

Meinecke, C. D., De Vos, L., Yilmaz, N., Steenkamp, E. T., Wingfield, M. J., Wingfield, B. D., Villari, C. 2022. A LAMP assay for rapid detection of the pitch canker pathogen *Fusarium circinatum*. Plant Dis.

Assessing *Fusarium circinatum* qPCR and LAMP assays for cross-reactivity with *F. pilosicola* DNA.

Materials and methods

Reactions using purified DNA

To assess whether other available *F. circinatum* molecular tests cross-react with the newly described *F. pilosicola*, we screened *F. pilosicola* DNA using the dual-labelled probe qPCR method developed by Iosifidis *et al.* (2009) and the probe-based LAMP method developed by Stehlíková *et al.* (2020).

The qPCR reaction mix (final volume of 20 μ L) consisted of 5.0 μ L of molecular grade water, 10 μ L of GoTaq[®] Probe qPCR Master Mix (Promega, Madison, Wisconsin, USA), 1.0 μ L of each FCIR-F and FCIR-R primers (10 μ M) and 1.0 μ L of the FCIR-P dual-labeled probe (10 μ M). The LAMP reaction (final volume 25 μ L) was prepared as described by Stehlíková *et al.* (2020), using identical reagents, primers, and reaction conditions. The DNA of *F. pilosicola* isolates CMWF 1183 and CMWF 1189 was tested by each assay. Each qPCR and LAMP run included the DNA of *F. circinatum* isolate CV-2020-006 (Table S1) as a positive control and two non-template negative controls. Each test of *F. pilosicola* or *F. circinatum* DNA was run in triplicate within the same run. All qPCR and LAMP reaction mixtures were assembled in eight-tube MicroAmp[®] Fast Reaction Tubes strips (Applied Biosystems TM).

All reactions were run on a StepOnePlus Real-Time PCR System (Applied Biosystems™, Foster City, California, USA). Reactions were run as described in loos *et al.* (2009) and Stehlíková *et al.* (2020) for the qPCR and LAMP tests, respectively.

Results

Reactions using purified DNA

Both isolates of the recently described *F. pilosicola* and the *F. circinatum* positive control tested positive by the previously published LAMP assay, whereas only *F. circinatum* DNA was amplified by the qPCR test (Table S1). The negative controls did not amplify.

Pathogenicity testing of *Fusarium pilosicola* on *Pinus* seedlings

Materials and methods

Inoculations

Nineteen-month-old *Pinus patula* x *Pinus tecunumanii* plants were housed at 25°C under the ambient day/night cycle. Plant stems were inoculated using a 10 mm cork borer to remove the bark, inserting an agar plug into the opening, and sealing the wound with Parafilm. Fungal isolates were grown on PDA at 25°C for 7 days. Trees were either inoculated with one of the only two known isolates of *Fusarium pilosicola* (isolates CMWF1183 and CMWF1189), *F. circinatum* (isolate FSP34) as a positive control, or sterile PDA as a negative control. Each treatment was applied to 26 trees. After 28 days, the bark surrounding the inoculation site was removed and the length of the lesion measured. Lesion lengths were compared between treatments using the Student's t-test.

Results

Inoculations

Neither isolate of *F. pilosicola* caused the formation of lesions on the pine seedlings ($P > 0.05$). *Fusarium circinatum* induced lesions that were significantly longer than those observed in the negative control and *F. pilosicola* inoculations ($P < 0.05$, Figure 1). Under the conditions tested, *F. pilosicola* is not virulent towards *Pinus patula* x *Pinus tecunumanii* seedlings.

Figure S1. Comparison of pathogenicity of *F. pilosicola* CMWF1183 and CMWF1189 and *F. circinatum* FSP34. Average lesion length was measured 28 days after inoculation on *P. patula* x *P. tecunumanii* seedlings. Error bars represent the standard deviation.

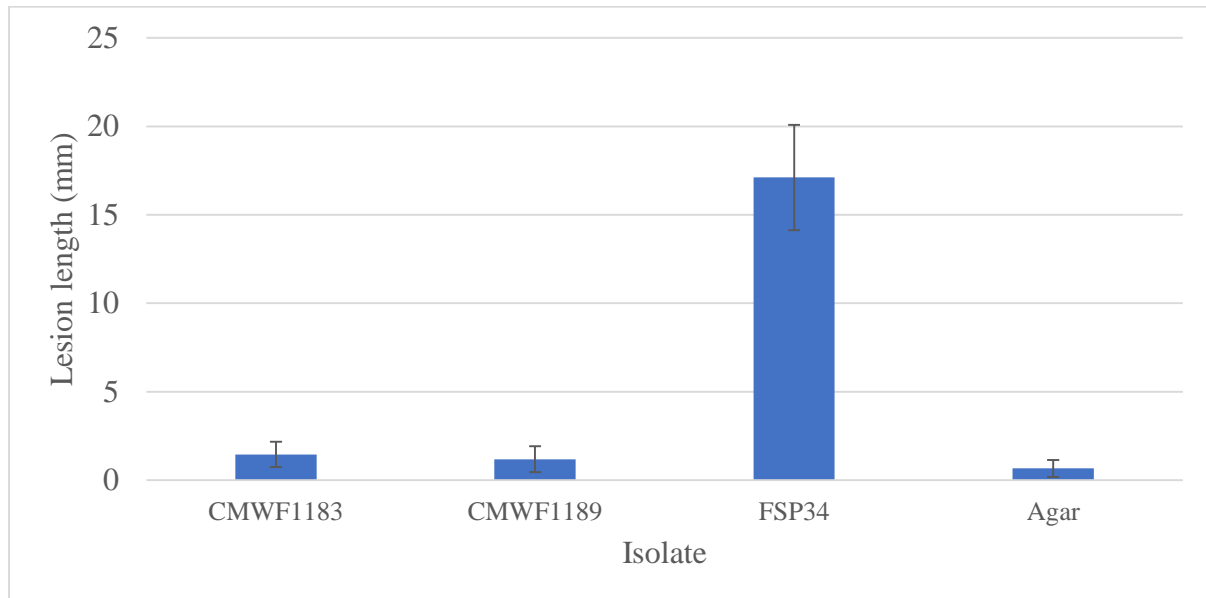


Table S1. Fungal isolates used to test available *Fusarium circinatum* qPCR and LAMP assays for cross-reactivity with the recently described *F. pilosicola*.

Fungal species	Collection Number ^a	Host Species	Country	qPCR result ^b	LAMP result
<i>Fusarium circinatum</i>	CV_2020_006	<i>Pinus taeda</i>	United States	3/3 40/26.41	3/3 13.6m
<i>Fusarium pilosicola</i>	CMWF 1183 = NRRL 29124 ^T	<i>Bidens pilosa</i>	United States	0/3	3/3 10.0m
<i>Fusarium pilosicola</i>	CMWF 1189 = NRRL 29123	<i>Bidens pilosa</i>	United States	0/3	3/3 10.3m

^a Abbreviations for the culture collections: the culture collection of Dr. Villari (CV) at the D.B. Warnell School of Forestry, University of Georgia, United States; the working collection of Fusarium isolates at FABI (CMWF) of the Forestry and Agricultural Biotechnology Institute (FABI), University of Pretoria, South Africa.

^b Positive qPCR and LAMP results were determined by amplification of at least 2 of 3 replicate reactions for each sample. Average reaction time is given (min for LAMP and min/Ct for qPCR, total reaction time 60 minutes). Positive results are indicated in bold.

^T Ex-type specimen

Table S2. Percent matches of each core LAMP primer binding site to the genomes of 50 *Fusarium* spp., representing the *Fusarium fujikuroi* species complex. Similarity of each primer binding sequence to genomic DNA was determined by BLAST comparison.

Species	Phylogeographic clade	Accession number	Isolate number	F3	B3	FIP	BIP
<i>Fusarium acutatum</i>	African A	GCA_012932015.1	NRRL 13308	0%	0%	0%	0%
<i>Fusarium andiyazi</i>	African A	ERX4077522	CBS 119857	0%	0%	0%	0%
<i>Fusarium brevicatenulatum</i>	African A	GCA_013363135.1	NRRL 25447	0%	0%	0%	0%
<i>Fusarium denticulatum</i>	African A	GCA_013396175.1	NRRL 25311	0%	0%	0%	0%
<i>Fusarium ficicrescens</i>	African A	ERX4078212	CBS 125178	0%	0%	0%	0%
<i>Fusarium lactis</i>	African A	ERX4077573	CBS 411.97	0%	0%	0%	0%
<i>Fusarium mundagurra</i>	African A	GCA_013396205.1	NRRL 66235	0%	0%	0%	0%
<i>Fusarium musae</i>	African A	GCA_019915245.1	F31	0%	0%	0%	0%
<i>Fusarium napiforme</i>	African A	GCA_013396005.1	NRRL 25196	0%	0%	0%	0%
<i>Fusarium nygamai</i>	African A	GCA_001262555.1	NRRL 66327	0%	0%	0%	0%
<i>Fusarium phyllophilum</i>	African A	GCA_013396025.1	NRRL 13617	0%	0%	0%	0%
<i>Fusarium pseudoanthophilum</i>	African A	GCA_013395995.1	NRRL 25211	0%	0%	0%	0%
<i>Fusarium pseudocircinatum</i>	African A	GCA_013396035.1	NRRL 36939	0%	0%	0%	0%
<i>Fusarium pseudonygamai</i>	African A	GCA_013186785.1	NRRL 13592	0%	0%	0%	0%
<i>Fusarium ramigenum</i>	African A	GCA_013186855.1	NRRL 25208	0%	0%	0%	0%
<i>Fusarium secorum</i>	African A	GCA_013363185.1	NRRL 62593	0%	0%	0%	0%
<i>Fusarium thapsinum</i>	African A	GCA_013186935.1	NRRL 22049	0%	0%	0%	0%
<i>Fusarium tjaetaba</i>	African A	GCA_013396195.1	NRRL 66243	0%	0%	0%	0%
<i>Fusarium udum</i>	African A	GCA_013186905.1	NRRL 25194	0%	0%	0%	0%
<i>Fusarium verticillioides</i>	African A	GCA_000149555.1	7600	0%	0%	0%	0%
<i>Fusarium xylarioides</i>	African A	GCA_013183765.1	KSU18978	0%	0%	0%	0%
<i>Fusarium xyrophilum</i>	African A	GCA_008711595.1	NRRL 62721	0%	0%	0%	0%
<i>Fusarium dlamini</i>	African B	GCA_013186775.1	NRRL 13164	0%	0%	0%	0%
<i>Fusarium agapanthi</i>	American	GCA_001654545.1	NRRL 54464	0%	0%	0%	0%
<i>Fusarium ananatum</i>	American	ERX4077703	CBS 118516	0%	0%	0%	0%
<i>Fusarium anthophilum</i>	American	GCA_013364935.1	NRRL 25214	0%	0%	0%	0%
<i>Fusarium bactridioides</i>	American	GCA_013623355.1	NRRL 66639	0%	0%	0%	0%

<i>Fusarium begoniae</i>	American	GCA_013186755.1	NRRL 25300	0%	0%	0%	0%
<i>Fusarium bulbicola</i>	American	GCA_013186765.1	NRRL 22947	0%	0%	0%	0%
<i>Fusarium circinatum</i>	American	GCA_000497325.3	FSP34	100%	100%	100%	100%
<i>Fusarium fracticaudum</i>	American	GCA_003353625.1	CBS 137234	0%	0%	0%	0%
<i>Fusarium guttiforme</i>	American	GCA_013186795.1	NRRL 22945	0%	0%	0%	0%
<i>Fusarium marasasianum</i>	American	GCA_022833035.1	CMW 25512	0%	0%	0%	0%
<i>Fusarium mexicanum</i>	American	GCA_013396015.1	NRRL 53147	0%	0%	0%	0%
<i>Fusarium pilosicola</i>	American	GCA_020615335.1	CMWF1183	100%	100%	95%	98%
<i>Fusarium pininemorale</i>	American	GCA_002165215.1	CMW 25243	0%	0%	0%	0%
<i>Fusarium sororula</i>	American	GCA_017579625.1	FCC 5425	0%	0%	0%	0%
<i>Fusarium sterilihyphosum</i>	American	GCA_013186845.1	NRRL 25623	0%	0%	0%	0%
<i>Fusarium subglutinans</i>	American	GCA_013396075.1	NRRL 66333	0%	0%	0%	0%
<i>Fusarium succisae</i>	American	GCA_013186925.1	NRRL 13298	0%	0%	0%	0%
<i>Fusarium temperatum</i>	American	GCA_001513835.1	CMWF389	0%	0%	0%	0%
<i>Fusarium tupiense</i>	American	GCA_013364945.1	NRRL 53984	0%	0%	0%	0%
<i>Fusarium annulatum</i>	Asian	GCA_019189775.1	F8_4S_2B	0%	0%	0%	0%
<i>Fusarium coicis</i>	Asian	GCA_013781345.1	NRRL 66233	0%	0%	0%	0%
<i>Fusarium concentricum</i>	Asian	GCA_014824425.1	NRRL 25181	0%	0%	0%	0%
<i>Fusarium fujikuroi</i>	Asian	GCA_900079805.1	IMI 58289	0%	0%	0%	0%
<i>Fusarium globosum</i>	Asian	GCA_013396165.1	NRRL 26131	0%	0%	0%	0%
<i>Fusarium mangiferae</i>	Asian	GCA_900044065.1	MRC7560	0%	0%	0%	0%
<i>Fusarium proliferatum</i>	Asian	GCA_900067095.1	ET1	0%	0%	0%	0%
<i>Fusarium sacchari</i>	Asian	GCA_017165645.1	FS66	0%	0%	0%	0%

Table S3. Results of sensitivity testing using purified *Fusarium circinatum* DNA and lysed spore suspension dilutions, and crude DNA extractions from known spore quantities.

DNA added	LAMP Results ^a		qPCR Results	
	Number of positive replicates	Reaction times (min) [total reaction time 60 minutes]	Number of positive replicates	Reaction times (Ct) [total reaction time 60 minutes]
1 ng μL^{-1}	3/3	12, 12, 12	3/3	15.85, 15.98, 15.93
100 pg μL^{-1}	3/3	14, 14, 13	3/3	25.13, 24.89, 24.99
10 pg μL^{-1}	2/3	16, 16	2/3	28.35, 28.26
1 pg μL^{-1}	0/3	-	1/3	39.25
100 fg μL^{-1}	0/3	-	0/3	-
Spores added				
1000 μL^{-1}	3/3	11, 12, 12	3/3	25.83, 25.93, 26.07
500 μL^{-1}	3/3	12, 12, 12	3/3	26.87, 25.98, 26.43
100 μL^{-1}	3/3	13, 12, 12	3/3	28.15, 28.19, 28.37
50 μL^{-1}	3/3	14, 14, 14	3/3	28.88, 28.77, 28.55
10 μL^{-1}	2/3	16, 16	3/3	31.12, 30.63, 30.50
5 μL^{-1}	1/3	47	3/3	33.19, 33.94, 33.39
1 μL^{-1}	0/3	-	0/3	-
Spores (crude)				
1000000	3/3	17, 18, 35	3/3	25.09, 25.74, 26.84
500000	1/3	47	3/3	27.19, 27.71, 27.91
100000	1/3	50	3/3	29.83, 29.05, 29.09
50000	0/3	-	3/3	35.42, 35.32, 33.25
10000	0/3	-	2/3	39.8, 39.0
5000	0/3	-	0/3	-

1000	0/3	-	0/3	-
500	0/3	-	0/3	-

^aPositive LAMP and qPCR results, indicated in bold, were determined by amplification of at least 2 of 3 replicate reactions for each dilution and are shown as the average amplification time (min) or the average amplification time and cycle threshold (Ct) for LAMP and qPCR, respectively.