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Proteolytic and amylolytic enzymes for bacterial biofilm control

By

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I hereby declare that the thesis entitled **“PROTEOLYTIC AND AMYLOLYTIC ENZYMES FOR BACTERIAL BIOFILM CONTROL”** which I hereby submit for the degree Philosophiae Doctor is my own original work and has not previously in its entirety or part been submitted at any university for a degree.

Signature: _____

Date: _____



Table of contents

List of tables.....	viii
List of figures.....	x
List of abbreviations	xii
Conference contributions	xiv
Publications.....	xiv
Acknowledgements.....	xv
Quotes	xvii
Dedications	xviii
Summary.....	1
Chapter 1	
Introduction.....	5
1.1 Microbial biofilms	5
1.2 Enzymes for biofilm control	7
1.3 References.....	9
Chapter 2	
Literature review.....	12
2.1 Defining biofilms	12

2.2 Biofilm formation and stages involving during biofilm development.....	13
2.2.1 The primary stage	13
2.2.2 The secondary stage.....	14
2.2.3 Micro colony formation	15
2.2.4 Formation of three dimensional structures	16
2.2.5 Biofilm maturation.....	16
2.2.6 Detachment and dispersal of biofilm cells.....	17
2.2.7 Summarized life cycle of biofilms.....	18
2.3 Factors affecting the growth and development of biofilms	19
2.3.1 Nutrients.....	19
2.3.2 Temperature effects	21
2.3.3 Surface condition	21
2.3.4 Velocity, turbulence and hydrodynamics	23
2.3.5 Effects of particles	25
2.3.6 Gene regulation.....	26
2.3.7 Quorum sensing (QS)	26
2.3.8 Properties of the cells.....	28
2.4 Description of extracellular polymeric structure (EPS).....	32
2.4.1 EPS production	32
2.4.2 EPS composition.....	33



2.4.3 EPS chemistry	36
2.4.4 Role of EPS	37
2.4.5 Heterogeneity of EPS structures	38
2.4.6 Cell structures associated with EPS	38
2.5 Biofilm producing enzymes	39
2.5.1 Extracellular enzymes	39
2.5.2 Enzyme activity in sludge flocs	39
2.5.3 Enzyme mediated resistance	40
2.5.4 Application of enzymes for biofilm control	41
Summarised literature review	52
2.7 References	54
Chapter 3	
Microplate assay for screening of proteolytic and amylase enzymes for biofilm removal	68
3.1 Abstract	68
3.2 Introduction	69
3.3 Materials and methods	72
3.3.1 Bacterial inoculum used for biofilm growth	72
3.3.2 Enzymes tested for biofilm removal	73
3.3.3 Micro plate assay for the evaluation of enzyme efficacy on biofilms	74



3.4 Results.....	76	
3.5 Discussion.....	80	
3.6 Conclusion	82	
3.7 References.....	83	
Chapter 4		
Spectrophotometric assay for the evaluation of proteolytic and amylase enzymes for biofilm removal.....		86
4.1 Abstract.....	86	
4.2 Introduction.....	87	
4.3 Materials and methods	90	
4.3.1 Bacterial inoculum used to grow biofilms	90	
4.3.2 Enzymes tested for biofilm removal	90	
4.3.4 Spectrophotometric assay for the assessment of enzymes for biofilm removal	90	
4.3.5 Quantitative determination of viable cells	91	
4.4 Results.....	93	
4.5 Discussion.....	100	
4.6 Conclusion	102	
4.7 References.....	104	

Chapter 5

The chemical composition of EPS in <i>Pseudomonas fluorescens</i> and mixed bacterial species biofilm and application of enzymes for EPS degradation.....	106
5.1 Abstract.....	106
5.2 Introduction.....	107
5.3 Materials and methods.....	109
5.3.1 Bacterial inoculum used for biofilm growth.....	109
5.3.2 Quantitative determination of viable cells.....	110
5.3.3 Extraction of extra cellular polymeric substances (EPS).....	110
5.3.4 Determination of the carbohydrate concentration in the EPS.....	110
5.3.5 Determination of the protein concentration in the EPS.....	111
5.3.6 Degradation of biofilm EPS.....	111
5.3.8 Testing of enzymes for the removal of biofilm cells on the glass wool.....	111
5.3.9 Sample preparation for Scanning Electron Microscopy.....	112
5.4 Results.....	113
5.5 Discussion.....	124
5.6 Conclusion.....	129
5.7 References.....	131
Chapter 6	
General discussion.....	136
Appendix.....	144



List of tables

Table 2.1 Available enzymes used for the control of biofilms	45
Table 2.2 The general composition of some bacterial extracellular polymeric substances (EPS) including humic substances.....	51
Table 3.3.1 Enzymes used for the removal of <i>Pseudomonas fluorescens</i> and mixed bacterial species biofilm	74
Table 3.4.1 Effects of enzymes on biofilms as measured by the mean percentage reduction	76
Table 4.3.1 Enzymes used for biofilm removal.....	92
Table 4.4.1 Spectrophotometric evaluation of cell density before and after enzymatic treatment tested at 1 U/ml.....	93
Table 4.4.2 Spectrophotometric evaluation of cell density before and after enzymatic treatment tested at 2 U/ml.....	94
Table 5.2.1 Enzymes used for biofilm removal.....	112
Table 5.4.1 Comparison of viable cells between fed and unfed <i>Pseudomonas fluorescens</i> biofilms	114
Table 5.4.2 Comparison of viable biofilm cells between fed and unfed mixed bacterial biofilms	114
Table 5.4.3 Comparison of extracted EPS, protein and carbohydrate concentrations produced from fed and unfed <i>Pseudomonas fluorescens</i> biofilm.....	115



Table 5.4.4 Comparison of extracted EPS, protein and carbohydrate concentrations produced from fed and unfed mixed bacterial species biofilm..... 115



List of figures

Fig. 2.1 Schematic illustrations of biofilm formation and development.	19
Fig. 2.2 Schematic illustration of processes involved in biofilm formation and development.....	21
Fig. 2.3 Schematic illustrations of the structure/ activity relationship in biofilms	25
Fig. 2.4 Quorum sensing processes in bacterial biofilms	28
Fig. 2.5 Flow chart of biofilm formation on a surface.....	31
Fig. 2.6 Illustrations of the processes taking place during the multimetal resistance and tolerance in microbial biofilms	41
Fig. 2.7 Schematic overview of the structural components of extracellular polymeric substances (EPS) involved in biofilm formation	44
Fig. 2.8 Illustrations of various α -amylases with applications to conservation practise ..	50
Fig. 3.4.1 Enzyme efficacy for removal of (A, C) <i>Pseudomonas fluorescens</i> biofilms and (B, D) mixed bacterial species biofilms treated at (A, B) 1 U/ml and (C, D) 2 U/ml.	77
Fig. 4.4.1 Cell density of (A, B) <i>Pseudomonas fluorescens</i> and (C, D) mixed bacterial species treated at (A, C) 1 U/ml and (B, D) 2 U/ml.....	95
Fig 4.4.2 (A) <i>Pseudomonas fluorescens</i> and (B) mixed bacterial species cells recovered after enzymatic treatment at 1 U/ml.....	98
Fig 4.4.3 (A) <i>Pseudomonas fluorescens</i> and (B) mixed bacterial species cells recovered after enzymatic treatment at 2 U/ml.....	98

Fig. 4.4.4 Photographs showing cells recovered after enzymatic treatment.....	100
Fig. 5.4.1 Biofilm growth of (A) <i>Pseudomonas fluorescens</i> and (B) mixed bacterial species.....	114
Fig. 5.4.2 Effects of enzymes on BSA and glucose (D) the non treated extracellular polymeric substance (EPS).....	118
Fig. 5.4.3 Degradation activity of protease enzymes on <i>Pseudomonas fluorescens</i> biofilms EPS.....	118
Fig. 5.4.4 Degradation activity of amylase enzymes tested individually on <i>Pseudomonas fluorescens</i> biofilm EPS.....	119
Fig. 5.4.5 Scanning Electron Microscope analysis of the degradation activity of enzymes on extra cellular polymeric substances (EPS) of 7d old <i>Pseudomonas fluorescens</i> biofilm attached to glass wool after 24h incubation at 26 ^o C.....	120
Fig. 5.4.6 Degradation activity of mixed enzymes on <i>Pseudomonas fluorescens</i> and mixed bacterial species biofilm EPS.....	121
Fig. 5.4.7 Scanning Electron Microscopy analysis of the degradation activity of mixed enzymes on extra cellular polymeric substances (EPS) of 7d old <i>Pseudomonas fluorescens</i> biofilms attached on the glass wool after 24h incubation at 26 ^o C.....	122
Fig. 5.4.8 Scanning Electron Microscope analysis of the degradation activity of mixed enzymes on extra cellular polymeric substances (EPS) of 7d old mixed bacterial species biofilm (Gram negative and positive bacteria) attached on the glass wool after 24h incubation at 30 ^o C.....	123



List of abbreviations

- AMG – Amyloglucosidase
- AMP – Ampicillin
- BAN – Bacterial Amylase Novo
- BRR – Biofilm Removal Reactor
- BSA – Bovine Serum Albumin
- CBA – Chlorobenzoic acid
- CF100XNB – Continuous fed with 100 times Nutrient Broth
- CFU – Colony Forming Units
- DNA – Deoxyribonucleic acid
- DS – Distribution System
- EPS – Extracellular Polymeric Substance
- HDPE – High density polyethelene
- HOC – Hydrophobic organic compound
- HUS – Hemolytic uremic syndrome
- kDa – kilodalton
- LPS – Lipopolysaccharide

- MIC – Microbiologically induced corrosion
- MIC – Minimum inhibitory concentration
- NAG – N – acetylglucosamide
- NaOH – Sodium Hydroxide
- OD – Optical density
- OMP – Outer Membrane Protein
- PE – Poly ethelene
- PIA – Polysaccharide intracellular adhesin
- PR – Percentage Reduction
- PVC – Polyvinyl chloride
- QS – Quarum Sensing
- SEM – Scanning Electron Microscopy
- TTP – Thrombocytopenic pupura
- UV – Ultra violet
- WAN – Without additional nutrients



Conference contributions

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Publications

Paper

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Quotes

- An aim in life is the only fortune worth finding

(Jacqueline Kennedy Onassis)

- All personal achievements start within the mind of the individual

(W Clement Stone)

- Every great success is an accumulation of thousands of ordinary efforts that no one sees or appreciates

(Brian Tracy)

- Selecting a challenge and meeting it creates a sense of self empowerment that becomes the ground for further successful challenges

(Julia Cameron)

- Treat people as if they were what they ought to be and you will help them become what they are capable of becoming.

(Johann Wolfgang Von Goethe)

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Dedications

*This work is dedicated to my son “Thuto” and my late brother “Douglas”
This is it brother, it happened! You really wanted to witness the ceremony
when I would be receiving my PhD degree certificate, but death destroyed
all of your plans. It is well brother and i promise to stick to those rules you
have coached me on “HOW TO LIVE A SIMPLE BUT LUXURIOUS LIFE.
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Summary

Biofilms are characterized by surface attachment, structural heterogeneity; genetic diversity; complex community interactions and an extracellular matrix of polymeric substances (EPS). Biofilms deposit and adhere to all surfaces that are immersed in aqueous environments. EPS serves many functions including: facilitation of the initial attachment of bacterial cells to a surface; formation and maintenance of the micro colony; enables the bacteria to capture nutrients; causes biofouling; cell-cell communication and enhances bacterial resistance antimicrobial agents. EPS also function as a stabilizer of the biofilm structure and as a barrier against hostile environments. Extracelullar polymeric substances are composed of a wide variety of materials including polysaccharides, proteins, nucleic acid, uronic acid, DNA, lipid and even humid substances.

EPS can be hydrophilic or hydrophobic depending on the structural components making up such EPS and the environmental conditions were the biofilms are developing. The exopolysachharides (EPS) synthesized by microbial cells vary greatly in their composition and in their chemical and physical properties within the bacterial strains. Due to variety in the structural components of the bacterial EPS, removal of biofilms by compounds that have no effects on the biofilm EPS would be difficult. Enzymes are proven to be effective in degrading biofilm EPS. The manner in which enzymes degrade the biofilm EPS is through binding and hydrolysis of the EPS components (proteins and carbohydrates) molecules and converting them into smaller units that can be transported through the cell membranes and then be metabolized.

The objectives of this study were to grow *Pseudomonas fluorescens* and mixed bacterial species biofilms in nutrient rich and nutrient limited medium conditions; to determine the EPS, protein and carbohydrate concentrations of the biofilm grown in rich and in limited nutrient conditions and to test the efficiency of protease and amylase enzymes for the degradation of the EPS and biofilm removal. In the results, there was a slight difference in the number of viable cells grown in biofilms that were fed than the cells of the unfed biofilms. As a result, the EPS, protein and carbohydrate concentrations were higher in the

fed biofilms than the unfed biofilms. There are contradictory reports about the composition of EPS especially with the ratio of carbohydrate to protein. Some of these reports indicate that certain biofilms EPS have bigger proportion of proteins and some found polysaccharides to be the dominant composition of the EPS of the biofilms. Nonetheless, the quantity and the composition of the EPS produced by bacterial biofilms depend on a number of factors such as microbial species, growth phase and the type of limiting substrate.

Enzymes were tested individually and in combination for the degradation of biofilm EPS. For efficient removal of biofilm, it is important that the structural components of the biofilm EPS should be known before application of the relevant enzymes. In this study, the test enzymes were effective for the degradation of the biofilm EPS except for the protease Polarzyme which had no activity. The reason for the inefficiency of Polarzyme may be due to its incompatibility with the specific protein structural components of the biofilm EPS tested in this study. The manner in which the enzymes degrade the biofilm EPS is through binding and hydrolysis of the protein and carbohydrate molecules and converting them into smaller units that can be transported through the cell membranes and then be metabolized. In addition, the mode of enzymatic action will depend on the specific EPS components and this in turn will determine its efficacy. The protease enzymes tested individually and in combination were most effective for EPS degradation. The efficiency of the proteases may be due to their broad spectrum activity in degrading a variety of proteins acting partly as the multi structural components of *Pseudomonas fluorescens* and mixed bacterial species biofilm EPS.

On the other hand, amylase enzymes tested individually and in combination was less effective for the EPS degradation. The structures of polysaccharides synthesized by microbial cells vary. Microbial exopolysaccharides are comprised of either homopolysaccharides or heteropolysaccharides. A number of lactic acid bacteria produce heteropolysaccharides and these molecules form from repeating units of monosaccharides including D- glucose, D- galactose, L- fructose, L- rhamnose, D- glucuronic acid, L- guluronic acid and D- mannuronic acid. The type of both linkages between monosaccharides units and the branching of the chain determines the physical

properties of the microbial heteropolysaccharides. Due to a wide range of linkages and the complexity of polysaccharides structures, it would therefore be difficult for the amylases to break down the bond linkages and the monomers making up polysaccharides which determine the physical and chemical structure of the EPS. It was therefore not surprising that the amylase enzymes tested for the degradation of *Pseudomonas fluorescens* and mixed bacterial species biofilms, were less effective than the proteases. Hence, when the amylase enzymes were tested in combination with the protease enzymes, efficiency improved. It was therefore concluded that the protease enzymes were the primary remedial compounds and the amylase enzymes were the secondary remedial compounds.

Conclusion

If a compound or compounds capable of destroying all the structural components of different EPS that are produced by different biofilms growing under different conditions is found then the “city of microbes” (biofilms) would be destroyed permanently. If only an enzyme or enzymatic mixture capable of shutting down or deactivating the quorum sensing systems of different biofilm EPS could be found, then there would not be any formation of biofilms. In this study, protease enzymes tested individually and in combination were the most effective in the degradation of biofilm EPS than the amylase enzymes resulting in the reduction of large population of the biofilm cells attached on the substratum.

Recommendation

Amylase enzymes tested individually and in combination were less efficient for the degradation of the biofilm EPS and biofilm removal. This may be due to the complex structure of the exopolysaccharides synthesized by different biofilms. Also, the bond linkages between monosaccharides units and the branching of the chain complex the structures and as a result confer in the physical properties of the microbial biofilms. Hence, when the amylase enzymes were tested in combination with the protease enzymes, activity improved. For efficient degradation of biofilm EPS, it is therefore recommended that, protease and amylase enzymes should be tested in combination. In

addition, the structure of the biofilm EPS should be investigated so that relevant enzymatic mixtures are tested for biofilm removal.