

# **FUNCTIONAL ANALYSIS OF CYCLIN D2;1-TYPE GENES EXPRESSED IN TRANSGENIC BANANA PLANTS**

By

DAVID TALENGERA

Thesis submitted in partial fulfillment of the requirement for the degree

PHILOSOPHIAE DOCTOR

Forestry and Agricultural Biotechnology Institute (FABI)

Department of Plant Science

Faculty of Natural and Agricultural Sciences

University of Pretoria

Supervisors:

PROF. K. KUNERT

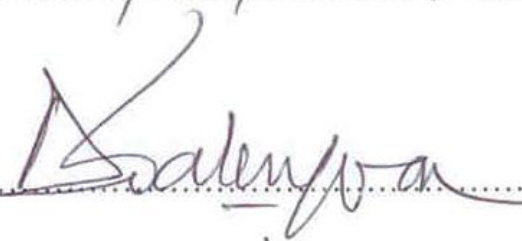
PROF. G.T.S. BEEMSTER

2011

## DECLARATION

This thesis is my original work and has not been presented for a degree in any other University.

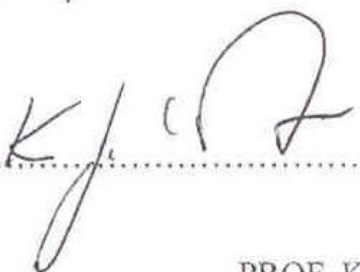
Date..... 23/2/2011 .....

Signed.....  .....

DAVID TALENGERA  
B.Sc. (Agric.), M.Sc. (Agric.) Makerere University, Kampala

This thesis has been submitted for examination with my approval as the University Supervisor.

Date..... 1/3/2011 .....

Signed.....  .....

PROF. KARL KUNERT

## DEDICATION

To my wife Susan Talengera, daughter Norah Nampiima, and sons Andrew Talengera and Simon Mukoka.

## **ACKNOWLEDGEMENTS**

I wish to express my profound gratitude to my supervisors Prof. Karl Kunert and Prof. Gerrit T.S. Beemster for their advice and guidance thorough my PhD study. Prof. Wilberforce K. Tushemereirwe, who is also the team leader of the National Banana Research Program, gave valuable advice and promptly availed the research requirements. All their dedication is highly appreciated.

I am also grateful to Prof. Dirk. Inzé, Director of the Department of Systems Biology of the University of Ghent, together with Prof. Michael J. Wingfield, the Director of Forestry and Agricultural Biotechnology Institute of the University of Pretoria, for the opportunity they offered me to work in their conducive research setups. For the period I worked in these institutes and at my home institute, the National Agricultural Research Laboratories Institute at Kawanda, I interacted with a number of people. Among them were Drs. Bart Rymen, Andrew Kiggundu, Geoffrey Arinaitwe, Charles Changa, Priver Namanya, Urte Shlüter, Getu Beyene, Juan Vorster and Ziaratnia Mahdi. Dr. Fabio Fiorani supplied me with useful constructs. These people together with colleagues of the molecular plant physiology group at the University Pretoria: Rosita Endah, Berhanu Amsalu Fenta, Priyan Pillay, Tsolofelo Kibido, Stefan Van Wyk, Rynhard Smit and Magdeleen DuPlessis are acknowledged for all their assistance and teamwork that made my research work enjoyable. I am very thankful to Drs. Deborah P. Delmer, Nancy Terryn and Rachel Chikwamba for their opinions and continuous encouragement that maintained my zeal to conduct this study.

All the staff of the National Banana Research Program and most especially the personnel of the Biotechnology Unit are commended for their cooperation and support. Mr. Bonny Oloka and Ms. Clara Samukoya who closely assisted me in my laboratory work are acknowledged for their effectiveness. I thank my wife Susan Talengera for her care and support. I appreciate her together with our children for their patience and for understanding my student life.

The Rockefeller Foundation funded this PhD work through the National Agricultural Research Organization under the project whose aim was to explore the potential of biotechnology to improve the East African highland banana architecture. This sponsorship is highly appreciated.

## SUMMARY

Early maturity is one of the most important aerial growth traits next to bunch size, in determining banana productivity. However, low seed fertility in banana and the lack of breeding lines limit the application of conventional breeding for this trait. Genetic transformation with a *CyclinD2*-type gene, responsible for the CyclinD2 protein sub unit, which modulates the cell cycle progression at the G1/S phase, enhanced growth of tobacco plants. On this basis, investigations were carried out on the possibility of increasing the growth rate of banana plants through transformation with and expression of a *CyclinD2*-type coding sequence. *Arabidopsis thaliana*; *CyclinD2;1* (*Arath*; *CycD2;1*) gene and its banana ortholog were over-expressed in banana to test their growth enhancement potential. The banana *CyclinD2;1* (*Musac*; *CycD2;1*) was isolated from an East African highland cooking banana (AAA) cultivar ‘Nakasabira’ and a cDNA was created by PCR using degenerated primers which was followed by genome walking. Characterization of the banana cyclin protein revealed an IWKVHAHY motif that was found to be conserved across the Musaceae family. Phylogenic analysis revealed a higher protein sequence identity of this banana cyclin to *CyclinD2;1* of monocot plants than that of *Arabidopsis*. This cyclin was also found to be expressed highly in meristematic tissue which linked it to the cell cycle. The coding sequence was submitted to the GeneBank under accession number HQ839770. *Arabidopsis* and banana *CyclinD2;1* gene coding sequences under the control of a constitutive promoter were used to transform embryogenic cells of the banana cultivar ‘Sukalindiizi’ (AAB) using the *Agrobacterium* transformation system. A higher relative expression of *Arath*; *CyclinD2;1* was found in the shoot than in the root apices and expression reduced transcript amounts of the endogenous banana *CyclinD2;1*. Plants of transformed banana line D2-41 had the highest *Arath*; *CyclinD2;1* transcript amount and exhibited a significantly faster leaf elongation rate, better root growth, faster first leaf opening and a bigger lamina composed of bigger epidermal cells than non-transformed control plants. Banana plants transformed with *Musac*; *CyclinD2;1* had a higher transcript amount of the transgene in the root apices when compared to the shoot apices. The higher transcript amount in the roots of plants of transformed line NKS-30 was related to faster

root growth and development of an extensive root system. Overall, this study has provided evidence that expression of cyclin coding sequences in transformed banana is related to growth promotion. Specifically, *Arath;CyclinD2;1* promoted shoot growth while the *Musa* homolog promoted root growth. Shoot and root growth phenotypes obtained in this study might have the potential to improve banana productivity in terms of short plant growth cycle, increased bunch weight, improved plant anchorage and increased plant resistance to root nematode damage. Future work should assess the produced plants in the field to allow transformed plants to exhibit their full potential and to be able to fully evaluate the vegetative and flowering phases.

## **Thesis composition**

*Chapter 1* describes the botany of the banana plants and the demand for a fast maturing banana plant and extensive root growth and the possible contribution of these characteristics to productivity. This chapter also contains the limitations of conventional methods to breeding for these traits and the molecular option targeting the *CyclinD2*-type gene to accelerate the cell cycle. *Chapter 2* covers the results obtained for isolation and characterization of the *CyclinD2;1*-type gene ortholog from banana including a phylogenetic analysis of the isolated banana cyclin gene sequence. *Chapter 3* describes the creation of Arabidopsis and banana *CyclinD2;1* gene constructs suitable for banana transformation, their delivery into transformation into banana cells using the Agrobacterium-based transformation system, regeneration of transformed plants and finally the detection of the transferred transgene in transformed banana using molecular biology tools. *Chapter 4* reports about the expression analysis of both the endogenous cyclin banana gene and of transferred cyclin transgenes in selected regenerated banana plants using the technique of quantitative RT-PCR. *Chapter 5* outlines the phenotypic evaluation of transformed banana plants to detect any changed shoot and root growth characteristics and to determine a possible relationship between changed characteristics and transcript amounts of transgenes. Finally, *chapter 6* summarizes the novel results found in this study and evaluates the set initial working hypothesis. Further, recommendations for further research work, based on the results obtained in this study, are outlined.



## Table of contents

DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENTS	iv
SUMMARY	vi
Thesis composition	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
CHAPTER ONE: INTRODUCTION	1
1.1 The banana plant	2
1.2 Banana breeding	2
1.2.1 Conventional breeding	2
1.2.2 Genetic engineering of banana	3
1.2.3 Factors influencing transgene expression	4
1.3 Banana growth and development	5
1.3.1 Banana leaf and root growth and productivity	6
1.3.2 Biotechnology and banana plant architecture	7
1.4 Plant cell cycle	7
1.4.1 Classification of cyclins	10
1.4.2 Regulation of cyclins	10
1.4.3 Role of the cell cycle in plant development	11
1.4.4 Cyclin expression studies	11
1.5 Working hypothesis and objectives	14
CHAPTER TWO: ISOLATION OF A BANANA <i>CYCLIN D2;1</i> GENE HOMOLOG	15
2.1 Introduction	16
2.2 Materials and Methods	17
2.2.1 RNA isolation	17
2.2.2 cDNA synthesis	17
2.2.3 PCR reactions and purification	18
2.2.4 Degenerated primer design	18
2.2.5 Genome walking	19
2.2.6 Preparation of restriction enzyme-digested genomic DNA	20
2.2.7 DNA cloning	21

2.2.8	DNA sequencing	22
2.2.9	Isolation of <i>cyclin</i> sequences	22
2.2.10	Bioinformatics	23
2.3	Results	24
2.3.1	Cyclin isolation	24
2.3.2	Molecular characterization of the coding sequence	26
2.3.3	Molecular characterization of the genomic sequence	30
2.4	Discussion	32
CHAPTER THREE: DESIGNING <i>CYCLIN</i> GENE CONSTRUCTS AND TRANSFORMATION OF BANANA		34
3.1	Introduction	35
3.2	Materials and Methods	36
3.2.1	Construction of transformation vector	36
3.2.2	Transformation of <i>Agrobacterium</i>	40
3.2.3	Transformation of banana cells	41
3.2.4	Molecular analysis of regenerated banana	44
3.3	Results	47
3.3.1	Gene construct	47
3.3.2	Plant transformation, selection and regeneration	51
3.3.3	Confirmation of transgene insertion	55
3.4	Discussion	59
CHAPTER FOUR: TRANSCRIPTION ANALYSIS OF CYLIN TRANSGENES		61
4.1	Introduction	62
4.2	Materials and Methods	63
4.2.1	RNA isolation and cDNA synthesis	63
4.2.2	Semi-quantitative PCR	64
4.2.3	Quantitative real-time PCR (qRT-PCR)	65
4.2.4	Primer design	66
4.3	Results	67
4.3.1	<i>Musac; CyclinD2;1</i> gene expression profiling	67
4.3.2	Integrity of <i>Arath; CyclinD2;1</i> transcripts	67
4.3.3	Gene expression analysis	69

4.4	Discussion	74
CHAPTER FIVE: PHENOTYPIC EVALUATION OF TRANSFORMED BANANA PLANTS EXPRESSING D-TYPE CYCLINS		76
5.1	Introduction	77
5.2	Materials and methods	78
5.2.1	Transgenic lines evaluated in the study	78
5.2.2	Establishment of transformed plantlets	78
5.2.3	Phenotypic evaluation of transgenic plants	79
5.2.4	Data analysis	82
5.3	Results	83
5.3.1	Leaf growth of <i>Arath;CyclinD2;1</i> expressing banana	83
5.3.2	Root growth of <i>Arath;CyclinD2;1</i> expressing banana	84
5.3.3	Leaf growth of <i>Musac;CyclinD2;1</i> expressing banana	89
5.3.4	Root growth of <i>Musac;CyclinD2;1</i> expressing banana	89
5.4	Discussion	94
CHAPTER SIX: GENERAL CONCLUSION AND RECOMMENDATIONS		96
REFERENCES		101
APPENDICES		117

## LIST OF TABLES

<b>Table 1.1</b>	Over-expression studies of cyclins and resultant phenotypes	13
<b>Table 2.1</b>	Banana <i>CyclinD2;1</i> gene structure represented by size and composition of the exons and intron, and splicing sites	31
<b>Table 3.1</b>	Regeneration frequency of Sukalindiizi cells transformed with <i>Arath;CycD2:1</i>	54
<b>Table 3.2</b>	Regeneration frequency of Sukalindiizi cells transformed with <i>Musac;Cyclin D2;1</i>	54
<b>Table 3.3</b>	Number of positive transformants from randomly selected regenerants	54
<b>Table 4.1</b>	Primers used in real-time qRT-PCR	66
<b>Table 4.1</b>	Summary of cyclin transcription levels in shoots and root apices	73
<b>Table 5.1</b>	Leaf growth characteristics of transgenic banana plants expressing <i>Arath;Cyclin D2;1</i>	86
<b>Table 5.2</b>	Comparison of plant and leaf growth between <i>Arath;CycD2;1</i> transgenic and non-transgenic control plants	87
<b>Table 5.3</b>	Root growth of banana transgenic plants expressing <i>Arath;Cyclin D2;1</i> and non-transgenic control plants	88
<b>Table 5.4</b>	Comparison of plant and leaf growth parameters of <i>Musac;CycD2;1</i> transgenic and non-transgenic control plants	90
<b>Table 5.5</b>	Root growth of transgenic banana plants expressing <i>Musac;CyclinD2;1</i> and non-transgenic	91

## LIST OF FIGURES

<b>Fig. 1.1</b>	Growth cycle of a banana plant	5
<b>Fig. 1.2</b>	Plant cell cycle	9
<b>Fig. 2.1</b>	Isolation of <i>CyclinD2;1</i> gene homolog from banana	25
<b>Fig. 2.2</b>	Nucleotide and deduced amino acid sequences of <i>Musac;CyclinD2;1</i> gene	27
<b>Fig. 2.3</b>	Multiple alignment of <i>CyclinD2;1</i> amino acid sequence of banana with <i>CyclinD2;1</i> from Arabidopsis and other representative species	28
<b>Fig. 2.4</b>	Phylogenic relationship between <i>Musac;CyclinD2;1</i> and other plant cyclins	29
<b>Fig. 2.5</b>	Presence of <i>Musac;Cyclin2;1</i> coding region in selected accessions after PCR reaction with genomic DNA	30
<b>Fig. 3.1</b>	Isolation of maize ubiquitin-1 promoter	48
<b>Fig. 3.2</b>	Construction of <i>Arath;CyclinD2;1</i> expression vector	49
<b>Fig. 3.3</b>	Construction of the <i>Musac;CyclinD2;1</i> expression vector	50
<b>Fig. 3.4</b>	<i>Arath;CyclinD2;1</i> gene cassette in expression vector pExpression B4-UBiB1-D2-B3 vector and <i>Musac;CyclinD2;1</i> gene cassette in expression vector pBIN:35S: <i>Musac;CyclinD2;1</i>	51
<b>Fig. 3.5</b>	Response of banana cells transformed with <i>Arath;CyclinD2;1</i> gene: selection and regeneration	52
<b>Fig. 3.6</b>	Response of banana cells transformed with <i>Musac;CyclinD2;1</i> gene: selection and regeneration	53
<b>Fig. 3.7</b>	PCR analysis of greenhouse potted <i>Arath;CyclinD2;1</i> regenerants	56
<b>Fig. 3.8</b>	Southern blot of <i>Arath;CyclinD2;1</i> putative transformed bananas	57
<b>Fig. 3.9</b>	PCR analysis of DNAs from greenhouse-grown <i>Musac;CyclinD2;1</i> transgenic plants	58
<b>Fig. 3.10</b>	Southern blot of banana plants transformed with <i>Musac;CyclinD2;1</i> coding sequence	59

<b>Fig. 4.1</b>	Expression of <i>Musac;CyclinD2;1</i> in different banana plant tissues	67
<b>Fig. 4.2</b>	Truncation of the <i>Arath;CyclinD2;1</i> transgene in transgenic banana	68
<b>Fig. 4.3</b>	Comparison of transcription of <i>Arath;CyclinD2;1</i> transgene in shoot and root apices of banana plants transformed with <i>Arath;CyclinD2;1</i> gene coding sequence	70
<b>Fig. 4.4</b>	Comparison of transcription of indigenous <i>Musac;CyclinD2;1</i> gene in shoots and root apices of banana plants transformed with <i>Arath;CyclinD;1</i> gene coding sequence	71
<b>Fig. 4.5</b>	Comparison of transcription of <i>Musac;CyclinD2;1</i> in shoot and root apices of banana plants transformed with a <i>Musac;CyclinD2;1</i> gene coding sequence	72
<b>Fig. 5.1</b>	Illustration of plant height and leaf growth measurements	80
<b>Fig. 5.2</b>	Leaf length profile and leaf elongation rate of banana plants transformed with <i>Arath;CyclinD2;1</i>	85
<b>Fig. 5.3</b>	<i>In vitro</i> growth of isolated roots of banana plants transformed with <i>Arath;CyclinD2;1</i> gene	92
<b>Fig. 5.4</b>	<i>In vitro</i> growth of isolated roots of banana plants transformed with the <i>Musac;CyclinD2;1</i> gene	92
<b>Fig. 5.5</b>	Visual comparison of root systems of three banana lines over-expressing the <i>Musac;CyclinD2;1</i>	93