Niche differentiation among invasive crayfish and their impacts on ecosystem

structure and functioning

Jackson, M.C. 1,2*, Jones, T. 1, Milligan, M. 1, Sheath, D. 1,3, Taylor, J. 1, Ellis, A. 4, England, J. 4 and

Grey, J.1*

1. School of Biological and Chemical Sciences, Queen Mary University of London, London, UK.

2. Centre for Invasion Biology, Department of Zoology and Entomology, University of Pretoria,

Hatfield, South Africa.

3. School of Applied Sciences, Bournemouth University, Poole, UK.

4. Environment Agency, Hatfield, Hertfordshire, UK.

* Corresponding authors: mjackson@zoology.up.ac.za; j.grey@qmul.ac.uk

Running head: Interactions among invasive crayfish

Key words: Invasive species, ecosystem functioning, stable isotopes, trophic cascades, crayfish.

Summary

1. Many aquatic ecosystems sustain multiple invasive species and interactions among them

have important implications for ecosystem structure and functioning. Here we examine

interactions among two pairs of invasive crayfish species, each established in separate

communities in close proximity in the Thames catchment, U.K. (signal, Pacifastacus

leniusculus and virile, Orconectes virilis; red swamp Procambarus clarkii and Turkish,

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Astacus leptodactylus) to address two questions: do sympatric invasive crayfish occupy a smaller niche than their allopatric counterparts due to potential resource competition; and do interactions among invasive species amplify or mitigate one another's impacts on the ecosystem?

- 2. Two fully-factorial mesocosm experiments (one for each crayfish pair) were used to investigate crayfish diet, and their impact on benthic invertebrate community structure, benthic algal standing stock and leaf litter decomposition rates, in allopatric and sympatric populations, compared to a crayfish free control. We used stable isotope analysis to examine crayfish diet in the mesocosms and in allopatric populations of each species in the Thames catchment.
- 3. Isotopic niche width did not vary significantly between allopatric and sympatric populations of crayfish pairs in the mesocosms and isotopic niche partitioning in all the wild populations suggests the invaders can coexist.
- 4. All four species altered benthic invertebrate community structure but with differing functional effects, often mediated via trophic cascades. Red swamp crayfish predation upon snails evidently promoted benthic algal standing stock via reduction in grazing pressure. However, a trophic cascade whereby the crayfish consumed native invertebrate shredders, causing a reduction in net leaf litter decomposition, was decoupled by red swamp and signal crayfish since they consumed leaf litter directly and thus moderated the cascade to a trickle when in sympatry with Turkish or virile crayfish, respectively.

- 5. Benthic invertebrate predator abundance was significantly reduced by sympatric red swamp and Turkish crayfish but not independently when in allopatry, indicating an amplified effect overall when in sympatry.
- 6. Our results suggest that the combined effect of multiple invasions on the ecosystem can reflect either an additive effect of their independent impacts, or an amplified effect, which is greater than the sum of their independent impacts. A lack of general pattern in their effects makes any potential management strategy more complex.

Introduction

Biological invasions are recognised as a significant driver of global environmental change with consequences from the individual, through to the whole ecosystem level of organisation (Sala *et al.*, 2000; Simon and Townsend, 2003; Ricciardi, 2007). Invasive species can severely disrupt the organisation of native communities by displacing native species or by reducing their abundance (Gurevitch and Padilla, 2004). These alterations in species assemblages and biodiversity can have major implications for ecosystem processes (Olden *et al.*, 2004; Hector and Bagchi, 2007; Pejchar and Mooney, 2009). As the pace of global change accelerates, many ecosystems sustain multiple invaders (e.g. Cohen and Carlton, 1998; Jackson and Grey, 2013; Ricciardi, 2006) and the interactions among them will have important consequences. In the UK alone, there are estimated to be almost 2000 non-native species, costing £1.7 billion in management and damage per annum (Roy *et al.*, 2012). With other environmental stressors such

as climate change facilitating invasions (Rahel and Olden, 2008), many more invaders are on the horizon (Gallardo and Aldridge, 2013a, 2013b).

Despite recent progress in invasion biology (e.g. Johnson *et al.*, 2009; Catford *et al.*, 2012; Sanders *et al.*, 2003), our knowledge regarding specific ecological impacts, and the mechanisms behind them, remain limited for most invaders. This is particularly true of animal invasions in aquatic habitats; there is a distinct bias in the literature towards studying terrestrial plant invasions (Lowry *et al.*, 2013). There is even less research on multi-species invasions involving complex interactions between invasive species and how those invaders integrate within food webs of native communities (Kuebbing, Nuñez and Simberloff, 2013).

Empirical evidence supports both facilitative and negative interactions between sympatric invaders. The Invasional Meltdown model predicts that disturbance caused by one invasive species will facilitate the establishment of further invaders (Simberloff and Von Holle, 1999; Simberloff, 2006) by, for example, removing a natural competitor or predator (Grosholz, 2005). Invasive species might act in synergy to magnify their independent impacts on ecosystem structure and functioning (Simberloff and Von Holle, 1999; Simberloff, 2006; Johnson *et al.*, 2009). On the contrary, there is the possibility that sympatric invasive species will moderate one another's impacts by each controlling the abundance of the other via competitive or predator-prey interactions (Lohrer and Whitlatch, 2002; Alonso and Martínez, 2006; Griffen, Guy and Buck, 2008). The impact of coexisting invaders can also be independent; for example, invasive signal crayfish (*Pacifastacus leniusculus*) and rainbow trout (*Oncorhynchus mykiss*) in Sweden had independent impacts on prey and hence their combined effects on ecosystem structure was additive (Nystrom *et al.*, 2001).

Decapods are extremely successful and disruptive invaders in many aquatic ecosystems (Karatayev et al. 2009). Some crayfish species in particular are widespread, conspicuous invaders with a range of impacts on ecosystem structure and functioning (Capinha, Leung and Anastacio, 2011). They are omnivores and often the largest invertebrates within a food web; omnivory can decouple trophic cascades and alter energy flow and hence, crayfish as invaders have a disproportional impact on food web dynamics (e.g. Lodge et al., 1994; Nyström, Brönmark and Granéli, 1999). Invasive species of crayfish regularly out-compete and replace native crayfish (Hill and Lodge, 1999; Alonso and Martínez, 2006; Dunn et al., 2009; Haddaway et al., 2012) but interactions among invasive crayfish are rarely examined despite the increasing likelihood of them occurring in sympatry. While competition has resulted in serial replacement of invasive crayfish in some instances (Hill and Lodge, 1999), co-existing populations are known (Bernardo et al., 2011; Nakata et al., 2005), but their combined interactive effect on the ecosystem is unknown. Furthermore, research focusing on the impact of invasive crayfish on biodiversity and ecosystem services is becoming increasingly important to inform management decisions as invasions become more widespread (Lodge et al., 2012).

There are seven species of crayfish with established populations at the present time in the United Kingdom, only one of which is native (Holdich, Rogers and Reynolds, 1999; Keller, Zu Ermgassen and Aldridge, 2009). Four of the invaders occur in the Thames catchment (Jackson and Grey, 2012) and yet little, if anything is known of how they might interact. Invasive species can have negative implications for biodiversity and the many valuable ecosystem services provided by freshwater environments (Pejchar and Mooney, 2009). Understanding how invaders are incorporated into communities, and unravelling how interactions among them impact upon

the ecosystem, will underpin our understanding of the whole ecosystem impacts of multiple invasions and, therefore, be valuable in management and policy decisions (Lodge *et al.*, 2012).

Populations of signal and virile (or northern, Orconectes virilis) crayfish occur in separate reaches of the Lee Navigation canal, North East London (Ahern, England and Ellis, 2008) and separate populations of red swamp (Procambarus clarkii) and Turkish (Astacus leptodactylus) crayfish occur in adjacent ponds in Hampstead Heath, North London (Ellis et al., 2012). At the time of study, all four species occurred in separate communities, with no range overlap. However, they have the potential to expand their range in the near future and, given the current close proximity of each crayfish pair in the two separate water bodies (signal and virile in the Lee Navigation and red swamp and Turkish in Hampstead Heath); this could result in sympatric populations of these pairs in the Thames catchment. Therefore, we aim to predict what the outcome of these range expansions might be. We used field data and mesocosm experiments to examine potential and actual interactions between signal and virile crayfish, and between red swamp and Turkish crayfish, to address the question: do interactions among invaders amplify or mitigate one another's impact on ecosystem structure and functioning? We also used stable isotope analyses to examine dietary interactions between the invaders in order to answer the question: do sympatric invasive species occupy a smaller niche than their allopatric counterparts due to potential resource competition?

Materials and methods

Field survey

Using a combination of nitrogen and carbon stable isotope ratios (15N:14N and 13C:12C) it is possible to establish an animal's food sources and trophic level relative to an isotopic baseline (Grey, 2006; Post, 2002). We collected samples of all crayfish species for stable isotope analysis from allopatric populations in the Thames Catchment using crayfish traps baited with fish pellets. Red swamp and Turkish crayfish were collected from ponds on Hampstead Heath in north London (the 'Bird Pond' and 'Hampstead 1', respectively). Signal crayfish and virile crayfish were collected from sites on the Lee Navigation in north east London (Carthagena Lock in Broxbourne and Pickett's Lock in Edmonton, respectively). We sampled 8-15 individuals (Syvaranta et al., 2013) from each population in the summer of 2009 and 2010. Muscle samples were dissected from the tail of each crayfish and oven dried overnight at 60 °C to constant weight before analysis at Queen Mary, University of London using an elemental analyser (Flash EA, 1112 series; Thermo-Finnigan) coupled to a continuous flow mass spectrometer (Finnigan MAT DeltaPlus; Thermo-Finnigan). Ratios of ¹⁵N: ¹⁴N and ¹³C: ¹²C are expressed in parts per mille (‰) using conventional delta notations (δ) relative to international standards (ammonium sulphate and sucrose, of known isotopic composition in relation to atmospheric nitrogen in air (N) and Pee Dee Belemnite (C); Ings et al., 2010).

Following Olsson et al. (2009), we corrected the $\delta^{15}N$ and $\delta^{13}C$ values of the crayfish from the field sites: for trophic position (TP) based on the nitrogen values of long-lived baseline primary consumers (native bivalve molluscs from each site; n=3 to 6); and for $\delta^{13}C$ ($\delta^{13}C_{corr}$) against the carbon values of a representative sample of benthic invertebrates (n=18 to 24),

respectively (see Appendix 1 in the Supporting Information). This allowed us to confidently compare the trophic ecology of each crayfish species over spatial and temporal scales, regardless of variations in the isotopic baseline.

Mesocosm experiment

Two outdoor mesocosm experiments, each of 42 days duration, were conducted during late summer and autumn 2010 using twenty fibreglass ponds (kidney shaped ponds; 0.4 m deep; ~1 m² benthic area). The first experiment examined interactions between signal and virile crayfish; the second, between red swamp and Turkish crayfish. Each mesocosm was prepared as follows: 2 cm depth of clean rock and gravel was added as a substrate and overlaid with 38cm depth of rainwater which was continually aerated by aquaria pumps with air-stones. A halfsection of drainpipe (20 cm length) was added as a crayfish refuge and a stack of three terracotta tiles (10 x 10 cm), each separated by 5mm, was added as an invertebrate refuge. Macrophytes (1 Mentha aquatica and 1 Callitriche sp.) were planted in each mesocosm and freshwater Daphnia, purchased from a local pet shop, were added in equal densities. Each mesocosm was then seeded with equal aliquots of kick samples of benthic invertebrates from the same sites from which the crayfish were sourced (Lee Navigation in experiment one; Hampstead Heath in experiment two) to mimic natural conditions and allowed to establish for 2 week before crayfish were added. Each experiment on each crayfish pair comprised 5 replicates of 4 treatments using a factorial design (i.e. crayfish A, crayfish B, crayfish A+B, no crayfish; Table 1). Each crayfish replicate had an equal density (4 crayfish m⁻²; within the range of signal crayfish densities found in the wild in the UK; Guan, 2000; Bubb, Thom and Lucas, 2004) and biomass of crayfish of the same sex (Table 1).

Dietary analysis

On the final day of each experiment, crayfish were frozen overnight and then thawed before dissecting a portion of muscle from the tail for the analysis of stable isotopes. Samples of leaf litter, macrophytes and invertebrates from each pond were also taken for stable isotope analysis and prepared as above. We were confident that the duration of the experiment was sufficient for isotopic turnover based upon data from controlled feeding experiments on aquatic species, including crayfish, under similar temperature conditions (natural fluctuations between 5 and 17 °C; e.g. Bosley *et al.*, 2002; Gamboa-Delgado *et al.*, 2011; Carolan *et al.*, 2012). However, we also tested for isotopic differences between those individuals caught from survey sites and corresponding individuals from the same survey sites subjected to the 42 days in the mesocosms, and checked for trajectory of change (Grey, Waldron and Hutchinson, 2004; Appendix 2).

Ecosystem structure and functioning

Leaf litter of *Alnus glutinosa* was collected and dried before being used in the experiments to examine decomposition rates. Three plastic mesh bags (aperture 1, 5 and 10 mm) containing a known mass (~ 3 g) of the air-dried leaf litter were fastened to the bottom of each mesocosm at the start of the experiments. Only the 10 mm bags were accessible to crayfish; the 5 mm bags were accessible to other smaller benthic invertebrates and the 1 mm bags were only accessible to microbial organisms. This allowed the direct and indirect impacts of crayfish on leaf litter breakdown to be quantified. The leaf litter remaining in each mesh bag after 6 weeks was washed and then dried at 60 °C to constant weight. The exponential decay rate coefficient (*k*)

was calculated for each treatment as a measure of the rate of leaf litter decomposition following Hieber and Gessner (2002) as:

$$k = \frac{1 \circ g \left(\frac{M_t}{M_0} \right)}{t}$$

where t is the duration of exposure (in days), M_0 is the initial dry mass (in grams) and M_t is the dry mass at time t.

We measured periphytic algal standing stock by placing a terracotta tile (5 x 5 cm) in each mesocosm at the start of the experiment. Tiles were removed on the final day of the experiment and all biofilm was scrubbed, washed off and filtered through GF/C filters (Whatman[®], Maidstone, UK) before adding 15 ml of 90% acetone. After 24 hours in the dark, the samples were centrifuged and the supernatant was used for spectrophotometry. We then used chlorophyll *a* concentration, quantified following Jeffery and Humphrey (1975) and expressed as mg cm⁻² as a measure of periphyton standing stock.

We quantified the benthic invertebrates at the end of the experiment by removing, counting and identifying all organisms in 25 L of filtered water, two 15 cm² sediment cores, leaf packs and invertebrate refugia (5 stacked tiles separated by 5mm each). All invertebrates from each mesocosm were merged as one sample and stored in 70% IMS prior to identification and counting. After identification, benthic invertebrates were assigned to functional feeding groups for further analysis.

Data analyses

To examine the trophic ecology of the crayfish from the field sites, we plotted standard ellipse areas (SEA_c; Jackson *et al.*, 2011; Jackson *et al.*, 2012) based on the distribution of individuals (8-15 per sample) in isotopic space as an estimate of each species core trophic niche using the SIAR package (Jackson *et al.*, 2011; Parnell *et al.*, 2010) in the 'R' computing programme (R Core Development Team 2012). The subscript 'c' indicates that a small sample size correction factor was used (Jackson *et al.*, 2011). Past studies have indicated that a sample size of >5 is appropriate to reveal population niche width using SEA_c (Jackson *et al.*, 2011; Syvaranta *et al.*, 2013).

Analysis of variance (ANOVA; Minitab $14^{\$}$) was used to test for differences in $\delta^{13}C$ and $\delta^{15}N$ between populations of crayfish in each experiment. Discriminant analysis was used to investigate if the species' classification could correctly predict the identity of each individual based on the stable isotope data.

We used the stable isotope-derived population metrics carbon range (CR_b) and nitrogen range (NR_b) as measures of the trophic niche width of crayfish from the mesocosm experiments (Layman *et al.*, 2007; Jackson *et al.*, 2011; Jackson *et al.*, 2012). CR_b and NR_b were calculated in the statistical package 'R' (R Core Development Team 2012) as the distance between the individuals with the highest and lowest $\delta^{13}C$ and $\delta^{15}N$ values, respectively. To enable comparison among variable sample sizes (3 individuals in the treatments with 2 species of crayfish and 6 individuals in the treatments with allopatric crayfish), the metrics were bootstrapped (n = 10000; indicated with a subscript 'b') based on the minimum sample size of 3. ANOVA was then used to test for difference in CR_b and NR_b between populations of crayfish. We used permutational multivariate analysis of variance (PERMANOVA; Anderson, 2001) to

test for effects of the experimental manipulations on the structure of benthic invertebrate assemblages using the PERMANOVA+ add-in to PRIMER® version 6.1 (PRIMER-E Ltd, Plymouth, UK). These analyses were done with 9999 permutations of the residuals under a reduced model (Anderson, Gorley and Clarke, 2008) and were based on Bray-Curtis similarity matrices calculated from log (X + 1)-transformed data. Similarity percentages analysis (SIMPER in PRIMER®; Clark and Warwick, 2001) was then used to determine the contribution of benthic invertebrate taxa to the mean Bray-Curtis dissimilarities between treatments using presence or absence of each crayfish species as factors. This method determines which taxa were affected most strongly by the presence of each species of crayfish in each experiment.

We used ANOVA to test for an effect of treatment on leaf litter breakdown, algal standing stock, benthic invertebrate richness and the total abundance of each benthic invertebrate functional feeding group. The Student-Newman-Keuls procedure was used following all ANOVA tests to make *post-hoc* comparisons among levels of significant terms. All analyses were balanced and variables were transformed where necessary prior to analysis to homogenise variances.

Finally, for clarity and to allow comparison between experiments, we calculated the standardised effect size of each treatment on each variable compared to the relevant experimental control, which had no crayfish. We used Hedges' d as a measure of effect size (Gurevitch, Morrison and Hedges, 2000); in our analysis, the effect size estimates the standardized mean difference between each treatment and the control for each variable we quantified (invertebrate abundance and richness, periphyton standing stock and decomposition) and we used it to highlight similarities and discrepancies in the effect of each invasive species on the ecosystem

(Sanders *et al.*, 2003). For each mean response variable (X) the individual effect size d of each crayfish treatment compared to the crayfish free control was calculated as:

$$\frac{X_i - X_u}{S}j$$

where j is a weighting factor based on the number of replicates (n) per treatment, calculated as:

$$1 - \frac{3}{4(n_i + n_u - 2) - 1}$$

and S is the pooled sample variation, calculated as:

$$\sqrt{\frac{(n_i - 1)\sigma_i^2 + (n_u - 1)\sigma_u^2}{n_i + n_u - 2}}$$

Finally, variance of d(Vd) was calculated as:

$$\frac{n_i + n_u}{n_i n_u} + \frac{d^2}{2(n_i + n_u)}$$

Results

Field survey

The core niche (SEA_c) of each invasive crayfish population from the field sites did not overlap in isotopic space suggesting niche partitioning between species (Fig. 2). The field survey revealed that virile crayfish had the largest dietary niche in both years (2009: 0.189 $\%^2$; 2010: 0.129 $\%^2$) followed by signal (2009: 0.076 $\%^2$; 2010: 0.055 $\%^2$), Turkish (2009: 0.036 $\%^2$; 2010: 0.037 $\%^2$) and red swamp crayfish (2009: 0.034 $\%^2$; 2010: 0.022 $\%^2$; Fig. 2).

Dietary interactions

In the mesocosms, the $\delta^{15}N$ was significantly lower in populations of signal compared to virile crayfish (F_{3,80} = 31.93, P < 0.001, Figure 1a), and Turkish compared to red swamp crayfish (F_{3,77} = 42.07, P < 0.001, Figure 1b) suggesting a degree of niche partitioning across trophic levels. Sympatric virile crayfish also had a significantly higher $\delta^{15}N$ than their allopatric counterparts (F_{3,80} = 31.93, P < 0.001; Figure 1a). The $\delta^{13}C$ was significantly lower in populations of allopatric signal crayfish compared to virile crayfish (F_{3,80} = 6.06, P = 0.001; Figure 1a) and red swamp crayfish compared to Turkish crayfish (F_{3,77} = 19.76, P < 0.001; Figure 1b), indicating niche partitioning across resources. Discrimination analysis correctly classified 88.1 % of signal crayfish (37 of 42), 83.3 % of virile crayfish (35 of 42), 90.48 % of red swamp crayfish (38 of 42) and 93.31% of Turkish crayfish (36 of 39) suggesting each species occupied a distinct area in isotopic space. All crayfish had changed isotopically during the experiment, assuming the field survey isotope values as a starting point, and in differing directions (Appendix 2).

Nitrogen and carbon range, both measures of isotopic niche width, were highest in virile and red swamp crayfish, respectively. However, niche width did not vary significantly between allopatric and sympatric populations of the same species (see Figure S1 in Supporting Information).

Assemblage composition

The benthic invertebrate community structure differed significantly between treatments in the signal and virile experiment (Pseudo- $F_{3,16} = 2.30$, P = 0.01). In both experiments, the

assemblage was the most similar in treatments with crayfish (average similarity between 65.2% and 76.6%) and, therefore, the control treatments were the most distinct. The same five taxa were affected the most by signal or virile crayfish and, in total they contributed almost 50% to the dissimilarity in the community assemblage (Table 2). Variation in just three taxa explained more than 50% of the dissimilarity in the community assemblage between both red swamp and Turkish crayfish absence or presence (Table 3).

The total abundance of grazers and scrapers was reduced in all crayfish treatments compared to the controls (experiment 1: $F_{3,16} = 2.72$, P = 0.079; experiment 2: $F_{3,16} = 4.75$, P = 0.015; Figure S2, Figure S3); the effect size was largest in treatments containing Turkish and red swamp crayfish (Figure 3a). Abundance of gatherers and shredders did not differ significantly between treatments in both experiments (Figure S2, Figure S3); however, the largest effect we recorded was a decline in abundance in the allopatric Turkish and virile treatments (Figure 3b). The largest effect on predator and carnivorous scavenger abundance was in the sympatric red swamp and Turkish crayfish treatment (Figure 3C), where abundance was significantly lower compared to the control ($F_{3,16} = 3.06$, P = 0.05; Figure S3). Signal and virile crayfish treatments also had significantly lower numbers of predators and carnivorous scavengers ($F_{3,16} = 8.78$, P = 0.001; Figure S2, Figure 3c) and Diptera ($F_{3,16} = 8.78$, P = 0.001; Figure S2, Figure 3d) compared to the control. Finally, benthic invertebrate taxon richness was highest in the treatment with no crayfish compared to all virile and signal crayfish treatments ($F_{3,16} = 7.43$, P = 0.002, Figure 3e).

Algal standing stock

Periphyton standing stock was significantly higher in the treatment with allopatric red swamp crayfish compared to the control ($F_{3,16} = 4.5$, P = 0.018; Figure 4a, Figure S4).

Leaf litter decomposition

Microbial leaf litter decomposition was reduced in all crayfish treatments compared to controls (Figure 4b); however, the effect was only significant in both treatments with Turkish crayfish present ($F_{3,16} = 6.86$, P = 0.003; Figure S5). In bags with a mesh aperture of 10mm, which allowed access by crayfish and other invertebrates, leaf litter decomposition rates were reduced in the allopatric virile and Turkish treatments and elevated in the allopatric signal and red swamp treatments (Figure 4d). This effect was significant in the allopatric virile treatment compared to the control and allopatric signal treatment ($F_{3,16} = 16.95$, P < 0.001; Figure S5); and in the allopatric Turkish treatment compared to the allopatric red swamp treatment ($F_{3,16} = 3.68$, P = 0.034; Figure S5). Leaf litter decomposition by smaller invertebrates (i.e. in 5mm aperture bags not accessible to crayfish) was reduced in all treatments bar those with signal crayfish present (Figure 4c). In these bags, decomposition rates were significantly lower in the control and virile crayfish treatments compared with the allopatric signal treatment ($F_{3,16} = 9.24$, P = 0.001; Figure S5); and in the allopatric Turkish treatment compared with the control ($F_{3,16} = 3.75$, P = 0.033; Figure S5).

Discussion

Our results demonstrate that multi-species crayfish invasions have different ecosystem level impacts compared to single-species invasions. Despite having generalist and omnivorous diets, we found dietary niche partitioning between each crayfish species, resulting in differences in their independent impacts on ecosystem structure and functioning. Niche partitioning suggests that interspecific competition will be low and, therefore, the niche width should be consistent between allopatric and sympatric populations (Gilbert, Srivastava and Kirby, 2008; Levine and HilleRisLambers, 2009). Accordingly, we found no evidence that crayfish niche width was reduced in the presence of a potential competitor. However, virile crayfish had significantly higher δ^{15} N, reflecting a higher trophic level, in the presence of signal crayfish, indicating that competitive interactions may have caused a shift in their diet.

These differences in diet preference expressed by each invasive crayfish species, consequently affected ecosystem functioning through trophic cascades to differing degrees (Carpenter *et al.*, 1987). Similarly, Rudnick and Resh (2005) found that differences in the diet of invasive red swamp crayfish and Chinese mitten crabs (*Eriocheir sinensis*) resulted in variation in their impacts on the ecosystem. We found that slight diet variation between crayfish species triggered opposing impacts on the ecosystem in both autochthonous- and allochthonous-based food chains. Red swamp crayfish had the largest negative effect on invertebrate grazer and scraper abundance (particularly on snails from the genus Lymnaea) and subsequently, were the only species to promote periphyton standing stock (see schematic in Figure 5). Such trophic cascades through the autochthonous-based food chain involving a reduction in grazing pressure (Charlebois and Lamberti, 1996; Nyström *et al.*, 1999) may have ramifications for whole

ecosystem functioning and alter primary productivity (Morin, Lamoureux and Busnarda, 1999; Carpenter *et al.*, 1987; Jones and Sayer, 2003).

Crayfish simultaneously affect intermediate consumers and their basal resources because they are omnivores (Usio, 2000). In our experiments, all four invasive species independently altered decomposition rates by two opposing mechanisms, causing variation in allochthonous leaf litter availability (see schematic in Figure 5). The three invertebrate taxa most affected by virile crayfish were all shredders or gatherers. In fact, both virile and Turkish crayfish had the greatest negative effect on invertebrate gatherer and shredder abundance which, via a trophic cascade, reduced leaf litter decomposition. In stark contrast, red swamp and signal crayfish promoted leaf litter decomposition by consuming leaf litter directly (reflected in their δ^{13} C values more closely associated with leaf litter) in preference to associated invertebrate prey, which decoupled the trophic cascade. Since Turkish and virile crayfish had an opposite effect on net decomposition rates to red swamp and signal crayfish, we observed an intermediate effect on net decomposition rates in the sympatric treatments (multi-species invasion scenarios), which were indistinguishable from the controls. This suggests that the impact of multi-species crayfish invasions will be the sum of their independent impacts. Therefore, if two sympatric invasive species have similar independent impacts on the ecosystem (i.e. red swamp and signal crayfish both elevate decomposition rates), their combined effect might be additive. For example, invasive rusty crayfish (Orconectes rusticus) and Chinese mystery snails (Bellamya chinensis) both independently reduce native snail biomass by consumption and competitive interactions, respectively; eliminating one species of native snail in the presence of both invaders (Johnson et al., 2009).

There was also evidence of an amplified outcome of interactions among invaders. Invertebrate predator and carnivorous scavenger abundance was reduced in the presence of signal and/or virile crayfish, consistent with an *in situ* experiment by Stenroth and Nyström (2003) on signal crayfish in Sweden. However, red swamp and Turkish crayfish had no independent effect and, instead, only reduced predator and carnivorous scavenger abundance in multi-species invasion scenarios. This indicates an amplified impact whereby the presence of both species synergistically increased competition (McCarthy *et al.*, 2006) and/or predator-prey links (Stenroth and Nyström, 2003) with native invertebrate predators resulting in the largest observed negative effect on their abundance (Figure 3c). There was also variation in the impact of invasive crayfish species on Dipteran larvae; abundance was reduced in the presence of signal and virile crayfish only, suggesting direct consumption had a larger effect than the positive effect of sediment sorting by crayfish (Usio and Townsend, 2004). Finally, only virile and signal crayfish treatments had lower taxon richness than the treatments with no crayfish, consistent with other studies on signal crayfish (e.g. Stenroth and Nyström, 2003).

Lodge *et al.* (2012) recently made a call for more research on the impact of crayfish invasions; here, we have shown how multi-species crayfish invaders interact to impact ecosystem structure and functioning. Some variation in the independent effects of invasive crayfish had implications for their combined synergistic impact, resulting in amplified, intermediate and additive effects on the ecosystem. Our results have serious implications for conservation of biodiversity and management of invasive species by indicating that multiple crayfish species are able to coexist due to resource partitioning, which could promote higher densities in sympatric populations (Siepielski *et al.*, 2011). Crayfish density is positively correlated with the magnitude of its impact on ecosystem structure and functioning (Parkyn, Rabeni and Collier, 1997) and,

therefore, we predict that multi-species crayfish invasions will have an amplified effect on the ecosystem as a result of higher total crayfish densities.

Acknowledgements

We are grateful to the Environment Agency for funding and to numerous colleagues for assistance, particularly Paul Fletcher. We are also indebted to Adrian Brooker and his colleagues at Hampstead Heath.

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Tables

Table 1. The replicated (n=5) treatments used in each experiment. The numbers in parentheses represent the number of individual crayfish used from each species in each replicate followed by the total biomass of those individuals. Crayfish abbreviations are as follows: red swamp crayfish, RSC; Turkish crayfish, TC; signal crayfish, SC and virile crayfish, VC.

Experiment 1	Experiment 2	
SC (6; 171 ± 10g)	RSC (6; 224 ± 10g)	
VC (6; 171 ± 10g)	TC $(6; 224 \pm 10g)$	
SC (3; $85 \pm 5g$) and VC (3; $85 \pm 5g$)	RSC (3; $112 \pm 5g$) and TC (3; $112 \pm 5g$)	
Neither species (0; 0)	Neither species (0; 0)	

Table 2. Results of SIMPER analysis indicating the primary benthic invertebrate taxa affected by the presence of signal (SC) and virile crayfish (VC). Mean relative abundance (\pm standard error) is displayed using the raw data for clarity however, the analysis was performed on Log(x+1)-transformed data. Please note, standard errors will be high since each mean covers two treatments; for instance abundances in SC absence will be an average of the neither species treatment and the VC only treatment.

Taxon	Mean abundance in SC absence	Mean abundance in SC presence	Contribution to dissimilarity (%)	Cumulative contribution (%)
Chironomidae	664.0 ± 125.0	193.0 ± 26.9	10.0	10
Limnephilidae	4.9 ± 1.1	12.5 ± 4.8	9.0	19.0
Corophiidae	10.1 ± 1.8	14.1 ± 4.2	8.8	27.8
Lymnaeidae	7.8 ± 3.2	3.0 ± 0.6	7.7	35.4
Gammaridae	5.0 ± 1.3	7.1 ± 2.7	7.47	42.9
Taxon	Mean abundance in VC absence	Mean abundance in VC presence	Contribution to dissimilarity (%)	Cumulative contribution (%)
Corophiidae	543.0 ± 149.0	314.0 ± 57.8	12.2	12.2
Limnephilidae	9.6 ± 3.6	7.0 ± 3.5	9.4	21.6
Gammaridae	16.0 ± 3.7	6.3 ± 1.3	9.0	30.6
Lymnaeidae	7.3 ± 3.2	5.8 ± 1.0	7.4	38.0
Chironomidae	7.7 ± 2.5	3.5 ± 0.6	6.5	44.5

Table 3. Results of SIMPER analysis indicating the primary benthic invertebrate taxa affected by the presence of red swamp (RSC) and Turkish crayfish (TC). Mean relative abundance (\pm standard error) is displayed using the raw data for clarity however, the analysis was performed on Log(x+1)-transformed data. Please note, standard errors will be high since each mean covers two treatments, for instance abundances in RSC absence will be an average of the neither species treatment and the TC only treatment.

Taxon	Mean abundance in RSC absence	Mean abundance in RSC presence	Contribution to dissimilarity (%)	Cumulative contribution (%)
Lymnaeidae Chironomidae Asellidae Turbellaria Oligochaeta	62.0 ± 90.7 157.0 ± 95.4 17.6 ± 21.7 9.9 ± 13.0 2.8 ± 4.21	11.7 ± 13.3 196.3 ± 106.1 14.5 ± 14.3 2.8 ± 6.1 1.6 ± 1.2	20.1 19.8 11.9 9.3 5.4	20.1 39.9 51.7 61.0 66.4
Taxon	Mean abundance in TC absence	Mean abundance in TC presence	Contribution to dissimilarity (%)	Cumulative contribution (%)
Chironomidae Lymnaeidae Asellidae Turbellaria Dytiscidae	175.1 ± 121.7 64.2 ± 89.8 19.6 ± 22.2 10.2 ± 13.23 2.1 ± 2.7	165.8 ± 79.9 11.0 ± 9.3 15.5 ± 12.7 3.7 ± 5.2 2.5 ± 3.5	20.6 20.2 12.1 9.7 5.3	20.6 40.1 52.9 62.5 67.8

Figure legends

- **Figure 1.** Stable isotope bi-plot of the mesocosm food webs. Closed squares present the average isotopic signature of resources from all ponds (mean \pm standard error).
- (a) Experiment 1; filled symbols represent individual signal crayfish and open symbols represent individual virile crayfish in allopatric (circles) and sympatric (triangles) populations.
- (b) Experiment 2; filled symbols represent individual red swamp crayfish and open symbols represent individual Turkish crayfish in allopatric (circles) and sympatric (triangles) populations.

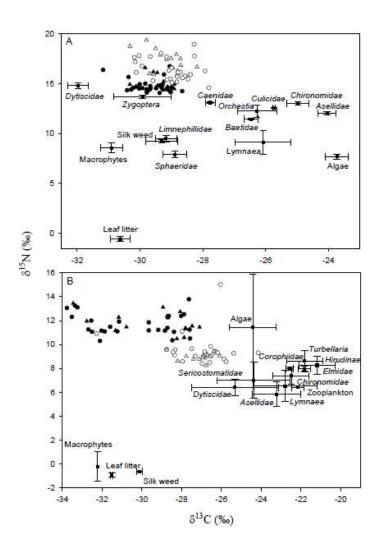


Figure 2. Isotopic bi-plot of trophic position (TP) and corrected δ^{13} C values (δ^{13} C_{corr}). Each ellipse encloses the core niche width (SEA_c) of signal (solid black), virile (dashed black), red swamp (solid grey) and Turkish (dashed grey) crayfish from the field sites in 2009 (a) and 2010 (b).

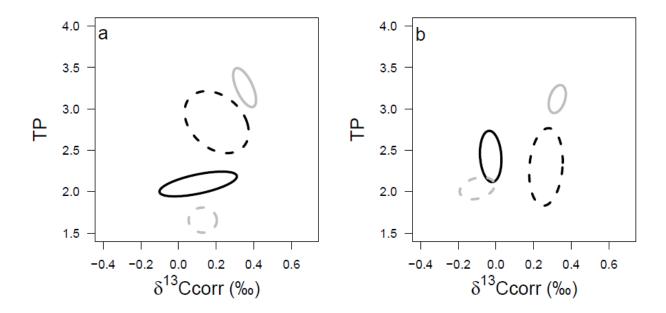


Figure 3. The effect size (Hedges d) of each variable in each treatment compared to the experimental control which had no crayfish. A positive d indicates an increase; negative d a decrease, in abundance or species richness relative to controls. Effects are statistically significant (P < 0.05) if confidence limits do not overlap. Gatherers and shredders (a), grazers and scrapers (b), predators and carnivorous scavengers (c), Diptera (d), taxon richness (e).

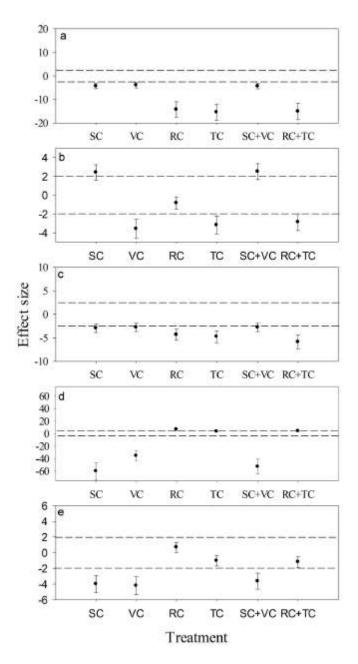


Figure 4. The effect size (Hedges d) of each variable in each treatment compared to the experimental control which had no crayfish. A positive d indicates an increase; negative d a decrease, in periphyton standing stock or decomposition relative to controls. Effects are statistically significant (P < 0.05) if confidence limits do not overlap. Periphyton standing stock (a), microbial decomposition (b), benthic invertebrate decomposition (c), crayfish decomposition (d).

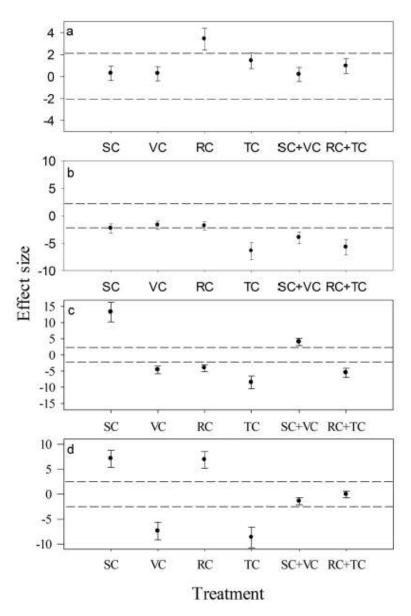


Figure 5. A schematic diagram illustrating the trophic cascades instigated by invasive crayfish in the experimental mesocosms. White arrows depict the direction of the cascade and grey arrows indicate the effect of the trophic cascade on biomass compared to the control with no crayfish. Crayfish caused a decline in grazer abundance and, therefore reduced grazing pressure on algae. In the red swamp crayfish treatments, this resulted in elevated benthic chlorophyll-a concentrations compared to the control (i). The crayfish also consumed invertebrates that shred and consume leaf litter; this impact was most evident in the virile and Turkish treatments and resulted in a decline in net decomposition rates (i.e. an increase in leaf litter biomass; ii). However, because crayfish are omnivores and will also consume leaf litter directly, they often decouple this trophic cascade resulting in elevated leaf litter loss and reduced leaf litter biomass; this effect was apparent in the signal and red swamp crayfish treatments (iii).

