

Monitoring of *Fusarium circinatum* spore loads in South African forest seedling nurseries

by

Lisa-Dan lle de Wet

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Department of Microbiology and Plant Pathology

University of Pretoria

Pretoria

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Promoter: Prof. E.T. Steenkamp

Co-promoter: Prof. M.J. Wingfield

Prof. B.D. Wingfield

Dr. N.B. Jones

DECLARATION

I, Lisa-Danélle de Wet declare that the dissertation, which I hereby submit for the degree *Magister Scientiae* at the University of Pretoria, is my own work and has not previously submitted by me for a degree at this or any other tertiary institution.

Signature

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TABLE OF CONTENTS

| | |
|--|-----------|
| ACKNOWLEDGEMENTS | 1 |
| PREFACE | 2 |
| Chapter 1 | 4 |
| Literature review: Techniques to identify <i>Fusarium</i> species | |
| INTRODUCTION | 5 |
| MORPHOLOGY AND OTHER CULTURAL TRAITS | 6 |
| HOST ASSOCIATIONS | 8 |
| SEXUAL COMPATIBILITY | 9 |
| VEGETATIVE COMPATIBILITY | 11 |
| METHODS BASED ON CHEMICAL TRAITS | 12 |
| Secondary metabolite profiling | 13 |
| Fatty acid methyl ester analysis | 14 |
| Infrared spectroscopy of cellular constituents | 14 |
| Isozyme analyses | 15 |
| ELISA-based methods | 15 |
| DNA-BASED IDENTIFICATION TECHNIQUES | 16 |
| DNA-fingerprinting methods | 16 |
| <i>Restriction fragment length polymorphism (RFLP)</i> | 16 |
| <i>Random amplified polymorphic DNA (RAPD)</i> | 17 |
| <i>Amplified fragment length polymorphisms (AFLPs)</i> | 17 |
| <i>Denaturing gradient gel electrophoresis (DGGE)</i> | 18 |
| DNA sequence analysis of specific genomic regions | 18 |
| <i>Sequence analysis of housekeeping loci</i> | 19 |
| <i>Species-specific PCR primers or probes</i> | 19 |
| <i>Microsatellites</i> | 20 |
| <i>Microarrays</i> | 21 |
| THE NEED FOR POLYPHASIC METHODS IN FUSARIUM DIAGNOSTICS | 21 |
| CONCLUSIONS | 22 |
| REFERENCES | 23 |
| Chapter 2 | 32 |
| Evaluation of the intergenic spacer (IGS) region as a diagnostic marker for <i>Fusarium circinatum</i> | |
| INTRODUCTION | 33 |
| MATERIALS AND METHODS | 36 |
| Fungal isolates | 36 |
| DNA extraction, PCR and sequencing | 36 |
| IGS sequence analysis | 37 |
| RESULTS | 38 |
| DISCUSSION | 39 |
| REFERENCES | 42 |
| FIGURES AND TABLES | 46 |

| | |
|--|------------|
| Chapter 3 | 50 |
| Monitoring spore loads of the pitch canker fungus, <i>Fusarium circinatum</i> , in a pine seedling nursery | |
| INTRODUCTION | 51 |
| MATERIALS AND METHODS | 53 |
| Experimental pine nursery and spore traps | 53 |
| Preparation of DNA standards | 54 |
| DNA isolation from spores | 54 |
| Real-time PCR | 56 |
| RESULTS | 57 |
| DNA isolation from spores | 57 |
| Real-time PCR | 57 |
| Monitoring <i>F. circinatum</i> in a pine seedling nursery | 58 |
| DISCUSSION | 58 |
| REFERENCES | 61 |
| TABLES | 63 |
| | |
| SUMMARY | 66 |
| | |
| APPENDIX 1 | A 1 |

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PREFACE

Fusarium circinatum is an important pathogen since it is a pathogen of pine trees worldwide, causing extensive destruction of both native pine forests and non-native plantations. The severity of the pitch canker disease caused by this fungus justifies understanding its distribution and spread better. The techniques used for *Fusarium* identification and detection are numerous, however, the quantification techniques for this genus, in particular *F. circinatum*, are limited. The main goal of this study was to detect and quantify the pine pathogen *F. circinatum*, using the intergenic spacer (IGS) region as a diagnostic marker.

Techniques used for the identification of *Fusarium* species can be divided into two groups i.e. non-DNA-based and DNA-based methods. The first chapter reviews the most popular and widely used techniques for *Fusarium* identification. Each group of techniques have similar advantages and disadvantages, although the groups of techniques are better suited to different types of questions and methodologies. Many years ago before the advent of molecular biology, non-DNA-based techniques, based on the morphology of an isolate, were popular. At present DNA-based techniques are more popular as these methods can be automated and considers a species on its most basic level, i.e., its DNA blueprint. DNA-based techniques are, however, usually combined with techniques based on morphology for control purposes and the various techniques also complement each other.

Many specific *F. circinatum* primers have been designed targeting the IGS region, due to this region's fast evolution. The IGS region is evaluated as a diagnostic marker in the second chapter to determine whether this region is indeed a good diagnostic marker. The specificity of the *F. circinatum* specific primers targeting the IGS were compared with the species for comparison including *F. circinatum* and other *Fusarium* species belonging to the *Gibberella fujikuroi* complex. The results indicate that the IGS region is a reliable diagnostic marker as a large part of this region is species-specific. Phylogenetic analysis of the IGS region showed that the isolates group together similarly to the groupings of previous studies that included the same isolates.

The dispersal of *F. circinatum* spores is mostly responsible for the spreading of the pitch canker disease between different individual pines. The disease is found in the whole life cycle of the pine tree, including the seed, seedling and adult stage. A technique to detect *F. circinatum* and quantify the spores can assist in the management of the disease in plantations

and seedling nurseries. In chapter three real-time PCR is the chosen technique for detection and quantification of *F. circinatum* spores in a seedling nursery as the success of this technique has been demonstrated in similar studies in the past. Four DNA extraction (from the spores) methods were also compared to increase the effectiveness and sensitivity of the whole detection and quantification process. Over a 12-month period temperature readings were recorded to determine whether there was a correlation between climate and spore production. The spore traps were also placed at different levels to determine whether air movement has an effect on spore dispersal. It was found that some of the DNA extraction methods are better than others, real-time PCR can detect low spore concentrations and temperature has an effect on spore production. This study has proven the effective use of real-time PCR for the detection and quantification of *F. circinatum* spores in a pine seedling nursery.

CHAPTER 1

LITERATURE REVIEW:

TECHNIQUES TO IDENTIFY *FUSARIUM* SPECIES

Abstract: Correct identification of *Fusarium* species, especially those harmful to humans, animals and plants will always be important. Only a few decades ago, the number of identification techniques for these fungi was limited due to technological and knowledge limitations. Since the discovery of the structure of DNA by Watson and Crick in 1953 and the improvement of associated technologies, the number of identification techniques available for fungi has increased remarkably. Originally, the only method to identify and study these fungi was that involving morphological features of isolates. This approach continues to be used, but usually in conjunction with other identification techniques. During the last decade, DNA-based identification techniques have become increasingly valuable as a result of being amenable to automation and the resolving power they provide. This review examines *Fusarium* identification techniques, discussing the properties of each technique, as well as their advantages and limitations. The final section considers the need and value of a polyphasic approach to *Fusarium* identification.

INTRODUCTION

Fusarium resides in the Order Hypocreales and in the Phylum Ascomycota (Seifert and Lévesque, 2004; Hibbett *et al.* 2007; Kristensen *et al.*, 2007b). Knowledge of this genus goes back as far as 1842, when *Fusarium solani* was first described as *Fusisporium solani* (Snyder and Hansen, 1941). *Fusarium* species are widely distributed across the globe where they commonly occur as saprobes or pathogens of higher eukaryotes (Leslie and Summerell, 2006). As plant pathogens, *Fusarium* species are frequently associated with a wide variety of hosts, including cereals and vegetable crops, as well as fruit and tree species (Leslie and Summerell, 2006). As human and animal pathogens, members of this genus are often linked with a range of opportunistic infections (Nelson *et al.*, 1994). *Fusarium* species are also abundant in the soil where they form an important component of microbial communities (Nelson *et al.*, 1994). In addition, *Fusarium* species are capable of producing a variety of toxic secondary metabolites or mycotoxins, which have been implicated in numerous animal and human diseases (Nelson *et al.*, 1994; Desjardins, 2003;). The agricultural, medical and veterinary importance of *Fusarium* species, therefore, makes their early detection and correct identification crucially important for effective control and/or management.

An important consideration for the identification process of any fungi is the application of an appropriate Species Concept (Mayden, 1997). The Species Concept determines which

experimental approaches or techniques should be used during the identification process, and also how the results of these experiments should be interpreted. Of the numerous Species Concepts available (reviewed by Mayden, 1997), the Morphological, Biological and Phylogenetic Species Concepts (MSC, BSC and PSC, respectively) are most widely used in *Fusarium* taxonomy (Leslie and Summerell, 2006).

Of the three Species Concepts, the MSC is the most extensively used technique, with the description of most *Fusarium* species including morphological diagnoses for differentiation and identification. However, *Fusarium* species are generally associated with a limited number of unique morphological traits (Leslie *et al.*, 2001), which drastically reduces the value of the MSC in *Fusarium* species diagnostics. The BSC concentrates on whether two isolates are capable of crossing sexually to produce fertile progeny (Dobzhansky 1935; Mayr 1996). However, the BSC also has limited application, as it is restricted to sexual species. The third Species Concept, PSC, forms part of most contemporary *Fusarium* classifications and identifications, because it focuses on the lineage of a species, its history (ancestors), future (descendants) and its evolution (how it originated) (Avice and Wollenberg, 1997). A number of excellent reviews on the Species Concepts used for *Fusarium* have been published (e.g., Nelson *et al.*, 1994; Leslie *et al.*, 2001; Summerell *et al.*, 2003; Leslie and Summerell 2006) and the details will not be discussed further in this review.

A second important aspect of the identification process that requires attention is the actual techniques used to diagnose a species. Most modern diagnostic procedures are based on DNA sequence information, although so-called “molecular methods” are sometimes used in conjunction with morphology. These techniques may vary greatly with regards to technical difficulty, financial considerations, time and level of resolution. However, for the purposes of routine species diagnostics, an effective identification technique should be rapid, economical, and quantitative, as well as allowing for direct sample analysis, minimum sample preparation and automation (Nie *et al.*, 2007). In this literature review, methods that have been or are currently used to identify species or strains of *Fusarium* are considered. Their value and suitability for routine diagnostic purposes is also discussed.

MORPHOLOGY AND OTHER CULTURAL TRAITS

Before 1935, hundreds of *Fusarium* species had been described as every isolate that was discovered on a new host was reported to be a new species (Leslie *et al.*, 2001). More order

in the identification and classification of *Fusarium* species was introduced by Wollenweber and Reinking (1935). The principles introduced at that time are still the foundation of modern *Fusarium* taxonomy. They divided the approximately thousand *Fusarium* species into sixteen sections, sixty-five species and seventy-five varieties (Wollenweber and Reinking, 1935; Nelson *et al.*, 1994). Snyder and Hansen (1956) concluded that there were only nine *Fusarium* species, but modern taxonomic treatments have shown that this is far from true (Leslie *et al.*, 2001; Seifert and Lévesque, 2004). Booth (1971) produced a synoptic key for the identification of *Fusarium* species based on many of the principles of Wollenweber and Reinking (Leslie *et al.*, 2001). Later, Gerlach and Nirenberg (1982) and Nelson *et al.* (1983) considered the work of Wollenweber and Reinking (Wollenweber and Reinking, 1935), Snyder and Hansen (Snyder and Hansen, 1940) and Booth (Booth, 1971) in order to develop strategies for the identification of *Fusarium* species based on morphological characteristics (Leslie *et al.*, 2001; Summerell *et al.*, 2003; Seifert and Lévesque, 2004). Many of these identification procedures are still widely used today.

A variety of nutrient media are used during the *Fusarium* identification process based on species morphology. The most important are carnation leaf agar (CLA) (Fischer *et al.*, 1982; Nelson *et al.*, 1994; Summerell *et al.*, 2003), potato dextrose agar (PDA), peptone-pentachloronitrobenzene (PPA; also known as *Fusarium* selective medium) (Summerell *et al.*, 2003) and synthetic low nutrient agar (SNA) (Nirenberg and O'Donnell, 1998; Summerell *et al.*, 2003). Isolates grown on CLA are more inclined to sporulate and instead of producing mycelial growth. Isolates produce macroconidia and chlamydospores making this medium valuable because both structures are important for identification under the microscope (Fischer *et al.*, 1982; Nelson *et al.*, 1994; Nirenberg and O'Donnell, 1998). PDA is a nutrient-rich medium that encourages mycelial growth causing isolates to produce colony pigmentation that is useful in identification (Nelson *et al.*, 1994; Summerell *et al.*, 2003). On the other hand, SNA is a low nutrient medium that can maintain strains for long periods of time and encourages the production of microconidia that can be used for identification when visualised under a microscope (Nirenberg and O'Donnell, 1998; Summerell *et al.*, 2003).

A pictorial guide (Summerell *et al.*, 2003) can be used to help with identification according to different spore types and the dimensions of spores, as these characteristics can be unique to a species. Other important characteristics include colony morphology and hyphal growth (Nelson *et al.*, 1994; Summerell *et al.*, 2003). However, a common problem with

morphological identification of *Fusarium* species is that different species may have similar or identical morphological characteristics. These are especially well documented for plant pathogenic species such as those in the *Gibberella zeae* (Burgess *et al.*, 1994), *G. fujikuroi* (Kvas *et al.*, 2009) and *F. solani* (Matuo and Snyder, 1972) species complexes.

HOST ASSOCIATIONS

Fusarium species are associated with many plant species and often cause serious diseases (Summerell *et al.*, 2003). As pathogens, they are also capable of infecting different parts of a plant (Summerell *et al.*, 2003). Because of the possible economic losses that may be associated with *Fusarium* diseases of plants, correct species identification is essential for the establishment of appropriate management strategies. This is because identification of the *Fusarium* species found on a specific plant host is important to determine the possible outcome of the disease, assessing and predicting the extent of the damage and whether the *Fusarium* species are in fact the cause of the disease.

Many *Fusarium* species are host-specific. Thus, the host on which an isolate of *Fusarium* is found may represent an important diagnostic character. *F. oxysporum* is a species that cause disease (commonly vascular wilt syndrome) in many different hosts including banana, cotton, legumes and vegetables all of which are economically important crops. *F. oxysporum* strains that cause vascular wilt syndrome are often host specific, infecting only a limited number of hosts. These strains are termed formae speciales as they are divided on a subspecies level, *F. oxysporum* f. sp. *cubense* infecting banana, *F. oxysporum* f. sp. *vasinfectum* infecting cotton and *F. oxysporum* f. sp. *lycopersici* infecting tomato to name a few (Summerell, *et al.* 2003). *F. solani* is another species than contain strains that are host specific (Summerell, *et al.* 2003). Therefore, pathogenicity tests are typically conducted to verify the correct identification and to determine whether a fungus is able to infect a plant and would thus be recognised as the causal agent of a disease.

The steps followed in pathogenicity tests are based on Koch's postulates (Koch, 1891; Fredricks and Relman, 1996; Grimes, 2006). According to these postulates, a pathogen must always be present during disease, the suspected pathogen must be isolated from the diseased host and cultured, a healthy host inoculated with the pathogen must show signs of disease with symptoms identical to the original host's symptoms and it should be possible to isolate the pathogen from the newly diseased host.

When testing for *Fusarium* host associations, the first step is the isolation of the possible pathogen and preparation of pure cultures on isolation medium. These cultures are then used to produce inoculum, which may be in various forms, including spore suspensions (e.g., Benyon *et al.*, 1996; Amatulli *et al.*, 2010) and mycelium obtained from growth media (Pérez-Sierra, 2007). The inoculum is then applied to healthy plants, for example, to the roots (e.g., Cafri *et al.*, 2005), kernels (e.g., Yates and Sparks, 2008) or stems (e.g., Catti *et al.*, 2007). The presence or absence of symptoms identical to those of the original host's symptoms is recorded to determine whether the organism in question is the causal agent of disease (Cafri *et al.*, 2005; Catti *et al.*, 2007; Jacobs *et al.*, 2007; Yates and Sparks, 2008). Finally, the pathogen is isolated from the diseased tissue to show that it was the causal agent of the observed symptoms.

SEXUAL COMPATIBILITY

The most important advantage of evaluating sexual compatibility is that it allows direct testing of the BSC (Dobzhansky, 1935; Mayr, 1996). If two individuals are capable of interacting sexually to produce fertile offspring, they probably represent members of the same species. The major disadvantages associated with this approach for *Fusarium* identifications pertain to issues surrounding asexuality and infertility. The teleomorph form of many species has never been found and most *Fusarium* species are known only in their asexual or anamorph forms. This, in turn, may be because the species is truly asexual, having lost its sexual phase as has been suggested for *F. oxysporum* (Leslie and Summerell, 2006). Alternatively, the inability to detect the sexual stage may be due to the inclusion of isolates in the test that are not sufficiently fertile under suitable laboratory conditions (for example Covert *et al.*, 1999) or due to the test being conducted at unsuitable laboratory conditions. Also, many isolates of *Fusarium* are known to have lost their female-fertility (the ability to act as female and produce the progeny bearing fruiting structure) and can only act as males (nuclear donors) in sexual crosses (Leslie and Summerell, 2006). Therefore, “negative results” (failure to induce sexual reproduction and the formation of mature fruiting structures and progeny) do not necessarily indicate that the interacting individuals belong to separate species. The latter is also true for “positive results” (successful induction of sexual reproduction and the formation of fruiting structures that bear fertile progeny), as closely related species may be able to mate (Dobzhansky, 1935; Mayr, 1996). Indeed, the genus *Fusarium* includes numerous examples of such cross-species or hybrid encounters, for

example *F. circinatum* and *F. subglutinans* (Steenkamp *et al.*, 2000), and *F. proliferatum* and *F. fujikuroi* (Leslie *et al.*, 2004).

Among the sexual species in *Fusarium* (those that have teleomorphs in the genera *Gibberella* and *Haemanectria*), some groups are heterothallic, while others are homothallic (Leslie and Summerell, 2006). Heterothallic species require the interaction of two fertile isolates of opposite mating types to induce the formation of sexual structures and to produce fertile progeny (Leslie and Summerell, 2006). Well-known examples of heterothallic species include those in the *G. fujikuroi* complex. On the other hand, the individuals of a homothallic species can produce sexual fruiting structures bearing fertile progeny, alone, but this form of reproduction results in progeny that are largely clones of the parent (Leslie and Summerell, 2006). Certain species of *Haemanectria* and species in the *G. zae* complex have this form of sexual reproduction (Hornok *et al.*, 2007).

The sexual species in *Fusarium* are generally dimictic, where sexual reproduction is determined by two alleles found at one mating type or MAT locus (Leslie and Summerell, 2006). The two alleles share little sequence similarity and were accordingly designated as the *MAT-1* and *MAT-2* idiomorphs (Cooke, 1958; Kerényi *et al.*, 1999). It has also been shown that the overall structure of this locus in asexual *Fusarium* species resembles that of sexual species (Yun *et al.*, 2000). The only difference between heterothallic and homothallic species, is that the two idiomorph alleles are located adjacent to one another on the same DNA molecule at the MAT locus of the homothallic *Fusarium* species studied to date, while the MAT locus of heterothallic species harbours only one of the idiomorph alleles (reviewed by Leslie and Summerell, 2006). Therefore, by exploiting the sequences at this locus in heterothallic species, it was possible to develop PCR-based approaches for determining the mating type of a specific isolate (Steenkamp *et al.*, 2000; Wallace and Covert, 2000; Yun *et al.*, 2000).

In the laboratory, sexual crosses with *Fusarium* species typically involve culturing the female strains on a medium such as V8 or carrot agar (Britz *et al.*, 1999; Covert *et al.*, 2007) and the male strains on a medium that allows abundant vegetative growth and sporulation (e.g. complete medium or PDA) (Britz *et al.*, 1999; Kerényi *et al.*, 1999; Leslie and Summerell, 2006; Covert *et al.*, 2007). Spores are then harvested from the male parent and used to “spermatise” the female culture. The fertilised cultures are then incubated for several months and regularly examined for the presence of perithecia and ascospores. To verify progeny

viability, ascospores may be germinated on water agar, after which the resulting cultures should display growth typical of the parental strains. Ideally, the progeny should also be subjected to sexual crosses to verify that they are indeed fertile. Such viable and fertile progeny generally result only from an interaction between conspecific, sexually compatible parents (Britz *et al.*, 1999; Kerényi *et al.*, 1999; Leslie and Summerell, 2006; Covert *et al.*, 2007).

The widespread occurrence of homothallism in *Fusarium* species complicates the use of sexual compatibility assays to test the BSC (Leslie and Summerell, 2006). This is because these fungi are able to complete the sexual cycle by themselves without the need for an interacting partner (Covert *et al.*, 2007; Hornok *et al.*, 2007), although mating between two isolates (outcrossing) is also possible (Bowden and Leslie, 1999; Hornok *et al.*, 2007). It is therefore, possible to use sexual compatibility tests effectively and where suitable controls are used and the results are interpreted appropriately. For example, to ensure that the emergence of sexual fruiting structures is due to an interaction between mated individuals and not the result of self-fertilisation, the progeny may be examined for the presence of recombinants (Covert *et al.*, 2007). If they display the characters or carry the alleles of only one of the parents, that parent probably gave rise to them through homothallic recombination. If they display traits or carry alleles of both parents, the parents are likely to be conspecific because they both participated in the cross. In addition, improved efficiency of sexual compatibility tests with homothallic species (increased appearance of heterozygous fruiting structures) can be achieved by using female fertile tester strains in which one of the two idiomorph alleles has been inactivated (Leslie and Summerell, 2006).

VEGETATIVE COMPATIBILITY

Vegetative compatibility is typically used to study the population biology of fungal species because this trait is determined by multiple genes or genetic loci (Leslie, 1993). Fungal species are thought to have between ten and fifteen of these loci (reviewed by Leslie and Summerell, 2006). For individuals to be compatible (the hyphae of two individuals fuse to form a stable heterokaryon), they must have similar alleles at all of these loci. Dissimilarity at a single locus usually results in incompatibility, as a stable heterokaryon is not maintained (Leslie, 1993). Due to the multigenic basis of this phenomenon, the progeny of a sexual cross will very rarely be compatible with the parental strains. Therefore, for a fungal species believed to primarily undergo asexual reproduction, vegetative compatibility tests represent a

powerful identification technique (Leslie and Summerell, 2006). This is because the lack of meiosis and sexual recombination precludes reshuffling of allelic combinations underpinning a specific vegetative compatibility group. As a result, vegetative compatibility groups form an integral part of the diagnostic procedure for species such as *F. oxysporum* (Puhalla, 1985).

To determine the vegetative compatibility group of an isolate, two isolates are paired and if their hyphae are able to fuse, the isolates are defined as belonging to the same vegetative compatibility group or as vegetatively compatible (Leslie, 1993). This test is facilitated by the use of nitrate non-utilising (*nit*) mutants and pairing of compatible mutants (Correll *et al.*, 1987; Klittich and Leslie, 1988). If wild type growth is produced in the area of contact between the two isolates, a stable heterokaryon is formed, which indicates that the paired isolates belong to the same vegetative compatibility group. If no growth is produced in the area of contact between the two isolates and a stable heterokaryon are not formed, the two isolates are vegetatively incompatible (Correll *et al.*, 1987; Klittich and Leslie, 1988).

METHODS BASED ON CHEMICAL TRAITS

Chemotaxonomy focuses on the molecules produced by a fungal species and includes the analysis of fatty acids, proteins, secondary metabolites (SMs) and carbohydrates (Frisvad *et al.*, 2008). Chemotaxonomy refers to the use of these chemical compound characteristics for classification or identification purposes (Frisvad *et al.*, 2007; Frisvad *et al.*, 2008). To identify a species based on these properties, a profile for the specific compound in a particular isolate is determined and compared to the profiles of known species in a database. The accuracy of identifications is thus dependant on the species-richness of the reference database, which also represents one of the major limitations associated with using chemotaxonomic approaches for species diagnoses. In addition, identification can be complicated by the ability of some species that are not closely related, producing similar compounds (De Nijs *et al.*, 1997). Other complications may include individuals that are not equally capable of producing the full range of compounds associated with that particular species (De Nijs *et al.*, 1997). Another factor that should also be considered is that the growth phase of the isolate may have an effect on the compounds produced. During sporulation certain secondary metabolites are produced that are not produced during other growth phases (Frisvad *et al.*, 2008).

Despite the various shortcomings associated with chemical compound profiling and chemotaxonomic analysis, these approaches are generally easy to standardise for application

in routine identification (De Nijs *et al.*, 1997). They are usually not time-consuming or laborious, although they may be dependent on the use of specialised equipment, which could limit their use for diagnostic purposes. In contrast, methodologies that involve more complex biochemical procedures (e.g., isozyme analysis; see below) or that require specialised material (e.g., antibodies used in ELISA; see below) are of limited use for routine diagnostic purposes. In fact, DNA-based procedures have now largely replaced the use of both ELISA and isozyme analyses for identification procedures.

Secondary metabolite profiling

Secondary metabolic compounds (SM) are not essential under normal growth circumstances (Frisvad *et al.*, 2008). They may, however, become necessary under abnormal circumstances, including a change in food sources available and in the presence of predators/competitors (Frisvad *et al.*, 2008). Secondary metabolites, most commonly produced by fungi, which are used for identification include terpenes, polyketides and non-ribosomal peptides (Jegorov *et al.*, 2006; Frisvad *et al.*, 2008). It is widely believed that SMs can be used for the classification and identification of fungi due to their inconsistent distribution in the fungal kingdom (De Nijs *et al.*, 1997; Frisvad *et al.*, 2008).

Mycotoxins represent an important group of SMs in fungi and *Fusarium* in particular. Knowledge of toxins produced on plants dates back to ancient Egypt as recorded in the Dead Sea Scrolls, but mycotoxins were only recognised as associated with plants in the 1900s (Richard, 2007). Mycotoxins commonly appear in the food chain, either directly through the consumption of contaminated food crops, or indirectly when livestock ingest affected feed. Secondary metabolites or mycotoxins of *Fusarium* are almost always associated with severe human and animal diseases (Nelson *et al.*, 1994; Desjardins 2003; Richard, 2007), which further emphasises the need for accurate species diagnosis (Richard, 2007).

Secondary metabolite profiles usually differ between fungal species, because every species produces a unique combination or quantity of small organic compounds most of which are secondary metabolites (Frisvad *et al.*, 2007; Frisvad *et al.*, 2008). Although analyses of all these compounds produced by a species may harbour taxonomically useful information (Frisvad *et al.*, 2008), the association of SMs (mycotoxins in particular) with human and animal disease has emphasised the use of SM profiles for the identification of fungal species in the medical and veterinary environments. *Fusarium* species are known to produce a large

range of SMs and mycotoxins such as trichothecenes, fumonisin, moniliformin, zearalenones and enniatins (Kristensen *et al.*, 2007a). Examples of *Fusarium* species that have been differentiated based on SM profiles include *F. culmorum* and *F. avenaceum* (De Nijs *et al.*, 1997), as well as *F. langsethiae*, *F. poae*, *F. sporotrichioides* and *F. kyushuense* (Thrane *et al.*, 2004).

To determine the SM profile of an isolate or species, the presence of toxins, antibiotics and other outwardly directed compounds produced during fungal growth under specific conditions are studied (Fischer *et al.*, 2000). For this purpose, isolates are typically grown on complex media that are rich in glucose and sucrose, as well as minerals and trace elements (Fischer *et al.*, 2000), for example PDA (Frisvad *et al.*, 2008), malt extract agar (MEA) (Fischer *et al.*, 2000; Frisvad *et al.*, 2008) and yeast extract-sucrose agar (YES) (Fischer *et al.*, 2000; Frisvad *et al.*, 2008). The chemical characteristics of the isolate may then be determined using one of three widely used techniques: chemical spot test, paper chromatography or thin layer chromatography (Frisvad *et al.*, 2008). Although thin layer chromatography is still used as a precursor technique during screening, high-pressure liquid chromatography diode array detection mass spectroscopy (hplc-dad-ms) is currently the technique of choice for fungal chemotaxonomy (Frisvad *et al.*, 2008).

Fatty acid methyl ester analysis

Fatty acid methyl ester (FAME) analysis is a chemotaxonomic technique that concentrates on the quantity and type of fatty acids produced by a fungus (Aye *et al.*, 2008). To achieve identification, the fatty acids are extracted from an isolate and separated by gas chromatography. FAME analysis allows identification to genus, species or strain levels (Aye *et al.*, 2008). However, when it was applied to the fungi associated with rice blight, the method did not allow for differentiation between *F. roseum* and *F. solani*, although it did differentiate the *Fusarium* species from *Rhizopus* and *Pythium* species (Aye *et al.*, 2008).

Infrared spectroscopy of cellular constituents

Fourier transform infrared spectroscopy determines the presence or absence of functional groups that may be unique to an isolate by irradiating the isolate with infrared light at different wavelengths (McMurry, 1998; Kos *et al.*, 2002; Nie *et al.*, 2007). Infrared spectroscopic techniques that have been used in the past to detect fungi include photoacoustic spectroscopy (PAS), diffuse reflectance spectroscopy (DRS) and attenuated total reflectance

spectroscopy (ATR) (Kos *et al.*, 2002). Infrared spectroscopy is based on the fact that different molecules absorb electromagnetic energy differently, which may be reflected in the potential energy curves of the molecules caused by the interaction of the molecules with infrared radiant energy (McMurry, 1998; Illingworth and Cullerne, 2000). Thus, during electromagnetic radiation, an exposed organic compound absorbs energy of certain wavelengths, which allows the generation of an absorption spectrum for the compound at various irradiation wavelengths. This is accomplished by using the Fourier transform mathematical algorithm, which allows the conversion of the raw absorbance data into the specific spectrum (*i.e.*, the absorbance units are functionally expressed and possibly related to wave numbers that are also functionally expressed) (Illingworth and Cullerne, 2000). A number of studies have used the Fourier transform infrared spectroscopy technique successfully to identify *F. graminearum*, *F. moniliforme*, *F. nivale*, *F. oxysporum* and *F. semitectum* (Kos *et al.*, 2002; Nie *et al.*, 2007).

Isozyme analyses

The protein constituents of an unknown species may be examined for identification purposes. Isozyme analysis represents an example of such a protein-based method that has been widely used in fungal (Micales *et al.*, 1986), and especially *Fusarium* taxonomy (Huss *et al.*, 1996; Yli-Mattila *et al.*, 1996; Abd-Elaah, 1998; Mohammadi *et al.*, 2004). The method exploits the fact that different species generally produce unique combinations of isozymes (Mohammadi, *et al.*, 2004). To evaluate this, the isozymes produced by a sample/isolate are extracted and separated by polyacrylamide gel electrophoresis. The isozyme bands are then visualised with appropriate stains and enzyme-buffer systems (Huss *et al.*, 1996; Yli-Mattila *et al.*, 1996; Mohammadi *et al.*, 2004). Isozyme profiles are then compared to a reference database to allow identification of the unknown sample/isolate.

ELISA-based methods

Enzyme-Linked Immunosorbent Assay (ELISA) may be used to detect a particular microbial species within a specific sample. ELISA is based on the detection of surface antigens or secreted antigens by an antibody with a detectable enzyme linked to the antibody (McCartney *et al.*, 2003; Jegorov *et al.*, 2006). Polyclonal antibodies (Aldwell *et al.*, 1983; Kaufman *et al.*, 1997; Ward *et al.*, 2004), monoclonal antibodies (Park *et al.*, 2003; Ward *et al.*, 2004; Hill *et al.*, 2006) and antibodies produced by phage display (Ward *et al.*, 2004) are the three types

of antibodies that can be used for identification and detection. Antigens can be detected by direct detection (Ward *et al.*, 2004; Hill *et al.*, 2006), indirect detection (Aldwell *et al.*, 1983; Ward *et al.*, 2004) or by “sandwich” detection where antibodies attach to two sides of the antigen (Ward *et al.*, 2004). A basic ELISA technique for detecting and quantifying *Fusarium* species found on barley was developed (Hill *et al.*, 2006) and used to study the fungi associated with *Fusarium* Head Blight (FHB) (Hill *et al.*, 2008). As with the use of other cellular constituents for the identification of species, the main limitation of ELISA is the availability of reference information, in this case appropriate antibodies to allow accurate identifications.

DNA-BASED IDENTIFICATION TECHNIQUES

Identification methods based on DNA information can generally be separated into two main groups, those that result in fingerprints, and those that access the sequence information directly. These two groups of methods differ in terms of their applicability for routine species identification. Fingerprinting methods, for example, suffer from the same limitations as those associated with chemical compound profiles (see above), as any particular DNA fingerprint must be compared to a database of reference fingerprints for known species. Thus, where the database is limited or if the unknown sample is new to Science, positive identifications are impossible. Also, with DNA-fingerprinting the possibility of homoplasy cannot be excluded, because a specific band in a fingerprint of two species is not necessarily underpinned by orthologous DNA information (Yli-Mattila *et al.*, 1996). Methods that are based on the actual DNA information are much more powerful in this regard, because species are usually distinguishable based on the uniqueness of their particular DNA sequence. In the following sections, the various DNA-based approaches used for the identification of fungi and *Fusarium* species, in particular, are briefly discussed.

DNA-fingerprinting methods

Restriction fragment length polymorphism (RFLP)

The first DNA-based method that was widely applied in fungal taxonomic studies was restriction fragment length polymorphism (RFLP). For this method, genomic DNA is digested with endonucleases, separated by electrophoresis and polymorphisms visualised following Southern hybridization with specific probes (Niesters *et al.*, 1993; Barve *et al.*, 2001). Because this method is tedious and labour-intensive, it now has limited application for

the identification of *Fusarium* species. Instead, a variant of the RFLP technique, PCR-RFLP, is used much more often for taxonomic purposes. PCR-RFLP entails the amplification of a specific genomic region using PCR, followed by digestion of the amplicon with restriction endonucleases and visualisation of the polymorphisms with standard gel electrophoresis procedures (Paavanen-Huhtala *et al.*, 1999; Kim *et al.*, 2001; Hinojo *et al.*, 2004; Llorens *et al.*, 2006a). Because of the simplicity and robustness of PCR-RFLPs, the approach has been used widely for the identification of *Fusarium* species (Young-Mi *et al.*, 2000; Konstantinova and Yli-Mattila, 2004) such as those in the *G. fujukuroi* complex (Hinojo *et al.*, 2004) and *F. redolens* (Bogale *et al.*, 2007), as well as formae speciales of *F. oxysporum* (Manicom *et al.*, 1986; Appel and Gordon, 1995).

Random amplified polymorphic DNA (RAPD)

RAPD is a PCR-based method in which single, short oligonucleotide primers are used to amplify arbitrary genomic regions, followed by visualisation of polymorphisms with standard gel electrophoresis methodologies (Williams *et al.*, 1990). RAPD techniques have been used extensively for the identification of *Fusarium* species such as *F. oxysporum*, in particular (Chiocchetti *et al.*, 1999; Paavanen-Huhtala *et al.*, 1999; Del Mar Jiménez *et al.*, 2001; El-Fadly *et al.*, 2008). However, the RAPD technique is notoriously sensitive, which limits its reproducibility within and among laboratories (Vos *et al.*, 1995). As a result, contemporary *Fusarium* identification procedures seldom use this method.

Amplified fragment length polymorphisms (AFLPs)

AFLPs harness the best qualities of genomic RFLPs and RAPDs to produce a simple and robust DNA fingerprinting method (Vos *et al.*, 1995). In principle, restricted genomic DNA fragments are ligated to adaptors (with known sequences) and amplified, to allow the amplification of fingerprints, which are visualised with standard electrophoresis methods (Vos *et al.*, 1995; Blears *et al.*, 1998). Since its introduction, AFLP has been applied for taxonomic purposes in many fungi (Abdel-Satar *et al.*, 2003). In fact, *Fusarium* isolates of a single species have been shown to display AFLP fingerprints that share 70 % or more similarity (Summerell *et al.*, 2003). This method has been used to study *Fusarium* species such as *F. langsethiae* (Schmidt *et al.*, 2004), *F. avenaceum*, *F. chylamdiosporum*, *F. oxysporum*, *F. moniliforme* and *F. solani* (Abdel-Satar *et al.*, 2003).

Denaturing gradient gel electrophoresis (DGGE)

DGGE is a popular technique for microbial ecology, as it solves the problem associated with most DNA-fingerprinting techniques of being unable to separate non-identical DNA fragments of the same size (Muyzer and Smalla, 1998). This is because these fragments are each characterised by their own denaturation properties. Thus, by applying such fragments to DGGE analysis (which uses polyacrylamide gel electrophoresis and a DNA denaturant such as urea and formamide with a linear gradient), it should be possible to separate such non-identical fragments from one another. These products can then be visualised with an appropriate DNA stain, and each band of an individual fingerprint will represent a unique sequence type (Muyzer and Smalla, 1998). To obtain identifications, these individual fragments are compared to the DGGE fragments of known reference species, or individual fragments can be sequenced directly to allow putative identification. Because this method allows for a culture-independent examination of the species potentially present in a particular environment, DGGE is gaining in popularity (Theelen *et al.*, 2001; Mach *et al.*, 2004). In the case of *Fusarium* it has, for example, been used to address ecological questions pertaining to Asparagus (*Asparagus officinalis*) root and crown rot (Yergeau *et al.*, 2005).

DNA sequence analysis of specific genomic regions

The utilisation of DNA sequence information for species identification typically involves phylogenetic tree-based and non-tree-based analyses. For tree-based identifications, DNA sequence information for the test isolates is subjected to phylogenetic analyses, after which conclusions regarding species identities are made from phylogenetic trees (Felsenstein, 1981). This approach is widely used in *Fusarium* taxonomy where it has allowed the systematic analysis of species in the *G. fujikuroi* (O'Donnell *et al.*, 1997; O'Donnell *et al.*, 2000), *G. zaeae* (O'Donnell *et al.*, 2004b) and *F. oxysporum* (O'Donnell *et al.*, 2004a) complexes.

DNA sequence information-based identifications that are dependent on non-tree-based analysis procedures typically involve the calculation of sequence similarities. This can, for example, be achieved by making use of BLAST (Basic Local Alignment Search Tool; <http://www.ncbi.nlm.nih.gov>) and public domain databases such as the nucleotide database (GenBank) of the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov>). The detection of specific single nucleotide polymorphisms (SNPs) (Rafalski, 2002; Kristensen *et al.*, 2007a) can also be used (Rafalski, 2002).

Sequence analysis of housekeeping loci

Markers commonly used for *Fusarium* diagnostics and identification include the genes encoding β -tubulin, histone H3, translation elongation factor 1 alpha (TEF) and the ribosomal RNA (rRNA) subunits (O'Donnell and Cigelnik, 1997; O'Donnell *et al.*, 1998; Geiser *et al.*, 2004), as well as the rRNA intergenic spacer (IGS) region (Kim *et al.*, 2001). In fact, comparisons have shown that phylogenetic trees inferred from the mitochondrial small subunit (mtSSU) rDNA, the nuclear 28S rDNA, β -tubulin and TEF are congruent (O'Donnell and Cigelnik, 1997; O'Donnell *et al.*, 1998). The only exception is in the rRNA internal transcribed spacer 2 (ITS2), which appears to have non-orthologous origins (O'Donnell and Cigelnik, 1997). The ITS regions are, therefore, not used for *Fusarium* identifications. This is despite the fact that the ITS region is the preferred DNA barcoding region for fungi (Pryce, *et al.*, 2003). Instead, the TEF region has been proposed for use in routine identifications of *Fusarium*, as this gene region appears to evolve sufficiently fast (harbours enough variation to allow species differentiation) and it does not harbour non-orthologous copies (Geiser *et al.*, 2004).

Species-specific PCR primers or probes

Species-specific PCR primers and probes have been developed by making use of diagnostic motifs occurring in specific genes or regions (e.g., IGS, β -tubulin, TEF, and mating type genes) (McCartney *et al.*, 2003; Ma and Michailides, 2007). They can also be developed by making use of the results of DNA fingerprinting studies, where diagnostic RAPD or AFLP fragments are sequenced and further characterised by determining the sequence of up- and downstream regions (McCartney *et al.*, 2003; Ma and Michailides, 2007). For *Fusarium* species, various species-specific primer sets or probes are available and are most commonly based on the sequence information for the rRNA IGS (Jurado *et al.*, 2005) and the genes encoding Tox5 (Demeke *et al.*, 2005) and 28S rDNA (Edel *et al.*, 2000).

To increase the speed and efficiency of species identification with species-specific PCR primers and probes, electrochemiluminescence PCR (Wei and Zhou, 2008), indirect real-time PCR (Ioos *et al.*, 2009) and direct real-time PCR (Schweigkofler *et al.*, 2004; Garbelotto *et al.*, 2008) have been used. For electrochemiluminescence PCR, each primer has a universal fragment attached to its tail, resulting in the amplicons containing known sequences. After PCR, hybridisation of two probes (biotin labelled probe and TBR labelled probe) complementary to the universal/known sequences occurs followed by capturing by magnetic

beads. During the capturing process, biotin and streptavidin interact and are detected by electrochemiluminescence. This technique is sensitive, simple and specific (Wei and Zhou, 2008). It has been used to detect *Fusarium* wilts caused by *F. oxysporum* f.sp. *cubense* (Wei and Zhou, 2008).

During indirect real-time PCR, fluorescence is generated during primer extension of the amplification stage of PCR, using the hydrolysis probe system (Leišova *et al.*, 2006). The hydrolysis probe is digested by the 5' activity of DNA polymerase causing a fluorescence quencher and reporter to become separated, resulting in the emission of light (the quencher cannot quench the fluorescence if not in close proximity to the reporter). During direct real-time PCR, a fluorescent molecule binds to, or is incorporated into, the amplification product, the result being the emission of light (Walker, 2001). In both direct and indirect real-time PCR, fluorescence or the emission of light reveals that the DNA for the target species is included in the sample mixture.

Examples of both types of real-time PCR are available for the pine pitch canker pathogen, *F. circinatum*. Indirect real-time PCR was used to detect this fungus in pine seeds by using a dual-labelled probe unique to *F. circinatum*'s IGS (Ioos *et al.*, 2009). For detection and quantification of this fungus with direct real-time PCR, species-specific primers, are used to amplify a portion of the IGS in the presence of the fluorescence dye SYBR Green (Schweigkofler *et al.*, 2004; Garbelotto *et al.*, 2008).

Microsatellites

Microsatellites are short tandem repeats, usually 1-10 bp long, that differ in length between isolates due to their high mutation rate (Jarne and Lagoda, 1996; Powell *et al.*, 1996). Sequences adjacent to the microsatellites are used to produce primers that allow the amplification of the microsatellites with PCR, after which the amplicons are separated on a gel by electrophoresis to reveal polymorphisms (Longato and Bonfante, 1997; Barve *et al.*, 2001; Santana *et al.*, 2009). Although microsatellites are usually used for population genetic studies, they are also useful for identifying the formae speciales and races of *F. oxysporum* (Barve *et al.*, 2001).

Microarrays

For large-scale identification of specific groups of fungi such as *Fusarium*, microarray technology has been used. Microarrays are based on the hybridisation of two complementary sequences, during which a known DNA sequence fragment of an organism is attached to a solid support, usually a glass slide (Wilson *et al.*, 2002; Spiess *et al.*, 2007) or silica chip (Campa *et al.*, 2008). The target genomic region is then amplified with PCR from the unknown isolates, labelled, denatured and hybridised to the DNA on the slide. Following wash steps to remove the unbound DNA, hybridisation is detected by the emittance of fluorescent light (Wilson *et al.*, 2002; Yoo *et al.*, 2009). This technology is commonly used for the diagnosis and identification of human fungal pathogens (Spiess *et al.*, 2007; Campa *et al.*, 2008) but also for the diagnosis and identification of *Fusarium* plant pathogens and those that produce mycotoxins in grain crops (Nicolaisen *et al.*, 2005; Kristensen *et al.*, 2007b).

THE NEED FOR POLYPHASIC METHODS IN *FUSARIUM* DIAGNOSTICS

Polyphasic identification involves the characterisation of a species at many levels, including the genotypic, phenotypic and phylogenetic characteristics of an organism (Vandamme *et al.*, 1996). The reason this approach has grown in popularity is that different techniques complement each other, where the one technique overcomes the shortcomings of others and vice versa. *Fusarium* identification could thus include DNA-based, as well as information regarding morphology, host, etc. (Llorens *et al.*, 2006b).

For *Fusarium* species, the polyphasic approach is also expanded to the actual Species Concept used. Morphological characteristics that are often used during application of the Morphological Species Concept (MSC) for identifying *Fusarium* species are limited in number and are often affected by environmental conditions, possibly resulting in a change in these characteristics (Leslie *et al.*, 2001). As discussed previously, the Biological Species Concept (BSC) can also not be used to identify all *Fusarium* species. Although application of the Phylogenetic Species Concept (PSC) generally overcomes problems associated with the other identification criteria, it may complicate identifications in the absence of additional data. Therefore, the PSC is typically used in conjunction with the MSC and/or the BSC.

CONCLUSIONS

Most *Fusarium* identification techniques have their own set of advantages and disadvantages. Advantages include specificity, sensitivity, speed, ease of automation and low cost. Disadvantages include labour intensity, high level of expertise needed, time needed, high costs and unreliability. DNA-based techniques have received the most attention in recent years as they can be automated and consider a species at its most basic level, its genetic sequence. This is also seen in the range of DNA-based methods used for the identification of *Fusarium* species. I think the technique that warrants more attention is microarrays as it allows for the identification of large numbers of isolates and species simultaneously. However, improvements in next generation sequencing technologies (Mardis, 2008; Morozova and Marra, 2008; Schuster, 2008) should make high- throughput sequence analysis for large-scale species identifications also feasible in the near future.

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CHAPTER 2

EVALUATION OF THE INTERGENIC SPACER (IGS) REGION AS A DIAGNOSTIC MARKER FOR *FUSARIUM CIRCINATUM*

Abstract: The pathogen *Fusarium circinatum*, causal agent of pitch canker in pine (*Pinus* spp.) is a threat to both native pine forests and non-native plantations worldwide. The dissemination of the pathogen is believed to be facilitated by various factors including wind, water splash, insect vectors and silvicultural practices. Effective monitoring and detection of the pathogen is thus important for disease management. Although a variety of DNA markers are commonly used for *Fusarium* diagnostics, markers based on the ribosomal RNA intergenic spacer (IGS) are used most frequently for identification and quantification of the pitch canker fungus. The aim of this study was to evaluate the suitability of the IGS region as a diagnostic marker for identifying *F. circinatum*. For this purpose, the IGS region was sequenced and the various primer and probe target areas were compared among different isolates of the fungus. The *F. circinatum* sequences were also compared to those obtained for other *Fusarium* species. These included species that are known to occur in the same environment as *F. circinatum* (e.g., *F. oxysporum*, *F. proliferatum*), as well as species that are closely related to this fungus (e.g., *F. subglutinans*). Phylogenetic analysis revealed three distinct clades for the *Gibberella fujikuroi* complex. Single nucleotide polymorphisms (SNPs) and indels were present within the IGS region of *F. circinatum* and the other *Fusarium* species included in this study. It was, however, found that none of the polymorphisms had a significant effect on the *F. circinatum* target sites. The IGS region is, therefore, valuable not only for diagnostics but also for population studies as this region has polymorphisms that are significant at an intraspecies level.

INTRODUCTION

Pitch canker of pine is caused by the ascomycete fungus, *Fusarium circinatum* (Schweigkofler *et al.*, 2004; European and Mediterranean Plant Protection Organization, 2005) that was first described in 1946 in the southern United States (Hepting and Roth, 1946). Since then the pathogen has been reported from various pine growing regions around the world (Wikler and Gordon, 2000; Ramsfield *et al.*, 2008) where it affects a large number of *Pinus* species (EPPO, 2005; Wingfield *et al.*, 2008) and *Pseudotsuga menziesii* (Wingfield *et al.*, 2008; Ioos *et al.*, 2009). The pathogen is thought to spread via the movement of untreated wood, wood products, infected seedlings, and contaminated growth medium (Mireku and Simpson, 2002; Wingfield *et al.*, 2008), as well as pine-associated insects such as *Rhyacionia* spp. (Matthews, 1962; Runion and Bruck, 1985), *Contarina* sp. (Dwinell *et al.*, 1985) and *Brachyderes incanus* (Romón *et al.*, 2007) to name a few. Globally, *F. circinatum* is regarded

as one of the most important pathogens of *Pinus* species and has been associated with great economic losses due to seedling and tree mortality, as well as reduction in wood quality and production (Mireku and Simpson, 2002; Wingfield *et al.*, 2008).

The pitch canker pathogen was initially thought to represent a *forma specialis* of *Fusarium subglutinans* because of its resemblance to this fungus and its specific association with *Pinus* species (Britz *et al.*, 2001; Wingfield *et al.*, 2008). *Fusarium subglutinans* f. sp. *pini* was later elevated to species level and named *F. circinatum* (teleomorph = *Gibberella circinata*) (Britz *et al.*, 2001; Wingfield *et al.*, 2008), which forms part of the *G. fujikuroi* species complex (Nirenberg and O'Donnell, 1998; Britz *et al.*, 1999) belonging to mating population H (Britz *et al.*, 1999; Desjardins, 2003). The most important culture-based identification methods for *F. circinatum* include morphology (Nirenberg and O'Donnell, 1998) and sexual compatibility tests (Britz *et al.*, 1998; Britz *et al.*, 2005). However, such culture-based identification procedures are time-consuming and can lead to ambiguous identifications as different *Fusarium* species may have similar morphological characteristics (Matuo and Snyder, 1972; Burgess *et al.*, 1994; Kvas *et al.*, 2009). Consequently, all diagnoses of the pitch canker pathogen now include some sort of DNA-based procedure, as these allow for much more rapid and robust identifications.

The markers most commonly used for the diagnosis of *F. circinatum* and other *Fusarium* species include regions of protein-coding genes such as β -tubulin (O'Donnell *et al.*, 2000), histone H3 (Steenkamp *et al.*, 1999), and translation elongation factor 1-alpha (TEF) (Geiser *et al.*, 2004). Although the ribosomal RNA (rRNA) genes and spacer regions have been used to some extent for studying certain *Fusarium* species (Gaudet *et al.*, 1989; Peterson and Logrieco, 1991), their conserved (Ganley and Kobayashi, 2007) and non-orthologous nature (O'Donnell and Cigelnik, 1997) generally preclude applications in diagnostics, especially for species in the *G. fujikuroi* complex. In contrast the rRNA intergenic spacer (IGS) apparently contains various regions with species-specific polymorphisms that allow the separation of some *Fusarium* species (Kim *et al.*, 2001; Ganley and Kobayashi, 2007).

Currently, there are three DNA-based strategies available for identifying an isolate as *F. circinatum*. TEF-barcode analysis involves sequencing of a specific portion of the TEF gene and comparisons against a database of voucher sequences (Geiser *et al.*, 2004). The H3 PCR-RFLP method uses restriction endonucleases for targeting *F. circinatum*-specific polymorphisms in the histone *H3* gene (Steenkamp *et al.*, 1999; Schweigkofler *et al.*, 2004).

The third approach involves the application of *F. circinatum*-specific real-time PCR primers (Schweigkofler *et al.*, 2004; Ramsfield *et al.*, 2008; Ioos *et al.*, 2009). The latter approach has the advantages of being more sensitive and specific than conventional PCR and allows for quantification of *F. circinatum* DNA in the sample (McCartney *et al.*, 2003; Ioos *et al.*, 2009). Three sets of species-specific PCR primers are available for the pathogen (Schweigkofler *et al.*, 2004; Ramsfield *et al.*, 2008; Ioos *et al.*, 2009), of which two sets (Schweigkofler *et al.*, 2004; Ioos *et al.*, 2009) specifically target the rRNA IGS.

The multicopy nature of the IGS region makes it valuable in diagnostics. All eukaryotic genomes, including that of *F. circinatum*, harbour numerous copies (sometimes thousands) of rRNA repeat regions (Rooney and Ward, 2005; Ganley and Kobayashi, 2007) arranged head-to-tail and separated by non-coding spacers (ITS and IGS) (Kim *et al.*, 2001) (Figure 1). The sequences of these repeats are kept intact through the processes of homogenisation (Peterson and Logrieco, 1991; Simon and Weiß, 2008) and gene conversion (Peterson and Logrieco, 1991; Jeffreys and May, 2004). In general the functional rRNA gene regions tend to have limited sequence variation due to selective constraints (Boffelli *et al.*, 2004), while spacers (containing promoter and non-coding regions) are apparently more tolerant to substitutions and indels (insertions and deletions) (Britten *et al.*, 2003; Mak *et al.*, 2006; Ganley and Kobayashi, 2007). Therefore, the IGS appears to be the fastest evolving non-coding rRNA region (Kim *et al.*, 2001) resulting in this region being more variable between different species than other regions of the rRNA (Fernández *et al.*, 2000; Kim *et al.*, 2001; Llorens *et al.*, 2006; Ganley and Kobayashi, 2007).

The objective of this study was to evaluate the suitability of the rRNA IGS region as a diagnostic marker for *F. circinatum*. For this purpose, comparisons of the complete sequence of the IGS region for the various biological species in the *G. fujikuroi* complex were used. The information obtained will enable the design of *F. circinatum* specific primers and/or probes to be used in future as a routine diagnostic procedure for the detection and monitoring of the pitch canker pathogen in a specific environment.

MATERIALS AND METHODS

Fungal isolates

The strains used in this study included *F. circinatum* isolates from different geographical regions to ensure that the various species-specific primers/probes are indeed specific to this species irrespective of an isolate's origin (Table 1). *Fusarium oxysporum*, *F. proliferatum* and *F. subglutinans* (Table 1) isolates were also included as *F. oxysporum* and *F. proliferatum* are commonly found in pine nurseries and *F. subglutinans* is closely related to *F. circinatum*.

DNA extraction, PCR and sequencing

All isolates were subcultured on potato dextrose agar (PDA) (Merck, Wadeville, Gauteng) at 25 °C for 7 days. Genomic DNA was isolated from the individual *Fusarium* isolates by making use of an extraction method based on CTAB (N-Cetyl-N,N,N-Trimethyl ammonium bromide) (Merck) (O'Donnell and Cigelnik, 1997). The complete *F. circinatum* IGS region was amplified in three overlapping regions (Figure 1) with primers GCNS7r (5' GTTGTTAAGAGGCGCGGTGTC 3'), GCNS3 (5' CTGCAAAGCTGTACAGAGGG 3') (Figure 1), CIRC4A (5' ACCTACCCTACACCTCTCACT 3') (Schweigkofler *et al.*, 2004) and FCIR-F (5' TCGATGTGTCGTCTCTGGAC 3') (Ioos *et al.*, 2009). For *F. oxysporum*, *F. proliferatum* and *F. subglutinans*, the total IGS region was amplified in two overlapping regions with primers GCNS7r (5' GTTGTTAAGAGGCGCGGTGTC 3'), GCNS3 (5' CTGCAAAGCTGTACAGAGGG 3') (Figure 1), NL11 (5' CGTAACGCCTCTAAGTCAG 3') (Figure 1) and CNS1 (5' GAGACAAGCATATGACTAC 3') (Figure 1) (Schweigkofler *et al.*, 2004).

Each 25- μ l PCR mixture contained *ca.* 1 ng template DNA, 0.63 mM MgCl₂ (Roche, Germany), 0.25 mM dNTPs (Fermentas, Vilnius, Lithuania), 0.25 μ M of each primer (Inqaba Biotech, South Africa), 1.25 U/ μ l *Taq* DNA polymerase (Roche, Germany) and reaction buffer (FABI, Pretoria, South Africa). All PCRs were conducted on a BioRad iCycler (Hercules, California, USA). The cycling parameters were similar to those used by Schweigkofler *et al.* (2004) and were as follows: 94 °C for 90 s, 40 cycles of 94 °C for 30 s, 52 °C for 30 s and 68 °C for 3 min, followed by 68 °C for 5 min and 4 °C until removed. The presence of PCR products was verified by gel electrophoresis. The resulting PCR products were then purified with G50 Sephadex columns (Sigma, Steinheim, Germany) and sequenced

by making use of an ABI PRISM BigDye v3.0 Cycle Sequencing Kit (Applied Biosystems, Foster City, California) and a 3730 DNA Analyser (Applied Biosystems, Foster City, California). Raw sequence data were visualized and corrected where necessary with Chromas Lite version 2.01 (Technelysium Pty. Ltd.) and BioEdit version 7.0.5.3 (Hall, 1999).

IGS sequence analysis

The full-length IGS sequences of all the isolates were aligned using the web-based alignment tool MAFFT (multiple sequence alignment based on the fast Fourier transform) version 6 with the L-INS-I (iterative refinement method using local pairwise alignment with affine gap cost) (Kato, *et al.* 2002; <http://align.bmr.kyushu-u.ac.jp/mafft/online/server/>) function.

BioEdit was used to identify and compare within this alignment, the various sequences for PCR primers, *F. circinatum*-specific primers CIRC1A (5' CTTGGCTCGAGAAGGG 3'), CIRC4A (5' ACCTACCCTACACCTCTCACT 3') (Schweigkofler *et al.*, 2004), FCIR-F (5' TCGATGTGTCGTCTCTGGAC 3') and FCIR-R (5' CGATCCTCAAATCGACCAAGA 3') (Ioos *et al.*, 2009) and probe FCIR-P (5' CGAGTCTGGCGGGACTTTGTGC 3') (Ioos *et al.*, 2009).

The IGS alignment was extended to include the IGS sequences for species in the *Gibberella fujikuroi* complex that were included in the Schweigkofler *et al.* (2004) study. These sequences were obtained from the *Fusarium* database (<http://fusarium.cbio.psu.edu>, and included *F. anthophilum* NRRL 13602, *F. bactridioides* NRRL 20476, *F. begoniae* NRRL 25300, *F. bulbicola* NRRL 13618, *F. circinatum* NRRL 25331, *F. concentricum* NRRL 25303, *F. fractiflexum* NRRL 28852, *F. fujikuroi* NRRL 13566, *F. globosum* NRRL 26131, *F. oxysporum* NRRL 22902, *F. proliferatum* NRRL 22944, *F. pseudoanthophilum* NRRL 25206, *F. subglutinans* NRRL 22016, *F. succisae* NRRL 13613, *Fusarium* sp. NRRL 25195, *Fusarium* sp. NRRL 25204, *Fusarium* sp. NRRL 25346, *Fusarium* sp. NRRL 25622, *Fusarium* sp. NRRL 25623, *Fusarium* sp. NRRL 25807, *Fusarium* sp. NRRL 26756, *Fusarium* sp. NRRL 26757, *Fusarium* sp. NRRL 29123, *F. thapsinum* NRRL 22045, *F. verticillioides* NRRL 22172). The resulting dataset was subjected to Maximum-likelihood phylogenetic analysis using jModelTest 0.1 (Posada, 2008) and PhyML 3.0 (Guindon and Gascuel, 2003).

RESULTS

The lengths of the PCR amplicons ranged from 450 base pairs (bp) to 970 bp, depending on the primer pair used. Primer pairs NL11 and CNS1 generated fragments with an average length of 970 bp, those produced with GCNS7r and GCNS3 were on average 650 bp in length, and primers FCIR-F and CIRC4A produced fragments of about 750 bp in length. The final IGS sequences produced in this study ranged from 1029 bp in length for *F. subglutinans* (MRC 2802) to 2650 bp for *Fusarium* sp. (NRRL 25623) (see Appendix 1 for an alignment of the complete IGS).

Phylogenetic analysis of the IGS sequences separated the sequences included in this study into three well-supported clades (Figure 2). These clades correspond to the so-called “Asian”, “African” and “American” clades of O’Donnell *et al.* (1998). This analysis further revealed that all the *F. circinatum* isolates grouped together within the “American” clade (Figure 2). Within the *F. circinatum* clade there was, however, no apparent grouping of isolates according to their geographic origins. This is because isolates from Florida and South Africa group together and California, Florida, Mexico and Spain group together.

In addition to hundreds of single nucleotide polymorphisms (SNPs), comparison of all the IGS sequences included in this study revealed the presence of several indels ranging in size from 2 bp to 18 bp. The *F. circinatum* isolates harboured seven indels at positions 314-322 bp, 396-397 bp, 405-411 bp, 646-647 bp, 716-724 bp, 1157-1167 bp, 1669-1679 bp. All the *F. oxysporum* isolates had two indels that were between positions 362 and 372 and positions 389 and 399 bp in the alignment. *Fusarium succisae* (NRRL 13613) had an insertion between positions 400 and 408 bp, and a deletion between position 1161 and 1178 bp. Isolate NRRL 25206 (*F. pseudoanthophilum*) had an insertion and a single-base deletion at between positions 489 and 495 bp and at position 733 bp, respectively. Between positions 804 and 811 bp, five isolates [*F. concentricum* (NRRL 25303), *F. fractiflexum* (NRRL 28852), *F. fujikuroi* (NRRL 13566), *F. globosum* (NRRL 26131) and *F. proliferatum* (NRRL 22944)] harboured an insertion. No indels were detected among the remaining nine sequences [*F. begoniae* (NRRL 25300), *F. bulbicola* (NRRL 13618), *Fusarium* sp. (NRRL 25195), *Fusarium* sp. (NRRL 25622), *Fusarium* sp. (NRRL 25623), *Fusarium* sp. (NRRL 25807), *Fusarium* sp. (NRRL 26756), *Fusarium* sp. (NRRL 26757) and *F. verticillioides* (NRRL 22172)].

Upon closer inspection of the *F. circinatum* sequences, it was apparent that no intra-specific polymorphisms severely affected the target sites for the primers and probes examined here. This was the case for the 29 SNPs, as well as the indel (between positions 459 and 518 bp) contained within the sequence of the *F. circinatum* isolate from Spain. The Ioos *et al.* (2009) species-specific primers and probe were identical to IGS sequence motifs of all the *F. circinatum* isolates included in this study. The only exceptions were *F. circinatum* isolate CMW 28961 (that harboured a deletion in part of the target region for FCIR-R) and isolate FCC 2265 that only had 14 out of a total of 21 nucleotides identical to primer FCIR-R (Figure 3). Also, the sequence of FCIR-F was identical to the corresponding region in *F. bactridioides*, *F. begoniae*, *F. bulbicola* and *F. succisae*. The sequence of FCIR-R was identical to the corresponding region in *F. subglutinans* and 18 of its 21 nucleotides matched those in the target region of the *F. anthropilum* IGS. The sequences of the Schweigkofler *et al.* (2004) species-specific primers were identical to the corresponding IGS motifs of all the *F. circinatum* isolates included in this study (Figure 4). However, one *F. subglutinans* isolate (MRC 8553) harboured an IGS sequence motif identical to CIRC1A, and CIRC4A was also identical to the sequences of *F. subglutinans* (MRC 2082) and *F. succisae*.

DISCUSSION

Interspecies variation within the IGS region of *Fusarium* species is common (Appel and Gordon, 1995; Kim *et al.*, 2001; Schweigkofler *et al.*, 2004) and was also observed in this study. The phylogenetic tree, based on the IGS data, resembled the one produced by Schweigkofler *et al.* (2004), which was also similar to those inferred from TEF (Geiser *et al.*, 2005) and β -tubulin (O'Donnell *et al.*, 2000; Geiser *et al.*, 2005) sequences. In all cases, the *Fusarium* isolates that were included were separated into the three expected clades. In cases where there were exceptions (e.g., the position of *F. concentricum*, *F. fractiflexum* and *F. fujikuroi*) the relevant nodes lacked bootstrap-support. However, the IGS phylogeny produced in the present study was markedly different from those that have been previously inferred from rRNA ITS data (O'Donnell *et al.*, 2000). Although some groupings were supported by the IGS and ITS data (e.g. the sistergroup relationship between *F. circinatum* and *F. subglutinans*), they merely reflected the fact that the species in the group harbour orthologous ITS regions (O' Donnell *et al.*, 2000). Therefore, the overall phylogenetic history of the two *G. fujikuroi* complex rRNA regions is not congruent, because the IGS tree is concordant with those inferred from of the commonly used protein-coding genes. The IGS

region thus represents an additional and easily exploitable region for phylogenetic studies of *Fusarium* species in this complex.

Significant levels of polymorphism in the IGS were detected at the intraspecies level. This is consistent with what has been found for other species, especially *F. oxysporum* (Appel and Gordon, 1995; Kim *et al.*, 2001). The results presented here, therefore, suggest that, in addition to being useful for species diagnostic purposes, that the IGS region also holds valuable information at the population level.

Another important aspect of this study was to evaluate the specificity of the existing *F. circinatum*-specific primers. The so-called “American” species *F. bactridioides*, *F. begoniae*, *F. bulbicola* and *F. succisae* all had sequences identical to FCIR-F. The same was also true for *F. subglutinans* that had sequences identical to FCIR-R (Ioos *et al.*, 2009). Overall, the *F. circinatum*-specific primers and probes were all complementary to at least one isolate that was not *F. circinatum*, although both primers of a set were never complementary to the target regions in the IGS of one non-*F. circinatum* isolate. It is thus reasonable to expect that the application of these primers and probes will allow for the detection of *F. circinatum* only, although positive diagnoses would have to be verified.

A possible disadvantage of the Schweigkofler *et al.* (2004) method is that it is dependent on the interaction of only two oligonucleotides with the target DNA during PCR. In contrast, the Ioos *et al.* (2009) method requires that two oligonucleotide primers and the oligonucleotide probe interact with the target DNA regions, thus increasing the specificity of the reaction. Also, despite the fact that the CIRC1A and CIRC4A primers were complementary to all the *F. circinatum* isolates, the CIRC1A was identical to one *F. subglutinans* (MRC 8553) isolate and CIRC4A to another *F. subglutinans* (MRC 2802) isolate. Although *F. subglutinans* is a pathogen of maize and not expected to be found in the same niche as *F. circinatum*, the possibility exists that *F. subglutinans* may be detected with the *F. circinatum*-specific primers. This is especially true in regions where pine and maize growing areas overlap.

To ensure the specificity of the *F. circinatum* diagnosis, Schweigkofler *et al.* (2004) included in their study additional “American” clade species (*F. anthophilum*, *F. bacteridioides*, *F. begoniae*, *F. bulbicola*, *F. subglutinans* and *F. succisae*). They found that at an annealing temperature of 66 °C or higher produce the most accurate results because this temperature allows the amplification of a 360 bp fragment from the isolates for *F. circinatum* only. Sub

optimum diagnostic PCRs, may therefore yield false positives due to the presence of species in the *G. fujikuroi* complex that belong to the American clade. In such instances, sequencing as an additional step may be a crucial requirement for diagnoses. Alternatively, different combinations of the primers used by Ioos *et al.* (2009) and Schweigkofler *et al.* (2004) could be used or new primers may be developed to prevent the occurrence of false positives during the diagnostics of *F. circinatum*.

DNA-based diagnostics are important in the identification of *F. circinatum*, to overcome the problems associated with culture-based methods. The results of this study demonstrate that the IGS region is an excellent marker for *Fusarium* (including *F. circinatum*) diagnostics due to the fact that it contains numerous polymorphisms, many of which are species-specific. Undoubtedly, methods based on the IGS region will remain essential tools in the routine identification, monitoring and quantification of *F. circinatum* in the commercial forestry environment.

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<http://fusarium.cbio.psu.edu> (FUSARIUM-ID v. 1.0)

FIGURES AND TABLES

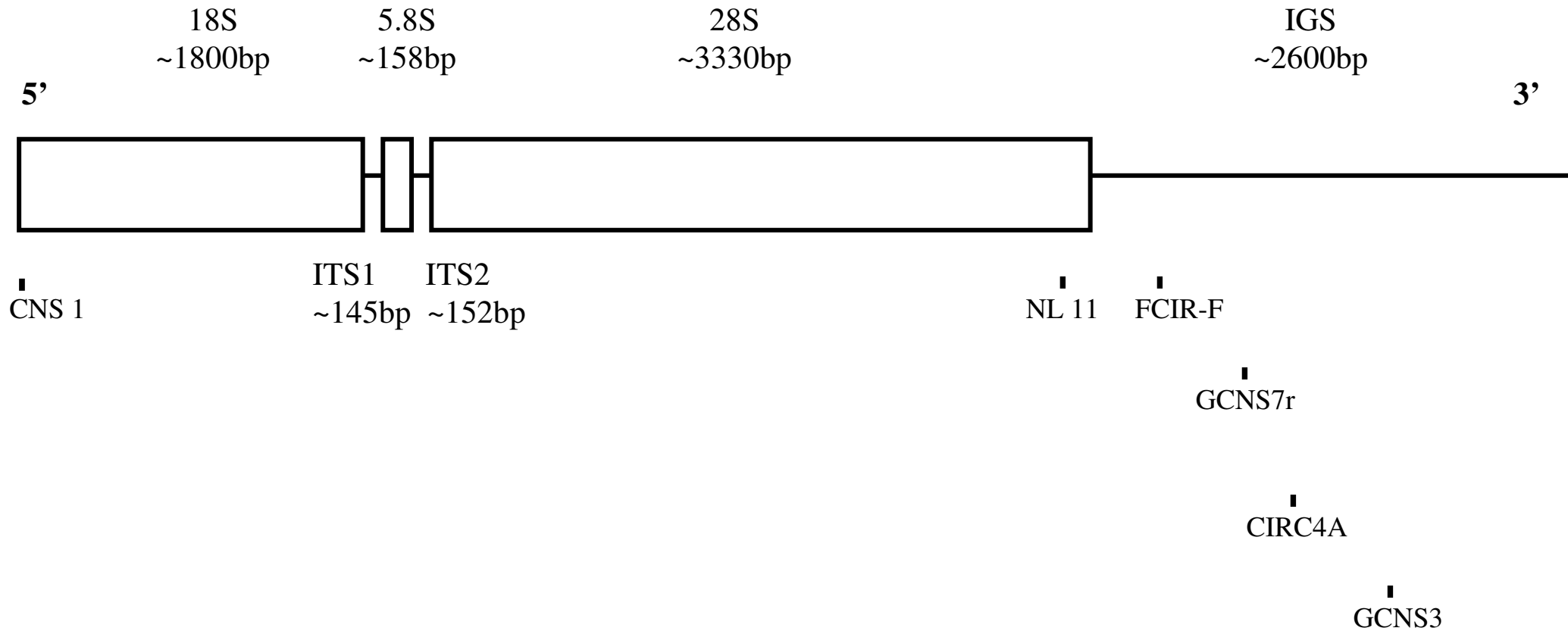


FIGURE 1: Ribosomal RNA region including the non-coding regions (on scale) and the regions where the primers used in this study attach. (Appel and Gordon, 1995; Burton *et al.*, 2005; Kane and Rollinson, 1998; Kim *et al.*, 2001; Lafontaine and Tollervey, 2001)

TABLE 1: *Fusarium* isolates included for IGS region sequences

| Isolate | Isolate number | Origin | Collector(s) |
|------------------------|----------------------------|------------------|--------------------------------------|
| <i>F. circinatum</i> | CMW 3198 ^a | USA (California) | G. Thomas |
| <i>F. circinatum</i> | FCC 2472 ^a | USA (California) | Unknown |
| <i>F. circinatum</i> | FCC 2483 ^a | USA (Florida) | Unknown |
| <i>F. circinatum</i> | FCC 2496 ^a | USA (Florida) | Unknown |
| <i>F. circinatum</i> | FCC 2497 ^a | USA | Unknown |
| <i>F. circinatum</i> | FCC 2248 ^a | Mexico | Unknown |
| <i>F. circinatum</i> | FCC 2265 ^a | Mexico | Unknown |
| <i>F. circinatum</i> | CMW 23565 ^a | Mexico | Unknown |
| <i>F. circinatum</i> | FCC 0010 ^a | South Africa | Unknown |
| <i>F. circinatum</i> | FCC 0041 ^a | South Africa | Unknown |
| <i>F. circinatum</i> | FCC 1834 ^a | South Africa | G. Hunter |
| <i>F. circinatum</i> | CMW 28961 ^a | Spain | E. Iturrityxa |
| <i>F. oxysporum</i> | CAV 105 ^{a†} | South Africa | A. Viljoen |
| <i>F. oxysporum</i> | CAV 602 ^{a†} | Australia | Unknown |
| <i>F. oxysporum</i> | CAV 617/1107 ^{a†} | Vietnam | I. Buddenhagen, N. Moore, S. Bentley |
| <i>F. oxysporum</i> | CAV 794 ^{a†} | Indonesia | Unknown |
| <i>F. oxysporum</i> | CAV 929 ^{a†} | Philippines | L. Magnaye |
| <i>F. proliferatum</i> | CMW 12864 ^a | USA | J. F. Leslie |
| <i>F. proliferatum</i> | CMW 12867 ^a | USA | J. F. Leslie |
| <i>F. proliferatum</i> | MRC 2301 ^a | USA (California) | J. Leslie |
| <i>F. proliferatum</i> | MRC 8549 ^a | USA (California) | Unknown |
| <i>F. proliferatum</i> | MRC 8550 ^a | Unknown | Unknown |
| <i>F. subglutinans</i> | MRC 8553 ^a | Unknown | Unknown |
| <i>F. subglutinans</i> | MRC 8554 ^a | USA (Illinois) | Unknown |
| <i>F. subglutinans</i> | MRC 6512 ^a | USA (Illinois) | J. F. Leslie |
| <i>F. subglutinans</i> | MRC 2802 ^a | South Africa | C. Schreuder |

^a*Fusarium* collection of the Tree Pathology Co-operative Programme (TPCP), Forestry and Agricultural Biotechnology Institute (FABI), University of Pretoria, Pretoria, South Africa

[†]Fourie *et al.*, (2009)

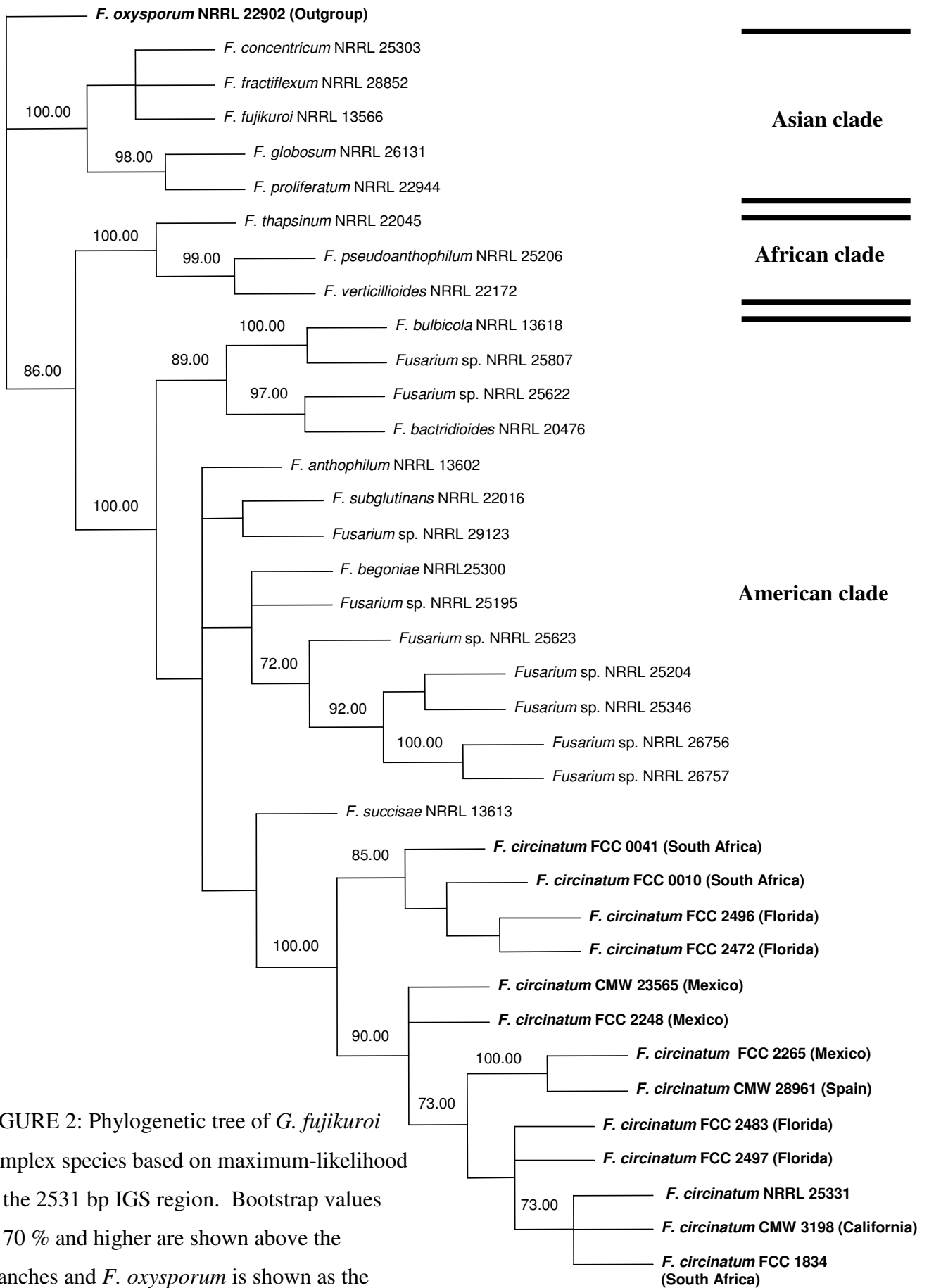


FIGURE 2: Phylogenetic tree of *G. fujikuroi* complex species based on maximum-likelihood of the 2531 bp IGS region. Bootstrap values of 70 % and higher are shown above the branches and *F. oxysporum* is shown as the outgroup.

| | 302 | 321 | 379 | 399 | 446 | 466 |
|--|------------------------|------|-----------------------|-------|-----------------------|-------|
| <i>F. circinatum</i> NRRL 25331 | TCGA-TGTGTCGTCTCTGGAC | --- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> CMW 3198 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 2483 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 2497 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> CMW 23565 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FI 120 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 0041 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 2248 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 0010 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 2472 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 1834 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> FCC 2265 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. circinatum</i> CMW 28961 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCGGGACTTTGTG | ---- | TCTTGGTCGAT | ----- |
| <i>F. anthropophilum</i> NRRL 13602 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. bactridioides</i> NRRL 20476 | TCGA-TGTGTCGTCTCTGGAC | ---- | TGAGTCTGGCAGGACTGTGTG | ---- | TCTTGGTCGATTTGGGGAGTG | |
| <i>F. begoniae</i> NRRL 25300 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. bulbicola</i> NRRL 13618 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. concentricum</i> NRRL 25303 | ---GA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. fractiflexum</i> NRRL 28852 | ---GA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. fujikuroi</i> NRRL 13566 | ---GA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. globosum</i> NRRL 26131 | ---GA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. oxysporum</i> NRRL 22902 | ---GA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGT | ----- | ACATGGTCGATTTGAGGATCG | |
| <i>F. proliferatum</i> NRRL 22944 | ---GA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. pseudoanthophilum</i> NRRL 25206 | TCGA-GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. subglutinans</i> NRRL 22016 | TCGATTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. succisae</i> NRRL 13613 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 25195 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 25204 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 25346 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 25622 | TCGA-TGTGTCGTCTCTGGAC | ---- | TGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 25623 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 25807 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 26756 | TCGA-TGCGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 26757 | TCGA-TGCGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>Fusarium</i> sp. NRRL 29123 | TCGA-TGTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. thapsinum</i> NRRL 22045 | TCGA-GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. verticillioides</i> NRRL 22172 | TCGA-GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. oxysporum</i> CAV 105 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGT | ----- | ACATGGTCGATTTGAGGATCG | |
| <i>F. oxysporum</i> CAV 602 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGT | ----- | ACATGGTCGATTTGAGGATCG | |
| <i>F. oxysporum</i> CAV 617 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGT | ----- | ACATGGTCGATTTGAGGATCG | |
| <i>F. oxysporum</i> CAV 794 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGT | ----- | ACATGGTCGATTTGAGGATCG | |
| <i>F. oxysporum</i> CAV 929 | C-GATGTCGTCTCTGGAC | ---- | GAGTCTGGT | ----- | ACATGGTCGATTTGAGGATCG | |
| <i>F. proliferatum</i> CMW 12864 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. proliferatum</i> CMW 12867 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. proliferatum</i> MRC 8549 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. proliferatum</i> MRC 2301 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. proliferatum</i> MRC 8550 | ---GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCATGGCTGTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. subglutinans</i> MRC 8553 | TCGATTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. subglutinans</i> MRC 8554 | TCGATTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. subglutinans</i> MRC 6512 | TCGATTGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| <i>F. subglutinans</i> MRC 2802 | TC-GATGTCGTCTCTGGAC | ---- | CGAGTCTGGCAGGACTTTGTG | ---- | TCTTGGTCGATTTGAGGATCG | |
| FCIR-F | TCGA-TGTGTCGTCTCTGGAC | | | | | |
| FCIR-P | | | CGAGTCTGGCGGGACTTTGTG | | | |
| FCIR-R | | | | | TCTTGGTCGATTTGAGGATCG | |

FIGURE 3: *Fusarium* species alignment for the portions of the IGS that harbour the Ios *et al.* (2009) oligonucleotides. Numbers at top of Figure indicate the position on the IGS; – Indicates sequence gaps or unknown nucleotides.

| | 1356 | 1371 | 1699 | 1719 |
|--|-------------------|-------|------------------------|------|
| <i>F. circinatum</i> NRRL 25331 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> CMW 3198 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 2483 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 2497 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> CMW 23565 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 2496 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 0041 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 2248 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 0010 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 2472 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 1834 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> FCC 2265 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. circinatum</i> CMW 28961 | CTTGGCTCGAGAAGGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>F. anthophilum</i> NRRL 13602 | CCTTGCTCGAGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>F. bactridioides</i> NRRL 20476 | CCTTGCTCGAGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>F. begoniae</i> NRRL 25300 | CCTTGCTCGAGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>F. bulbicola</i> NRRL 13618 | CTTGGCTCGAGAAACG | ---- | AGGAAGTGGTGTAGGGTAGGT | |
| <i>F. concentricum</i> NRRL 25303 | CCTTGCTCGAGAAACG | ---- | TTGATGTGGTATAGGGTAGGT | |
| <i>F. fractiflexum</i> NRRL 28852 | CCTTGCTCGAGAAACG | ---- | TTGAAAGTGGTATAGGGTAGGT | |
| <i>F. fujikuroi</i> NRRL 13566 | CCTTGCTCGAGAAACG | ---- | TTGAAAGTGGTATAGGGTAGGT | |
| <i>F. globosum</i> NRRL 26131 | CCTTGCTCGAGAAACG | ---- | TTGAAAGTGGTATAGGGTAGGT | |
| <i>F. oxysporum</i> NRRL 29202 | CCTTGCTCGAGAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. proliferatum</i> NRRL 22944 | CCTTGCTCGAGAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. pseudoanthophilum</i> NRRL 25206 | CCTTGCTCGAGAAACG | ---- | ATGCAGAGGTATAGGGTAGGT | |
| <i>F. subglutinans</i> NRRL 22016 | CCTTGCTCGAGAAACG | ---- | GGGGAGAGGTGTAGGGTAGGT | |
| <i>F. succisae</i> NRRL 13613 | CCTTGCTCGAGAAACG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 25195 | CTTTG--CGAGAAACG | ---- | AGGGAGAGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 25204 | CTTTGCACCGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 25346 | CTTTGCACCGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 25622 | CCTTGCTCGAGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 25623 | CCTTGCTCGAGAAACG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 25807 | CCTTGCTCGAGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 26756 | CCTTGCTCGAGAAACG | ---- | AGGGAGTGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 26757 | CCTTGCTCGAGAAACG | ---- | AGGGAGTGGTGTAGGGTAGGT | |
| <i>Fusarium</i> sp. NRRL 29123 | CCTTGCTCGAGAAACG | ---- | AGGTAGTGGTGTAGGGTAGGT | |
| <i>F. thapsinum</i> NRRL 22045 | TTTTGCTCGAGAAACG | ---- | ATGCAGAGGTATAGGGTAGGC | |
| <i>F. verticillioides</i> NRRL 22172 | CCTTGCTCGAGAAACG | ---- | GTGTGGAGGTATAGGGTAGGT | |
| <i>F. oxysporum</i> CAV 105 | --TTACCCGAGGGAGG | ---- | ATGTGGTGGTGTAGGGTAGGT | |
| <i>F. oxysporum</i> CAV 617 | TTTTACTCGAGGGAGG | ---- | ATGTGGTGGTGTAGGGTAGGT | |
| <i>F. oxysporum</i> CAV 794 | TTTTACTCGAGGGAGG | ---- | ATGTGGTGGTGTAGGGTAGGT | |
| <i>F. oxysporum</i> CAV 602 | TTTTATTCAAGGGAGG | ---- | ATGTGGTGGTGTAGGGTAGGT | |
| <i>F. oxysporum</i> CAV 929 | TTTTATTCAAGGAAGG | ---- | ATGTGGTGGTGCAGGGTAGGT | |
| <i>F. proliferatum</i> CMW 12864 | TTTTGTTTGA AAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. proliferatum</i> MRC 2301 | TTTTGTTTGA AAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. proliferatum</i> MRC 12867 | TTTTGCTTGA AAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. proliferatum</i> MRC 8550 | TTTTGCTTGA AAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. proliferatum</i> MRC 8549 | TTTTGCTTGA AAAACG | ---- | TTGAGGTGGTATAGGGTAGGT | |
| <i>F. subglutinans</i> MRC 8553 | CTTGGCTCGAGAAGGG | ---- | GGGGAGAGGTGTAGGGTAGGT | |
| <i>F. subglutinans</i> MRC 8554 | ----- | ----- | GGGGAGAGGTGTAGGGTAGGT | |
| <i>F. subglutinans</i> MRC 6512 | CTTGGCTCGAAAAGGG | ---- | GGGGAGAGGTGTAGGGTAGGT | |
| <i>F. subglutinans</i> MRC 2802 | CCTTGAAAAGAAAGG | ---- | AGTGAGAGGTGTAGGGTAGGT | |
| CIRC1A | CTTGGCTCGAGAAGGG | | | |
| CIRC4A | | | AGTGAGAGGTGTAGGGTAGGT | |

FIGURE 4: *Fusarium* species alignment for the positions of the IGS that harbour the Schweigkofler *et al.* (2004) oligonucleotides. Numbers at top of Figure indicate the position on the IGS; – Indicates sequence gaps or unknown nucleotides.

CHAPTER 3

MONITORING SPORE LOADS OF THE PITCH CANKER FUNGUS, *FUSARIUM CIRCINATUM*, IN A PINE SEEDLING NURSERY

Abstract: Detection and quantification are important steps in developing management strategies for pathogens. Many different techniques can be used to quantify the presence and concentrations of viruses, bacteria and fungi. Plant pathogens cause great economic losses, and these include *Fusarium circinatum* that causes a serious canker disease of pines. Two previous studies were conducted that focused on the quantification of *F. circinatum* spores in a native pine forest and one study that focused on the detection of this pathogen on pine seeds. In all three of these studies, real-time PCR was used for quantification or detection. The advantage that real-time PCR offers compared to conventional PCR is that the technique is capable of determining the original amount of DNA for organisms present at the start of the PCR cycle. The quantity can provide an indication of the extent of the infection and the steps that might be appropriate for its management. This study used real-time PCR to determine the spore concentrations of *F. circinatum* in a pine seedling nursery over a 12-month period and considered whether temperature had an effect on the spore concentration. The real-time PCR technique was successful in quantifying spore concentrations and it appears that temperature has an effect on spore concentrations, although the effect was less obvious than expected.

INTRODUCTION

Fusarium circinatum is an important pathogen of *Pinus* species worldwide (Gordon, 2006; Wingfield *et al.*, 2008). This pathogen can infect all parts of the host plant including the main trunk, branches, cones, seeds, and roots (Wingfield *et al.*, 2008). Typical symptoms include branch dieback, development of resinous cankers at the point of infection (Wingfield *et al.*, 2008) and the infection of roots that results in damping-off and root or collar disease (Wingfield *et al.*, 2008).

The dispersal of spores of the pitch canker pathogen is mediated by a variety of factors. These include wind, water splash, insect vectors and transmission through infected seed (Britz *et al.*, 2001; Wingfield *et al.*, 2008). Irresponsible silvicultural and nursery practices can also lead to the introduction of the pathogen into regions that were previously free of pitch canker (Wingfield *et al.*, 2008). Therefore the combination of sterile practises in nurseries, effective quarantine strategies and early identification of the pathogen are important for preventing the dispersal of the pathogen (European and Mediterranean Plant Protection Organization, 2005; Wingfield *et al.*, 2008).

In order to monitor the presence of a pathogen in a specific environment, a variety of detection techniques are available. These include antibody-based techniques (Ward *et al.*, 2004) and various nucleic acid-based techniques (McCartney *et al.*, 2003; Ward *et al.*, 2004). Of the latter group, real-time PCR methods based on non-specific fluorescent dyes such as SYBR Green and hydrolysis probes using TaqMan technology (Bowyer, 2007) are most commonly used. A typical real-time PCR determines the amount (number of copies) of amplicons after each cycle during PCR measured by fluorescence, which is proportional to the number of amplicons generated (Mackay *et al.*, 2002; Bustin, 2005). The cycle where the overall emission of fluorescence shifts from reflecting background noise to that of the actual DNA target is known as the threshold cycle (C_t). By making use of this C_t -value, the original amount of target DNA in the reaction can be determined (Mackay *et al.*, 2002; McCartney *et al.*, 2003; Bustin, 2005). Real-time PCR methods thus provide both qualitative and quantitative data and may be used to detect and quantify a pathogen in a specific environment.

Real-time PCR methods for the detection and quantification of pathogens in a specific environment have various advantages. These methods are exceptionally sensitive (Klein, 2002) and they can be conducted relatively rapidly (Burlakoti *et al.*, 2007; Ciglenc̆ki *et al.*, 2008; Bustin *et al.*, 2009). Most importantly, however, real-time PCR methods are not dependent on culturing of the target organism(s) (Nicolaisen *et al.*, 2009), provided that appropriate PCR targets and suitable DNA isolation methods are available. For *Fusarium* species, various DNA targets and DNA isolation methods are available for use in real-time PCR methodologies. Commonly used DNA targets include the ribosomal RNA intergenic spacer (IGS) region (Schweigkofler *et al.*, 2004; Ioos *et al.*, 2009) and the genes encoding *Tri5* (Burlakoti *et al.*, 2007), translation elongation factor 1 α (Nicolaisen *et al.*, 2009) and beta-tubulin (Reischer *et al.*, 2004). Popular DNA isolation methods include those using commercial kits (e.g., GeneClean and Qiagen DNeasy Minikit®) (Wallace and Covert, 2000; Schweigkofler *et al.*, 2004; Ioos *et al.*, 2009; Nicolaisen *et al.*, 2009) and those using surfactants such as hexadecyltrimethylammonium bromide (CTAB) (Lee and Taylor, 1990) and DNA isolation methods based on phenol-chloroform extraction (Atkins and Clark, 2004).

The use of real-time PCR methods based on SYBR Green and other non-specific DNA binding dyes (Bustin, 2005; Heid *et al.*, 2010) has several advantages over those based on hydrolysis probes and TaqMan technology. The most important of these is that SYBR Green-based methods are more flexible and cost effective (Bowyer, 2007). Although hydrolysis

probes are more specific because they only bind to target DNA (Sarlin *et al.*, 2006; Burlakoti *et al.*, 2007), these probes are usually expensive to design and produce (Bowyer, 2007). Although, the SYBR Green-based methods do not require these probes (Newby *et al.*, 2003), the use of SYBR Green and similar dyes for detection and quantification purposes are associated with a number of important limitations. For example, non-optimal PCRs may produce primer-dimers with which the non-specific dye will associate, thus resulting in false positives of inaccurate quantifications (Mackay *et al.*, 2002). Therefore, optimal PCR and cycling conditions are typically used in combination with melting curve analysis after real-time PCR (Bustin, 2005). The latter allows for differentiation between double stranded DNA of the target amplicons and primers-dimers (Mackay *et al.*, 2002; Bowyer, 2007), as primer-dimers are usually associated with lower melting temperatures (Bowyer, 2007; Ciglenc̆ki *et al.*, 2008).

The objective of this study was to develop a cost effective real-time PCR procedure to detect and quantify the spores of *F. circinatum* in the nursery environment. For this purpose, the specific aims were threefold: (i) to evaluate four widely used extraction procedures for their efficiency in isolating good-quality DNA from spores that have been deposited on filter paper traps, (ii) to design a real-time PCR protocol for the routine quantification of *F. circinatum* spores trapped in a pine seedling nursery, and (iii) to use this method to monitor the spore loads in the nursery environment over a 12-month period.

MATERIALS AND METHODS

Experimental pine nursery and spore traps

Two spore trap structures were placed in the experimental greenhouse, one at the front and the other the back of the greenhouse (12 x 15 m) based at the University of Pretoria experimental farm, Forestry and Agricultural Biotechnology Institute (FABI) nursery, Pretoria, South Africa. This greenhouse is used for routine pathogenicity tests with *F. circinatum* to evaluate the susceptibility of different pine species, families and hybrids to pitch canker. For these tests seedlings were inoculated with various strains (FCC 3977, FCC 3978 and FCC 3979), stored in the *Fusarium* collection of the Tree Protection Co-operative Programme (TPCP), at FABI, University of Pretoria, Pretoria, South Africa. Three 70 mm Whatman No.1 filter paper discs (Whatman BioSystems Ltd., Maidstone, Kent, England) were attached to each of

the spore trap structures at heights of 0.2 m, 0.8 m (tray level) and 2.0 m above ground level. These filter papers served as solid surfaces onto which the aerial spores of the pitch canker pathogen (derived from pathogenicity tests conducted at the time) could be deposited. All six of the spore traps were replaced every two weeks for a total period of 52 weeks by first wetting the filter paper with 4xTE buffer (pH 8; 40 mM Tris HCl, 4 mM EDTA) to increase spore-trapping capability. The minimum and maximum temperatures were recorded every second week.

Preparation of DNA standards

To determine the most efficient DNA isolation procedure and to generate standards for use in the real-time PCR runs, three isolates of *F. circinatum* were used. These were FCC 2254, FCC 2265 and FCC 2472; all obtained from the *Fusarium* collection of the TPCP. Other *Fusarium* species were not included, as the specificity of the CIRC1A and CIRC4A primers were evaluated in Chapter 2 and found to be specific for *F. circinatum*. The *F. circinatum* isolates were inoculated onto ½ strength potato dextrose agar (PDA, Merck, Wadeville, Gauteng) medium. After 7 days of incubation at 25 °C, cultures were flooded with 4 ml of autoclaved distilled water (dH₂O) and swirled to loosen spores. Spores were then quantified using a haemocytometer (Marienfeld, Germany) after which a serial dilution of 10⁰ spores/ml, 10¹ spores/ml, 10² spores/ml, 10³ spores/ml, 10⁴ spores/ml, 10⁵ spores/ml, 10⁶ spores/ml and 10⁷ spores/ml was prepared. For each of these dilutions, 4.5 ml dH₂O was used to flood a 70 mm Whatman No.1 filter paper, which was left to air dry for 2 hours.

DNA isolation from spores

Individual spore traps designed for the preparation of DNA standards were divided into quarters, with each quarter to be used for one of four DNA isolation protocols: (i) the Schweigkofler *et al.* (2004) method; (ii) the Walsh *et al.* (1991) method; (iii) the Aljanabi and Martinez (1997) method and (iv) a modified version of the O'Donnell and Cigelnik (1997) method. The Schweigkofler *et al.* (2004) method involves washing the spores from the quarter filter paper spore trap and harvesting by spinning at a low centrifugal force (1000 rcf). The harvested spores were then disrupted by homogenisation with glass beads after which DNA was isolated using the standard CTAB treatments and phenol-chloroform-isoamyl alcohol, which are briefly as follows. Filter paper spore traps were washed with 20 ml 4xTE buffer (65 °C) to remove the spores. The spores were resuspended in the 4xTE buffer by

vortexing at a high speed for 5 min followed by centrifugation (1000 rcf; 90 min) after the removal of the filter paper. The supernatant (top layer) was removed and DNA extracted from the pellet (100 µl that remained in the tube) by homogenising with a glass bead for 30 s. A sample was subjected to three freeze-thaw cycles (2 min on ice, 2 min at 75 °C, extending the last thaw to 30 min) after the addition of 300 µl CTAB extraction buffer. Phenol-chloroform-isoamyl alcohol (350 µl) was added to the tube and 250 µl of the supernatant was removed and placed into a new tube. The sample was finally purified using a GeneClean kit (Bio 101, Inc. Carlsbad, California). A volume of 3 µl of the supernatant was used directly for PCR.

The Walsh *et al.* (1991) method involved using a cation chelating agent, Chelex 100 Resin (Biorad, Hercules, USA). Briefly, this method entails the addition of the quarter filter paper spore trap to a total volume of 50 ml consisting of 5 g of the Chelex 100 Resin and sterilised distilled water. The sample with the Chelex slurry was then vortexed for 15 s at medium speed (speed 5) followed by centrifugation at a high speed (12 100 rcf) using a microcentrifuge for 30 s. The sample was incubated for 20 min at 95 °C, and vortex for 15 s. Finally, the sample was centrifuged (12 100 rcf) for 60 s. Of the final supernatant, 3 µl was used directly in PCR.

The Aljanabi and Martinez (1997) method for isolating DNA from the spores found on a quarter filter paper spore trap entails the homogenisation of the filter paper spore trap for 15 s in 400 µl of a salt buffer. This method entails incubation of the samples at 65 °C for 1 h after the addition of 40 µl 20 % sodium dodecyl sulfate (SDS) and 8 µl proteinase K (20 mg/ml). To each sample 300 µl NaCl (6 M) was added and vortexed for 30 s at maximum speed. The final volume supernatant of 100 µl was transferred to a new tube after the centrifugation of the sample for 30 min at 10 000 rcf. Hundred µl of isopropanol was added to the supernatant, mixed and incubated for 1 h at -20 °C. The precipitated nucleic acids were harvested by centrifugation for 20 min at 4 °C and 10 000 rcf. The pellet was washed with 70 % ethanol, dried and resuspended in 40 µl dH₂O. A volume of 3 µl of the supernatant was used directly for PCR.

The fourth method was a modification of the standard CTAB and phenol-chloroform isolation method as previously described by O'Donnell and Cigelnik (1997). This method entailed the addition of a quarter filter paper spore trap and glass beads to a 2 ml screw-cap tube. Also added to the tube was 15 µl proteinase K (10 µg/µl) and 750 µl TES buffer. Following

homogenisation in a homogeniser for 10 s, the tube was incubated for 1 h at 60 °C with occasional vortexing. After the addition of 270 µl NaCl (5 M) and 120 µl CTAB (10 % volume), the tube was again vortexed and incubated for 10 min at 65 °C. Following centrifugation (30 min; 4 °C; 19 357 rcf), the supernatant (400 µl) was removed and placed in a new tube. After the addition of 400 µl phenol-chloroform, the tube was vortexed and centrifuged for 30 min at 4 °C and 19 357 rcf. After repeating the phenol-chloroform extraction on 300 µl of the supernatant, a chloroform extraction was performed on 200 µl of the resulting supernatant to remove residual phenol. From the latter tube, 100 µl of the supernatant was transferred into a new tube and 60 µl isopropanol added to the supernatant, after which the sample was incubated overnight at -20 °C. The precipitated nucleic acids were then harvested by centrifugation (30 min; 4 °C; 19 357 rcf), washed with 70 % ethanol, air-dried and suspended in 40 µl dH₂O. A volume of 3 µl of the supernatant was used directly for PCR.

Real-time PCR

The first step to prepare a successful real-time PCR reaction is to determine the appropriate concentration and/or volumes of the fluorescent dye, DNA and primers as well as the annealing temperature. The volumes of fluorescent dye ranged from 6-10 µl and DNA from 1-3 µl. The primer mix concentrations ranged from 0.2-1 µM and the annealing temperature from 60-65 °C. The optimum real-time PCR reaction consisted of 3 µl DNA (higher volumes of DNA resulted in inconsistent amplification and lower volumes resulted in no amplification of lower spore concentrations), 0.2 µM primer mix (higher concentrations of primers mix allowed the generation of primer-dimers) and 10 µl fluorescent dye made up to a total volume of 20 µl with distilled water. The annealing temperature ranged from 60 to 65 °C. All of the temperatures allowed the amplification of *F. circinatum*, however, only at 65 °C were the PCR reaction specific only for *F. circinatum*.

The four DNA isolation protocols were compared by conducting three real-time PCR runs on each standard. All the filter paper spore traps underwent four real-time PCR runs (using each quarter as a replicate) and the average of the four runs was calculated for a final spore concentration. Each real-time PCR reaction (20 µl) contained 10 µl SsoFastTMEvaGreen® Supermix (BioRad, Hercules, California, USA), 0.75 µl CIRC1A (5' CTTGGCTCGAGAAGGG 3') and CIRC4A (5' ACCTACCCTACACCTCTCACT 3') (Schweigkofler *et al.*, 2004) primer

mix [Primer mix: 8 μ l of 10 mM CIRC1A (Inqaba Biotech, South Africa), 8 μ l of 10 mM CIRC4A (Inqaba Biotech, South Africa) and 4 μ l dH₂O], 3 μ l DNA and 6.25 μ l dH₂O.

All real-time PCR runs were performed on a BioRad CFX 96 real-time machine (Hercules, California, USA) with the reaction conditions as follows: 95 °C for 10 min, 44 cycles (95 °C for 10 s, 65 °C for 15 s, and 72 °C for 30 s). Amplification was followed by a melting curve analysis for which the reaction conditions were as follows: 95 °C for 1 min, 40 °C for 1 min and 65 °C to 95 °C with a 0.1 °C increment. A computer with a Windows XP/Vista operating system enabled analysis of the C_t and melting curve using the CFX Manager software (Hercules, California, USA). C_t was determined according to the relative fluorescence for each well at each cycle (McCartney *et al.*, 2003). The melting temperature was determined by making use of the decrease in fluorescence with the increase in temperature.

RESULTS

DNA isolation from spores

All the DNA isolation protocols proved to be highly efficient in isolating DNA from the filter papers inoculated with *F. circinatum* spores. They all allowed detection of spore concentrations among the standards to as low as 10³ spores/ml. The only exception was the Schweigkofler *et al.* (2004) protocol for which spores were only detected at 10⁷ spores/ml (Table 1). Both the modified CTAB method and the Aljanabi and Martinez (1997) protocol allowed detection of spore concentrations from 10² through to 10⁷ spores/ml, but took longer to conduct than the Walsh *et al.* (1991) protocol (Table 1). Thus the protocol chosen for DNA isolation of the naturally infected spore traps was the Walsh *et al.* (1991) method due to the protocol's sensitivity and the shorter time needed for DNA isolation.

Real-time PCR

Melting curve analysis was conducted to ensure that positive results were due to target DNA amplification and not due to primer-dimers. Target DNA and primer-dimers were distinguished using melt peaks that plot the regression of fluorescence (reduction in fluorescence) versus temperature. A positive peak is higher/above the threshold bar; a peak at a higher temperature indicating the target DNA and a peak at a lower temperature indicating a primer-dimer due to the primer's lower melting temperature. A positive peak at a higher temperature was thus considered a positive result.

Monitoring *F. circinatum* in a pine seedling nursery

Over the entire 52-week monitoring period, no spores were detected on any of the six spore traps during nine of the fortnightly sample collections, or the spore concentrations were too low to be detected. During eleven of the two-weekly sampling occasions, the average spore concentration was equal to or less than 10^3 spores/ml, while two of the samples had an average of 10^4 spores/ml and another two contained an average of 10^5 spores/ml. All the spore trap heights (positions) gave similar results, producing spore traps with concentrations from below 10^3 to 10^5 spores/ml, except for positions 3 and 4 that had a narrower spore concentration range (Table 2). The minimum temperatures had low variation over the 12-month period and the maximum temperatures showed greater variation (Table 2). The minimum temperatures ranged from 5 °C to 23 °C while the maximum temperatures ranged from 20 °C to 43 °C.

DISCUSSION

The real-time PCR used in this study was based on a non-specific dye, as it is more cost effective than those based on hydrolysis probes. Although previous authors (Schweigkofler *et al.*, 2004; Garbelotto *et al.*, 2008; Ioos *et al.*, 2009) were divided in their choice of detection system, both hydrolysis probes (Ioos *et al.*, 2009) and SYBR Green approaches (Schweigkofler *et al.*, 2004; Garbelotto *et al.*, 2008) are available for *F. circinatum*. Therefore, even though a hydrolysis probe system is generally believed to be more sensitive than a non-specific dye system, the latter can be used to reliably quantify the spores of the pitch canker fungus.

This study compared four DNA isolation protocols that differed not only in time required to complete the extractions but also in sensitivity. The Schweigkofler *et al.* (2004) protocol has been used in the past in a study that focussed on the detection and quantification of *F. circinatum* spores and the concentrations detected ranged from 10^3 - 10^6 spores/ml. In this study only very high spore concentrations (10^7 spores/ml) were detected with this method. This lack in sensitivity may be due to the extraction protocol being inferior (less effective) to other extraction protocols. The method also requires a variety of equipment and chemicals/reagents increasing the cost of this isolation method compared to other methods that require less equipment and chemicals/reagents. The Aljanabi and Martinez (1997) protocol and the newly developed, modified O'Donnell and Cigelnik (1997) method were the

most sensitive and allowed detection of spore concentrations as low as 10^2 /ml. However, Walsh *et al.* (1991) was the chosen method for extracting DNA from the spores found on the spore traps. Even though it is less sensitive than the Aljanabi and Martinez (1997) and the newly developed method, the Walsh *et al.* (1991) method was less time consuming and required fewer chemicals/reagents and less specialised equipment. The DNA isolation method (Walsh *et al.*, 1991) used during this study was cost-effective, sensitive and quick, a combination of characteristics not found in the other three isolation methods. It ultimately allowed detection of spore concentrations between 10^3 - 10^7 spores/ml.

The results of the current study agree with those reported by Schweigkofler *et al.* (2004). From the literature it also appears that the Ios *et al.* (2009) real-time PCR assay is more sensitive than the Schweigkofler *et al.* (2004) assay. This is because it allows detection of only 8 fg DNA compared to the lower limit of 10 pg DNA or 10^3 spores/ml that were detected with the CIRC1A and CIRC4A primer pair. However, the Ios *et al.* (2009) method differed not only in the fact that it included *F. circinatum* culturing/enrichment and a different DNA isolation method, but the authors also had different aims. The Schweigkofler *et al.* (2004) study focussed on detecting *F. circinatum* and developing an assay to trap and quantify the spore concentrations of the pathogen found on pine trees by using real-time PCR. The Ios *et al.* (2009) study focussed on comparing enrichment procedures and DNA extraction kits to enable the detection of *F. circinatum* on pine seeds using real-time PCR.

The annealing temperature during PCR is important as it determines the sensitivity of the real-time PCR assay. The Schweigkofler *et al.* (2004) study included *Fusarium* species belonging to the *Gibberella fujikuroi* species complex to determine the specificity of the CIRC1A and CIRC4A primer pair and it was found that the primers were specific for only *F. circinatum* at an annealing temperature of 66 °C and higher.

The temperature range in the greenhouse can be expected to be narrower than the temperature range under normal environmental conditions as the greenhouse is a closed controlled system. At the highest minimum temperature of 23 °C (week 6) the average spore concentration was 10^4 spores/ml and at the lowest minimum temperature of 5 °C (week 30) the average spore concentration was lower (10^3 spores/ml), similar to the study of Garbelotto *et al.* (2008) that found that sporulation is limited at low temperatures. Between week 6 and week 20, there appeared to be a variation in average spore concentration, while between week 24 and week 50 the average spore concentration was constant. A possible explanation for this may be that

the minimum and maximum temperatures between week 6 and week 20 were more stable (little variation) compared to the minimum and maximum temperatures between week 24 and week 50. The change in seedling numbers in the greenhouse over the 12-month period was not recorded, but could have had an effect on the variation in spore concentration over the study period.

The Garbelotto *et al.* (2008) study found that sporulation is higher under cool (not cold) wet conditions and warm conditions in the absence of rainfall. The pine seedling nursery that was the focus of this study is regularly irrigated, which may be the reason for an average spore concentration that did not exceed 10^4 spores/ml. Also the average temperatures ranged from mild to warm, temperatures that may be too high for enhancing sporulation.

The position of the spore traps appears to have had an effect on the spore concentrations detected. Spore concentrations were not the same between position 1 and 4 (2 m above ground) and between position 3 and 6 (0.2 m above ground). Spore concentration did, however, agree between positions 2 and 5 by having the same spore concentration range being detected ($\leq 10^3$ to 10^5). Position 2 and 5 were at the seedlings tray level and positions 1, 3, 4 and 6 were at levels where there is the possibility of more air movement, as there are no seedling trays to obstruct airflow. This may indicate that spores produced on the pine seedlings by *F. circinatum* are able to disperse by air but do not rise or fall much in level above ground during air transport.

The DNA isolation method used during this study proved to be sensitive, cost-effective and required limited chemical and equipment usage. Furthermore, the spore concentrations of *F. circinatum* could be successfully detected and quantified using real-time PCR. A non-specific dye was used due to its cost-effectiveness compared to a hydrolysis probe system and the success obtained during previous studies that used a similar dye. Thus the methods applied during this study can be refined to be used as a diagnostic technique in a commercial pine seedling nursery. In a commercial seedling nursery other data (humidity, wind speed, wind direction, spore trap height and human movement) should be included to ultimately determine the seasonal trends of *F. circinatum* for successful management strategies.

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TABLES

TABLE 1: Comparison between the DNA isolation protocols

| Protocol | Spore concentration (spores/ml) [†] | | | | | | Time require to process samples (hours) |
|------------------------------------|--|-----------------|-----------------|-----------------|-----------------|-----------------|---|
| | 10 ² | 10 ³ | 10 ⁴ | 10 ⁵ | 10 ⁶ | 10 ⁷ | |
| Schweigkofler <i>et al.</i> (2004) | (-) | (-) | (-) | (-) | (-) | (+) | 5 [‡] |
| Walsh <i>et al.</i> (1991) | (-) | (+) | (+) | (+) | (+) | (+) | < 1 |
| Aljanabi and Martinez (1997) | (+) | (+) | (+) | (+) | (+) | (+) | 3.5 |
| Newly developed protocol | (+) | (+) | (+) | (+) | (+) | (+) | > 24 |

(-) The *F. circinatum* target fragment was not detected

(+) The *F. circinatum* target fragment was detected

[†] No detection of the *F. circinatum* target fragment at spore concentrations of 10⁰ and 10¹

[‡] Only a limited number of samples can be processed simultaneously as a centrifuge can only spin 6 falcon tubes at a time

TABLE 2: Summary of spore concentrations detected in the FABI Greenhouse during the study

| Week | Start date | Spore trap structures and position† | | | | | | Temperature (°C) | | |
|------|------------|-------------------------------------|--|-------------------------|-----------------------|--|-------------------------|------------------|------|-----------------|
| | | 1 2 m above ground | Structure 1 2 0.8 m above ground | 3 0.2 m above ground | 4 2 m above ground | Structure 2 5 0.8 m above ground | 6 0.2 m above ground | Average | Min. | Max. |
| 2 | 10 Nov | * | * | * | * | * | * | | 18 | 26 |
| 4 | 24 Nov | * | * | * | * | * | * | | 18 | 31 |
| 6 | 8 Dec | * | 10 ⁴ | * | * | * | * | 10 ⁴ | 23 | 30 |
| 8 | 22 Dec | * | * | * | * | * | * | | 18 | 32 |
| 10 | 5 Jan | ≤10 ³ | 10 ⁴ | * | * | * | * | 10 ³ | 22 | 31 |
| 12 | 19 Jan | 10 ⁴ | 10 ⁴ | * | 10 ⁴ | * | 10 ³ | 10 ⁴ | 21 | 42 |
| 14 | 2 Feb | 10 ⁵ | 10 ⁵ | * | * | * | 10 ⁵ | 10 ⁵ | 17 | 29 |
| 16 | 16 Feb | * | ≤10 ³ | ≤10 ³ | * | * | ≤10 ³ | ≤10 ³ | 17 | 30 |
| 18 | 2 Mar | * | * | * | 10 ² | 10 ³ | 10 ² | 10 ² | 17 | 33 |
| 20 | 16 Mar | 10 ⁵ | * | * | * | * | * | 10 ⁵ | 17 | 28 |
| 22 | 30 Mar | * | * | * | * | * | * | | 18 | 41 |
| 24 | 13 Apr | ≤10 ³ | * | ≤10 ³ | 10 ³ | 10 ³ | 10 ³ | 10 ³ | # | # |
| 26 | 27 Apr | 10 ³ | ≤10 ³ | ≤10 ³ | 10 ³ | 10 ³ | * | 10 ³ | # | # |
| 28 | 11 May | ≤10 ³ | 10 ⁴ | 10 ³ | 10 ³ | 10 ⁴ | * | 10 ³ | 15 | 31 |
| 30 | 25 May | ≤10 ³ | * | * | 10 ³ | * | * | 10 ³ | 5 | 43 [‡] |
| 32 | 8 Jun | ≤10 ³ | 10 ³ | ≤10 ³ | 10 ³ | ≤10 ³ | * | ≤10 ³ | 15 | 30 |

| Week | Start date | Spore trap structures and position† | | | | | | Temperature (°C) | | |
|-------|------------|-------------------------------------|---|----------------------------|--------------------------|---|----------------------------|------------------|------|------|
| | | 1 2 m above ground | Structure 1 2 0.8 m above ground | 3 0.2 m above ground | 4 2 m above ground | Structure 2 5 0.8 m above ground | 6 0.2 m above ground | Average | Min. | Max. |
| 34 | 22 Jun | * | $\leq 10^3$ | 10^3 | * | 10^5 | * | 10^3 | 18 | 26 |
| 36 | 6 Jul | * | * | $\leq 10^3$ | * | * | * | $\leq 10^3$ | 18 | 20 |
| 38 | 20 Jul | * | * | * | * | * | * | | 8 | 21 |
| 40 | 3 Aug | * | * | * | * | * | * | | 10 | 25 |
| 42 | 17 Aug | * | * | $\leq 10^3$ | * | $\leq 10^3$ | * | $\leq 10^3$ | 10 | 29 |
| 44 | 31 Aug | * | * | * | * | * | * | | 11 | 31 |
| 46 | 14 Sept | * | * | * | * | * | * | | 16 | 34 |
| 48/50 | 12 Oct | * | $\leq 10^3$ | * | * | $\leq 10^3$ | $\leq 10^3$ | $\leq 10^3$ | 17 | 35 |
| 52 | 26 Oct | * | * | * | * | * | * | | 17 | 27 |

† All entries represent the average of four real-time PCR experiment results

* Indicates samples in which *F. circinatum* was not detected due to fungus absence or presence at a level below the real-time PCR detection limit

No data available

‡ At week 30 the highest maximum of 43 °C occurred, an uncharacteristic temperature during the winter season. This may be due to either a broken thermometer or the greenhouse fans that did not switch on when the inside temperature of the greenhouse reached 25 °C.

Weeks 48/50 were combined

SUMMARY

This project focussed on the pine pathogen *Fusarium circinatum* as this fungus is responsible for great economic losses and endangering pine forests worldwide. A better understanding of this pathogen could lead to improved disease management, thus limiting the dispersal of the fungus. A diagnostic technique, whether non-DNA-based or DNA-based, should be chosen with care to ensure obtaining accurate results, limiting expenses and time needed to complete a project. This study reviewed the techniques used for the identification and detection of *Fusarium* species. Non-DNA-based and DNA-based techniques focus on the morphological or chemical traits, and nucleotide sequence respectively. This review enabled the selection of an appropriate detection and quantification technique for *F. circinatum*, as the advantages and disadvantages of all techniques were considered and compared.

The intergenic spacer (IGS) region of the ribosomal RNA repeat module is a popular diagnostic marker for *Fusarium* species due to the region's multiple copies in the genome and fast evolution rate. This marker was evaluated to determine if it could be used for *F. circinatum* diagnostics. The results confirmed the IGS region to be a good diagnostic marker, due to the region's species specificity, by comparing the IGS region of *F. circinatum* and other *G. fujikuroi* complex species. The results of phylogenetic analysis of all the isolates included in this study were supported by those of previous studies that also focussed on the IGS region.

Real-time PCR was used successfully for the detection and quantification of *F. circinatum* during past studies. Due to these successes, real-time PCR was chosen as the technique for the quantification of *F. circinatum* spores in a pine seedling nursery. The results of this study showed that the real-time PCR technique employed was specific and sensitive as it allowed detection of low spore concentrations. The study also showed that there is a correlation between spore production and temperature and that spore dispersal is possibly, but not exclusively, due to air movement. Real-time PCR can thus be used for future studies that focus on the detection and quantification of *F. circinatum* as this study has shown this technique to be effective.

APPENDIX 1

| | 10 | 20 | 30 | 40 | 50 | 60 | 70 | 80 | 90 | 100 | 110 |
|--------------------------------------|--|--|----|----|----|----|----|----|----|-----|-----|
| <i>F. circinatum</i> NRRL25331 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | | ACTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | | ATTCTTCCACGGCGCTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | | ATTCTTCCACGGCACTCGAAGCGTGTCTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | | ATTCTTCCACGGCGCTCGAAGCGCGTCGTGGTATTTCGCGTATTGTAATTTCAACACGAGCGGGGTCAAATCCTTTGCAGACGACTTAGCTGTGCGAAACGGTCCTGTAA | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | | ----- | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | | ----- | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | | ----- | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | | ----- | | | | | | | | | |

| | 120 | 130 | 140 | 150 | 160 | 170 | 180 | 190 | 200 | 210 | 220 |
|--------------------------------------|--|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGAAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGAAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-AT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCCGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-TT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGA-CT | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-AC | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-CT | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-AT | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCTGGATTTCCGGAGACTTGTAGGGG-TTGTGGG-AT | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCTGGATTTCCGGAGACTTGTAGGGG-TTGTGGG-AT | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | GCAGTAGAGTAGCCTTGTTGTTACGATCTGCTGAGGGTAAGCCGTCCTTCGCCTCGATTTCCTCCGAATGGGTTCTCCGGATTTCTGGAGACTTGTAGGGG-TTGTGGG-AT | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | ----- | | | | | | | | | | |

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          230      240      250      260      270      280      290      300      310      320      330
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F. circinatum NRRL25331 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum CAL2511 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum FL2483 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum FL2497 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum MEX4900 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGARWTTYMT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum FL2496 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum SA0041 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum MEX2248 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum SA0010 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGKKGRAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum FL2472 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum SA1834 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum MEX2265 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. circinatum SP5498 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
F. subglutinans NRRL22016 TTTTTCGATTGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGCACTCTGTG-----
F. begoniae NRRL25300 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTG-----
Fusarium sp. NRRL25346 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGAACTTTGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL25204 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGGCTTTGTG-----
Fusarium sp. NRRL26756 TTTTTCGA-TGCGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGGCTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL26757 TTTTTCGA-TGCGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGGCTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL25623 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL25195 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
F. succisae NRRL13613 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGGACTTTGTGCGGCTGT
F. anthophilum NRRL13602 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTACAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL29123 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGCACTTTGTG-----
F. bulbicola NRRL13618 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL25807 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
Fusarium sp. NRRL25622 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
F. bacterioides NRRL20476 TTTTTCGA-TGTGTCGTCTCTGGACGGGCGGTTCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCAGGACTTTGTG-----
F. pseudoanthophilum NRRL2520 TTTTTCGA-GATGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TTCCTCGAGTCTGGCAGGACTTTGTG-----
F. verticillioides NRRL22172 TTTTTCGA-GATGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TTCCTCGAGTCTGGCAGGACTTTGTG-----
F. thapsinum NRRL22045 TTTTTCGA-GATGTCCTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TTCCTCGAGTCTGGCAGGACTTTGTG-----
F. concentricum NRRL25303 TTTT--GA-TGTGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCATGGCTTTGTG-----
F. fractiflexum NRRL28852 TTTT--GA-TGTGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCATGGCTTTGTG-----
F. fujikuroi NRRL13566 CTTT--GA-TGTGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCGGAGTTGTG-----
F. globosum NRRL26131 TTTT--GA-TGTGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCATGGCTTTGTG-----
F. proliferatum NRRL22944 TTTT--GA-TGTGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCATGGCTTTGTG-----
F. oxysporum NRRL22902 TTTT--GA-TGTGTCGTCTCCGGACGGGCGGTGCAGGGTAGTCGAGTTGGACTTGGTGGAAATTCAT-----TCCGTCGAGTCTGGCATGGCTTTGTG-----
F. subglutinans MRC8553 -----
F. subglutinans MRC8554 -----
F. subglutinans MRC6512 -----
F. subglutinans MRC2802 -----
F. proliferatum CMW 12864 -----
F. proliferatum MRC 2301 -----
F. proliferatum CMW 12867 -----
F. proliferatum MRC 8550 -----
F. proliferatum MRC 8549 -----
F. oxysporum CAV105 -----
F. oxysporum CAV617 -----
F. oxysporum CAV794 -----
F. oxysporum CAV602 -----
F. oxysporum CAV929 -----
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| | 340 | 350 | 360 | 370 | 380 | 390 | 400 | 410 | 420 | 430 | 440 |
|--------------------------------------|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTKCTTGGKCTTGGTCGATTTGAGGATCGATTTCGAGGGTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTCCATCCGTCGAGTCTGGCGGGAC--T-TTG-----TGCGGCTGTGTGYTGKAYKGT | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGAT----- | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTAAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GCAACTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTAAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GCAACTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GGAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTACTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | GTCGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAGCTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCAGGGCTGGGC--G-TTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCAGGGCTGGGC--G-TTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTCAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GCAACTTGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTCAGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGTCTTGGTCGATTTCAAAGATCGATTTCGAGGGCTGAGG--G-CCGGCCGTGAGTTGTGTGTGAGGGATGCCGT | | | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTACTCAGTCTTGGTCGATTTGAGGATCGATTTCGGGGCTGAGT--G-TCG-----ACTGTGTGTGAGGGATGGGGT | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTC----- | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GCG-----GTTAGTGTAGGTGGGTGCCGT | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-TCG-----AGTAAGTGTAGATGGGTGCCGT | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-TCG-----AGTAAGTGTAGATGGGTGCCGT | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGATGATCGATTTCGAGGACCGGCC--A-CAG-----AGGATGTGGATAGATGGGGT | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | --CGGCTGTGTGCTGGACGGTGCAGGGTAGGCTGCTTGGTCTTGGTCGATTTGAGGATCGATTTCGAGGGCTGGGC--G-CTG-----GTAAGTTCGAGGTGTATGCCGT | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | --CGGCTGTGTGCTGGACGGTGTAGGGTAGGCTGCTTGGACATGGTCGGTTCGAGGATCGATTTCGAGGGCTGGGC--TCG-----ATGATGTGTATGATGCCGT | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | ----- | | | | | | | | | | |

| | 450 | 460 | 470 | 480 | 490 | 500 | 510 | 520 | 530 | 540 | 550 |
|--------------------------------------|--|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| <i>F. circinatum</i> NRRL25331 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGRGTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | | GCAGGGTAGGCTGCTTGGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | | -----GGTGCAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCCGATCGAGGTTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCAAATCTGATGTCGGCTTCCGTGCGGACCAGATCGAGCTTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCTTGGTATAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGAGGTTGGCTTCCGTGCGGACCAGATCGAGCATGGTATAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAGTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCATGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAGTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCATGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCAAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCGTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATCTGATGTCGGCTTCCGTGCGGACCAGATCGAGCTTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | | CTAGGGTAGGCTGATTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCTTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGACC | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGCCAGATCGAGCTTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTTGA-- | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | | CTAGGGTAGGCTGGTTCGTC-TTGGTCAAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCTTGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | | CTAGGGTAAGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGGGCATGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGCCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | | CTAGGGTAAGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACCAGATCGAGCATGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGCCGATC | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | | CTAGGGTAGGCTAGTTTGTC-TTGGTCGAATTTGATATTGGCTTCCGTGCGGACTAGATCGGGCATGGTACAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTTGATGTTGGCTTCCGTGCGGACTAGATCGAGCGTGGTCCAGGTTAGGTACAGGGTAGGCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATCTGATGTCGGCTTCCGTGTAGAGCGCAGCGAGGTTGGTGCAGGTTAGGTACAGGGTAGGCGAGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATCTGATGTCGGCTTCCGTGTAGGACGGAACGATGGTGGTGCAGGTTAGCTATAGGGTAGGCGAGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | | -----AGACTTTGGTCGATC | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAGTTCGATGTCGGCTCTCGTGTAGAACAGAGCGAGCGTGGTTCAGGTTAGGTACAGGGTAGGCGAGCTTAGACTTTGGCCGATC | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAGTTCGATGTCGGCTTCCGTGTAGAACAGAGCGAAGCTGGTCCAGGTTAGGTACAGGGTAGGCGAGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTCGATGTCGGCTTCCGTGTGGAACAGAGCGAGAGTGGTCCAGGTTAGGTACAGGGTAGGCGAGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAACTCGATGTCGGCTTCCGTGCGAGACCAGAACGAGGTTGGTGCAGGTTAGGTACAGGGTAGGCGAGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | | CTAGGGTAGGCTGGTTTGTC-TTGGTCGAATTCGATGTCGGCTTCCGTGCGAGACCAGAGCGAGGTTGGTCCAGGTTAGGTACAGGGTAGGCGAGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | | -----AGCTCGGCTTAGACTTTGGTCGATC | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | | ----- | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | | ----- | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | | ----- | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | | ----- | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | | ----- | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | | ----- | | | | | | | | | |

| | 560 | 570 | 580 | 590 | 600 | 610 | 620 | 630 | 640 | 650 | 660 |
|--------------------------------------|---|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> SA20041 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | TGGATGTCGG--TTCTCTGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | TGGATGTCGG--TTCTCTGGCTGACGGATCCAGAGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | TGGATGTCGG--TTCTCCGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | TGGATGTCGG--TTCTCCGGCTGACGGATCTCAGAGCTTCGGAACGAGATGGGAGTAGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | TGGATGTCGG--TTCTCCGGCTGACGGATCTCAGAGCTTCGGAACGAGATGGGAGTAGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | TGGATGTTGG--TTCTCTGGCTGACGGATCTCAGAGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTCTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | TGGATGTTGG--TTCTCTGGCTGACGGATCTCAGAGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTCTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | TGGATGTCGG--TTCTCCGGCTGACGGATCTCAGAGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | TGGATGTCGG--TTCTCCGGCTGACGGATCTCAGAGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | TGGATGTCGG--TTCTCTGGCTGACGGTCTCAGAGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTAGCGTTGAG--ATG | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | -----TCTGGCTGACGGATCTCAGCGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTCGAG--GTA | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | TGGATGTCGG--TTCTCTGGCTGACGGATCCAGAGCTTCAGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGCGTTGAG--ATG | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | TGGGTGTCGG--TTCTCTGGCTGGCGGACCTCAGAGCTTCGGAACGAGATGGGAGTAGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGTTGAG--GTG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | TGGGTGTCGG--TTCTCTGCCTGGCGGACCTCAGAGCTTCGGAACGAGATGGGAGTAGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTCCGTTGAG--GTG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | TGGATGTCGG--TTCTCCGGCTGACGGATCTCAGGGCTTCGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTTGTATTAGG--ATG | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | TGGATGTTGT--TTCTCTGGCTGGCGGATCTCAGGGCTTTGGAACGAGATGGGAGTGGTGCAGGGTAGGCCA | ACTTCTGTCTTTGCCAGGATGCGATTTGTATTAGG--ATG | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | TGGATGTCGG--TTCTCTGGCTGACGGGTCTGAGGTCGTCGAAACGAGATGCGACTGGTGCAGGGTAAGCA | GCTTCCGTATCGGCCGGGAAGTGATTCGAATTGAGATATG | | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | TGGGTGTCGGTTTCCTGGCTGGCGGGTCTGAGGTCGTCGAGACGAGATACGAGTGGTGCAGGGTAAGCA | GCTTCTGTATCTGCCAGGAAGCGATTTCG----- | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | TGGATGTCGG--TTCTCTGGCTGACGGGTCTGAGGTCGCTGAAACGATACGGGGCTGGTGCAGGGTAGGCCA | ACTTCTGACT--GCCGGGAAGCGATTCCGAATCGAG--TTG | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | TGGATGTCGG--TTCTCTGGCTGATGGATCTGAGAGCGTCGAAACGAGATGCGGGCGCTGCAGGGTAGGTA | ACTTCTGTCT--GTCAGGAAGCGATTTGTATTAGG--ATG | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | TGGATGTCGG--TTCTCCGGCTGATGGATCTGAGACCGTCGAAACGAGATGCGGGCGCTGCAGGGTAGGTA | ACTTCTGTCTCTCGTCAAGGAGCGATTCCGTTGAG--ATG | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | TAGATGTCGG--TTCTCTGGCTGATGGATCTGAGACCGTCGAAACGAGATGCGGGCGCTGCAGGGTAGGTA | ACTTCTATCTTCGGCAGGAAGCGATTCCGTTGAG--ATG | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | TGGATGTTAG--TTCTCTGGCTGATGGATCTGAAAGCGTCAAACGAGATGCGGGCGCTGCAGGGTAGGTA | ACTTCTGTCTTCGTCAGGAAGCGATTTGTATTAGG--ATG | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | TGGATGTCGG--TTCTCTGGCTGATGGGTCTGGGACCGTCAAACGAGATGCGGGCGCTGCAGGGTAGGTA | ACTTCTGTCTTCGTCAGGAAGCGATTCCGTTGAG--ATG | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | TGGAGGTCGA--TTCTCCGGCTGGCGGATCTGACACTGTCGAAACGAGATGCGAGCGGTGTAGGGTAGGTA | GTTTCGTTCTCGCCAGGTTGCGATTTG--GACGAG--ATG | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | ----- | | | | | | | | | | |

| | 670 | 680 | 690 | 700 | 710 | 720 | 730 | 740 | 750 | 760 | 770 | |
|--------------------------------------|---|--|--|--|--|--|--|--|--|--|--|--|
| <i>F. circinatum</i> NRRL25331 | | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT | TGCCGT-CTAGGTTAGG-TACAGGGTAGGTAGAA-TCTCAG-TTTCGTCGCT-ACAG-TTTGCCA-----TAGGTGTAGGGTA-GGTACAGGGTAGGCAGATTTCT |
| <i>F. circinatum</i> CAL2511 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | | | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | | | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | | | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | | | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | | | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | | | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | | | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | | | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | | | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | | | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | | | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | | | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | | | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | | | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | | | | | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | | | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | | | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | | | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | | | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | | | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | | | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | | | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | | | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | | | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | | | | | | | | | | | | |

| | 780 | 790 | 800 | 810 | 820 | 830 | 840 | 850 | 860 | 870 | 880 |
|--------------------------------------|--|-------------------------|------------------|---------|---------|---------|---------|---------|---------|---------|---------|
| <i>F. circinatum</i> NRRL25331 | | | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> FL2483 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> FL2497 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> FL2496 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> SA0041 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> SA0010 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> FL2472 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> SA1834 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. circinatum</i> SP5498 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGA-TTTTTTGT | TTTGCCATA-AAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | ACTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGCGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAACGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATGAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGATTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATGAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTCTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAACGGTTTTGTGGCCGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATATAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | ACTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | CCTCCGGCCAGTACTTGCTTATGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | CCTCCGTCAGTACTTGTTTGTGCGTGCGAAATGGTTTTGTGGTCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | CTTCCGGCCAGTACTTGTTTGTGCGTGCGAAATGGTTTTCAAGGCTGGTGGGCGTGAGTCGATTTTTTTGT | TTTGCCATACAAATGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | TCCCCGGCCAGTACTTGCTATGCGTGCGAAACGGTTTTCGTGGTCCAGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATCGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | CCCCGGCCAGTACTGCGATGCGTGCGAAACGGTTTTGTGGTCCGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATCGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | CCCCAGGCCAGTACTTGCCCGTGGCGTGCGAAATAGTTTTGCGGTCTGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCGACTATTTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | -ATCCGGCCAGTACTGGCCATGCGTGCGAAATGGTTTTGCAGTCTGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | -ATCCGGCCAGTACTGGCCATGCGTGCGAAATGGTTTTGCAGTCTGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | -ATCCGGCCAGTACTGGCCATGCGTGCGAAATGGTTTTGCAGTCTGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | -ATCCGGCCAGTACTGGCCATGCGTGCGAAATGGTTTTGCAGTCTGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | -GTCCGGCCAGTACTGGCCATGCGTGCGAAATGGTTTTGCAGTCTGGTGGCCGTGAGTCGATTTTTTTGT | TTTGCCATACTATTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | -CTCCGGCCAGTACT-----GTCTGGTGGTCTGAGGCGATTTTTTTGT | TTTGCCATACTATTGAATTTTGC | GGGAAATCAAAAGTGG | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | ----- | | | | | | | | | | |

| | 890 | 900 | 910 | 920 | 930 | 940 | 950 | 960 | 970 | 980 | 990 |
|--------------------------------------|--|--|--|--|--|--|--|--|--|--|--|
| <i>F. circinatum</i> NRRL25331 | | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG | CCCACGAGTCGGGTCTGGCGTGCCTCCGACTGGAACATCGTCGGTGGACATATGAGAGGGAGCAATCCGGCCGGGCCGGGGCCCATCGCGACCTGCAGGGTAGGTAAAAG |
| <i>F. circinatum</i> CAL2511 | | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | | | | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | | | | | | | | | | | |

| | 1000 | 1010 | 1020 | 1030 | 1040 | 1050 | 1060 | 1070 | 1080 | 1090 | 1100 | |
|--------------------------------------|---|---|--|---|-----------------------|-----------------------|-------------------|-----------------------|-----------------------|----------------------|------|--|
| <i>F. circinatum</i> NRRL25331 | | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | |
| <i>F. circinatum</i> CAL2511 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> FL2483 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> FL2497 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> MEX4900 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> FL2496 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> SA0041 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> MEX2248 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> SA0010 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> FL2472 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> SA1834 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> MEX2265 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. circinatum</i> SP5498 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTTCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | TAAAAACAGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGA | GGT | GGAAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGA | GGT | TAGAAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | GGT | GAGGGCCATGAAGACTTG | --G-----A | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | GGT | GAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | GGT | GAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | GGT | GAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGA | GGT | GGAAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGA | GGT | GGAAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. succisae</i> NRRL13613 | TAAAAA | AGTTGTTAAAAGGCCCGGTGTCGGTGTGCTTGTATCCGAGGAAGAGAATTTGCTGGAGA | GGT | GGAAGGGCCATGAAGACTTG | --GACGGA----- | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | TTAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | GGT | GAGGGCCACGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGA | GGT | GGAAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTTCAGA | GGT | GGAAGGGCCATGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | TAAAAA | AGTTGATAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTTGAGA | GGT | GGAAGGGCCATGAAGACTCG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTCGAGGAAGAGAATTTGCTGGAGA | GGT | GAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTATCTGAGAG | GGT | GAGAGGGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | TAGATA | AGTTGTTAAAGAGGCCCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | TGG | AAGACGGCCAGGAAATCCGATGTTACCCGGGCACGGGAA | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAAGAGAATTTGCTGGAGG | TG | GAGGGCCAGGAAATCCGACGCTACCCGGGCATGGGAA | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGAGGAGAGAGAATTTGCTGGAGG | TGA | AGGGCGCCAGGAAATCCGATGTTACCCGGGCATGGGAA | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGGGGAAGAGAATTTCTGGAGG | TGG | AGGGAGGGAGCCGGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | CAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGGGGAAGAGAATTTGCTGGAGG | TGG | AGGGAGCCGGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGGGAAAGAGAATTTATGTGGAGG | TGG | AGGGAGCCGGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. globosum</i> NRRL26131 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGGGGAAGAGAATTTGCTGGAGG | TGG | AGGGAGCCAGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGGGGAAGAGAATTTATCTGGAGG | TGG | AGGGAGGGAGCCGGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | TAAAAA | AGTTGTTAAAGAGGCGCGGTGTCGGTGTGCTTGTATTTGCGGGAGAGAATTTATCTGGGG | TG | CGGGTAGCCGGGAAGACTTG | --GACGGATCTGGCCCGGGAA | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | ----- | ----- | ----- | ----- | GGGCATGAAA | ACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | ----- | ----- | ----- | ----- | GGGCATGAAA | ACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | ----- | ----- | ----- | ----- | GGCCCAA | AAACTC | --GACGGACTGGTCCGGG | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | ----- | ----- | ----- | ----- | GGAGGTGGAGG | GAAGCCCGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | ----- | ----- | ----- | ----- | CTGGAGGTGGAGG | GAAGCCCGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | ----- | ----- | ----- | ----- | GGAGGTGGAGG | GAAGCCCGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | ----- | ----- | ----- | ----- | GGAGGTGGAGG | GAAGCCCGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | ----- | ----- | ----- | ----- | GGAGGTGGAGG | GAAGCCCGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | ----- | ----- | ----- | ----- | GGAGGTGGAGG | GAAGCCCGGAAGACTTG | --GACGGATCTGGTCCGGGAA | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | ----- | ----- | ----- | ----- | CGGGAGGTGCTGGG | TACCCGAAA | AAACTTG | --AACGGATCTGGCCCGGGCC | | | |
| <i>F. oxysporum</i> CAV617 | ----- | ----- | ----- | ----- | ----- | TTATCTGGAGG | TGCTGGGCACCCGAAA | AAACTTG | --GACGGATCTGGCCCGGGAA | | | |
| <i>F. oxysporum</i> CAV794 | ----- | ----- | ----- | ----- | ----- | GGTGTGGG | TACCCGAAA | AAACTTG | --AACGGATCTGGCCCGGGAA | | | |
| <i>F. oxysporum</i> CAV602 | ----- | ----- | ----- | ----- | ----- | TCTGGGG | TGCTGGG | TACCCGAAA | AAACTTG | --AACGGATCTGGCCCGGAA | | |
| <i>F. oxysporum</i> CAV929 | ----- | ----- | ----- | ----- | ----- | TGCTGGGA | AACCCGAAA | AAACTTG | --AACGGATCTGGCCCGGACC | | | |

| | 1110 | 1120 | 1130 | 1140 | 1150 | 1160 | 1170 | 1180 | 1190 | 1200 | 1210 | |
|--------------------------------------|---|---|---|---|---|---|---|---|---|---|---|---|
| <i>F. circinatum</i> NRRL25331 | | CGGGTCTGGGCTCGGATTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGATTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG | CGGGTCTGGGCTCGGGTTTTGGGGTGGTGCAGGGTAGGCAGGTTTAGATAGACGGTC-CAGGGTAGGT-----ACAGGGTAGGCACAGGGTAG |
| <i>F. circinatum</i> CAL2511 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | | | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | | | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | | | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | | | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | | | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | | | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | | | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | | | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | | | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | | | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | | | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | | | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | | | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | | | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | | | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | | | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | | | | | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | | | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | | | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | | | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | | | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | | | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | | | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | | | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | | | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | | | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | | | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | | | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | | | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | | | | | | | | | | | | |

| | 1220 | 1230 | 1240 | 1250 | 1260 | 1270 | 1280 | 1290 | 1300 | 1310 | 1320 |
|--------------------------------------|---|--|------|------|------|------|------|------|------|------|------|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> SA20041 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | GTAGAAGTCT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | GTAAAATTTT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | GTAGAATTTT | -GGTAAATAGTATGGGCGGGTGTAGGGTAGGCTTGGATACCGTTTTGTTTCACGCCCTTACCTTGGCTCGAGAAGGGACAATCTTGG-CCGGGATCGAAG | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | CAAAAACCC | -GGTAAAC-CCGTGGGGGGTGTAGGGAAGGCTCGAAAGGCTTCCCTTTGGAGCCTTACCTTTGAAAAGAAAGGGGTTTTTTCG-CCGGTTCCCGG | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | GTAAAACCC | -GGTATATAGTAGGGGGGGTGTAGGGTAGGCTGAAATACCTTTTTGTTTGGCCCTTACTTTTGTGAAAACGACAATTTTGG-CCGGAATGAAGG | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | GTAAAACCC | -GGTATATAGTAGGGGGGGTGTAGGGTAGGCTGAAATACCGTTTTGTTTGGCCCTTACTTTTGTGAAAACGACAATTTTGG-CCGGAATGAAGG | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | GTAAAACCC | -GGTATATAGTAGGGGGGGTGTAGGGTAGGCTGAAATACCGTTTTGTTTGGCCCTTACTTTTGTGAAAACGACAATTTTGG-CCGGAATGAAGG | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | GTAAAACCC | -GGTATATAGTAGGGGGGGTGTAGGGAAGGCTGAAATACCGTTTTGTTTGGCCCTTACTTTTGTGAAAACGACAATTTTGG-CCGGAATGAAGG | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | GTAAAACCC | -GGTATATAGTAGGGGGGGTGTAGGGAAGGCTGAAATACCGTTTTGTTTGGCCCTTACTTTTGTGAAAACGACAATTTTGG-CCGGAATGAAGG | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | GTAGAACCC | -GGTATATAGTAGGGGGGGTGTAGGGAAGGCTGAAATACCGTTTTGTTTGGCCCTTACTTTTGTGAAAACGACAATTTTGG-CCGGAATGAAGG | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | --AAAATCT | -GGTTATAGTA-GGGGGGGTGTGGGTAGGCTGGACACCGTTTTTCCAAATCCCC--TTACCCGAGGGAGGACAATCTTGG-CTGGTTGGAGG | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | --AAAATCT | -GGTATATAGTA-GGGGGGGTGTGGGTAGGCTGGACACCGTTTTTCCAAATCCCC--TTACCCGAGGGAGGACAATCTTGG-CTGGATGGAGG | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | --AAAATCT | -GGTACATATA--GGGGGGTGTAGGGAAGGCTGGACACCGTTTTTCCAAATCCCC--TTACCCGAGGGAGGACAATCTTGG-CTGGATGGAGG | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | --AGACTTC | -GGTATATATA--GGGGGGTGTGGGTAGGCTGGACACCGTTTTTCCAAATCCCC--TTACCCGAGGGAGGACAATCTTGG-CTGGATGGAGG | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | --AAACTTC | -GGTTTTTATTT--GGGGGGTGTAGGGAAGGCTGGACCCCTTTCCAAATCCCC--TTACCCGAGGGAGGACAATCTTGG-CCGGAACGGAGG | | | | | | | | | |

| | 1330 | 1340 | 1350 | 1360 | 1370 | 1380 | 1390 | 1400 | 1410 | 1420 | 1430 |
|--------------------------------------|---|--|--|------|------|------|------|------|------|------|------|
| <i>F. circinatum</i> NRRL25331 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | TCTAGGGTAGGCTGAATCTATGATTATGTAAGCTGTACTGTAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | TCTAGGGTAGGCTGAATCTATGATTATGTAAGCTGTACTGTACTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | TCTAGGGTAGGCTGAATCTATGATTATGTAAGCTGTACTGTACTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | TCTAGGGTAGGCTGAATCTATGATCATGTAAGCTGTACTCCAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | TCTAGGGTAGGCTGAATCTATGATCATGTAAGCTGTACTCCAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | TCTAGGGTAGGCTGAATCTATGATTATGTAAGCTGTACTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | TCTAGGGTAGGCTGATTTTATGATTATGTAAGCTGTACTCTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGTGGAGAGAT | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | TCTAGGGTAGGCTGAATTTATGATTATGCAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCCCATAACAAATCGGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | TCTAGGGTAGGCTGAATCCATGATTATGTAAGCTGTATAA | CTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | -----TCTAGGGTAGGCTGAATTTATGATTATGTAAGCTGTATAA | CTAGGGTAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | TCTAGGGTAGGCTGATTTTATGATTGTGTACGCTACATTACTCTAGGGTAGGTA | AAAAATCTCACATAAACTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | TCTAGGGTAGGCTGATTTTATGATTATGTAAGCTACATTACTCTAGGGTAGGTA | AAAAATCTCATATAAACTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | TCTAGGGTAGGCTAAATGTGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCTCATACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | TCTAGGGTAGGCTGAATCTGTAGTTTGTAGCTGTATAGCTCTAGGGTAGGTA | AAAAATCTTATATAAACTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | TCTAGGGTAGGCTGGATTCATGATTACATAAGCTGTACTGCTCTAGGGTAGGTA | GAAAGACCCATAACAAATCTGATCAGAAACGGTGAAGAAGTGTATTTGCTCGGGAGAT | | | | | | | | | |
| <i>F. verticillioide</i> NRRL22172 | TCTAGGGTAGGCTGAATCTATGATTATATAAGCTATACTGCTCTAGGGTAGGTA | GAAAGATCCCATAGAAATCTGATCAGAAATGGTGACGAAGTGTATTTGCTCGGAAGAT | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | TCTAGGGTAGGCTGAATTTATGATTATATAAGCTGATTACTCTAGGGTAGGTA | GAAAAATCCCATATAAGCTGACCAACTGGTGAAGATGTGATTTGCTCGGAAGAT | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | TCTAGGGTAGGCTGAATTTATGATTATATAAGCTGGATAGCTCTAGGGTAGGTA | GAAAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAGAT | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | TCTAGGGTAGGCTGAATCTATGATTAGATAAGCTAGATTACTCTAGGGTAGGTA | GAAAAATCCGATATAAACTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAGAT | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | TCTAGGGTAGGCTGAATCTATAATTGTATAAGCTAGACTACTCTAGGGTAGGTA | GAAAAATCCCATCTAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAGTT | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | TCTAGGGTAGGCTGAATTTATGATTATATAAGCTGGATAGCTCTAGGGTAGGTA | GAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAGAT | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | TCTAGGGTAGGCTGAATTTATGATTATATAAGCTGGATAGCTCTAGGGTAGGTA | GAAAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAGAT | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | TGTAGGGTAAGCTTAAATTTACGATTACATGATCTGTGTCCTCTAGGGTAGGTA | GAAAAATCCCATATACTGATCACATTTGGTGAAGAGGTTGGTTGGCTGGTGAAGAT | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | TCTAGGGTAGGCTGAATGTATGATTATGTAAGCTGTATAGCTCTGGGTA | GAGGTA | AAAAATCCCATAACAAATCTGATTGGAATTGGTGAAGATGTGATTTGGCTGGAGAGAT | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | -----TCTAGGGTAGGCTGAATGTTTATGATTATGTAAGCTGTATAGCTCTAGGGTAGGTA | A | AAAAATCCCATAACAAATCTGATTGGAATTGGGAAAAATGTGATTTGGCTGGARAGAT | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | CCCCGGTTAGGGTAGCCTGGATTTATCCCTCTGTGTACCCTGGGGAAGGGGAAAA | ATCCCGGGCCAAACCCAAAAGGAATTGGGGGGATGTTTTTGTGTTGCCGAAAT | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | TCTAGGGTAGGTTGATTTTATGATTATATAAGCTGGATAGCTCTAGGGAA | GAGTTAAAAATCCCATAACAAATCTGGTTGCAATTGGGGAATTTGTGATTTGCTGTGAAAAAT | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | TCTAGGGTAGGCTGATTTTATGATTATATAAGCTGGATAGCTCTAGGGTAGGTA | GAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAAAAT | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | TCTAGGGTAGGTTGATTTTATGATTATATAAGCTGGATAGCTCTAGGGAA | GAGTTAAAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAAAAT | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | TCTAGGGTAGGTTGATTTTATGATTATATAAGCTGGATAGCTCTAGGGTAGGTA | GAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAAAAT | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | TCTAGGGTAGGTTGATTTTATGATTATATAAGCTGGATAGCTCTAGGGAA | GAGTTAAAAATCCCATAACAAATCTGGTTGCAATTGGTGAAGTTGTGATTTGCTGTGAAAAAT | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | TGTTGGGTAGGCTTAACTTACAATTACTTTTCTGTGTCCTCTAGGGAA | GAGGGGAAAAATCCCATATATATCTGATCACATTTGGTGAAGAGGTTGGTTGGCTGGTGAAGAT | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | TGTAGGGTAGGCTTAACTTACAATTACTTTGATCTGTGTCCTCTAGGGTA | GAGGTA | AAAAATCCCATAATTTCTAACTACTTTGGTGAAGAGGTTGGTTGGCTGGTGAAGAT | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | TGTAGGGTAGGTTTATTTTACAATTACTTTGATCTGTGTCCTCTAGGGTA | GAGGTA | AAAAATCCCATAATTTCTAACTACTTTGGGAAAAAGGTTGGTTGGCTGGGAAAT | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | TGTAGGGTAGGTTTATTTTACAATTACTTTGATCTGTGTCCTCTAGGGAA | GAGGTA | AAAAATCCCATAATTTCTAACTACTTTGGGAAAAAGGTTGGTTGGGGAAT | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | TGTAGGGAAGGCTTACTTTACAATTACTTTGATCTGGGCCACTCTGGGTA | GAGGTA | AAAAATCCCATAATTTTGTGCCCTTTGGGAAAAAGGTTGTTGCTGGGAAAT | | | | | | | | |

| | 1440 | 1450 | 1460 | 1470 | 1480 | 1490 | 1500 | 1510 | 1520 | 1530 | 1540 |
|--------------------------------------|---|------------|------------|------------|-----------|-----------|-------------|-------------|-------------|-------------|------------|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAGATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> CAL2511 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAGATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> FL2483 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAGATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> FL2497 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAGATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> MEX4900 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> FL2496 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> SA20041 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> MEX2248 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> SA0010 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> FL2472 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> SA1834 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> MEX2265 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. circinatum</i> SP5498 | GGACGAAAGT | GCAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. subglutinans</i> NRRL22016 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. begoniae</i> NRRL25300 | GGAGAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>Fusarium</i> sp. NRRL25346 | GGAGAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>Fusarium</i> sp. NRRL25204 | GAAGAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>Fusarium</i> sp. NRRL26756 | GGAGAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>Fusarium</i> sp. NRRL26757 | GGAGAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>Fusarium</i> sp. NRRL25623 | GGAGAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>Fusarium</i> sp. NRRL25195 | GGAGGAAA | AGTGTAGTGC | AGAACTGTAT | GTGATTTTG | CAAAAGT | GAGTGTAAA | ATG-GAAAGTC | GATGTTCC | CGAGCAG | TTGGAG | CACGTTT |
| <i>F. succisae</i> NRRL13613 | GGATAAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCCCG | AGCAGGTG | GAGAGCAC | GTTTGGAG |
| <i>F. anthophilum</i> NRRL13602 | GGAGGAAA | AGTGCAA | TGCAGAA | CTGTATGT | GATTTTG | CAAAAGT | GAGTGTAAA | ATG-GAAAGTC | CAATGTT | CCCGAG | CAGTTG |
| <i>Fusarium</i> sp. NRRL29123 | GGACGAAA | AGTGTAA | TGCAGAA | CTGTATGT | GATTTTG | CAAAAGT | GAGTGTAAA | ATG-GAAAGTC | CGGATG | AGAGCA | CGTTT |
| <i>F. bulbicola</i> NRRL13618 | GGATAAAAGT | GTGATGCAGA | ACTGTGTGTG | ATTTTGCAAA | AGTGAGTGC | GAAAATG- | GAAAATCA | ATGTTCC | CGAGCAG | GCGAG | CACGTTT |
| <i>Fusarium</i> sp. NRRL25807 | GGATAAAAGT | GCAGCGCA | GAACCTGT | ATGTGAT | TTTTCAA | AAAGT | GAGTGC | GAAAATG- | GAAAGTT | AATGTT | CCCTGAG |
| <i>Fusarium</i> sp. NRRL25622 | GGAGAAAAGT | GTGATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCC | CGAGCAG | TTGAG | ACGTTT |
| <i>F. bacterioides</i> NRRL20476 | GGAGGAAA | AGTGTAA | TGCAGAA | CTGTATGT | GATTTTG | CAAAAGT | GGGTGTAAA | ATG-GAAAGTC | CGATGTT | CCCGAG | CAGTTG |
| <i>F. pseudoanthophilum</i> NRRL2520 | GCAGAAAAGT | GTGATGCAGA | ACTGTATGTG | ACTTTTG | CAAAAGT | GAGTGC | GAAAATG- | GAAAGTC | GGACTT | CCCGAG | CAGATG |
| <i>F. verticilliioides</i> NRRL22172 | GCAGAAAAGT | GTAACGCAGA | ACTGTATGTG | GCTTTTG | CAAAAGT | GAGTGC | GAAAATG- | AAAAGTC | GGACTT | CCCGAG | CGGTTG |
| <i>F. thapsinum</i> NRRL22045 | GGAAATAAGT | GTGATGTG | GAAAATG | ATGTGACT | TTTGT | AAAAGT | GAGTGTAAA | ATG-GAAAGTC | CAGACTT | CCCGAG | CAGATG |
| <i>F. concentricum</i> NRRL25303 | GAAGAAAAGT | GTAATGC | GGAATCG | TATGTG | ACTTTT | GCAAAAGT | GGGTGTAAA | ATG-GAAAGTC | CGGTTT | CCCTG | AGACAG |
| <i>F. fractiflexum</i> NRRL28852 | GGAGAAAAGT | GTGATGC | GGAATT | GTATGTG | ACTTTT | GTA | AAAGTGG | GTGAAA | ATG-GAAAGTC | CGGTTT | CCCGAG |
| <i>F. fujikuroi</i> NRRL13566 | GGAGAAAAGT | GTAATGTG | GGAATT | GTATGTG | ACTTTT | GCA | AAAGTGG | GTGAAA | ATG-GAAAGTC | CGGTTT | CCCGAG |
| <i>F. globosum</i> NRRL26131 | GGAGAAAAGT | GTGATGC | GGAATT | GTATGTG | ACTTTT | GCA | AAAGTGG | GTGAAA | ATG-GAAAGTC | CGGTTT | CCCGAG |
| <i>F. proliferatum</i> NRRL22944 | GGAGAAAAGT | GTGATGC | GGAATT | GTATGTG | ACTTTT | GTA | AAAAGT | GGATGTG | GAAA | ATG-GAAAGTC | CGGTTT |
| <i>F. oxysporum</i> NRRL22902 | GGACAAAAGT | GCAATGT | AGAATTA | TATGTG | ATTTT | GCAAA | AGTGG | GTGAAA | ATG-GAAAGTC | CGGTTT | CCCGAC |
| <i>F. subglutinans</i> MRC8553 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCC | CGAGCAG | TTAAG | AGCACG |
| <i>F. subglutinans</i> MRC8554 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCC | CGAGCAG | TTAAG | AGCACG |
| <i>F. subglutinans</i> MRC6512 | GGACGAAAGT | GTAATGCAGA | ACTGTATGTG | ATTTTGCAAA | AGTGAGTGT | AAAAATG- | GAAAGTCG | ATGTTCC | CGAGCAG | TTAAG | AGCACG |
| <i>F. subglutinans</i> MRC2802 | GGAGAAAAGT | GTAATGCAGA | GCTGTATGTG | ATTTTGCAAA | AGTGAGTGT | GAAAATG- | GAAAGTCG | ATGTTCC | CGACCAG | ATGAG | AGCACG |
| <i>F. proliferatum</i> CMW 12864 | GGAAAAAAGT | GTGATGC | AAAATT | GTAGGTG | ACTTTG | TAAAAG | GGAAATG | GAAA | ATG-GAAAGTC | CGGTTT | CCCGAG |
| <i>F. proliferatum</i> MRC 2301 | GGAAAAAAGT | GTGATGC | AAAATT | GTAGGTG | ACTTTG | TAAAAG | GGAAATG | GAAA | ATG-GAAAGTC | CGGTTT | CCCGAG |
| <i>F. proliferatum</i> MRC 12867 | GGAAAAAAGT | GTGGAC | CCAAAAT | TGTTGTG | ATTTT | GTA | AAAAGG | GAATG | GAAA | ATG-GAAAGTC | CGGTTT |
| <i>F. proliferatum</i> MRC 8550 | GGAAAAAAGT | GTGATGC | AAAATT | GTAGGTG | ACTTTG | TAAAAG | GGAAATG | GAAA | ATG-GAAAGTC | CGGTTT | CCCGAG |
| <i>F. proliferatum</i> MRC 8549 | GGAAAAAAGT | GTGAC | CCAAAAT | TGTTGTG | ATTTT | GTA | AAAAGG | GAATG | GAAA | ATG-GAAAGTC | CGGTTT |
| <i>F. oxysporum</i> CAV105 | GGACAAAAGT | GCAATGT | AGAATTA | TATGTG | ATTTT | GCAAA | AGTGG | GTGAAA | ATG-GGAAAT | C | CGGTTT |
| <i>F. oxysporum</i> CAV617 | GGATAAAAGT | GCACCGT | AAAATTA | TTTGTG | ATTTT | GCAAA | ATAG | GTGAAA | ATG-GGAAAT | C | CGGTTT |
| <i>F. oxysporum</i> CAV794 | GGACAAAAGT | GCAATGT | AGAATTA | TATGTG | ATTTT | GCAAA | AGTGG | GTGAAA | ATG-GGAAAT | C | CGGTTT |
| <i>F. oxysporum</i> CAV602 | GAACAAAAGT | GCAATGT | AAAATTA | TTTGTG | ATTTT | GCAAA | AGGG | GTGAAA | ATTTG | GAA | ATG-GGAAAT |
| <i>F. oxysporum</i> CAV929 | GAACAAAAGT | GCAATATA | AAATTTTT | TTTGTG | TTTTT | GCAAA | AGGG | GGTAAA | ATTTG | GAA | ATG-GGAAAT |

| | 1550 | 1560 | 1570 | 1580 | 1590 | 1600 | 1610 | 1620 | 1630 | 1640 | 1650 | |
|--------------------------------------|---|---------|-----------|----------|----------|----------|---------|----------|---------|---------|-----------|---------|
| | | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> CAL2511 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> FL2483 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> FL2497 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> MEX4900 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> FL2496 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> SA0041 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> MEX2248 | RCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> SA0010 | RCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> FL2472 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> SA1834 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> MEX2265 | RCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. circinatum</i> SP5498 | GCCTTGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. subglutinans</i> NRRL22016 | GCCTCGT | CTTGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. begoniae</i> NRRL25300 | GCCTTGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL25346 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL25204 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL26756 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL26757 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL25623 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL25195 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. succisae</i> NRRL13613 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. anthophilum</i> NRRL13602 | GCCTCGT | CGAGCCG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL29123 | GCCTCGT | CGAGCCG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. bulbicola</i> NRRL13618 | GCCTCGT | CGAGATG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL25807 | GCCTCGT | CGAGATG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>Fusarium</i> sp. NRRL25622 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. bacterioides</i> NRRL20476 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. pseudoanthophilum</i> NRRL2520 | GCCTCGT | CGAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. verticillioides</i> NRRL22172 | GCCTCAT | CGAGACG | AGCACAAC | CGGTATAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. thapsinum</i> NRRL22045 | GCCTCGCC | GAGACG | ATCACAAC | CGGTATAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. concentricum</i> NRRL25303 | GCCTTTT | CGAGCCG | TACACAG | CGTATG | TATGTG | TGCGG | ACTCCT | -GTTTGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. fractiflexum</i> NRRL28852 | GTCTTTT | CGAGGCG | GCACAG | CGGTGC | AGCTTAT | GTGTG | CGTGCC | AGACTCCT | -GTGTG | TCCCTGT | ACGGCGGGG | AAGTAGA |
| <i>F. fujikuroi</i> NRRL13566 | GCCTTTT | CGAGACG | GTACACAAC | CGGTGC | AGCTTAT | GTGTG | TGCC | AGACTCCT | -GTGTG | TCCCTGT | ACGGCGGGG | AAGTAGA |
| <i>F. globosum</i> NRRL26131 | GCCTTTT | CGAGGCG | GTACAG | CGGTGC | AGCTTAT | GTGTG | TCCG | GACTCCT | -GTGTG | TCCCTGT | ACGGCGGGG | AAGTAGA |
| <i>F. proliferatum</i> NRRL22944 | GCCTTTT | CGAGGCG | GTACACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. oxysporum</i> NRRL22902 | ACCTCTC | CGAGACG | ACTCAAC | CGGTACC | ACCCATG | TGTTGG | TCCGG | CTCCT | -GTGCG | GCCGTCC | AGGGCGGGG | AAGTAGA |
| <i>F. subglutinans</i> MRC8553 | GCCTCGT | CAAGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. subglutinans</i> MRC8554 | ----- | GTCTTG | ACGAGC | ACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA |
| <i>F. subglutinans</i> MRC6512 | GCCTCGT | CTTGACG | AGCACAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. subglutinans</i> MRC2802 | GCCTTTT | CGAGACG | AGCAGAAC | CGGTGTAG | CCCATGTG | TGTGCAC | GACTCCT | -GTGCGT | CCTCCAG | GGCGGGG | AAGTAGA | |
| <i>F. proliferatum</i> CMW 12864 | CCYTTTT | CRAGGCG | GTACACAAC | CGGKGC | ASCCATG | TGTGTG | TGCC | GACTCCT | -CGTGT | TCCGTCT | AGGGCGGGG | AAGTAGA |
| <i>F. proliferatum</i> MRC 2301 | GCCTTTT | CRAGGCG | GTACACAAC | CGGKGC | ASCCATG | TGTGTG | TGCC | GACTCCT | -CGTGT | TCCGTCT | AGGGCGGGG | AAGTAGA |
| <i>F. proliferatum</i> MRC 12867 | CCTTTTT | CAAGSGG | TATAAK | GGKGC | AGYTTAT | GTGTG | TGCC | GATTCCCT | -CGTGT | TCCGTCT | AGGGCGGGG | AAGTAGA |
| <i>F. proliferatum</i> MRC 8550 | CCTTTTT | CRAGSGG | TATAAK | GGKGC | ASYYTTAT | GTGTG | TGCC | GATTCCCT | -CGTGT | TCCGTCT | AGGGCGGGG | AAGTAGA |
| <i>F. proliferatum</i> MRC 8549 | CCYTTTT | CRAGSGG | TATAAK | GGKGC | ASYTTAT | GTGTG | TGCC | GATTCCCT | -CGTGT | TCCGTCT | AGGGCGGGG | AAGTAGA |
| <i>F. oxysporum</i> CAV105 | ACCTCTT | GAGACG | ACCTCAAC | CGGTACC | ACCGATG | TGTTGG | TCCGG | CTCCT | -GTGCG | GCCGTCC | AGGGCGGGG | AAGTAGA |
| <i>F. oxysporum</i> CAV617 | ACCTTTCC | GAGACG | ACCTCAAC | CGGTACC | ACCGATG | TGTTGG | TCCGG | CTCCT | -GTGCG | GCCGTCC | AGGGCGGGG | AAGTAGA |
| <i>F. oxysporum</i> CAV794 | ACCTCTC | CGAGACG | ACCTCAAC | CGGTACC | ACCGATG | TGTTGG | TCCGG | CTCCT | -GTGCG | GCCGTCC | AGGGCGGGG | AAGTAGA |
| <i>F. oxysporum</i> CAV602 | ACCTCTC | CGAGACG | ACCTCAAC | CGGTACC | ACCCATG | TGTTGG | TCCGG | CTCCT | -GTGCG | GCCGTCC | AGGGCGGGG | AAGTAGA |
| <i>F. oxysporum</i> CAV929 | ACCTTTT | CGAGACG | ACCTCAAC | CGGTACC | ACCCATG | TGTTGG | TCCGG | CTCCT | -GTGCG | GCCGTCC | AGGGCGGGG | AAGTAGA |

| | 1660 | 1670 | 1680 | 1690 | 1700 | 1710 | 1720 | 1730 | 1740 | 1750 | 1760 |
|--------------------------------------|---|-------|---|------|------|------|------|------|------|------|------|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> FL2483 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> FL2497 | CACCCCGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | CACCCMCGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> FL2496 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> SA0041 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | CACCCCGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> SA0010 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> FL2472 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> SA1834 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | CACCCCGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGTCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. circinatum</i> SP5498 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGTCCTTTAGAT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | CACCCACGATCCGATCACCCGCTCCGTCGATTCGCTTTGATAT | ----- | GCGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | CACCCACAATCCGATCACCTCGTCCGCTGATTCACCTTGATAT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGCTTTGATAT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGCTTTGATAT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGCTTTGATAT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGACTTTATCT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGTAACCTTCCAACCT | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGTCCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAGCTTCCAACCT | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGCTCCGCTGGTAT | ----- | GTGAGGACAGGCTAGGGTAGGTTAAAGTCAGGTCTAGGGTAGGCCAGCTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | CACCCACGATCCGATCACCTCGTCCGTCGATTCGCTTTGATAT | ----- | GTGAGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | CACCCACGGTCGGATCACCTCGTCCGTCGATTCGCTTTATCT | ----- | GTGGACAGGCTAGGGTAGGCCAGAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | CACCCACGGTCGGATCACCTCGTCCGTCGATTCGCTTTATCT | ----- | GTGGACAGGCTAGGGTAGGCCAGAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | CACCCACGACCCGATCACCTCGTCCGTCGATTCGCTTTGTTTC | ----- | GTGGACAGGCTAGGGTAGGCCAGAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | CACCTACGATCCGATCACCTCGTCCGTCGATTCGCTTTGATAT | ----- | GTGAGA-----CAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | CACCCACGACCCGCTCACCTCGTCCGTCGATTCGCTTTCAAAT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGCCAACTTCCCACCT | | | | | | | | |
| <i>F. verticillioides</i> NRRL22172 | CACCCACGA-CCGTTACCTCGTCCGTCGATTCGCTCCTCATAT | ----- | GCGGGACAGGCTAGGGTAGTCCAGAGTCGGGTCTAGGGTAGGCCAACTTCCCACCT | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | CACCCACGACCGGTTACCTTGCTGTGTCGATTCGCTTTCAAATCGTCTTCAAGAGT | ----- | GAGACTGGTCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCCACCT | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | TACCCACAGTCCGATCACCTCGCCGTCGATTCGCTTTGGGT | ----- | GAGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | CACCCATGGTCGAATCACCTTGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGTACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | CACCCACGGTCGACTCACCTCGCCGTCGATTCGCTTTGGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAGAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | CACCCACAGTCGATTCACCTCGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | CACCCACGGTCGAATCACCTCGCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | GGTCCAGGGTAGGTTTCGCCAAATCCGTCATCCGGCTTGAATC | ----- | GAAGGACAGGCTAGGGTAGGCCAGAGTCGGGTCTAGGGTAGGCCAACCTCACCCT | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | CACCCACGACCCGATCACCTCGTCCGTCGATTTGTCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | CACCCACGACCCGATCACCTCGTCCGTCGATTTGTCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | CACCCACGACCCGATCACCTCGTCCGTCGATTTGTCTTTATCT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | CACCCCGACCCGATCACCTCGTCCGTCGATTCGCTTTGATAT | ----- | GTGGGACAGGCTAGGGTAGGCCAAAGTCAGGTCTAGGGTAGGCCAACTTCCAACCT | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | CACCCACGGTCGAATCACCTCGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGAACTTCCAGCCT | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | CACCCACGGTCGAATCACCTCGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGAACTTCCAGCCT | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | CACCCACGGTCGAATCACCTCGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGAACTTCCAGCCT | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | CACCCACGGTCGAATCACCTCGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGAACTTCCAGCCT | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | CACCCACGGTCGAATCACCTCGCCCGTCGATTCGCTTTGAGGT | ----- | GAGGGACAGGCTAGGGTAGGCCAAAGTCGGGTCTAGGGTAGGAACTTCCAGCCT | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | GGTCCAGGGTAGGTTCTCCAAATCCGTCATCCGGCTTGAATC | ----- | GAAGGACAGGCTAGGGTAGGCCAGAGTCGGGTCTAGGGTAGGCAGCTCTCACCCT | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | GGTCCAGGGTAGGTTCTCCAAATCCGTCATCCGGCTTGAATC | ----- | GAAGGACAGGCTAGGGTAGGCCAGAGTCGGGTCTAGGGTAGGCAGCTCTCACCCT | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | GGTCCAGGGTAGGTTCTCCAAATCCGTCATCCGGCTTGAATC | ----- | GAAGGACAGGCTAGGGTAGGCCAGAGTCGGGTCTAGGGTAGGCAGCTCTCACCCT | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | GGTCCAGGGTAGGTTCTCCAAATCCGTCATCCGGCTTGAATC | ----- | GAAGGACAGGTTAGGGTAGGCCAGAGTCGGGTTTAGGGTAGGCCAACCTCACCCT | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | GGTCCAGGGTAGGTTCTCCAAATCCGTCATCCGGCTTGAATC | ----- | GAAGGACAGGTTAGGGTAGGCCAGAGTCGGGTTTAGGGTAGGCCAACCTCACCCT | | | | | | | | |

| | 1770 | 1780 | 1790 | 1800 | 1810 | 1820 | 1830 | 1840 | 1850 | 1860 | 1870 |
|--------------------------------------|---|------|------|------|------|------|------|------|------|------|------|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCCCCCCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | CAGAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACTACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGTACGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | CAAAGCTGCCTACCCGGGAGTCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGATACCCCCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | CGAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | CAAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | CAAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGAGACCACCTCCATCGCATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | CAAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGCATTGCGCGTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | CGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | CGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | CGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | CGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | CGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | CGAAGTGGTCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCATGACGACGGGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | CAAAGCTGCCTACCCGGGAGCCAATTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | TGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | CGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 12867 | TGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | TGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGGATGCGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | TGAAGCTGCCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCAGGAGACGGGACCACCTCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | CGAAGTGGTCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCATGACGACGGGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | CGAAGTGGTCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCATGACGACGGGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | CGAAGTGGTCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCATGACGACGGGACCACCACCATTGAACTCGCCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | CGAAGTGGTCTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGCATGACGACGGGACTACCACCATTGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | CGAAGTGGTTTACCCGGTAGTCAACTTCAATCGCCTCTCAGCCTCCACAGACCTCGAAGACGACGGGACTACCCCATCGGATTGCGCTTGGTCGAAATAGTCGGTA | | | | | | | | | | |

| | 1990 | 2000 | 2010 | 2020 | 2030 | 2040 | 2050 | 2060 | 2070 | 2080 | 2090 |
|--------------------------------------|---|---------------------|--|------|------|------|------|------|------|------|------|
| <i>F. circinatum</i> NRRL25331 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> FL2483 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> FL2497 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> FL2496 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> SA0041 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> SA0010 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> FL2472 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> SA1834 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. circinatum</i> SP5498 | | ACGAGGCGGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGTGAGGACAAAACCGGGCGAGCAACCTC | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGGAAAACCGGACAAGCAACCTC | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | | ACGAAGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGCGAGGACAAAACCGGGCGAGCAACCTC | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | | ACGAAGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGTGAGGACAAAACCGGGCGAGCAACCTC | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | | ACGAAGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGTGAGGACAAAACCGGGCGAGCAACCTC | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGTGAGGACAAAACCGGGCGAGCAACCTC | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | | ACGAGGCAGTCTGGCGTGC | CGGCCACTAAAACGGTCTCGGAGGTATATGAGAAGGGAGCAAATCCGGCCGAGCCTGAAAGGGTGAGGACAAAACCGGGCGAGCAACCTC | | | | | | | | |

| | 2320 | 2330 | 2340 | 2350 | 2360 | 2370 | 2380 | 2390 | 2400 | 2410 | 2420 |
|--------------------------------------|--|------|------|------|------|------|------|------|------|------|------|
| | | | | | | | | | | | |
| <i>F. circinatum</i> NRRL25331 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. verticillioideus</i> NRRL22172 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | CCTGAGCAACGGGAGGTAACCTCTCGCCGTGGACACCGGAATGGTAGAAGCGGCGTGCTGCGTCCTCCTCTTGGGGCCCCTAAGCCACACCTCCCACAGCGGGTTCGGTG | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | ----- | | | | | | | | | | |

| | 2430 | 2440 | 2450 | 2460 | 2470 | 2480 | 2490 | 2500 | 2510 | 2520 | 2530 |
|--------------------------------------|--|------|------|------|------|------|------|------|------|------|------|
| <i>F. circinatum</i> NRRL25331 | | | | | | | | | | | |
| <i>F. circinatum</i> CAL2511 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> FL2483 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> FL2497 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> MEX4900 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> FL2496 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> SA0041 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> MEX2248 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> SA0010 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTC-CCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> FL2472 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> SA1834 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> MEX2265 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. circinatum</i> SP5498 | CGGCGGACGGACGCCCTGAGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGCTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. subglutinans</i> NRRL22016 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. begoniae</i> NRRL25300 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25346 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25204 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26756 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL26757 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25623 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25195 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. succisae</i> NRRL13613 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. anthophilum</i> NRRL13602 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL29123 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. bulbicola</i> NRRL13618 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25807 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>Fusarium</i> sp. NRRL25622 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. bacterioides</i> NRRL20476 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. pseudoanthophilum</i> NRRL2520 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. verticilliioides</i> NRRL22172 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. thapsinum</i> NRRL22045 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. concentricum</i> NRRL25303 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. fractiflexum</i> NRRL28852 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. fujikuroi</i> NRRL13566 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. globosum</i> NRRL26131 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. proliferatum</i> NRRL22944 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. oxysporum</i> NRRL22902 | CGGCGGACGGACGCCCTGGGGAATTTAGAGGGGGAAAGCGGATTGCCCTAGCGGTGTTGTTGGCCCTGCCGACCTCACTGCGAAAGGCGCGACTTCACCGTCGCCACCCA | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8553 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC8554 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC6512 | ----- | | | | | | | | | | |
| <i>F. subglutinans</i> MRC2802 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12864 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 2301 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> CMW 12867 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8550 | ----- | | | | | | | | | | |
| <i>F. proliferatum</i> MRC 8549 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV105 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV617 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV794 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV602 | ----- | | | | | | | | | | |
| <i>F. oxysporum</i> CAV929 | ----- | | | | | | | | | | |

| | 2540 | 2550 |
|--------------------------------------|---------------------------------|------|
| | | |
| <i>F. circinatum</i> NRRL25331 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> CAL2511 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> FL2483 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> FL2497 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> MEX4900 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> FL2496 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> SA0041 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> MEX2248 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> SA0010 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> FL2472 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> SA1834 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> MEX2265 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. circinatum</i> SP5498 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. subglutinans</i> NRRL22016 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. begoniae</i> NRRL25300 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL25346 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL25204 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL26756 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL26757 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL25623 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL25195 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. succisae</i> NRRL13613 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. anthophilum</i> NRRL13602 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL29123 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. bulbicola</i> NRRL13618 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL25807 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>Fusarium</i> sp. NRRL25622 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. bacterioides</i> NRRL20476 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. pseudoanthophilum</i> NRRL2520 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. verticillioides</i> NRRL22172 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. thapsinum</i> NRRL22045 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. concentricum</i> NRRL25303 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. fractiflexum</i> NRRL28852 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. fujikuroi</i> NRRL13566 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. globosum</i> NRRL26131 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. proliferatum</i> NRRL22944 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. oxysporum</i> NRRL22902 | GTAACCTTGTCTCTCCGGCGCCTCAG | |
| <i>F. subglutinans</i> MRC8553 | ----- | |
| <i>F. subglutinans</i> MRC8554 | ----- | |
| <i>F. subglutinans</i> MRC6512 | ----- | |
| <i>F. subglutinans</i> MRC2802 | ----- | |
| <i>F. proliferatum</i> CMW 12864 | ----- | |
| <i>F. proliferatum</i> MRC 2301 | ----- | |
| <i>F. proliferatum</i> CMW 12867 | ----- | |
| <i>F. proliferatum</i> MRC 8550 | ----- | |
| <i>F. proliferatum</i> MRC 8549 | ----- | |
| <i>F. oxysporum</i> CAV105 | ----- | |
| <i>F. oxysporum</i> CAV617 | ----- | |
| <i>F. oxysporum</i> CAV794 | ----- | |
| <i>F. oxysporum</i> CAV602 | ----- | |
| <i>F. oxysporum</i> CAV929 | ----- | |