

**Enantioselective sensors and biosensors for clinical analysis**

by

**R'afat Mahmoud Nejem**

Submitted in partial fulfilment of the requirements for the degree

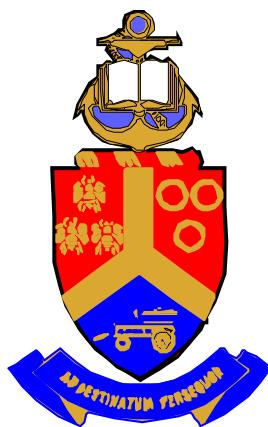
**PHILOSOPHIAE DOCTOR**

in the Faculty of Natural and Agricultural Sciences

University of Pretoria

Pretoria

May 2004



## Dedications

*In deep appreciations, I would like to dedicate this work to: my lovely wife, Grace, for her patience in the durable era of the country state, and for her extensive encouragement and incredible endurance; my children, Aysha, Ameera, Khaldon and Mohamad of being apart of them, promise I will compensate you all the times you missed me; my parents, Mahmoud and Aysha, for their blessings and pray for my success; my brother, Moien, for his full support to my dreams, dreams become true; and my brothers and sisters for continuous support.*

### Acknowledgments

I would like to acknowledge and extend my sincere gratitude to many people for their contribution and assistance to make this work comes to light. Dr. Raluca Stefan thank you very much for your valuable time, your experience, guidance and full support gave to me in this thesis.

I am thankful to Prof. J. F. van Staden for his encouragement, cooperation and support.

To Prof H. Y. Aboul-Enein, deep appreciation is the right words for your assistance.

My colleagues, Dr. Ph. Fletcher, Dr. A. Rat'ko, Dr. K. Ozoemena, Dr. N. Beyene, L. Popovic, V. Mulaudzi, S. Bairu and R. Bokretzion, thank you for being a good company in my hard times.

NRF sponsor and UP bursary my dreams become real by your generous financial support.

I would like to express my deep appreciation to the Ministry of Higher Education (Palestine), Palestinian ministry of higher education, Al-Aqsa University, The Palestinian employee council for giving me the opportunity to come to South Africa and verify my dreams.

Special thanks and appreciations for Prof. Dr. Y. Abu Dayyah, Dr. Abdul-Aziz Abo Share'ah, Dr. Abdoul-Jalil Sarsour, Dr. M. Issa, Dr. N. Al-Abadllah, Dr. A. Shaat, Dr. A.

Al-Qutshan, I. Al-Astal T. Dahman, A. Khalaf, K.Abo Shab for their continuous encouragement and help.

I would like to acknowledge my closest friends who encouraged me a lot in my farness, Mr. Riyad Siedm, Nusirat Social Rehabilitation Center and Mr. Ayman Al-Sisi.

Special thanks, for those who stand beside me with their real care and help, my wife, children for scarifying the time I spent away from them, my parents, brothers and sisters thank you for your encouragement.

And finally above all I thank ALLAH almighty for giving me the power, the peace and the faith to achieve this work.

**Enantioselective sensors and biosensors for clinical analysis**

**by**

**R'afat Mahmoud Nejem**

**Supervisor: Dr Raluca-Ioana Stefan**

**Department of Chemistry**

**University of Pretoria**

**Degree: Philosophiae Doctor**

**SYNOPSIS**

The enantioanalysis of compounds of biological importance with a chiral moiety is very important because each enantiomer is a marker for a different disease. Accordingly, very reliable methods of enantioanalysis should be employed for the correct diagnosis of the diseases. The utilization of amperometric biosensors and enantioselective, potentiometric membrane electrodes made the assay of a single enantiomer faster, easier and more reliable if one compare with the chromatographic techniques which are widely proposed for this kind of analyses.

Monocrystalline diamond was proposed as matrix for amperometric electrodes and amperometric biosensors design. The advantages of using such material for electrode design are: (a) lower background currents and noise signals, which lead to improve S/B and S/N ratios, and lower detection

limits; (b) good electrochemical activity (pre-treatment is not necessary); (c) wide electrochemical potential window in aqueous media; (d) very low capacitance; (e) extreme electrochemical stability; and (f) high reproducibility of analytical information.

The design selected for the electrodes is simple, fast and reproducible. The carbon or diamond powders were mixed with paraffine oil to give the carbon or diamond paste which can act alone as electroactive material in the electrodes or it can be modified with a chiral selector (e.g., cyclodextrins, maltodextrins or antibiotics) or enzyme (e.g., L(D)-aminoacid oxidase, L-lysine oxidase). The results obtained by employing the amperometric electrodes and biosensors and the enantioselective, potentiometric membrane electrodes proved a high sensitivity, selectivity, accuracy and high reliability. These characteristics made them suitable to be used for the enantioanalysis of different compounds of biological importance (e.g., pipecolic acid, glyceric acid, 2-hydroxyglyceric acid, fucose, L-vesamicol and L-lysine) in serum and/or urine samples.

The features of the proposed enantioselective, amperometric and potentiometric electrodes proposed in this thesis are their utilization for *in vivo* measurements and as detectors in flow systems (flow injection analysis or/and sequential injection analysis). This will simplify the enantioanalysis and will improve considerable the reliability of the analytical information favorizing a fast and accurate diagnosis of the diseases associated with the marker determined.

**Enantioselective sensors en biosensors vir kliniese analyse  
deur**

**R'afat Mahmoud Nejem**

**Studieleier: Dr Raluca-Ioana Stefan**

**Department Chemie**

**Universiteit van Pretoria**

**Graad: Philosophiae Doctor**

**SAMEVATTING**

Die enantioanalise van verbindings van biologiese belang met 'n chirale deel is baie belangrik omdat elke enantiomeer 'n merker vir verskillende siektes is. Gevollik moet uiters betroubare enantioanalitiese metodes gebruik word vir die korrekte diagnose van die siektes. Die aanwending van amperometriese biosensors en enantioselektiewe, potensiometriese membraanelektrodes maak die essai van 'n enkele enantiomeer vinniger, makliker en meer betroubaar as dit met chromatografiese tegnieke, wat algemeen vir die tipe analises voorgestel word, vergelyk word.

Monokristallyne diamant is as matrys vir die ontwerp van amperometriese elektrodes en amperometriese biosensors voorgestel. Die voordele om sulke materiaal vir elektrode-ontwerp te gebruik, is: (a) laer agtergrondstroom en geraas seine, wat lei tot verbeterde S/B en S/N

verhoudings, en laer deteksielimiete; (b) goeie elektrochemiese aktiwiteite (voorafbehandeling is nie nodig nie); (c) 'n wye elektrochemiese potensiaalvenster in waterige media; (d) baie lae kapasitansie; (e) ekstreem elektrochemiese stabiliteit; en (f) hoë reproducerebaarheid van analitiese inligting.

Die ontwerp van die elektrodes is eenvoudig, vinnig en reproducerebaar. Die koolstof - of diamantpoeiers word met paraffienolie gemeng om 'n koolstof of diamantpasta te gee wat alleen as elektroaktiewe materiaal in die elektrodes kan optree of gemodifiseer kan word met 'n chiraal selekteerde (byvoorbeeld, siklodekstriene, maltodekstriene of antibiotika) of ensiem (byvoorbeeld, L(D)-aminosuur oksidase, L-lisien oksidase). Die resultate wat met die amperometriese elektrodes en biosensors en die enantioselektiewe, potensiometriese membraanelektrodes verkry word toon 'n hoë sensitiwiteit, selektiwiteit, akkuraatheid en betroubaarheid. Hierdie kenmerke maak hulle uiters gesik in die gebruik van die enantioanalise van verskillende verbinding wat biologies belangrik is (byvoorbeeld, pipekoliensuur, gliseriensuur, 2-hidroksigliseriensuur, fukose, L-vesamikol en L-lisien) in serum en/of urinemonsters.

Die uitstaande kenmerke van die voorgestelde enantioselektiewe, amperometriese en potensiometriese elektrodes wat in hierdie tesis voorgestel word, is hulle toepassing vir *in vivo* metings en as detektore in vloeisisteme (vloei-inspuitanalise of/en sekvensiele inspuitanalise). Dit sal enantioanalise vereenvoudig en die betroubaarheid van analitiese inligting heelwat verbeter wat vinnige en akkurate diagnose van siektes wat met die merker bepaal word, bevoordeel.

## Table of contents

<b>Synopsis</b>	<b>i</b>
<b>Samevatting</b>	<b>iii</b>
<b>Dedications</b>	<b>v</b>
<b>Acknowledgements</b>	<b>vi</b>
<b>Table of contents</b>	<b>viii</b>
<b>Introduction</b>	<b>1</b>
<b>Chapter 1 Chirality in clinical analysis</b>	<b>4</b>
1.1    Introduction	4
1.2    Chirality and configuration	6
1.3    Descriptors of chiral molecules (Nomenclature)	9
1.3.1    The L and D designations	9
1.3.2    The Cahn-Ingold-Prelog designations (S and R designations)	11
1.3.3    (-) and (+) designations ( <i>l</i> or <i>d</i> )	13
1.3.4    Helicity ( <i>M</i> or <i>P</i> )	14
1.4    Enantiomeric purity	15
1.5    Sources of chiral compounds	15
1.6    Importance of chiral molecules	16

1.6.1	Chirality and clinical diagnosis	17
1.6.2	Importance of chirality for pharmaceutical compounds	19
1.7	Method of chiral recognition	24
1.7.1	Polarimetry	24
1.7.2	Chromatographic methods	24
1.7.3	Capillary electrophoresis	27
1.7.4	Nuclear magnetic resonance spectroscopy	28
1.7.5	Circular dichroism	29
1.7.6	Ferroelectric liquid crystals	29
1.8	Molecular recognition of enantiomers using electrochemical electrodes	31
1.8.1	Molecular recognition of enantiomers using enantioselective, potentiometric membrane electrodes (EPMEs)	31
1.8.2	Molecular recognition of enantiomers using amperometric biosensors	32
1.8.3	Molecular recognition of enantiomers using amperometric immunosensors	32
1.9	Electrodes as detectors in flow or sequential injection analysis (FIA or SIA)	33
1.10	References	38
<b>Chapter 2 Enantioselective, potentiometric membrane electrodes</b>		<b>45</b>
2.1	Introduction	45
2.2	Design of enantioselective, potentiometric membrane electrodes	47
2.2.1	Modified paste electrode design	48

2.2.1.1 Cyclodextrins as chiral selectors in the EPMEs design	48
2.2.1.2 Maltodextrins as chiral selectors in the EPMEs design	50
2.2.1.3 Macrocyclic antibiotics as chiral selectors in the EPMEs design	51
2.2.2 Plastic membrane based electrode design	53
2.3 Response characteristics of EPME	54
2.3.1 Standard electrode potentials, $E^{\circ}$	55
2.3.2 Response of EPME	56
2.3.3 Limit of detection	57
2.3.4 Linear concentration range	58
2.3.5 Influence of pH	58
2.3.6 Influence of the temperature on the response of the electrode	59
2.3.7 Response time	59
2.3.8 Ionic strength and activity coefficients	60
2.4 Selectivity of enantioselective potentiometric membrane electrodes	60
2.4.1 Mixed solution method	62
2.4.2 Separate solution method	62
2.5 Direct potentiometric method	63
2.6 References	66
<b>Chapter 3 Amperometric electrodes for enantioanalysis</b>	<b>70</b>
3.1 Introduction	70
3.2 Design of amperometric electrodes	71

3.2.1	Design of carbon paste based amperometric electrodes	71
3.2.2	Design of diamond based amperometric electrodes	72
3.3	Response characteristics of the amperometric electrodes	74
3.3.1	Slope (Response) of the electrode	76
3.3.2	Limit of detection	77
3.3.3	Linear concentration range	78
3.3.4	pH range	78
3.3.5	Ionic strength and activity coefficients	79
3.3.6	Response time	79
3.3.7	Influence of the temperature on the response of the electrodes	80
3.4	Selectivity of the amperometric electrodes	80
3.4.1	Mixed solution method	81
3.4.2	Separate solution method	82
3.5	Direct amperometric method	83
3.6	Differential pulse voltammetry	83
3.6.1	Potential pulse amplitude	88
3.6.2	The scan rate	89
3.6.3	Peak area and peak height	89
3.7	References	90
<b>Chapter 4 Amperometric biosensors for enantioanalysis</b>		<b>93</b>
4.1	Introduction	93
4.2	Design of amperometric biosensors	94

4.2.1	Physical immobilization	97
4.2.1.1	Biosensors based on plastic membranes	98
4.2.1.2	Biosensors based on carbon and diamond paste	98
4.2.2	Chemical immobilization	100
4.2.2.1	Plastic based amperometric biosensors	100
4.2.2.2	Carbon paste based amperometric biosensors	101
4.3	Response characteristics of amperometric biosensors	102
4.3.1	pH range	102
4.3.2	Life time ( $t_L$ )	102
4.3.1	Michaelis-Menten constant ( $K_M$ )	103
4.4	Selectivity of the amperometric biosensors and immunosensors	104
4.5	Chronoamperometry	104
4.6	Direct amperometry	106
4.7	References	107
<b>Chapter 5</b>	<b>Enantioanalysis of L- and D-pipecolic acid in biological samples</b>	<b>109</b>
5.1	Introduction	109
5.2	Reagents and chemicals	112
5.3	Amperometric electrode for enantioselective analysis of pipecolic acid	113
5.3.1	Apparatus	113
5.3.2	Electrode design	113
5.3.3	Recommended procedure: Direct DPV assay	113
5.3.4	Results and discussion	114

5.3.4.1	Electrode response	114
5.3.4.2	Selectivity of the diamond paste based electrode	115
5.3.4.3	Analytical applications	116
5.4	Enantioselective, potentiometric membrane electrodes for the determination of L-pipecolic acid in serum	118
5.4.1	Apparatus	118
5.4.2	Electrode design	118
5.4.3	Recommended procedure: Direct potentiometry	119
5.4.4	Results and discussion	119
5.4.4.1	Response characteristics of the EPMEs	119
5.4.4.2	The influence of pH on the responses of the electrodes	121
5.4.4.3	Selectivity of the EPMEs	121
5.4.4.4	Analytical applications	122
5.5	Amperometric biosensors for the enantioselective analysis of L- and D- pipecolic acids in biological fluids	124
5.5.1	New amperometric biosensors based immobilization of L- and D-amino acid oxidases on diamond paste for the determination of L- and D-pipecolic acids in serum samples	124
5.5.1.1	Apparatus	124
5.5.1.2	Amperometric biosensors design	124
5.5.1.3	Recommended procedure: Direct DPV assay	125
5.5.1.4	Preparation of the modified diamond paste	125
5.5.1.5	Results and discussion	125

5.5.1.5.1 Amperometric biosensors response	125
5.5.1.5.2 Selectivity of the Amperometric biosensors	126
5.5.1.5.3 Analytical applications	127
5.5.2 Carbon paste based amperometric biosensors for the enantioselective analysis of pipecolic acid	129
5.5.2.1 Apparatus	129
5.5.2.2 Amperometric biosensors design	129
5.5.2.2.1 Monoenzyme amperometric biosensors	130
5.5.2.2.2 Bienzyme amperometric biosensors	130
5.5.2.3 Recommended procedure: Direct amperometry	130
5.5.2.4 Determination of L- and D-pipecolic acids in serum samples	130
5.5.2.5 Results and discussion	131
5.5.2.5.1 Amperometric biosensors response	131
5.5.2.5.2 Enantioselectivity of the amperometric biosensors	132
5.5.2.5.3 Analytical applications	133
5.5.3 Diamond paste based amperometric biosensors based on L-AAOD and D-AAOD for the determination of L- and D-pipecolic acids	135
5.5.3.1 Apparatus	135
5.5.3.2 Amperometric biosensors design	135
5.5.3.3 Recommended procedure: Direct amperometry	136

5.5.3.4	Determination of L- and D-pipecolic acids in serum samples	136
5.5.3.5	Results and discussion	136
5.5.3.5.1	Response characteristics of amperometric biosensors	136
5.5.3.5.2	Enantioselectivity of the amperometric biosensors	137
5.5.3.5.3	Analytical applications	138
5.6	Sequential injection analysis utilizing amperometric biosensors as detectors for the simultaneous determination of L- and D-pipecolic acids	139
5.6.1	Apparatus	139
5.6.2	Biosensors's design	140
5.6.3	Sequential injection system	140
5.6.4	Results and discussion	142
5.6.4.1	The response characteristics of the biosensors in the SIA system	142
5.6.4.2	Selectivity of the amperometric biosensors	144
5.6.4.3	Analytical applications	145
5.7	Conclusion	147
5.8	References	149

<b>Chapter 6 Diamond paste-based electrodes for the determination of L- and D-fucose using differential pulse voltammetry</b>	<b>152</b>
6.1      Introduction	152
6.2      Experimental section	155
6.2.1    Apparatus	155
6.2.2    Diamond paste electrode design	155
6.2.3    Recommended procedures	155
6.2.3.1    Cyclic voltammetry	155
6.2.3.2    Direct differential pulse voltammetry	156
6.2.4    Reagents and materials	156
6.2.5    L-fucose samples	157
6.3      Results and discussion	157
6.3.1    Optimization of working conditions	157
6.3.2    Response of diamond paste based electrodes	161
6.3.3    Selectivity of the diamond paste electrodes	162
6.3.4    Analytical applications	163
6.3.5    Statistical comparison between diamond paste electrodes and the standard method for fucose analysis	165
6.4      Conclusion	166
6.5      References	167

<b>Chapter 7 Enantioselective, potentiometric membrane electrodes for the determination of L- and D-glyceric acids</b>	<b>170</b>
7.1 Introduction	170
7.2 Reagents and materials	173
7.3 Enantioselective, potentiometric membrane electrode based on maltodextrins	174
7.3.1 Apparatus	174
7.3.2 Electrodes design	174
7.3.3 Recommended procedure	175
7.3.4 Results and discussion	175
7.3.4.1 EPMEs response characteristics	175
7.3.4.2 Effect of pH on the responses of the electrodes	177
7.3.4.3 Selectivity of the electrodes	178
7.3.4.4 Analytical applications	179
7.4 Enantioselective, potentiometric membrane electrode based on cyclodextrins	181
7.4.1 Apparatus	181
7.4.2 Electrodes design	181
7.4.3 Recommended procedure	182
7.4.4 Results and discussion	182
7.4.4.1 EPMEs response characteristics	182
7.4.4.2 Effect of pH on the responses of the electrodes	184
7.4.4.3 Selectivity of the electrodes	184
7.4.4.4 Analytical applications	186

7.5	Enantioselective, potentiometric membrane electrode based on macrocyclic antibiotics	188
7.5.1	Apparatus	188
7.5.2	Electrodes design	189
7.5.3	Recommended procedure	189
7.5.4	Results and discussion	190
7.5.4.1	Response characteristics of EPMEs	190
7.5.4.2	The influence of pH on the responses of the EPMEs	191
7.5.4.3	Selectivity of the EPMEs	192
7.5.4.4	Analytical applications	192
7.6	Conclusion	194
7.7	References	196
<b>Chapter 8</b>	<b>Diagnosis of L- and D-2-hydroxyglutaric acidurias using enantioselective, potentiometric membrane electrodes</b>	<b>199</b>

8.1	Introduction	199
8.2	Reagents and materials	202
8.3	Enantioselective, potentiometric membrane electrode based on maltodextrins	203
8.3.1	Equipments and apparatus	203
8.3.2	Electrodes design	204
8.3.3	Recommended procedure	204
8.3.4	Results and discussion	205
8.3.4.1	EPMEs response	205

8.3.4.2	The pH influence on the responses of the EPMEs	206
8.3.4.3	Selectivity of the electrodes	207
8.3.4.4	Analytical applications	207
8.4	Enantioselective, potentiometric membrane electrode based on cyclodextrins for the determination of L- and D-2-hydroxyglutaric acid in urine samples	209
8.4.1	Equipments and apparatus	209
8.4.2	Electrodes design	209
8.4.3	Recommended procedure	210
8.4.4	Results and discussion	210
8.4.4.1	EPMEs response	210
8.4.4.2	The pH influence on the responses of the EPMEs	212
8.4.4.3	Selectivity of the electrodes	213
8.4.4.4	Analytical applications	213
8.5	Determination of D-2-hydroxyglutaric acid in urine ample using enantioselective, potentiometric membrane electrodes based on antibiotics	216
8.5.1	Apparatus	216
8.5.2	Electrodes design	216
8.5.3	Recommended procedure	217
8.5.4	Results and discussion	217
8.5.4.1	Response characteristics of EPMEs	217
8.5.4.2	The influence of pH on the responses of the EPMEs	218
8.5.4.3	Selectivity of the EPMEs	219
8.5.4.4	Analytical applications	220

8.6	Conclusion	222
8.7	References	224

**Chapter 9 Enantioanalysis of L-vesamicol in serum sample using  
enantioselective, potentiometric membrane electrodes** **227**

9.1	Introduction	227
9.2	Reagents and materials	229
9.3	Enantioselective, potentiometric membrane electrodes based on maltodextrins	230
9.3.1	Equipments and apparatus	230
9.3.2	Electrodes design	230
9.3.3	Recommended procedure	231
9.3.4	Results and discussion	231
9.3.4.1	EPMEs response characteristics	231
9.3.4.2	Effect of pH on the responses of the electrodes	232
9.3.4.3	Selectivity of the electrodes	233
9.3.4.4	Analytical applications	234
9.4	Cyclodextrins based enantioselective, potentiometric membrane electrodes	235
9.4.1	Apparatus	235
9.4.2	Electrodes design	235
9.4.3	Recommended procedure	236
9.4.4	Results and discussion	236
9.4.4.1	EPMEs response characteristics	236

9.4.4.2	Influence of pH on the responses of the electrodes	237
9.4.4.3	Selectivity of the electrodes	238
9.4.4.4	Analytical applications	239
9.5	Enantioselective, potentiometric membrane electrode based on macrocyclic antibiotics	240
9.5.1	Apparatus	240
9.5.2	EPMEs design	240
9.5.3	Recommended procedure	241
9.5.4	Results and discussion	241
9.5.4.1	The response characteristics of EPMEs	241
9.5.4.2	Effect of pH on the response of the EPMEs	243
9.5.4.3	Selectivity of the electrodes	243
9.5.4.4	Analytical applications	244
9.6	Conclusion	245
9.7	References	246
<b>Chapter 10 Amperometric biosensor for the enantioanalysis of L-lysine in serum samples</b>		<b>248</b>

10.1	Introduction	248
10.2	Reagents and materials	250
10.3	Diamond paste based amperometric biosensor	251
10.4	Apparatus	251
10.5	Recommended procedures	252

10.6	Determination of L-lysine in serum samples	252
10.7	Results and discussion	252
10.7.1	Response characteristics of the amperometric biosensors	252
10.7.2	Enantioselectivity of the amperometric biosensor	253
10.7.3	Analytical applications	254
10.8	Conclusion	255
10.9	References	256
<b>Chapter 11 Conclusions</b>		<b>259</b>
<b>Appendix</b>		<b>263</b>
<b>Appendix A Publications</b>		<b>264</b>
<b>Appendix B Presentations</b>		<b>268</b>

## Introduction

The availability of continuous monitoring of metabolic substances is very important in the intensive care units and can be a great aid to patients care. Chirality has been reported as an important issue for different compounds of biological importance. The presence of chiral compounds in human fluids (serum, urine, spinal fluids) as normal metabolites of human metabolism or drug metabolism give the vitality for monitoring levels of these molecules in biofluids. The existence of higher or lower levels of these specific molecules is a marker indicator of human body abnormalities. The normal concentration level changes of molecules in humans mostly referred to the deficiency of some enzymes. Amino acids, carbohydrates and urinary organic acids are excreted in human and their changes can cause different types of diseases such as inborn metabolic disorders and cancer.

Many diseases are caused by substances existing in enantiomeric form and each enantiomer causes a different disease, so to diagnose these illnesses it is very important to find an analytical method that can discriminate between the L- and D-enantiomers. These methods will be very helpful and should exhibit reliable analytical information, fast analysis and could be applied for the continuous monitoring of the enantiomers in biological fluids.

Molecular recognition plays the main role in chiral discrimination around an asymmetric center. Due to the importance of enantiomeric discrimination, there has been substantial need for the discovery of substances used in the enantioanalysis for the diagnosis,

prevention, and treatment of human diseases. These developments have resulted in increased demand for sensitive and specific analytical methods.

The instrumental methods for quantitation which are most commonly used in clinical enantioanalysis are structural analysis, chromatographic and electrochemical methods. Electrochemical sensors are a very good alternative for structural analysis because of their high reliability that is given by high precision, high reproducibility and rapidity. The precision obtained using electrochemical sensors is higher than that obtained using chromatographic methods due to the fact that electrochemical sensors can be used directly for measurements of the compounds in solution.

The aim of this thesis is to construct reliable enantioselective electrodes (amperometric electrodes, amperometric biosensors, enantioselective, potentiometric membrane electrodes) to be applied in diagnosis. Carbon and diamond pastes are proposed as matrices for the sensors' design. Chiral recognition principles based on selected binding as well as on catalyst selectivity must be considered for the selection of the best chiral selector or enzyme.

Differential pulse voltammetry, chronoamperometry and potentiometry can be used for the direct assay of enantiomers in the serum and/or urine samples. An analysis of the performances of the electrodes has shown that the selection of the type of the electrode and matrix of its membrane should be done in accordance with the complexity of the structure of the enantiomer to be determined. Also, the analytical information obtained in

the enantioanalysis using electrochemical sensors is more reliable than that obtained using conventional or chromatographic methods.