

## Occurrence, effects, and ecological risks of chemicals in sanitizers and disinfectants: A review


 Ndeke Musee<sup>a,\*</sup>, Phephile Ngwenya<sup>a</sup>, Lenah Kagiso Motaung<sup>a</sup>, Kgalifi Moshuhla<sup>a</sup>, Philiswa Nomngongo<sup>b</sup>
<sup>a</sup> Emerging Contaminants Ecological Risk Assessment (ECERA) Group, Department of Chemical Engineering, University of Pretoria, Pretoria, South Africa

<sup>b</sup> Department of Science and Innovation National Research Foundation South African Research Chair Initiative (DSI-NRF SARChI) in Nanotechnology for Water, University of Johannesburg, Doornfontein 2028, South Africa

### ARTICLE INFO

#### Keywords:

 Ecotoxicity  
 COVID-19  
 Aquatic systems  
 Wastewater  
 Soil  
 Freshwater  
 Risk assessment  
 Risk quotient  
 Risk management

### ABSTRACT

In response to the novel coronavirus referred to as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) – a virus that causes COVID-19 disease has led to wide use of sanitizers and disinfectants. This, in turn, triggered concerns on their potential deleterious effects to human health and the environment due to numerous chemicals incorporated in both product categories. Here, the current state of science regarding the occurrence and ecological effects of different classes of chemicals in these products (e.g., ultraviolet filters, fragrances, etc.) are summarized in different natural (e.g., rivers) and engineered (e.g., wastewater treatment plants) systems. Data collected in the literature suggests chemicals incorporated in sanitizers and disinfectants are present in the environment, and a large portion are toxic to fish, algae, and daphnia. Using the risk quotient approach based on occurrence data, we found eight chemicals that posed the highest risk to aquatic organisms in freshwater systems were benzalkonium chloride, 4-chloro-m-cresol, sodium ortho phenyl phenate, hydrogen peroxide, 1, 2-propanediol, 4-Methyl-benzilidene-camphor, ethylhexyl methoxy cinnamate, and octocrylene. Considering limited occurrence and effects information for most chemicals, further studies on environmental monitoring and potential consequences of long-term exposure in aquatic ecosystems are recommended.

### Contents

1. Introduction . . . . .	63
2. Materials and methods . . . . .	63
3. Occurrence and toxicity of sanitizers and disinfectants . . . . .	64
3.1. Solvents . . . . .	64
3.2. Oxidants and antioxidants. . . . .	65
3.3. Antimicrobials. . . . .	67
3.4. UV filters . . . . .	68
3.5. Fragrances . . . . .	70
3.6. Emulsifiers . . . . .	71
4. Environmental risk characterization. . . . .	72
5. Concluding remarks. . . . .	74
Declaration of Competing Interest . . . . .	74
Acknowledgements . . . . .	74
Supplementary data . . . . .	74
References . . . . .	74

\* Corresponding author.

 E-mail address: [ndeke.musee@up.ac.za](mailto:ndeke.musee@up.ac.za) (N. Musee).


Production and hosting by Elsevier on behalf of KeAi

<http://dx.doi.org/10.1016/j.enceco.2023.01.003>

Received 1 December 2022; Received in revised form 19 January 2023; Accepted 23 January 2023

Available online 4 February 2023

 2590-1826/© 2023 The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co. Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

The emergence of the novel severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) which causes coronavirus disease (COVID-19) outbreak triggered collective efforts by local authorities, governments, and public health institutions across the globe to implement effective preventive measures [1,2]. Among these measures were the non-pharmaceutical interventions (NPIs) (e.g., washing of hands, wearing masks, and social distancing) [2]. Washing of hands and regular decontamination of surfaces led to increased disinfection campaigns on public facilities, community-shared spaces, and homes. Concomitantly, this has led to sharp increase and unprecedented [3] use of sanitizers, and disinfectants across the globe [4–6]. For example, Choi and colleagues have found a rise in handwashing patterns and usage of hygiene product (e.g., hand sanitizers and soaps) during pre- and post-COVID-19 era in South Korea [6]. In addition, another work documented an exponential increase in use of sanitizers and disinfectants to the extent producers and/or manufacturers could not meet the extremely high market demand for both product categories especially at the beginning of the pandemic [7]. Therefore, the increase in use of these product categories has resulted to their constituent chemicals release into natural (rivers, lakes, and groundwater), and technical (e.g., wastewater treatment plants (WWTPs) and tap water) systems. For example, Nason and colleagues qualitatively identified upward trends of disinfectant concentrations (e.g., three ultraviolet filters) in sludge, reflecting increased use during the initial wave of the COVID-19 pandemic [8].

The NPIs have proven to be effective remedy to prevent the spread of COVID-19. However, they have been a source of chemicals incorporated in sanitizers and disinfectants into the aquatic environment, but largely with unknown risks to aquatic biota. Therefore, to investigate and quantify risks associated with sanitizers and disinfectants, systematic identification of their constituent chemicals under variant product categories is required. Remarkably, although there are distinctive differences between sanitizers and disinfectants [9,10], but certain ingredients and classes of chemicals are used in both product categories (Fig. 1). In brief, disinfection refers to the destruction or irreversible inactivation of infectious viruses, germs

and bacteria on surfaces or objects using variant chemicals [2,9,10]. Conversely, sanitization entails use of chemicals to reduce microorganisms on inanimate surfaces, for example, in this case prone to the transmission of COVID-19 infection [1,2,10,11].

The chemicals widely incorporated in sanitizers and disinfectants, for example, found in the South African commerce (Fig. 1) are broadly solvents, oxidants and antioxidants, moisturizers, UV-filter, antimicrobials, emulsifiers, and fragrances. Each chemical used is aimed to achieve single or multiple functions in a given product. For instance, due to their strong activities against microbes and pathogens, antimicrobial agents and antiseptics are incorporated in sanitizers and disinfectants. Furthermore, to achieve effective or improved potency of sanitization and disinfection, two or more active ingredients are combined generally in a given product.

To elucidate the implications of chemicals widely incorporated in sanitizers and disinfectants, first, we systematically reviewed the occurrence, hazard, and risks of the constituent chemicals in the natural and technical systems. Second, we outline a summary of key findings including knowledge gaps related to the sources, occurrence, toxicity, and risks of sanitizers and disinfectants to the environment, and recommendations for plausible future research.

## 2. Materials and methods

To identify the occurrence and effects of chemicals incorporated in sanitizers and disinfectants in different environmental compartments, a literature review was conducted using sources including Google Scholar, Science Direct (published papers, book chapters, and reviews), a dataset of European Chemical Agency (ECHA), and technical documents e.g., material safety database sheet (MSDS) for a given product. Only articles in English were included in this study. The literature search was performed using a suite of keywords (Table 1) following a framework proposed by Hartmann et al. [12]. Keywords either singularly, or as combinations were used in the search. An example of combined keywords is “UV-Filters review surface water”, among many others used in this study. Each chemical was given a symbol for ease of updating the information and a list of the chemicals are in Table S1 in the supporting information (SI).

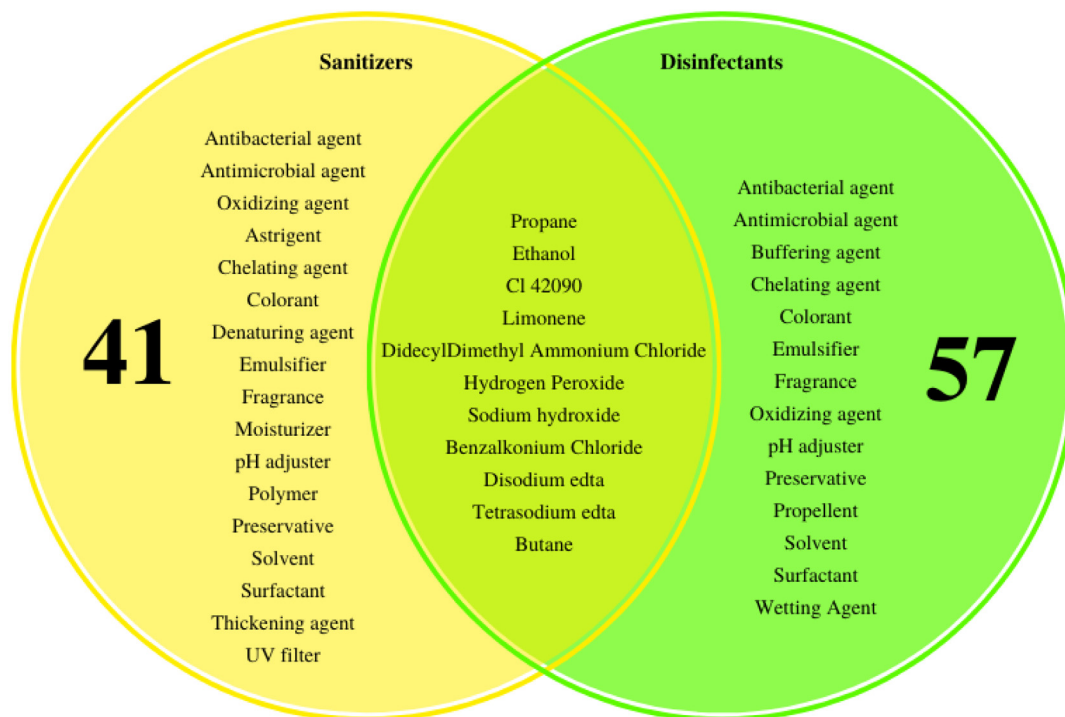


Fig. 1. Constituent chemical classes, and those commonly incorporated in sanitizers and disinfectants identified in South Africa commerce from October 2020 to January 2021 [Source: data collected by researchers in South African market].

**Table 1**

Search words and or their combinations used to retrieve occurrence and effects data of chemicals investigated in the study. These were used with, without spaces, and the AND search function.

Concept 1: Class of chemicals OR specific chemicals in the given class <sup>a</sup> OR product category <sup>a</sup>	Concept 2: Occurrence	Concept 3: Toxicity	Concept 4: Aquatic environment
“Antimicrobials” OR “Fragrances” OR “UV filters” OR “Solvents” OR “Oxidants/antioxidants” OR “Emulsifiers” OR “Moisturizers” OR “disinfectants” OR “sanitizers” OR “chemical name <sup>a</sup> ”	“occurrence” OR “detection” OR “analysis” OR “fate” OR “concentration” OR “measured”	“effects” OR “toxicity” OR “NOEC” OR “EC50” OR “LC50” OR “ecotoxicity” OR “MSDS”, OR “fish” OR “algae” OR “daphnia”	“ecosystem” OR “fresh water” OR “freshwater” OR “surface water” OR “wastewater” OR “wastewater” OR “effluent” OR “river” OR “influent” OR “sludge” OR “aquatic environment” OR “treated water” OR “soil” OR “sediment” OR “lake” OR “groundwater”

<sup>a</sup> Chemical names and product categories in place of classes.

Following the described procedure, a summary of sources per class of chemicals found relevant for the effects and occurrence data used is set out in Table S2. Sources that reported the effects and occurrence for all chemicals were in total 331 (with 150 and 181 for occurrence and effects, respectively). To gain insights on the occurrence data distribution, analysis was done based on continent and environmental compartment (Table S3). Most occurrence data were in Europe ( $n = 56$ ) and North America ( $n = 29$ ), but only three in Africa (Table S3). These findings are in good agreement with those previously reported for chemicals incorporated in personal care products where their occurrence data were severely lacking in Africa and South America [13].

### 3. Occurrence and toxicity of sanitizers and disinfectants

Chemicals used in sanitizers and disinfectants are widely detected in aquatic, terrestrial and engineered environmental systems ranging from very low to high concentrations (ng/L– $\mu$ g/L). The distribution in the environment were observed to be dependent on use patterns, access, or lack thereof to services (e.g., sanitation), standards of living, and geographical loci across the globe. For example, geographical distribution may be dependent on: (i) regulatory regime of a given country or region, (ii) categories of sanitizers and disinfectants in commerce in a specific region or country over a certain period, and (iii) the extent to which certain chemicals have been banned or recommended for use under strict conditions. Here, chemicals incorporated in sanitizers and disinfectants were critically reviewed for their occurrence, sources, ecotoxicity, and plausible impacts on the environmental systems.

#### 3.1. Solvents

Solvents are carbon-based organics that include alcohols, ketones, amines, ethers, esters and aliphatic, cyclic, and halogenated hydrocarbons [14]. These chemicals have homogeneous physicochemical properties, and are further grouped linked to their polarity, and ability to donate a hydrogen bond during interactions with solutes. Solvents play a key role in the manufacturing industry due to limitations set by physicochemical properties of substances, for safety, and control of selectivity during chemical reactions [15].

An estimated 20 million tonnes of solvents are produced yearly [16], and commonly used in sanitizers and disinfectants, and these includes 1,2-propanediol, benzyl alcohol, isopropanol, ethanol, isobutane, Polyethylene glycol (PEG)-40 hydrogenated castororbate-60. This has led to the presence of certain chemicals in soils, surface water, groundwater, and wastewater [17–20]. For example, concentrations of isopropanol were high in Greece of up to 40.6  $\mu$ g/L in wastewater [21]. The high concentrations were due to use of isopropanol as a solvent for photo-resistant stripping and cleaning following the etching of silicon wafers in the semiconductor industry by companies located in the industrial science park near the sampling area. In addition, Lee and colleagues detected isopropanol in WWTP effluents (292 ng/L) and lake water (3.10  $\mu$ g/L) [22].

Benzyl alcohol is used as a solvent in different product categories including paints, adhesives, perfumes, and sanitizers [23]. Benzyl alcohol was detected in wastewater treatment plants (WWTPs) in China at

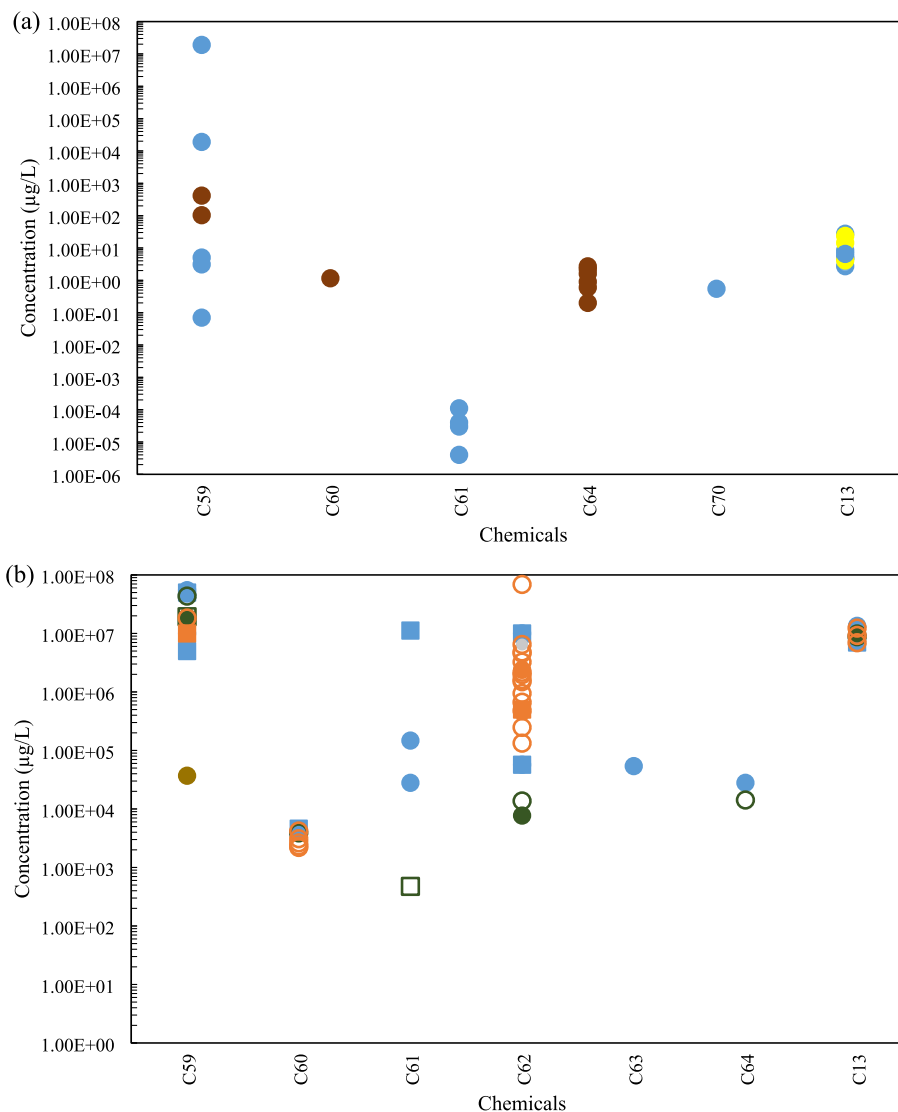
concentrations of 500–11,000 and 150–400 ng/L, respectively, in the influents and effluents [20]. Data suggest ethanol is the most commonly detected solvent (Fig. 2a). For example, Avery et al. [24] reported concentrations of 2.73 and 23.59  $\mu$ g/L for ethanol in surface waters and lakes, respectively, in USA (Fig. 2a). Studies suggest that frequent and continued use of sanitizers may pose severe human health risks including cardiac dysrhythmias, respiratory arrest hypothermia, cancer, among others [25,26], especially due their extended period of use as was the case during peak COVID-19 pandemic.

In the USA, 1,2-propanediol was detected at very high concentrations in the surface water (19  $\mu$ g/L), but very low in the groundwater (4 ng/L) [27] (Fig. 2a). It is likely the elevated quantities of 1,2-propanediol were from de-icing agents as these values were detected in storm water runoff at an airport in Utah, USA [27]. Further, propane and butane were detected in soil samples in India at concentrations of 1–34  $\mu$ g/L and 1–9  $\mu$ g/L, respectively [28]. Conversely, solvents were detected in the air due to their high volatility. For example, in Canada 199, 80.23, and 62.03 ng/L of butane, isopropane, and isobutane, respectively, were detected in air; and as a result increased the likelihood for direct exposure to humans.

Effects of solvents to different taxa (fish, algae, *Daphnia magna*, and bacteria) under different test durations of 24 to 96 h for acute studies have been reported for a number of solvents [29–31]. For example, 96-h lethal concentration 50 (96-h LC<sub>50</sub>) values of benzyl alcohol on three different fish species: *Leuciscus idus*, *Pimephales promelas* (fathead minnow), and *Lepomis macrochirus* (Bluegill) were established at 646, 460, and 10 mg/L, respectively. A study by Mattson [32] reported 96-h LC<sub>50</sub> acute toxicity test on *Pimephales promelas juveniles* of 460 mg/L nominal concentration. Twenty-four- and forty-eight-h effective concentration 50 (EC<sub>50</sub>) values on immobilization of *D. magna* were, respectively, 55 mg/L [33], and 230 mg/L [34]. Moreover, studies using *Scenedesmus quadricauda* as exposure organism yielded 96-h EC<sub>50</sub> value on growth inhibition at 640 mg/L. These results indicate benzyl alcohol can potentially exert harmful to non-harmful effects to fish.

Toxicity data reviewed herein show alkyl dimethylamine oxide (dodecyl dimethylamine oxide or lauryl dimethylamine oxide) as most toxic solvent to different taxa e.g., fish, algae, etc. [35] dependent on exposure duration and organism type. For example, documented 96-h LC<sub>50</sub> and no observed effect concentration (NOEC) values for *Pimephales promelas* (fathead minnow) fish were 3.46 mg/L and 0.420 mg/L, respectively. In addition, a low NOEC of 1.30 mg/L was recorded for *Oryzias latipes* [36]. In another work, *Danio rerio* were found to be the most tolerant species assessed with LC<sub>50</sub> of 31.8 mg/L [37].

Similarly, results of *D. magna* [38] demonstrated very low NOECs (0.700 mg/L) on epical endpoints (production and survival). In addition, algae *P. subcapitata* species effects data for the production and survival endpoints yielded 72-h ErC<sub>50</sub> values of 0.153 [30], and 0.250 mg/L [35] respectively. However, certain solvents (e.g., 1,2-propanediol and isopropanol) were observed to be non-harmful to different taxa with E(L) C<sub>50S</sub> > 1000 mg/L (Fig. 2b). For example, Bridie et al. [39] reported LC<sub>50</sub> > 5000 mg/L on *Carassius auratus* species (goldfish); whereas West and colleagues [40] reported 48-h LC<sub>50</sub> of 10,000 mg/L following exposure of *Lebistes reticulatus* (guppy) to 1,2-propanediol. These authors also conducted 24-h studies on the immobilization of *D. magna* which yielded



**Fig. 2.** (a) Occurrence of chemicals classified as solvents in sanitizers and disinfectants in variant environmental matrices (surface water (blue), lake water (yellow), and groundwater (brown)). And (b) their E(L)C<sub>50</sub>s tested on fish (blue), algae (green) and daphnia (orange) at different exposures (24 h (full squares), 48 h (open circles), 72 h (open squares) and 96 h (full circles)). In the entire paper, the following colors are used for consistency to represent variant exposure durations in this report in toxicity graphs. There are: black (0.083 h), brown (0.25 h), purple (0.5 h), pink (3 h), red (17 h); green (24 h); navy blue (48 h); orange (72 h), and sky blue (96 h).

EC<sub>50</sub> > 10,000 mg/L [40]. The low toxicity of 1,2-propanediol can be attributed to its rapid degradation in different environmental media with half-lives of 1–4, and 0.8 d in water and air, respectively [41].

The glycerol and glycerin are among widely used solvents. Herein, results of both chemicals were grouped together owing no specificity on toxicity reported as in certain cases it included their derivatives [42]. For example, Perales and colleagues documented effects of different glycerol derivatives with EC<sub>50</sub> values >100 mg/L for different species, and therefore were broadly non-toxic except for the solvent code 444 on *D. magna* which yielded a EC<sub>50</sub> of 13.7 mg/L [42]. For sanitizers and disinfectants containing polyalkylene glycols, it was not possible to classify their toxicity and occurrence data based on a specific chemical as the term includes a broad number of compounds e.g., polyethylene glycol, polypropylene glycol, among others.

For alcohol-based solvents, data indicates that they are non-harmful to the aquatic taxa (Fig. 2b). For example, isopropanol (isopropyl alcohol) exerted no toxicity on fish *Carassius auratus* mortality as 24-h EC<sub>50</sub> values were > 5000 mg/L, and 7060 mg/L over 7-d chronic studies on *Poecilia reticulata* (guppy) [37]. Results on exposure of *D. magna* to isopropanol showed similar trends with very low EC<sub>50</sub> of 3010 mg/L as well as NOECs

of 2100 mg/L and 757 mg/L for the reproduction and growth endpoints, respectively. These results and other data summarized in Fig. 2b on effects of solvents point to highly varied effects from non-harmful (e.g., 1,2-propanediol or isopropyl alcohol) (irrespective of organisms' trophic level) to high toxic (e.g., dodecyltrimethylamine oxide to algae).

### 3.2. Oxidants and antioxidants

Antioxidants are used to preserve lipid components from quality deterioration through the prevention of oxidation with an estimated global production of 1.25 million tons by 2016 [43]. These chemicals are classified broadly into three categories based on their antioxidant mechanisms, namely; primary, secondary, and tertiary antioxidants [44,45]. Their mechanisms of action include neutralizing or scavenging reactive nitrogen (RNS) or oxygen species (ROS); or inhibition on the formation of RNS or ROS; or removing O<sub>2</sub> and binding the metal ions required for the generation of ROS. Antioxidants are widely used in product categories including personal care products (PCPs), foods, sanitizers, plasticizers, textiles, and other household goods [46]. Conversely, oxidizing agents are used in fabric bleaches, water purification, fuel combustion, and rubber vulcanization.

Data demonstrates the widespread occurrence of oxidants and antioxidants in surface waters, wastewater, soils, and sludge [47–50], but this is only for limited chemicals different environmental matrixes (Fig. 3a). Results herein indicate oxidants and antioxidants commonly detected are tocopheryl acetate and hydrogen peroxide (Fig. 3a). For example, tocopheryl acetate was detected in sludge with maximal concentrations reaching 1.10 mg/g [51]; whereas in surface water measured hydrogen peroxide was 3.67  $\mu\text{g/L}$  [52]. Further, in Germany elevated concentrations of tocopheryl acetate were detected in WWTPs effluent in the range of 6–25  $\mu\text{g/L}$  [51]. Blum and colleagues detected 660  $\mu\text{g/kg}$  and 110  $\text{ng/L}$  of tocopheryl acetate, respectively, in soils and WWTP effluents in Sweden [53]. Therefore, the release of effluent with high concentrations in the  $\mu\text{g/L}$  range of tocopheryl acetate may end up into surface waters unless it undergoes several hundred-fold dilution factors, or are effectively removed in the WWTPs. This, in turn may pose reasonably high risks to aquatic organisms at different biological levels of organization.

Methylisothiazolinone a heterocyclic compound [54,55] was detected at concentrations (mean and standard deviation with  $n = 4$  independent experiments) of  $1.21 \pm 0.46$  in untreated sewage as well as in the beach (at two different locations i.e., Leba and chlapowo) at levels of  $2.19 \pm 0.47$  and  $4.48 \pm 1.04$   $\mu\text{g/L}$ , and soil at levels of  $1.04 \pm 0.06$ – $10.8 \pm 1.07$   $\mu\text{g/kg}$ , respectively, [48]. However, in the seawater and river samples the concentrations of methylisothiazolinone were below the limit of quantification (LOQ) [48] likely due to high dilution in both environmental matrixes. In addition, the presence of oxidants in tap water samples have been documented (Fig. 3a). This implies ineffective portable water treatment technologies and concomitant increasing use of sanitizers and disinfectants may lead to human exposure to their constituent chemicals through drinking water. However, most chemicals permissible exposure threshold values especially to vulnerable populations (e.g., elderly, children, etc.) remain yet to be established.

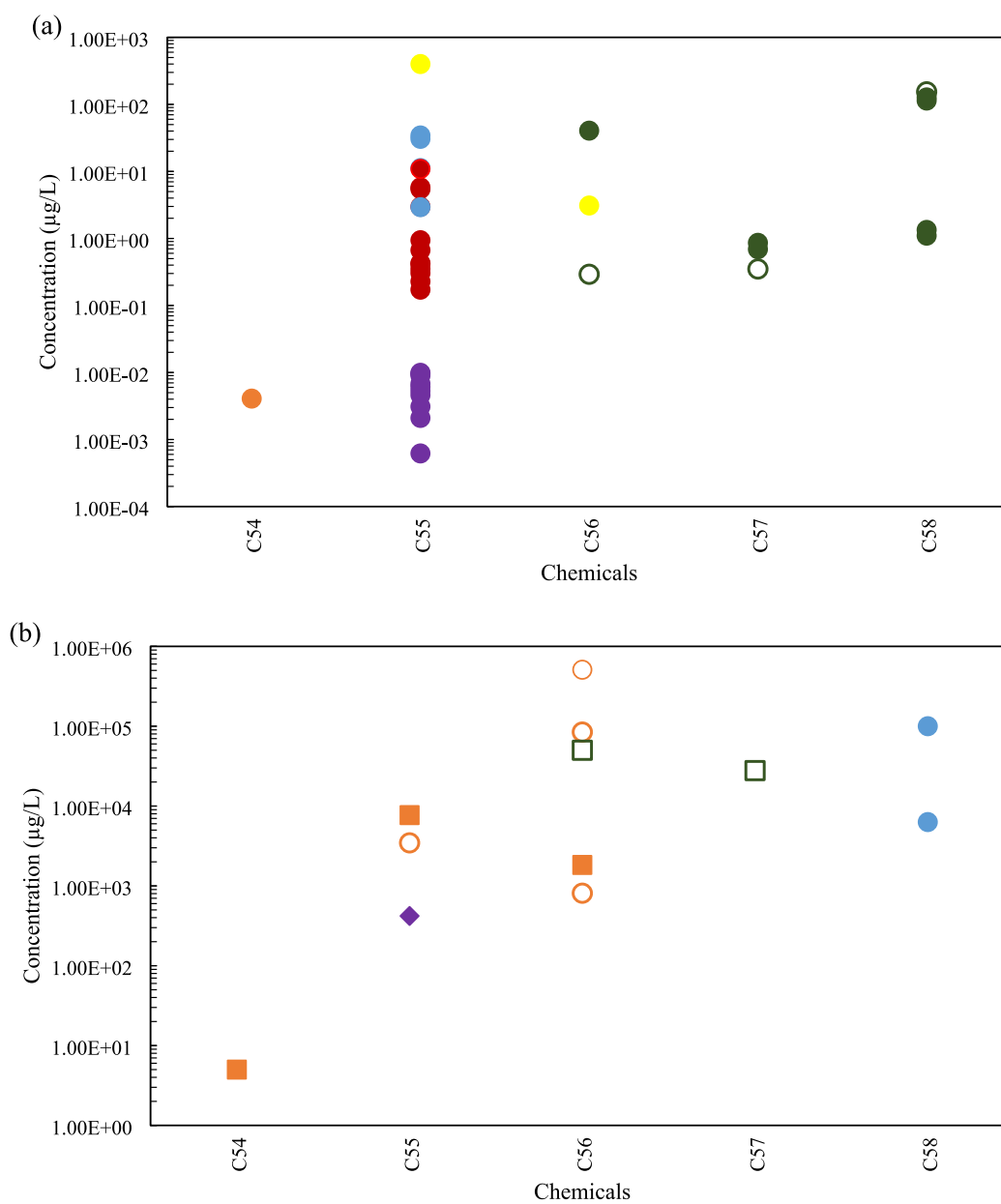


Fig. 3. (a) Occurrence of chemicals classified as oxidants and antioxidants in sanitizers and disinfectants in variant environmental matrixes (WWTP influent (full green circles) and effluent (open green circles), surface water (blue), river water (red), tap water (orange), lake water (yellow) and marine water (purple). And (b) their  $E(L)C_{50}$ s tested on fish (blue), algae (green) daphnia (orange) and bacteria (purple) at different exposures (0.5 (full diamonds), 24 h (full squares), 48 h (open circles), 72 h (open squares) and 96 h (full circles).

Effects data of oxidants and antioxidants on aquatic organisms at different levels of biological organization are summarized in Fig. 3b. Similar to their occurrence data (Fig. 3a), the ecotoxicity studies of these compounds are limited (Fig. 3b). In this review, the assessed data indicate hypochlorous acid as the most toxic oxidant (Fig. 3b) with 48-h  $EC_{50}$  values for crustaceans and for fish as 5 [56], and 40  $\mu\text{g/L}$  [57], respectively.

Recent increasing interest in peracetic acid (PAA) as a sustainable disinfectant for use in aquaculture due to its low concentrations (as low as 1 mg/L) required, and knowledge deficits on its toxicity to fish has made it to be the most investigated chemical in this class [58,59]. PAA releases into the environment and various waste streams are due to wide usage as a disinfectant, and from the production facilities. There are propositions that PAA may not pose toxicity effects when diluted in water to its effective concentration [60,61]. However, Straus et al. [62] exposure study on Channel catfish (*Ictalurus punctatus*) found that 24-h  $LC_{50}$  values of 2.60 and 1.60 mg/L for yolk-sac fry and swim-up fry, respectively. Furthermore, NOECs of 2.20 and 1.30 mg/L for yolk-sac fry and swim-up fry, respectively, were observed. Similarly, Straus et al., (2012) [63] observed 48- and 72-h  $EC_{50}$  values in the ranges of 0.152–1.10 mg/L, and 35.0–350  $\mu\text{g/L}$  on *D. magna* and *Scenedesmus subspicatus*, respectively. Elsewhere, several researchers [64,65] reported the  $EC_{50}$  values of 1.38 and 0.18 over 72 and 120 h, respectively, on *Selenastrum capricornutum*. In addition, Hicks and colleagues [64] observed very low NOEC of 130  $\mu\text{g/L}$  for the same species. Taken together, reviewed data point to high toxicity of PAA to fish, crustaceans, and unicellular organisms, and therefore, contradicts the proposition that PAA is non-toxic to aquatic organisms.

Other works have reported high toxic effects of solvents on aquatic organisms [66,67]. For example, sodium dichloroisocyanurate dihydrate was found to be very toxic to daphnia with 48 h  $EC_{50}$  of 0.19 mg/L [66]. Further, terpeneol has also been found to induce toxic effects to salmonid fish species with observed 96-h  $LC_{50}$  values of 6.30 and 6.60 mg/L for coho salmon and rainbow trout, respectively [67]. Conversely, hydrogen peroxide and L-Lactic acid were non-harmful to aquatic organisms with most  $EC_{50}$  values >100 mg/L [68,69]. Overall, data suggest PAA and hypochlorous acid to be toxic or very toxic irrespective of the taxonomical class as results summarized in Fig. 3b as both chemicals has mostly  $E(L)C_{50}$  values <10 mg/L. Data suggest fish and daphnia as most sensitive species with several  $E(L)C_{50}$  values <1 mg/L [62,65,66]. Due to data deficit on the occurrence and/or effects of oxidants and antioxidants compounds impede establishing their risks to aquatic organisms. This, in turn raises the need for occurrence and effects studies of oxidants and antioxidants to aid their risk assessment in the aquatic environments.

### 3.3. Antimicrobials

Antimicrobials are chemicals or their mixtures thereof employed to destroy, deter, render harmless, prevent the action of, or otherwise exert a controlling effect on any harmful organism by any means other than mere physical or mechanical action [49]. Different classes of chemicals are used as antimicrobials, and these include alcohols, [70], aldehydes [71], biguanides [72], and halogen-releasing agents [73] with N-halamines as most dominant [74]. Recent advances have led to the replacement of iodine by more effective antimicrobials [75] including peroxygens and other forms of oxygen (e.g., hydrogen peroxide, ozone), peracetic acid, and chlorine dioxide [76], and phenols [77].

Thirty-two antimicrobials were reviewed in this study, and their concentrations varied widely from low 0.60 ng/L (South Korea) [78] to 3.87 mg/L (Ghana) [79] in the influent and effluent. Notably, the exceptionally high concentrations of benzalkonium chloride (BAC) detected in Ghana were attributed to its wide use in household cleaning products [80,81]. BAC is the widely detected antimicrobial in wastewaters across the globe in countries including Austria, Germany, Sweden, USA [82], France [49], and Ghana [79]. For example, BAC was detected in river water at concentrations of 0.67–1.60 ng/L in China [83], and 1220 ng/L in USA [84]. Further, BAC was detected in digested sludge at very high concentration of 210–89,000 ng/g [85]. These results demonstrate the release

of BAC to water resources and soil compartments through effluents, and sludge application in agricultural fields, respectively.

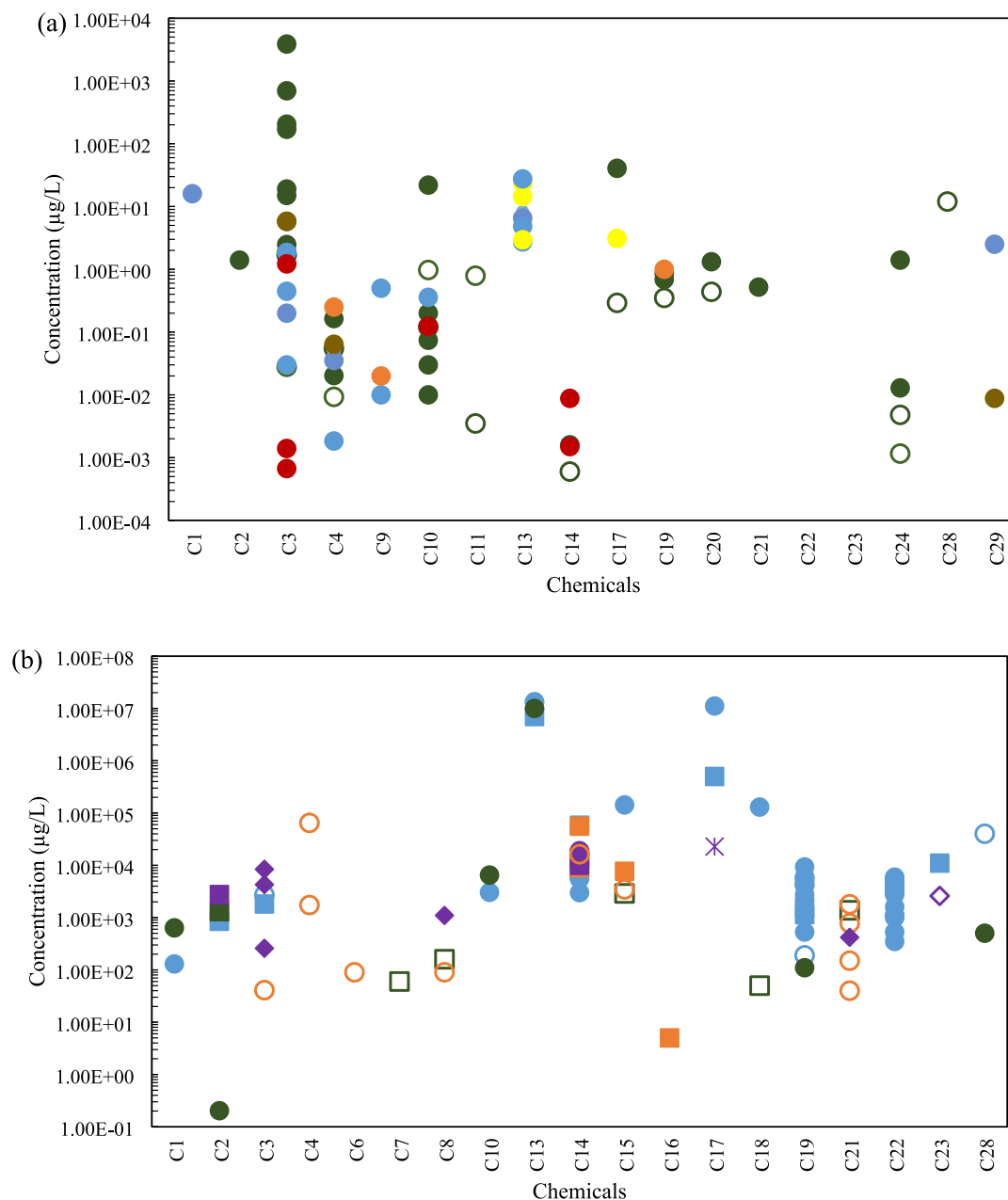
Following the incomplete removal of antimicrobials in wastewater, have led to their eventual presence in surface water. Seven antimicrobials were detected in surface water in five different countries. For example, BAC was detected in urban runoff in France [86], chloroxylenol in the UK [87,88], sodium ortho phenyl phenate in China [22], and isopropanol (isopropyl alcohol) in Korea [22]. Compounds with the maximum concentrations recorded in surface water were chloroxylenol at 21.9  $\mu\text{g/L}$  (UK) [87] and isopropanol (isopropyl alcohol) (Korea) at 3.10  $\mu\text{g/L}$  [22]. This is possibly because both chemicals are widely used as antiseptics and disinfectants in cosmetics and personal care products, leading to their consequent high concentrations in surface waters linked to incomplete removal in WWTPs.

Although antimicrobials are widely used, but their toxicological effects on non-target environmental microorganisms [89], and other aquatic organisms are severely lacking. Susceptibility to antimicrobial agents is due on their ability to kill microorganisms and/or inhibit growth. This poses challenges to comparing toxicity results between those conducted in laboratory settings versus in actual environment matrixes as testing conditions significantly influence the observed outcome effects [90–92]. Results in Fig. 4a summarize the toxicity of antimicrobials on different aquatic taxonomical groups. Of the 17 antimicrobials documented in this review, four were found to induce very high acute toxicity (4-chloro-m-cresol, methylisothiazolinone, peracetic acid, benzalkonium chloride, and sodium ortho phenyl phenate) relative to the rest.

In addition, a large portion of chemicals were found to exert effects ranging from very toxic to harmful on different taxa except in certain cases e.g., glutaraldehyde (on fish), and methylisothiazolinone (on daphnia), with algae and daphnia as the most sensitive species. For example,  $EC_{50}$  values of 4-chloro-m-cresol and sodium ortho phenyl phenate on fish *Acartia tonsa* (planktonic copepod) indicates they are very toxic at 130 and 116  $\mu\text{g/L}$ , respectively, on the same species [88]. Elsewhere, results of Leung [93] demonstrated that glutaraldehyde was toxic to different fish species including *Oncorhynchus mykiss*, *Pimephales promelas*, *Lepomis macrochirus*, and *Salmo salar* with  $EC_{50}$  values of 11, 5–6, 9–11, and 3 mg/L, respectively. Other works by Pereira and colleagues [94] observed glutaraldehyde to be toxic to *Danio rerio* with  $EC_{50}$  of 5.80 mg/L.

Other organisms including *D. magna* [95], *Ceriodaphnia dubia* [96], and *Neocardina denticulata* [97] were found to be sensitive to antimicrobials, nine were identified to be highly toxic with toxicity values ranging from 0.04 to 1.80 mg/L. These chemicals included 4-chloro-m-cresol, BAC [98], benzisothiazolinone [99], chlorine [65], chlorophenols [100], chloroxylenol [88], hypochlorous acid [56], methylisothiazolinone [83], PAA [65,101], and sodium ortho phenyl phenate [88]. The CCR report [102] documented the chronic toxicity of glutaraldehyde exposed to *D. magna* for 21 d with NOEC and LOEC values of 2.13 and 4.25 mg/L, respectively. For BAC, Lavorgna et al. [103] reported the chronic toxicity tested over 21 d on *D. magna*, and 7 d on *C. dubia* with high  $EC_{50}$ s of 1  $\mu\text{g/L}$  and 44  $\mu\text{g/L}$ , respectively. In addition, BAC induced DNA damage on *D. magna*, and *C. dubia* at an  $EC_{50}$  at low concentrations of 38.2 and 403.7  $\mu\text{g/L}$  [103], respectively. Results of BAC demonstrate that both chronic and acute observed effects fall within the same range and highlight its high toxicity to the crustacean daphnia.

Studies on algae showed these organisms are sensitive to antimicrobials, with reported  $EC_{50}$ s in the range of 0.01–10 mg/L (Fig. 4b). As an example, BAC tested on *P. subcapitata* had  $EC_{50}$  of 41  $\mu\text{g/L}$  [98], whilst benzisothiazolinone tested on green algae had an  $EC_{50}$  of 150  $\mu\text{g/L}$  [104]; therefore, both chemicals are very toxic to algae. Both BAC and octyldecyl dimethyl ammonium chlorides had a higher growth inhibition effect on *Chlorella vulgaris* with an  $EC_{50}$  of 203  $\mu\text{g/L}$  and 110 mg/L [105], respectively. Utsunomiya and colleagues [106] also found BAC to be highly toxic on *Chlorella pyrenoidosa*. Notably, most individual antibacterial compounds are predominantly toxic. This, in turn raises the likelihood of resultant chemical mixtures with common



**Fig. 4.** (a) Occurrence of antimicrobial chemicals in sanitizers and disinfectants in variant environmental matrices (WWTP influent (full green circles) and effluent (open green circles), surface water (blue), river water (red), tap water (orange), lake water (yellow), and groundwater (brown)). And (b) their  $E(L)C_{50}$ s tested on fish (blue), algae (green) daphnia (orange) and bacteria (purple) at different exposures (0.083 h (asterisk), 0.25 h (open diamond), 0.5 h (full diamond), 24 h (full squares), 48 h (open circles), 72 h (open squares) and 96 h (full circles)).

mechanisms of action with higher combined deleterious effects to the aquatic organisms [107].

Reviewed data also showed high bactericidal effects of antimicrobials (Fig. 4b). For example, Chhetri and co-workers [65] observed very high  $EC_{50}$  values following 30 min bioluminescence exposures of PAA, hydrogen peroxide, chlorine dioxide, and chlorine on *Vibrio fischeri* at 0.42, 5.67, 1.10, and 1.10 mg/L, respectively. These results indicate PAA as most toxic to bacteria. In another study, 24 h chronic studies following exposure of chlorhexidine gluconate on different bacterial species *Bacillus subtilis*, *Staphylococcus aureus*, and *Escherichia coli* induced inhibition at concentrations of 3.46 mg/L, 4.42 mg/L and 2.33 mg/L, respectively, thus indicative of *Escherichia coli* being the most sensitive bacterial species.

### 3.4. UV filters

Organic ultraviolet (UV) filters are used in numerous consumer products chiefly to provide protection against damage from UV irradiation [108] with eventual increasing release into the environment from different sources. These sources include human recreational activities (e.g., swimming and bathing), industrial wastewater discharges, construction (e.g., in paints), and laundry activities. There are two classes of UV filters, viz.: organic and inorganic filters, and account for approximately 20% of the sunscreen products in commerce [109]. Here, the focus is on the organic category as they are incorporated in sanitizers and disinfectants found in the South African commerce. Examples of UV filters identified were benzophenone-3 (BP-3), benzophenone-4 (BP-4), 4-Methyl-

benzilidene-camphor (4MBC), ethylhexyl methoxy cinnamate (EHMC), and octocrylene (OC), and are widely used globally [110].

Until now, UV filters have been detected in natural aquatic, terrestrial environments, and engineered systems (e.g., WWTPs) across the globe. Data demonstrates wide variability of UV filters concentrations (0.505–2196  $\mu\text{g/L}$ ) in wastewater [87,111–115]. This may be due to a combination of factors including low water solubility, high lipophilicity, low degradability, and high organic content [116,117]. Thus, UV filters are likely to accumulate in the aquatic environment [116].

BP-3 is the most widely measured UV filters in environmental systems including rivers, surface waters, and wastewater, but few detections in lakes, groundwater, and seawater (Fig. 5a). Owing to the high use of BP-3 elevated concentrations have been detected e.g., in sludge (Norway; 0.824–2.12  $\mu\text{g/g}$  [113], wastewater effluent (Italy; 46.9  $\mu\text{g/L}$  [118], and sea water (Central Pacific Ocean; 34.3  $\mu\text{g/L}$  [119]. Additionally, maximum concentrations of 5.72, 3.35, and 44  $\mu\text{g/L}$  have been measured in river waters in Spain [120], China [121], and the UK [87], respectively. In another

study, BP-3 was detected in Spain in surface water and tap water at concentrations of 5429 as well as 10 to 295  $\text{ng/L}$ , respectively, [111].

Other chemicals incorporated in UV filters have also been detected in the aquatic environment including EHMC (3.0–27.1  $\mu\text{g/L}$  [119,122], and OC (4.4  $\mu\text{g/L}$ ) in lake water (Fig. 5a). The occurrences for the BP-4 were very high measuring in certain cases over 1000  $\text{mg/L}$  both in the influent and effluent (Fig. 5a), and river water samples in the UK [87]. The higher BP-4 concentrations in the effluent were attributed to the inefficacy of WWTPs; thus, pointing to the treatment systems' inability to completely remove the UV filter [87]. At present, only few studies have documented the occurrence of UV filters in tap- and ground-waters with most concentrations <100  $\text{ng/L}$  [111,122]. Other works have documented UV filters presence in sediments and sludge at elevated concentrations in the  $\mu\text{g/g}$  range for BP-3 (2.1), EHMC (4.7), and OC (41.6) [113], and BP-4 (5.0) [114]. Differences in concentrations of UV filters in river water, and their presence in sludge were dependent on variability of removal efficiencies based on technologies adopted in a given WWTP. For example, activated sludge

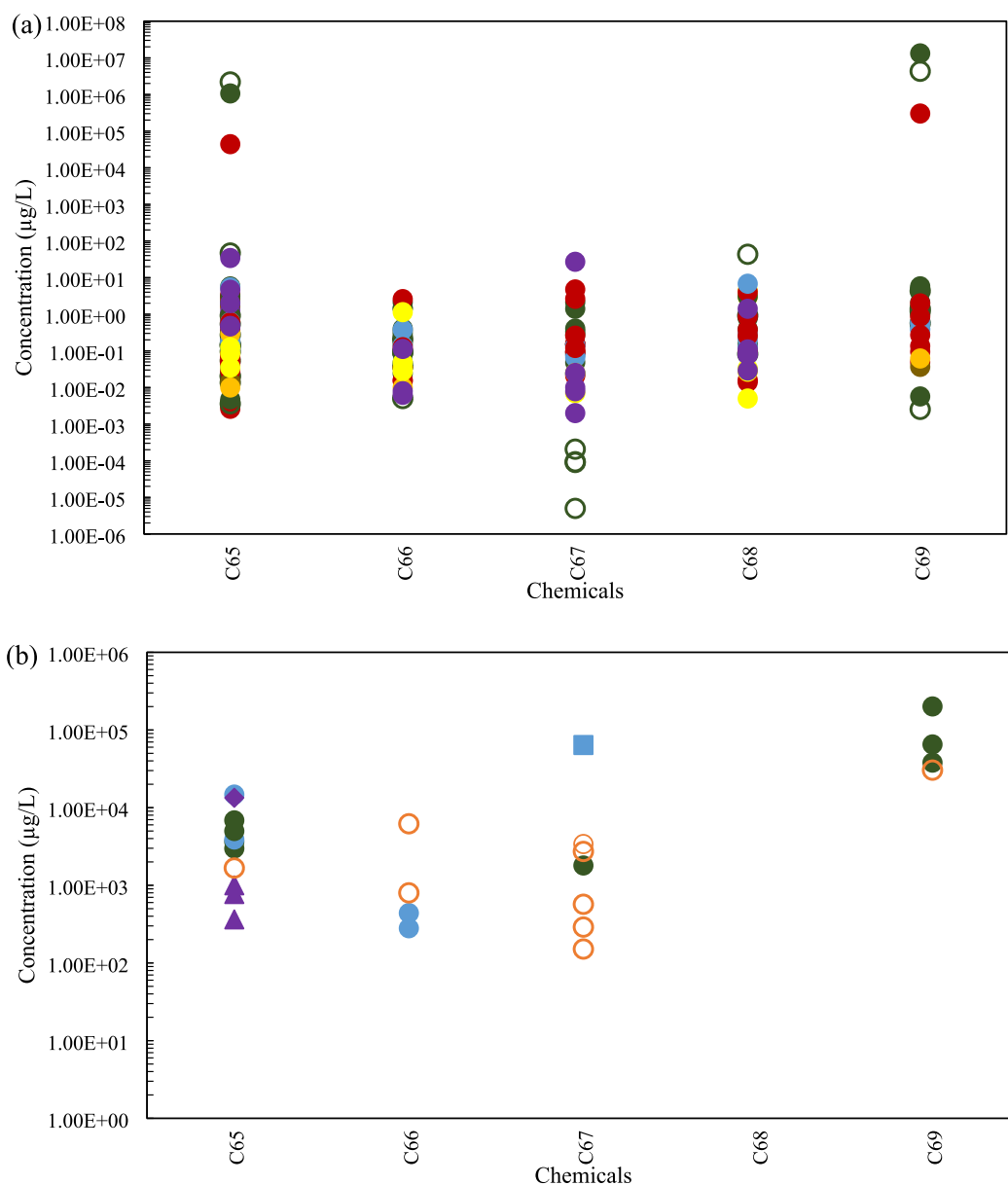


Fig. 5. (a) Occurrence of chemicals incorporated in UV filters in sanitizers and disinfectants in variant environmental matrices (WWTP influent (full green circles) and effluent (open green circles), surface water (blue), river water (red), lake water (yellow), marine water (purple) and groundwater (brown)). And (b) their E(L)C<sub>50</sub>s tested on fish (blue), algae (green) and daphnia (orange) at different exposures (0.5 (full diamonds), 24 h (full squares), 48 h (open circles), 72 h (open squares) and 96 h (full circles)).



treatment was inefficient to remove UV filters compared to other treatment processes e.g., filter beds [87]. Therefore, owing to increasing and high concentrations of UV-filters detected in the aquatic environments, they have been identified as among chemicals of emerging environmental concern [123].

Data indicates UV filters can induce different levels of effects on the aquatic life (Fig. 5b). Due to deleterious effects on coral reefs, some chemicals used as in UV filters have been banned in several countries. For instance, 4MBC has been banned as a sunscreen component in the USA [124]. Other countries like Hawaii, Key West, and the United States Virgin Islands (USVI) have recently banned the use of sunscreens containing BP-3 and EHMC owing to associated correlation between adverse effects and the coral reef bleaching [117,125]. Several studies have demonstrated the degradation of UV filters through the photolysis processes; however, the effects of the formed transformation products (TPs) remain unquantified [124]. For example, 4-HB (4-hydroxybutyrate) a TP of benzophenones was observed to induce adverse effects on reproduction, and interference with aquatic and terrestrial organisms' growth [126].

Toxicity studies for UV filters are largely for the crustacean species with observed effects ranging from moderate to high (<10 mg/L) except for BP-4 and OC. In addition, BP-4 and OC data showed they had harmful effects with 48 h LC<sub>50s</sub> (Fig. 5b) (30.4–50 mg/L) [126–128] and very high toxicity (48-h EC<sub>50</sub> 0.03 mg/L) [129], respectively. The mechanism of toxicity mostly entailed hormone interference as well as bioaccumulation effects [130]. For instance, exposure of *D. magna* to OC induced behavioral impairment on their photo-tactic response, and delayed mortality up to seven days post-exposure at environmentally relevant concentrations (0.20–200 µg/L) [129]. Similarly, BP-3 and EHMC were observed to induce moderate toxicity to algae (96 h EC<sub>50</sub> of 2.98 mg/L and 72 h EC<sub>50</sub> of 1.80 mg/L) [126,127]. In addition, BP-4 can cause slight to non-toxicity (96 h EC<sub>50</sub> for growth of 38.0 mg/L) [131,132]. However, no toxic effects on algae were observed following exposure to 4MBC and OC. Additionally, OC and BP-4 were non-toxic on fish, but BP-3 and 4MBC induced moderate to high toxicity with results for EHMC indicative of low toxicity.

In another study, results indicated BP-3 can reduce egg production, and with subsequent fewer hatchings and feminization of male fish. As a result, this has deleterious implications on reproduction and ultimate fish population [133]. EHMC was observed to induce increased malformation, heart rate, and hatching delay in fish embryos [134] whereas BP-4 interfered with expression of genes involved in hormonal pathways and steroidogenesis following exposure to zebra fish [135]. In other works, 4MBC was observed to impair motility, and also the induction of morphological abnormalities during embryonic development of fish [136,137]. In addition, 4MBC was found to increase the production of vitellogenin and chorigenin in male fish (*Oryzias latipes*) [138], increases in hatching time, reduction on hatching rate, and retarded growth [139]. Overall, the toxicity of UV filters on aquatic organisms especially on bacteria is limited with data accessible only for BP-3. Results indicate BP-3 is highly toxic to bacteria (*Bacteroidetes* and *Proteobacteria*) (48 h EC<sub>50</sub> 0.36–1.00 mg/L) [140], and toxic to *V. fischeri* (15 min EC<sub>50</sub> 13.4 mg/L) [141]. Therefore, high release of UV filters from sanitizers and disinfectants may have serious implications to microbes e.g., bacteria.

### 3.5. Fragrances

Fragrances are organic compounds with pleasant odours, and widely incorporated in numerous daily products including perfumes, hair care products, baby care products, lotions, essential oils, air fresheners, detergents, and as a flavor in bakery items [142,143]. As they are PCPs subclass [144,145]; and are classified according to their source, chemical structure, and their provided note. Natural fragrances consist of two classes, namely; aroma and musk compounds differentiated by their plant and animal sources, respectively [146]. The largest class of fragrances are synthetics but are not included in this work as none were identified in sanitizers and disinfectant product categories. In early 2010s, with increasing demand

globally, fragrances production volumes were projected above 1000 t annually in the EU [147], and are on the rise.

To date, however, there are limited studies on measured environmental concentrations of different fragrances especially the organic forms in comparison to synthetic ones including polycyclic musks such as galaxolide (HHCB) and tonalide (AHTN), and bicyclic hydrocarbon [148]. Occurrence data for most fragrances ranged from low (4.95 ng/L) to high (238 µg/L) concentrations, but none >100 mg/L in different matrixes except for two chemicals: benzyl salicylate and methylpropional (Fig. 6a). Benzyl Salicylate to date has been widely reported in different environmental matrixes as demonstrated by data retrieved from the published literature (Fig. 6a).

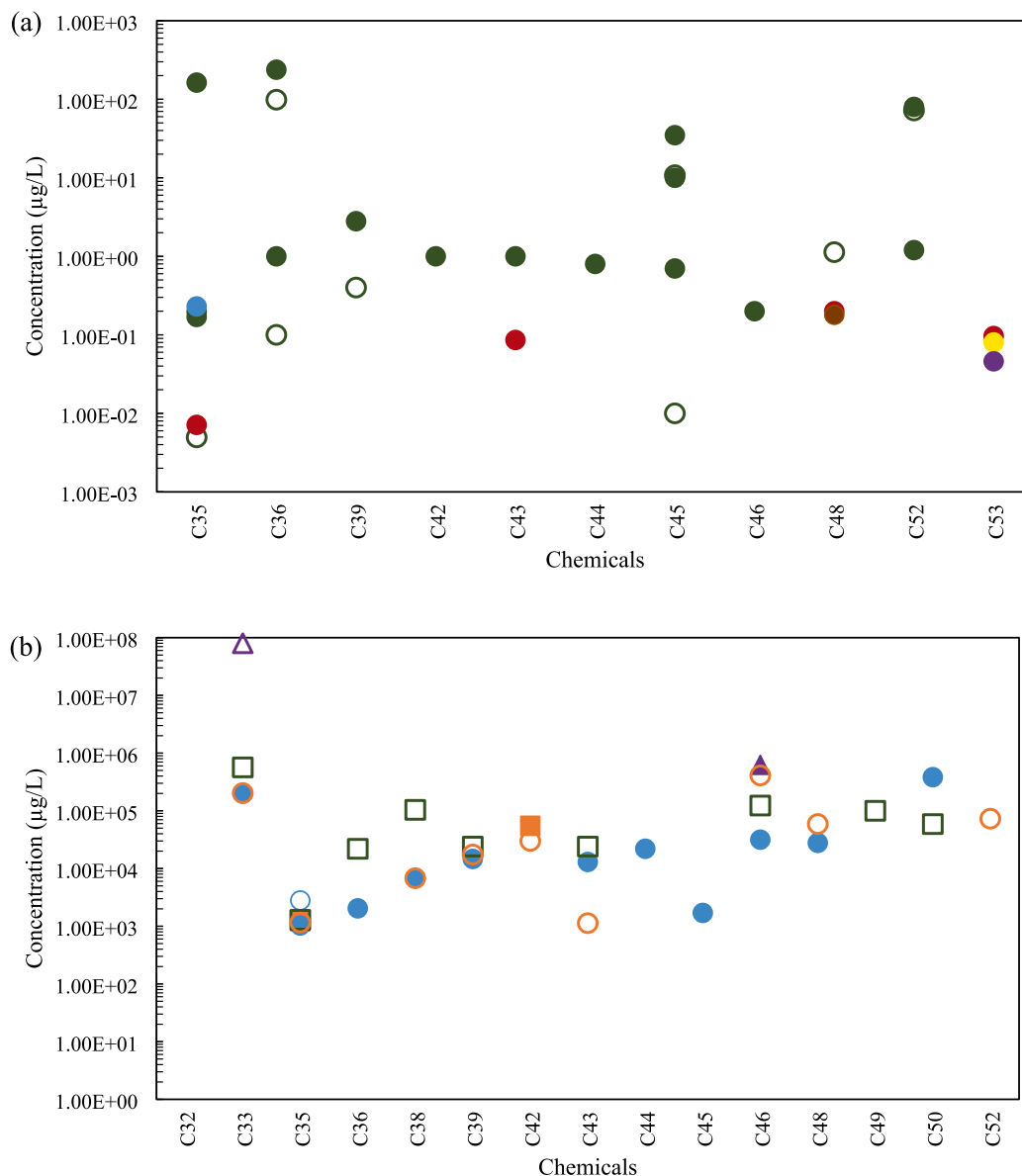
Concentrations of Benzyl salicylate ranged from <LQL (lowest quantifiable limit) to 127 µg/L in wastewater effluent [149], and potentially high levels were released into the surface waters. Further, this compound has been detected in WWTP effluent leading into seawater in Terra Nova Bay, Antarctica at low concentrations of 2.20–4.50 ng/L [150]. In other works, for example, D-Limonene and Alpha-isomethyl ionone were qualitatively detected, but unquantified in WWTPs effluents in Henares River located in the Tajo River Basin, Spain [151]. Similarly, other fragrances have been detected at low concentrations in the aquatic environments including lakes, rivers, and coastal systems [150,152–154]. For example, Alvarez et al. [152] reported limonene concentrations ranging from undetectable limits to 46 ng/L in the coastal waters of San Francisco Bay and the Southern California Bight, USA.

Although certain fragrances are pseudo-persistent in the aquatic environments through continuous release from technical systems (e.g., WWTPs), however, evidence point to their effective removal through wastewater treatment processes. For example, Lillial was detected at low concentrations of 10.0 to 57.0 ng/L in German WWTPs, and the subsequent receiving waters [147]. In addition, hexylcinnamaldehyde was measured at high concentration of 10.0 µg/L in the influent; but was reduced to <10.0 ng/L (>99.99% removal efficiency) in the effluent [147]. Hence, WWTPs can achieve high removal efficiencies (≥90%) for certain fragrances in efficient and effective functioning treatment systems.

Alpha-isomethyl ionone (AIM) is widely used in product cosmetics, toiletry products, household cleaners, and detergents in excess of 100 t annually [155]. At present, AIM was identified as active ingredient in sanitizer brands commercialized in South African although they have been banned in certain jurisdictions. Alpha-ionone was also identified as an active ingredient in sanitizers; but no detected concentrations have been reported in the literature. Measured concentration of ionones (only for beta-ionone was detected in two WTPs (XWTP and SWTP) in S City (subtropical maritime climate), Guangdong Province, China in the ranges of 7.90 to 28.5 ng/L in raw water; but none was detected in the effluent [156]. The removal of ionones is associated to its hydrophobic character, and ease of degradation through oxidation, heating, and irradiation processes [157].

Limited studies have documented the effects of organic fragrances as demonstrated by results depicted in Fig. 6b for different taxa in the aquatic environment. Benzyl salicylate has EC<sub>50</sub> of 2.80 µg/L, and NOEC and lowest observed effects concentration (LOEC) of 0.1 and 1 µg/L (48 h) on copepod *Acartia tonsa*, respectively [158]. These concentrations are within the range detected in surface water systems such as river water (Fig. 6a). Therefore, high use of sanitizers and disinfectants may exacerbate the adverse effects of Benzyl salicylate in the environment. Effects studies of citral on fish, daphnia, and algae yielded were 6.78, 6.8, and 103.8 mg/L, respectively (Fig. 6b), thus being toxic to fish and daphnia. Notably, these values are above the reported EC<sub>50s</sub> for fragrances (Fig. 6b). However, according to the classification of chemicals, herein most fragrances had their effects in the order of harmful to toxic. As such, more studies are required to ascertain their effects on aquatic species, particularly at sub-lethal effects where apical effects would not be observable but may induce deleterious effects particularly at molecular level with long-term impact to ecological integrity.

The 72 h EC<sub>50</sub> toxicity of AIM were 2.89 (under growth phase), 3.23 (number of cells) and 7.47 mg/L (growth rate) for algae, whilst the NOEC for algae was 0.404 mg/L. Similarly, fish 96 h EC<sub>50</sub> of 10.9 mg/L and 1.9 mg/L for *Orochlorhynchus mykiss* and zebrafish, respectively [159] have



**Fig. 6.** (a) Occurrence of chemicals classified as fragrances in sanitizers and disinfectants in variant environmental matrices (WWTP influent (full green circles) and effluent (open green circles), surface water (blue), river water (red), tap water (orange), lake water (yellow), marine water (purple) and groundwater (brown). And (b) their E(L)C<sub>50</sub>s tested on fish (blue), algae (green) daphnia (orange) and bacteria (purple) at different exposures (3 h (open triangles), 17 h (full triangles), 24 h (full squares), 48 h (open circles), 72 h (open squares) and 96 h (full circles).

been observed. Other fragrances such as Limonene showed concentration-dependent effects on the growth inhibition and photosynthetic abilities of the algae *Chlorella vulgaris*, but no ECs were provided [160].

### 3.6. Emulsifiers

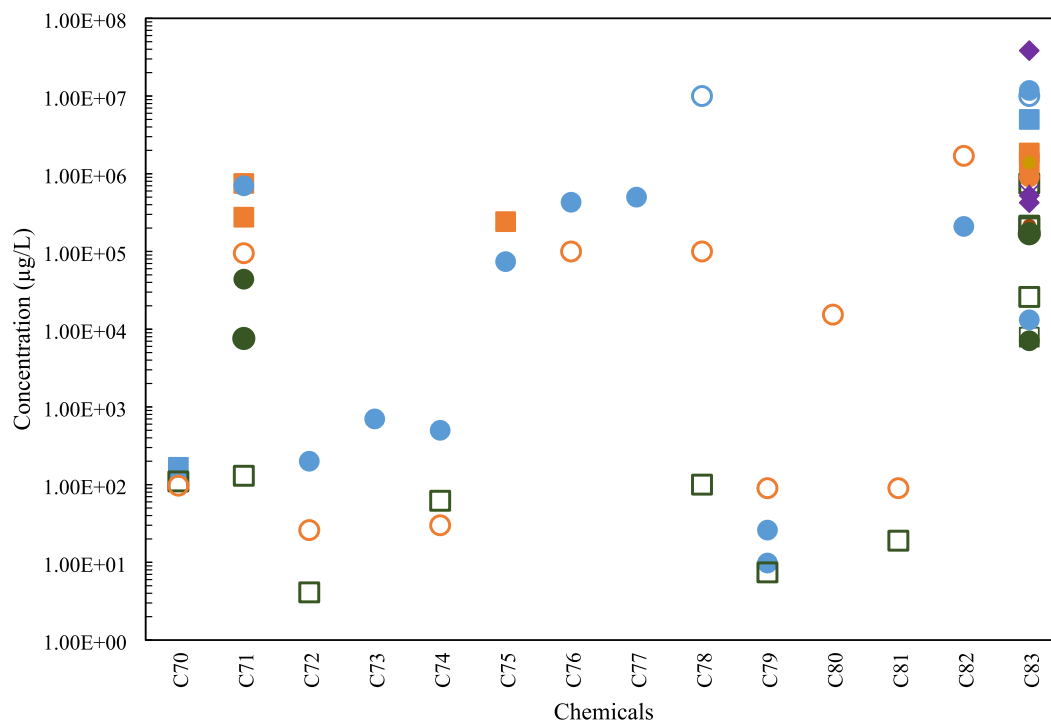
Emulsifiers consist of polymers and proteins among other chemical classes, and are essential stabilizers utilized in formulations that require emulsion, and exhibits both hydrophobic and hydrophilic properties [161,162]. According to Tcholakova and colleagues [163] the physicochemical properties of emulsifiers are dependent on the genesis of the emulsifier in question. As such, emulsions are predominantly formulated using an amalgamation of emulsifiers to improve their functionality [164].

Numerous studies have demonstrated that use of an amalgamation of emulsifiers to form emulsion-based antimicrobials has the potential to enhance antimicrobial activity and physical stability [165,166]. At present, however, data on the environmental occurrence and toxicity of emulsifiers are largely lacking, and/or defined by numerous data deficits, but also

highly contradictory [167]. This, in turn, limits our collective ability to estimate the ecological risk of emulsifiers in different environmental matrices.

The data retrieved from the literature indicates triethanolamine as the most investigated emulsifier with over 20 E(L)C<sub>50</sub>s (41%) across the three taxa (Fig. 7). Further, data showed triethanolamine to be toxic with E(L)C<sub>50</sub>s of 7.10 mg/L to *Scenedesmus subspicatus* following 48 h exposure [168]. However, other works have ranked the chemical as non-toxic to all taxa [167,169,170]. Results in Fig. 7 also show that cetrimonium bromide and polymeric biguanide hydrochloride are highly toxic to algae with E(L)C<sub>50</sub>s of 4.00 mg/L and 19.0 µg/L, respectively [171].

About two decades ago, Tişler and colleagues highlighted lack of documentation on the ecological toxicity data for emulsifiers (e.g., cetrimonium bromide), and our review results indicate these knowledge gaps have remained largely undressed although emulsifiers are widely used in numerous product categories. Yet, other studies have categorized cetrimonium bromide as extremely toxic to aquatic organisms [172]. Toxicity data for the emulsifiers are characterized by contradictions. An example of data



**Fig. 7.**  $E(L)C_{50}$  of chemicals classified as emulsifiers in sanitizers and disinfectants values tested on fish (blue), algae (green) daphnia (orange) and bacteria (purple) at different exposures (0.083 h (asterisk), 0.25 h (open diamond), 0.5 (full diamonds) 24 h (full squares), 48 h (open circles), 72 h (open squares) and 96 h (full circles).

contradiction is that the United States Environmental Protection Agency [173] classified polymeric biguanide hydrochloride among chemicals of low environmental risks. However, recent findings have demonstrated biguanide hydrochloride to be toxic to *Biomphalaria glabrata* (freshwater snails) at all different growth stages [174].

#### 4. Environmental risk characterization

To gain an understanding on the likely implications of chemicals incorporated in sanitizers and disinfectants on different environmental systems. Here, a chemical risk was deterministically calculated in surface waters, lakes, and rivers. For each chemical, its ecological risk was determined for three taxa: fish, algae, and daphnia by calculating the risk quotient (RQ). RQ is a ratio of measured environmental concentration (MEC) to the predicted no effect concentration (PNEC). PNEC was determined by dividing  $E(L)C_{50}$  values with an assessment factor (AF). The AF aids to account for inter- and intra-species effects variability. For each chemical, the PNEC was calculated using the least toxicity among the three taxa (fish, algae, or daphnia) [175]. To identify the hot spots (i.e., waters with increased risk), the retrieved occurrence data for each chemical was used (as MEC), and the results are summarized in Fig. 8.

Owing to lack of ecotoxicity chronic effects datasets for a large portion of chemicals reviewed herein, acute toxicity was used to determine the PNECs. Since PNEC was determined using acute toxicity an  $AF = 1000$  [175] was used. For chemicals with published  $E(L)C_{50}$  values, calculated PNEC ranges for the fish, daphnia, and algae taxonomical groups were 0.03–5000, 0.001–10,000, and 0.11–19,000  $\mu\text{g/L}$ , respectively. Low PNECs suggest that even at low MECs, the MEC in question may exceed the threshold toxicity, and in turn, pose risk to the aquatic organisms. Conversely, very high PNECs signify a chemical does not pose risk to aquatic organisms unless released at very high concentration(s) into the aquatic environments.

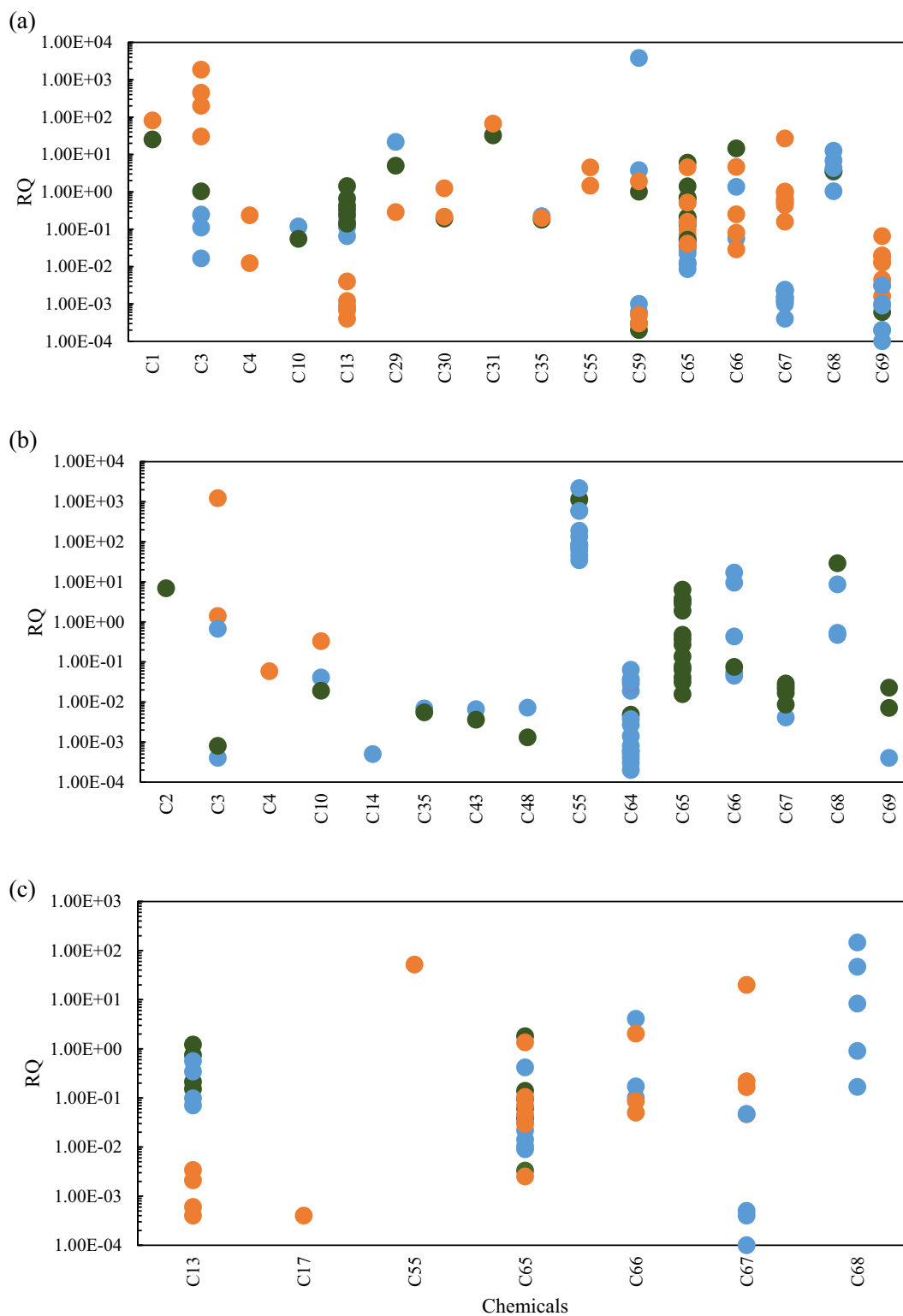
According to Technical Guideline Document on risk assessment [176], if  $RQ \geq 1$ , risk is considered unacceptable for the aquatic habitat; whereas if  $RQ < 1$  no risk is posed to aquatic organisms. Here, framework by Lemly [177] for risk classification was employed, and consists of four classes, thus:

$RQ < 0.1$ ,  $0.1 \leq RQ < 1$ ,  $1 \leq RQ < 10$ , and  $RQ \geq 10$  signifying none, low, moderate, and high risks, respectively. Results summarized in Fig. 8 indicate risks of chemicals (having both toxicity and occurrence data) ranked from none to very high. As the occurrence data used in the model were MECs for specific locations, RQ values  $> 1$  indicate moderate to high risks of chemicals in question to the aquatic environment. These results are important as can aid to prioritize chemicals of concern using the derived risk scores, and secondly, identify hot spots/hot events.

Using the safety threshold of  $RQ \geq 10$  for a chemical risk to be ranked as high – priority of chemicals of concern were identified per category (e.g., fragrances, UV filters, etc.). These spanned in various classes including antimicrobials (4-chloro-m-cresol, benzalkonium chloride, and sodium ortho phenyl phenate in surface water), solvents (1,2-propanediol in surface water), oxidants and antioxidants (hydrogen peroxide in lakes and rivers), and UV filters (4-Methylbenzilidene-camphor, ethylhexyl methoxy cinnamate, and octocrylene) across all three compartments. This assumption is justifiable because most published ecotoxicity data are highly variable, and broadly, reported observed effects are at individual level for the taxonomical group on focus.

This is because individual effects are not necessarily reflective of population-level effects as the later are more important as they offer insights into realistic risks of a given chemical to the environment. The toxicity numerous endpoints (e.g., growth, reproduction, hatching rate, etc.) in which  $E(L)C_{50}$  values were determined and used here; however, in most cases we could not ascertain how the actual experiments were conducted – as this information was not documented in many reviewed articles. For example, it was not possible to establish if the toxicity retrieved from the literature reflected population-level effects; therefore, the threshold RQ for high risk was set at  $\geq 10$ . This approach, however, should be used with caution. The reason is because it reflects only the current state-of-knowledge on exposure and effects of the chemicals with available data, and therefore, likely to be refined in future as more occurrence and toxicological data become accessible.

Results listed in Fig. 8 demonstrate daphnia as the most sensitive species as evidenced by high RQ values across different chemical classes. This does not imply that under all circumstances, the daphnia will be the most



**Fig. 8.** Summary of risk characterization of sanitizers and disinfectants using measure concentrations in the aquatic environments: (a) surface water, (b) river water, and (c) lake water across the globe relative to no-effect concentrations for fish (blue), algae (green) and daphnia (orange) taxonomical groups.

affected organism. Rather, use of PNECs for specific taxonomical groups offer a practical approach owing to its high variability driven by chemical composition (both spatial and temporal) in a given aquatic environment, exposure variability based on changes in concentration as well as underlying drivers of the observed toxicity for specific taxa (e.g., age, trophic level, mode of action, experimental setup, etc.).

Of the 22 chemicals with data where RQs were calculated, nine had their risks ranked as *none* and/or *low* ( $RQ < 1$ ) across three aquatic environments, and taxonomical groups. These chemicals were antimicrobials (benzothiazoline, chloroxylenol, ethanol, glutaraldehyde, and isopropanol), oxidants and antioxidants (hydrogen peroxide), fragrances (benzyl salicylate, eugenol, and linalool), and UV-filters (B-4). Notably, all fragrances

with occurrence and effects data exhibited no risks to the aquatic organisms. In addition, nine chemicals in different classes, namely: antimicrobials (4-chloro-m-cresol benzalkonium chloride, and sodium ortho phenyl phenate), oxidants and antioxidants (hydrogen peroxide), solvents (1,2-propanediol), emulsifiers (didecylammonium chloride), and UV-filters (4-Methyl-benzilidene-camphor, ethylhexyl methoxy cinnamate, and octocrylene) posed high risks to different taxonomical groups as RQ values were >10 (Fig. 8).

Therefore, these nine chemicals were identified as of high priority based on their risk scores. Authors acknowledge that due to limited occurrence and toxicity data, environmental risks of most chemicals (78.8%) identified in the five classes (78.8% of total 99 chemicals) were indeterminable. This means the top list of chemicals of concern are likely to be higher as data become accessible. Furthermore, to fill these information deficits for these chemicals and generate data to aid their risk assessment and management in freshwater systems; we propose use of modelling approaches. For example, occurrence data can be derived using material flow analysis approaches to derive the predicted environmental concentrations, whereas *in silico* techniques (e.g., quantitative structure-activity relationships (QSARs), etc. can be used to generate environmental toxicity effects to diverse taxa across variant compartments and taxonomy. This is because modelling approaches at present are well established and their derived data are accepted for legislative functions e.g., jurisdictions in the USA and European Union. These models have been applied successfully for risk assessment of chemicals incorporated in pharmaceuticals and personal care products. The modelling approaches can thus be used to carry out rapid first tier risk assessment of chemicals incorporated in sanitizers and disinfectants. The proposed approaches are neither tedious, costly, nor time consuming as well as eliminates use and sacrificing of numerous organisms unlike the experimental-based approaches.

In South Africa, only a single study has reported UV filters occurrence in the aquatic environment [178]. Their calculated aquatic risks for benzophenones (with MEC based on combined concentrations of BP-1, BP-3, and BP-4 in river water) had RQ of 0.07, and effects data were determined using the ECOSAR model. This RQ value for South Africa is very low relative to most values obtained for individual benzophenones of BP-3 and BP-4 results listed in Fig. 8. This is likely due to differences in UV filters consumption patterns per country or region, standards of living, wastewater treatment efficacies, and geographical loci across the globe especially based on likely dilution of the pollutants in the environment.

## 5. Concluding remarks

Published literature indicates most chemicals widely incorporated in sanitizers and disinfectants, are used in other product categories. As a result, they are highly ambiguous in different aquatic environments both in the natural (lakes, rivers, seas/oceans), and engineered matrixes (wastewater, tap water) with largely potential consequences to humans and the environment. Many of these chemicals are of environmental concern as their concentrations vary greatly from low (<1 ng/L) to very high (>100 µg/L), and therefore, are pseudo-persistent owing to their continuous release into the environment – although their long-term consequences remain largely unquantified.

This review has also dealt with the ecotoxicity of these chemicals to taxonomical groups in natural and engineered systems. Results demonstrate the effects are dependent on factors including taxa under consideration, their habitat (freshwater, marine, etc.), experimental setup, chemical in question, life-cycle phase of exposure organism (eggs, juveniles, etc.), among others. Moreover, most chemicals lacked chronic data, and even a large portion had neither acute nor chronic data. Further, it is unclear whether chemicals in sanitizers and disinfectants may be transferred from the aquatic environments to humans because of biomagnification through water and food chain pathways. This is urgent owing to their wide use in sanitizers and disinfectants especially in response to COVID-19, but also since many were already at elevated environmental concentrations above toxicity threshold(s) to variant aquatic taxonomical groups even during

pre-pandemic period. Yearly, large tonnages of these chemicals are discharged into the environment; therefore, both parent and/or their TPs can induce deleterious effects especially at sub-lethal level, though at present remain unquantified.

Of the chemicals assessed benzalkonium chloride, 4-chloro-m-cresol, sodium ortho phenyl phenate, hydrogen peroxide, 1,2-propanediol, 4-Methyl-benzilidene-camphor, didecylammonium chloride, ethylhexyl methoxy cinnamate, and octocrylene posed high risks to different taxa in the aquatic environment. Several of these chemicals are persistent in the environment even when removed via WWTPs to very low concentrations at ng/L levels. This point to the importance of controlling their flows, e.g., through improved treatment efficacies, and consideration of use of substitutes developed using green chemistry-based alternatives among other approaches to mitigate subtle environmental consequences.

From the reviewed literature, most occurrence studies were in the Organization for Economic Co-operation Development (OECD) countries and China, and least in South America and Africa. This raises the need to monitor and detect constituent chemicals incorporated in sanitizers and disinfectants in the environment with emphasis in the later countries. Second, there is need to achieve a fine balance between the benefits of incorporating certain chemicals in products against their risks. Such data should be used as a guiding principle in regulatory framework, but also to consider banning certain chemicals. Thirdly, there is research gap that need redress especially to develop greener chemicals for use in sanitizers and disinfectants. For example, increasing release of antimicrobial agents e.g., chlorhexidine and quaternary ammonium compounds may induce emergence of tolerant, resistant, and cross-resistant microbial strains even at sub-inhibitory concentrations.

Finally, a national database of chemicals used in domestic and industrial applications is existentially beneficial if a holistic and robust risk assessment valuable to achieve effective chemicals management in the environment. For instance, due diligence was done to identify chemicals reviewed herein incorporated in sanitizers and disinfectants in South Africa commerce. However, due to lack of chemicals national database it was improbable to ascertain whether all major chemicals were included in this review. This drawback can be remedied by the development and maintaining of widely accessible national databases of variant categories of products, and their constituent chemicals.

## Declaration of Competing Interest

The authors declare that they have no known competing financial or personal interests that plausibly appeared to influence the work on this paper.

## Acknowledgements

The authors are grateful for the financial assistance provided by the Water Research Commission (South Africa, grant number C2020/2021\_00387), and Department of Science and Innovation-National Research Foundation-South African Research Chair Initiative (DSI-NRF SARCHI) programme, grant number 91230. The authors wish to acknowledge the Universities of Pretoria, and Johannesburg, Department of Science and Innovation/National Research Foundation-South African Research Chair Initiative.

## Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.enceco.2023.01.003>.

## References

- [1] D. Pradhan, P. Biswasroy, P. Kumar Naik, G. Ghosh, G. Rath, A review of current interventions for COVID-19 prevention, *Arch. Med. Res.* 51 (2020) 363–374, <https://doi.org/10.1016/j.arcmed.2020.04.020>.

- [2] M.H. Al-Sayah, Chemical disinfectants of COVID-19: an overview, *J. Water Health* 18 (2020) 843–848, <https://doi.org/10.2166/wh.2020.108>.
- [3] A. Daverey, K. Dutta, COVID-19: eco-friendly hand hygiene for human and environmental safety, *J. Environ. Chem. Eng.* 9 (2021), 104754, <https://doi.org/10.1016/j.jece.2020.104754>.
- [4] Future Market Insights, Hand Sanitizer Market, (n.d.). <https://www.futuremarketinsights.com/reports/hand-sanitizer-market> (accessed January 4, 2023).
- [5] Markets and Markets, Surface Disinfectant Market - Global Growth Drivers & Opportunities, Markets and Markets. (n.d.). <https://www.marketsandmarkets.com/Market-Reports/surface-disinfectant-market-231286043.html> (accessed January 4, 2023).
- [6] K. Choi, S. Sim, J. Choi, C. Park, Y. Uhm, E. Lim, A.Y. Kim, S.J. Yoo, Y. Lee, Changes in handwashing and hygiene product usage patterns in Korea before and after the outbreak of COVID-19, *Environ. Sci. Eur.* 33 (2021) 79, <https://doi.org/10.1186/s12302-021-00517-8>.
- [7] A. Berardi, D.R. Perinelli, H.A. Merchant, L. Bisharat, I.A. Basheti, G. Bonacucina, M. Cespi, G.F. Palmieri, Hand sanitizers amid CoViD-19: a critical review of alcohol-based products on the market and formulation approaches to respond to increasing demand, *Int. J. Pharm.* 584 (2020), 119431 <https://doi.org/10.1016/j.ijpharm.2020.119431>.
- [8] S.L. Nason, E. Lin, B. Eitzer, J. Koelmel, J. Peccia, Changes in sewage sludge chemical signatures during a COVID-19 community lockdown, part 1: traffic, drugs, mental health, and disinfectants, *Environ. Toxicol. Chem.* 41 (2022) 1179–1192, <https://doi.org/10.1002/etc.5217>.
- [9] J.L.J. Jing, T. Pei Yi, R.J.C. Bose, J.R. McCarthy, N. Tharmalingam, T. Madheswaran, Hand sanitizers: a review on formulation aspects, adverse effects, and regulations, *Int. J. Environ. Res. Public Health* 17 (2020), E3326 <https://doi.org/10.3390/ijerph17093326>.
- [10] D. Koenig, W.S. Martin, The cutting edge of clean: understanding residual sanitization and disinfection, *Knowl. Libr.* (2021) 1–13 [http://community.ifma.org/knowledge\\_library/m/members\\_only\\_content/1058537](http://community.ifma.org/knowledge_library/m/members_only_content/1058537) (accessed October 28, 2022).
- [11] G. Kampf, D. Todt, S. Pfaender, E. Steinmann, Persistence of coronaviruses on inanimate surfaces and their inactivation with biocidal agents, *J. Hosp. Infect.* 104 (2020) 246–251, <https://doi.org/10.1016/j.jhin.2020.01.022>.
- [12] J. Hartmann, S. Wuijts, J.P. van der Hoek, A.M. de Roda Husman, Use of literature mining for early identification of emerging contaminants in freshwater resources, *Environ. Evid.* 8 (2019) 33, <https://doi.org/10.1186/s13750-019-0177-z>.
- [13] A.J. Ebele, M. Abou-Elwafa Abdallah, S. Harrad, Pharmaceuticals and personal care products (PPCPs) in the freshwater aquatic environment, *Emerg. Contam.* 3 (2017) 1–16, <https://doi.org/10.1016/j.emcon.2016.12.004>.
- [14] D.R. Joshi, N. Adhikari, An overview on common organic solvents and their toxicity, *J. Pharm. Res. Int.* (2019) 1.
- [15] K. Grodowska, A. Parczewski, Organic solvents in the pharmaceutical industry, *Acta Pol. Pharm.* 67 (2010) 3–12.
- [16] C.J. Clarke, W.-C. Tu, O. Levers, A. Bröhl, J.P. Hallett, Green and sustainable solvents in chemical processes, *Chem. Rev.* 118 (2018) 747–800, <https://doi.org/10.1021/acs.chemrev.7b00571>.
- [17] H. Gulyas, M. Reich, Organic constituents of oil reclaiming wastewater, *J. Environ. Sci. Health Part A*. 35 (2000) 435–464, <https://doi.org/10.1080/10934520009376981>.
- [18] L. Kong, K. Kadokami, H.T. Duong, H.T.C. Chau, Screening of 1300 organic micropollutants in groundwater from Beijing and Tianjin, North China, *Chemosphere*. 165 (2016) 221–230, <https://doi.org/10.1016/j.chemosphere.2016.08.084>.
- [19] O.V. Poliakov, A.T. Lebedev, O. Hänninen, Organic pollutants in snow of urban and rural Russia and Finland, *Toxicol. Environ. Chem.* 75 (2000) 181–194, <https://doi.org/10.1080/02772240009358903>.
- [20] J. Wang, Z. Tian, Y. Huo, M. Yang, X. Zheng, Y. Zhang, Monitoring of 943 organic micropollutants in wastewater from municipal wastewater treatment plants with secondary and advanced treatment processes, *J. Environ. Sci. China*. 67 (2018) 309–317, <https://doi.org/10.1016/j.jes.2017.09.014>.
- [21] B.-Z. Wu, T.-Z. Feng, U. Sree, K.-H. Chiu, J.-G. Lo, Sampling and analysis of volatile organics emitted from wastewater treatment plant and drain system of an industrial science park, *Anal. Chim. Acta* 576 (2006) 100–111, <https://doi.org/10.1016/j.aca.2006.03.057>.
- [22] S. Lee, C. Liao, G.-J. Song, K. Ra, K. Kannan, H.-B. Moon, Emission of bisphenol analogues including bisphenol A and bisphenol F from wastewater treatment plants in Korea, *Chemosphere*. 119 (2014) 1000–1006, <https://doi.org/10.1016/j.chemosphere.2014.09.011>.
- [23] H. Mehlhorn, Blood sucking and chewing lice, in: N. Rezaei (Ed.), *Encycl. Infect. Immun.*, Elsevier, Oxford 2022, pp. 994–1014, <https://doi.org/10.1016/B978-0-12-818731-9.00012-4>.
- [24] G. Avery, L. Foley, A. Carroll, A. Roebuck, A. Guy, R. Mead, R. Kieber, J. Willey, S. Skrabal, J. Felix, K. Mullaugh, J. Helms, Surface waters as a sink and source of atmospheric gas phase ethanol, *Chemosphere*. 144 (2015) 360–365, <https://doi.org/10.1016/j.chemosphere.2015.08.080>.
- [25] A. Mahmood, M. Egan, S. Pervez, H.A. Alghamdi, A.B. Tabinda, A. Yasar, K. Brindhadevi, A. Pugazhendhi, COVID-19 and frequent use of hand sanitizers; human health and environmental hazards by exposure pathways, *Sci. Total Environ.* 742 (2020), 140561 <https://doi.org/10.1016/j.scitotenv.2020.140561>.
- [26] V.K. Pal, S. Lee, M. Naidu, C. Lee, K. Kannan, Occurrence of and dermal exposure to benzene, toluene and styrene found in hand sanitizers from the United States, *Environ. Int.* 167 (2022), 107449 <https://doi.org/10.1016/j.envint.2022.107449>.
- [27] R.D. Sills, P.A. Blakeslee, The environmental impact of deicers in airport stormwater runoff, *Chemical Deicers and the Environment*, 1992 <https://trid.trb.org/view/363861>, (accessed October 28, 2022).
- [28] M.A. Rasheed, M. Lakshmi, M.S. Kalpana, D.J. Patil, A.M. Dayal, Recognition of hydrocarbon microseepage using microbial and adsorbed soil gas indicators in the petroliferous region of Krishna–Godavari Basin, India, *Curr. Sci.* 112 (2017) 560–568.
- [29] W.W. Johnson, M.T. Finley, *Handbook of Acute Toxicity of Chemicals to Fish and Aquatic Invertebrates: Summaries of Toxicity Tests Conducted at Columbia National Fisheries Research Laboratory, 1965–78*, U.S. Fish and Wildlife Service, 1980 <https://pubs.er.usgs.gov/publication/rp137> (accessed October 26, 2022).
- [30] D. Scheerbaum, Metazachlor—Alga, growth inhibition test with *Scenedesmus subspicatus*, 72h, ReportSso68811 Lab. Für Angew. Biol. Sarstedt Ger, 2000.
- [31] G.W. Stratton, C.T. Corke, Toxicity of the insecticide permethrin and some degradation products towards algae and cyanobacteria, *Environ. Pollut. Ser. Ecol. Biol.* 29 (1982) 71–80, [https://doi.org/10.1016/0143-1471\(82\)90055-1](https://doi.org/10.1016/0143-1471(82)90055-1).
- [32] V. Mattson, R.J. Arthur, W.C. Walbridge, Acute toxicity of selected organic compounds to fathead minnows, Environmental Protection Agency, Office of Research and Development, Environmental Research Laboratory, 1976 [https://cfpub.epa.gov/si/si\\_public\\_record\\_report.cfm?Lab=NHEERL&dirEntryId=43246](https://cfpub.epa.gov/si/si_public_record_report.cfm?Lab=NHEERL&dirEntryId=43246) (accessed October 26, 2022).
- [33] Smart-Lab MSDS 040 Indonesia, MSDS\_BENZYL\_ALCOHOL.pdf, [https://smartlab.co.id/assets/pdf/MSDS\\_BENZYL\\_ALCOHOL.pdf](https://smartlab.co.id/assets/pdf/MSDS_BENZYL_ALCOHOL.pdf) 2017 (accessed October 28, 2022).
- [34] Henkel MSDS 157262 V1.3 1 Australia, 00000000000378302\_000000173014\_MSDS\_UT\_US\_EN.PDF, 2021 [https://mysds.henkel.com/SAP\\_GATEWAY/odata/SAP/YPSSWH\\_DOO\\_SRV//DocContentSet\(Appid='YPSSW\\_SDSUA',Matnr='00000000000378302',Matnrcomp='','Subid='000000173014',Subidcomp='','Sbglvid='MSDS\\_UT\\_US',Laiso='EN'\)/DocContentData/\\$value](https://mysds.henkel.com/SAP_GATEWAY/odata/SAP/YPSSWH_DOO_SRV//DocContentSet(Appid='YPSSW_SDSUA',Matnr='00000000000378302',Matnrcomp='','Subid='000000173014',Subidcomp='','Sbglvid='MSDS_UT_US',Laiso='EN')/DocContentData/$value).
- [35] ECHA, Registration Dossier - ECHA, <https://echa.europa.eu/da/registration-dossier/-/registered-dossier/14748/6/3> 2021 (accessed October 26, 2022).
- [36] SIAM 22, SIDS Initial Assessment Report, Amine Oxides, [https://www.cleaninginstitute.org/sites/default/files/research-pdfs/SIDS\\_Amine\\_Oxides.pdf](https://www.cleaninginstitute.org/sites/default/files/research-pdfs/SIDS_Amine_Oxides.pdf) 2006 (accessed October 28, 2022).
- [37] USEPA, Preliminary Data Summary: Airport Deicing Operations (Revised), Office of Water, United States Environmental Protection Agency, Washington, DC, EPA-821-R-00-016, 2000 <https://www.epa.gov/sites/default/files/2015-06/documents/airport-deicing-pds-2000.pdf> (accessed November 4, 2022).
- [38] A. Maki, Correlations between *Daphnia magna* and fathead minnow (*Pimephales promelas*) chronic toxicity values for several classes of test substances, *J. Fish. Res. Board Can.* 36 (1979) 411–421, <https://doi.org/10.1139/f79-061>.
- [39] A.L. Bridie, C.J.M. Wolff, M. Winter, The acute toxicity of some petrochemicals to goldfish, *Water Res.* 13 (1979) 623–626, [https://doi.org/10.1016/0043-1354\(79\)90010-1](https://doi.org/10.1016/0043-1354(79)90010-1).
- [40] R. West, M. Banton, J. Hu, J. Klapacz, The distribution, fate, and effects of propylene glycol substances in the environment, *Rev. Environ. Contam. Toxicol.* 232 (2014) 107–138, [https://doi.org/10.1007/978-3-319-06746-9\\_5](https://doi.org/10.1007/978-3-319-06746-9_5).
- [41] ATSDR, Toxicological Profile for Ethylene Glycol, 1997 305.
- [42] E. Perales, J.L. García, E. Pires, L. Aldea, L. Lomba, B. Giner, Ecotoxicity and QSAR studies of glycerol ethers in *Daphnia magna*, *Chemosphere*. 183 (2017) 277–285, <https://doi.org/10.1016/j.chemosphere.2017.05.107>.
- [43] R. Mishra, S. Bisht, Antioxidants and their characterization 44 (2011) 91–680.
- [44] B. Daramola, G.O. Adegoke, Chapter 25 - bitter kola (*Garcinia kola*) seeds and health management potential, in: V.R. Preedy, R.R. Watson, V.B. Patel (Eds.), *Nuts Seeds Health Dis. Prev.*, Academic Press, San Diego 2011, pp. 213–220, <https://doi.org/10.1016/B978-0-12-375688-6.10025-8>.
- [45] D.B. Hermund, Antioxidant properties of seaweed-derived substances, *Bioact. Seaweeds Food Appl. - Nat. Ingred. Healthy Diets*, Academic Press 2018, pp. 201–221.
- [46] B. Halliwell, Antioxidants: the basics-what they are and how to evaluate them, in: H. Sies (Ed.), *Adv. Pharmacol.*, Academic Press 1996, pp. 3–20, [https://doi.org/10.1016/S1054-3589\(08\)60976-X](https://doi.org/10.1016/S1054-3589(08)60976-X).
- [47] M.J. Hopwood, I. Rapp, C. Schlosser, E.P. Achterberg, Hydrogen peroxide in deep waters from the Mediterranean Sea, South Atlantic and South Pacific Oceans, *Sci. Rep.* 7 (2017) 43436, <https://doi.org/10.1038/srep43436>.
- [48] M. Nowak, Z. Zawadzka, K. Lisowska, Occurrence of methylisothiazolinone in water and soil samples in Poland and its biodegradation by *Phanerochaete chrysosporium*, *Chemosphere*. 254 (2020), 126723, <https://doi.org/10.1016/j.chemosphere.2020.126723>.
- [49] C. Pajjens, A. Bressy, B. Frère, D. Tedoldi, R. Mailler, V. Rocher, P. Neveu, R. Moilleron, Urban pathways of biocides towards surface waters during dry and wet weathers: assessment at the Paris conurbation scale, *J. Hazard. Mater.* 402 (2021), 123765, <https://doi.org/10.1016/j.jhazmat.2020.123765>.
- [50] M.O. Sunday, W.A. Jadoon, T.T. Ayeni, Y. Iwamoto, K. Takeda, Y. Imaizumi, T. Arakaki, H. Sakugawa, Heterogeneity and potential aquatic toxicity of hydrogen peroxide concentrations in selected rivers across Japan, *Sci. Total Environ.* 733 (2020), 139349, <https://doi.org/10.1016/j.scitotenv.2020.139349>.
- [51] R.P. Eganhouse, I.R. Kaplan, alpha-Tocopheryl acetate as an indicator of municipal waste contamination in the environment, *Environ. Sci. Technol.* 19 (1985) 282–285, <https://doi.org/10.1021/es00133a014>.
- [52] W.J. Cooper, R.G. Zika, Photochemical formation of hydrogen peroxide in surface and ground waters exposed to sunlight, *Science*. 220 (1983) 711–712, <https://doi.org/10.1126/science.220.4598.711>.
- [53] K.M. Blum, P.L. Andersson, G. Renman, L. Ahrens, M. Gros, K. Wiberg, P. Haglund, Non-target screening and prioritization of potentially persistent, bioaccumulating and toxic domestic wastewater contaminants and their removal in on-site and large-scale sewage treatment plants, *Sci. Total Environ.* 575 (2017) 265–275, <https://doi.org/10.1016/j.scitotenv.2016.09.135>.
- [54] C. Burnett, W. Bergfeld, D. Belsito, C. Klaassen, R. Marks, S. Shank, S. Tj, A.A.F. Pw, Final report of the safety assessment of methylisothiazolinone, *Int. J. Toxicol.* 29 (2010), <https://doi.org/10.1177/1091581810374651>.
- [55] J. Fewings, T. Menné, An update of the risk assessment for methylchloroisothiazolinone/methylisothiazolinone (MCI/MI) with focus on rinse-off products, *Contact Dermatitis* 41 (1999) 1–13, <https://doi.org/10.1111/j.1600-0536.1999.tb06200.x>.

- [56] P.A. Taylor, An evaluation of the toxicity of various forms of chlorine to *Ceriodaphnia dubia*, Environ. Toxicol. Chem. 12 (1993) 925–930, <https://doi.org/10.1002/etc.5620120517>.
- [57] J.S. Mattice, S.C. Tsai, M.B. Burch, Comparative toxicity of hypochlorous acid and hypochlorite ions to mosquitofish, Trans. Am. Fish. Soc. 110 (1981) 519–525, [https://doi.org/10.1577/1548-8659\(1981\)110<519:CTOHA>2.0.CO;2](https://doi.org/10.1577/1548-8659(1981)110<519:CTOHA>2.0.CO;2).
- [58] J. Davidson, S. Summerfelt, D.L. Straus, K.K. Schrader, C. Good, Evaluating the effects of prolonged peracetic acid dosing on water quality and rainbow trout *Oncorhynchus mykiss* performance in recirculation aquaculture systems, Aquac. Eng. 84 (2019) 117–127, <https://doi.org/10.1016/j.aquaeng.2018.12.009>.
- [59] D. Liu, S. Behrens, L.-F. Pedersen, D.L. Straus, T. Meinelt, Peracetic acid is a suitable disinfectant for recirculating fish-microalgae integrated multi-trophic aquaculture systems, Aquac. Rep. 4 (2016) 136–142, <https://doi.org/10.1016/j.aqrep.2016.09.002>.
- [60] S.S. Block, Disinfection, Sterilization, and Preservation, Lippincott Williams & Wilkins, 2001.
- [61] S. Gad, Encyclopedia of Toxicology, ScienceDirect, 2014 <http://www.sciencedirect.com:5070/referencework/9780123864550/encyclopedia-of-toxicology> (accessed October 29, 2022).
- [62] D.L. Straus, T. Meinelt, D. Liu, L.-F. Pedersen, Toxicity of peracetic acid to fish: variation among species and impact of water chemistry, J. World Aquac. Soc. 49 (2018) 715–724, <https://doi.org/10.1111/jwas.12475>.
- [63] D.L. Straus, T. Meinelt, B.D. Farmer, B.H. Beck, Acute toxicity and histopathology of channel catfish fry exposed to peracetic acid, Aquaculture. 342–343 (2012) 134–138, <https://doi.org/10.1016/j.aquaculture.2012.02.024>.
- [64] L. Hicks, T. Ziegler, J. Bucksath, Acute toxicity of 5% peracetic acid (Vigor Ox) to *Selenastrum capricornutum* Printz, Unpublished report 42866, Princeton NJ, USA, 1996.
- [65] R.K. Chhetri, S. Di Gaetano, A. Turolla, M. Antonelli, H.R. Andersen, Ecotoxicity evaluation of pure peracetic acid (PAA) after eliminating hydrogen peroxide from commercial PAA, Int. J. Environ. Res. Public Health 17 (2020) 5031, <https://doi.org/10.3390/ijerph17145031>.
- [66] S. Gheorghie, D.N. Mitroi, M.S. Stan, C.A. Staicu, M. Cicirna, I.E. Lucaci, M. Nita-Lazar, A. Dinischiotu, Evaluation of sub-lethal toxicity of benzethonium chloride in *Cyprinus carpio* liver, Appl. Sci. 10 (2020) 8485, <https://doi.org/10.3390/app10238485>.
- [67] J. Stroh, M.T. Wan, M.B. Isman, D.J. Moul, Evaluation of the acute toxicity to juvenile Pacific coho salmon and rainbow trout of some plant essential oils, a formulated product, and the carrier, Bull. Environ. Contam. Toxicol. 60 (1998) 923–930, <https://doi.org/10.1007/s001289900716>.
- [68] R.R. Clayton, R.C. Summerfelt, Toxicity of hydrogen peroxide to fingerling walleyes, J. Appl. Aquac. 6 (1996) 39–49, [https://doi.org/10.1300/J028v06n03\\_04](https://doi.org/10.1300/J028v06n03_04).
- [69] eCA Germany, Regulation (EU) No 528/2012 concerning the making available on the market and use of biocidal products, Evaluation of active substances Assessment report L (+) Lactic acid, <https://echa.europa.eu/documents/10162/25096aa4-a3c7-bed9-a119-efd3a5f6cee9> 2017 (accessed October 31, 2022).
- [70] N.I. Thaddeus, E.C. Francis, O.O. Jane, A.C. Obunname, E.C. Okechukwu, Effects of some common additives on the antimicrobial activities of alcohol-based hand sanitizers, Asian Pac. J. Trop. Med. 11 (2018) 222, <https://doi.org/10.4103/1995-7645.228437>.
- [71] S.E. Walsh, J.-Y. Maillard, A.D. Russell, C.E. Catrenich, D.L. Charbonneau, R.G. Bartolo, Development of bacterial resistance to several biocides and effects on antibiotic susceptibility, J. Hosp. Infect. 55 (2003) 98–107, [https://doi.org/10.1016/s0195-6701\(03\)00240-8](https://doi.org/10.1016/s0195-6701(03)00240-8).
- [72] F. Chen, J. Moat, D. McFeely, G. Clarkson, I.J. Hands-Portman, J.P. Furner-Pardoe, F. Harrison, C.G. Dowson, P.J. Sadler, Biguanide iridium(III) complexes with potent antimicrobial activity, J. Med. Chem. 61 (2018) 7330–7344, <https://doi.org/10.1021/acs.jmedchem.8b00906>.
- [73] Y. Sun, Y. Zhang, Y. Xia, T. Fan, M. Xue, Enkhbayar Bulgan, C. Harnood, A. Dong, Evaluation of physicochemical properties and bactericidal activity of efficient chemical germicidal water (CGW), LWT - Food Sci. Technol. 59 (2014) 1068–1074, <https://doi.org/10.1016/j.lwt.2014.06.026>.
- [74] X. Fan, X. Ren, T.-S. Huang, Y. Sun, Cytocompatible antibacterial fibrous membranes based on poly(3-hydroxybutyrate-co-4-hydroxybutyrate) and quaternarized N-halalime polymer, RSC Adv. 6 (2016) 42600–42610, <https://doi.org/10.1039/C6RA08465F>.
- [75] P. Durani, D. Leaper, Povidone-iodine: use in hand disinfection, skin preparation and antiseptic irrigation, Int. Wound J. 5 (2008) 376–387, <https://doi.org/10.1111/j.1742-481X.2007.00405.x>.
- [76] G. McDonnell, Peroxygens and other forms of oxygen: their use for effective cleaning, disinfection, and sterilization, New Biocides Dev, American Chemical Society 2007, pp. 292–308, <https://doi.org/10.1021/bk-2007-0967.ch013>.
- [77] W.B. Hugo, Phenols: a review of their history and development as antimicrobial agents, Microbios. 23 (1978) 83–85.
- [78] S.J. Lim, C.-K. Seo, T.-H. Kim, S.-W. Myung, Occurrence and ecological hazard assessment of selected veterinary medicines in livestock wastewater treatment plants, J. Environ. Sci. Health B 48 (2013) 658–670, <https://doi.org/10.1080/03601234.2013.778604>.
- [79] B. Dwumfour-Asare, P. Adantey, K.B. Nyarko, E. Appiah-Effah, Greywater characterization and handling practices among urban households in Ghana: the case of three communities in Kumasi Metropolitan, Water Sci. Technol. 76 (2017) 813–822, <https://doi.org/10.2166/wst.2017.229>.
- [80] S.M. Choi, T.H. Roh, D.S. Lim, S. Kacew, H.S. Kim, B.-M. Lee, Risk assessment of benzalkonium chloride in cosmetic products, J. Toxicol. Environ. Health B Crit. Rev. 21 (2018) 8–23, <https://doi.org/10.1080/10937404.2017.1408552>.
- [81] L. del C. Velázquez, N.B. Barbini, M.E. Escudero, C.L. Estrada, A.M.S. de Guzmán, Evaluation of chlorine, benzalkonium chloride and lactic acid as sanitizers for reducing *Escherichia coli* O157:H7 and *Yersinia enterocolitica* on fresh vegetables, Food Control (2009), <https://doi.org/10.1016/j.foodcont.2008.05.012> (accessed October 27, 2022).
- [82] O.W. Barber, E.M. Hartmann, Benzalkonium chloride: a systematic review of its environmental entry through wastewater treatment, potential impact, and mitigation strategies, Crit. Rev. Environ. Sci. Technol. 52 (2022) 2691–2719, <https://doi.org/10.1080/10643389.2021.1889284>.
- [83] W.-L. Li, Z.-F. Zhang, C. Sparham, Y.-F. Li, Validation of sampling techniques and SPE-UPLC/MS/MS for home and personal care chemicals in the Songhua Catchment, Northeast China, Sci. Total Environ. 707 (2020), 136038, <https://doi.org/10.1016/j.scitotenv.2019.136038>.
- [84] I. Ferrer, E.T. Furlong, Accelerated solvent extraction followed by on-line solid-phase extraction coupled to ion trap LC/MS/MS for analysis of benzalkonium chlorides in sediment samples, Anal. Chem. 74 (2002) 1275–1280, <https://doi.org/10.1021/ac010969l>.
- [85] M. Östman, R.H. Lindberg, J. Fick, E. Björn, M. Tysklind, Screening of biocides, metals and antibiotics in Swedish sewage sludge and wastewater, Water Res. 115 (2017) 318–328, <https://doi.org/10.1016/j.watres.2017.03.011>.
- [86] A. Van de Voorde, C. Lorgeoux, M.-C. Gromaire, G. Chebbo, Analysis of quaternary ammonium compounds in urban stormwater samples, Environ. Pollut. 164 (2012) 150–157, <https://doi.org/10.1016/j.envpol.2012.01.037>.
- [87] B. Kasprzyk-Hordern, R.M. Dinsdale, A.J. Guwy, The removal of pharmaceuticals, personal care products, endocrine disruptors and illicit drugs during wastewater treatment and its impact on the quality of receiving waters, Water Res. 43 (2009) 363–380, <https://doi.org/10.1016/j.watres.2008.10.047>.
- [88] D. Montes-Grajales, M. Fennix-Agudelo, W. Miranda-Castro, Occurrence of personal care products as emerging chemicals of concern in water resources: a review, Sci. Total Environ. 595 (2017) 601–614, <https://doi.org/10.1016/j.scitotenv.2017.03.286>.
- [89] A.K. Fahimipour, S. Ben Maamar, A.G. McFarland, R.A. Blaustein, J. Chen, A.J. Glawe, J. Kline, J.L. Green, R.U. Halden, K. Van Den Wymelenberg, C. Huttenhower, E.M. Hartmann, Antimicrobial chemicals associate with microbial function and antibiotic resistance indoors, MSystems 3 (2018) e00200–e00218, <https://doi.org/10.1128/mSystems.00200-18>.
- [90] S. Buffet-Bataillon, P. Tattevin, M. Bonnaure-Mallet, A. Jolivet-Gougeon, Emergence of resistance to antibacterial agents: the role of quaternary ammonium compounds—a critical review, Int. J. Antimicrob. Agents 39 (2012) 381–389, <https://doi.org/10.1016/j.ijantimicag.2012.01.011>.
- [91] M.D. Johnston, E.A. Simons, R.J. Lambert, One explanation for the variability of the bacterial suspension test, J. Appl. Microbiol. 88 (2000) 237–242, <https://doi.org/10.1046/j.1365-2672.2000.00951.x>.
- [92] G. Nicoletti, V. Boghossian, F. Gurevitch, R. Borland, P. Morgenroth, The antimicrobial activity in vitro of chlorhexidine, a mixture of isothiazolinones (“Kathon” CG) and cetyl trimethyl ammonium bromide (CTAB), J. Hosp. Infect. 23 (1993) 87–111, [https://doi.org/10.1016/0195-6701\(93\)90014-q](https://doi.org/10.1016/0195-6701(93)90014-q).
- [93] H.W. Leung, Ecotoxicology of glutaraldehyde: review of environmental fate and effects studies, Ecotoxicol. Environ. Saf. 49 (2001) 26–39, <https://doi.org/10.1006/eesa.2000.2031>.
- [94] S.P.P. Pereira, R. Oliveira, S. Coelho, C. Musso, A.M.V.M. Soares, I. Domingues, A.J.A. Nogueira, From sub cellular to community level: toxicity of glutaraldehyde to several aquatic organisms, Sci. Total Environ. 470–471 (2014) 147–158, <https://doi.org/10.1016/j.scitotenv.2013.09.054>.
- [95] L. Guilhermino, T. Diamantino, M.C. Silva, A.M. Soares, Acute toxicity test with *Daphnia magna*: an alternative to mammals in the prescreening of chemical toxicity? Ecotoxicol. Environ. Saf. 46 (2000) 357–362, <https://doi.org/10.1006/eesa.2000.1916>.
- [96] J. McLaughlin, J.-C.J. Bonzongo, Effects of natural water chemistry on nanosilver behavior and toxicity to *Ceriodaphnia dubia* and *Pseudokirchneriella subcapitata*, Environ. Toxicol. Chem. 31 (2012) 168–175, <https://doi.org/10.1002/etc.720>.
- [97] H.-H. Sung, Y.-W. Chiu, S.-Y. Wang, C.-M. Chen, D.-J. Huang, Acute toxicity of mixture of acetaminophen and ibuprofen to Green Neon Shrimp, *Neocaridina denticulata*, Environ. Toxicol. Pharmacol. 38 (2014) 8–13, <https://doi.org/10.1016/j.etap.2014.04.014>.
- [98] N. Kreuzinger, M. Fuerhacker, S. Scharf, M. Uhl, O. Gans, B. Grillitsch, Methodological approach towards the environmental significance of uncharacterized substances — quaternary ammonium compounds as an example, Desalination. 215 (2007) 209.
- [99] M.-H. Li, Comparative toxicities of 10 widely used biocides in three freshwater invertebrate species, Chem. Ecol. 35 (2019) 472–482, <https://doi.org/10.1080/02757540.2019.1579311>.
- [100] Y. Sung, K.E. Fletcher, K.M. Ritalahti, R.P. Apkarian, N. Ramos-Hernández, R.A. Sanford, N.M. Mesbah, F.E. Löffler, *Geobacter lovleyi* sp. nov. strain SZ, a novel metal-reducing and tetrachloroethene-dechlorinating bacterium, Appl. Environ. Microbiol. 72 (2006) 2775–2782, <https://doi.org/10.1128/AEM.72.4.2775-2782.2006>.
- [101] D. Burgess, A. Forbis, Acute Toxicity of Oxonia Active to *Daphnia Magna*. Iowa, USA, unpublished report 552 report report 30724 <https://www.ecetoc.org/wp-content/uploads/2014/08/JACC-040.pdf> 1983 (accessed October 28, 2022).
- [102] CCR, Influence of Pirox 850 on the Reproduction of *Daphnia magna*. Project 164002, Cytotest Cell Research GmbH & Co., Rossdorf, Germany, 1990, <https://corpora.tika.apache.org/base/docs/govdocs1/128/128968.pdf>.
- [103] M. Lavorgna, C. Russo, B. D’Abrosca, A. Parrella, M. Isidori, Toxicity and genotoxicity of the quaternary ammonium compound benzalkonium chloride (BAC) using *Daphnia magna* and *Ceriodaphnia dubia* as model systems, Environ. Pollut. Barking Essex 1987 (210) (2016) 34–39, <https://doi.org/10.1016/j.envpol.2015.11.042>.
- [104] T. Madsen, H.B. Boyd, D. Nylén, A. Pedersen, G. Petersen, F. Simonsen, Environmental Health Assessment of Substances in Household Detergents and Cosmetic

- Detergent Products, <https://www2.mst.dk/udgiv/publications/2001/87-7944-596-9/pdf/87-7944-597-7.pdf> 2001 (accessed October 26, 2022).
- [105] M. Zhu, F. Ge, R. Zhu, X. Wang, X. Zheng, A DFT-based QSAR study of the toxicity of quaternary ammonium compounds on *Chlorella vulgaris*, *Chemosphere*. 80 (2010) 46–52, <https://doi.org/10.1016/j.chemosphere.2010.03.044>.
- [106] A. Utsunomiya, T. Watanuki, K. Matsushita, M. Nishina, I. Tomita, Assessment of the toxicity of linear alkylbenzene sulfonate and quaternary alkylammonium chloride by measuring 13C-glycerol in *Dunaliella* sp, *Chemosphere*. 35 (1997) 2479–2490, [https://doi.org/10.1016/S0045-6535\(97\)00316-0](https://doi.org/10.1016/S0045-6535(97)00316-0).
- [107] M.D. Hernando, M. Petrovic, A.R. Fernández-Alba, D. Barceló, Analysis by liquid chromatography-electrospray ionization tandem mass spectrometry and acute toxicity evaluation for beta-blockers and lipid-regulating agents in wastewater samples, *J. Chromatogr. A* 1046 (2004) 133–140.
- [108] Y. Yang, Y.S. Ok, K.-H. Kim, E.E. Kwon, Y.F. Tsang, Occurrences and removal of pharmaceuticals and personal care products (PPCPs) in drinking water and water/sewage treatment plants: a review, *Sci. Total Environ.* 596–597 (2017) 303–320, <https://doi.org/10.1016/j.scitotenv.2017.04.102>.
- [109] M. Bilal, S. Mehmood, H.M.N. Iqbal, The beast of beauty: environmental and health concerns of toxic components in cosmetics, *Cosmetics*. 7 (2020) 13, <https://doi.org/10.3390/cosmetics7010013>.
- [110] S. Kar, H. Sanderson, K. Roy, E. Benfenati, J. Leszczynski, Ecotoxicological assessment of pharmaceuticals and personal care products using predictive toxicology approaches, *Green Chem.* 22 (2020) 1458–1516, <https://doi.org/10.1039/C9GC03265G>.
- [111] A. Jurado, P. Gago-Ferrero, E. Vázquez-Suñé, J. Carrera, E. Pujades, M.S. Díaz-Cruz, D. Barceló, Urban groundwater contamination by residues of UV filters, *J. Hazard. Mater.* 271 (2014) 141–149, <https://doi.org/10.1016/j.jhazmat.2014.01.036>.
- [112] O. Golovko, V. Kumar, G. Fedorova, T. Randak, R. Grabic, Removal and seasonal variability of selected analgesics/anti-inflammatory, anti-hypertensive/cardiovascular pharmaceuticals and UV filters in wastewater treatment plant, *Environ. Sci. Pollut. Res.* 21 (2014) 7578–7585, <https://doi.org/10.1007/s11356-014-2654-9>.
- [113] K.H. Langford, M.J. Reid, E. Fjeld, S. Øxnevad, K.V. Thomas, Environmental occurrence and risk of organic UV filters and stabilizers in multiple matrices in Norway, *Environ. Int.* 80 (2015) 1–7, <https://doi.org/10.1016/j.envint.2015.03.012>.
- [114] C. Plagellat, T. Kupper, R. Furrer, L.F. de Alencastro, D. Grandjean, J. Tarradellas, Concentrations and specific loads of UV filters in sewage sludge originating from a monitoring network in Switzerland, *Chemosphere*. 62 (2006) 915–925, <https://doi.org/10.1016/j.chemosphere.2005.05.024>.
- [115] M.M.P. Tsui, H.W. Leung, P.K.S. Lam, M.B. Murphy, Seasonal occurrence, removal efficiencies and preliminary risk assessment of multiple classes of organic UV filters in wastewater treatment plants, *Water Res.* 53 (2014) 58–67, <https://doi.org/10.1016/j.watres.2014.01.014>.
- [116] K.I. Ekpeghere, U.-J. Kim, S.-H. O, H.-Y. Kim, J.-E. Oh, Distribution and seasonal occurrence of UV filters in rivers and wastewater treatment plants in Korea, *Sci. Total Environ.* 542 (2016) 121–128, <https://doi.org/10.1016/j.scitotenv.2015.10.033>.
- [117] S. Narla, H.W. Lim, Sunscreen: FDA regulation, and environmental and health impact, *Photochem. Photobiol. Sci.* 19 (2020) 66–70, <https://doi.org/10.1039/C9PP00366E>.
- [118] E. Magi, M.D. Carro, C. Scapolla, K.T.N. Nguyen, Stir bar sorptive extraction and LC-MS/MS for trace analysis of UV filters in different water matrices, *Chromatographia*. 17–18 (2012) 973–982, <https://doi.org/10.1007/s10337-012-2202-z>.
- [119] A. Goksoyr, K.E. Tollefsen, M. Grung, K. Loken, E. Lie, A. Zenker, K. Fent, M. Schlabach, S. Huber, Balsa raft crossing the Pacific finds low contaminant levels, *Environ. Sci. Technol.* 43 (2009) 4783–4790, <https://doi.org/10.1021/es900154h>.
- [120] L. Mandarić, E. Diamantini, E. Stella, K. Cano-Paoli, J. Valle-Sistac, D. Molins-Delgado, A. Bellin, G. Chiogna, B. Majone, M.S. Diaz-Cruz, S. Sabater, D. Barcelo, M. Petrovic, Contamination sources and distribution patterns of pharmaceuticals and personal care products in Alpine rivers strongly affected by tourism, *Sci. Total Environ.* 590–591 (2017) 484–494, <https://doi.org/10.1016/j.scitotenv.2017.02.185>.
- [121] M. Ma, H. Wang, M. Zhang, Q. Zhen, X. Du, Facile fabrication of polyaniline coated titania nanotube arrays as fiber coatings for solid phase microextraction coupled to high performance liquid chromatography for sensitive determination of UV filters in environmental water samples, *Anal. Methods* 9 (2017) 211–221, <https://doi.org/10.1039/C6AY02632J>.
- [122] R. Rodil, M. Moeder, R. Altenburger, M. Schmitt-Jansen, Photostability and phytotoxicity of selected sunscreen agents and their degradation mixtures in water, *Anal. Bioanal. Chem.* 395 (2009) 1513, <https://doi.org/10.1007/s00216-009-3113-1>.
- [123] D. Vione, R. Caringella, E. De Laurentiis, M. Pazzi, C. Minero, Phototransformation of the sunlight filter benzophenone-3 (2-hydroxy-4-methoxybenzophenone) under conditions relevant to surface waters, *Sci. Total Environ.* 463–464 (2013) 243–251, <https://doi.org/10.1016/j.scitotenv.2013.05.090>.
- [124] E. Paredes, S. Perez, R. Rodil, J.B. Quintana, R. Beiras, Ecotoxicological evaluation of four UV filters using marine organisms from different trophic levels *Isochrysis galbana*, *Mytilus galloprovincialis*, *Paracentrotus lividus*, and *Siriella armata*, *Chemosphere*. 104 (2014) 44–50, <https://doi.org/10.1016/j.chemosphere.2013.10.053>.
- [125] D. Finverson, N. Sabzevari, S. Qiblawi, J. Blitz, B.B. Norton, S.A. Norton, Sunscreens: UV filters to protect us: part 2-increasing awareness of UV filters and their potential toxicities to us and our environment, *Int. J. Womens Dermatol.* 7 (2021) 45–69, <https://doi.org/10.1016/j.ijwd.2020.08.008>.
- [126] D. Molins-Delgado, P. Gago-Ferrero, M.S. Díaz-Cruz, D. Barceló, Single and joint ecotoxicity data estimation of organic UV filters and nanomaterials toward selected aquatic organisms. Urban groundwater risk assessment, *Environ. Res.* 145 (2016) 126–134, <https://doi.org/10.1016/j.envres.2015.11.026>.
- [127] Y. Du, W.-Q. Wang, Z.-T. Pei, F. Ahmad, R.-R. Xu, Y.-M. Zhang, L.-W. Sun, Acute toxicity and ecological risk assessment of benzophenone-3 (BP-3) and benzophenone-4 (BP-4) in ultraviolet (UV)-filters, *Int. J. Environ. Res. Public Health* 14 (2017) E1414, <https://doi.org/10.3390/ijerph14111414>.
- [128] K. Fent, P.Y. Kunz, A. Zenker, M. Rapp, A tentative environmental risk assessment of the UV-filters 3-(4-methylbenzylidene-camphor), 2-ethyl-hexyl-4-trimethoxycinnamate, benzophenone-3, benzophenone-4 and 3-benzylidene camphor, *Mar. Environ. Res.* 69 (Suppl) (2010) S4–S6, <https://doi.org/10.1016/j.marenvres.2009.10.010>.
- [129] A. Boyd, C.B. Stewart, D.A. Philibert, Z.T. How, M.G. El-Din, K.B. Tierney, T.A. Blewett, A burning issue: the effect of organic ultraviolet filter exposure on the behaviour and physiology of *Daphnia magna*, *Sci. Total Environ.* 750 (2021), 141707, <https://doi.org/10.1016/j.scitotenv.2020.141707>.
- [130] H. Wang, H. Xi, L. Xu, M. Jin, W. Zhao, H. Liu, Ecotoxicological effects, environmental fate and risks of pharmaceutical and personal care products in the water environment: a review, *Sci. Total Environ.* 788 (2021), 147819, <https://doi.org/10.1016/j.scitotenv.2021.147819>.
- [131] M. Esperanza, M. Seoane, C. Rioboo, C. Herrero, Á. Cid, Differential toxicity of the UV-filters BP-3 and BP-4 in *Chlamydomonas reinhardtii*: a flow cytometric approach, *Sci. Total Environ.* 669 (2019) 412–420, <https://doi.org/10.1016/j.scitotenv.2019.03.116>.
- [132] Y. Huang, L. Luo, X.Y. Ma, X.C. Wang, Effect of elevated benzophenone-4 (BP4) concentration on *Chlorella vulgaris* growth and cellular metabolites, *Environ. Sci. Pollut. Res. Int.* 25 (2018) 32549–32561, <https://doi.org/10.1007/s11356-018-3171-z>.
- [133] S.L. Schneider, H.W. Lim, Review of environmental effects of oxybenzone and other sunscreen active ingredients, *J. Am. Acad. Dermatol.* 80 (2019) 266–271, <https://doi.org/10.1016/j.jaad.2018.06.033>.
- [134] B. Nataraj, K. Maharajan, D. Hemalatha, B. Rangasamy, N. Arul, M. Ramesh, Comparative toxicity of UV-filter Octyl methoxycinnamate and its photoproducts on zebrafish development, *Sci. Total Environ.* 718 (2020), 134546, <https://doi.org/10.1016/j.scitotenv.2019.134546>.
- [135] S. Zucchi, N. Blüthgen, A. Ieronimo, K. Fent, The UV-absorber benzophenone-4 alters transcripts of genes involved in hormonal pathways in zebrafish (*Danio rerio*) eluthero-embryos and adult males, *Toxicol. Appl. Pharmacol.* 250 (2011) 137–146, <https://doi.org/10.1016/j.taap.2010.10.001>.
- [136] V.W.T. Li, M.P.M. Tsui, X. Chen, M.N.Y. Hui, L. Jin, R.H.W. Lam, R.M.K. Yu, M.B. Murphy, J. Cheng, P.K.S. Lam, S.H. Cheng, Effects of 4-methylbenzylidene camphor (4-MBC) on neuronal and muscular development in zebrafish (*Danio rerio*) embryos, *Environ. Sci. Pollut. Res. Int.* 23 (2016) 8275–8285, <https://doi.org/10.1007/s11356-016-6180-9>.
- [137] C. Quintaneiro, B. Teixeira, J.L. Benedé, A. Chisvert, A.M.V.M. Soares, M.S. Monteiro, Toxicity effects of the organic UV-filter 4-methylbenzylidene camphor in zebrafish embryos, *Chemosphere*. 218 (2019) 273–281, <https://doi.org/10.1016/j.chemosphere.2018.11.096>.
- [138] M. Inui, T. Adachi, S. Takenaka, H. Inui, M. Nakazawa, M. Ueda, H. Watanabe, C. Mori, T. Iguchi, K. Miyatake, Effect of UV screens and preservatives on vitellogenin and choriogenin production in male medaka (*Oryzias latipes*), *Toxicology*. 194 (2003) 43–50, [https://doi.org/10.1016/S0300-483X\(03\)00340-8](https://doi.org/10.1016/S0300-483X(03)00340-8).
- [139] M. Liang, S. Yan, R. Chen, X. Hong, J. Zha, 3-(4-Methylbenzylidene) camphor induced reproduction toxicity and antiandrogenicity in Japanese medaka (*Oryzias latipes*), *Chemosphere*. 249 (2020), 126224, <https://doi.org/10.1016/j.chemosphere.2020.126224>.
- [140] C. Lozano, S. Matallana-Surget, J. Givens, S. Nouet, L. Arbuckle, Z. Lambert, P. Lebaron, Toxicity of UV filters on marine bacteria: combined effects with damaging solar radiation, *Sci. Total Environ.* 722 (2020), 137803, <https://doi.org/10.1016/j.scitotenv.2020.137803>.
- [141] Q. Zhang, X. Ma, M. Dzakpasu, X.C. Wang, Evaluation of ecotoxicological effects of benzophenone UV filters: luminescent bacteria toxicity, genotoxicity and hormonal activity, *Ecotoxicol. Environ. Saf.* 142 (2017) 338–347, <https://doi.org/10.1016/j.ecoenv.2017.04.027>.
- [142] G. Abedi, Z. Talebpour, F. Jamechenarboo, Survey of analytical methods for sample preparation and analysis of fragrances in cosmetics and personal care products, *Trends Anal. Chem.* (2018), <https://doi.org/10.1016/j.trac.2018.01.006> (accessed October 26, 2022).
- [143] M.B. Tahir, A. Ahmad, T. Iqbal, M. Ijaz, S. Muhammad, S.M. Siddeeq, Advances in photo-catalysis approach for the removal of toxic personal care product in aqueous environment, *Environ. Dev. Sustain.* 22 (2020) 6029–6052, <https://doi.org/10.1007/s10668-019-00495-1>.
- [144] C. Lange, B. Kuch, J.W. Metzger, Occurrence and fate of synthetic musk fragrances in a small German river, *J. Hazard. Mater.* 282 (2015) 34–40, <https://doi.org/10.1016/j.jhazmat.2014.06.027>.
- [145] Y. Luo, W. Guo, H.H. Ngo, L.D. Nghiem, F.I. Hai, J. Zhang, S. Liang, X.C. Wang, A review on the occurrence of micropollutants in the aquatic environment and their fate and removal during wastewater treatment, *Sci. Total Environ.* 473–474 (2014) 619–641, <https://doi.org/10.1016/j.scitotenv.2013.12.065>.
- [146] A. Salvador, A. Chisvert, Analysis of Cosmetic Products, 2nd edition, 2011 <https://www.elsevier.com/books/analysis-of-cosmetic-products/salvador/978-0-444-63508-2> (accessed October 28, 2022).
- [147] U. Klaschka, P.C. von der Ohe, A. Bschorer, S. Krezmer, M. Sengl, M. Letzel, Occurrences and potential risks of 16 fragrances in five German sewage treatment plants and their receiving waters, *Environ. Sci. Pollut. Res. Int.* 20 (2013) 2456–2471, <https://doi.org/10.1007/s11356-012-1120-9>.
- [148] J. Margot, L. Rossi, D.A. Barry, C. Holliger, A review of the fate of micropollutants in wastewater treatment plants, *WIREs, Water*. 2 (2015) 457–487, <https://doi.org/10.1002/wat2.1090>.
- [149] S.L. Simonich, W.M. Begley, G. Debaere, W.S. Eckhoff, Trace analysis of fragrance materials in wastewater and treated wastewater, *Environ. Sci. Technol.* 34 (2000) 959–965, <https://doi.org/10.1021/es991018g>.



- [150] M. Vecchiato, S. Cremonese, E. Gregoris, E. Barbaro, A. Gambaro, C. Barbante, Fragrances as new contaminants in the Venice lagoon, *Sci. Total Environ.* 566–567 (2016) 1362–1367, <https://doi.org/10.1016/j.scitotenv.2016.05.198>.
- [151] S. Gutiérrez, C. Fernandez, C. Barata, J.V. Tarazona, Forecasting risk along a river basin using a probabilistic and deterministic model for environmental risk assessment of effluents through ecotoxicological evaluation and GIS, *Sci. Total Environ.* 408 (2009) 294–303, <https://doi.org/10.1016/j.scitotenv.2009.09.053>.
- [152] D.A. Alvarez, K.A. Maruya, N.G. Dodder, W. Lao, E.T. Furlong, K.L. Smalling, Occurrence of contaminants of emerging concern along the California coast (2009–10) using passive sampling devices, *Mar. Pollut. Bull.* 81 (2014) 347–354, <https://doi.org/10.1016/j.marpolbul.2013.04.022>.
- [153] A.K. Baldwin, S.R. Corsi, L.A. De Cicco, P.L. Lenaker, M.A. Lutz, D.J. Sullivan, K.D. Richards, Organic contaminants in Great Lakes tributaries: prevalence and potential aquatic toxicity, *Sci. Total Environ.* 554–555 (2016) 42–52, <https://doi.org/10.1016/j.scitotenv.2016.02.137>.
- [154] D. Relić, A. Popović, D. Đorđević, J. Časlavský, Occurrence of synthetic musk compounds in surface, underground, waste and processed water samples in Belgrade, Serbia, *Environ. Earth Sci.* 76 (2017) 122, <https://doi.org/10.1007/s12665-017-6441-z>.
- [155] V.T. Politano, A.A. Lapczynski, G. Ritacco, A.M. Api, Ninety-day toxicity study of alpha-iso-methylionone in rats, *Int. J. Toxicol.* 31 (2012) 595–601, <https://doi.org/10.1177/1091581812466116>.
- [156] X. Bai, T. Zhang, C. Wang, D. Zong, H. Li, Z. Yang, Occurrence and distribution of taste and odor compounds in subtropical water supply reservoirs and their fates in water treatment plants, *Environ. Sci. Pollut. Res. Int.* 24 (2017), <https://doi.org/10.1007/s11356-016-7966-5>.
- [157] M. Hattori, A. Watabe, K. Takahashi,  $\beta$ -Lactoglobulin protects  $\beta$ -ionone related compounds from degradation by heating, oxidation, and irradiation, *Biosci. Biotechnol. Biochem.* 59 (1995) 2295–2297, <https://doi.org/10.1271/bbb.59.2295>.
- [158] M. Picone, G.G. Distefano, D. Marchetto, M. Russo, M. Vecchiato, A. Gambaro, C. Barbante, A.V. Ghirardini, Fragrance materials (FMs) affect the larval development of the copepod *Acartia tonsa*: an emerging issue for marine ecosystems, *Ecotoxicol. Environ. Saf.* 215 (2021), 112146, <https://doi.org/10.1016/j.ecoenv.2021.112146>.
- [159] A.M. Api, D. Belsito, S. Biserta, D. Botelho, M. Bruze, G.A. Burton, J. Buschmann, M.A. Cancellieri, M.L. Dagli, M. Date, W. Dekant, C. Deodhar, A.D. Fryer, S. Gadhia, L. Jones, K. Joshi, A. Lapczynski, M. Lavelle, D.C. Liebler, M. Na, D. O'Brien, A. Patel, T.M. Penning, G. Ritacco, F. Rodriguez-Roperio, J. Romine, N. Sadekar, D. Salvito, T.W. Schultz, F. Siddiqi, I.G. Sipes, G. Sullivan, Y. Thakkar, Y. Tokura, S. Tsang, RIFM fragrance ingredient safety assessment, methyl ionone (mixture of isomers), CAS registry number 1335-46-2, *Food Chem. Toxicol.* 134 (2019), 110716, <https://doi.org/10.1016/j.fct.2019.110716>.
- [160] J. Zhao, L. Yang, L. Zhou, Y. Bai, B. Wang, P. Hou, Q. Xu, W. Yang, Z. Zuo, Inhibitory effects of eucalyptol and limonene on the photosynthetic abilities in *Chlorella vulgaris* (Chlorophyceae), *Phycologia* 55 (2016) 696–702, <https://doi.org/10.2216/16-38.1>.
- [161] M.R.S. Jain, T.G. Jain, S.S. Gavitt, M.N.S. Patil, Natural Hand Sanitizer: Effective Product, 2020 1.
- [162] D.J. McClements, S.M. Jafari, Improving emulsion formation, stability and performance using mixed emulsifiers: a review, *Adv. Colloid Interf. Sci.* 251 (2018) 55–79, <https://doi.org/10.1016/j.cis.2017.12.001>.
- [163] S. Tcholakova, N.D. Denkov, A. Lips, Comparison of solid particles, globular proteins and surfactants as emulsifiers, *Phys. Chem. Chem. Phys.* 10 (2008) 1608–1627, <https://doi.org/10.1039/B715933C>.
- [164] A. Forgiairini, J. Esquena, C. González, C. Solans, Formation and stability of nano-emulsions in mixed nonionic surfactant systems, in: P.G. Koutsoukos (Ed.), *Trends Colloid Interface Sci.* XV, Springer, Berlin, Heidelberg 2001, pp. 184–189, [https://doi.org/10.1007/3-540-45725-9\\_42](https://doi.org/10.1007/3-540-45725-9_42).
- [165] J.S. Franklyne, A. Mukherjee, N. Chandrasekaran, Essential oil micro- and nanoemulsions: promising roles in antimicrobial therapy targeting human pathogens, *Lett. Appl. Microbiol.* 63 (2016) 322–334, <https://doi.org/10.1111/lam.12631>.
- [166] L. Salvia-Trujillo, R. Soliva-Fortuny, M.A. Rojas-Graü, D.J. McClements, O. Martín-Belloso, Edible nanoemulsions as carriers of active ingredients: a review, *Annu. Rev. Food Sci. Technol.* 8 (2017) 439–466, <https://doi.org/10.1146/annurev-food-030216-025908>.
- [167] G. Libralato, A. Volpi Ghirardini, F. Avezzù, Seawater ecotoxicity of monoethanolamine, diethanolamine and triethanolamine, *J. Hazard. Mater.* 176 (2010) 535–539, <https://doi.org/10.1016/j.jhazmat.2009.11.062>.
- [168] OECD, OECD's Work on Co-operating in the Investigation of High Production Volume Chemicals - Home, <https://hpvchemicals.oecd.org/ui/Default.aspx> 2001 (accessed January 18, 2023).
- [169] M.M. Fiume, B. Heldreth, W.F. Bergfeld, D.V. Belsito, R.A. Hill, C.D. Klaassen, D. Liebler, J.G. Marks, R.C. Shank, T.J. Slaga, P.W. Snyder, F.A. Andersen, Safety assessment of triethanolamine and triethanolamine-containing ingredients as used in cosmetics, *Int. J. Toxicol.* 32 (2013) 59S–83S, <https://doi.org/10.1177/1091581813488804>.
- [170] M.A. Liebert, 8 final report on the safety assessment of triethanolamine, diethanolamine, and monoethanolamine, *J. Am. Coll. Toxicol.* 2 (1983) 183–235, <https://doi.org/10.3109/10915818309142006>.
- [171] ECHA, Tocopherols, Endpoint Summary, <https://echa.europa.eu/da/registration-dossier/-/registered-dossier/12703/6/2/1> (accessed October 31, 2022).
- [172] T. Tišler, J. Zagorc-Končan, M. Cotman, A. Drolc, Toxicity potential of disinfection agent in tannery wastewater, *Water Res.* 38 (2004) 3503–3510, <https://doi.org/10.1016/j.watres.2004.05.011>.
- [173] USEPA, United States Environmental Protection Agency Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms (2002), 2002 232.
- [174] A. de Oliveira Melo, D.B.D. Santos, L.D. Silva, T.L. Rocha, J.C.B. Bezerra, Molluscicidal activity of polyhexamethylene biguanide hydrochloride on the early-life stages and adults of the *Biomphalaria glabrata* (Say, 1818), *Chemosphere* 216 (2019) 365–371, <https://doi.org/10.1016/j.chemosphere.2018.10.035>.
- [175] ECHA, Guidance on Information Requirements and Chemical Safety Assessment Chapter R.7b: Endpoint specific guidance Draft Version 4.0, [https://echa.europa.eu/documents/10162/23047722/ir\\_csa\\_r7b\\_pbt\\_caracal\\_draft\\_en.pdf/1526c738-afa2-8bce-f233-4379f0b697a4](https://echa.europa.eu/documents/10162/23047722/ir_csa_r7b_pbt_caracal_draft_en.pdf/1526c738-afa2-8bce-f233-4379f0b697a4) 2017 (accessed November 18, 2022).
- [176] European Chemical Bureau (ECB), Institute for Health and Consumer Protection, European Commission: Technical guidance document on risk assessment in support of commission directive 93/67/EEC on risk assessment for new notified substances, commission regulation (EC) No. 1488/94 on risk assessment for existing substances, and directive 98/8/EC of the European Parliament and of the Council Concerning the Placing of Biocidal Products on the Market. Part II. EUR 20418 EN/2, Joint Research Centre, Ispra, Italy, 2003 [https://echa.europa.eu/documents/10162/987906/tgdpart2\\_2ed\\_en.pdf/138b7b71-a069-428e-9036-62f4300b752f](https://echa.europa.eu/documents/10162/987906/tgdpart2_2ed_en.pdf/138b7b71-a069-428e-9036-62f4300b752f) (accessed October 31, 2022).
- [177] A.D. Lemly, Evaluation of the hazard quotient method for risk assessment of selenium, *Ecotoxicol. Environ. Saf.* 35 (1996) 156–162, <https://doi.org/10.1006/eesa.1996.0095>.
- [178] E. Archer, B. Petrie, B. Kasprzyk-Hordern, G.M. Wolfaardt, The fate of pharmaceuticals and personal care products (PPCPs), endocrine disrupting contaminants (EDCs), metabolites and illicit drugs in a WWTW and environmental waters, *Chemosphere* 174 (2017) 437–446, <https://doi.org/10.1016/j.chemosphere.2017.01.101>.