Pegylis*, but from a page-priority perspective they appear on

the same page. Furthermore, to-date five species have been described in *Hypopholis*, while 30 species have been described in *Pegylis*, with the remaining two species originally being placed in *Adoretopsis* or *Pegylidius* (Appendix 1 provides a checklist). Thus, to correct their taxonomy, but at the same time maintain nomenclatural stability *Pegylis* is regarded as having been more widely used in the literature than *Hypopholis*, and thus *Pegylis* takes priority over *Hypopholis*.

The analysis and discussion provided above indicates that there is no justification for retaining *Hypopholis* as a genus, and thus *Hypopholis* is synonymized with *Pegylis* here, which introduces the following new combinations:

Pegylis sommeri (Burmeister, 1855) **comb. n.**;

P. uelensis (Burgeon, 1946) **comb. n.**,

P. vittata (Fåhraeus in Boheman, 1857) **comb. n.**

A checklist of all the current taxa in the subtribe Pegylina is provided as Appendix 1. The newly reconstituted Pegylina includes two genera, *i.e.* *Eupegylis* Duvivier, 1892 (1 sp.) and *Pegylis* Erichson, 1847 (36 spp).

ACKNOWLEDGEMENTS

I thank all of the following: curators from the museums listed in Harrison (2014) for access to specimens and especially for the loan (M. Kerley BMNH) and passage (M. Krüger TMSA) of the type series of *Stenopegylis*; Paul Schoolmeesters, M.L. Jameson and D. Keith for literature; A. Ballerio, A. Bezděk, A. Szitó, A.V. Evans, W. Warner, C.P.T.D. Gillett, D. Keith, E. Ricchiardi, G. Mynhardt and M.L. Jameson for information on the presence of a mesosternal process in other scarab taxa; M.J. Wingfield and C.H. Scholtz (both UP) provided NRF and Mellon Foundation funding, respectively; K. Balkwill's (WITS) writing retreats to the Jackson Field Station at WITS' Pullen Nature Reserve for writing time; and G. Goodman-Cron (WITS) and three anonymous reviewers for insightful comments that improved the paper.

REFERENCES

- AHRENS, D. 2005. The phylogeny of Sericini and their position within the Scarabaeidae based on morphological characters. *Systematic Entomology* 31: 113–144.
- AHRENS, D. & VOGLER, A.P. 2008. Towards the phylogeny of chafers (Sericini): Analysis of alignment-variable sequences and the evolution of segment numbers in the antennal club. *Molecular Phylogenetics and Evolution* 47: 783–798.
- ARROW, G.J. 1901. LI. – Remarks on secondary sexual differences in rutelid Coleoptera, with descriptions of some new forms. *Annals and Magazine of Natural History* 7(7): 393–401.
- ARROW, G.J. 1902. XIX. – On rutelid and melolonthid beetles from Mashonaland and East Africa. *Annals and Magazine of Natural History* 9(7): 89–101.
- ARROW, G.J. 1943. LXVI. – Systematic notes on the

- melolonthine beetles belonging to the genus *Lepidiota* and some related genera. *Annals and Magazine of Natural History* 10(11): 773–785.
- BARAUD, J. & BRANCO, T. 1990. Révision des *Chasmatopterus* Latreille, 1825. *Coleopterological Monographs* 1: 9–55.
- BEZDĚK, A. 2004a. Catalogue of Diplotaxini of the Old World. *Zootaxa* 463: 1–90.
- BEZDĚK, A. 2004b. Revision of the genus *Ceratogonia* Kolbe, 1899. *Annales Zoologici* 54(4): 797–801.
- BOUCHARD, P., BOUSQUET, Y., DAVIES, A.E., ALONSO-ZARAZAGA, M.A., LAWRENCE, J.F., LYAL, C.H.C., NEWTON, A.F., REID, C.A.M., SCHMITT, M., SLIPINSKI, S.A. & SMITH, A.B.T. 2011. Family-group names in Coleoptera. *Zookeys* 88: 1–972.
- BRANCO, T. 2004. Description of a new species of *Chasmatopterus* Dejean, 1821, from Portugal, with a note on the subfamily Chasmatopterinae (Coleoptera: Melolonthidae). *Elytron* 17–18: 27–36.
- BUNALSKI, M. 2002. Melolonthidae (Coleoptera: Scarabaeoidea) of the Palearctic and Oriental Regions. I. Taxonomic remarks on some genera of Melolonthinae. *Polskie Pismo Entomologiczne* 71(4): 401–413.
- BURGEON, L. 1946. Melolonthini et Pachydemini du Congo belge (suite) (Coleopt. Scarab. Melolonthinae). *Revue de Zoologie et de Botanique africains, Bruxelles* 39(4): 339–366.
- BURMEISTER, H. 1855. *Handbuch der Entomologie*, Berlin. 4(2): 1–569.
- COCA-ABIA, M.M. 2007. Phylogenetic relationships of the subfamily Melolonthinae. *Insect Systematics & Evolution* 38(4): 447–472.
- DUVIVIER, A. 1946. Mélanges Entomologiques. X. Diagnoses de Coléoptères nouveaux du Congo. *Annales de la Société Entomologique de Belgique* 36: 56–60.
- EDMONDS, W.D. & ZÍDEK, J. 2004. Revision of the Neotropical dung beetle genus *Oxysternon* (Scarabaeidae: Scarabaeinae: Phanaeini). *Folia Heyrovskyana Supplementum* 11: 1–58.
- ENDRÖDI, S. 1985. *The Dynastinae of the world*. Dr. W. Junk Publishers, The Hague as volume 28 in the Series Entomologica, Budapest, pp. 1–800; plates I–XLVI.
- ERICHSON, W.F. 1847. *Naturgeschichte des Insecten Deutschland. I. Coleoptera, Scarabaeides*. Nicolaische Buchhandlung, Berlin. 1(3) parts 4–5: 481–800.
- EVANS, A.V. 2003. A checklist of the New World chafers. *Zootaxa* 211: 1–458.
- EVANS, A.V. & SMITH, A.B.T. 2009. *An electronic checklist of the New World Chafers (Coleoptera: Scarabaeidae: Melolonthinae)*. Version 3. Electronically published, Ottawa, Canada.
- FÄHRHRAEUS in BOHEMAN, C.H. 1857. *Insecta Caffrariae annis 1838–1845 a J.A. Wahlberg collecta, Coleoptera, Holmiae*. 2: 1–395 + 1 plate.
- GERSTAECKER, C.E.A. 1867. Beitrag zur Insekten-fauna von Zanzibar, nach dem während der Expedition des Baron v.d. Decken gesammelten Material zusammengestellt. *Archiv für Naturgeschichte, Berlin* 33(1): 1–49.
- GERSTAECKER, C.E.A. 1873. *Die Gliedertier-Fauna des Sansibar-Gebietes* Decken C.C. Baron Carl Claus von der Decken's Reisen in Ost Afrikas, Berlin. 1–542.
- GOLOBOFF, P.A., 1997. NONA Version 2.0 (for Windows). Computer software and documentation. Published by the author, Instituto Miguel Lillo, Miguel Lillo 205, 400 Sierra Madre de Tucuman, Argentina.
- HARDY, C.R., LIPSCOMB, D.L., NIXON, K.C. & OCHOTERENA, H. 2003. *WinClada ver. 1.0, a basic user's manual*. Available upon request from Kevin C. Nixon, Bailey Hortorium, 462 Mann Library, Cornell University, Ithaca, NY 14853, U.S.A. 1–29.
- HARRISON, J. du G. 2004. Revision of the endemic southern African genus *Rhabdopholis* Burmeister, 1855 (Coleoptera: Scarabaeidae: Melolonthinae). *African Entomology* 12(1): 39–54.
- HARRISON, J. du G. 2009. A taxonomic revision of the African leaf chafer genus *Asthenopholis* Brenske, 1898 (Coleoptera: Scarabaeidae: Melolonthinae): a SEM study. *Zootaxa* 2225: 1–48.
- HARRISON, J. du G. 2014. Review of the South African species of *Pegylis* Erichson, 1847 (Coleoptera: Scarabaeidae: Melolonthinae) commonly known as large wattle chafers. *African Entomology* 22: 685–713.
- HOLM, E. & MARAIS, E. 1992. *Fruit Chafers of Southern Africa (Scarabaeidae: Cetoniini)*. Ekogilde, Hartebeespoort, Pretoria, South Africa. 1–326 + 32 plates.
- HOWDEN, H.F. 1954. A review of the genus *Podolasia* Harold (Coleoptera: Scarabaeidae). *American Museum Novitates* 1661: 1–11.
- HOWDEN, H.F. 1997. Podolasiini, new tribe, and a revision of the included genera, *Podolasia* Harold and *Podostena* Howden, new genus (Coleoptera: Scarabaeidae: Melolonthinae). *The Coleopterists Bulletin* 51(3): 223–255.
- JAMESON, M.L. 1997. Phylogenetic analysis of the sub-tribe Rutelina and revision of the *Rutela* generic groups. *Bulletin of the University of Nebraska State Museum* 14: 1–184.
- JAMESON, M.L. & WADA, K. 2004. Revision of the genus *Peltonotus* Burmeister (Coleoptera: Scarabaeidae: Dynastinae) from South-eastern Asia. *Zootaxa* 502: 1–66.
- JAMESON, M.L. & WADA, K. 2009. Five new species of *Peltonotus* Burmeister (Scarabaeidae: Dynastinae: Cyclocephalini) from Southeast Asia. *Insecta Mundi* 0102: 1–16.
- KATOVICH, K. 2008. A generic-level phylogenetic review of the Macroductylini (Coleoptera: Scarabaeidae: Melolonthinae). *Insecta Mundi* 23: 1–78.
- KEITH, D. 2003. Le genre *Brachyllus* Brenske, 1896 en Chine. Nouvelles acquisitions pour la faune chinoise (Coleoptera, Melolonthidae). *Bulletin de la Société Entomologique de France* 108(3): 307–311.
- KEITH, D. 2005. Remarque's taxonomiques sur quelques Rhizotroginae orientaux et description de nouvelles espèces (Col. Scarabaeoidea Melolonthidae). *Symbioses* 12: 23–32.
- KEITH, D. 2007. Trios nouvelles espèces du genre *Brachyllus* Brenske, 1896 (Insecta: Coleoptera: Scarabaeoidea, Melolonthidae). In: Hartmann, M. & Weipert, J. *Biodiversität und Naturlausstattung im Himalaya II*: 419–424.

- KIM, J.I. 1996. Taxonomic study of Korean Rutelidae (Coleoptera) IV. Genus *Mimela*. *Korean Journal of Entomology* **26**(3): 235–242.
- KOLBE, H.J. 1894. Beiträge zur Kenntniss der Melolonthiden. I. Uebersicht der Melolonthinen, Leucopholinen und Schizonychen Africas. *Annales de la Société Entomologique de Belgique* **38**(10): 548–577.
- KRIKKEN, J. 1984. A new key to the suprageneric taxa in the beetle family Cetoniidae. *Zoologische Verhandlungen Leiden* **210**: 1–75.
- KUIJTEN, P.J. 1992. A revision of the genus *Parastasia* in the Indo-Australian Region (Coleoptera: Scarabaeidae: Rutelinae). *Zoologische Verhandlungen Leiden* **275**: 1–176.
- LACROIX, M. 1989. Insectes Coléoptères Melolonthidae (1^{re} partie). Muséum National d'Histoire Naturelle, Paris. *Faune de Madagascar* **73**(1): 1–302 + 785 figs.
- LACROIX, M. 2001. Pachydeminae de l'Est Africain (Kenya et Tanzanie) (Coleoptera, Melolonthidae). *Coléoptères* **7**(13): 173–226.
- LACROIX, M. 2007. *Pachydeminae du Monde* (Scarabaeoidea, Melolonthidae). *Genera et Catalogue Commenté*. Collection Hannetons, Lacroix, Paris, France. 1–450 + 264 plates.
- LACROIX, M. 2010. *Melolonthinae afrotropicaux* (Coleoptera, Melolonthidae) *Genera et Catalogue Commenté*. 1–277 + 140 plates & figures + 52 colour plates. Editions M., Lacroix, Paris, France.
- MYNHARDT, G. & WENZEL, J.W. 2010. Phylogenetic analysis of the myrmecophilous *Cremastocheilus* Knoch (Coleoptera, Scarabaeidae, Cetoniinae), based on external adult morphology. *ZooKeys* **34**: 129–140.
- NIXON, K.C. 2002. WinClada ver. 1.00.08. Published by the author, Ithaca, New York, U.S.A.
- NIXON, K.C. & CARPENTER, J.M., 1993. On outgroups. *Cladistics* **9**: 413–426.
- PÉRINGUEY, L. 1904. Descriptive catalogue of the Coleoptera of South Africa (Lucanidae and Scarabaeidae). *Transactions of the South African Philosophical Society* **13**: 1–293 + 4 plates.
- PHILIPS, T.K., EDMONDS, W.D. & SCHOLTZ, C.H. 2004. A phylogenetic analysis of the New World tribe Phanaeini (Coleoptera: Scarabaeidae: Scarabaeinae): hypotheses on relationships and origins. *Insect Systematics & Evolution* **35**(1): 43–63.
- PRICE, D.L. 2007. A phylogenetic analysis of the dung beetle genus *Phanaeus* (Coleoptera: Scarabaeidae) based on morphological data. *Insect Systematics & Evolution* **38**(1): 1–18.
- PRICE, D.L. 2009. Phylogeny and biogeography of the dung beetle genus *Phanaeus* (Coleoptera: Scarabaeidae). *Systematic Entomology* **34**(1): 137–150.
- SMITH, A.B.T. 2006. A review of the family-group names for the Superfamily Scarabaeoidea (Coleoptera) with corrections to nomenclature and a current classification. *Coleopterists Society Monograph* **5**: 144–204.
- SMITH, A.B.T. 2008. South American Melolonthinae classification and nomenclature: some problems and solutions. *Insecta Mundi* **60**: 1–28.

Appendix 1. Revised checklist of the genera and species of Melolonthini: Pegylina Lacroix, 1989, following nomenclatural changes implemented here. An asterisk indicates taxa included in the phylogenetic study.

Genus *Eupegylis* Duvivier, 1892 (1 sp.)

confusa Duvivier, 1892*

Genus *Pegylis* Erichson, 1847 (35 spp.)

= *Adoretopsis* Fairmaire, 1887

= *Hypopholis* Erichson, 1847 (**syn. n.**)

= *Pegylidius* Péringuey, 1904

Type species: *Pegylis morio* Blanchard, 1850*

angolensis Moser, 1915*

bennigseni Brenske, 1898*

burgeoni Decelle, 1968*

conspurcata (Gerstaecker, 1867 – *Hypopholis*)*

ertli Moser, 1919*

gestroi Brenske, 1895*

giraudetae Decelle, 1968*

gracilis Burgeon, 1946

hauseri Brenske, 1898

kenyensis Decelle, 1968*

kigonserana Moser, 1919

lindiana Moser, 1919*

lineata Lacroix, 2008

lukulediana Moser, 1919

maculipennis Lansberge, 1882*

mashuna (Péringuey, 1904 – *Pegylidius*)*

microchaeta Moser, 1919*

morio Blanchard, 1850*

= *brevior* Fairmaire, 1884

neumanni Kolbe, 1894*

pilosa Lacroix, 2008*

pondoensis Arrow, 1943*

rufolineata Kolbe, 1894*

rufomaculata Linell, 1896*

salaama Brenske, 1898

salernei Lacroix, 2008

sommeri (Burmeister, 1855 – *Hypopholis*) **comb. n.***

= *sulcicollis* (Boheman, 1857 – *Hypopholis*)

tchadensis Lacroix, 2008

tenuitarsis (Fairmaire, 1887 – *Adoretopsis*)*

uelensis (Burgeon, 1946 – *Hypopholis*) **comb. n.***

ugandensis Lacroix, 2008

usambarae Brenske, 1898

vestita Brenske, 1895

vittata (Fähræus in Boheman, 1857 – *Hypopholis*) **comb. n.***

wernerii Lacroix, 2008

zavattarii Gridelli, 1940
