

Supplementary Information for

Multiclonal human origin and global expansion of an endemic bacterial pathogen of livestock

Gonzalo Yebra^{1*}, Joshua D. Harling-Lee^{1*}, Samantha Lycett¹, Frank M. Aarestrup², Gunhild Larsen², Lina Cavaco³, Keun Seok Seo⁴, Sam Abraham⁵, Jacqueline M. Norris⁶, Tracy Schmidt⁷, Marthie M. Ehlers^{7,8}, Daniel O. Sordelli⁹, Fernanda R. Buzzola⁹, Wondwossen A. Gebreyes^{10,11}, Juliano L. Gonçalves¹², Marcos V. dos Santos¹², Zunita Zakaria¹³, Vera L. M. Rall¹⁴, Orla M. Keane¹⁵, Dagmara A. Niedziela¹⁵, Gavin K. Paterson^{1,16}, Mark A. Holmes¹⁷, Tom C. Freeman^{1,18}, and J. Ross Fitzgerald^{1 †}

(1) The Roslin Institute, University of Edinburgh, Edinburgh, UK; (2) The National Food Institute, Technical University of Denmark, Lyngby, Denmark; (3) Statens Serum Institute, Copenhagen, Denmark; (4) Department of Basic Sciences, College of Veterinary Medicine, Mississippi State University, Starkville, MS, United States; (5) Antimicrobial Resistance and Infectious Diseases Laboratory, College of Science, Health, Engineering and Education, Murdoch University, Murdoch, WA, Australia; (6) Sydney School of Veterinary Science, University of Sydney, Sydney, Australia; (7) Department of Medical Microbiology, University of Pretoria, Pretoria, South Africa; (8) Department of Medical Microbiology, Tshwane Academic Division, National Health Laboratory Service, Pretoria, South Africa; (9) Instituto de Investigaciones en Microbiología y Parasitología Médica, University of Buenos Aires-CONICET, Buenos Aires, Argentina; (10) Molecular Epidemiology, College of Veterinary Medicine, the Ohio State University, Columbus, USA; (11) Department of Large Animal Clinical Sciences, College of Veterinary Medicine, Michigan State University, East Lansing, MI, USA; (12) Department of Nutricion and Animal Production, School of Veterinary Medicine and Animal Sciences, University of São Paulo, Pirassununga, SP, Brazil; (13) Institute of Bioscience, Universiti Putra Malaysia, Serdang, Malaysia; (14) Department of Chemical and Biological Sciences, Institute of Biosciences, São Paulo State University, Botucatu-SP, Brazil, (15) Animal & Bioscience Department, Teagasc, Grange, Dunsany, Co. Meath, Ireland; (16) R(D)SVS, University of Edinburgh, Edinburgh, UK; (17) Department of Veterinary Medicine, University of Cambridge, Cambridge, UK; (18) Janssen Immunology, Spring House, PA, USA.

* These authors contributed equally

† Corresponding author: J. Ross Fitzgerald (ross.fitzgerald@roslin.ed.ac.uk)

This PDF file includes:

Supplementary Materials

Figures S2 to S7

Tables S1 to S6

Supplementary Materials

Accessory Genome & Geographical Analysis

To identify genes significantly enriched in specific geographic locations, we performed adjusted Fisher's tests within each CC dataset. Our primary aim was to identify any genes enriched in the same location across multiple CCs, as this would provide evidence for genes inhabiting a geographic niche. Were this true, we would expect subsequent acquisition of such genes by a foreign CC upon migration into that niche. However, we found just 36 genes positively associated (Bonferroni corrected $p < 0.05$) with the same location in two of the seven CCs, and none in three or more CCs (**SI Dataset 5**). Of those 36, 15 are positively associated with Norwegian CC130 and CC133 isolates. We also identify just 19 genes negatively associated (Bonferroni corrected $p < 0.05$) with a single location; 11 of these are negatively associated with Norwegian CC130 and CC133 isolates (**SI Dataset 5**).

Supplementary Figures

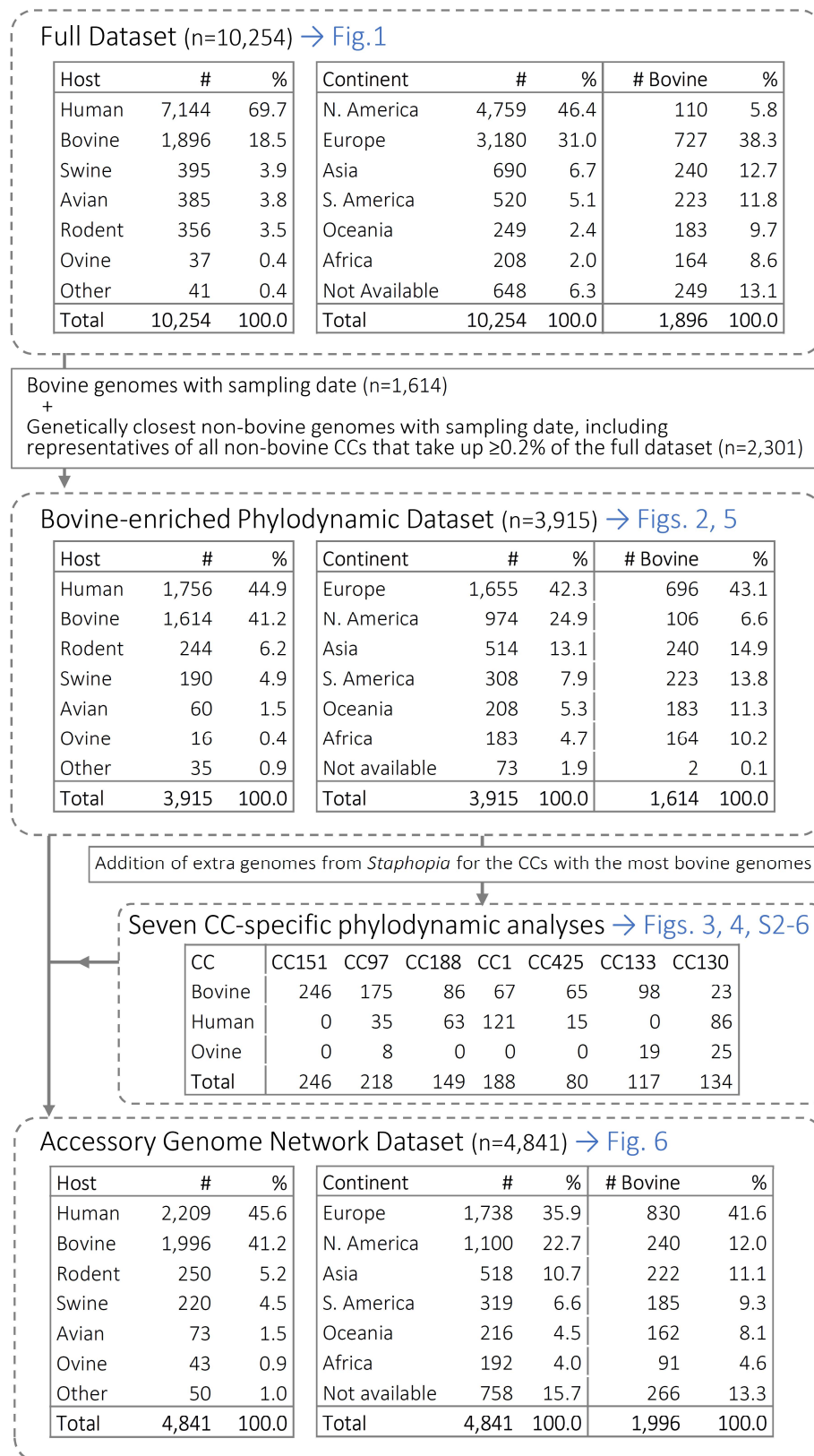


Fig. S1: Schematic description of the datasets used in this study and the relationships between them. Summary tables for each dataset in terms of host and location (continent) is included, as well as in which Figures each dataset is used.

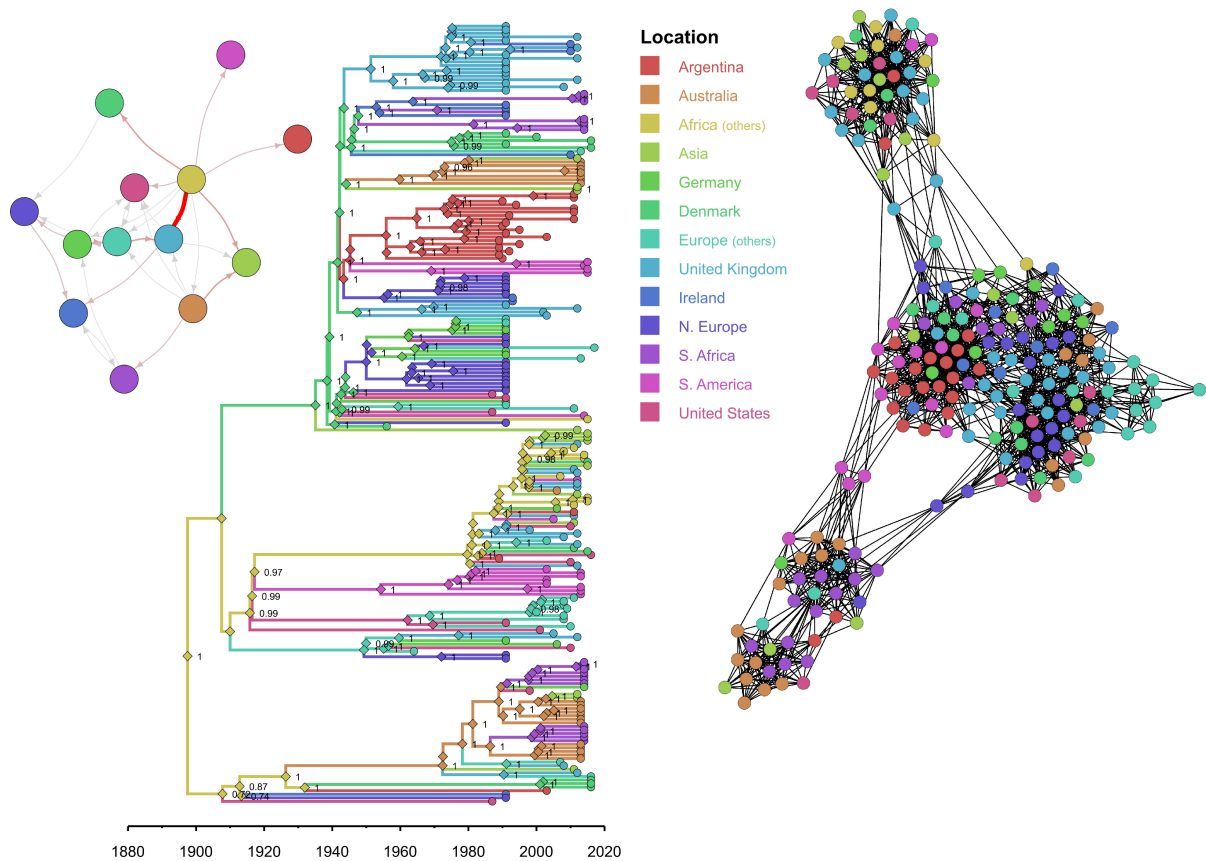


Fig. S2: Phylogeographic analysis of the multi-host-associated *S. aureus* CC97 based on core and accessory genome. Bayesian time-stamped tree from a core genome alignment (1,936,889bp, of which were 24,458 variable sites) of CC97 genome sequences, with branches coloured according to the reconstructed location in the discrete trait analysis (left); and network of accessory genome of the same sequences and colours (right, based on 1,221 accessory genes defined as genes in more than 1 genome, and not in all genomes). Inset next to the tree: graphic summary of migrations between countries, in which the thickness and colour (grey->red) of arrows is proportional to the number of migration events inferred.

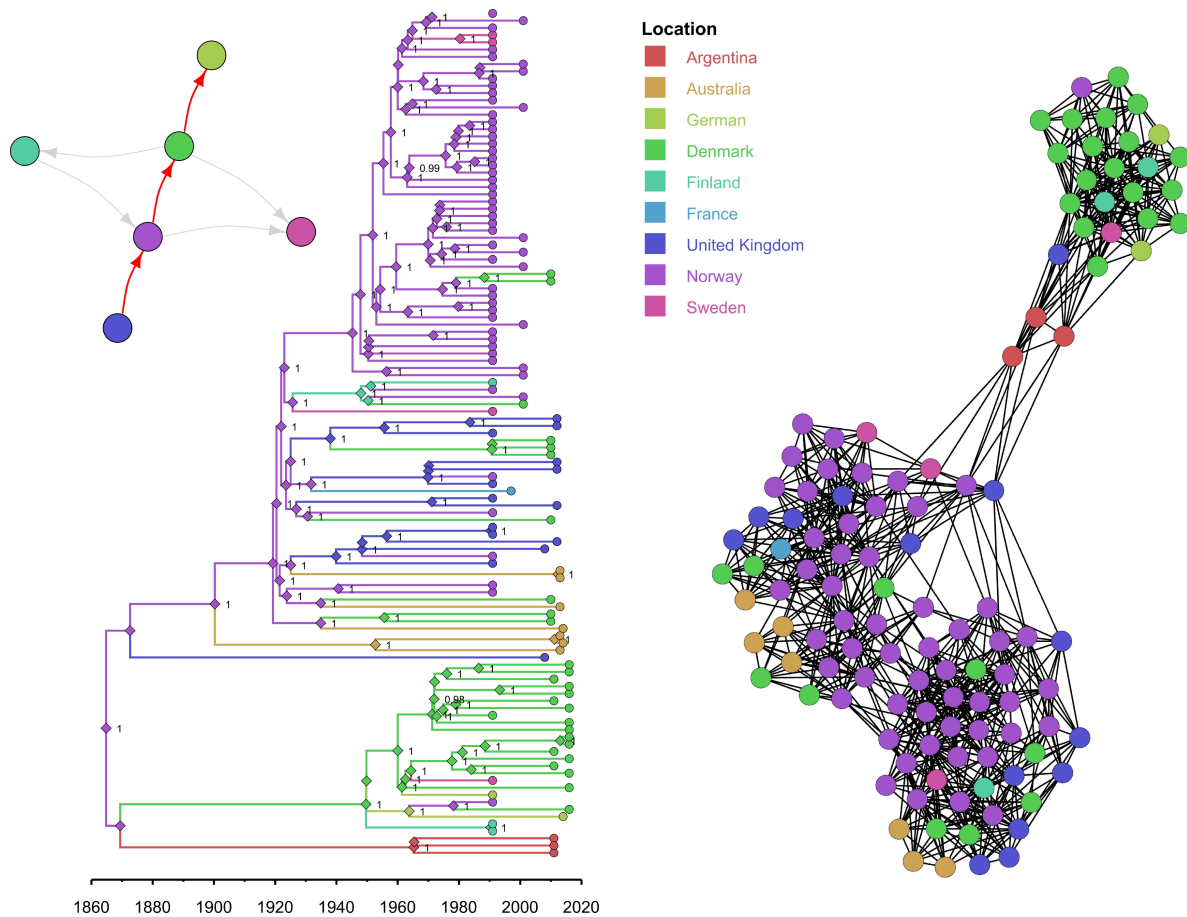


Fig. S3: Phylogeographic analysis of the multi-host-associated *S. aureus* CC133 based on core and accessory genome. Bayesian time-stamped tree from a core genome alignment (2,363,270bp, of which 14,435bp were variable sites) of CC133 genome sequences, with branches coloured according to the reconstructed location in the discrete trait analysis (left); and network of accessory genome of the same sequences and colours (right, based on 604 accessory genes defined as genes in more than 1 genome, and not in all genomes). Inset next to the tree: graphic summary of migrations between countries, in which the thickness and colour (grey→red) of arrows is proportional to the number of migration events inferred.

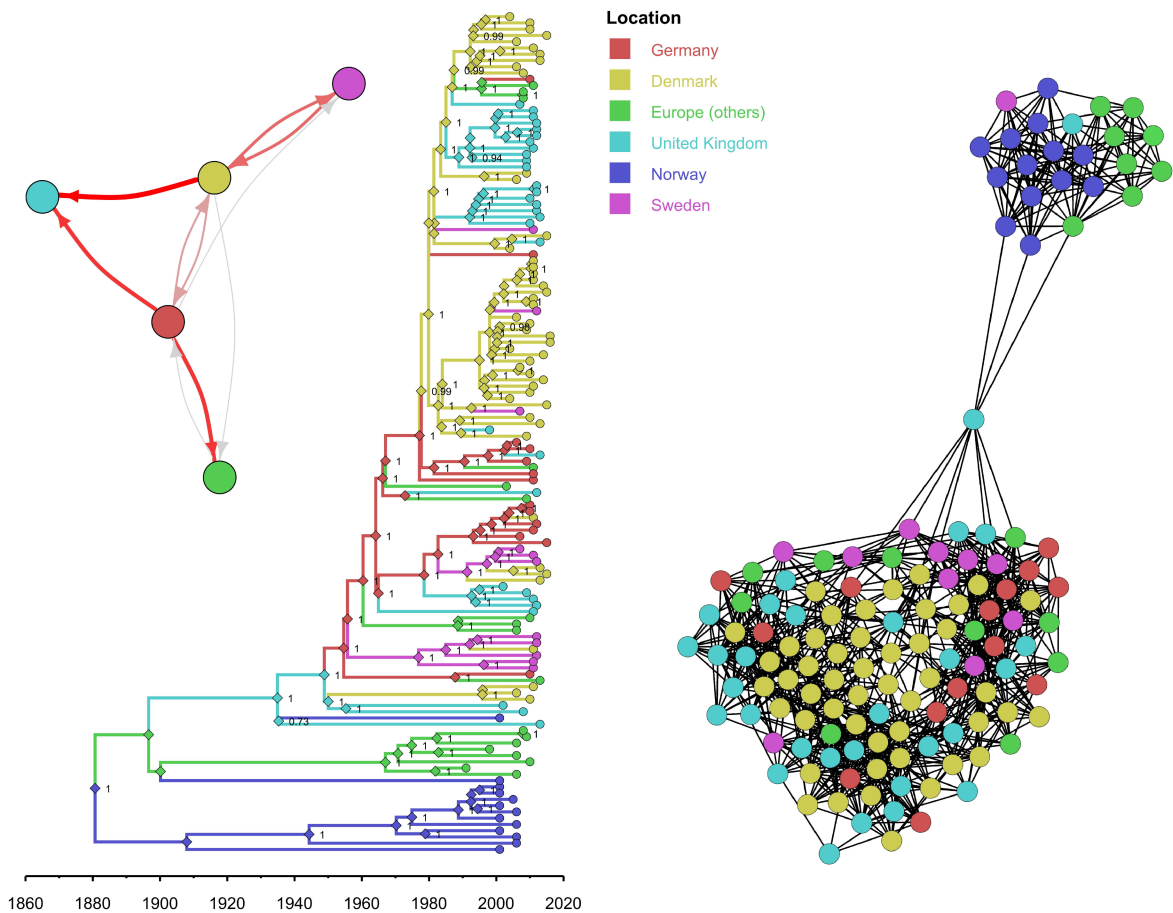


Fig. S4: Phylogeographic analysis of the multi-host-associated *S. aureus* CC130 based on core and accessory genome. Bayesian time-stamped tree from a core genome alignment (2,368,277bp, of which 15,123bp were variable sites) of CC130 genome sequences, with branches coloured according to the reconstructed location in the discrete trait analysis (left); and network or accessory genome of the same sequences and colours (right, based on 1,042 accessory genes defined as genes in more than 1 genome, and not in all genomes). Inset next to the tree: graphic summary of migrations between countries, in which the thickness and colour (grey->red) of arrows is proportional to the number of migration events inferred.

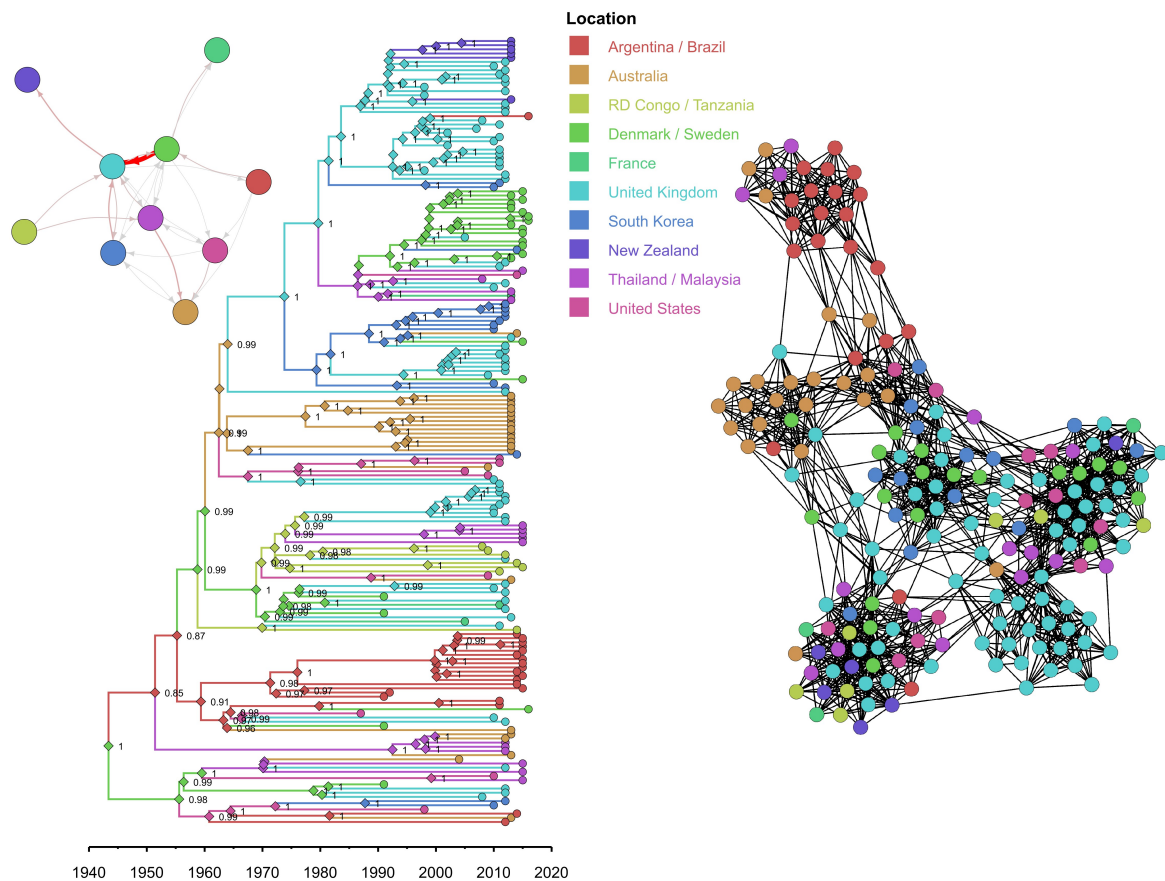


Fig. S5: Phylogeographic analysis of the multi-host-associated *S. aureus* CC1 based on core and accessory genome. Bayesian time-stamped tree from a core genome alignment (1,930,409bp, of which 18,984bp were variable sites) of CC1 genome sequences, with branches coloured according to the reconstructed location in the discrete trait analysis (left); and network of accessory genome of the same sequences and colours (right, based on 1,186 accessory genes defined as genes in more than 1 genome, and not in all genomes). Inset next to the tree: graphic summary of migrations between countries, in which the thickness and colour (grey→red) of arrows is proportional to the number of migration events inferred.

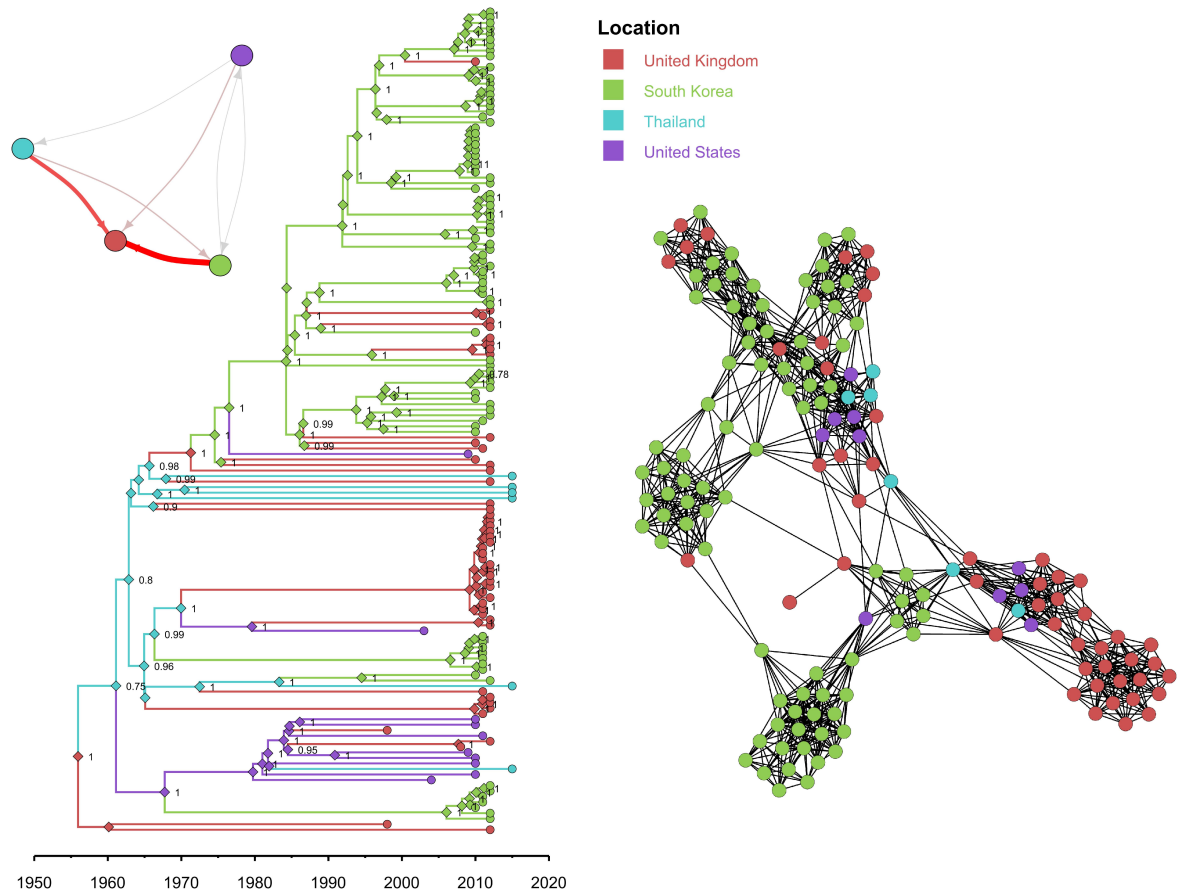


Fig. S6: Phylogeographic analysis of the multi-host-associated *S. aureus* CC188 based on core and accessory genome. Bayesian time-stamped tree from a core genome alignment (2,451,373bp, of which 7,378bp were variable sites) of CC188 genome sequences, with branches coloured according to the reconstructed location in the discrete trait analysis (left); and network of accessory genome of the same sequences and colours (right, based on 717 accessory genes defined as genes in more than 1 genome, and not in all genomes). Inset next to the tree: graphic summary of migrations between countries, in which the thickness and colour (grey→red) of arrows is proportional to the number of migration events inferred.

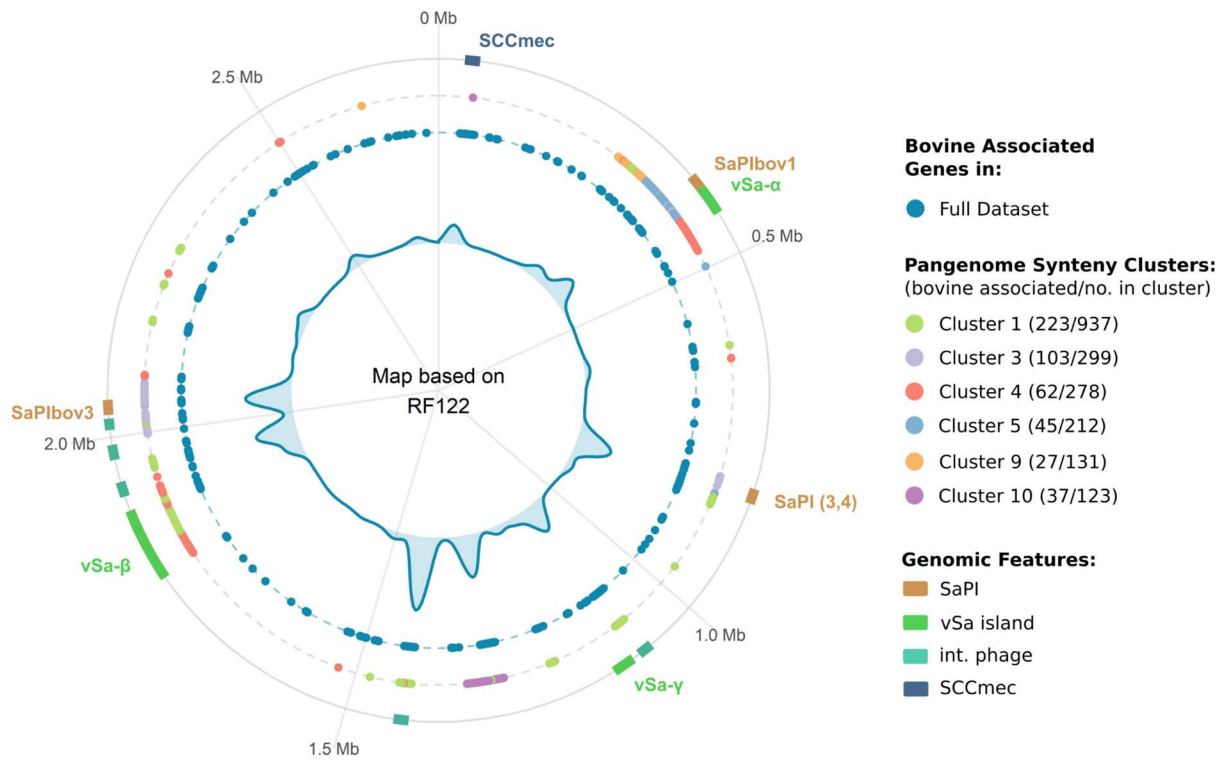


Fig. S7. Pangenome synteny clusters significantly enriched in bovine *S. aureus* associated genes, mapped to the reference genome RF122. Central density plot displays distribution of bovine-associated genes. Circle dot plots represent the location of bovine associated genes (inner), pangenome synteny clusters (middle), and known genomic features of interest (outer).

Supplementary Tables

Table S1. Distribution of inferred host changes in the global bovine-enriched *S. aureus* phylogeny from the SIMMAP analyses.

State change	Median number of changes (95%HPD)
Others -> Human	277.5 (251-313)
Human -> Others	188.5 (159-208)
Human -> Bovine	182 (160-202)
Bovine -> Others	131.5 (114-149)
Others -> Bovine	114 (95-133)
Bovine -> Human	63.5 (46-77)

Table S2. Distribution of inferred location changes (i.e. migrations) in the global bovine-enriched *S. aureus* phylogeny from the SIMMAP analyses. The table shows only those with median ≥ 10 changes for simplicity.

State change	Median number of changes (95%HPD)
N America → SS Africa	41 (30-52)
Ireland → Switzerland	32.5 (23-44)
N America → Ireland	31 (22-38)
UK → Ireland	28 (20-36)
N America → Switzerland	27 (18-36)
Ireland → N America	26.5 (18-35)
Switzerland → N America	26 (14-34)
Australia → SE Asia	24 (13-32)
Denmark → Sweden	19 (11-30)
S America → Sweden	15 (8-25)
Germany → N America	13 (8-19)
Denmark → Norway	13 (6-19)
UK → Denmark	13 (9-18)
Germany → Switzerland	12 (7-18)
Germany → Denmark	12 (6-17)
N America → S Europe	11 (6-19)
Sweden → Switzerland	11 (4-18)
Sweden → S Europe	11 (6-17)
Switzerland → Ireland	10 (5-17)
N America → Germany	10 (5-17)
Norway → UK	10 (5-15)
Sweden → Finland	10 (5-15)

Table S3. Analysis of Association Index ratio of phylogenetic distribution and trait (host, location, clustering based on accessory genome) performed with BaTS. The AI ratio ranges from 0 (perfect association) to 1 (no association).

Clonal Complex	# Hosts	Host AI	# Locations	Location AI	# of MCL Clusters (*)	MCL AI
CC151	-	n/a	11	0.13 (0.13-0.14)	5	0.42 (0.42-0.44)
CC97	2	0.33 (0.29-0.36)	13	0.35 (0.34-0.36)	5	0.21 (0.21-0.22)
CC1	2	0.25 (0.24-0.28)	10	0.37 (0.37-0.37)	5	0.59 (0.57-0.61)
CC188	2	0.04 (0.04-0.05)	4	0.1 (0.09-0.11)	5	0.26 (0.26-0.26)
CC133	2	0.39 (0.31-0.49)	9	0.24 (0.24-0.25)	3	0.51 (0.48-0.52)
CC130	3	0.67 (0.58-0.79)	6	0.20 (0.20-0.20)	3	0.34 (0.31-0.38)
CC425	2	0.26 (0.19-0.38)	-	n/a	2	0.41 (0.35-0.51)

(*) Clusters of accessory genomes defined using $i = 1.40$

Table S4: Goodman-Kruskal tau ($GK\tau$) values for association between accessory genome clusters and host/location. The values range from 0 (no predictability) to 1 (full predictability), i.e. the higher the value the better clustering matches/predicts the metadata variable.

Clonal Complex	Cluster threshold (MCLi)	# clusters	# hosts	#locations	G-Kτ Host	G-Kτ Location
CC1	1.40	5	2	14	0.569	0.163
	2.10	7	2	14	0.579	0.211
CC97	1.40	5	3	13	0.607	0.086
	2.00	6	3	13	0.611	0.097
CC130	1.40	3	3	7	0.275	0.138
	2.10	4	3	7	0.282	0.143
CC133	1.40	3	2	9	0.066	0.169
	2.10	4	2	9	0.150	0.18
CC151	1.40	8	1	11	n/a	0.312
	2.10	11	1	11	n/a	0.418
CC188	1.40	5	2	4	0.355	0.310
	2.10	8	2	4	0.565	0.435
CC425	1.40	2	2	4	0.000	0.012
	2.10	5	2	4	0.112	0.605

Table S5: Distribution of the *mecA* gene among selected Clonal Complexes (CCs) and Host species

	Bovine	Human	Swine	Ovine	Other	Total
CC398	58/82 (70.73%)	332/417 (79.62%)	55/91 (60.44%)	na	46/70 (65.71%)	491/660 (74.39%)
CC5	10/26 (38.46%)	229/326 (70.25%)	35/45 (77.78%)	na	0/57 (0%)	274/408 (67.16%)
CC8	10/38 (26.32%)	254/363 (69.97%)	na	na	10/20 (50%)	274/421 (65.08%)
CC45	55/69 (79.71%)	10/62 (16.13%)	na	na	0/3 (0%)	65/134 (48.51%)
CC1	1/119 (0.84%)	39/139 (28.06%)	0/2 (0%)	na	1/36 (2.78%)	41/296 (13.85%)
CC97	20/624 (3.21%)	6/63 (9.52%)	4/8 (50%)	na	4/7 (57.14%)	34/702 (4.84%)
CC130	1/114 (0.88%)	0/93 (0%)	0/16 (0%)	0/24 (0%)	0/9 (0%)	1/256 (0.39%)
CC133	0/106 (0%)	0/1 (0%)	na	1/19 (5.26%)	0/4 (0%)	1/130 (0.77%)
CC151	0/276 (0%)	na	na	na	na	0/276 (0%)
CC188	0/92 (0%)	1/66 (1.52%)	1/1 (100%)	na	0/8 (0%)	2/167 (1.20%)
CC425	1/126 (0.79%)	0/15 (0%)	na	na	0/2 (0%)	1/143 (0.70%)
Other	21/324 (6.48%)	245/664 (36.90%)	11/57 (19.30%)	na	4/203 (1.97%)	281/1,248 (22.52%)
Total	177/1,996 (8.87%)	1,116/2,209 (50.52%)	106/220 (48.18%)	1/43 (2.33%)	65/373 (17.43%)	1,465/4,841 (30.26%)

Table S6: Distribution of the *mecC* gene among selected Clonal Complexes (CCs) and Host species

	Bovine	Human	Swine	Ovine	Other	Total
CC130	109/114 (95.61%)	92/93 (98.92%)	16/16 (100%)	6/24 (25%)	2/36 (22.22%)	225/256 (87.89%)
CC425	101/126 (80.16%)	14/15 (93.33%)	na	na	0/2 (0%)	115/143 (80.42%)
CC133	1/106 (0.94%)	0/1 (0%)	na	1/19 (5.26%)	0/4 (0%)	2/130 (1.54%)
CC151	1/276 (0.36%)	na	na	na	na	1/276 (0.36%)
CC1	1/119 (0.84%)	0/139 (0%)	0/2 (0%)	na	0/36 (0%)	1/296 (0.34%)
Other	3/1,255 (0.24%)	1/1,888 (0.05%)	0/202 (0%)	na	1/295 (0.34%)	6/3,740 (0.36%)
Total	216/1,996 (10.82%)	107/2,209 (4.84%)	16/220 (7.27%)	7/43 (16.28%)	3/373 (0.80%)	349/4,841 (7.21%)