

# Expression of a $\beta$ -1,3-glucanase from a biocontrol fungus in transgenic pearl millet

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## ABSTRACT

*Sclerospora graminicola* is an Oomycete (heterotrophic Stramenopiles), fungal-like obligate phytopathogen, the causal agent of downy mildew in pearl millet (*Pennisetum glaucum* [L.] R. Br.), and a major constraint in the production of this cereal crop. In this study a hydrolytic enzyme,  $\beta$ -1,3-glucanase (*gluc78*), from the biocontrol fungus *Trichoderma atroviride*, was introduced into the genome of a pearl millet breeding line, 842B, by particle bombardment. Constructs were prepared containing the *gluc78* gene, encoding the 78 kDa  $\beta$ -1,3-glucanase protein, downstream of either the constitutive ubiquitin promoter or the wound inducible potato proteinase inhibitor IIK gene promoter (*pin2*). The positive selectable marker gene, *manA*, encoding mannose-6-phosphate isomerase (phosphomannose isomerase) under the control of the ubiquitin promoter, was used for co-transformation. Transgenic plants were obtained harbouring the *manA* selectable marker gene and the antifungal gene *gluc78* downstream of either the ubiquitin or *pin2* promoter. Full constructs or minimal transgene expression cassettes containing the genes of interest were successfully introduced into the genome of pearl millet. Progeny of stably transformed plants, harbouring the *gluc78* transgene which is driven by the *pin2* promoter and followed by the rice *Act1* intron

sequences, were subjected to pathogenicity trials. One transgenic event exhibited a reduction of 58% in the incidence of *S. graminicola* infection, however other transgenic pearl millet events showed no resistance to this phytopathogen. The event conferring decreased susceptibility to *S. graminicola* had high levels of the glucanase transcript especially in transgenic plants showing higher levels of downy mildew infection.

**Keywords:** Fungal resistance,  $\beta$ -1,3-glucanase, *Sclerospora graminicola*, transgenic pearl millet.

## 1. INTRODUCTION

Pearl millet (*Pennisetum glaucum* formerly *Pennisetum americanum*) is a staple food for the world's poorest and most food-insecure people in Africa and India. Pearl millet is the only major staple cereal that reliably produces both grain and forage on poor, sandy soils under hot, dry conditions of Africa and Asia (Goldman *et al.*, 2003). The worst scourge of pearl millet is the downy mildew disease caused by the fungal phytopathogen *Sclerospora graminicola* (Sacc.) J. Schröt. It causes systemic infection in pearl millet that manifests itself as foliar symptoms and as green ear disease in the panicle (Sarosh *et al.*, 2005).

*Trichoderma atroviride* (Sacc.) J. Schröt. (previously *T. harzianum*) is a soil-borne filamentous fungus, capable of parasitising several plant pathogenic fungi, and is a well characterised biocontrol agent (Ait-Lahsen *et al.*, 2001; Emani *et al.*, 2003). *T. atroviride* is therefore a potential source of powerful resistance genes against fungal induced plant diseases. Secretion of lytic enzymes, mainly glucanases and chitinases, is considered the most crucial step of the mycoparasitic process (Cohen-Kupiec *et al.*, 1999). The lytic enzymes degrade the cell walls of the pathogenic fungi, enabling *Trichoderma* to utilize both their cell walls and cellular contents for nutritional purposes.

It is well documented that acidic and basic plant  $\beta$ -1,3-glucanases are associated with fungal infection (Gheong *et al.* 2000; Ji *et al.* 2000; Jung and Hwang, 2000). Induction of a  $\beta$ -1,3-

glucanase occurs in seedlings of pearl millet in response to infection by *S. graminicola* and high levels of a 30 kDa  $\beta$ -1,3-glucanase are present in downy mildew resistant pearl millet cultivars but absent in susceptible cultivars (Kini *et al.*, 2000a,b). These studies indicate that  $\beta$ -1,3-glucanases play an important role in the defence mechanism of pearl millet plants against *S. graminicola* infection. In addition, transgenic potato plants expressing a soybean  $\beta$ -1,3-endoglucanase gene exhibited increased resistance to the oomycete *Phytophthora infestans* (Mont.) de Bary (Borkowska *et al.*, 1998).

Broad spectrum resistance against soil-borne and foliar pathogens was achieved in potato plants expressing an antifungal endochitinase from *T. atroviride* (*chit42*) (Lorito *et al.*, 1998). The high level and broad spectrum of resistance obtained against these pathogens with a single constitutively expressed endochitinase gene, *chit42*, improved the previously limited efficacy of transgenic expression of chitinase genes from plants and bacteria in plants (Lorito *et al.*, 1998). In addition, a 78 kDa  $\beta$ -1,3-glucanase (EC 3.2.1.58) from *T. atroviride* exhibited activity ( $ED_{50}$  of 0.06-0.26  $\mu\text{g ml}^{-1}$ ) against *Phytophthora spp.* in an antifungal bioassay when applied in combination with microbial toxins and chemical fungicides (Fogliano *et al.*, 2002). The aim of the presented study was to introduce *gluc78*, a 78 kDa  $\beta$ -1,3-glucanase enzyme from *T. atroviride* into the genome of pearl millet to improve resistance to *S. graminicola*. Since *gluc78* had no sequence homology to plant or bacterium glucanases, it was hypothesised that stable integration and expression of this single glucanase gene in pearl millet could significantly contribute to increased resistance to downy mildew.

*Agrobacterium*-mediated transformation of crops usually generates transgenic plants carrying a single-copy transgene, flanked by the left and the right T-DNA border sequences. Nevertheless, a number of reports have shown the presence of multiple transgene copies and vector sequences in up to 75% of transformants (Kononov *et al.*, 1997; Lange *et al.*, 2006). Therefore, the approach of introducing minimal transgene expression cassettes (MTECs) by biolistic transformation with the goal of enhancing low copy number integration of transgenes, and minimising transgene rearrangements and gene silencing (Fu *et al.*,

2000; Kohli *et al.*, 1998), was employed. Vector backbones have the tendency to promote transgene rearrangements, since the vector backbone has been shown to provide a number of recombination hotspots. The removal of these sequences would possibly limit the influence of recombinogenic elements on the process of integration (Fu *et al.*, 2000).

In this study, scutellum cells of pearl millet immature zygotic embryos (IZEs) were co-transformed with the positive selectable marker gene, *manA* and the *gluc78* transgene either under the control of the *pin2* or ubiquitin promoter. Seedlings produced from selfed seed of transgenic T<sub>2</sub> pearl millet of selected transformation events were tested for resistance to *S. graminicola*.

## 2. MATERIALS AND METHODS

### 2.1 Plasmid construction

A gene (*gluc78*) encoding an antifungal glucan 1,3- $\beta$ -glucosidase was cloned from strain P1 of the biocontrol fungus *Trichoderma atroviride* (Donzelli *et al.*, 2001). The gene encodes for a protein of 770 aa, including a 40 aa signal peptide. The *gluc78/nos* terminator fragment was ligated downstream of the constitutive ubiquitin promoter within pAHC25 (Christensen and Quail, 1996) and the product was designated pUBI*gluc78* (10 131 bp) (Fig. 1). A construct was also prepared containing the *gluc78* gene driven by the *pin2* promoter followed by an AMV leader sequence (pGEM*gluc78*; 6 584bp). Construct pPA*gluc78* (7 148bp) was prepared with the *pin2* promoter, and the rice *Act1* intron sequence upstream of the *gluc78* gene (Fig. 1). The ubiquitin promoter was inserted upstream of the *manA* gene in the multiple cloning region of the promoterless construct pNOV3604 (Syngenta Seeds AG). The resulting 6210 bp construct was designated pNOV3604ubi (Fig. 1).

Fig 1

In order to assess the sequence integrity of expression vectors, plasmid DNA was digested with various restriction enzymes both individually and in combinations to verify that the expected fragment sizes were obtained for the four prepared constructs. In addition, sequence integrity of the vectors and inserts across ligation regions was verified by

sequence analysis (ABI PRISM™ dye terminator cycle sequencing) using the ready reaction kit with AmpliTaq® DNA polymerase (The Perkin Elmer Corporation, Norwalk, USA).

## 2.2 Pearl millet plant transformation

Medium J was used for all pearl millet transformations as previously described (O’Kennedy *et al.*, 2004a). MTECs were isolated as follows: *Hind* III isolated fragment of 4613 bp from pUBI*gluc78*, *Hind* III/*Sph* I isolated fragments of 3845bp or 4109bp from pGEM*gluc78* or pPA*gluc78*, respectively and a *Hind* III/*Asp* 718 fragment of 3559 bp from pNOV3604ubi. The fragments were gel purified with a QIAquick (QIAGEN, Hilden, Germany) gel extraction kit or GeneClean II kit (Q BIOgene, USA) under sterile conditions. Proliferating IZEs were co-bombarded with pNOV3604ubi and one of the constructs containing the *gluc78* gene (Table 1) at a ratio (molar) of 1:2 respectively. IZEs were pre-cultured for 6-8 days, bombarded at a helium pressure of 900-1200 kPa and then cultured for 4-6 days on osmoticum free medium before being transferred to mannose-containing selection medium, of 2 g l<sup>-1</sup> maltose and 15 g l<sup>-1</sup> mannose.

## 2.3 PCR analysis of putative transgenic plants

The *manA* specific primers (PMI fwd: 5’-CGT TGA CTG AAC TTT ATG GTA TGG-3’ and PMI as: CAC TCT GCT GGC TAA TGG TG-3’) were used to amplify a 965 Kb mannose gene fragment, from genomic DNA preparations of putative transgenic pearl millet plantlets. The *gluc78* specific primers (*gluc78* fwd: 5’-ATG ATG GGT CTC TCA ACC GTC-3’ and *gluc78* as: 5’-TAG CAC CGC CAT TGA GAA TGG-3’) were used to amplify a 1100 Kb fragment, from genomic DNA preparations. The *bla* gene (conferring ampicillin resistance) present in the vector backbones was amplified with the following primers: *bla* fw 5’-TGCTTAATCAGTGAGGCACC-3’ and *bla* as1 5’-AGATCAGTTGGGTGCACG-3’ to produce a PCR product of 757 bp.

## 2.4 Southern blot analysis of transgenic plants

Genomic DNA was extracted from putative transgenic pearl millet leaf material using the mini extraction procedure of Dellaporta *et al.* (1983). Five micrograms of pearl millet genomic DNA were digested with selected restriction enzymes and the fragments separated on an agarose gel and analysed by Southern blot analysis as previously described (O’Kennedy *et al.* 2004b). Internal fragments of the *manA* gene of pNOV3604ubi and the *gluc78* gene of pPA*gluc78* were labelled with digoxigenin (DIG) by the PCR DIG probe synthesis kit as described by the supplier (Roche Diagnostics GmbH, Mannheim, Germany). The DIG labelled molecular weight marker III from was included on all blots in order to estimate fragment sizes. An internal DNA probe of *gluc78* was also prepared using redivue™ <sup>32</sup>P-dCTP labelling and the mega prime™ labelling kit (RPN1605) (Amersham Biosciences, UK) and southern blot analysis (DNA target) conducted as described for northern blot analysis.

Infection with *Sclerospora graminicola*.

## 2.5 Northern and Western blot analysis of transgenic pearl millet plants

In order to determine expression levels of the *gluc78* transgene, RNA and protein extractions, of selected non-transgenic control plants and transgenic pearl millet inoculated with *S. graminicola*, were prepared 21 days after inoculation and probed with a labelled DNA fragment or antibody, respectively. Wound elicited expression of *gluc78* was assessed in similar age transgenic and non-transgenic control plants by wounding with forceps at 1 cm intervals along a selected leaf. Leaves were harvested 24 hours after wounding.

Total RNA was extracted using the QIAGEN RNeasy plant mini kit (QIAGEN) according to the manufacturer’s recommendation. The RNA was separated on a denaturing formaldehyde gel as described in the booklet of the QIAGEN RNeasy plant mini kit. Eight microgram of RNA was loaded per lane. The RNA was transferred overnight via capillary action to a positively charged nylon membrane (Roche Diagnostics GmbH) using 10 X SSC (Sambrook *et al.*, 1989) and then fixed to the dried membrane for 30 min at 80°C followed by exposure to UV light for three minutes.

An internal DNA probe of *gluc78* was prepared using redivue™ <sup>32</sup>P-dCTP labelling and the mega prime™ labelling kit (RPN1605) (Amersham Biosciences, UK). Unincorporated nucleotides were removed with a QIAquick® Nucleotide Removal kit (QIAGEN) and the probe was denatured by boiling before addition to the blots. Blots were treated for 4h in prehybridisation solution (100 mg l<sup>-1</sup> denatured Salmon sperm DNA, 50 mM Tris pH 8.0, 10 mM EDTA, 5 X SSC, 20 mg each of BSA, Ficoll 400 and PVPP, 0.2% SDS [m/v]) followed by 16h in hybridisation solution (100 mg l<sup>-1</sup> denatured Salmon sperm DNA, 500 g l<sup>-1</sup> [m/v] dextran sulfate, 50 mM Tris pH 8.0, 10 mM EDTA, 5 X SSC, 20 mg l<sup>-1</sup> each of BSA, Ficoll 400 and PVPP, 0.2% SDS [m/v]) at 65°C in a Techne Hybridiser HB-1D (Techne). Post hybridisation washes were performed as follows: 5 min at room temperature (2 X SSC, 0.5% SDS [m/v]), a second more stringent wash for 5 min at room temperature (0.1 X SSC, 0.1% SDS) and 10-15 min at 65°C (0.1 X SSC, 0.1% SDS). Blots were exposed to X-ray film (Hyperfilm™ MP, Amersham Pharmacia Biotech) in the presence of intensifier screens for 3-5 days at -80°C.

Protein extractions were carried out according to Kini and co-workers (2000a) with the addition of PVPP and protease inhibitor cocktail (Sigma P9599) for protection against protease activity. Leaf material was homogenised with a mortar and pestle in extraction buffer (50 mM sodium acetate buffer pH 5.2), containing PVPP and protease inhibitor cocktail, according to the manufacturer's recommendation. The extracts were centrifuged at 4°C for 10 minutes at 12 000 rpm. The supernatant was frozen at -20°C until SDS-PAGE analysis. The protein concentration of each sample was determined by Bio-Rad protein dye (Bio-Rad), reagent, with bovine serum albumin as the standard. For each sample, 2 µg aliquots of protein were separated by polyacrylamide gel electrophoresis (PAGE) on a 12% separating, and 6% stacking gel according to the method of Laemmli (1970). Proteins were transferred to MSI PVDF-plus transfer membranes (0.45 micron) (Amersham Biosciences) by means of a Hoefer TE22 blot system. Western blots were probed with antiserum (1:750) to the glucanase protein using the ECL RPN2108 western blotting kit (Amersham

BioSciences). For biotinylated molecular marker detection (ECL RPN2107, Amersham BioSciences) streptavidin-horseradish peroxidase conjugate (ECL RPN1231, Amersham BioSciences) was used.

## 2.6 Pathogenicity trials of transgenic progeny

Transgenic pearl millet seedlings containing the antifungal gene, *gluc78*, from *T. atroviride* (Donzelli *et al.*, 2001), and selectable marker gene, *manA* (Syngenta Crop Protection AG), were identified by germination on mannose containing medium. The transgenic germinating plantlets were transplanted to pots of peat and sand compost at the glasshouse facility at Pen-y-Ffridd Experimental Station, Bangor, Wales, prior to inoculation with *S. graminicola*. *S. graminicola* isolates from Niger, Senegal and Mali were maintained in the glasshouse on the highly susceptible pearl millet cultivar 7042(S).

All the transgenic plants were derived from the cultivar 842B. The seedlings were planted in low-nutrient compost consisting of 2 parts fine peat to one part sharp sand with added nutrients (Chempak Seed Base, Chempak Products, UK: NPK 25-39-30 mg l<sup>-1</sup>). Pots were placed on flood benching in a controlled environment glasshouse providing a 16-h day length with light intensity between 500 and 1200  $\mu\text{mol Em}^{-2} \text{s}^{-1}$  and a temperature of 25-30°C from 06:00-18:00 and 20°C from 18:00-06:00. The plants were watered by filling the benches to a depth of 1-2 cm for 30 min. Plants were fed twice weekly with a liquid fertilizer (Vitax Vitafeed 214, Leicester, UK). When the seedlings were at the coleoptile to one-leaf stage, inoculum was prepared. Leaves from 2-3 month old infected plants (Niger, Senegal and Mali isolates in a 1:1:1 ratio) were collected mid-morning, after plants had been photosynthesising for at least 4 h, wiped clean of old sporangia using moist laboratory roll (Kimwipes Roll, Kimberly Clark, Kent, UK) and incubated in sealed plastic boxes lined with moist laboratory roll for 7-8h at 20°C in the dark. Freshly produced sporangia were collected by shaking the leaves in cold distilled water (0-2°C). The resulting sporangial suspension was filtered through muslin to remove debris and adjusted to  $2 \times 10^{-5}$  sporangia/ml with cold distilled water. Each pot of seedlings was sprayed with approximately 4 ml of the inoculum using a compressed-air



cylinder-fed sprayer (Kestrel Eqpt. Ltd., London, UK). The inoculum was maintained on ice throughout inoculation to prevent zoospore release and so ensure a uniform sporangial concentration over time (Jones *et al.*, 2001). The pots were covered with a polythene sheet to maintain a high level of humidity and incubated in the glasshouse at 20°C overnight. The development of downy mildew on treated plants were determined 21 days post infection and compared to wild type control plants. All residual plant material was incinerated after assessment. Care was taken to destroy the plant material before flowering occurred.

## 2.7 Reverse transcription quantitative PCR (RT-qPCR) analysis of transgenic plants

Gene specific primers were synthesised for an endogenous pearl millet  $\beta$ -glucanase (AF488414) and *gluc78* (AF253421) from *T. atroviride*. Primers designed to a pearl millet 18S rRNA gene were used as an internal reference control for normalisation (Crampton *et al.* 2009). Optimal primer design was performed using Primer3 software (<http://frodo.wi.mit.edu>), and internal primer secondary structure (hairpins, dimers, palindromes, repeats) was assessed using Net Primer software ([www.PremierBioSoft.com](http://www.PremierBioSoft.com)). Primers used in RT-qPCR were as follows: 18S rRNA (18S fw 5'-GCCATCGCTCTGGATACATT-3' and 18S as 5'-TCATTACTCCGATCCCGAAG-3'), and *T. atroviride* transgene glucanase expression (Gluc78 fw 5'-CTGACCCTGACTTTTCCAAGTG-3' and Gluc78 as 5'-GAAGAAGCTGTACATGCCAGTG-3'), producing PCR products of 83, 101 and 113 bp, respectively.

DNase 1-treated (QIAGEN) total RNA from each of the treated plants was reverse transcribed using a Transcriptor First Strand cDNA Synthesis Kit (Roche Diagnostics). Each 20  $\mu$ l reaction contained 200 ng total RNA, 60  $\mu$ M random hexamer primer, 1 X Transcriptor RT reaction buffer, 20 U Protector RNase Inhibitor, 1 mM dNTP mix, 10 U Transcriptor reverse transcriptase. Reactions were incubated at 55°C for 30 min, after which the Transcriptor reverse transcriptase was inactivated by heating to 85°C for 5 min. Reactions were stored at -20°C until use in real time PCR reactions.

Conditions for all PCR reactions were optimised in a GeneAmp PCR System 2400 thermocycler (Applied Biosystems, Foster City, CA, USA) with regard to forward and reverse primer concentrations and annealing temperature. Optimised results were applied to the LightCycler PCR protocol. Real time PCR reactions were prepared using a LightCycler® FastStart DNA Master<sup>PLUS</sup> SYBR Green 1 kit (Roche Diagnostics). Each 20 µl reaction contained 0.25 µM forward primer, 0.25 µM reverse primer, 1 X LightCycler Master Mix and 2 µl cDNA (undiluted). Reactions were added to glass capillaries and placed into the LightCycler rotor (Roche Diagnostics). The following LightCycler experimental run protocol was used: denaturation cycle (95°C for 10 min), amplification and quantification cycle repeated 45 times (95°C for 10 s, 60°C for 5s, 72°C for 6s with a single fluorescence measurement), melting curve cycle (65-95°C with a heating rate of 0.1°C per second and continuous fluorescence measurement), and finally a cooling step to 40°C. LightCycler software 3.3 (Roche Diagnostics) was employed to calculate crossing points ( $C_T$ ) for each transcript.  $C_T$  is defined as the point at which the fluorescence rises appreciably above the background fluorescence (Pfaffl, 2001).

### 3. RESULTS

#### 3.1 Production of transgenic pearl millet

The *gluc78* gene, encoding  $\beta$ -1,3-glucanase from *T. atroviride* (*gluc78*), in constructs pGEM*gluc78* and pPA*gluc78* was driven by the potato proteinase inhibitor IIK wound inducible promoter, (Fig. 1) to elicit expression during wounding and/or pathogen attack. In addition, the ubiquitin constitutive promoter, was ligated upstream of the selectable marker gene, *manA* (pNOV3604ubi), or *gluc78* (pUBI*gluc78*). Two-direction sequence analysis confirmed that sequence integrity was maintained across ligation sites and that the ATG start codon of *gluc78* was in the desired reading frame. The *gluc78* gene was introduced as part of complete plasmids or as MTECs (fragments containing only the promoter, open reading frame and terminator sequences). The *gluc78* genes were co-transformed with the *manA* selectable marker gene.

Ten transgenic plants contained the *gluc78* transgene from *T. atroviride* (Table 1), driven either by the constitutive ubiquitin promoter (pUB*gluc78*) or the *pin2* wound inducible promoter (pPA*gluc78*), whereas one plant contained only the selectable marker gene, *manA* (event 10.4A, data not shown). Genomic Southern blot analysis of T<sub>0</sub> and T<sub>1</sub> transformation events showed that events 7.10 1,2,3 and A,B,C were clones, whereas 7.7-2, 7.9, 2.1-A, 10.4-A and 7.6-B were unique transformation events (Figs 2 and 3). Twenty two and 111 pg of plasmid DNA represent 2 and 10 copies of the transgenes, respectively. Identical integration patterns were obtained for respective T<sub>1</sub> progeny (data not shown). The integration pattern of transformation events 7.10 1-3 and A-C were also confirmed by hybridisation with a radioactive labelled internal fragment of *gluc78* (Fig 2c). Transformation events 2.1-A and 7.6-B were obtained by co-transformation of the *manA* selectable marker gene and the antifungal gene *gluc78* driven by the constitutive promoter, ubiquitin (Fig. 3). Transformation events 2.1-A and 7.6 B were obtained introducing either full constructs or minimal transgene expression cassettes (MTECs), respectively. Restriction enzymes Hind III alone or Hind III in combination with Eco RI isolates the gene of interest including the promoter and terminator sequence (Figs 1 and 3). Southern blot fragments larger and smaller than the expected sizes are potentially concatamers or truncated versions of the introduced genes (Fig 3, lanes 11-13). DNA of transformation event 2.1A was digested with *Hind* III, which theoretically releases a 4.6 Kb fragment containing ubiquitin promoter plus *gluc78* plus nos terminator (lanes 4-8, Fig. 3A) and 2.2 Kb fragment (*Hind* III/*Eco* RI digest) containing the ubiquitin intron plus *manA* transgene (lane 9, Fig. 3B). The *Eco* RI site in the ubiquitin intron upstream of *gluc78* is potentially methylated as was the case for construct pAHC25 it originates from (Christensen and Quail, 1996). Therefore, the *Hind* III/*Eco* RI digest of transformation event 2.1A also resulted in a 4.6 Kb fragment (lane 9, Fig 3A) instead of the expected 3 Kb fragment..Transformation event 10.4-A contained only the selectable marker gene *manA*, but not the *gluc78* transgene (data not shown). (A co-transformation efficiency of 91% was therefore obtained.

An estimated 6-12 copies of the minimal transgene expression cassette containing *gluc78* were inserted in transformation event 7.6B. Apart from the expected 4.6 Kb fragment, numerous larger and smaller band sizes were obtained indicating truncated versions, concatamers, tandem repeat inserts or rearrangement of the transgene *gluc78* (Fig. 3a). PCR amplification of the antibiotic resistance gene, *bla*, was negative for transgenic line 7.6-B, illustrating that vector backbone was absent and only the linear minimal cassette integrated.

In order to determine the Mendelian segregation of the transgenic events (T<sub>1</sub> to T<sub>4</sub> progeny) obtained in this study, progeny of transgenic events were germinated on mannose containing medium (Table 2) as described previously (O'Kennedy *et al.*, 2004b). Often less than 50% of the seeds germinated on the selection medium, therefore not reflecting a typical Mendelian segregation during the first four generations.

### 3.2 Pathogenicity trials with *S. graminicola*

Table 2

Selected transgenic plants were subsequently inoculated with *S. graminicola*. T<sub>2</sub> transgenic pearl millet plants designated 7.10-B, 7.10-1 and 7.10-2 were inoculated by *S. graminicola* to screen for downy mildew resistance. Downy mildew disease was assessed 20 days after inoculation based on the incidence of infected plants within each pot (Table 3). The disease score for each treatment was the infection percentage for individual pot replicates. Thus, the disease score for each entry could range from 0-100% and was a continuous variable. Symptoms of systemic infection were seen as distinct chlorosis of infected leaves, white downy growth on both leaf surfaces (the asexual sporangiophores emerging from stomata) and stunted growth.

Transgenic event 7.10-2 (T<sub>2</sub> progeny) inoculated with *S. graminicola*, gave a disease score reduction of almost 58% when compared to inoculated control 842B plants (Table 3). Progeny of transgenic events 7.10-B and 7.10-1 however, gave a higher disease incidence (1.5 and 1.7 fold, respectively) when compared to control plants.

Table 3

### 3.3 Molecular analysis of the expression of the *gluc78* transgene

RNA and protein analyses were conducted to determine if a correlation existed between the expression levels of *gluc78*, and altered *S. graminicola* disease susceptibility. Expression of the *gluc78* transgene in leaves was analysed by RNA gel northern blotting and by protein gel western blotting for selected control and transgenic plants (Fig. 4 and 5). Wound-induced expression of the glucanase transcript of transformation event 7.10-2 were 2-4 times the intensity of the unwounded control leaf material 2, 6, 18, 24 and 48h after wounding (northern blot analysis not shown). Under the stated assay conditions, faint signals of the *gluc78* transcript were detected in both inoculated and non-inoculated non-transgenic plants (Fig. 4a, b and c; lanes 1 and 2). High levels of expression were obtained in infected transgenic plants, and slightly lower levels of expression were obtained in inoculated uninfected transgenic plants, when compared to control plants (Fig. 4). In addition, the protein encoded by *gluc78* was clearly expressed in transgenic plants inoculated with *S. graminicola* (Fig. 5), whereas the protein was absent in non-transgenic control plants whether the plants were inoculated with the pathogen or not.

RT-qPCR confirmed the occurrence of an increased level of transcript of the transgene in individually infected transgenic pearl millet events when infected by *S. graminicola* (Fig. 6). Transgene *gluc78* expression levels in control 842B (plant 1, non-challenged; plant 4, challenged but non-infected and plant 5, challenged and infected with *S. graminicola*) were absent. Plants 10, 12, 15 and 16 indicated by the downward diagonal pattern on the bars showed *S. graminicola* infection symptoms whereas the rest did not show any signs of downy mildew infection although challenged with the pathogen. RT-qPCR results (Fig 6) indicate that *S. graminicola* infected transgenic plants displayed higher *gluc78* expression levels than non-infected plants.

Fig. 6

#### 4. DISCUSSION

This is the first report introducing antifungal genes in a pearl millet parental breeding line which were previously used in breeding programmes for resistance to downy mildew. Our rationale was to further improve the moderately resistant breeding line 842B through the introduction of the antifungal gene *gluc78*. In previous studies, antifungal genes were introduced in a highly susceptible inbred line 7042S (Girgi *et al.*, 2006; Latha *et al.*, 2006).

Using a biolistic-mediated transformation protocol both complete constructs and MTECs were successfully introduced into line 842B. The overall goal of using MTECs was to enhance low copy number integration of transgenes, and minimise transgene rearrangements and gene silencing as previously reported (Fu *et al.*, 2000; Kohli *et al.*, 1998). Vector backbones have the tendency to promote transgene rearrangements, since the vector backbone has been shown to provide a number of recombination hotspots (Fu *et al.*, 2000). The removal of these sequences would possibly limit the influence of recombinogenic elements during the process of integration (Fu *et al.*, 2000). In this study, an estimated 6-12 copies of the transgene *gluc78* were inserted in pearl millet transformation event 7.6B, probably due to the high DNA concentration used for bombardment (183 ng per shot). Recently, Lowe and co-workers (2009) produced more than 1600 maize (inbred H99) events consistently yielding single copy events at high transformation frequencies using only 2.5 ng cassettes DNA per shot.

Generally, transgenic pearl millet plants expressing *gluc78* were more susceptible to *S.graminicola* and displayed severe disease symptoms relative to wild-type 842B plants. *In vivo* and *in vitro* infection of the same transgenic pearl millet lines with *Puccinia substriata* also showed an increase in disease incidence (data not shown). However, T<sub>2</sub> progeny of transgenic event 7.10-2, inoculated with *S. graminicola*, resulted in a disease score reduction of almost 58%, whereas transgenic events 7.10-B and 7.10-1 gave a significantly higher disease incidence when compared to control plants. Quantitative real time PCR confirmed the occurrence of an increased level of the *gluc78* transcript of the transgene in individually

infected transgenic pearl millet, as well as plants that were infected with *S. graminicola* but did not confer resistance to these pathogens. Sanders and co-workers (2002) reported that silencing of a heterologous glucanase, lead to diminishing levels of the plant's endogenous glucanase activity, and increased disease susceptibility. The authors reported that this could be due to post-transcriptional gene silencing (PTGS) induced among glucanase genes that share some homology. The glucanase from *T. atroviride* shares no sequence homology with glucanases of plants or bacteria and therefore PTGS is unlikely to be the cause of inefficiency of the introduced gluc78.

Callose, a sugar polymer that is composed of (1,3)- $\beta$ -D-glucan, is deposited between the plasma membrane and the cell wall in close proximity to the invading pathogen as part of the HR to infection by fungi and viruses (Donofrio and Delaney, 2001). Accumulation of callose in response to insect and pathogenic attacks has been well documented (Flors *et al.*, 2007; Kruger *et al.*, 2002; Saheed *et al.*, 2009), and indicates that callose deposition constitutes a defence mechanism against insect and pathogen invasion. Iglesias and Meins (2000) proved that the movement of plant viruses is delayed in a  $\beta$ -1,3-glucanase-deficient tobacco mutant, showing an enhanced resistance related to augmented deposition of callose. Inhibition of callose synthesis by infiltrating 2-DDG, a callose synthesis inhibitor (Flors *et al.*, 2005; Ton and Mauch-Mani, 2004), reversed the enhanced resistance of the transgenic plants lacking  $\beta$ -1,4-glucanases when compared to the wild-type levels. Similarly, transformation of tobacco and *Nicotiana sylvestris* expressing an antisense construct of a class I  $\beta$ -1,3-glucanase from the latter species (homologous to tobacco PR-2d) became less diseased than non-transformed control plants. This result is interpreted as being due to enhanced callose accumulation at the plasmodesmata, leading to more effective blockage of virus transport from cell to cell (Bucher *et al.*, 2001; Fridborg *et al.*, 2003). In the present study, it is therefore possible that expression of the gluc78 transgene negatively modulates callose deposition through gluc78 hydrolysis of  $\beta$ -1,3 glycosidic bonds in linear or branched glucans of the plant.

Production of lytic enzymes by *Trichoderma* spp. and their effect on the growth of phytopathogenic fungi is well established (Schirmböck *et al.*, 1994; Lorito *et al.*, 1998; Fogliano *et al.*, 2002) but not the effect on plant glucan and callose formation. The effect of using *Trichoderma* spp. on the endogenous glucan and callose levels in plants still needs to be elucidated.

This is the first report of an antifungal  $\beta$ -1,3-glucanase from *T. atroviride* that is stably expressed in a transgenic cereal crop. Preliminary pathogenicity trials and molecular analysis indicated that the expression of this protein during wounding, *P. substriata* and *S. graminicola* infection, increased the glucanase transcript levels, but promotes disease susceptibility rather than improve disease tolerance. Preliminary results expressing  $\beta$ -1,3-glucanase from *Trichoderma atroviride* (*gluc78*) in transgenic pearl millet, indicate a failure to confer enhanced resistance to *S. graminicola*. Despite significant progress in elucidating defence mechanisms in cereal crops (Salzman *et al.* 2005; Zhu-Salzman *et al.* 2004; Ayliffe and Lagudah, 2004), and particularly in pearl millet (Crampton *et al.* 2009), the mechanism that governs resistance to downy mildew remains an enigma.

## **ACKNOWLEDGEMENTS**

pAHC25 was a kind gift from Prof. Peter H. Quail, Director of Research, PGEC, Albany, California. pBIN-GLUC, containing the *gluc78* gene, was obtained from Prof Matteo Lorito from the University of Naples, Italy. The project was co-funded by the European Commission contract number: ERBIC18 CT98 0316. The positive selectable marker gene *pmi*, encoding mannose-6-phosphate isomerase (phosphomannose isomerase) was obtained from Syngenta Seeds AG under a material transfer agreement signed by the CSIR Biosciences. We acknowledge Dr. J. Wilson and Dr W. de Miliano for providing the *P. substriata* strains and Gillian Smith, Guguletho Ngwenya, Rufus Khonothi, Jeffrey Mathabe and Moses Mokoena for technical assistance. Some results presented in this document are outputs from several research projects of the Plant Sciences Research Programme funded by the UK Department for International Development (DFID) and administered by the Centre for Arid



Zone Studies (CAZS) for the benefit of developing countries. The views expressed are not necessarily those of DFID.

## REFERENCES

- Ait-Lahsen, H., Soler, A., Rey, M., de la Cruz, J., Monte, E. and Llobell, A., 2001. An antifungal exo- $\alpha$ -1,3-Glucanase (AGN13.1) from the biocontrol fungus *Trichoderma harzianum*. *Applied Environmental Microbiology* 67(12): 5833-5844.
- Ayliffe, M.A. and Lagudah, E.S., 2004. Molecular genetics of disease resistance in cereals. *Annals of Botany* 94: 765-773.
- Borkowska, M., Krzymowska, A., Talarczyk, A., Awan, M.F., Yakovleva, L., Kleckowski, K., Wielgat, B., 1998. Transgenic potato plants expressing soybean beta-1,3-endoglucanase gene exhibit an increased resistance to *Phytophthora infestans*. *Zeitschrift für Naturforschung* 53 (11-12):1012-1026.
- Bucher, G.L., Tarina, C., Heinlein, M., Di Serio, F., Meins, F. and Iglesias, V.A., 2001. Local expression of enzymatically active class I  $\beta$ -1,3-glucanase enhances symptoms of TMV infection in tobacco. *Plant Journal* 28(3): 361-369.
- Christensen, A.H. and Quail, P.H., 1996. Ubiquitin promoter-based vectors for high-level expression of selectable and/or screenable marker genes in monocotyledonous plants. *Transgenic Research* 5: 213-218.
- Cohen-Kupiec, R., Broglie, K.E., Friesem, D., Broglie, R.M., Chet, I., 1999. Molecular characterization of a novel beta-1,3-exoglucanase related to mycoparasitism of *Trichoderma harzianum*. *Gene* 226 (2): 147-154.
- Crampton, B.G., Hein, I., Berger, D.K., 2009. Salicylic acid confers resistance to a biotrophic rust pathogen, *Puccinia substriata* in pearl millet, *Pennisetum glaucum*, *Molecular Plant Pathology*, 10 (2): 291-304.
- Dellaporta, S.L., Wood, J. and Hicks, J.B., 1983. A plant DNA miniprep: Version II. *Plant Molecular Biology Reports* 1: 19-21.
- Donofrio, N.M. and Delaney, T.P., 2001. Abnormal callose response phenotype and hypersusceptibility to *Peronospora parasitica* in defense-compromised *Arabidopsis* nim1-1

and salicylate hydroxylase-expressing plants. *Molecular Plant Microbe Interaction* 14, 439–450.

Donzelli, B.G., Lorito, M., Scala, F. and Harman, G.E., 2001. Cloning, sequence and structure of a gene encoding an antifungal glucan 1,3-beta-glucosidase from *Trichoderma atroviride* (T. harzianum). *Gene* 17 (1-2): 199-208.

Emani, C., Garcia, J.M., Lopato-Finch, E., Pozo, M.J., Uribe, P., Kim, D-J., Sunilkumar, G., Cook, D.R., Kenerley, C.M. and Rathore, K.S., 2003. Enhanced fungal resistance in transgenic cotton expressing an endochitinase gene from *Trichoderma virens*. *Plant Biotechnology Journal* 1: 321-336.

Flors, V., Ton, J., Jakab, G., and Mauch-Mani, B., 2005. Abscisic Acid and Callose: Team Players in Defence Against Pathogens? *Journal of Phytopathology* 153, 377–383.

Flors, V., Leyva, Mde, L., Vicedo, B., Finiti, I., Real, M.D., García-Agustín, P., Bennett, A.B., González-Bosch, C., 2007. Absence of the endo-beta-1,4-glucanases Cel1 and Cel2 reduces susceptibility to *Botrytis cinerea* in tomato. *Plant Journal* 52(6):1027-40.

Fogliano, V., Ballio, A., Gallo, M., Woo, S., Scala, F. and Lorito, M., 2002. *Pseudomonas lipodepsipeptides* and fungal cell wall degrading enzymes act synergistically in biocontrol. *Molecular Plant-Microbe Interactions* 15:323-333.

Fridborg, I., Grainger, J., Page, A., Coleman, M., Findlay, K. and Angell, S. 2003. TIP, a novel host factor linking callose degradation with the cell-to-cell movement of potato virus X. *Molecular Plant-Microbe Interactions* 16 (2): 132-140.

Fu, X., Duc, L-T., Fontana, S., Bong, B.B., Tinjuangjun, P., Sudhakar, D., Twyman, R.M., Christou, P. and Kohli, A., 2000. Linear transgene constructs lacking vector backbone sequences generate low-copy-number transgenic plants with simple integration patterns. *Transgenic Research* 9: 11-19.

Gheong, Y.H., Kim, C.Y., Chun, H.J., Moon, B.C., Park, H.C., Kim, J.K., Lee, S., Han, C., Lee, S.Y. and Cho, M.J., 2000. Molecular cloning of a soybean class III beta-1,3-glucanase gene that is regulated both developmentally and in response to pathogen infection. *Plant Science* 154(1): 71-81.

Girgi, M., Breese, W.A., Lörz, H., Oldach, K.H., 2006. Rust and downy mildew resistance in pearl millet (*Pennisetum glaucum*) mediated by heterologous expression of the *afp* gene from *Aspergillus giganteus*. *Transgenic Research* 15, 313–324.

Goldman, J.J., Hanna, W.W., Fleming, G., Ozias-Akins, P., 2003. Fertile transgenic pearl millet [*Pennisetum glaucum* (L.) R. Br.] plants recovered through microprojectile bombardment and phosphinothricin selection of apical meristem-, inflorescence-, and immature embryoderived embryogenic tissues. *Plant Cell Reports* 21, 999–1009.

Iglesias, V.A. and Meins, F., 2000. Movement of plant viruses is delayed in a beta-1,3-glucanase-deficient mutant showing a reduced plasmodesmatal size exclusion limit and enhanced callose deposition. *Plant Journal* 21, 157–166.

Ji, C., Norton, R.A., Wicklow, D.T. and Dowd, P.F., 2000. Isoform patterns of chitinase and beta-1,3-glucanase in maturing corn kernels (*Zea mays* L.) associated with *Aspergillus flavus* milk stage infection. *Journal of Agricultural and Food Chemistry* 48(2): 507-511.

Jones, E.S., Breese, W.A. and Shaw, D.S., 2001. Inoculation of pearl millet with the downy mildew pathogen, *S. graminicola*: Chilling inoculum to delay zoospore release and avoid spray damage to zoospores. *Plant Pathology* 50: 310-316.

Jung, H.W. and Hwang, B.K., 2000. Pepper gene encoding a basic beta-1,3-glucanase is differentially expressed in pepper tissue upon pathogen infection and ethephon or methyl jasmonate treatment. *Plant Science* 159(1): 97-106.

Kini, K.R., Vasanthi, N.S. and Shetty, H.S., 2000a.. Induction of  $\beta$ -1,3-glucanase in seedlings of pearl millet in response to infection by *S. graminicola*. *European Journal of Plant Pathology* 106: 267-274.

Kini, K.R., Vasanthi, N.S., Umesh-Kumar, S. and Shetty, H.S., 2000b. Purification and properties of a major isoform of  $\beta$ -1,3-glucanase from pearl millet seedlings. *Plant Science* 150: 139-145.

Kohli, A., Leech, M., Vain, P., Laurie, D.A. and Christou, P., 1998. Transgene organization in rice engineered through direct DNA transfer supports a two-phase integration mechanism mediated by the establishment of integration hot spots. *Proceedings of the National Academy of Science. USA* 95: 7203-7208.

Kononov, M.E., Bassuner, B. and Gelvin, S.B., 1997. Integration of T-DNA binary vector 'backbone' sequences into the tobacco genome: evidence for multiple complex patterns of integration. *Plant Journal* 11(5): 945-957.

Kruger, W.M., Carver, T.L.W. and Zeyen, R.J., 2002. Effects of inhibiting phenolic biosynthesis on penetration resistance of barley isolines containing seven powdery mildew resistance genes or alleles. *Physiol Mol Plant Pathology* 61, 41–51.

Laemmli, U.K., 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227: 680-685.

Lange, M., Vince, E., Møller, M.G. and Holm, P.B., 2006. Molecular analysis of transgene and vector backbone integration into the barley genome following *Agrobacterium*-mediated transformation. *Plant Cell Reports* 25: 815-820.

Latha, A.M., Rao, K.V., Reddy, T.P. and Reddy, V.D., 2006. Development of transgenic pearl millet (*Pennisetum glaucum* (L.) R. Br.) plants resistant to downy mildew. *Plant Cell Reports* 29, 927–935.

Lorito, M., Woo, S.L., Garcia, I., Colucci, G., Harman, G.E., Pintor-Toto, J.A., Filippone, E., Muccifora, S., Lawrence, B., Zoina, A. and Scala, F., 1998. Genes from mycoparasitic fungi as a source for improving plant resistance to fungal pathogens. *Proceedings of the National Academy of Science USA* 95(14): 7860-7865.

Lowe, B.A., Prakash, N.S., Way, M., Mann, M.T., Spencer, T.M. and Boddupalli R.S., 2009. Enhanced single copy integration events in corn via particle bombardment using low quantities of DNA. *Transgenic Research* 18:831-840.

O’Kennedy, M.M., Smith, G. and Botha, F.C., 2004a. Improved regeneration efficiency of a pearl millet (*Pennisetum glaucum*) breeding line. *South African Journal of Botany* 70(4): 502-508.

O’Kennedy, M.M., Burger, J.T. and Botha, F.C., 2004b. Pearl millet transformation system using the positive selectable marker gene phosphomannose isomerase. *Plant Cell Reports* 22(9): 684-690.

Saheed, S.A., Cierlik, I., Larsson, K.A., Delp, G., Bradley, G., Jonsson, L.M. and Botha, C.E., 2009. Stronger induction of callose deposition in barley by Russian wheat aphid than

bird cherry-oat aphid is not associated with differences in callose synthase or beta-1,3-glucanase transcript abundance. *Physiology Plantarum* 135(2):150-61.

Salzman, R.A., Brady, J.A., Finlayson, S.A., Buchanan, C.D., Summer, E.J., Sung, F., Klein, P.E., Klein, R.R., Pratt, L.H., Cordonnier-Pratt, M. and Mullet, J.E., 2005. Transcriptional profiling of sorghum induced by methyl jasmonate, salicylic acid and aminocyclopropane carboxylic acid reveals cooperative regulation and novel gene responses. *Plant Physiology* 138: 352-368.

Sambrook, J., Fritsch, E.F. and Maniatis, T. 1989. *Molecular Cloning: a Laboratory Manual*, 2<sup>nd</sup> edn., Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.

Sanders, M., Maddelein, W., Depicker, A., Van Montagu, M., Cornelissen, M. and Jacobs, J., 2002. An active role for endogenous  $\beta$ -1,3-glucanase genes in transgene-mediated co-suppression in tobacco. *EMBO Journal* 21: 5824-5832.

Sarosh, B.R., Sivaramakrishnan, S. and Shetty, H.S. 2005. Elicitation of defense related enzymes and resistance by L-Methionine in pearl millet against downy mildew disease caused by *Sclerospora graminicola*. *Plant Physiol Biochem* 43: 808-815.

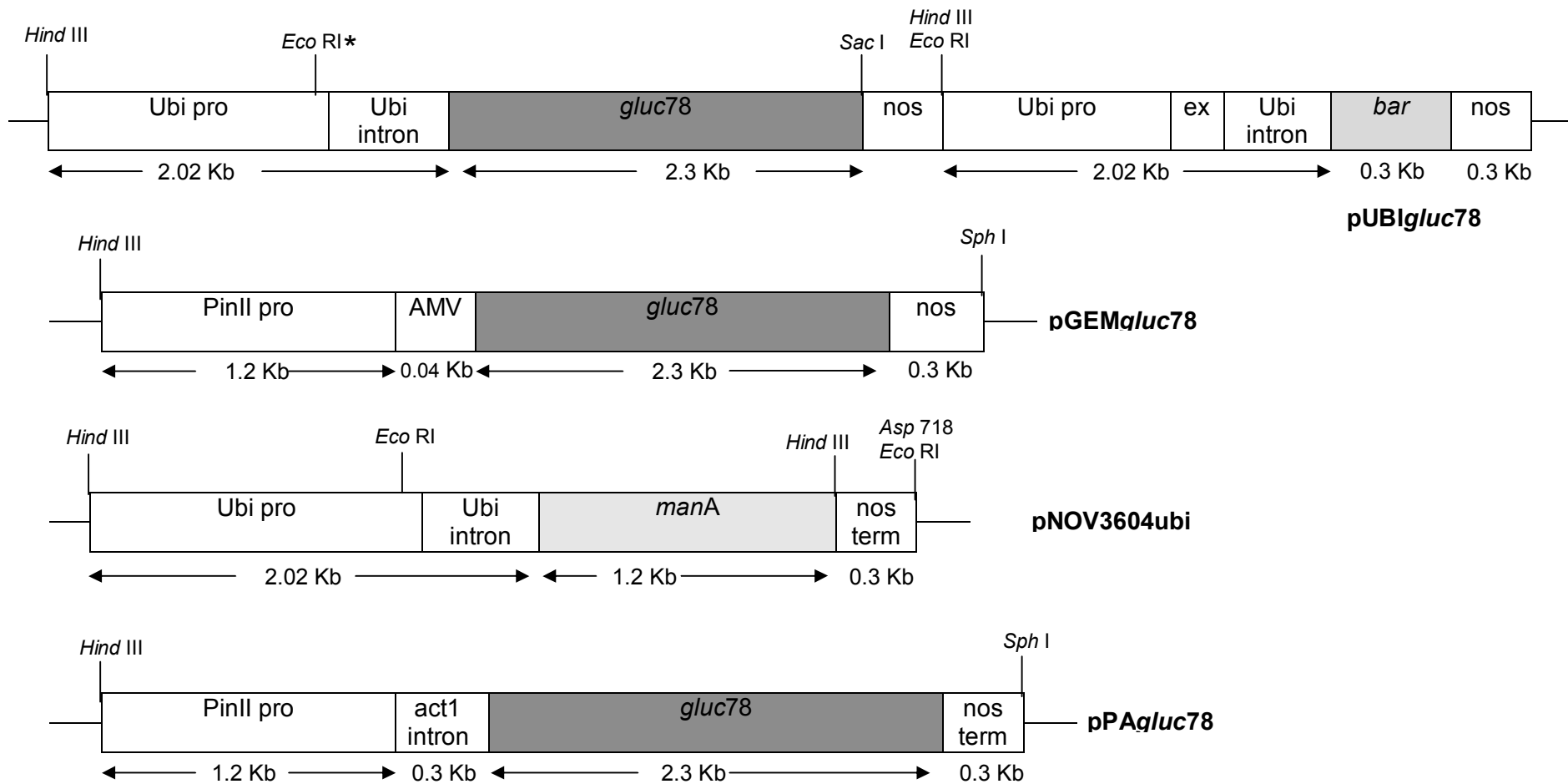
Schirmböck, M., Lorito, M., Wang, Y-L., Hayes, C.K., Arisan-Atac, I., Scala, F., Harman, G.E. and Kubicek, C.P., 1994. Parallel formation and synergism of hydrolytic enzymes and peptaibol antibiotics, molecular mechanism involved in the antagonistic action of *Trichoderma harzianum* against phytopathogenic fungi. *Applied and Environmental Microbiology* 60 (12): 4364-4370).

Ton, J. and Mauch-Mani, B., 2004.  $\beta$ -amino-butyric acid-induced resistance against necrotrophic pathogens is based on ABA-dependent priming for callose. *The Plant Journal* 38(1): 119-130.

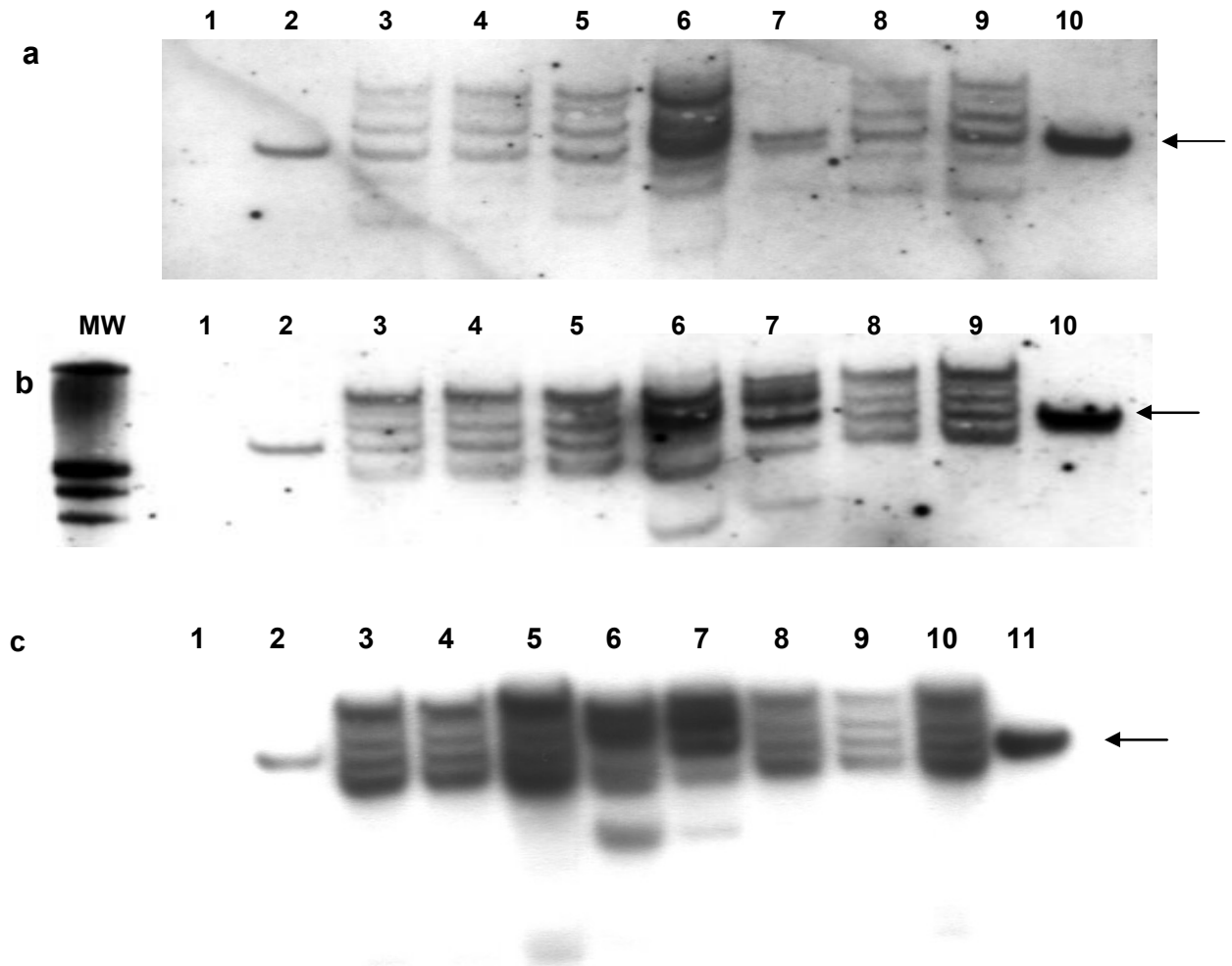
Wilson, J.P., Hanna, W.W., Gascho, G.J., 1996. Pearl millet grain yield loss from rust infection. *Journal of Production Agriculture* 80: 543–545.

Xu, D., McElroy, D., Thornburg, R.W., and Wu, R., 1993. Systemic induction of a potato *pin2* promoter by wounding, methyl jasmonate, and abscisic acid in transgenic rice plants. *Plant Molecular Biology* 22: 573 – 588.

Zu-Salzman, K., Salzman, R.A., Ahn, J-E. and Koiwa, H., 2004. Transcriptional regulation of sorghum defense determinants against a phloem-feeding aphid. *Plant Physiology* 134: 420-431.

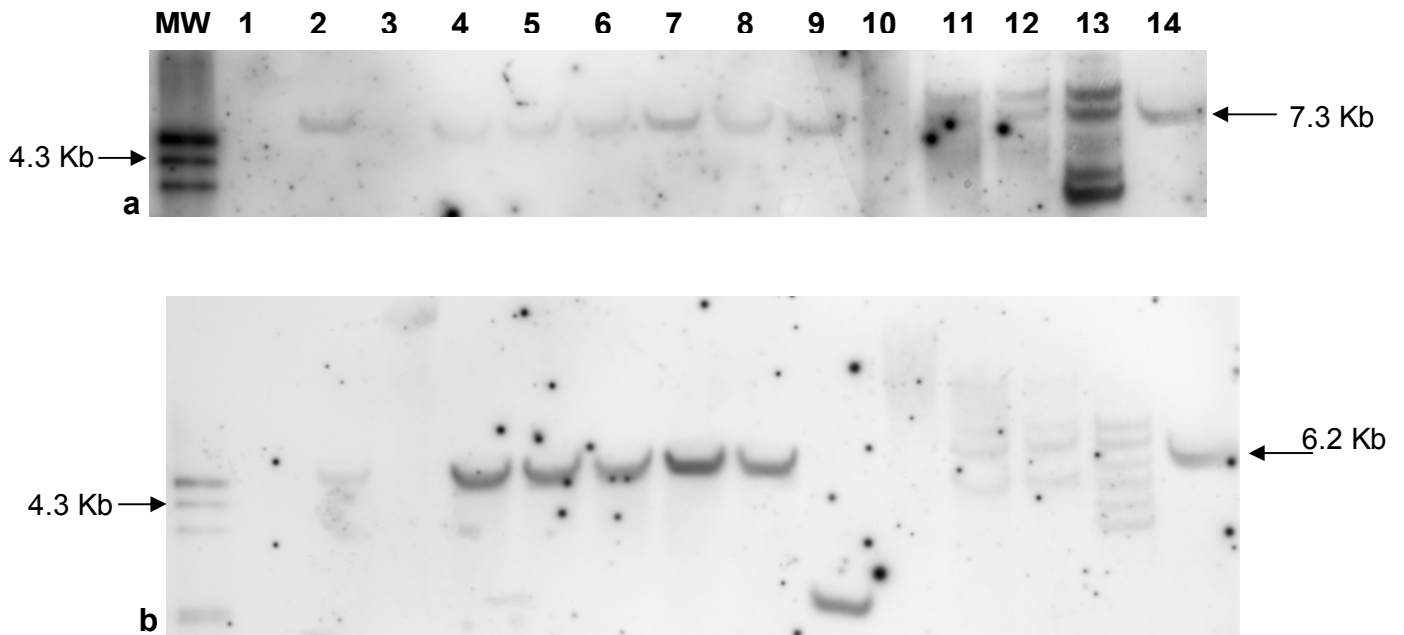


**Fig. 1** Constructs containing either the selectable marker genes *manA* or *bar*, or the antifungal gene *gluc78*. The *Eco RI* site in the ubiquitin intron upstream of *gluc78* is potentially methylated (indicated by an asterisks, \*) as was the case for construct pAHC25 it originates from Christensen and Quail (1996).

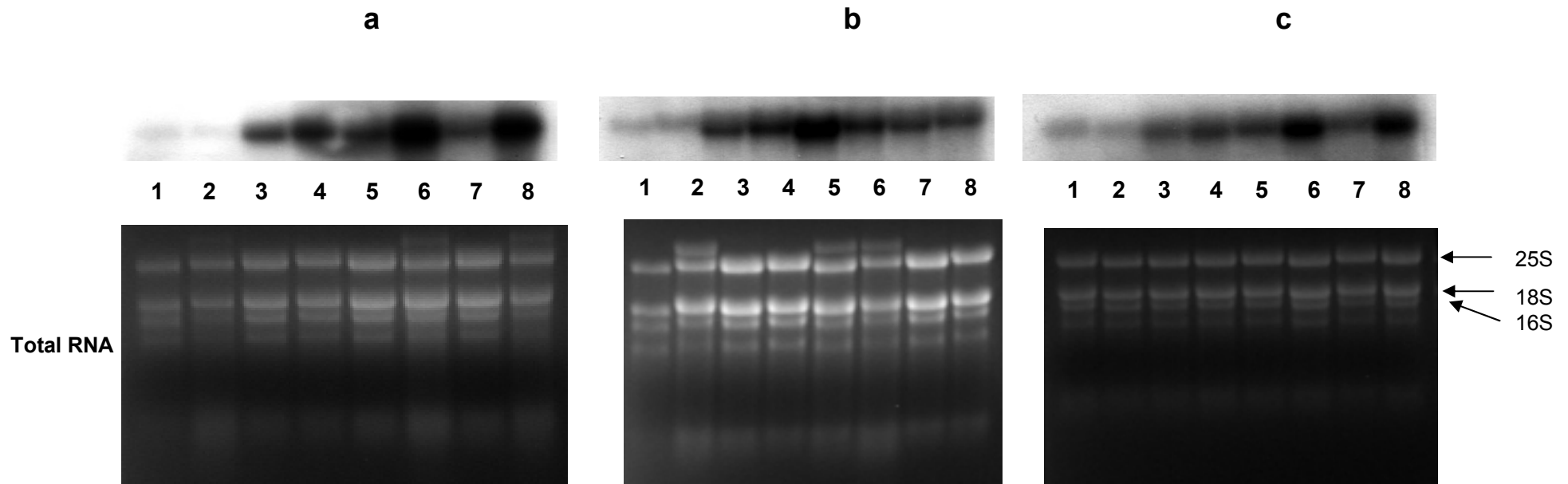


**Fig. 2:** Southern blot analysis of independent  $T_0$  transformation events of pearl millet genotype 842B. Genomic DNA was purified from plant leaf material, restricted with Hind III and resolved in a 0.8% agarose gel, transferred to a nylon membrane, then probed with an internal DNA fragment of the *manA* (blot a) or *gluc78* (blots b and c) transgene, either DIG labelled (blots a and b) or radioactive labelling (blot c). Blots a and b: MW, DIG labelled molecular weight marker III (Roche Diagnostics South Africa); lane 1, untransformed pearl millet leaf material of genotype 842B; lanes 2 and 10, represent untransformed pearl millet spiked with 2 and 10 transgene copies respectively; lane 3, plant 7.10A; lane 4, plant 7.10B; lane 5, plant 7.10C; lane 6, plant 7.7.2; lane 7, plant 7.9; lane 8, plant 7.10.1; lane 9, plant 7.10.3. Blot c: lane 1, untransformed control; lanes 2 and 11, represent untransformed pearl millet spiked with 2 and 10 transgene copies respectively; lane 3, plant 7.10A; lane 4, plant 7.10B; lane 5, plant 7.10C; lane 6, plant 7.7.2; lane 7, plant 7.9; lane 8, plant 7.10.1; lane 9, plant 7.10.2; lane 10, plant 7.10.3. The arrows indicate the size of pNOV3604ubi (6.2 kb) or pPAgluc78 (7.15 kb) linearised with Hind III.

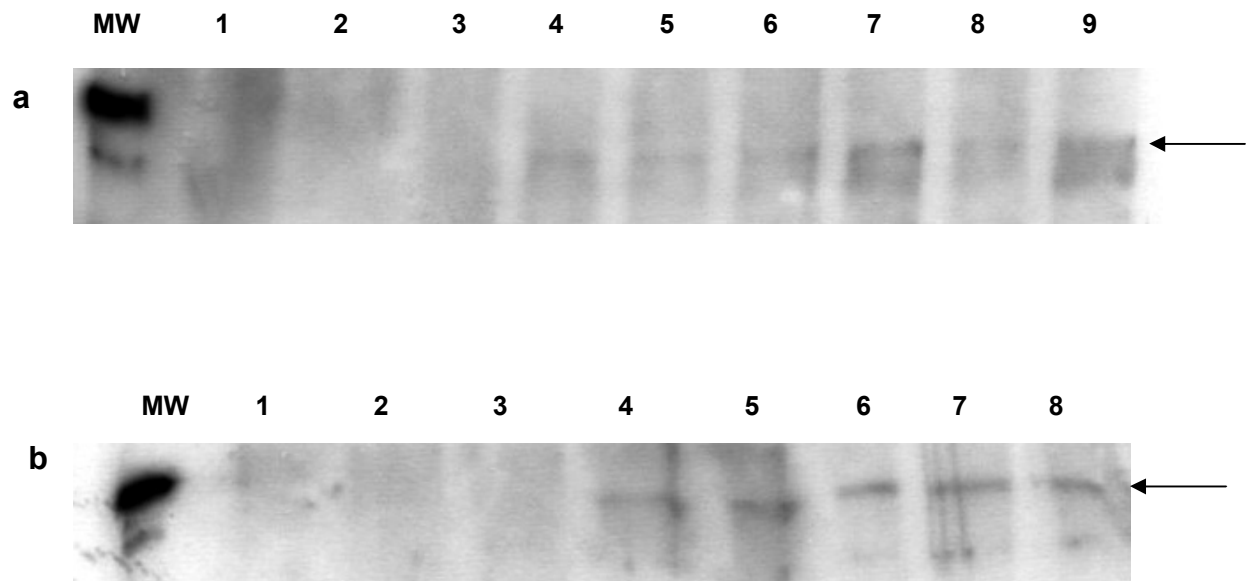




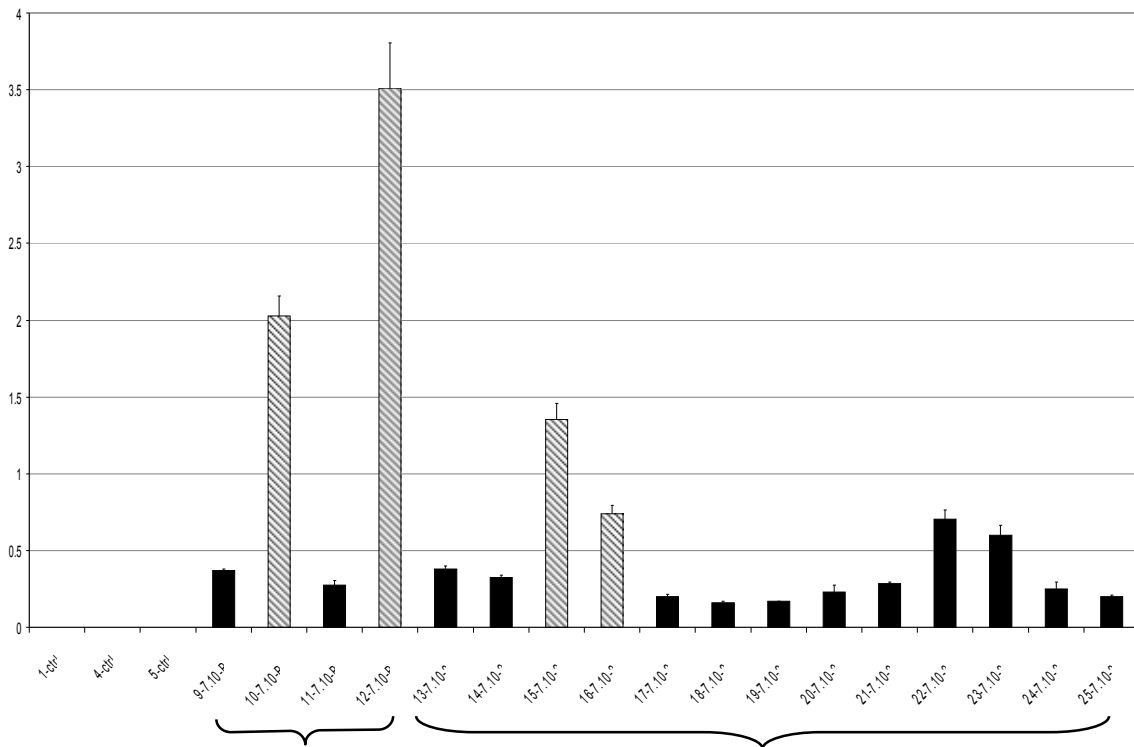
**Fig. 3:** Southern blot analysis of independent T<sub>0</sub> transformation events of pearl millet genotype 842B transformed with full constructs (2.1A) or minimal transgene cassettes (7.6B). Genomic DNA was purified from plant leaf material, resolved in a 0.8% agarose gel, transferred to a nylon membrane and then probed with a DIG labelled internal DNA fragment of *gluc78* (blot a) or *man A* (blot b). MW, DIG labelled molecular weight marker III; lane 1, DNA of untransformed pearl millet leaf material of genotype 842B; lanes 2 and 14, represent DNA of untransformed 842B spiked with 2 and 10 transgene copies respectively; lane 3, undigested DNA of plant 2.1A T<sub>0</sub>; lane 4, 2.1A T<sub>0</sub> digested with Hind III; lanes 5–8, genomic DNA of individual T<sub>1</sub> progeny plants of 2.1A digested with Hind III; lane 9, genomic DNA of 2.1A digested with Hind III/Eco RI; lane 10, genomic DNA of T<sub>1</sub> 7.6B undigested; lanes 11 and 12, genomic DNA of individual T<sub>1</sub> progeny plants of 7.6B digested with Hind III; lane 13, genomic DNA of an individual T<sub>1</sub> plant of 7.6B digested with Hind III/Eco RI.



**Fig. 4:** Detection of the *gluc78* *T. atroviride* glucanase transcript in infected and non-infected selfed progeny T<sub>2</sub> generation transgenic and control untransformed pearl millet genotype 842B. Total RNA was extracted from leaf material and 8 µg of RNA loaded/lane and blotted to a nylon membrane. The RNA gel blot was hybridised with *agluc78* internal DNA fragment probe. Northern blot analysis was performed on a: lane 1, non-infected non-challenged with *S. graminicola* control plant; lane 2, infected control plant; lanes 3 and 4, two individual infected plants from transformation event 7.10.1; lanes 5 and 7, non-infected 7.10B; lanes 6 and 8, infected 7.10B; b: lane 1, non-infected control plant; lane 2, infected control plant; lanes 3, 4, 7 and 8, non-infected challenged transgenic plants of event 7.10.2; lanes 5 and 6, infected challenged transgenic plants of event 7.10.2; c: lane 1, non-infected non-challenged control plant; lane 2, non-infected challenged control plant; lanes 3–8, non-infected challenged transgenic plants of event 7.10.2. Ethidium bromide stained gels show that equal amounts of total RNA were loaded in each lane, in each set a, b and c.



**Fig. 5:** Western blot analysis of total soluble proteins from leaves of transgenic and non-transgenic control plants. A two microgram aliquot of protein was separated in each lane, the blot immunoprobed with polyclonal anti-*gluc78* antibodies (dilution 1:750) and visualised with anti-rabbit secondary antibodies (dilution 1:5000). Blots a and b: MW, ECL molecular weight marker; lane 1, non-infected non-challenged with *S. graminicola* control plant; lane 2, non-infected *S. graminicola* challenged plant; lane 3, infected control plant. Blot a: lanes 4 and 5, two individual infected plants from transformation event 7.10.1; lanes 6 and 8, non-infected 7.10B; lanes 7 and 9, infected 7.10B. Blot b: lanes 4, 5 and 8, non-infected challenged transgenic plants of event 7.10.2; lanes 6 and 7, infected challenged transgenic plants of event 7.10.2. The arrows indicate the protein size of approximately 78 kDa.



**Fig. 6:** Transcript levels of the transgene *gluc78* from *T. atroviride* determined by RT-qPCR in transgenic pearl millet and non-transgenic 842B control plants following infections. The Y-axis indicates the  $\beta$ -glucanase expression relative to 18S RNA levels and the X-axis shows the 842B control plants followed by individual infected plants. Individual progeny plants (bars) of transgenic events (7.10-2 and 7.10B) are grouped together by brackets and compared to control 842B plants. The plant numbering for each bar is given as the plant number for each individual experiment followed by the transgenic event number (7.10-2 or 7.10B). The error bars represent three technical replicates.  $\beta$ -Glucanase expression in control 842B (plant 1, non-challenged; plant 4, challenged but non-infected and plant 5, challenged and infected with *S. graminicola*) compared to transgenic events 7.10B and 7.10-2, four and thirteen plants respectively, inoculated with *S. graminicola*. Plants 10, 12, 15 and 16 indicated by the downward diagonal pattern on the bars (▨) showed symptoms of *S. graminicola* infection, whereas plants indicated by the black boxes showed no symptoms of *S. graminicola* infection.

**Table 1:** Conditions for bombardment of pearl millet IZEs of genotype 842B pre-cultured on medium J and subsequently bombarded with either full constructs (experiment 1 and 2) or minimal transgene expression cassette (experiments 3 and 4) of pUbi-gluc78 (ubi gluc), pGEM gluc78 (AMV gluc) or pPA gluc78 (PA gluc) (containing the beta-1,3-glucanase 78) and the *manA* gene as selectable marker gene.

Exp. #	Helium pressure (kPa)	Constructs	DNA introduced (ng per shot)	Transgenic events
1	1000	<i>manA</i> :pPAgluc	64:104	7.7.2
	1200			7.9
				7.10 (1-3) 7.10 (A-C)
2	1000	<i>manA</i> :ubi gluc	85:170	2.1A
3	900	<i>manA</i> :ubi gluc	70:113	7.6B
4	900	<i>manA</i> :AMV gluc	66:182	10.4A

**Table 2:** Seed germination on mannose containing medium (1 g/l maltose and 15 g/l mannose) or maltose alone for 842B control plants.

Pearl millet transformation events	Number of seed <i>manA</i> pos : number of seeds <i>manA</i> neg			
	T <sub>1</sub> + : -	T <sub>2</sub> + : -	T <sub>3</sub> + : -	T <sub>4</sub> + : -
842B control				44 : 14 19 : 21
7.6B		59 : 377		
2.1A		23 : 21	7 : 5	
7.10 A	4 : 142	24 : 76		
7.10 B	1 : 11	3 : 6	26 : 14	30 : 15 11 : 29
7.10 C	7 : 163	12 : 22 21 : 29		33 : 10 21 : 19
7.10.1	70 : 772	24 : 76 33 : 16		18 : 27 10 : 30
7.10.2	16 : 408	50 : 165 5:72		34 : 11 16 : 24
7.10.3	21 : 196	12 : 88 9 : 32 8 : 32		
7.9	1 : 32	36 : 421 20:80	3 : 9	
7.7.2	2 : 74	35 : 36 2:34	10 : 38 2 : 10	

**Table 3:** Transgenic and non-transgenic control plants screened for downy mildew susceptibility by inoculation with *S. graminicola*.

Experimental Pearl Millet plant material	Total number of plants treated	Number of plants infected	Overall Disease Incidence (%)
Uninoculated, non-transgenic, 842B	36	0	0.0
Inoculated, non-transgenic, 842B	41	17	41.5
Inoculated, transgenic event 7.10.B	34	24	70.6
Inoculated, transgenic event 7.10.1	5	3	60.0
Inoculated, transgenic event 7.10.2	57	10	17.5