

Decoding the synergistic potential of MAZ-51 and zingerone as therapy for melanoma treatment in alignment with sustainable development goals

Kganya Letsoalo¹  | Evangeline Nortje¹  | Sean Patrick²  | Trevor Nyakudya¹  | Yvette Hlophe¹ 

¹Department of Physiology, University of Pretoria, Pretoria, South Africa

²Environmental Chemical Pollution and Health Research Unit, University of Pretoria, Pretoria, South Africa

Correspondence

Yvette Hlophe, Department of Physiology, University of Pretoria, Private Bag X323, Arcadia, Pretoria 0007, South Africa.
Email: yvette.hlophe@up.ac.za

Funding information

National Research Foundation, Grant/Award Number: N1F580A1F685; University of Pretoria Postgraduate Funding Scheme

Abstract

Melanoma, an invasive class of skin cancer, originates from mutations in melanocytes, the pigment-producing cells. Globally, approximately 132,000 new cases are reported each year, and in South Africa, the incidence stands at 2.7 per 100,000 people, signifying a worrisome surge in melanoma rates. Therefore, there is a need to explore treatment modalities that will target melanoma's signalling pathways. Melanoma metastasis is aided by ligand activity of transforming growth factor-beta 1 (TGF- β 1), vascular endothelial growth factor-C (VEGF-C) and C-X-C chemokine ligand 12 (CXCL12) which bind to their receptors and promote tumour cell survival, lymphangiogenesis and chemotaxis. (3-(4-dimethylaminonaphthelen-1-ylmethylene)-1,3-dihydroindol-2-one) MAZ-51 is an indolinone-based molecule that inhibits VEGF-C induced phosphorylation of vascular endothelial growth factor receptor 3 (VEGFR-3). Despite the successful use of conventional cancer therapies, patients endure adverse side effects and cancer drug resistance. Moreover, conventional therapies are toxic to the environment and caregivers. The use of medicinal plants and their phytochemical constituents in cancer treatment strategies has become more widespread because of the rise in drug resistance and the development of unfavourable side effects. Zingerone, a phytochemical derived from ginger exhibits various pharmacological properties positioning it as a promising candidate for cancer treatment. This review provides an overview of melanoma biology and the intracellular signalling pathways promoting cell survival, proliferation and adhesion. There is a need to align health and environmental objectives within sustainable development goals 3 (good health and well-being), 13 (climate action) and 15 (life on land) to promote early detection of skin cancer, enhance sun-safe practices, mitigation of environmental factors and advancing the preservation of biodiversity, including medicinal plants. Thus, this review discusses the impact of cytostatic cancer drugs on patients and the environment and examines the potential use of phytochemicals as adjuvant therapy.

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial-NoDerivs](https://creativecommons.org/licenses/by-nc-nd/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2024 The Authors. *Cell Biochemistry and Function* published by John Wiley & Sons Ltd.

KEYWORDS

chemotherapeutics, MAZ-51, melanoma, phytochemicals, sustainable development goals, zingerone

1 | INTRODUCTION

Cancer is a heterogeneous disorder characterised by unchecked cell growth and proliferation, with transformed cells invading surrounding tissue and mobilising to distant sites.¹ The World Health Organization (WHO) 2020 statistics recorded cancer as the second leading cause of death globally, with approximately 10 million deaths reported.² South Africa reported a total of 107,467 new cases and 57,373 cancer-related deaths in 2018.² Breast, lung, colon and rectum, prostate, and nonmelanoma skin and stomach cancers are the most widespread types of cancers.²

Skin cancer manifests as the uncontrolled growth of mutated skin cells,³ with over 1.5 million cases estimated worldwide in 2020.⁴ South Africa ranks second in skin cancer incidence, with 20,000 new cases and 700 fatalities recorded annually.⁵ The condition is categorised into two types: melanoma and nonmelanoma skin cancer, with melanoma constituting merely 1% of cases but contributing to as much as 90% of skin cancer-related deaths.⁶ The next section of the review delves into a detailed discussion of metastatic melanoma providing its biology.

2 | MELANOMA

2.1 | Epidemiology

Malignant melanoma is a neoplasm characterised by the malignant transformation of melanocytes.⁷ Albeit contributing to 1% of skin cancer cases, melanoma contributes to 90% of skin cancer-related deaths.⁶ Global estimates recorded an average of 132,000 new cases annually, while South Africa recorded an incidence of 2.7 per 100,000 individuals.⁵ Melanoma incidence is increasing globally with an alarming element being the discrepancy in age and gender. While males exhibit a 1.5-fold greater incidence, younger females aged 20–45 report higher incidence rates. However, a sharp incline in incidence rate is recorded in males greater than 50 years.^{8,9} Younger females possess a greater predisposition to melanoma as they are more likely to be exposed to ultraviolet (UV) radiation, directly from the sun or from artificial sources.¹⁰ Additionally, the difference in metabolic activity of males and females in response to androgens and oestrogens influences melanoma development.¹⁰

The occurrence of malignant melanoma is dependent on the synergistic interplay between risk factors such as common or atypical nevi, the use of cosmetics, chronic sun exposure (particularly during childhood) and geographical zone.^{7,11} Incidence rates are particularly elevated in Australia, New Zealand, Europe and Northern America while lower rates are observed in Africa.⁷ This disparity is concordant

Significance statement

- Cytostatic drugs, despite being effective treatment agents, induce nontarget effects harming noncancerous cells.
- This review highlights the use of phytochemicals as adjuvant therapy in cancer treatment to combat the nontarget effects of cytostatic drugs.
- Additionally, phytochemicals as adjuvants decrease the environmental burden incurred by cytostatic drugs, promoting environmental sustainability.

with ethnicity, lifestyle as well as genetic composition. Populations with fair skin are frequently diagnosed with cutaneous melanoma whereas, individuals from Asia and Africa develop acral and mucosal melanomas at lower incidence rates.⁷

2.2 | Melanoma etiology

2.2.1 | The role of UV radiation exposure

UV radiation is the most prominent carcinogenic factor inducing melanoma and nonmelanoma skin cancer. The UV spectrum consists of UV-A (315–400 nm), UV-B (280–315 nm) and UV-C (100–280 nm).¹² UV-C despite having the shortest wavelength, does not penetrate the ozone layer and therefore, has no observed effect on melanoma genesis.¹² Approximately 5% of UV-B and 95% of UV-A rays reach the earth's surface however, UV-B is the most potent form as it directly induces DNA mutations whereas, UV-A penetrates deeper layers of the skin (dermal stratum papillare) and results in indirect damage resulting in oxidative stress.^{3,12,13} In instances where UV-induced mutations affect genes dictating signalling pathways such as cell cycle, proliferation, apoptosis or DNA repair then malignancy may be induced.¹³

Consequent to UV-radiation-induced DNA damage, skin keratinocytes produce melanocyte stimulating hormone (MSH) which in turn binds to melanocortin 1 receptor (MC1R) inducing the production of melanin and thus preventing UV-induced DNA damage.¹⁴

Melanin contains pro-oxidant and antioxidant properties; its transformation from an antioxidant to a pro-oxidant due to UV-radiation induces carcinogenesis.¹⁵ Its pro-oxidant properties increase intracellular oxygen radical concentrations promoting DNA mutations of melanocytes. These mutations prompt the overactivation of various signalling pathways resulting in uncontrolled cell proliferation.¹⁵

2.2.2 | The role of cosmetics

Cosmetics such as facial makeup are frequently applied to the skin and thus their constituents are exposed to and absorbed by the skin for extended periods of time.¹¹ Facial products are expected to consist of ingredients that are safe for consumer use and comply with regulatory standards. However, potential carcinogens such as parabens, ethoxylated compounds, formaldehyde donors, urea and silica have been detected in facial products.¹¹ The carcinogens are categorised into two groups mainly genotoxic and nongenotoxic compounds. Genotoxic compounds induce direct DNA damage following application or proceeding xenobiotic metabolism.¹¹ Conversely, nongenotoxic compounds result in indirect DNA damage by inducing the formation of reactive oxygen species (ROS), immunosuppression and inducing inflammation.¹¹

2.2.3 | The role of hereditary factors

Melanoma is typically a consequence of somatic mutations however, a shared family history increases disease risk because of inherited mutations and common sun exposure habits.¹⁶

Germline mutations in cyclin dependent kinase inhibitor 2 A (CDKN2A) and to a lesser extent cyclin dependent kinase 4 (CDK4), are genetic impairments affiliated with familial melanoma. Inheritance of mutated CDKN2A allele increases an individual's risk of developing melanoma. Seventy percent of cutaneous melanoma cases reported a mutation in the CDKN2A gene.¹⁶ This gene is situated on chromosome 9p21 and encodes p16^{INK4A} and p14^{ARF}, tumour suppressor proteins. Under normoxic conditions, these proteins promote cell-cycle arrest however, mutations inhibit their cancer-suppression activity promoting uncontrolled cell proliferation.^{16,17} In addition, p16^{INK4A} negatively regulates cell cycle progression by suppressing the activity of CDK4 inhibiting Gap 1-synthesis checkpoint progression whereas, p14^{ARF} positively regulates p53, a tumour suppressor protein, by inhibiting the phosphorylation of murine double minute 2-a p53 regulator- thus preventing cell cycle progression.¹⁷ Therefore, impairment of CDKN2A by deletions, promoter silencing or mutations results in uncontrolled cell growth and proliferation.¹⁷

Cdk 4, located on chromosome 12q3, binds to, and is regulated by p16. Mutations in its binding domain inhibit it from associating with p16 therefore, promoting cell cycle progression.¹⁷

3 | MELANOMA DIAGNOSIS AND STAGING

Melanoma classification is outlined by the American Joint Committee on Cancer (AJCC) based on the tumour thickness, nodal involvement and metastasis (TNM) system.^{18,19} Tumour thickness, within the TNM framework, is characterised by the primary tumour's thickness and ulceration. Conversely, the Breslow measurement

considers the depth of invasion of the neoplasm.^{6,19} Nodal involvement signifies whether the tumour has progressed to proximal lymph nodes. In the TNM system, the 'M' assesses dissemination to distal lymph nodes and organs, with the brain, lungs and liver being the preferred metastatic sites.^{6,19}

Melanoma staging may further be classified into stages 0–IV.⁵ Stage 0, also known as in situ, denotes cancer cells that are confined to the epidermis lacking infiltration into the dermis, lymph nodes or distant organs. Stage I is characterised by a tumour measuring approximately 1 mm in thickness with or without ulceration.⁵ Stage II is characterised by a tumour of 4 mm thickness without metastatic evidence.^{5,20} Stage III is characterised by a tumour size ≥ 4 mm, coupled with dissemination to proximal lymph nodes.^{5,20} Stage IV arises when melanoma cells metastasize to distant organs such as the brain, lungs and liver, in addition to distal lymph nodes.^{5,20}

4 | MELANOMA SUBTYPES

Melanoma is subcategorised into four subtypes: acral lentiginous, superficial spreading, lentigo malignant and nodular melanoma.^{5,21} Understanding the characteristics of each subtype is crucial for early detection, accurate diagnosis and tailored treatment strategies.

4.1 | Superficial spreading melanoma

Superficial spreading melanoma is the most prevalent form²² accounting for half to three-quarters of diagnosed malignancies.²¹ Superficial spreading melanoma stems from pre-existing moles or nevi and is restricted to the epidermis, for a period before the vertical growth phase takes place. Nevi are ubiquitously positioned around the body with increased density on the back and legs in females, and the back and trunk in men.^{5,21}

4.2 | Nodular melanoma

Nodular melanoma is the most invasive class of melanoma, contributing to around 15%–20% of all South African cases, and is renowned for its ability to rapidly penetrate the skin.⁵ It is characterised by the emergence of moles that exhibit dark blue–black, blue–red and occasionally colourless hues, typically found on the neck, head and trunk.^{19,21}

4.3 | Lentigo melanoma

Lentigo melanoma is a less prevalent subtype of melanoma responsible for 5%–15% of cases, with lesions located on areas of the body susceptible to photodamage including the face, ears, arms or upper torso.^{5,19} This form of melanoma is less invasive with an extended radial growth phase. It is further characterised by blue,

black, tan or brown lesions that appear to be flat with irregular borders.²¹

4.4 | Acral lentiginous melanoma

Acral lentiginous melanoma is an infrequent form of melanoma, but frequently diagnosed in individuals with African ancestry.²¹ It presents as tan to brown–black patches with uneven borders and is customarily located on the underside of nail plates, soles of the feet and the palms.^{5,19,21}

Through a comprehensive understanding of these melanoma subtypes, healthcare professionals can enhance their diagnostic accuracy and treatment strategies, ultimately leading to improved outcomes for patients. In the subsequent sections of this review, we will assess the emerging therapeutic approaches, shedding light on the intricacies of melanoma management.

5 | CURRENT MELANOMA TREATMENT OPTIONS

Cancer is characterised as heterogenous, tissue-specific and evolutionary. Consequently, several treatment modalities have been developed and are currently devised to eradicate tumour cells and mitigate the risk of recurrence.¹

5.1 | Surgical excision

The primary treatment approach for early-stage melanoma involves surgical excision of both the neoplastic cells and the neighbouring noncancerous tissues. This approach is supported by a 92% overall survival rate. Furthermore, subsequent steps often involve sentinel lymph node (SLN) biopsy and the excision of surrounding lymph nodes.^{7,14,23}

Surgical excision during the initial stages of pathogenesis results in improved prognosis. However, with a 10% chance of a 5-year survival rate, metastatic cells demonstrate resistance to surgical intervention. Consequently, advanced stages of melanoma necessitate additional treatment plans, such as chemotherapy.^{7,23}

5.2 | Chemotherapy

Over the recent decades, chemotherapy has been the conventional therapeutic approach. The first drug approved by the Food and Drug Administration (FDA) for metastatic melanoma was dacarbazine (DITC), receiving approval in 1975.²³ DITC is an alkylating agent that induces DNA damage by inserting alkyl groups in guanine bases resulting in cell death.²⁴ Despite being regarded as one of the most potent chemotherapeutic drugs, DITC is minimally effective, yielding a median survival ranging from 5 to 11 months, coupled with a 1-year

survival rate of 27%.²³ The side effects associated with DITC include nausea, vomiting, leukopenia and anaemia.²⁵ Temozolomide (TMZ), an analogue of DITC, presents similar pharmacological activities to DITC however, unlike DITC, TMZ can cross the blood-brain barrier targeting central nervous system (CNS) metastasis.²⁶ Retrospective studies by Agarwala and colleagues and Paul and colleagues consisting of 122 patients presenting with intracranial disease demonstrated an overall response rate of 7% with a 77% decrease in CNS metastasis.^{27,28}

Combination therapy targeting multiple cell cycle components is a common therapeutic strategy to combat tumour resistance and to reduce adverse side effects associated with monotherapy.²⁶ Polytherapy of DITC with other cytostatic drugs such as vinblastine, vindesine, cisplatin, carboplatin and taxane has been investigated however, no significant overall survival rates have been recorded when comparing DITC monotherapy to DITC polytherapy.⁷

With the advancement of research, the underlying molecular mechanisms dictating melanoma initiation, growth and progression have been explored and detailed derailing treatment modalities from cytotoxic drugs to more specific treatment strategies such as targeted therapy.²⁴ A phase III clinical trial comparing vemurafenib, a BRAF inhibitor, to DITC in patients with V600E- mutant metastatic melanoma observed a response rate of 48% in vemurafenib and 5% for DITC.²⁹ Additionally, vemurafenib-treated patients exhibited a progression-free survival (PFS) of 5.3 months compared to a PFS of 1.6 months in DITC.²⁹ A median PFS of 4.8 months was observed in trametinib-treated patients compared with 1.5 months for the chemotherapy group. An overall survival rate of 81% for trametinib and 67% for chemotherapy was recorded at 6 months.²⁹

5.3 | Targeted therapy

The mitogen-activated protein kinase (MAPK) pathway is a signalling cascade that modulates cell survival, differentiation and proliferation. The pathway constitutes of RAS, RAF, MEK and ERK intermediary kinase proteins transducing outside-in signalling. Approximately 90% of melanomas display irregularities within the MAPK pathway. The most prevalent genetic anomalies linked to disrupted MAPK signalling in melanoma involve mutations in the BRAF and NRAS gene.¹⁴ Currently, no specific targeted therapy for NRAS mutations have been identified however, the impact of BRAF inhibitors, specifically vemurafenib and dabrafenib, on disease outcomes has been investigated. The findings demonstrate improved survival rates and increased tolerance however, patients eventually develop resistance to the treatments, resulting in the resurgence of the MAPK pathway.^{7,23} To counteract treatment resistance and relapse, combination therapy with MEK inhibitors, cobimetinib and trametinib, were explored. Preclinical studies documented increased apoptosis and a decrease in treatment resistance. Nevertheless, significant side effects have been recorded with the use of MAPK pathway inhibitors.⁷

5.4 | Challenges associated with conventional cancer therapies

Conventional cytotoxic therapies have shown favourable patient outcomes. However, it is important to note that cytostatic drugs do not distinguish between cancerous and noncancerous cells thus, resulting in undesired side effects associated with their use.¹ Table 1 shows drugs used for cancer treatment and their side effects.

Chemotherapy has been proven to eliminate cancer cells however, its cytotoxic activity is observed in noncancerous cells resulting in adverse side effects.¹ The most prevalent side effects recorded are vomiting, nausea, fatigue, malaise, diarrhoea, headaches, rashes, pain, infections, mucositis, alopecia and loss of appetite.³⁹

Chemotherapeutic drug utilisation results in immunosuppression, as cytotoxic agents target dividing hematopoietic cells, resulting in neutropenia and cytopenia. This increases the susceptibility to infections, including those caused by oncogenic viruses.³⁹ Additionally, cytotoxic drugs induce both epigenetic and genetic damage. Consequently, their use not only adversely affects healthy rapidly dividing cells, but also increases an individual's vulnerability to secondary malignancies and other diseases.³⁹ Cytotoxic drugs can lead to neurological side effects, including memory loss, cognitive dysfunction, vision impairment, seizures, dementia and cerebral infarctions.³⁹ These effects impact a significant proportion of patients, ranging from 4% to 75% following the completion of treatment.³⁹ Moreover, treatment protocols include the use of combination and adjuvant chemotherapy; long-term use of the latter culminates in chronic fatigue, sexual dysfunction, musculoskeletal abnormalities and skin changes.³⁹ Although combination chemotherapy targets multiple pathways, it is associated with the development of multidrug resistance. Therefore, it is imperative to explore alternative forms of treatment.^{39,40}

The development of melanoma is an intricate process involving various intracellular signalling pathways. Knowledge in the weak

points of these signalling pathways allows researchers to enhance treatment strategies moving from current conventional therapeutics to more effective and efficient treatment strategies that inhibit cancer cell growth without inducing nontarget effects. The following section reviews signalling pathways involved in melanoma, describing the aberrations resulting in malignancy.

6 | ROLE OF THE TUMOUR MICROENVIRONMENT IN CANCER PROGRESSION

The tumour microenvironment consists of tumour cells and stroma namely cancer-associated fibroblasts, tumour-associated macrophages, tumour endothelial cells, leukocytes and pericytes that mould an environment that supports tumour growth and progression.^{41,42}

Tumour cells secrete regulatory factors such as cytokines, growth factors and chemokines that recruit stromal, immune cells and enzymes that remodel the extracellular matrix (ECM) to construct a favourable environment that promotes tumour growth, proliferation and metastasis.⁴²

Cytokines have been observed to promote melanoma cell growth, proliferation and survival.⁴³ In vitro studies have recorded that several melanoma cell lines secrete cytokines and growth factors that function in an autocrine and/or paracrine manner to mediate growth, invasion and angiogenesis.⁴³ Additionally, cytokines operate as adhesion molecules and have been shown to possess antiapoptotic properties.⁴³ Dysregulation of melanoma cytokine levels affect malignancy by altering the sensitivity to therapeutics and mitigating disease progression.⁴⁴ Interleukin (IL)-1, fibroblast growth factor (bFGF), transforming growth factor-beta (TGF- β) and platelet-derived growth factor (PDGF) stimulate neovascularisation required for melanoma cell growth; these factors are elevated in melanoma

TABLE 1 List of food and drug administration approved synthetic cancer drugs and their associated side effects.

Drug name	Mode of action	Side effect	References
Dacarbazine	Inhibits DNA and RNA synthesis. BRAF inhibitor.	Fatigue, loss of appetite, nausea, vomiting, kidney and liver malfunction	Milijašević et al. ³⁰
Paclitaxel	Induces a metaphase/anaphase halt and cell death	Alopecia, nausea and vomiting, mucositis, neutropenia, leukopenia, anaemia	Kampan et al. ³¹
Cisplatin	Promotes DNA damage and induces apoptosis	Nephrotoxic, cardiotoxic, and hepatotoxic	Dasari et al. ³²
Vemurafenib	Inhibits activation of the MAPK, inhibiting proliferation	Photosensitivity, maculopapular eruptions and hyperkeratosis	Trunzer et al. ³³
Temozolomide	Deposits methyl group on DNA guanine bases	Nausea and vomiting	Wesolowski et al. ³⁴
Ipilimumab	Inhibits cytotoxic T-lymphocyte antigen-4	Dermatological, gastrointestinal/hepatic, endocrine and pulmonary system abnormalities	Darnell et al. ³⁵ and Youssef et al. ³⁶
Nivolumab	Inhibits programmed cell death-1 receptor and its ligand	Colitis, hepatitis, skin toxicities, hypophysis and thyroid dysfunction	Koppolu et al. ³⁷ and Spain et al. ³⁸

patients.⁴⁴ Elevated concentrations of IL-10 have been detected in stage II and IV melanoma patients. IL-10 downregulates antitumour responses and functions as a tumour growth factor.⁴⁴ The cytokine, IL-6, is a pleiotropic cytokine secreted by multiple cells such as lymphocytes, monocytes, macrophages, keratinocytes, tumour and endothelial cells.⁴⁵ In vitro studies, elevated IL-6 concentrations inhibit melanocyte growth however, IL-6 promotes the growth and proliferation of cells isolated from metastatic melanoma.⁴⁵ Melanoma patients with elevated IL-6 concentrations present with a poor response to treatment, have a lower survival rate and are resistant to IL-2 therapy.⁴⁴ Therefore, it is evident that cytokines play a significant role in promoting melanoma cell growth, proliferation and survival.

7 | GROWTH FACTORS THAT ACTIVATE INTRACELLULAR SIGNALLING PATHWAYS

7.1 | Transforming growth factor-beta signalling in melanoma

TGF- β is a pleiotropic cytokine forming part of a family that consists of more than 30 members in mammals and manages several cellular processes including apoptosis, angiogenesis, wound healing, embryonic development, immune surveillance and tumour biology.^{46,47}

TGF- β 's three isoforms (TGF- β 1, 2 and 3), are synthesized as inactive complexes and transported to the extracellular matrix (ECM) where activation takes place through proteolysis.^{47,48} TGF- β mediates signal transduction through nonsmad (noncanonical) and smad

(canonical) pathways. Active TGF- β 1 binds to its receptors (T β R II and T β R I) with serine/threonine kinases. T β R II binds to and phosphorylates T β R I transducing intracellular signalling through the activity of smad proteins, Figure 1.^{41,47}

Signalling may conversely be conveyed through nonsmad pathways such as protein extracellular signal-regulated kinase (Erk) 1/2, kinase B (Akt), p38 MAPK and phosphoinositide 3-kinase (PI3K) prompting cancer cell progression, invasion and metastasis, Figure 1.^{48,49}

TGF- β 1 functions dichotomously; it acts as a tumour suppressor in melanocytes and premalignant cells by modulating cyclin-dependent kinase inhibitors, p21 and p15, and c-myc, an oncogene that endorses cell proliferation.⁴⁷ However, the accrual of genetic and epigenetic modifications procures TGF- β 1 insensitive to its inhibitory effects.⁵⁰

Paracrine secretion induces cancer cell growth and invasion through the modification of the tumour microenvironment (TME) resulting in the activation of stromal fibroblasts and their conversion to myofibroblasts.⁴⁹ Overexpressed TGF- β 1 prompts ECM stiffening through increased production of collagen I and III and fibronectin, proteins involved in ECM adhesion, augmenting communication between the ECM and fibroblast.⁴⁹ Additionally, fibroblast-induced signalling is conveyed through integrins initiating downstream signalling cascades, prompting malignancy.⁵¹

TGF- β 1 advances tumour angiogenesis by upregulating the secretion of vascular endothelial growth factors (VEGFs) and interleukin 8.^{47,52} A study performed by Kyung and colleagues observed TGF- β 1 induced expression of vascular endothelial growth factor-C (VEGF-C) by activating the smad pathway in a gastric cancer cell line.⁵³ Furthermore, TGF- β 1 induces epithelial cell growth

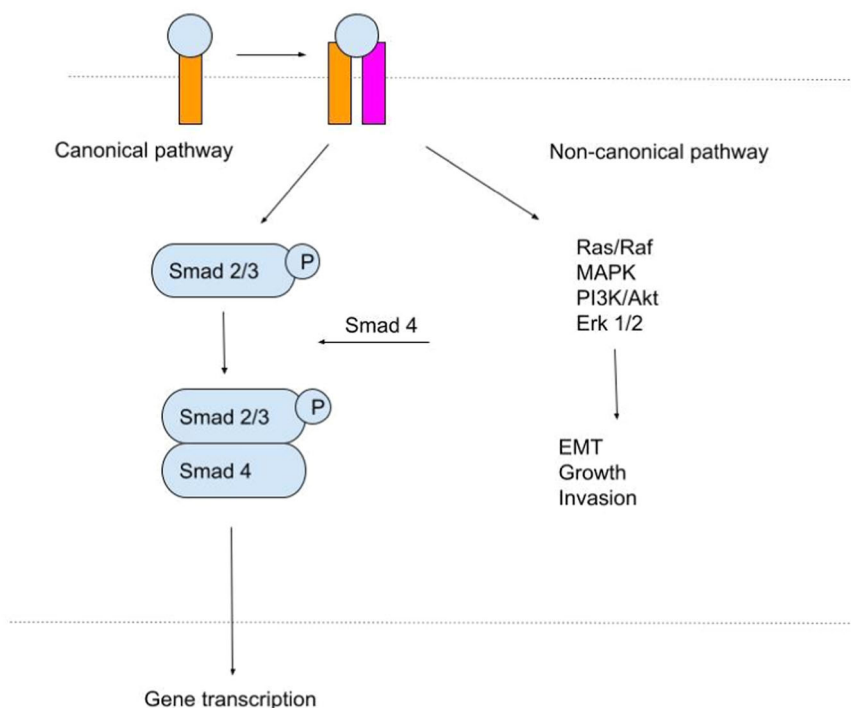


FIGURE 1 Transforming growth factor-beta signalling pathway. TGF- β 1 binds to T β R II (receptor II) which recruits and phosphorylates T β R I (receptor I) resulting in the activation of the canonical and noncanonical signalling pathways. Active canonical pathway results in the recruitment and phosphorylation of smad 2/3 and smad 4 proteins which migrate to the nucleus and induce gene transcription. Noncanonical signalling includes Ras/Raf, MAPK, PI3K/Akt and Erk1/2 signalling pathways which promote cell growth, proliferation and survival.³⁶⁻³⁹ (Image was designed by K. Letsoalo in Microsoft Word 2016). EMT, epithelial mesenchymal transition; MAPK, mitogen activated protein kinase; PI3K, phosphoinositide 3-kinases.

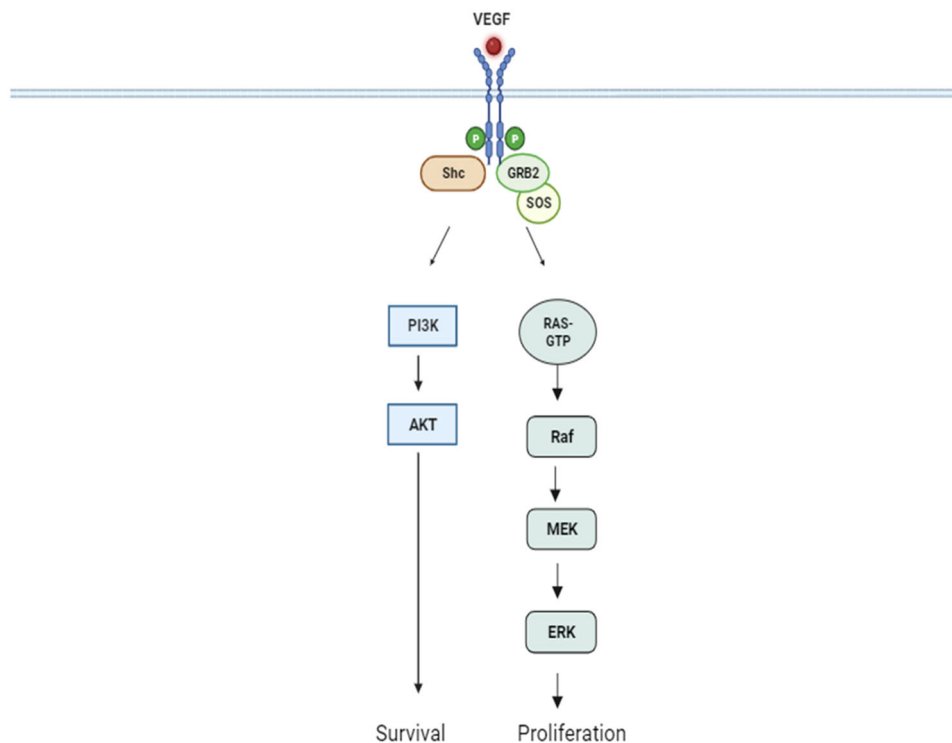


FIGURE 2 VEGFC signalling pathway. VEGF-C binds to its receptors resulting in receptor homo or heterodimerization. Receptor tyrosine residues are phosphorylated creating docking sites for adapter proteins. Ligand binding transduces signalling through the MAPK and PI3K/Akt signalling cascades which promote tumour cell proliferation and survival.⁵⁸ (Image was designed by K. Letsoalo with [BioRender.com](#)). GRB2, growth factor receptor binding protein 2; Shc, src homology containing protein; SOS, son of sevenless protein; PI3K, phosphoinositide 3-kinases; VEGF, vascular endothelial growth factor.

prompting the development of peritumoral neo-vasculature thus, prompting tumour growth and metastasis.⁵²

7.2 | Vascular endothelial growth factors and their receptors in melanoma signalling

The differentiation of endothelial cells during the embryonic period, postnatal vasculature development and sprouting of vessels during pathology is reliant on VEGFs and their corresponding receptors (VEGFRs).⁵⁴ VEGFs are structurally related angiogenic and lymphangiogenic factors that are constituents of the PDGF subgroup of the growth factor cystine knot group.⁵⁵ In mammals the VEGF family consists of VEGF-A, B, C and D, placental growth factor (PlGF) as well as the homologs that are expressed in Orf viruses including VEGF-E and VEGF-F expressed in snake venom.^{55–57} Alternative splicing and proteolytic processing yield various isoforms of the proteins.^{55,56}

VEGFs induce signalling through their receptor tyrosine kinases, VEGFR 1,2 and 3 with ligand binding yielding receptor homo or heterodimerisation, conformational changes and auto or transphosphorylation of tyrosine residues, ultimately activating signalling cascades that promote cell proliferation, survival and migration.^{55,58}

The primary mechanism of mortality in cancer patients is the dissemination of tumours to secondary locations through the

vasculature.⁵⁹ Tumour dissemination to secondary locations utilises three mechanisms mainly: direct spread (invasion of surrounding tissues and organs), hematogenous (metastasis to secondary locations using the bloodstream) and lymphatic metastasis (metastasis to secondary locations and lymph nodes utilising the lymphatic vasculature).⁶⁰ VEGF-C binding to VEGFR-3 promotes lymphangiogenesis, providing a metastatic route for melanoma cells.

7.2.1 | VEGFR-3/VEGF-C signalling pathways

VEGF-C is a lymphangiogenic growth factor signalling through VEGFR-2 and 3.⁵⁶ VEGF-C and its receptors are predominantly expressed by endothelial cells however, their expression is observed in tumour cells thus paracrine/autocrine signalling between tumour cells, vasculature and nonendothelial cells activates the VEGFR-3/VEGF-C axis inducing biological responses including tumour growth, proliferation and migration.⁵⁶

Ligand binding yields receptor dimerization and autophosphorylation of tyrosine residues Y1230/Y1231, recruiting adapter proteins: growth factor receptor binding protein 2 (grb-2) and src homology containing protein (Shc).⁶¹ Active receptors activate the MAPK and PI3K/Akt pathway (Figure 2).⁶¹

7.2.2 | Role of VEGF-C/VEGFR-3 signalling pathway in promoting lymphangiogenesis

Tumour growth and metastasis to regional lymph nodes is the initial step in melanoma dissemination and serves as a significant indicator of disease prognosis.⁶² In melanoma lymphangiogenesis is the emergence of lymphatic vessels from pre-existing vessels and is induced by VEGF-C binding to its receptor VEGFR-3 expressed by endothelial cells, inducing lymphatic endothelial cell (LEC) proliferation and the development of peritumoral and intratumoral vessels thus, promoting metastasis to lymph nodes.⁶³

In the past, limited knowledge and research of lymphatic molecular markers impeded research that would allow researchers to distinguish blood vessels from lymphatic vasculature in the TME and thus elucidate lymphatic metastasis.^{59,62} However, the emergence of VEGF-C and D, their receptors and lymphatic vessel endothelial hyaluronan receptor-1 (LYVE-1), a lymphatic specific marker, has contributed to the significance of the lymphatic vasculature in promoting metastasis in various malignancies including melanoma.^{59,62}

A study performed by Liu and colleagues established the contribution of VEGF-C and D in promoting lymphangiogenesis and lymph node metastasis.⁶⁴ The continuous overexpression of these factors by tumour cells and stroma promotes lymph vessel growth cultivating metastasis.⁵⁹ In a melanoma clinical study, VEGF-C overexpression correlated with SLN spread.⁶⁵ Additionally, a melanoma animal model observed a positive relation between overexpressed VEGF-C and lymphatic vessel quantity and peritumoral vessel diameter.⁵⁹ However, a soluble VEGFR-3 inhibitor minimised tumour induced lymphangiogenesis and metastasis.⁵⁹ Several tumours express elevated concentrations of growth factor C and/or D with growth factor expression paralleling lymphatic vessel density (LVD), lymphatic metastasis and disease outcome.⁶⁶ From these studies it is evident that the VEGF-C/VEGFR-3 signalling pathway is significant in fostering a metastatic niche by promoting lymphatic metastasis.

Tumours procure a premetastatic niche at the SLN before metastasis taking place.⁶⁷ The phenomenon was initially observed and exemplified by Michael Detmar and his colleagues in a skin-cancer animal model.⁵⁹ SLN metastasis is coupled with increased distant metastatic prevalence as well as hematogenous spread justifying that hematogenous and lymphatic spread act synergistically to promote distant metastasis.⁵⁹ Primary tumour cells secrete VEGF-C/A which are transported via the lymphatic vessels to SLN where they procure a premetastatic niche. Arrival of the tumours at the SLN results in elevated concentrations of VEGF-C/A which journey to distant lymph nodes resulting in the increased capacity of the lymph vessel network.^{66,67} SLN lymphangiogenesis is correlated with increased metastasis at distant lymph nodes however, the absence of lymph node metastasis impedes distant organ metastasis thus, substantiating the contribution of the lymph nodes and vessel in tumour dissemination.⁶⁶ Additionally, chemokines secreted by LEC employ the lymphatic vasculature to generate a gradient inducing

directional migration of tumour cells from the primary location to secondary sites promoting lymph node and distant metastasis.⁶⁶

8 | CONTRIBUTION OF CHEMOKINE SIGNALLING IN LYMPHATIC METASTASIS

Chemokines are cytokines consisting of a wide-range family of small (8–15 kDa) proteins characterised by their chemotactic properties and similar composition, facilitating heparin binding and the regulation of the host's defence system.⁶⁸ Chemokines are characterised by their amino acid sequence and positioning of the cysteine residues within the protein.⁶⁹ Variation in the N-terminal cysteine organisation gives rise to four subgroups/families mainly: CC, CXC, CX₃C and XC with C denoting the N-terminal domain and X denoting an amino acid.^{69,70} Initially chemokines were named according to their function however, in the year 2000 a systemic nomenclature system was described that includes subfamily designation (CC, CXC, CX₃C and XC) followed by L (representing the ligand) and a number representing the gene it was initially isolated from.⁷⁰

C-X-C chemokine receptor 4 (CXCR4) and its corresponding ligand C-X-C chemokine ligand 12 (CXCL12) are the most widespread chemokine receptor/chemokine pair in various malignancies including melanoma.^{5,71} Under normoxic conditions, chemokine signalling is responsible for foetal development, tissue repair and the homing of hematopoietic cells and leukocytes to specific sites and tissues. However, tumourigenesis is characterised by the directional migration of receptor-expressing tumour cells to ligand-expressing metastatic sites.⁶⁶

CXCR4 upregulation is a key metastatic factor in various malignancies including human melanoma. CXCL12 binds to CXCR4 and activates a plethora of signalling cascades that promote melanoma growth, adhesion, angiogenesis and migration.⁷² Kim and colleagues observed the role of chemokine receptors in melanoma and colorectal cancer liver metastasis.⁷³ Microarray analysis classified CXCR4 as the most prominent receptor in both cell lines; 89% of melanoma patients and 97% of colorectal cancer patients expressed CXCR4. Additionally, *in vitro* treatment with CXCL12 increased cell migration in the cell lines.⁷³ These findings correlate with other cancer research outputs classifying CXCR4 expression as a prognostic factor. In addition, inhibition of CXCR4 signalling pathway abrogates cell migration in metastatic melanoma.^{74,75} Therefore, the CXCR4/CXCL12 signalling pathway is prominent in melanoma metastasis and thus, targeting this pathway provides a therapeutic approach to prevent migration.

Lymph nodes express elevated concentrations of CXCL12 driving a gradient that facilitates the attraction of CXCR4-expressing malignant cells, therefore, promoting directional migration toward the lymph nodes. Moreover, tumour-associated lymphatic vasculature and not normal lymphatics express CXCL12, justifying the role of the lymphatic endothelium in metastatic spread.⁶⁶ Evidently, the CXCR4/CXCL12 gradient induces distant metastasis mainly to CXCL12-expressing tissue such as the lungs, liver and bone-melanoma

metastatic sites.⁶⁶ A study performed by Kim and colleagues observed the proximity of CXCR4⁺ melanoma cells with CXCL12-producing lymphatic vessels in metastatic lymph nodes and lung tissue with CXCR4⁺/CD133⁺ cells presenting a greater metastatic activity than CXCR4/CD133⁻ cells.⁷⁶ Moreover, inhibition of CXCR4 abolished melanoma growth and metastasis thus, justifying the role of the CXCR4/CXCL12 axis in promoting lymph node and distant metastasis by adopting the lymphatic vasculature.⁷⁶

8.1 | CXCR4/CXCL12 signalling in melanoma

Signalling through CXCR4/CXCL12 prompts downstream signalling cascades resulting in multiple responses such as chemotaxis, cell survival, proliferation and gene transcription.^{77,78} Ligand binding initiates G-protein activation through the exchange of guanine diphosphate (GDP) for guanine triphosphate (GTP) resulting in the dissociation into GTP-bound and α and $\beta\gamma$ subunits.⁷⁸ Dissociated $\beta\gamma$ subunits initiate the activation of phospholipase-c- β (PLC- β) and PI3K. Phospholipase-c- β cleaves phosphatidylinositol (4,5)-bisphosphate into secondary messengers mainly: inositol (1,4,5)-triphosphate (IP3) and diacylglycerol (DAG). Inositol (1,4,5)-triphosphate prompts intracellular Ca²⁺ release through binding to its endoplasmic reticulum receptors, Figure 3.⁷⁹

G α and G $\beta\gamma$ subunits drive PI3K activation resulting in the phosphorylation of focal adhesion proteins such as focal adhesion kinase (FAK), cytoskeletal protein paxillin and proline-rich kinase-2 and thus facilitate cell migration by reorganisation of the actin cytoskeleton.⁶⁸ Active PI3K promotes the rapid production of phosphatidylinositol (3,4,5)-triphosphate prompting Akt pathway activation.⁸⁰ Moreover, the active Akt pathway induces the activation of Bcl-2 associated agonist of cell death (BAD), an antagonist of B-cell lymphoma 2 (BCL-2), thus contributing to cell survival.⁶⁸ Additionally, CXCR4 signalling through Akt diminishes the activity of glycogen synthase kinase-3 beta (GSK3 β)⁸⁰ and initiates the stabilization of β -catenin which translocates to the nucleus and prompts gene transcription and proliferation, Figure 3.⁶⁸

9 | INTRACELLULAR ADHESION PROTEINS AND INTEGRINS ACTIVATED BY CHEMOKINES BINDING THEIR SPECIFIC RECEPTORS

9.1 | Integrins

Integrins are large, complex and heterodimeric glycoproteins that bridge signal communication between the internal and external environments.⁸¹ These proteins are composed of alpha and beta subunits forming noncovalent heterodimers. In mammals, there are 18 alpha and 18 beta subunits, giving rise to 24 distinct integrin heterodimers.⁵ Integrins primarily bind to various components of the ECM such as vitronectin, fibronectin, laminin or collagen. This binding provides essential anchorage for cell adhesion and invasion

processes.⁸¹ Integrin binding to the ECM induces their clustering in the membrane plane and enables them to recruit and activate several signalling and adapter protein such as Src family kinases (SFKs), FAK and scaffolding molecules such as p130CRK-associated substrate to assemble focal adhesions (FA).⁸² Moreover, they integrate the ECM into the actin cytoskeleton by recruiting proteins such as paxillin, talin, α -actinin, vinculin and tensin. Thus, FA protein recruitment and activity is regulated by integrin activity and directs cell adhesion and migration.⁸¹ Integrins serve as mediators of bidirectional signalling where intracellular signalling transforms extracellular process (inside-out signalling); conversely, extracellular ligand binding activates intracellular signalling cascades (outside-in signalling) by activating the Ras/Rho signalling pathways.⁸³

9.2 | Focal adhesion kinase background

FAK is a 125kDa⁸⁴ nonreceptor tyrosine kinase whose activity is regulated by integrin signalling, GPCRs, cytokines and growth factors.⁸⁵ FAK mediates multiple cellular processes including proliferation, survival, adhesion and migration.⁵ Furthermore, FAK advances tumour stemness, epithelial mesenchymal transition, chemotherapeutic resistance tumour angiogenesis and fibrosis in the stroma. FAK is a ubiquitously expressed protein with three domains: an N-terminal domain, a central kinase domain and a C-terminal focal adhesion targeting (FAT) domain.⁸⁵ The C-terminal domain associates with FA-associated proteins such as talin and paxillin. Subsequent to growth factor or integrin signalling, FAK is autophosphorylated at Y397 resulting in the formation of a Src binding site which further phosphorylates other tyrosines on FAK therefore, yielding additional binding sites for Src homology 2 (SH2) domain-containing proteins.^{84,85} Moreover, an active FAK/Src complex activates downstream proteins including paxillin.⁸⁴

9.2.1 | Focal adhesion kinase in the development of melanoma

The FAK promoter region contains a p53 binding site where wild-type p53 inhibits FAK transcription.⁸⁶ However, mutant p53 which is observed during malignancy is incapable of binding to the promoter region and thus, displays no inhibitory effects on FAK promoter activity, therefore, promoting continuous FAK transcription and overexpression.⁸⁶

Anoikis is a form of programmed cell death initiated because of obstructions between the cell and the ECM and the loss of FAK activity.^{87,88} Overexpression of FAK encourages resistance to anoikis despite the detached state of cells from the ECM. Furthermore, increased activity of the FAK/Src complex stimulates the induction of PI3K/Akt and MEK/Erk 1/2 signal transduction, promoting cell survival in the detached conformation.⁸⁷ TGF- β stimulates FAK and Akt expression utilising smad3 and p38 MAPK respectively, conferring anoikis resistance and promoting tumour survival.⁸⁹

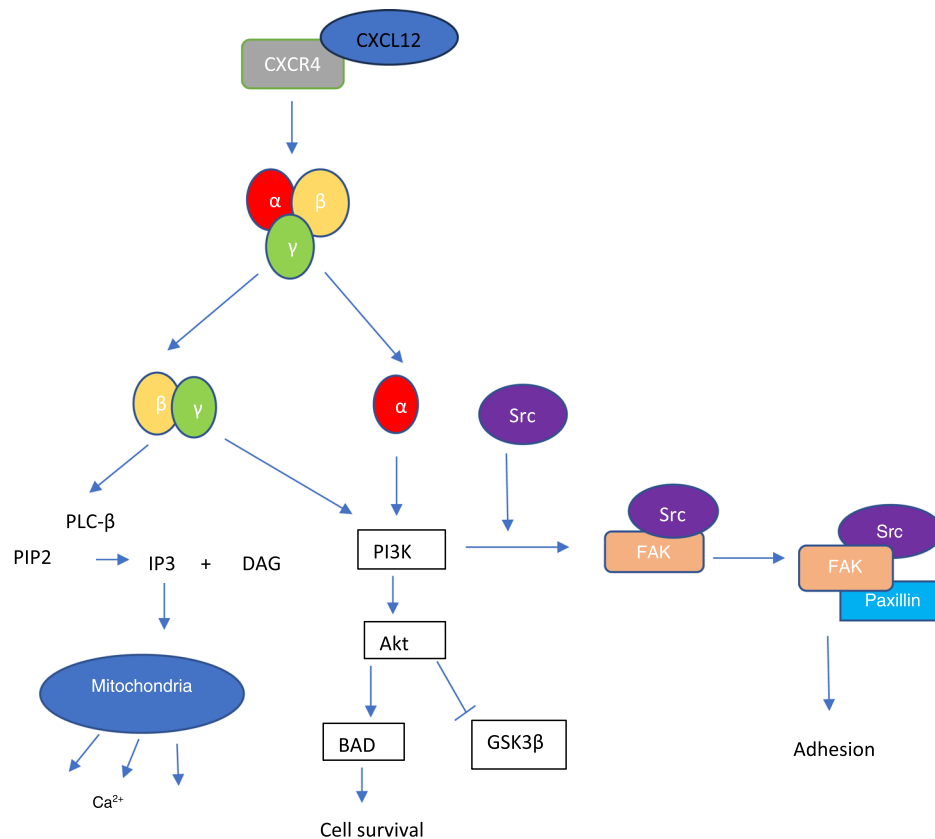


FIGURE 3 CXCR4/CXCL12 signalling pathway. Active CXCR4/CXCL12 results in the exchange of GDP for GTP resulting in the dissociation of G α and G $\beta\gamma$ subunits. G α and G $\beta\gamma$ activate PI3K driving adhesion through the activity of FAK and paxillin. PI3K additionally inhibits GSK3 β and activates BAD protein promoting tumour cell survival. G $\beta\gamma$ subunits induce the production of IP3 and DAG with IP3 promoting mitochondrial calcium secretion.^{67,74,76} (Image was designed by K. Letsoalo in Microsoft Word 2016). BAD, Bcl-2 associated agonist of cell death; CXCL12, CXC chemokine ligand 12; CXCR4, CXC chemokine receptor 4; DAG, diacylglycerol; FAK, focal adhesion kinase; GSK3 β , glycogen synthase kinase-3 beta; IP3, inositol (1,4,5)-triphosphate; PI3K, Phosphoinositide 3-kinases; PIP2, phosphatidylinositol (4,5)-bisphosphate; PLC- β , phospholipase-c- β .

Additionally, FAK overexpression inhibits the induction of caspase-3-mediated apoptosis whereas FAK inhibition induces apoptosis.⁸⁷

FAK overexpression and phosphorylation is associated with cell-cycle progression through modulation of cell-cycle proteins, this further reiterates the role of FAK in tumour cell survival.⁸⁷ Overexpressed FAK facilitates cyclin D1 expression and the inhibition of p21 therefore, prompting cell-cycle progression through the G1 phase. Overexpressed FAK is delineated to regulate the E26 transformation specific (ETS) binding site located within the cyclin D1 promoter which further modulates the transcription of cyclin D1.⁸⁷ FAK further promotes cell-cycle progression by inhibition of p53 tumour suppressor mediated apoptosis.⁸⁷ Therefore, FAK regulates cancer cell proliferation by modulating cell-cycle molecules or promoting turnover of tumour suppressor proteins.

9.3 | Paxillin background

Paxillin is the principle constituent of FAs which plays a critical role in signal transduction following the interaction between the ECM and

integrins.^{90,91} Paxillin is a scaffolding protein and recruits kinases and phosphatases, co-factors, structural and oncoproteins whose activity is required for intracellular signalling.⁹¹ Activation of the above-mentioned proteins reorganises the actin cytoskeleton and induces assembly/disassembly of FAs that are essential for cell adhesion, metastasis and migration.⁹¹ Paxillin positively regulates cell adhesion as it is recruited to nascent FAs at the anterior of the cells inducing the assembly of adhesion complexes; conversely, it is attributed to FA disassembly at the rear end of the cell during cell migration.⁹¹ Paxillin activity is not only localised to FAs, but also to the cytoplasm and nucleus with its activity exerted on gene transcription. Therefore, it bridges signal transduction from the plasma membrane and cytoskeleton to the nucleus.⁹¹ Despite its interaction with protein complexes and enzymes, paxillin does not present any enzymatic activity itself. Instead, paxillin serves as a docking site for other proteins, facilitating the assembly of multiprotein complexes.⁹¹

Paxillin activity and localisation is tightly regulated by its phosphorylation in response to diverse stimuli. Tyr31 and Tyr118 are the well-known phosphorylation sites however, paxillin may also be phosphorylated at various serine and tyrosine residues.⁹¹ In

response to integrin association with the ECM, paxillin is phosphorylated prompting the recruitment of proteins such as talin, vinculin, tensin and FAK that are required for FA assembly.⁹¹ Paxillin phosphorylation at the tyrosine residue provides a scaffold for the recruitment of FAK and Src. Furthermore, Src-induced phosphorylation at Tyr88 and Tyr118 as well as phosphorylation at Tyr118 and Tyr31 by FAK at the N-terminus initiates paxillin interaction with downstream effectors inducing p130Crk-associated substrate (p130cas) extracellular signal transduction into cellular responses mediated by MAPK.⁹¹

9.3.1 | The role of paxillin in cancer development

Paxillin acquires gain of function mutations which are associated with tumour progression in several malignancies including melanoma.⁹² Paxillin functions in conjunction with other adhesion proteins to regulate cell migration and adhesion thus, enhancing metastasis.^{91,92} To evaluate the role of paxillin in melanoma metastasis Velasco-Velazquez and colleagues transfected B16-F10 melanoma cells with paxillin-siRNA.⁹³ Transfection significantly inhibited the metastatic potential of the B16-F10 cells.⁹³ Furthermore, increased levels of phospho-paxillin have been observed in melanoma cells compared to melanocytes.⁹³ To further determine the role of paxillin in melanoma metastasis, B16-F10 cells were treated with 4-hydroxycoumarin (4-HC). 4-HC reduced α and β isoforms of paxillin mRNA levels.⁹³ Additionally, 4-HC mitigated paxillin signalling pathways resulting in decreased phosphorylation of FAK and GTP-bound Rac-1.⁹³ Therefore, it is evident that paxillin is a significant contributing factor of melanoma metastasis.

10 | POTENTIAL SYNTHETIC COMPOUND TO INHIBIT MELANOMA CELL PROLIFERATION

10.1 | MAZ-51(3-(4-dimethylaminonaphthalen-1-ylmethylene)-1,3-dihydroindol-2-one)

MAZ-51 (3-(4-dimethylaminonaphthalen-1-ylmethylene)-1,3-dihydroindol-2-one) is an indolinone-based molecule synthesised to inhibit VEGFR-3 phosphorylation in endothelial cells.^{94,95} Indolinones contain distinct amino acid components at position three that form part of adenosine triphosphate-competitive inhibitors of receptor kinases and have been proven to bind to specific receptor tyrosine kinases such as fibroblast growth factor, epidermal growth factor, PDGF and VEGF.^{90,94} Indolinone derivatives exhibit anti-proliferative properties and induce apoptosis in cancer and endothelial cell lines. MAZ-51 competitively binds to VEGFR-3 inhibiting VEGF-C induced phosphorylation of VEGFR-3 therefore, inactivating signalling cascades that promote cell survival and proliferation in endothelial and cancer cell lines.⁹⁰ To our knowledge, limited studies have observed the effect of MAZ-51

on melanoma cell proliferation. However, melanoma is a VEGFR-3-expressing tumour and has been proven to utilise the VEGFR-3/VEGF-C signalling pathway to promote lymphatic metastasis thus, we hypothesize that VEGFR-3 expressed by melanoma cells will be sensitive to MAZ-51 treatment.^{90,96} A study conducted by Lee and colleagues observed the effect of MAZ-51 on VEGFR-3 inhibition in B16-F10 melanoma cell line.⁹⁵ MAZ-51 treated cells exhibited a decrease in intratumoural lymphatic vessels in the lungs, decrease in tumour size as well as a decrease in the expression of VEGF-C, VEGFR-3 and Prox-1.⁹⁵ Therefore, from the available literature, it is evident that MAZ-51 is a potential therapeutic strategy for melanoma by targeting the lymphangiogenic pathway.

11 | THE ROLE OF PHYTOCHEMICALS AS POTENTIAL CHEMOTHERAPEUTIC AGENTS

11.1 | Implications of using phytochemicals/plants for cancer treatment

Harmonious to the limited efficacy of conventional cancer treatments, the use of medicinal plants and their bioactive compounds as anticancer agents has been accepted as a form of medical intervention.⁹⁷ The use of medicinal plants for primary healthcare is a long-standing practice with approximately 80% of populations in developing countries being reliant on phytomedicine for their primary health care.^{98,99} Their accessibility, low costs and reduced side effects contribute to their increased consumption in comparison to synthetic drugs.⁹⁸ The health-promoting effects of phytochemicals are attributed to their biological properties such as antioxidants, anti-inflammatory, antimicrobial and anticancer activities.¹ With the increase in efficacious experimental findings, phytochemicals are postulated to possess significant anticancer capabilities.⁹⁹

Carcinogenesis is dependent on the aberrant activation of signalling pathways that promote cell growth, proliferation and inhibit the sensitivity of pathways to regulatory molecules that maintain a homeostatic balance.¹⁰⁰ The proposed mechanism of action utilised by phytochemicals is their ability to regulate signal transduction pathways by increasing proapoptotic proteins and decreasing antiapoptotic proteins, increasing expression of regulatory proteins thus, promoting cell cycle arrest, decreasing the sensitivity of cells to mitogens and mitigating invasion and metastasis by inhibiting epithelial mesenchymal transition (EMT).^{99,101} Several signalling pathways such as the MAPK/Erk and PI3K/Akt are dysregulated during pathogenesis promoting cancer cell growth and metastasis.¹⁰¹ Targeting these pathways provides a solid foundation to prevent and treat malignancies. Table 2 provides a list of phytochemicals that have been investigated for melanoma and provides their mode of action in combating malignancy.

Both in vitro and in vivo studies advocate for phytochemicals as alternative treatment modalities for cancer. However, clinical studies supporting the sole use of phytochemicals are limited due to challenges such as their low bioavailability and the high doses

required for effective treatment outcomes.^{111,112} The combined application of medicinal plants with existing cytotoxic drugs has garnered significant attention and widespread acceptance. This approach is favoured for its potential to decrease the chemotherapeutic drug dose, reduce toxicity, enhance drug bioavailability, lower resistance development and capitalise on the synergistic effects of phytochemicals and synthetic drugs to jointly inhibit tumour cell proliferation.¹¹³

One such phytochemical that has been identified as a potential anticancer agent is zingerone.

11.2 | Zingerone

Zingerone (4-(4-hydroxy-3-methoxyphenyl)-2-butanone) is a non-toxic bioactive phytochemical compound that has pharmacological activities.¹¹⁴ It is a ketone that is a 4-phenylbutan-2-one. It belongs to a group of compounds called methoxyphenols where a methyl group is attached to a benzene ring. Zingerone contains pharmacological properties including antidiabetic, anti-inflammatory, antidiarrheic, antipolytic, antispasmodic and anticancer properties.^{114,115}

Zingerone's chemoprotective properties have been observed in *in vitro* and *in vivo* studies where it induces cell cycle arrest in neuroblastoma cells, prevents angiogenesis and its antiproliferative

effects investigated in rat colon cancer, ovarian cancer and colorectal cancer.^{116,117} Zingerone possesses several pharmacological properties that render it a suitable chemopreventative and chemotherapeutic agent. The following section details the pharmacological properties of zingerone in cancer management.

11.2.1 | Mechanism of action of zingerone against melanoma development

Zingerone's chemotherapeutic properties have been observed in multiple malignancies however,¹¹⁶⁻¹¹⁸ to our knowledge zingerone's ability to induce melanoma cell death has not been studied. Nevertheless, acetyl zingerone—a derivative of zingerone—has been shown to ameliorate DNA mutations in melanocytes following sun exposure.¹¹⁹⁻¹²¹ Cyclobutane pyrimidine dimers (CPDs) are UV-radiation-induced photoproducts resulting in DNA lesions and mutations and are strongly correlated with melanoma and non-melanoma skin cancer.^{119,121} Additionally, CPDs are correlated to photoaging and the production of immunomodulatory cytokine tumour necrosis factor alpha which has been shown to promote melanoma signalling pathways.¹¹⁹ Studies by Chaudhuria and colleagues and Srivastava and colleagues observed that acetyl zingerone is effective in inhibiting the formation of CPDs in

TABLE 2 List of phytochemicals utilised for melanoma treatment.

Phytochemical name	Source	Mode of action	References
Fisetin	Apples, onions, grapes, cucumbers, and strawberries	Promotes mesenchymal to epithelial transition and targets the NFκB and MAPK signalling	Chandra Pal et al. ^{21,102}
Indole-3-carbinol	Broccoli and brussels sprouts	Stabilises PTEN. Induces G1 cell cycle arrest and apoptosis.	Aronchik et al. ¹⁰³
Epigallocatechin gallate	Green tea	Inhibits expression of PD-L1 and PD-L2. Induces cell cycle arrest and apoptosis.	Chandra Pal et al. ²¹ and Ravindran Menon et al. ¹⁰⁴
Eugenol	Cloves, bay leaf and cinnamon leaf	Induces S-phase cell cycle arrest and apoptosis. Inhibits E2F1.	Ghosh et al. ¹⁰⁵
Resveratrol	Grapes, mulberries, peanuts, eucalyptus, and cranberries	G1/S cell-cycle arrest. Upregulates p53.	Chandra Pal et al. ²¹ and Pourhanifeh et al. ¹⁰⁶
Capsaicin	Chilli peppers	Activates caspase 3, 8 and 9. Downregulates Bcl-2.	Chandra Pal et al. ²¹
Apigenin	Parsley, celery, artichokes	Downregulates ERK 1/2 and PI3K/Akt signalling	Chandra Pal et al. ²¹
Genistein	Soybeans	Inhibits angiogenesis, proliferation, and metastasis and promotes apoptosis. Upregulates p53, p21 and checkpoint kinase 2.	Chandra Pal et al. ²¹
Curcumin	Turmeric	Targets Akt, NFκB and AP-1	Chandra Pal et al. ²¹
Silymarin	<i>Silybum marianum</i> L. Gaertn	Induces apoptosis and cell cycle arrest. Downregulates Bcl-2 and upregulates Bax.	Chandra Pal et al. ²¹ and Vaid et al. ¹⁰⁷
Procyanidin	Cocoa, berries, apples, grapes	Targets 67 kDa Laminin receptor signalling	Bae et al. ¹⁰⁸
Luteolin	Broccoli, raw brussels sprouts, carrots, peppers and parsley	Inhibits expression of MMP-2 and 9. Targets PI3K/Akt pathway.	Rocchetti et al. ¹⁰⁹ and Yao et al. ¹¹⁰

melanocytes following UV exposure.^{119,121} Acetyl zingerone inhibits the formation of CPDs by upregulating the expression of nucleotide excision repair pathway, decreasing reactive oxygen species (ROS) and neutralizing free radicals and scavenging peroxynitrite.^{119,121}

According to the literature, zingerone promotes cancer cell death by inhibiting the disintegration of the ECM, inhibiting angiogenesis, promoting cell cycle arrest and apoptosis.^{108,122,123} However, in melanoma cells, zingerone targets oxidative stress by inhibiting the formation of ROS. In melanocytes, melanin suppresses the formation of ROS. However, during malignancy melanogenesis is a source of oxidative stress. Overexpressed ROS induces a melanocyte homeostatic imbalance thus compromising their viability and fostering their malignant transformation.¹²⁴ Taking into consideration the antioxidant activity of zingerone and acetyl zingerone and the role of ROS in promoting melanoma, we hypothesize that zingerone is a suitable therapeutic strategy to target oxidative stress and thus, inhibit melanoma genesis.

Current cancer treatment approaches utilise cytostatic cancer drugs to halt cancer cell growth and proliferation. However, these drugs are inefficiently metabolised and are excreted into environmental water systems through bodily waste. This raises concerns about their impact on the environment, with drug residues detected in water bodies and soil. Striking a balance between effective cancer treatment and minimising environmental harm is a pressing challenge, urging the development of more eco-friendly treatment strategies. The next section of the review provides a detailed discussion of the impact of cytostatic cancer drugs on the environment.

12 | THE EFFECTS OF CYTOSTATIC CANCER DRUGS ON THE ENVIRONMENT

Cancer is the second leading cause of death with its global burden on the rise, 9.6 million cancer-related deaths were recorded in 2018¹²⁵ with an estimation of 29.5 million cases will be recorded in 2040.¹²⁶ The increase in cancer incidence is positively correlated with an increase in the prescription of chemotherapeutic drugs and the presence of antineoplastics (APs), anticancer drugs, in the aquatic environment.¹²⁵

Anticancer drugs are inefficiently metabolised by the human body and are excreted either as the parental compound or its metabolite via urine and faeces, some traces are found in sweat and vomit.¹²⁶ Cancer treatment frequently takes place at the hospital with wastewater derived from hospitals and pharmaceutical factories being noteworthy contributors of anticancer drugs in the aquatic environment.¹²⁶

Elevated concentrations of AP agents,¹²⁵ reaching levels on the order of micrograms per liter ($\mu\text{g/L}$), have been identified in hospital effluents.¹²⁷ Despite this, there are limited environmental regulations overseeing their safety thresholds and wastewater treatment options for these compounds.¹²⁵ The presence of APs in untreated wastewater may result in acute and chronic effects in vulnerable aquatic species such as zebrafish and crustaceans.¹²⁷

Chronic exposure to APs alters the genetic composition and cell-cycle of aquatic flora and fauna with researchers deeming them as pseudo-resistant pollutants.¹²⁸ Individual APs are present at low concentrations with limited aquatic effects however, their combined effect results in additive and synergistic effects.¹²⁹ The combination of cyclophosphamide (CP), ifosfamide (IF) and their metabolites in alga *Synechococcus leopoliensis* brought about increased growth inhibition in comparison to the individual drugs.¹¹⁶ A study performed by Elserk and colleagues indicates that a single high dose of APs is comparable to the combination of multiple APs at low concentrations.¹²⁸ Additionally, bioaccumulation and biomagnification of APs may further increase their concentration in the aquatic environment.¹²⁸

The presence of APs in the environment is a well-known phenomenon however, information about their biological effects on organisms is scarce.^{126,127} Anticancer agents are cytotoxic and cytostatic drugs that target cancerous and noncancerous cells thus, eukaryotic organisms are prone to the side effects.¹²⁷ With the global incidence of cancer on the rise, several AP agents have been detected in the aquatic environment with the five most widely studied being: CP, IF, methotrexate, tamoxifen and 5-fluorouracil (5-FU).¹²⁵

Tamoxifen, an oestrogen receptor antagonist, is used individually or in combination with other drugs to treat hormone receptor-positive breast cancer.^{125,127} A study performed by Pagano and colleagues observed the effects of tamoxifen on fertilization, embryogenesis and mitotic effects in the sea urchin.¹²⁷ Researchers observed the effects of tamoxifen on developmental toxicity resulting in early embryonic death. CP is an alkylating agent utilised for the treatment of various malignancies including ovarian and breast cancer, retinoblastoma, multiple myeloma and mycosis fungoides to name a few.¹³⁰ Li and colleagues observed the effects of CP at concentrations of 0.5–50 $\mu\text{g/L}$ on zebrafish.¹³⁰ Results indicated that exposure to CP induced malformation, histopathological alterations in the retina and liver and decreased swimming mobility of the zebrafish.¹³⁰ Additionally, CP hampered the transcription of genes required for the MAPK signalling pathway.¹³⁰ Novak and colleagues studied the individual effects of CP and IF as well as in combination with 5-FU and cisplatin (CDDP) in zebrafish liver cell lines.¹³¹ Individually, CP and IF induced minimal cytotoxicity however, in combination with 5-FU and CDDP, the compounds induced DNA strand breaks.¹³¹

Chemotherapeutics, although required for cancer treatment enter the wastewater system and endanger the aquatic life with their combinations acting additively and/or synergistically.¹²⁹ The intersection of cancer treatment and ecological preservation underscores a call for sustainable progress, aligning with multiple UN Sustainable Development Goals (SDGs).

13 | CONTRIBUTION OF CHEMOTHERAPEUTICS TO THE SDGs

In cancer treatment, residual chemotherapeutic agents infiltrating aquatic systems impact SDGs 14 and 15 (life below water and on land). Nanotechnology offers promise, aligning predominantly with SDGs 3 (good health) and 9 (innovation). Engineered nanoparticles,

targeting drug delivery, reinforce SDG 6 (clean water). Addressing contamination pathways resonates with SDGs 6 and 12 (responsible consumption). Enhanced wastewater management and innovative treatment resonate with SDGs 6 and 11 (sustainable cities). The synthesis of medical exigency and ecological imperatives advances SDG 3 while nurturing the planet (SDG 13).

13.1 | Sustainable development goal 3—Good health and well-being

Anticancer drugs are administered both within hospitals and to outpatients as a result, both hospital and domestic wastewater are a source of AP agents, entering and contaminating the aquatic ecosystem.¹²⁶ Residential sewage treatment plants are inadequately equipped to process AP waste and thus, serve as a contamination route for surface and groundwater.¹²⁵ Moreover, several developed countries utilise septic tank systems which are ineffective in the removal of pharmaceuticals suggesting an additional route of contamination for the soil and groundwater.¹²⁵

Although concentrations of APs are nonsignificant in drinking water, vulnerable populations such as foetuses, children and breastfeeding individuals are at risk.¹³² Chemotherapeutics target rapidly multiplying cells sensitising foetuses, babies and children to their adverse side effects due to the rapid growth experienced by these groups. However, there is a knowledge gap about the risk associated with exposure through drinking water, skin and oral exposure.¹³²

Chemotherapeutics are genotoxic resulting in the occurrence of malignancy when they interface proto-oncogenes or tumour suppressor genes.¹³³ Furthermore, exposure to genotoxic agents induces sequences of events resulting in the occurrence of adverse side effects. The beneficial effects of chemotherapeutics outweigh the detrimental risks in patients however, healthcare workers and caregivers are exposed to the drugs and are subjected to the side effects without any beneficial activity of the drug.¹³³ Baniasodi and colleagues studied the urinary concentrations of CP and IF in healthcare workers involved in the preparation and administration of chemotherapeutic drugs.¹³⁴ Results observed the presence of CP in five preshift and nine postshift samples. IF was detected in one preshift and four postshift samples whereas, the drugs were not detected in the control group. Additionally, the healthcare workers reported a headache as the most frequent adverse effect.¹³⁴ A study performed by Elshaer recorded the effects of APs on exposed nurses. Thirty one percent of exposed nurses presented with infertility and 36.36% recorded oral ulcers.¹³⁵ Additionally, white blood cell counts were reduced whereas, there was an increase in creatine levels.¹³⁵ Exposure to APs is associated with a twofold increase in spontaneous abortions, congenital malformations, infertility and the development of acute myelogenous leukaemia and myelodysplastic syndrome.¹³³ The use of chemotherapeutics albeit being beneficial for the initial malignancy results in adverse side effects and the development of secondary malignancies. Moreover, exposure of APs to healthcare

workers is associated with the development of several disorders. APs not only affect the health and well-being of patients but also of their caregivers thus, calling for research for alternative treatment options.

13.2 | Sustainable development goal 8—Decent work and economic growth

Limited studies have been conducted to characterise the effects of APs on the aquatic ecosystem.¹²⁶ It is postulated that APs are mutagenic and genotoxic affecting various trophic levels of aquatic life resulting in a decrease in the quality and population of the aquatic ecosystem.¹³¹

Countless people worldwide are dependent on healthy aquatic ecosystems as sources of food and employment. Additionally, aquatic ecosystems sustain economic growth, modulate the climate and aid in the prosperity of coastal communities.¹³⁶ Therefore, it is of utmost importance to sustain and manage this natural resource. The Organisation for Economic Co-operation and Development (OECD) has reported that oceans contribute approximately US\$1.5 trillion annually to economic growth.¹³⁶ According to the Food and Agricultural Organization (FAO), around 58.5 million individuals worldwide are employed in primary fish production, with women accounting for 21% of this workforce.¹³⁶ The department of agriculture, fisheries and forestry states that the fisheries sector has a net worth of approximately R8 billion annually with 28,000 people employed in the commercial sector in South Africa.¹³⁷ Even though the South African fisheries industry contributes less than 1% of the total gross domestic product (GDP) and only 5% of the Western Cape's provincial GDP, this sector is a significant contributor to food security, employment and environmental impact.¹³⁷

Given the fisheries sector's role in ensuring food security and providing employment opportunities, contamination of aquatic ecosystems and oceans can profoundly impact the economic growth and development of countries, particularly developing countries. Many communities in these regions depend on commercial and recreational fishing for both sustenance and job security, making them especially vulnerable to the negative consequences of ecosystem contamination.¹³⁶ Additionally, the decrease in aquatic populations and increase in genetic aberrations affects research and development as several aquatic organisms such as the zebrafish are involved in experimental scientific research.¹³¹ Thus, pharmaceutical contamination of aquatic ecosystems is a cause for concern affecting various sectors that affect economic growth and development.

13.3 | Sustainable development goal 9—Target 9.5, indicator 9.5.1: Research and development spending

Cancer cells are heterogenous exhibiting frequent mutations and insensitivity toward traditional treatment options.¹³⁸ Considering the challenges associated with current chemotherapy, the use of plant-based anticancer agents is on the rise. Although the use of

phytochemicals is gaining attention with several researchers advocating for their employment, they present with low bioavailability, limited genotoxic profiling and variable immune response.¹³⁸

One of the outcomes stemming from the combined use of chemotherapeutics and phytochemicals is the potential for herb-drug interactions. This poses a particular concern for patients undergoing treatment for chronic conditions, as they may be at a higher risk for such interactions.¹³⁹ The most prevalent mechanism behind herb-drug interactions involves the obstruction and/or induction of drug-metabolising enzymes and transport proteins by herbs, resulting in the reduced efficacy of the chemotherapeutic drug.¹³⁹ Accurate analysis of phytochemical-drug interaction and increased bioavailability to the target tissue using advanced imaging tools pave a pathway for successful anticancer drug intervention.¹³⁸

Current knowledge of phytochemical-drug interaction rests on in vitro, in vivo and in silico models with minimal clinical trials studies. Therefore, research of phytochemical interaction in human studies and the development of advanced imaging tools will enhance the understanding of mechanisms of action and predict clinical effects.¹⁴⁰

14 | CONCLUSION

This paper reviewed melanoma, detailing its biology, and signalling pathways. Additionally, it explored the current treatment strategies providing their mechanism of action and their associated side effects and explored the potential use of phytochemicals as adjuvant therapy.

Medicinal plants and their derivatives are gaining traction as potential therapeutic agents due to their reduced toxicity, affordability and capacity to alleviate side-effects often seen with the use of synthetic drugs. Research is ongoing to understand their effectiveness and mechanisms of action. These natural compounds, phytochemicals, are preferred over their synthetic counterparts because of their lower toxic profiles, affordability and ability to minimise side effects. Furthermore, they have a positive environmental impact, contrasting with synthetic drugs that can have unintended consequences, jeopardizing patients, caregivers and our ecosystem. The nontarget effects of synthetic drugs can be detrimental to both human health and the environment.

Although phytochemicals have shown promising results in laboratory settings, their limited bioavailability and stability have hindered their advancement in clinical trials. Contemporary cancer treatments often combine various methods, including surgery, radiation and chemotherapy. However, the concurrent use of multiple synthetic drugs can result in increased resistance to these drugs and unintended harm to healthy cells. Recent research has highlighted the potential benefits of pairing phytochemicals with traditional cancer treatments, suggesting that this combination can reduce the need for synthetic drugs, decrease toxicity and combat drug resistance. This combined approach could lead to more effective and targeted treatments for cancer, ultimately resulting in the destruction of cancer cells.

Incorporating environmental contaminants and SDGs 3 (good health and well-being), 9 (industry, innovation, and infrastructure) and 8 (decent work and economic growth), the findings of this review underscore the importance of sustainable and environmentally friendly treatment options. Using medicinal plants and phytochemicals aligns with these SDGs by promoting health, fostering innovation in treatment modalities and ensuring that the means of production do not harm the environment or the workforce. Further research will be crucial to unlock the full potential of phytochemicals in clinical practice and to address challenges related to their bioavailability and stability. This paves the way for a new era of cancer treatment that is not only more effective but also environmentally conscious.

ACKNOWLEDGEMENTS

The research supporting this review was funded by the National Research Foundation (NRF) of South Africa (N1F580A1F685) and the University of Pretoria's Postgraduate funding scheme.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

No additional data is available.

ORCID

Kganya Letsoalo  <http://orcid.org/0000-0003-1679-5293>

Evangeline Nortje  <http://orcid.org/0000-0002-0042-4466>

Sean Patrick  <http://orcid.org/0000-0002-0364-0068>

Trevor Nyakudya  <http://orcid.org/0000-0003-1872-9257>

Yvette Hlophle  <http://orcid.org/0000-0002-3112-2436>

REFERENCES

- Garcia-Oliveira P, Otero P, Pereira AG, et al. Status and challenges of plant-anticancer compounds in cancer treatment. *Pharmaceuticals*. 2021;14(2):157. doi:10.3390/ph14020157
- The World Health Organization. *Cancer*. The World Health Organization. https://www.who.int/health-topics/cancer#tab=tab_1
- Gordon R. Skin cancer: an overview of epidemiology and risk factors. *Semin Oncol Nurs*. 2013;29(3):160-169. doi:10.1016/j.soncn.2013.06.002
- Arnold M, Singh D, Laversanne M, et al. Global burden of cutaneous melanoma in 2020 and projections to 2040. *JAMA Dermatol*. 2022;158(5):495-503. doi:10.1001/jamadermatol.2022.0160
- Nkandeu DS, Basson C, Joubert AM, et al. The involvement of a chemokine receptor antagonist CTCE-9908 and kynurenine metabolites in cancer development. *Cell Biochem Funct*. 2022;40:608-622. doi:10.1002/cbf.3731
- Eddy K, Chen S. Overcoming immune evasion in melanoma. *Int J Mol Sci*. 2020;21(23):8984. doi:10.3390/ijms21238984
- Juszczak AM, Wöelfle U, Končić MZ, Tomczyk M. Skin cancer, including related pathways and therapy and the role of luteolin derivatives as potential therapeutics. *Med Res Rev*. 2022;42(4):1423-1462. doi:10.1002/med.21880

8. Dika E, Patrizi A, Lambertini M, et al. Estrogen receptors and melanoma: a review. *Cells*. 2019;8(11):1463. doi:10.3390/cells8111463
9. Morgese F, Sampaolesi C, Torniai M, et al. Gender differences and outcomes in melanoma patients. *Onco Ther*. 2020;8(1):103-114. doi:10.1007/s40487-020-00109-1
10. Roh MR, Eliades P, Gupta S, Grant-Kels JM, Tsao H. Cutaneous melanoma in women. *Int J Womens Dermatol*. 2015;1:21-25. doi:10.1016/j.ijwd.2015.01.001
11. Balwierz R, Biernat P, Jasińska-Balwierz A, et al. Potential carcinogens in makeup cosmetics. *Int J Environ Res Public Health*. 2023;20(6):4780. doi:10.3390/ijerph20064780
12. Budden T, Bowden N. The role of altered nucleotide excision repair and UVB-induced DNA damage in melanomagenesis. *Int J Mol Sci*. 2013;14(1):1132-1151. doi:10.3390/ijms14011132
13. Rass K, Reichrath J. UV damage and DNA repair in malignant melanoma and nonmelanoma skin cancer. *Adv Exp Med Biol*. 2008;624:162-178. doi:10.1007/978-0-387-77574-6_13
14. Leonardi G, Falzone L, Salemi R, et al. Cutaneous melanoma: from pathogenesis to therapy (Review). *Int J Oncol*. 2018;52(4):1071-1080. doi:10.3892/ijo.2018.4287
15. Strashilov S, Yordanov A. Aetiology and pathogenesis of cutaneous melanoma: current concepts and advances. *Int J Mol Sci*. 2021;22(12):6395. doi:10.3390/ijms22126395
16. Burns D, George J, Aucoin D, et al. The pathogenesis and clinical management of cutaneous melanoma: an evidence-based review. *J Med Imaging Radiat Sci*. 2019;50(3):460-469. doi:10.1016/j.jmir.2019.05.001
17. Paluncic J, Kovacevic Z, Jansson PJ, et al. Roads to melanoma: key pathways and emerging players in melanoma progression and oncogenic signaling. *Biochim Biophys Acta-Mol Cell Res*. 2016;1863(4):770-784. doi:10.1016/j.bbamcr.2016.01.025
18. Gershenwald JE, Scolyer RA, Hess KR, et al. Melanoma staging: evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. *CA Cancer J Clin*. 2017;67(6):472-492. doi:10.3322/caac.21409
19. Bunnell AM, Nedrud SM, Fernandes RP. Classification and staging of melanoma in the head and neck. *Oral Maxillofac Surg Clin North Am*. 2022;34(2):221-234. doi:10.1016/j.coms.2021.12.001
20. Ward WH, Lamberton F, Goel N, Yu JQ, Farma JM. Clinical presentation and staging of melanoma. In: Ward WH, Farma JM, eds. *Cutaneous Melanoma: Etiology and Therapy*. Codon Publications; 2017.
21. Chandra Pal H, Marchiony Hunt K, Diamond A, Al. Elmets C, Afaq F. Phytochemicals for the management of melanoma. *Mini Rev Med Chem*. 2016;16(12):953-979. doi:10.2174/1389557516666160211120157
22. El Sharouni MA, van Diest PJ, Witkamp AJ, Sigurdsson V, van Gils CH. Subtyping cutaneous melanoma matters. *JNCI Cancer Spectr*. 2020;4(6):pkaa097. doi:10.1093/jncics/pkaa097
23. Davis LE, Shalin SC, Tackett AJ. Current state of melanoma diagnosis and treatment. *Cancer Biol Ther*. 2019;20(11):1366-1379. doi:10.1080/15384047.2019.1640032
24. Luke JJ, Schwartz GK. Chemotherapy in the management of advanced cutaneous malignant melanoma. *Clin Dermatol*. 2013;31(3):290-297. doi:10.1016/j.clindermatol.2012.08.016
25. Jiang G, Li RH, Sun C, Liu YQ, Zheng JN. Dacarbazine combined targeted therapy versus dacarbazine alone in patients with malignant melanoma: a meta-analysis. *PLoS One*. 2014;9(12):e111920. doi:10.1371/journal.pone.0111920
26. Pham JP, Joshua AM, da Silva IP, Dummer R, Goldinger SM. Chemotherapy in cutaneous melanoma: is there still a role? *Curr Oncol Rep*. 2023;25(6):609-621. doi:10.1007/s11912-023-01385-6
27. Agarwala SS, Kirkwood JM, Gore M, et al. Temozolomide for the treatment of brain metastases associated with metastatic melanoma: a phase II study. *J Clin Oncol*. 2004;22(11):2101-2107. doi:10.1200/jco.2004.11.044
28. Paul MJ, Summers Y, Calvert AH, et al. Effect of temozolomide on central nervous system relapse in patients with advanced melanoma. *Melanoma Res*. 2002;12(2):175-178. doi:10.1097/00008390-200204000-00011
29. Khattak M, Fisher R, Turajlic S, Larkin J. Targeted therapy and immunotherapy in advanced melanoma: an evolving paradigm. *Ther Adv Med Oncol*. 2013;5(2):105-118. doi:10.1177/1758834012466280
30. Milijašević B, Stefanović D, Lalić-Popović M, et al. Acute toxic effects of single dose dacarbazine: hematological and histological changes in an animal model. *Biotech Histochem*. 2014;89(8):583-590. doi:10.3109/10520295.2014.918653
31. Kampan NC, Madondo MT, McNally OM, Quinn M, Plebanski M. Paclitaxel and its evolving role in the management of ovarian cancer. *Biomed Res Int*. 2015;2015:413076. doi:10.1155/2015/413076
32. Dasari S, Tchounwou PB. Cisplatin in cancer therapy: molecular mechanisms of action. *Eur J Pharmacol*. 2014;740:364-378. doi:10.1016/j.ejphar.2014.07.025
33. Trunzer K, Pavlick AC, Schuchter L, Gonzalez R, McArthur GA, Hutson TE, et al. Pharmacodynamic effects and mechanisms of resistance to vemurafenib in patients with metastatic melanoma. *J Clin Oncol*. 2013;31(14):1767-1774. doi:10.1200/jco.2012.44.7888
34. Wesolowski JR, Rajdev P, Mukherji SK. Temozolomide (Temodar). *AJNR Am J Neuroradiol*. 2010;31(8):1383-1384. doi:10.3174/ajnr.A2170
35. Darnell EP, Mooradian MJ, Baruch EN, Yilmaz M, Reynolds KL. Immune-related adverse events (irAEs): diagnosis, management, and clinical pearls. *Curr Oncol Rep*. 2020;22(4):39. doi:10.1007/s11912-020-0897-9
36. Youssef G, Dietrich J. Ipilimumab: an investigational immunotherapy for glioblastoma. *Expert Opin Investig Drugs*. 2020;29(11):1187-1193. doi:10.1080/13543784.2020.1826436
37. Koppolu V, Rekha Vasigala VK. Checkpoint immunotherapy by nivolumab for treatment of metastatic melanoma. *J Cancer Res Ther*. 2018;14(6):1167-1175. doi:10.4103/jcrt.JCRT_1290_16
38. Spain L, Diem S, Larkin J. Management of toxicities of immune checkpoint inhibitors. *Cancer Treat Rev*. 2016;44:51-60. doi:10.1016/j.ctrv.2016.02.001
39. Sak K. Chemotherapy and dietary phytochemical agents. *Chemother Res Pract*. 2012;2012:1-11. doi:10.1155/2012/282570
40. Bukowski K, Kciuk M, Kontek R. Mechanisms of multidrug resistance in cancer chemotherapy. *Int J Mol Sci*. 2020;21(9):3233. doi:10.3390/ijms21093233
41. Kodama S, Podyma-inoue K, Uchihashi T, et al. Progression of melanoma is suppressed by targeting all transforming growth factor- β isoforms with an Fc chimeric receptor. *Oncol Rep*. 2021;46(3):197. doi:10.3892/or.2021.8148
42. Yuan Y, Jiang YC, Sun CK, Chen QM. Role of the tumor microenvironment in tumor progression and the clinical applications (review). *Oncol Rep*. 2016;35(5):2499-2515. doi:10.3892/or.2016.4660
43. Elias EG, Hasskamp JH, Sharma BK. Cytokines and growth factors expressed by human cutaneous melanoma. *Cancers*. 2010;2(2):794-808. doi:10.3390/cancers2020794
44. Wang X, Montoyo-Pujol YG, Bermudez S, et al. Serum cytokine profiles of melanoma patients and their association with tumor progression and metastasis. *J Oncol*. 2021;2021:1-9. doi:10.1155/2021/6610769

45. Hoejberg L, Bastholt L, Schmidt H. Interleukin-6 and melanoma. *Melanoma Res.* 2012;22(5):327-333. doi:10.1097/CMR.0b013e3283543d72
46. Chen W, Ten Dijke P. Immunoregulation by members of the TGF β superfamily. *Nat Rev Immunol.* 2016;16(12):723-740. doi:10.1038/nri.2016.112
47. Syed V. TGF- β signaling in cancer. *J Cell Biochem.* 2016;117(6):1279-1287. doi:10.1002/jcb.25496
48. Liu S, Chen S, Zeng J. TGF- β signaling: a complex role in tumorigenesis (Review). *Mol Med Rep.* 2017;17(1):699-704. doi:10.3892/mmr.2017.7970
49. Papageorgis P, Stylianopoulos T. Role of TGF β in regulation of the tumor microenvironment and drug delivery (review). *Int J Oncol.* 2015;46(3):933-943. doi:10.3892/ijo.2015.2816
50. Seoane J, Gomis RR. TGF- β family signaling in tumor suppression and cancer progression. *Cold Spring Harbor Perspect Biol.* 2017;9(12):a022277. doi:10.1101/cshperspect.a022277
51. Margadant C, Sonnenberg A. Integrin-TGF- β crosstalk in fibrosis, cancer and wound healing. *EMBO Rep.* 2010;11(2):97-105. doi:10.1038/embor.2009.276
52. Javelaud D, Alexaki VI, Mauviel A. Transforming growth factor- β in cutaneous melanoma. *Pigm Cell Melanoma Res.* 2008;21(2):123-132. doi:10.1111/j.1755-148X.2008.00450.x
53. Pak KH, Park KC, Cheong JH. VEGF-C induced by TGF- β 1 signaling in gastric cancer enhances tumor-induced lymphangiogenesis. *BMC Cancer.* 2019;19(1):799. doi:10.1186/s12885-019-5972-y
54. Viillard C, Larrivée B. Tumor angiogenesis and vascular normalization: alternative therapeutic targets. *Angiogenesis.* 2017;20(4):409-426. doi:10.1007/s10456-017-9562-9
55. Ye X, Gaucher JF, Vidal M, Broussy S. A structural overview of vascular endothelial growth factors pharmacological ligands: from macromolecules to designed peptidomimetics. *Molecules.* 2021;26(22):6759. doi:10.3390/molecules26226759
56. Chen JC, Chang YW, Hong CC, Yu YH, Su JL. The role of the VEGF-C/VEGFRs axis in tumor progression and therapy. *Int J Mol Sci.* 2012;14(1):88-107. doi:10.3390/ijms14010088
57. Apte RS, Chen DS, Ferrara N. VEGF in signaling and disease: beyond discovery and development. *Cell.* 2019;176(6):1248-1264. doi:10.1016/j.cell.2019.01.021
58. Koch S, Claesson-Welsh L. Signal transduction by vascular endothelial growth factor receptors. *Cold Spring Harbor Perspect Med.* 2012;2(7):a006502. doi:10.1101/cshperspect.a006502
59. Pastushenko I, Conejero C, Carapeto FJ. La linfangiogenesis. Sus implicaciones en el diagnóstico, tratamiento y pronóstico del melanoma. *Actas Dermosifiliogr.* 2015;106(1):7-16. doi:10.1016/j.ad.2014.02.013
60. Wang M, Xu Y, Wen GZ, Wang Q, Yuan SM. Rapamycin suppresses angiogenesis and lymphangiogenesis in melanoma by downregulating VEGF-A/VEGFR-2 and VEGF-C/VEGFR-3 expression. *Oncotargets Ther.* 2019;12:4643-4654. doi:10.2147/ott.S205160
61. Wissmann C, Detmar M. Pathways targeting tumor lymphangiogenesis. *Clin Cancer Res.* 2006;12(23):6865-6868. doi:10.1158/1078-0432.Ccr-06-1800
62. Streit M, Detmar M. Angiogenesis, lymphangiogenesis, and melanoma metastasis. *Oncogene.* 2003;22(20):3172-3179. doi:10.1038/sj.onc.1206457
63. Raica M, Jitariu AA, Cimpean AM. Lymphangiogenesis and anti-lymphangiogenesis in cutaneous melanoma. *Anticancer Res.* 2016;36(9):4427-4436. doi:10.21873/anticancer.10986
64. Liu B, Ma J, Wang X, et al. Lymphangiogenesis and its relationship with lymphatic metastasis and prognosis in malignant melanoma. *Anat Rec.* 2008;291(10):1227-1235. doi:10.1002/ar.20736
65. Cianfarani F, Mastroeni S, Odorisio T, et al. Expression of vascular endothelial growth factor-C in primary cutaneous melanoma predicts sentinel lymph node positivity. *J Cutan Pathol.* 2012;39(9):826-834. doi:10.1111/j.1600-0560.2012.01955.x
66. Cheresch DA. Monographs editor. *Genes Cancer.* 2011;2:1071. doi:10.1177/1947601911431886
67. Gillot L, Baudin L, Rouaud L, Kridelka F, Noël A. The pre-metastatic niche in lymph nodes: formation and characteristics. *Cell Mol Life Sci.* 2021;78(16):5987-6002. doi:10.1007/s00018-021-03873-z
68. Chatterjee S, Behnam Azad B, Nimmagadda S. The intricate role of CXCR4 in cancer. *Adv Cancer Res.* 2014;124:31-82. doi:10.1016/b978-0-12-411638-2.00002-1
69. Jacquetot N, Duong CPM, Belz GT, Zitvogel L. Targeting chemokines and chemokine receptors in melanoma and other cancers. *Front Immunol.* 2018;9:2480. doi:10.3389/fimmu.2018.02480
70. Hughes CE, Nibbs RJB. A guide to chemokines and their receptors. *FEBS J.* 2018;285(16):2944-2971. doi:10.1111/febs.14466
71. Wang Y, Xie Y, Oupický D. Potential of CXCR4/CXCL12 chemokine axis in cancer drug delivery. *Curr Pharmacol Rep.* 2016;2(1):1-10. doi:10.1007/s40495-015-0044-8
72. Mitchell B, Mahalingam M. The CXCR4/CXCL12 axis in cutaneous malignancies with an emphasis on melanoma. *Histol Histopathol.* 2014;29(12):1539-1546. doi:10.14670/hh-29.1539
73. Kim J, Mori T, Chen SL, et al. Chemokine receptor CXCR4 expression in patients with melanoma and colorectal cancer liver metastases and the association with disease outcome. *Ann Surg.* 2006;244(1):113-120. doi:10.1097/01.sla.0000217690.65909.9c
74. André ND, Silva VAO, Ariza CB, Watanabe MAE, De Lucca FL. In vivo knockdown of CXCR4 using jetPEI/CXCR4 shRNA nanoparticles inhibits the pulmonary metastatic potential of B16-F10 melanoma cells. *Mol Med Rep.* 2015;12(6):8320-8326. doi:10.3892/mmr.2015.4487
75. O'Boyle G, Swidenbank I, Marshall H, et al. Inhibition of CXCR4-CXCL12 chemotaxis in melanoma by AMD11070. *Br J Cancer.* 2013;108(8):1634-1640. doi:10.1038/bjc.2013.124
76. Karaman S, Detmar M. Mechanisms of lymphatic metastasis. *J Clin Invest.* 2014;124(3):922-928. doi:10.1172/jci71606
77. Teicher BA, Fricker SP. CXCL12 (SDF-1)/CXCR4 pathway in cancer. *Clin Cancer Res.* 2010;16(11):2927-2931. doi:10.1158/1078-0432.Ccr-09-2329
78. Stone M, Hayward J, Huang C, E. Huma Z, Sanchez J. Mechanisms of regulation of the chemokine-receptor network. *Int J Mol Sci.* 2017;18(2):342. doi:10.3390/ijms18020342
79. Zhao R, Liu J, Li Z, Zhang W, Wang F, Zhang B. Recent advances in CXCL12/CXCR4 antagonists and nano-based drug delivery systems for cancer therapy. *Pharmaceutics.* 2022;14(8):1541. doi:10.3390/pharmaceutics14081541
80. Bianchi ME, Mezzapelle R. The chemokine receptor CXCR4 in cell proliferation and tissue regeneration. *Front Immunol.* 2020;11:2109. doi:10.3389/fimmu.2020.02109
81. Desgrosellier JS, Cheresch DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer.* 2010;10(1):9-22. doi:10.1038/nrc2748
82. Cooper J, Giancotti FG. Integrin signaling in cancer: mechanotransduction, stemness, epithelial plasticity, and therapeutic resistance. *Cancer Cell.* 2019;35(3):347-367. doi:10.1016/j.ccell.2019.01.007
83. Bendas G, Borsig L. Cancer cell adhesion and metastasis: selectins, integrins, and the inhibitory potential of heparins. *Int J Cell Biol.* 2012;2012:1-10. doi:10.1155/2012/676731
84. Mousson A, Legrand M, Steffan T, et al. Inhibiting FAK-paxillin interaction reduces migration and invadopodia-mediated matrix degradation in metastatic melanoma cells. *Cancers.* 2021;13(8):1871. doi:10.3390/cancers13081871
85. Murphy JM, Rodriguez YAR, Jeong K, Ahn EYE, Lim STS. Targeting focal adhesion kinase in cancer cells and the tumor

- microenvironment. *Exp Mol Med.* 2020;52(6):877-886. doi:10.1038/s12276-020-0447-4
86. Golubovskaya VM, Finch R, Kweh F, et al. p53 regulates FAK expression in human tumor cells. *Mol Carcinog.* 2008;47(5):373-382. doi:10.1002/mc.20395
 87. Tai YL, Chen LC, Shen TL. Emerging roles of focal adhesion kinase in cancer. *BioMed Res Int.* 2015;2015:1-13. doi:10.1155/2015/690690
 88. Paoli P, Giannoni E, Chiarugi P. Anoikis molecular pathways and its role in cancer progression. *Biochim Biophys Acta Mol Cell Res.* 2013;1833(12):3481-3498. doi:10.1016/j.bbamcr.2013.06.026
 89. Horowitz JC, Rogers DS, Sharma V, et al. Combinatorial activation of FAK and AKT by transforming growth factor- β 1 confers an anoikis-resistant phenotype to myofibroblasts. *Cell Signal.* 2007;19(4):761-771. doi:10.1016/j.cellsig.2006.10.001
 90. Hlophe YN, Joubert AM. Vascular endothelial growth factor-C in activating vascular endothelial growth factor receptor-3 and chemokine receptor-4 in melanoma adhesion. *J Cell Mol Med.* 2022;26(23):5743-5754. doi:10.1111/jcmm.17571
 91. López-Colomé AM, Lee-Rivera I, Benavides-Hidalgo R, López E. Paxillin: a crossroad in pathological cell migration. *J Hematol Oncol.* 2017;10(1):50. doi:10.1186/s13045-017-0418-y
 92. Liu W, Huang X, Luo W, Liu X, Chen W. The role of paxillin aberrant expression in cancer and its potential as a target for cancer therapy. *Int J Mol Sci.* 2023;24(9):8245. doi:10.3390/ijms24098245
 93. Velasco-Velázquez MA, Salinas-Jazmín N, Mendoza-Patiño N, Mandoki JJ. Reduced paxillin expression contributes to the antimetastatic effect of 4-hydroxycoumarin on B16-F10 melanoma cells. *Cancer Cell Int.* 2008;8:8. doi:10.1186/1475-2867-8-8
 94. Park JH, Shin YJ, Riew TR, Lee MY. The indolinone MAZ51 induces cell rounding and G2/M cell cycle arrest in glioma cells without the inhibition of VEGFR-3 phosphorylation: involvement of the RhoA and Akt/GSK3 β signaling pathways. *PLoS One.* 2014;9(9):e109055. doi:10.1371/journal.pone.0109055
 95. Lee JY, Hong SH, Shin M, Heo HR, Jang IH. Blockade of FLT4 suppresses metastasis of melanoma cells by impaired lymphatic vessels. *Biochem Biophys Res Commun.* 2016;478(2):733-738. doi:10.1016/j.bbrc.2016.08.017
 96. Špirić Z, Eri Ž, Erić M. Significance of vascular endothelial growth factor (VEGF)-C and VEGF-D in the progression of cutaneous melanoma. *Int J Surg Pathol.* 2015;23(8):629-637. doi:10.1177/1066896915583694
 97. Sagbo IJ, Otang-Mbeng W. Plants used for the traditional management of cancer in the eastern cape province of South Africa: a review of ethnobotanical surveys, ethnopharmacological studies and active phytochemicals. *Molecules.* 2021;26(15):4639. doi:10.3390/molecules26154639
 98. Sofowora A, Ogunbodede E, Onayade A. The role and place of medicinal plants in the strategies for disease prevention. *Afr J Tradit Complement Altern Med.* 2013;10(5):210-229. doi:10.4314/ajtcam.v10i5.2
 99. Choudhari AS, Mandave PC, Deshpande M, Ranjekar P, Prakash O. Phytochemicals in cancer treatment: from preclinical studies to clinical practice. *Front Pharmacol.* 2020;10:1614. doi:10.3389/fphar.2019.01614
 100. Sever R, Brugge JS. Signal transduction in cancer. *Cold Spring Harbor Perspect Med.* 2015;5(4):a006098. doi:10.1101/cshperspect.a006098
 101. Ayaz M, Nawaz A, Ahmad S, et al. Underlying anticancer mechanisms and synergistic combinations of phytochemicals with cancer chemotherapeutics: potential benefits and risks. *J Food Quality.* 2022;2022:1-15. doi:10.1155/2022/1189034
 102. Pal HC, Sharma S, Strickland LR, et al. Fisetin inhibits human melanoma cell invasion through promotion of mesenchymal to epithelial transition and by targeting MAPK and NF κ B signaling pathways. *PLoS One.* 2014;9(1):e86338. doi:10.1371/journal.pone.0086338
 103. Aronchik I, Kundu A, Quirit JG, Firestone GL. The antiproliferative response of indole-3-carbinol in human melanoma cells is triggered by an interaction with NEDD4-1 and disruption of wild-type PTEN degradation. *Mol Cancer Res.* 2014;12(11):1621-1634. doi:10.1158/1541-7786.Mcr-14-0018
 104. Ravindran Menon D, Li Y, Yamauchi T, et al. EGCG inhibits tumor growth in melanoma by targeting JAK-STAT signaling and its downstream PD-L1/PD-L2-PD1 axis in tumors and enhancing cytotoxic T-cell responses. *Pharmaceuticals.* 2021;14(11):1081. doi:10.3390/ph14111081
 105. Ghosh R, Nadiminty N, Fitzpatrick JE, Alworth WL, Slaga TJ, Kumar AP. Eugenol causes melanoma growth suppression through inhibition of E2F1 transcriptional activity. *J Biol Chem.* 2005;280(7):5812-5819. doi:10.1074/jbc.M411429200
 106. Pourhanifeh MH, Abbaszadeh-Goudarzi K, Goodarzi M, et al. Resveratrol: a new potential therapeutic agent for melanoma? *Curr Med Chem.* 2021;28(4):687-711. doi:10.2174/0929867326666191212101225
 107. Vaid M, Singh T, Prasad R, Katiyar SK. Silymarin inhibits melanoma cell growth both in vitro and in vivo by targeting cell cycle regulators, angiogenic biomarkers and induction of apoptosis. *Mol Carcinog.* 2015;54(11):1328-1339. doi:10.1002/mc.22208
 108. Bae J, Kumazoe M, Murata K, Fujimura Y, Tachibana H. Procyanidin C1 inhibits melanoma cell growth by activating 67-kDa laminin receptor signaling. *Mol Nutr Food Res.* 2020;64(7):e1900986. doi:10.1002/mnfr.201900986
 109. Rocchetti MT, Bellanti F, Zadorozhna M, Fiocco D, Mangieri D. Multi-faceted role of luteolin in cancer metastasis: EMT, angiogenesis, ECM degradation and apoptosis. *Int J Mol Sci.* 2023;24(10):8824. doi:10.3390/ijms24108824
 110. Yao X, Jiang W, Yu D, Yan Z. Luteolin inhibits proliferation and induces apoptosis of human melanoma cells in vivo and in vitro by suppressing MMP-2 and MMP-9 through the PI3K/AKT pathway. *Food Funct.* 2019;10(2):703-712. doi:10.1039/c8fo02013b
 111. Russo M, Spagnuolo C, Tedesco I, Russo GL. Phytochemicals in cancer prevention and therapy: truth or dare? *Toxins.* 2010;2(4):517-551. doi:10.3390/toxins2040517
 112. Rizeq B, Gupta I, Ilesanmi J, AlSafran M, Rahman MM, Ouhtit A. The power of phytochemicals combination in cancer chemoprevention. *J Cancer.* 2020;11(15):4521-4533. doi:10.7150/jca.34374
 113. Rodriguez S, Skeet K, Mehmetoglu-Gurbuz T, et al. Phytochemicals as an alternative or integrative option, in conjunction with conventional treatments for hepatocellular carcinoma. *Cancers.* 2021;13(22):5753. doi:10.3390/cancers13225753
 114. Ahmad B, Rehman MU, Amin I, et al. A review on pharmacological properties of zingerone (4-(4-hydroxy-3-methoxyphenyl)-2-butanone). *Sci World J.* 2015;2015:1-6. doi:10.1155/2015/816364
 115. Amer R. The possible protective role of zingerone on ethanol induced enterotoxicity of jejunum in adult albino rats: light and scanning electron microscopic study. *J Microsc Ultrastruct.* 2020;8(2):69-74. doi:10.4103/jmau.jmau_55_19
 116. Choi JS, Ryu J, Bae WY, et al. Zingerone suppresses tumor development through decreasing cyclin D1 expression and inducing mitotic arrest. *Int J Mol Sci.* 2018;19(9):2832. doi:10.3390/ijms19092832
 117. Qian S, Fang H, Zheng L, Liu M. Zingerone suppresses cell proliferation via inducing cellular apoptosis and inhibition of the PI3K/AKT/mTOR signaling pathway in human prostate cancer PC-3 cells. *J Biochem Mol Toxicol.* 2021;35(1):e22611. doi:10.1002/jbt.22611
 118. Lee J, Oh SW, Shin SW, Lee KW, Cho JY, Lee J. Zingerone protects keratinocyte stem cells from UVB-induced damage. *Chem-Biol Interact.* 2018;279:27-33. doi:10.1016/j.cbi.2017.11.004

119. Chaudhuri RK, Meyer T, Premi S, Brash D. Acetyl zingerone: an efficacious multifunctional ingredient for continued protection against ongoing DNA damage in melanocytes after sun exposure ends. *Int J Cosmet Sci.* 2020;42(1):36-45. doi:10.1111/ics.12582
120. Meyer TA, Swindell WR, Chaudhuri RK. Acetyl zingerone: a photostable multifunctional skincare ingredient that combats features of intrinsic and extrinsic skin aging. *Antioxidants.* 2023;12(6):1168. doi:10.3390/antiox12061168
121. Srivastava J, Young MM, Yadav VK, et al. The role of acetyl zingerone and its derivatives in inhibiting UV-induced, incident, and delayed cyclobutane pyrimidine dimers. *Antioxidants.* 2023;12(2):278. doi:10.3390/antiox12020278
122. Bhaskar Chandra Sahoo SS, Nayak S, Kar B. Pharmacological activity and biochemical interaction of Zingerone: a flavour additive in spice food. *Plant Sci Today.* 2022;9(1):81-88.
123. Su P, Veeraraghavan VP, Krishna Mohan S, Lu W. A ginger derivative, zingerone—a phenolic compound—induces ROS-mediated apoptosis in colon cancer cells (HCT-116). *J Biochem Mol Toxicol.* 2019;33(12):e22403. doi:10.1002/jbt.22403
124. Sagwal SK, Bekeschus S. ROS pleiotropy in melanoma and local therapy with physical modalities. *Oxid Med Cell Longevity.* 2021;2021:1-21. doi:10.1155/2021/6816214
125. Wormington AM, De Maria M, Kurita HG, Bisesi JH Jr., Denslow ND, Martyniuk CJ. Antineoplastic agents: environmental prevalence and adverse outcomes in aquatic organisms. *Environ Toxicol Chem.* 2020;39(5):967-985. doi:10.1002/etc.4687
126. Li D, Chen H, Liu H, et al. Anticancer drugs in the aquatic ecosystem: environmental occurrence, ecotoxicological effect and risk assessment. *Environ Int.* 2021;153:106543. doi:10.1016/j.envint.2021.106543
127. Ribeiro F, Costa-Lotufo L, Loureiro S, Pavlaki MD. Environmental hazard of anticancer drugs: state of the art and future perspective for marine organisms. *Environ Toxicol Chem.* 2022;41(8):1793-1807. doi:10.1002/etc.5397
128. Yadav A, Rene ER, Mandal MK, Dubey KK. Threat and sustainable technological solution for antineoplastic drugs pollution: review on a persisting global issue. *Chemosphere.* 2021;263:128285. doi:10.1016/j.chemosphere.2020.128285
129. Heath E, Filipič M, Kosjek T, Isidori M. Fate and effects of the residues of anticancer drugs in the environment. *Environ Sci Pollut Res.* 2016;23(15):14687-14691. doi:10.1007/s11356-016-7069-3
130. Li D, Sun W, Chen H, et al. Cyclophosphamide affects eye development and locomotion in zebrafish (*Danio rerio*). *Sci Total Environ.* 2022;805:150460. doi:10.1016/j.scitotenv.2021.150460
131. Novak M, Žegura B, Modic B, Heath E, Filipič M. Cytotoxicity and genotoxicity of anticancer drug residues and their mixtures in experimental model with zebrafish liver cells. *Sci Total Environ.* 2017;601-602:293-300. doi:10.1016/j.scitotenv.2017.05.115
132. Jureczko M, Kalka J. Cytostatic pharmaceuticals as water contaminants. *Eur J Pharmacol.* 2020;866:172816. doi:10.1016/j.ejphar.2019.172816
133. Viegas S, Ladeira C, Costa-Veiga A, Perelman J, Gajski G. Forgotten public health impacts of cancer—an overview. *Arch Ind Hyg Toxicol.* 2017;68(4):287-297. doi:10.1515/aiht-2017-68-3005
134. Baniyasi S, Alehashem M, Yunesian M, Rastkari N. Biological monitoring of healthcare workers exposed to antineoplastic drugs: urinary assessment of cyclophosphamide and ifosfamide. *Iran J Pharm Res.* 2018;17(4):1458-1464.
135. Elshaer N. Adverse health effects among nurses and clinical pharmacists handling antineoplastic drugs: adherence to exposure control methods. *J Egypt Public Health Assoc.* 2017;92(3):144-155.
136. The World Bank. *Blue Economy.* The World Bank; 2022. <https://www.worldbank.org/en/topic/oceans-fisheries-and-coastal-economies>
137. SADC-EU EPA. *South African Fisheries and the SADC-EU Economic Partnership Agreement.* SADC-EU EPA; 2017. <https://sadc-epa-outreach.com/images/files/sadc-eu-epa-fisheries-july-2017.pdf>
138. Dhupal M, Chowdhury D. Phytochemical-based nanomedicine for advanced cancer theranostics: perspectives on clinical trials to clinical use. *Int J Nanomedicine.* 2020;15:9125-9157. doi:10.2147/ijn.S259628
139. Fasinu PS, Rapp GK. Herbal interaction with chemotherapeutic drugs—a focus on clinically significant findings. *Front Oncol.* 2019;9:1356. doi:10.3389/fonc.2019.01356
140. Yeung KS, Gubili J, Mao JJ. Herb-drug interactions in cancer care. *Oncology.* 2018;32(10):516-520.

How to cite this article: Letsoalo K, Nortje E, Patrick S, Nyakudya T, Hlophe Y. Decoding the synergistic potential of MAZ-51 and zingerone as therapy for melanoma treatment in alignment with sustainable development goals. *Cell Biochem Funct.* 2024;42:e3950. doi:10.1002/cbf.3950