

Improving *in vitro* propagation of *Protea cynaroides* L. (King Protea) and the
roles of starch and phenolic compounds in the rooting of cuttings

by

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Protea cynaroides (Photograph: G. Bredenkamp)

DECLARATION

I hereby certify that this thesis is my own work, except where duly acknowledged. I declare that this thesis that I hereby submit at the University of Pretoria has not previously been submitted by me for degree purposes at any other university.

Signature:.....

(How-Chiun Wu)

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LIST OF ABBREVIATIONS

2,4-D	:	2,4-Dichlorophenoxyacetic acid
2iP	:	N ⁶ -(2-isopentyl)adenine
ABA	:	Abscisic acid
AND	:	Anderson (1975) medium
BA	:	Benzyladenine
BAP	:	6-Benzylaminopurine
GA ₃	:	Gibberellic acid
IAA	:	3-indolyl-acetic acid
IBA	:	3-indolebutyric acid
MS medium	:	Murashige and Skoog (1962) medium
NAA	:	1-Naphthalene acetic acid
NOA	:	2-Naphthylloxyacetic acid
TDZ	:	Thidiazuron
WPM	:	Woody plant medium (Loyd and McCown, 1981)
IEDC	:	Induced embryogenic determined cells
PEDC	:	Pre-embryogenic determined cells
CARD	:	Curve-fitting to allelochemical response data
PAR	:	Photosynthetic active radiation
PEG	:	Polyethylene glycol
mOsm.kg ⁻¹	:	MilliOsmol kilogram ⁻¹
ACN	:	Acetonitrile
HPFC	:	High performance flash chromatography
HPLC	:	High performance liquid chromatography
NMR	:	Nuclear magnetic resonance
MS	:	Mass spectrophotometer
TLC	:	Thin layer chromatography
UV	:	Ultraviolet

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Improving *in vitro* propagation of *Protea cynaroides* L. (King Protea) and the roles of starch and phenolic compounds in the rooting of cuttings

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Abstract

Protea cynaroides L. (King Protea) is a well known cutflower. Seeds and stem cuttings are commonly used to propagate *P. cynaroides*. However, the success rate and rooting rate of seeds and cuttings, are inconsistent and slow. The potential of *in vitro* propagation as an alternative method to produce *P. cynaroides* plantlets was investigated. *In vitro* studies consisted of *in vitro* germination of mature zygotic embryos, micrografting and direct somatic embryogenesis of zygotic embryos and excised cotyledons. In the germination study, temperature was the most important factor in obtaining a high germination percentage. Alternating temperatures of $21\pm 2^{\circ}\text{C}/12\pm 2^{\circ}\text{C}$ (light/dark) was suitable for germination and over 90% of embryos germinated, while the germination percentage of embryos at $25\pm 2^{\circ}\text{C}$ was poor. Plantlets were successfully established in *ex vitro* conditions when planted in a peat/coir/sand mixture. Micrografting of *P. cynaroides* was done by grafting microshoots (microscion), which was taken from *in-vitro*-established nodal explants, onto roots of decapitated *in-vitro*-germinated seedlings. After the graft union formed, buds on the microscion sprouted. A protocol to induce direct somatic embryogenesis was developed. Direct somatic embryogenesis was achieved on both *P. cynaroides* mature zygotic embryos and excised cotyledons. The addition of auxins such as NAA and 2,4-D singly or in combination with TDZ, BAP or kinetin suppressed the formation of somatic embryos. Formation of somatic embryos was observed in medium lacking growth regulators. Germination of somatic embryos was highest in medium containing GA_3 . The roles of starch and phenolic compounds in the rooting of *P. cynaroides* cuttings were also studied. Starch and total soluble phenol analyses

results revealed a positive correlation between high root formation and increased starch and phenolic content. NMR and MS analyses identified high amounts of 3,4-dihydroxybenzoic acid in stems of *P. cynaroides*. *In vitro* bioassay showed that 3,4-dihydroxybenzoic acid stimulated and inhibited root growth of *P. cynaroides* explants, depending on the concentration. A link was made between the endogenous concentration levels of 3,4-dihydroxybenzoic acid and rooting of *P. cynaroides* stem cuttings. Findings of this study contribute towards a better understanding of the roles starch and phenolic compounds play in the rooting of *P. cynaroides*.

Keywords: *Protea cynaroides*, *in vitro* germination, micrografting, somatic embryogenesis, phenolic compounds, starch

INTRODUCTION

Protea cynaroides (King Protea) is a multi-stemmed, upright shrub that grows to between 0.3 and 2 m tall. It has sparse branches, with hairless stems (Rebelo, 2000). The leaves are round, oval or narrowly elliptic, ranging from 50 to 120 mm in length and 50 to 75 mm in width. The flowerhead sizes range from 120 mm to 300 mm in diameter and the colour of the bracts, which are either hairy or hairless, range from pink to creamy-white (Patterson-Jones, 2000).

The King Protea is the national flower of South Africa. They are widely spread throughout the south-western and southern parts of South Africa. Its magnificent inflorescence is a well known cutflower in many parts of the world. The most common methods of propagation are by seeds and stem cuttings. However, propagation by both seeds and stem cuttings have limitations in large-scale commercial production. Seed germination is usually inconsistent, even when the seed is treated with seed primers. However, the main problem with plants derived from seeds is genetic variation. This is particularly problematic when uniform blooms of a specific cultivar are highly sought after in the market place. Vegetative propagation of protea cuttings has become more common, nevertheless, this has its own limitations. The difficulty in inducing quick and consistent rooting of stem cuttings has not been overcome. At the moment, *P. cynaroides* cuttings take four to six months to root in the mistbed, prior to being transplanted to the field. After transplanting, it takes several years for the first high quality flower to be produced, which makes it an expensive flower to produce. Currently, some flowers are still picked from the wild, which firstly, do not always adhere to international standards, and secondly, cannot maintain a consistent flow of quality floral products to the floriculture industry (Coetzee, 2000).

Even with the abovementioned problems in the propagation of *P. cynaroides*, the cultivation of proteas in general has gradually increased over the years, mainly through the increase in area being planted. Although the majority is grown in the southern hemisphere, cultivation areas in the northern hemisphere have increased from 800 ha in 2000 to 900 ha in 2004 (Anonymous, 2005). South Africa is the world

leader regarding the total area of Proteaceae grown, which in 2004 was 3,853 ha, of which 2795 ha was broadcast sown. This is followed by Australia with an estimated 1,230 ha, while in the northern hemisphere, California leads the way with approximately 405 ha, followed by Israel with 270 ha. The amount of fresh Proteaceae flowers exported by South Africa has steadily increased from approximately 2,100 tons in the early nineties to over 4,200 tons in 2004 (Anonymous, 2005).

Very few studies on the *in vitro* propagation of *P. cynaroides* have been reported. Nevertheless, the establishment (Ben-Jaacov and Jacobs, 1986) and multiplication (Wu, 2001) of *P. cynaroides* explants have been investigated, where explants were successfully cultured *in vitro*. *In vitro* rooting of these explants were however, not successful. Furthermore, the use of other *in vitro* propagation methods, such as somatic embryogenesis and zygote culture to obtain rooted plantlets has not been reported.

In numerous studies, reviewed in Chapter 1, it has been shown that micropropagation techniques, whether through *in vitro* embryo culture, somatic embryogenesis, organogenesis or micrografting, can overcome common problems such as slow germination, low regeneration rate and poor rooting capacity of various plant species. The main objective of the research reported in this thesis was to introduce alternative propagation methods for *P. cynaroides*, which included *in vitro* culture of zygotic embryos, micrografting and somatic embryogenesis. These *in vitro* propagation practices could ultimately be used in practice to breed and mass propagate one of the most valuable *Protea* species. Furthermore, the aim of this study was to contribute new knowledge towards the understanding of the roles of starch and phenolic compounds in the root formation of *P. cynaroides*, through allelopathy and biochemical studies.