

# **Novel 3-Trifluoromethyl-1,2,4-Oxadiazole Analogues of Astemizole with Multi-Stage Antiplasmodium Activity and In vivo Efficacy in a *Plasmodium berghei* Mouse Malaria Infection Model**

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**ABSTRACT:** Iterative medicinal chemistry optimization of an ester containing astemizole (AST) analogue **1** with an associated metabolic instability liability led to the identification of a highly potent 3-trifluoromethyl-1,2,4-oxadiazole analogue **23** (*Pf*NF54 IC<sub>50</sub> = 0.012 μM; *Pf*K1 IC<sub>50</sub> = 0.040 μM) displaying high microsomal metabolic stability (HLM CL<sub>int</sub> < 11.6 μl.min<sup>-1</sup>.mg<sup>-1</sup>), and >1,000-fold higher selectivity over hERG compared to AST. In addition to asexual blood stage activity, the compound also shows activity against liver and gametocyte life cycle stages and demonstrates *in vivo* efficacy in *P. berghei*-infected mice at 4 × 50 mg.kg<sup>-1</sup> oral dose. Preliminary interrogation of the mode of action using live-cell microscopy and cellular heme speciation revealed that the **23** could be affecting multiple processes in the parasitic digestive vacuole, with the possibility of a novel target at play in the organelles associated with it.

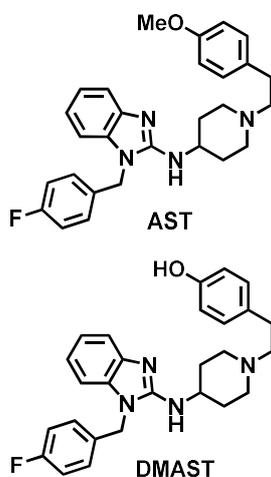
Keywords: Astemizole; repositioning; *Plasmodium falciparum*; *Plasmodium berghei*; human *ether-á-go-go*-related gene (hERG); Live-cell microscopy.

## INTRODUCTION

Malaria is a life-threatening infectious disease, affecting almost half of the world's population. It is caused by *Plasmodium falciparum* (*P. falciparum*, *Pf*) parasites and transmitted to humans by the female *Anopheles* mosquito.<sup>1</sup> Approximately 241 million global cases of malaria were recorded in 2020, with a mortality of 627,000. The Sub-Saharan Africa region accounted for 96% of total reported deaths,<sup>2</sup> and children under the age of 5 years were still among the most vulnerable groups. Notably, a steady decline in the global malaria mortality has been recorded between the year 2000 and 2019. This has been achieved through malaria elimination and prevention programs, increased investments in malaria research, improved diagnostic tools, and treatment among other malaria control initiatives.<sup>2</sup> However, malaria service disruptions during the COVID-19 pandemic resulted in a 12% rise in malaria deaths in 2020 compared to 2019.<sup>2</sup>

To date, the first line of treatment has been the effective artemisinin-based combination therapies (ACTs). However, reports of the emergence of resistance towards ACTs has continued to rise in western Cambodia to the Greater Mekong Subregion (GMS) and Africa, posing a serious threat to the global management and control of malaria.<sup>3,4</sup> The development of novel, pan active and affordable chemotherapies with no cross resistance to existing medicines to contribute to the pipeline of drugs under development is therefore imperative.

Astemizole (AST, Figure 1) is a well-known second-generation antihistamine. However, it was withdrawn from the market in most countries due to its rare but fatal side effect, implicated by QTc prolongation, torsades de pointes, cardiac arrhythmia due to hERG K<sup>+</sup> channel blockade.<sup>5,6</sup> AST and its principle metabolite Desmethylastemizole (DMAST, Figure 1) were first identified to possess antimalarial properties by Chong and co-workers.<sup>7</sup>



**Figure 1:** Chemical structures of astemizole (AST) and desmethylastemizole (DMAST)

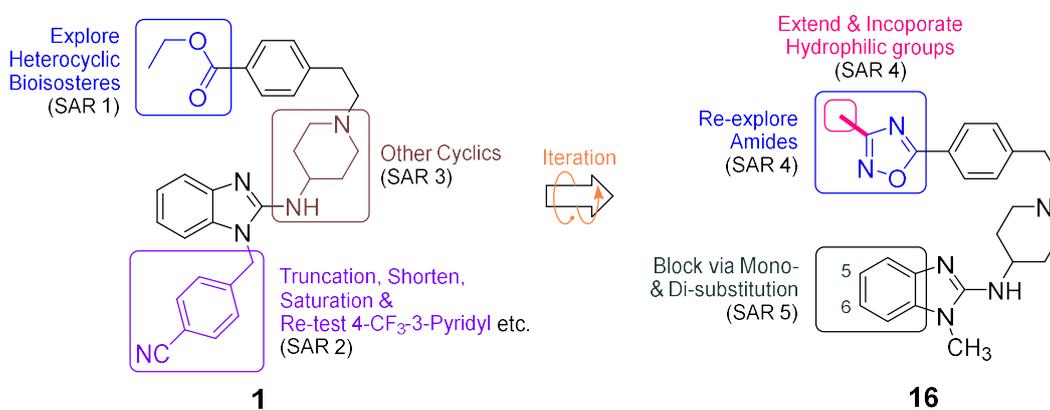
Medicinal chemistry efforts aimed at addressing the hERG liability and optimising antimalarial activity have previously been undertaken by a few groups and ourselves. Musonda and co-workers demonstrated the potential to overcome *Pf* resistance to chloroquine (CQ) via a CQ-AST hybridization approach and identified hybrids with high *in vivo* efficacy in the *P. berghei*

(*Pb*) mouse infection model of malaria.<sup>8</sup> Furthermore, Roman and co-workers (2013),<sup>9</sup> and De Jonghe and co-workers (2017),<sup>10</sup> separately revealed crucial antiplasmodium structure-activity relationships (SAR) of AST, in which the latter group identified a potent analogue (*Pf*3D7 IC<sub>50</sub> = 0.030 μM) with reduced hERG channel activity (hERG IC<sub>50</sub> = 0.030 μM; SI = 110). However, the poor drug-likeness characterized by high lipophilicity and poor solubility of the analogues would present development challenges.

To build on this work, our group initiated a project aimed at repositioning AST by expanding the existing antiplasmodium SAR around AST and improving drug-like properties. Initially, we reported AST analogues with improved potency, enhanced solubility and potential for multi-stage activity against asexual blood stages (ABS, *Pf*NF54 IC<sub>50</sub>'s = 0.033 – 1.9 μM), liver stages (*Pb* IC<sub>50</sub> = 0.210 μM), and late-stage gametocytes (stage IV/V; *Pf*LG IC<sub>50</sub>'s = 1.9 – 4.1 μM).<sup>11</sup> Furthermore, our work implicated intracellular inhibition of hemozoin formation by these analogues within the parasite as a contributing mode of action, which supported previous findings of interference of AST with the heme detoxification pathway.<sup>7,11</sup> With the exception of the hybridization work reported by Musonda and co-workers,<sup>8</sup> all studies so far have not progressed any analogues to proof-of-concept (PoC) *in vivo* efficacy studies. Herein, we report SAR studies leading to novel structural analogues of AST with *in vivo* PoC in mice and mechanistic insights using live-cell microscopy, providing significant advances over previous studies.

An ester-containing AST analogue **1** (Figure 2) emerged with high *in vitro* *Pf* activity and favourable solubility from our recent work; however, unsurprisingly it had poor microsomal metabolic stability.<sup>12</sup> This prompted an optimization campaign to improve microsomal metabolic stability towards identifying an AST analogue for PoC studies. While various strategies are

known to address ester group metabolic instability, bioisosterism is one of the most common approaches for hydrolysis-labile groups such as esters.<sup>13</sup> In this regard, we focused on ring replacements for the ethyl ester group in compound **1**, particularly exploring 5-membered heterocycles to maintain spatial geometry and increase structural rigidity (Figure 2, SAR 1).<sup>14,15</sup> Following the identification and fixing of a more stable ester surrogate in SAR 1, we then sought to explore other parts of the molecule in an effort to enhance potency, solubility as well as further reduce the hERG affinity and deliver a compound for *in vivo* efficacy studies.



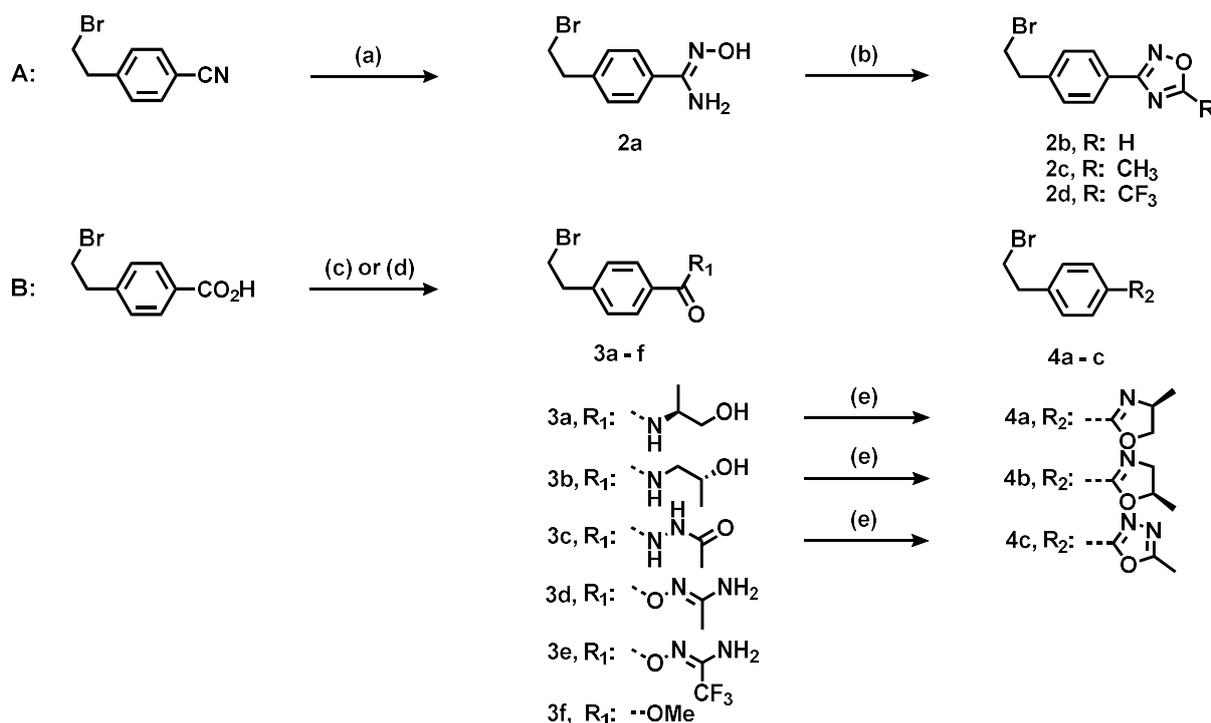
**Figure 2:** Progressive SAR design towards improving metabolic stability of compound **1**

In SAR 2, we revisited the previous SAR around the benzyl moiety, this time including ring saturation (Figure 2). In SAR 3, the 4-aminopiperidine linker was replaced with other cyclics not previously explored. Based on learnings from SARs 1 – 3, we iteratively replaced the 4-CN-benzyl group with *N*-methyl and explored SAR at the 3-position of the 1,2,4-oxadiazole moiety, including replacement with various amides (SAR 4, Figure 2). CYP-mediated hydroxylation of DMAST at benzimidazole position 6 is known to generate the 6-hydroxyldesmethylastemizole (6-OH-DMAST) metabolite *in vivo*, thereby contributing to the clearance of AST and DMAST.<sup>5</sup> For this reason, we sought to either block or sterically hinder the 6-benzimidazole position by exploring various substituents at both positions 5 and 6 (SAR 5, Figure 2).

## CHEMISTRY

The synthesis of target compounds commenced with the preparation of various phenethyl bromide intermediates (**2** – **4**) using modified literature methods (Scheme 1).<sup>16–20</sup>

### Scheme 1. Synthetic Approach for Intermediates **2** – **4**<sup>a</sup>

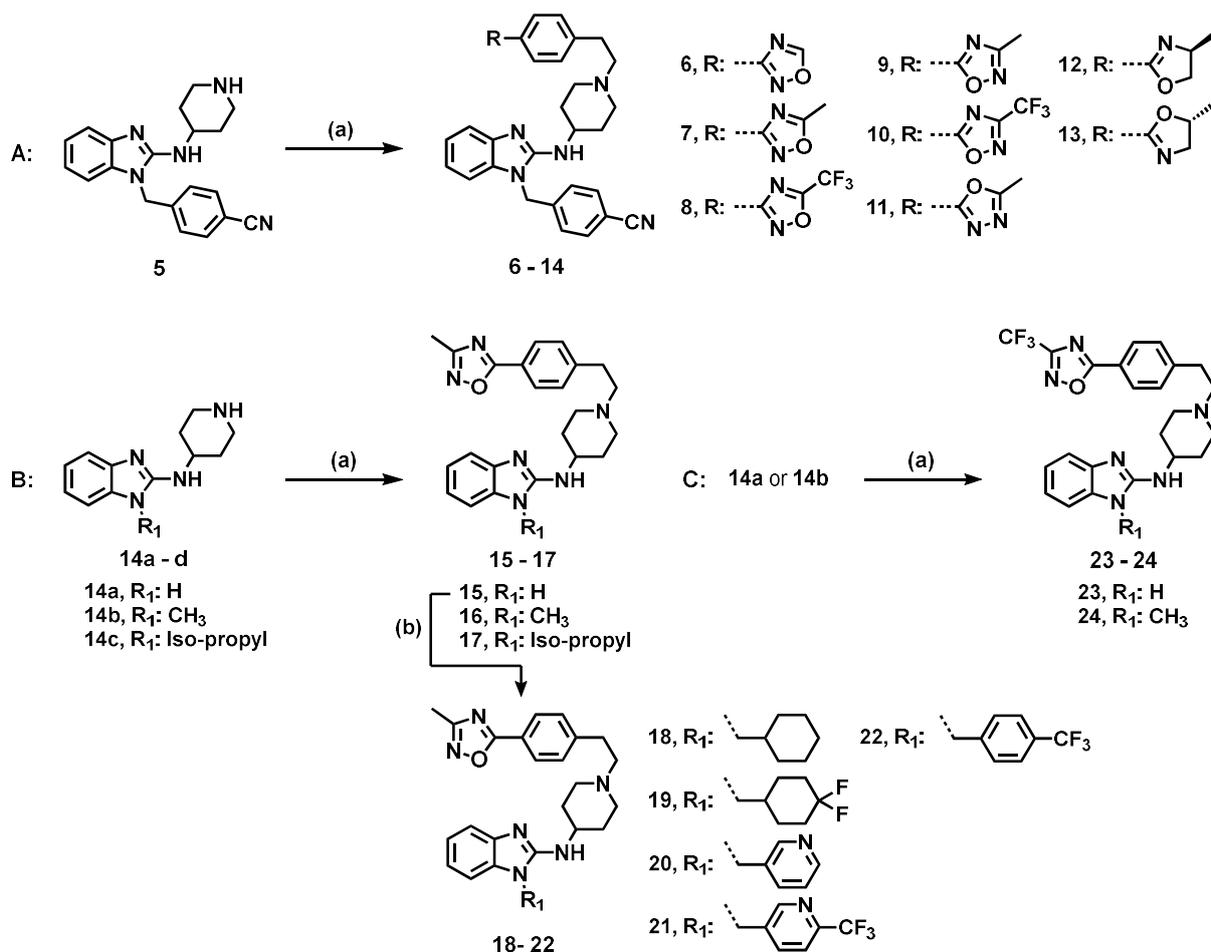


*Reagents and conditions:* (a) (i)  $\text{NH}_2\text{OH}\cdot\text{HCl}$ , 8-hydroxyquinolone,  $\text{Et}_2\text{N}$ ,  $\text{EtOH}$ ,  $79^\circ\text{C}$ , 1.5 h, (ii)  $21^\circ\text{C}$ , 10%  $\text{HCl}$ , pH 3 (82%); (b) for **2b**:  $\text{CH}(\text{EtO})_3$ ,  $\text{BF}_3\cdot\text{OEt}_2$ , Pyridine,  $80^\circ\text{C}$ , 1 h (51%); for **2c**: Acetyl chloride,  $50^\circ\text{C}$ , 0.5 h (80%); for **2d**:  $(\text{CF}_3\text{CO})_2\text{O}$ ,  $\text{DCM}$ , Pyridine,  $21^\circ\text{C}$ , 20 min (76%); (c) Amine or amidoxime,  $\text{EDCI}$ ,  $\text{DMAP}$ ,  $\text{DCM}$ ,  $18^\circ\text{C}$ , 2 h (65 – 91%); (d)  $\text{TMS}\cdot\text{CHN}_2$ ,  $\text{MeOH}$ , Toluene,  $21^\circ\text{C}$ , 0.5 h (**3f**, 97%); (e)  $\text{TsCl}$ ,  $\text{Et}_3\text{N}$ ,  $\text{DCM}$ ,  $30^\circ\text{C}$ , 0.5 h (**4a**, 70%; **4b**, 85%; **4c**, 88%).

Analogues **6** – **13** (SAR 1), and **15** – **24** (SAR 2) were synthesized by coupling phenethyl bromides **2** – **4**, or commercially sourced alkylating agents to previously reported amine intermediates **5** and **14a** – **b**, respectively (Scheme 1).<sup>12</sup> Compound **17** was prepared from

isopropyl intermediate **14c**, which had been synthesized following previously reported synthetic procedures.<sup>12</sup>

**Scheme 2. Synthetic Approach for Analogues 6 – 25 (SAR 1 & 2)**

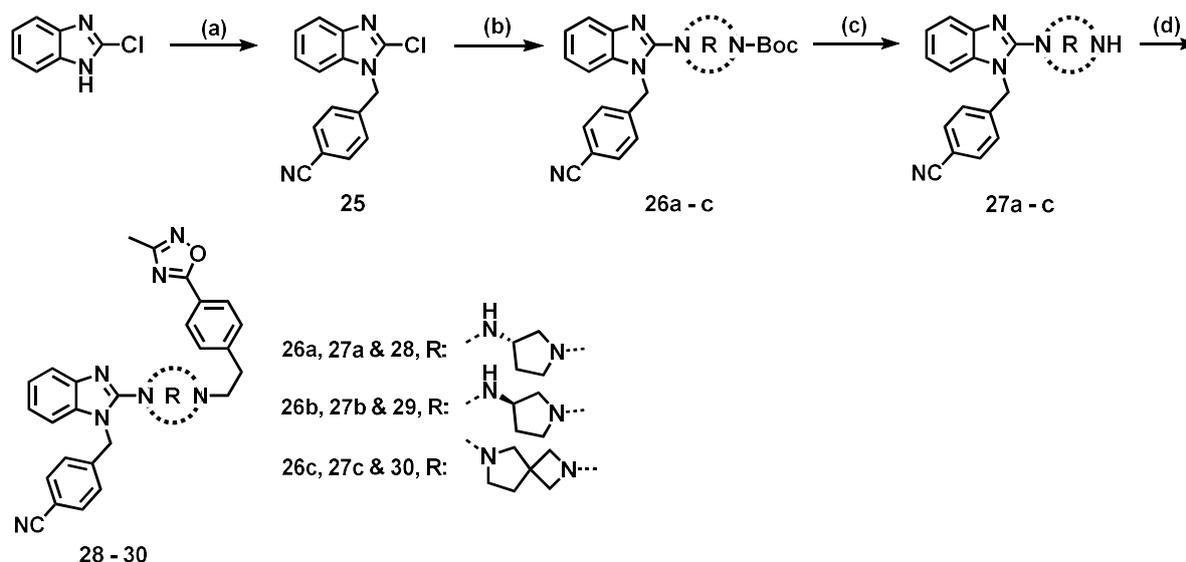


*Reagents and conditions:* (a) Phenethyl bromide (**2b-d**, **3d-e**, or **4a-c**), K<sub>2</sub>CO<sub>3</sub>, 80 °C, 5 – 12 h (48 – 91%); (b) Alkyl bromide, K<sub>2</sub>CO<sub>3</sub>, DMF, 70 °C, 12 h (58 – 80%).

Coupling of 2-chlorobenzimidazole and 4-(bromomethyl)benzotrile, followed by microwave-assisted nucleophilic aromatic substitution (S<sub>N</sub>Ar) with a series of commercially sourced mono *N*-Boc-protected diamines produced intermediates **26** in good yields (Scheme 3). *N*-Boc

deprotection of **26** resulted in free amine intermediates **27**, which were subjected to a final *N*-alkylation step with **3d** to afford analogues **28** – **30** in high yields.

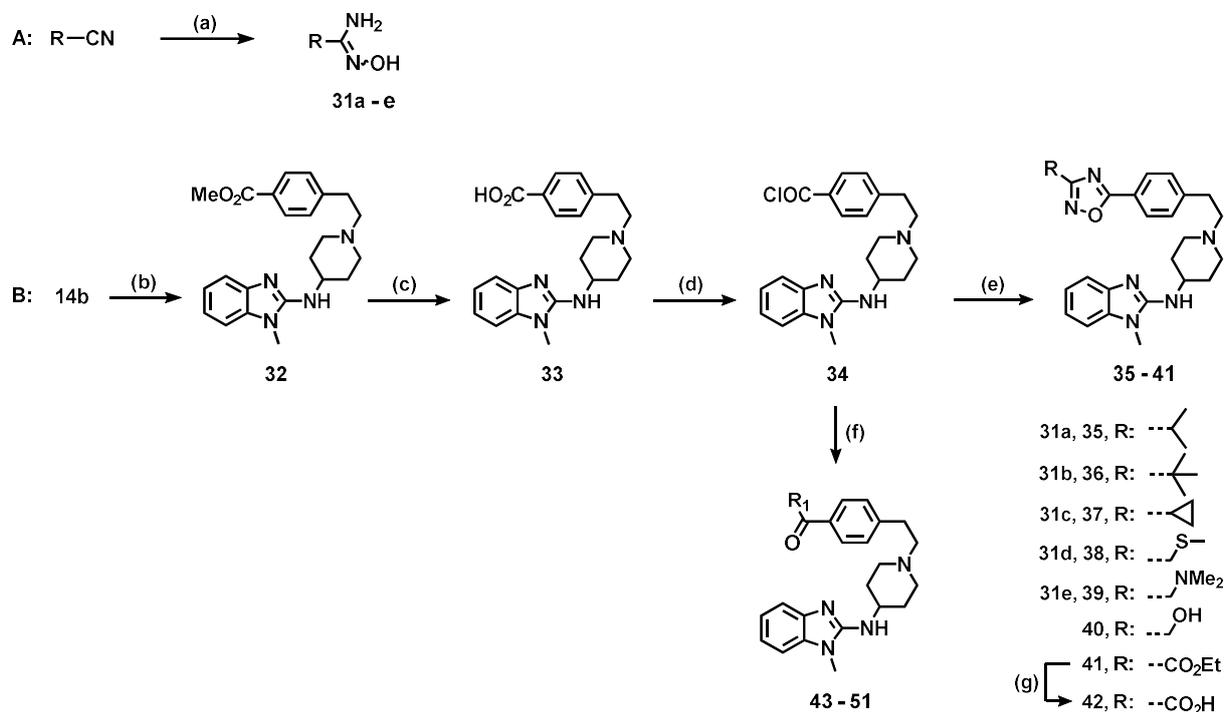
### Scheme 3. Synthetic Approach for Analogues **28** – **30** (SAR 3)



*Reagents and conditions:* (a) 4-(bromomethyl)benzonitrile,  $K_2CO_3$ , acetone, 23 °C, 2 h (98%); (b) *N*-Boc-amine,  $Et_3N$ , Toluene, 150 °C,  $\mu W$ , 5 – 30 min (55 – 78%); (c) TFA, DCM, 23 °C, 2 h (75 – 98%); (d) **3d**,  $K_2CO_3$ , 80 °C, 10 – 24 h (67 – 80%).

In SAR 4, various amidoximes (**31**) were initially prepared from the respective nitriles using hydroxylamine hydrochloride in EtOH (Scheme 4A). Secondly, a nucleophilic substitution reaction between **14b** and **3f** afforded ester **32**, which was hydrolysed to **33**, followed by treatment with thionyl chloride to give acyl chloride **34**, quantitatively. Amidoximes (**31**) and amines were coupled to **34** in the presence of  $Et_3N$  to produce *O*-acyl amidoximes and amides (**43** – **51**), respectively. The 3-substituted-1,2,4-oxadiazoles (**35** – **41**) were directly obtained from the respective *O*-acyl amidoximes *via*  $K_2CO_3$ -mediated cyclo-condensation in MeCN at 85 °C, while acid **42** was prepared from hydrolysis of ester **41** (Scheme 4B).

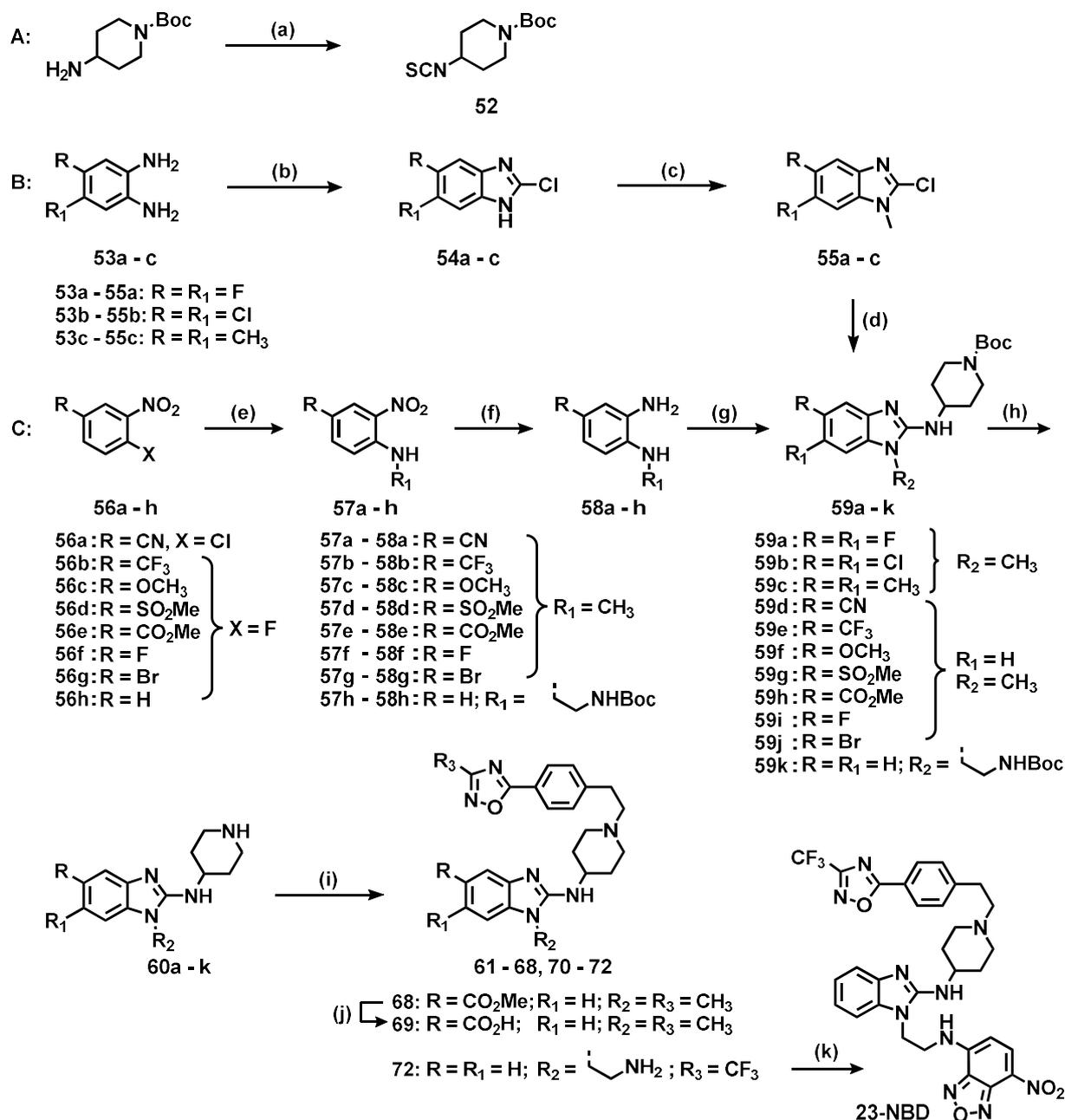
#### Scheme 4. Synthetic Approach for Analogues 35 – 50 (SAR 4)



*Reagents and conditions:* (a)  $NH_2OH \cdot HCl$ , 8-hydroxyquinolone,  $Et_3N$ , Ethanol,  $79^\circ C$ , 2 h, (70 – 95%); (b) **3f**,  $K_2CO_3$ ,  $80^\circ C$ , 5 h (68%); (c) 2M NaOH, MeOH,  $78^\circ C$ , 2 h, then 3N HCl, pH 2,  $20^\circ C$  (98%); (d)  $SOCl_2$ ,  $80^\circ C$ , 2 h (99%); (e) (i) Amidoxime,  $Et_3N$ , dry THF,  $20^\circ C$ , 2 – 10 h; (ii)  $K_2CO_3$ , MeCN,  $85^\circ C$ , 16 h (44 – 60% over two steps); (f) Amine,  $Et_3N$ , dry THF,  $23^\circ C$ , 6 – 10 h (66 – 98%); (g) 2N NaOH, EtOH,  $80^\circ C$ , 2 h; then 3N HCl, pH 2,  $20^\circ C$  (96%).

SAR 5 analogues were prepared *via* two converging synthetic routes (Scheme 5B and 5C) to access 5,6-disubstituted and 5-substituted benzimidazole analogues. For 5,6-disubstituted analogues, cyclization of commercially sourced 4,5-disubstituted-1,2-diamines (**53a–c**) with 1,1-carbonyldiimidazole (CDI), followed by chlorination using  $POCl_3$  produced corresponding 5,6-disubstituted-2-chloro-1*H*-benzimidazoles **54a–c**.

**Scheme 5. Synthetic Approach for Analogues 61 – 71 & 23-NBD (SAR 5)**



Reagents and conditions: (a) 1,1-Thiocarbonyldiimidazole, DMF, 23 °C, 12 h (97%); (b) (i) CDI, DMAP, THF, 20 °C, 12 h (quant’); (ii) POCl<sub>3</sub>, 110 °C, 12 h (58–93%); (c) CH<sub>3</sub>I, Acetone, K<sub>2</sub>CO<sub>3</sub>, 23 °C, 2 h (90–97%); (d) *tert*-butyl 4-aminopiperidine-1-carboxylate, Et<sub>3</sub>N, 150 °C, 2–12 h (66–85%); (e) 2M CH<sub>3</sub>NH<sub>2</sub> in THF (for **57a-g**) or *N*-Boc-ethylenediamine (for **57h**), Et<sub>3</sub>N, MeCN, 65 °C, 4–18 h, (66–96%); (f) H<sub>2</sub>,

balloon, 10% Pd/C, 1:1 MeOH/EtOAc, 21 °C, 12 h (89–96%); (g) **52**, DCC, TEA, MeCN, 85 °C, 12 h (80–95%); (h) TFA, DCM, 21 °C, 3 h (71–98%); (i) **3d** (for **61 – 71**) or **3e** (for **72**), MeCN, 85 °C, 8–12 h (46–78%); (j) 2M NaOH, EtOH, 25 °C, 12 h; then 3N aq. HCl, pH 2, 20 °C (94%); (k) 4-Chloro-7-nitrobenzoxadiazole, NaHCO<sub>3</sub>, 1:1 MeCN/H<sub>2</sub>O, 65 °C, 12 h (68%).

*N*-Methylation of **54a–c** using methyl iodide afforded 5,6-disubstituted-1-methyl-2-chlorobenzimidazoles **55a–c**, which were subjected to S<sub>N</sub>Ar coupling with *N*-Boc-4-aminopiperidine in Et<sub>3</sub>N at 150 °C, to give **59a–c** in good yields (Scheme 5B).

On the other hand, preparation of 5-substituted analogues was initiated by S<sub>N</sub>Ar reaction between methylamine (2M in THF) or *N*-Boc-ethylenediamine and an appropriately substituted *o*-halo-nitrobenzene (**56a–h**) in the presence Et<sub>3</sub>N at 65 °C in MeCN. The resulting 1-amino-2-nitrobenzenes (**57a–h**) were subjected to a reduction [H<sub>2</sub>, Pd/C] step to afford 1,2-diamines **58a–h** in high yields (Scheme 5C). *N,N'*-Dicyclohexylcarbodiimide (DCC)-mediated cyclization of the diamines **58a–h** with previously prepared isothiocyanate **52** (Scheme 5A) in MeCN produced 2-amino benzimidazoles **59d–k** in moderate yields (48 – 79%). *N*-Boc deprotection of **59a–k** using TFA afforded free amines **60a–k**, which were subsequently coupled (S<sub>N</sub>2) with **3d** to afford final compounds **61–68** and **70–71** in good yields. Carboxylic acid analogue **69** was obtained *via* hydrolysis of ester **68**, while 7-nitrobenzoxadiazole (NBD) tagged fluorescent probe **23-NBD** was prepared *via* S<sub>N</sub>Ar between corresponding amine **72** and 4-chloro-7-nitrobenzoxadiazole (NBD-Cl).

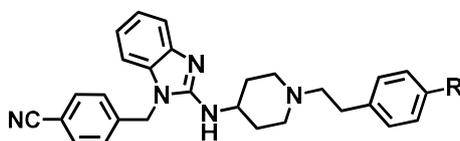
## RESULTS AND DISCUSSION

### *In vitro* Asexual Blood-Stage Antiplasmodium Activity and Solubility

All target compounds were evaluated for their *in vitro* antiplasmodium activity (Table 1) against the drug-sensitive strain of *P. falciparum* NF54 (*Pf*NF54) and aqueous turbidimetric kinetic solubility at pH 7.4. Compounds showing high activity ( $IC_{50} < 0.20 \mu M$ ) were further screened against the multi-drug resistant (MDR) strain *Pf*K1, to assess the potential of cross-resistance with existing antimalarial drugs.

**Bioisosteric Replacement of Ester Group (SAR 1).** Heterocyclic 1,2,4-oxadiazole and oxazoline isosteres produced analogues with general retention of potency (*Pf*-NF54  $IC_{50}$ 's = 0.021 – 0.104  $\mu M$ , Table 1) relative to the ester compound (**1**, *Pf*-NF54  $IC_{50}$ : 0.043  $\mu M$ ).

**Table 1:** *In vitro* antiplasmodium activity and solubility of SAR 1 analogues.



Compound	R	$PfIC_{50}$ ( $\mu M$ ) <sup>a</sup>		RI <sup>b</sup>	Sol. <sup>c</sup> ( $\mu M$ )	Compound	R	$PfIC_{50}$ ( $\mu M$ ) <sup>a</sup>		RI <sup>b</sup>	Sol. <sup>c</sup> ( $\mu M$ )
		NF54	K1					NF54	K1		
AST	OMe	0.086	0.370	4.3	40	<b>9</b>		0.097	0.178	1.8	20
<b>1</b>		0.043	0.052	1.2	40	<b>10</b>		0.022	0.027	1.2	10
<b>6</b>		0.089	0.399	4.5	40	<b>11</b>		0.519	ND		20
<b>7</b>		0.051	0.174	3.4	40	<b>12</b>		0.074	0.263	3.6	20
<b>8</b>		0.021	0.049	2.3	20	<b>13</b>		0.104	0.352	3.4	60
CQ		0.004	0.14	35	-						

<sup>a</sup>Mean from  $n \geq 2$  independent experiments with sensitive (NF54) and multidrug-resistant (K1) strains of *P. falciparum*; <sup>b</sup>Resistance Index (RI) =  $[(PfK1\ IC_{50})/(PfNF54\ IC_{50})]$ ; <sup>c</sup>Turbidimetric kinetic solubility at pH 7.4; ND = not determined; CQ = Chloroquine.

Interestingly, a ~2-fold increase in potency was consistently observed following the introduction of methyl (5-CH<sub>3</sub>, **7**, *Pf*-NF54 IC<sub>50</sub> = 0.051 μM) and trifluoromethyl (5-CF<sub>3</sub>, **8**, *Pf*-NF54 IC<sub>50</sub> = 0.021 μM) substituents relative to 5-unsubstituted 1,2,4-oxadiazole analogue **6** (IC<sub>50</sub> = 0.089 μM). Similarly, reversed 3-substituted-1,2,4-oxadiazoles displayed a 4-fold potency difference between 3-CH<sub>3</sub> (**9**, *Pf*-NF54 IC<sub>50</sub> = 0.097 μM) and 3-CF<sub>3</sub> (**10**, *Pf*-NF54 IC<sub>50</sub> = 0.022 μM) analogues.

Compared to 1,2,4-oxadiazoles, activity was diminished by 12-fold in 1,3,4-oxadiazole analogue **11** (*Pf*-NF54 IC<sub>50</sub> = 0.519 μM), an observation that may be attributed to the differences in the level of aromaticity between 1,2,4-oxadiazoles and 1,3,4-oxadiazoles. Being less aromatic, 1,2,4-oxadiazoles are more closely related to esters than 1,3,4-oxadiazole congeners are, and as a result, this potentially translates into differences in intrinsic biomacromolecule interaction between the two isomeric forms.<sup>21</sup> 4-Methyl-oxazoline **12** (*Pf*-NF54 IC<sub>50</sub> = 0.074 μM) displayed comparable activity with the 5-methyl-oxazoline congener **13** (*Pf*-NF54 IC<sub>50</sub> = 0.104 μM). All analogues from this SAR displayed low to moderate solubility (10 – 60 μM).

**Benzyl Group and Diamine Linker (SARs 2 and 3).** Next, we retained the 3-CH<sub>3</sub>-1,2,4-oxadiazole ring (**9**) and re-explored the SAR at the benzimidazole N-1 position. Consistently, activity was retained following replacement of the 4-CN-benzyl group with 4-CF<sub>3</sub>-containing aromatic moieties (**21**, *Pf*-NF54 IC<sub>50</sub> = 0.044 μM and **22**, *Pf*-NF54 IC<sub>50</sub> = 0.030 μM), and saturated (cyclohexyl)methyl (**18**, *Pf*-NF54 IC<sub>50</sub> = 0.093 μM, Table 2).

**Table 2:** *In vitro* antiplasmodium activity and solubility of SAR 2 analogues and NBD-probe **72**.

Compound	R	R <sub>1</sub>	<i>Pf</i> IC <sub>50</sub> (μM) <sup>a</sup>		RI <sup>b</sup>	Sol. <sup>c</sup> (μM)
			NF54	K1		
<b>15</b>	H		0.055	0.176	3.2	80
<b>16</b>	CH <sub>3</sub>		0.033	0.082	2.5	160
<b>17</b>			0.702	-	1.3	20
<b>18</b>			0.093	0.156	1.7	80
<b>19</b>		CH <sub>3</sub>	0.460	-		20
<b>20</b>			0.304	-		ND
<b>21</b>			0.044	0.173	3.9	40
<b>22</b>			0.030	0.104	3.5	10
<b>23</b>	H		0.012	0.040	3.3	100
<b>24</b>	CH <sub>3</sub>		0.066	0.048	0.7	160
<b>71<sup>d</sup></b>		CF <sub>3</sub>	0.090	0.036	0.40	>200
<b>23-NBD<sup>d</sup></b>			0.140	0.370	2.64	120
<b>CQ</b>			0.004	0.14	35.0	-

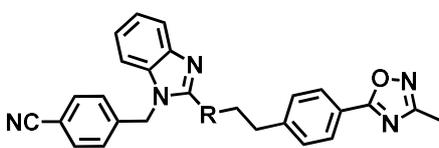
<sup>a</sup>Mean from  $n \geq 2$  independent experiments with sensitive (NF54) and multidrug-resistant (K1) strains of *P. falciparum*; <sup>b</sup>Resistance Index (RI) =  $[(Pf)K1 IC_{50}) / (Pf)NF54 IC_{50}]$ ; <sup>c</sup>Turbidimetric kinetic solubility at pH 7.4; <sup>d</sup>these compounds were tested at Swiss TPH using the 72 h [<sup>3</sup>H] hypoxanthine incorporation assay, ND = not determined; CQ = Chloroquine.

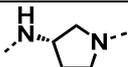
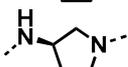
Interestingly, *N*-methyl analogue (**16**, *Pf*-NF54 IC<sub>50</sub> = 0.033 μM) and truncation of the benzyl group (**15**, *Pf*-NF54 IC<sub>50</sub> = 0.055 μM) produced compounds with high activity and solubility (>80 μM). Previously, this change drastically reduced activity in analogues containing a cyano (CN) group in place of the oxadiazole ring.<sup>12</sup>

Conversely, isopropyl (**17**) and (4,4-difluorocyclohexyl)methyl (**19**) analogues displayed reduced activities (Table 2) and low solubility (20 μM). Inspired by the identification of **15** and **16**, and the superior activity of 3-/5-CF<sub>3</sub> containing 1,2,4-oxadiazole analogues (**8** and **10**), we synthesized compounds **23** and **24**. Gratifyingly, **23** demonstrated a significant (4.6-fold) increase in potency (*Pf*-NF54 IC<sub>50</sub> = 0.012 μM) compared to **15**, while a 2-fold potency drop was observed in **24** (*Pf*-NF54 IC<sub>50</sub> = 0.066 μM) relative to **16**, albeit maintaining high solubility profiles (160 μM).

Using **9** as the benchmark, we next re-explored the 4-aminopiperidine linker SAR by replacement with (R)- and (S)-3-amino pyrrolidine, and 2,6-diazaspiro[3.4]octane (**28** – **29**, Table 3). All three analogues had comparable activities (**28** – **30**, Table 3), albeit ~5-fold lower compared to **9**. Additionally, all analogues in this SAR showed low to moderate solubility (40 – 80 μM).

**Table 3:** *In vitro* antiplasmodium activity and solubility of SAR 3 analogues.



Compound	R	<i>Pf</i> NF54 IC <sub>50</sub> ( $\mu$ M) <sup>a</sup>	Sol. <sup>c</sup> ( $\mu$ M)
28		0.515	80
29		0.448	80
30		0.446	40
CQ		0.004	-

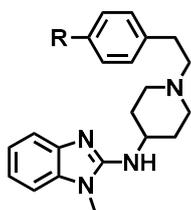
<sup>a</sup>Mean from  $n \geq 2$  independent experiments with sensitive (NF54) and multidrug-resistant (K1) strains of *P. falciparum*; <sup>b</sup>Turbidimetric kinetic solubility at pH 7.4; ND = not determined; CQ = Chloroquine.

**Exploring 3-position of 1,2,4-oxadiazole Moiety and Amidation (SAR 4):** In this SAR, compound **16** was iteratively used as a template to derive further analogues. We envisaged that *N*-methyl containing analogues would possess lower molecular weight, aromatic character, and lipophilicity ( $clogP$ ), which would potentially translate into favorable solubilities, and reduced hERG channel inhibition.

Functionalization of the 1,2,4-oxadiazole moiety at the 3-position by introducing both alkyl and polar atom (group)-containing moieties produced analogues with up to >15-fold lower activity (**35**, *Pf*-NF54 IC<sub>50</sub> = 0.492  $\mu$ M, Table 4), compared to **16**. However, the *tert*-butyl analogue **36** (*Pf*-NF54 IC<sub>50</sub> = 0.064  $\mu$ M) retained high activity and was equipotent to **24** (*Pf*-NF54 IC<sub>50</sub> = 0.066  $\mu$ M). This is not entirely surprising as the *tert*-butyl group has been widely used as an effective CF<sub>3</sub>-group surrogate. No apparent difference in activity was observed between cyclopropyl and isopropyl (**35** and **37**) groups, attributable to subtle steric differences between

the two groups.<sup>22,23</sup> Incorporation of a carboxylic acid moiety at this position was detrimental to activity (**42**, *Pf*-NF54 IC<sub>50</sub> > 6.0 μM), a consistent trend observed for all carboxylic acid-containing analogues of AST (i.e., **33**). Solubility was consistently lower in analogues containing alkyl groups compared to those with polar atoms (Table 4).

**Table 4:** *In vitro* antiplasmodium activity and solubility of SAR 4 analogues.



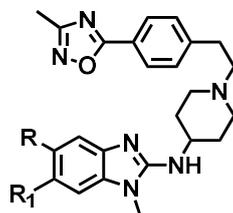
Compound	R	<i>Pf</i> IC <sub>50</sub> (μM) <sup>a</sup>		RI <sup>b</sup>	Sol. <sup>c</sup> (μM)	Compound	R	<i>Pf</i> NF54 IC <sub>50</sub> (μM) <sup>a</sup>	Sol. <sup>c</sup> (μM)
		NF54	K1						
<b>32</b>		1.450	-		120	<b>42</b>		4.17	120
<b>32b</b>		>6	-		160	<b>43</b>		1.73	60
<b>34</b>		0.492	-		60	<b>44</b>		>6	100
<b>35</b>		0.064	0.30	4.6	60	<b>45</b>		2.61	80
<b>36</b>		0.484	-		60	<b>46</b>		3.99	80
<b>37</b>		0.383	-		80	<b>47</b>		>6	80
<b>38</b>		0.831	-		120	<b>48</b>		>6	80
<b>39</b>		0.567	-		100	<b>49</b>		3.41	120
<b>41</b>		>6	-		120	<b>50</b>		1.28	60
<b>CQ</b>		0.004	0.14	35	-				

<sup>a</sup>Mean from n ≥ 2 independent experiments with sensitive (NF54) and multidrug-resistant (K1) strains of *P. falciparum*; <sup>b</sup>Resistance Index (RI) = [(*Pf*K1 IC<sub>50</sub>)/(*Pf*NF54 IC<sub>50</sub>)]; <sup>c</sup>Turbidimetric kinetic solubility at pH 7.4; ND = not determined; CQ = Chloroquine.

All amides displayed low antiplasmodium activities (**42** – **50**, *Pf*-NF54 IC<sub>50</sub> > 1.70 μM), with an unclearly discernable antiplasmodium SAR within the set of analogues prepared. Coupled with the observed high activity derived from analogues **15** and **16**, this suggested the apparent existence of SAR between the presence (or absence) of the benzyl group at *N*-1, relative to an appropriate functionality at the 4-position of the lateral phenyl group. All amides exhibited moderate to high solubility (60 – 180 μM).

**Benzimidazole Ring Substitution (SAR 5):** We next investigated substitution of the benzimidazole ring at positions 5 and 6 in front runner **16**. Activity was generally diminished across the SAR (Table 5).

Noteworthy, 5,6-dimethyl analogue **61** (*Pf*-NF54 IC<sub>50</sub> = 0.163 μM) displayed the highest activity overall, and in comparison, with halogen match pairs 5,6-F **62** (*Pf*-NF54 IC<sub>50</sub> = 1.16 μM) and 5,6-Cl **63** (*Pf*-NF54 IC<sub>50</sub> = 0.514 μM). Chloro substitution was better tolerated than fluoro substitution, resulting in ~2-fold higher activity in both the mono- and disubstituted fluoro congeners (**70**, *Pf*-NF54 IC<sub>50</sub> = 0.834 μM and **62**, *Pf*-NF54 IC<sub>50</sub> = 1.161 μM). Consistently, activity was lost in carboxylic acid derivative **69** (*Pf*-NF54 IC<sub>50</sub> > 6 μM) albeit displaying high solubility (Sol. = 160 μM).

**Table 5:** *In vitro* antiplasmodium activity and solubility of SAR 5 analogues.

Compound	R	R <sub>1</sub>	<i>Pf</i> NF54 IC <sub>50</sub> (μM) <sup>a</sup>	Sol. <sup>c</sup> (μM)	Compound	R	R <sub>1</sub>	<i>Pf</i> NF54 IC <sub>50</sub> (μM) <sup>a</sup>	Sol. <sup>c</sup> (μM)
<b>60</b>	CH <sub>3</sub>	CH <sub>3</sub>	0.163	60	<b>66</b>	SO <sub>2</sub> Me		4.651	60
<b>61</b>	F	F	1.161	80	<b>67</b>	CO <sub>2</sub> Me		0.496	120
<b>62</b>	Cl	Cl	0.514	80	<b>68</b>	CO <sub>2</sub> H	H	>6	160
<b>63</b>	CN		0.611	80	<b>69</b>	F		0.834	80
<b>64</b>	CF <sub>3</sub>	H	0.422	40	<b>70</b>	Br		0.720	80
<b>65</b>	OMe		0.309	80	<b>CQ</b> <sup>f</sup>			0.004	

<sup>a</sup>Mean from  $n \geq 2$  independent experiments with sensitive (NF54) and multidrug-resistant (K1) strains of *P. falciparum*; <sup>b</sup>Resistance Index (RI) =  $[(PfK1\ IC_{50})/(PfNF54\ IC_{50})]$ ; <sup>c</sup>Turbidimetric kinetic solubility at pH 7.4; ND = not determined; CQ = Chloroquine.

No potential for cross-resistance with existing antimalarial drugs was observed in analogues from this work based on the low resistance indices ( $RI < 5$ ) with the *Pf*K1 strain (Tables 1 – 2 and 4).

**Metabolic Stability.** Active compounds ( $Pf$ NF54-IC<sub>50</sub> < 0.10 μM) with moderate solubility (> 20 μM) were evaluated for their microsomal metabolic stability using mouse and human liver microsomes (MLM and HLM, Table 6).<sup>24</sup> Following the replacement of the labile ethyl ester (**1**, MLM CL<sub>int</sub> = 323 μl.min<sup>-1</sup>.mg<sup>-1</sup>) group with heterocyclic surrogates, 3-unsubstituted 1,2,4-oxadiazole **6** (MLM CL<sub>int</sub> = 266 μl.min<sup>-1</sup>.mg<sup>-1</sup>) still showed a high metabolic instability profile

compared to 3-CH<sub>3</sub>-(**7**, MLM CL<sub>int</sub> = 67.4 μl.min<sup>-1</sup>.mg<sup>-1</sup>) and 3-CF<sub>3</sub>-(**8**, MLM CL<sub>int</sub> = 26.4 μl.min<sup>-1</sup>.mg<sup>-1</sup>) substituted match pairs in which the 3-position is blocked. Oxazoline derivative **12** (MLM CL<sub>int</sub> = 71.3 μl.min<sup>-1</sup>.mg<sup>-1</sup>) also showed marked improvement, albeit sub-optimal (MLM CL<sub>int</sub> > 12 μl.min<sup>-1</sup>.mg<sup>-1</sup>; ≤ 95% remaining) especially in the rodent strain (MLM) across the SAR. This was, however, circumvented following the truncation of the benzyl group and retaining the 3-CF<sub>3</sub>-1,2,4-oxadiazole moiety (**23** and **24**, MLM CL<sub>int</sub> < 11.6 μl.min<sup>-1</sup>.mg<sup>-1</sup>).

**Table 6: In vitro Microsomal Metabolic Stability of Selected Analogues**

Compound	Intrinsic Clearance, CL <sub>int</sub>		clogP <sup>a</sup>
	MLM	HLM	
<b>AST</b>	147	107	5.70
<b>1</b>	373.2	431.3	5.22
<b>6</b>	266	167.9	4.80
<b>7</b>	67.4	22.8	5.12
<b>8</b>	26.4	19.2	5.42
<b>9</b>	86.4	23	5.12
<b>12</b>	71.3	88.9	5.23
<b>15</b>	<11.6	<11.6	4.08
<b>16</b>	43.6	19.7	4.28
<b>18</b>	123.9	200.4	5.88
<b>21</b>	40.7	31.9	5.08
<b>22</b>	18.9	11.6	5.94
<b>23</b>	<11.6	<11.6	4.30
<b>24</b>	<11.6	<11.6	4.51
<b>35</b>	18.4	15.2	5.15

MLM = mouse liver microsomes; and HLM = human liver microsomes, expressed as percent (%) of drug remaining after incubation with microsomes for 30 minutes. CL<sub>int</sub> = predicted intrinsic clearance in μl.min<sup>-1</sup>.mg<sup>-1</sup>. Mean from n ≥ 2 independent experiments. <sup>a</sup>Determined using StarDrop Software, Version 6.11.

Unaided by metabolite identification experiments, this suggested that microsomal metabolism (i.e., phase I) also occurred in the benzyl moiety, in addition to the ester group in compound **1**. Consistent with literature, 3-/5-**CH**<sub>3</sub>-1,2,4-oxadiazole containing analogues (i.e., **7** and **16**) generally exhibited lower stability compared to 3-/5-**CF**<sub>3</sub>-1,2,4-oxadiazole containing analogues (i.e., **8** and **24**, Table 6).

***In vitro* Antigametocytic and Liver-Stage Activity:** To assess activity of the analogues during other *Plasmodium* life-cycle stages in line with target candidate profile 3 (TCP3) for new antimalarials,<sup>25</sup> and based on previously observed multi-stage activity of AST and some of its analogues,<sup>11,26</sup> selected analogues from this series were assessed for their antiplasmodium sexual (gametocyte) and liver-stage properties.

20 analogues representing the structural diversity of each SAR were subjected to a dual-point screen against early and late-stage gametocytes (EGs, II/III; LGs, IV/V) at 1.0  $\mu$ M and 5.0  $\mu$ M (Supporting Information Table S1) in a luciferase assay that allows for the determination of stage-specific gametocytocidal activity. None of the tested compounds showed high inhibitory activity (< 50% inhibition) in both EGs and LGs at 1.0  $\mu$ M. However, 9 analogues showed moderate (50 – 70% inhibition) to good activity (>70% inhibition) at 5  $\mu$ M, and predominantly against EGs (Table S1). IC<sub>50</sub> values were only determined for compounds showing activities >80% in the dual point assay. Notably, compounds devoid of the 4-CN-benzyl (or an aromatic moiety at benzimidazole *N*-1 position, i.e., AST) displayed higher activity against EG's than against LGs. Structurally related frontrunners **23**, **24**, and **36** showed the highest activities with IC<sub>50</sub>'s in the low micromolar range (*Pf*EG IC<sub>50</sub> = 1.18 – 1.67  $\mu$ M, Table 7) and > 2-fold higher compared to AST's activity against LGs (*Pf*LG IC<sub>50</sub> = 3.35  $\mu$ M, Supporting Information Table S1).<sup>11</sup>

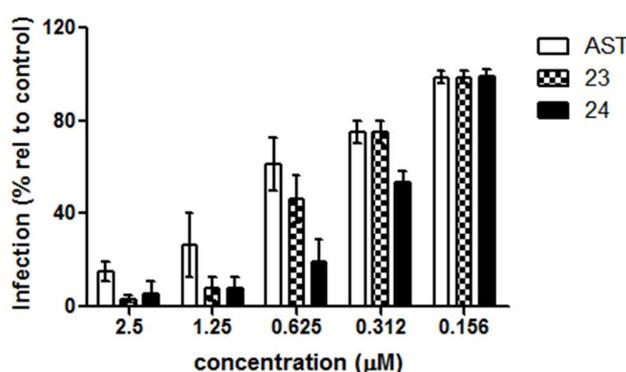
**Table 7: *In vitro* antiplasmodium Life-Cycle Stage Activity Data**

Compound	IC <sub>50</sub> , (μM)	
	<i>Pf</i> EG <sup>a</sup>	<i>Pb</i> HepG2 <sup>b</sup>
AST	-	0.59 ± 0.21
<b>23</b>	1.52 ± 0.25	0.49 ± 0.18
<b>24</b>	1.67 ± 0.28	0.21 ± 0.09
<b>36</b>	1.18 ± 0.28	-

<sup>a</sup>Early stage Gametocytes (EG Stages I/III), reference drug methylene blue (*Pf*EG IC<sub>50</sub> = 0.2 μM);

<sup>b</sup>*Plasmodium berghei* (*Pb*)-infected HepG2 cells, reference drug Primaquine (IC<sub>50</sub> = 6.0±1.4 μM).

Activity against liver stage infection was assessed using *P. berghei* in the HepG2 cell line. A preliminary single-point (1.0 μM) screen of only compounds with high *in vitro* ABS activity (*Pf*-NF54 IC<sub>50</sub> < 0.1 μM) and high microsomal metabolic stability (**15**, **23**, **24**, and **36**) showed that only **23** and **24** exhibited >75% reduction of parasite load with IC<sub>50</sub>'s of 0.49 and 0.21 μM, respectively (Table 7 and Figure 3). HepG2 cell confluency was assessed in parallel for each compound, as a measure of cytotoxicity (Supporting Information Table S2). Gratifyingly, no cytotoxicity was observed for all four compounds. (SI's > 80).



**Figure 3.** Concentration dependent inhibition of *P. berghei*-sporozoite infection in HepG2 cells. Values are mean, and error bars are standard deviation normalized from untreated cells (negative control)

***In vitro* hERG Channel Inhibition Activity.** Representative analogues from the five SARs were profiled for the potential for cardiotoxicity risk by assessing their hERG K<sup>+</sup> channel inhibitory activity (Supporting Information Table S3). The selection was made to best represent the various approaches used towards attenuating the hERG liability.

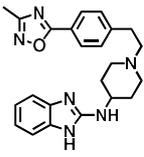
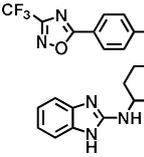
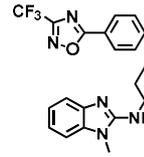
Although hERG inhibition is still an issue, all tested compounds showed lower hERG inhibitory concentrations (IC<sub>50</sub> > 0.10 μM) compared to AST. Zwitterion furnishing compounds (**42**, **69**) and amides (**44**, **50**: hERG IC<sub>50</sub> ≥ 30 μM) displayed the least hERG channel inhibitory activity, albeit accompanied by poor selectivity's (hERG SI < 18), as a result of equally low *in vitro* *Pf* activity. Similarly, removal of an aromatic moiety (i.e., **16**), ring saturation (i.e., **18**) and other lipophilicity lowering strategies (i.e., **67**) proved ineffective towards achieving high selectivity over hERG. Generally, high *Pf* inhibitors also produced relatively high hERG channel inhibition and vice-versa. However, higher selectivity's (hERG SI ≥ 19) were generally achieved in analogues with exceptional potency against *Pf*. These notably include 1,2,4-oxadiazole derivatives containing a CF<sub>3</sub> (**23** and **10**) and *t*-butyl (**36**) groups. Frontrunner compound **23** (hERG IC<sub>50</sub> = 0.63 μM; SI = 53) represents ~1,071-fold higher selectivity compared to AST (hERG IC<sub>50</sub> = 0.0042 μM).

**Cytotoxicity.** Analogues showing high antiplasmodium activity (*Pf*NF54-IC<sub>50</sub> < 0.10 μM) were profiled for cytotoxicity against the Chinese Hamster Ovary (CHO) cell line. No cytotoxicity was observed in this cell line, as all the tested compounds displayed ideal selectivity margins (SI > 34, Supporting Information Table S3).

***In vivo* Efficacy in Mice.** Compounds **15**, **23** and **24** (Table 8) were assessed for their *in vivo* efficacy in the *P. berghei* mouse infection model of malaria based on their high *in vitro* activity (*Pf*NF54 IC<sub>50</sub> < 0.10 μM), solubility and metabolic stability (MLMs CL<sub>int</sub> < 11.6 μl.min<sup>-1</sup>.mg<sup>-1</sup>).

In a standard quadrupole oral dose regimen of 50 mg.kg<sup>-1</sup>, **15**, **23** and **24** showed 40%, 99.5% and 90% reduction in parasitemia, respectively, relative to untreated mice (Table 8).

**Table 8: In vivo Efficacy after Oral Dosing in *P. Berghei*-infected Mice at 4 × 50 mg.kg<sup>-1</sup>**

Parameter	<b>15</b>	<b>23</b>	<b>24</b>
			
dose (mg.kg <sup>-1</sup> )	4 × 50	4 × 50	4 × 50
activity (%)	40	99.5	90
MSD*	4 <sup>a</sup>	14	9

\*MSD = mean survival days. <sup>a</sup>Mice were euthanized on day 4 in order to prevent expected death otherwise occurring at day 6 due to high parasitemia.

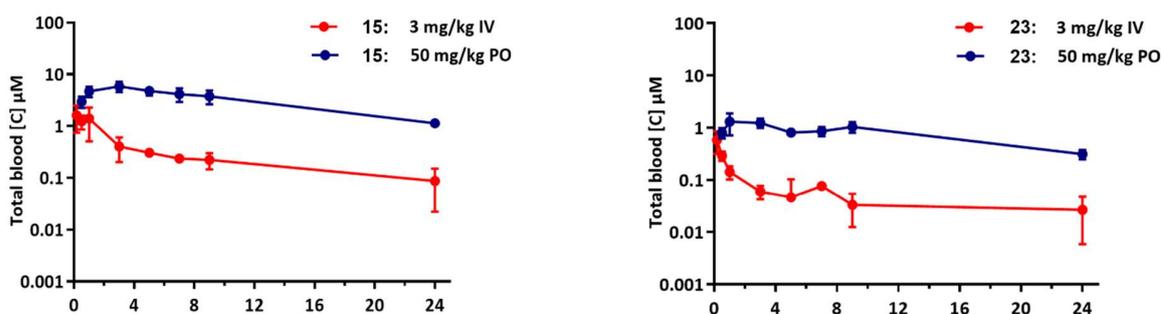
The high efficacy of **23** correlated to its high *in vitro* potency (*Pf*NF54 IC<sub>50</sub> 0.012 μM), although curative effects were not observed at this dose (mouse mean survival of < 30 days, Table 8). Chloroquine (CQ) was used as the reference drug, achieving 99.9% reduction in parasitemia when dosed orally at 4 × 30 mg.kg<sup>-1</sup>, with a mouse mean survival of 24 days.

**Pharmacokinetic Studies in Mice.** When dosed intravenously (IV, 3 mg.kg<sup>-1</sup>), **15** showed low clearance from blood (16.6 ml.min<sup>-1</sup>.kg<sup>-1</sup>, Table 9) with moderate tissue distribution (13.6 L.kg<sup>-1</sup>) resulting in a long half-life (11.6 h). Oral dosing of **15** at 10 mg.kg<sup>-1</sup> revealed rapid absorption (*T*<sub>max</sub> = 1.5 h) with moderate bioavailability (29.3%).

**Table 9: Mouse Pharmacokinetic Parameters of 15 and 23.**

Parameter	15			23		
	iv	oral		iv	oral	
dose (mg.kg <sup>-1</sup> )	3	10	50	3	10	50
C <sub>max</sub> (μM)	-	1	5.9	-	0.3	1.5
T <sub>max</sub> (h)	-	1.5	3.0	-	0.7	1.7
apparent t <sub>1/2</sub> (h)	11.6	10.3	9.2	4.2	1.1	10.2
CL <sub>int</sub> (ml.min <sup>-1</sup> .kg <sup>-1</sup> )	16.6	-	-	71.7	-	-
V <sub>d</sub> (L.kg <sup>-1</sup> )	13.6	-	-	26.6	-	-
AUC <sub>0-∞</sub> (μM.min <sup>-1</sup> )	550	535	5548	93.4	48	1429
F (%)	-	29.3	61	-	15.6	93.3

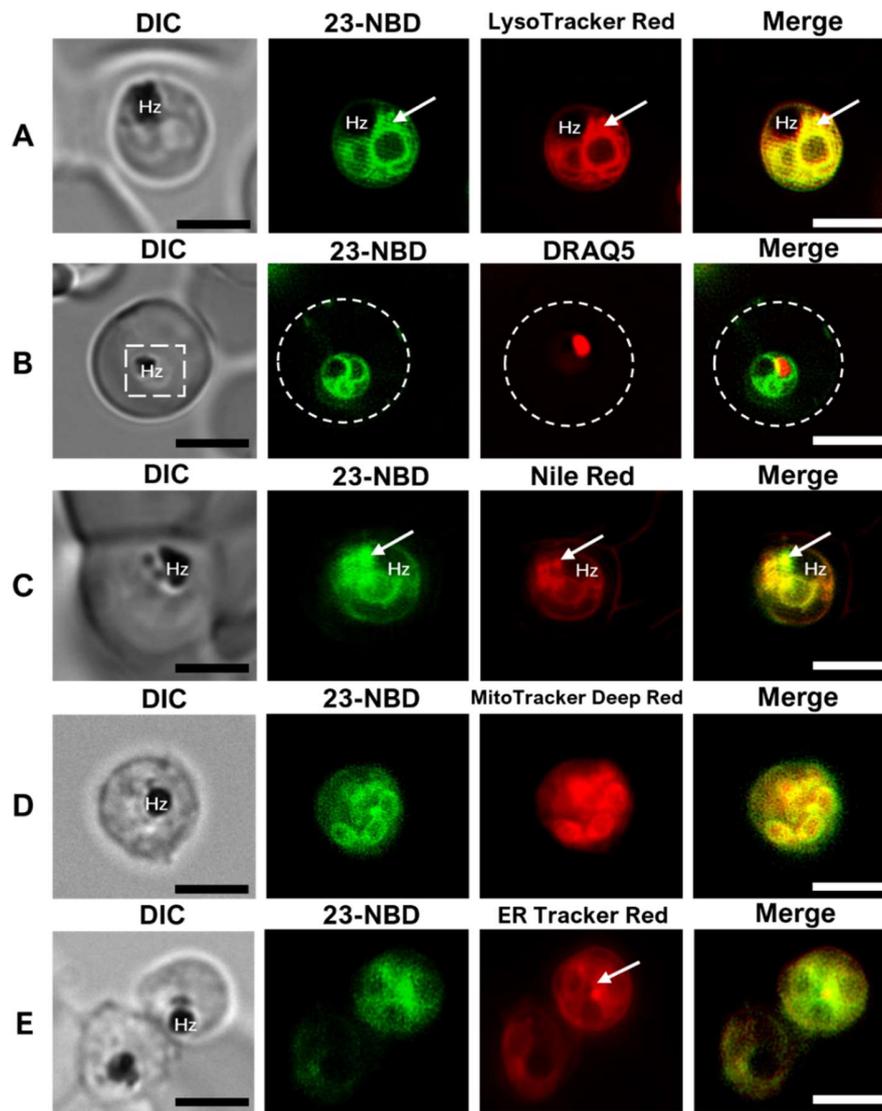
On the other hand, IV dosing of **23** at 3 mg.kg<sup>-1</sup> showed rapid clearance (71.7 ml.min<sup>-1</sup>.kg<sup>-1</sup>), albeit tissue distribution was high (26.6 L.kg<sup>-1</sup>) with a moderate half-life (4.2 h). Orally, **23** (T<sub>max</sub> = 0.7 h) was absorbed 2-fold faster than **15** at 10 mg.kg<sup>-1</sup>, and bioavailability was equally 2-fold lower and suboptimal (15.6%) at that dose. However, **23** displays a dose-dependent PK profile, as demonstrated by a greater than dose proportional increase in oral exposure at 50 mg.kg<sup>-1</sup> relative to the 10 mg.kg<sup>-1</sup> dose.



**Figure 4:** Blood concentrations of **15** and **23** following *intravenous* (*iv*) and oral (*p.o.*) dosing in healthy BalbC mice

This increased *in vivo* exposure, combined with the higher *in vitro* whole-cell *Pf* potency against ABS (**23** IC<sub>50</sub> = 0.012 μM vs **15** IC<sub>50</sub> = 0.055 μM ) likely explains the better *in vivo* efficacy of **23**.

**Mechanistic Studies.** *Live-cell Microscopy Assay.* To shed light on the mechanism of action (MoA) of this class of compounds, **23** was initially investigated for its intrinsic fluorescence for *P. falciparum* live-cell imaging of using a fluorimeter. Excitation between 200 – 400 nm and emission between 400 – 800 nm at 1.0  $\mu$ M showed no significant response compared to the blank in both DCM and DMSO (Figure S3). This prompted the attachment of an extrinsic fluorophore to enable live-cell imaging. NBD was chosen based on its ready availability, small size, and stability over a biologically relevant pH range.<sup>27</sup> Its attachment point to **23** at *N*-1 was solely guided by SAR studies already described. Gratifyingly, the NBD-labeled derivative (probe) retained sub-micromolar *in vitro Pf* activity (**23**-NBD, *Pf*-NF54  $IC_{50}$  = 0.140  $\mu$ M; Table 2). The probe was subjected to super-resolution structured illumination microscopy (SR-SIM) to assess sub-cellular accumulation in *Pf*-infected cells. This was carried out using commercially available organelle trackers LysoTracker Red, MitoTracker Deep Red, DRAQ5, ER-Tracker Red and Nile Red (Figure 4A – E).



**Figure 4:** Live-cell SR-SIM of *P. falciparum*-infected erythrocyte treated with 23-NBD (Green, Panels A – E), with LysoTracker Red (Red, Panel A), with nuclear marker DRAQ5 (Red, Panel B), with Lipid Marker Nile Red (Red, Panel C), with MitoTracker Deep Red (Red, Panel D), and with ER Tracker Red (Red, Panel E). White arrows indicate areas of intense localization of the dye and regions of complete overlap are shown in the merged image (Yellow). Scale Bars: 5 micrometres ( $\mu\text{m}$ ). Hz: Hemozoin; DIC: differential interference contrast.

These fluorescent dyes are capable of illuminating respective membranes of acidic organelles like the parasite's digestive vacuole (DV), the mitochondrion, nucleus, endoplasmic reticulum, and lipids, respectively.<sup>28-30</sup>

The result revealed complete accumulation between **23**-NBD and the LysoTracker Red (Figure 4A). Regions of highest accumulation were observed around the hemozoin (Hz) crystals, thereby implicating the parasite's digestive vacuole as a plausible site of action of **23**-NBD. Conversely, no colocalization was observed between **23**-NBD and DRAQ5, eliminating the nucleus as a possible site of action (Figure 4B).

Lipids have been shown to play vital roles during the asexual blood stage of the parasite's life cycle. Precisely, neutral lipids have been implicated in the formation of  $\beta$ -hematin *in vitro* and have been found at the site of Hz formation *in vivo*. It is believed that neutral lipids may serve as a site for Hz biocrystallization.<sup>31</sup> Following significant accumulation of **23**-NBD around the Hz crystals, accumulation with the parasite's neutral lipids was also assessed. Neutral lipid droplets were found in close proximity to the Hz crystals and these droplets colocalize with **23**-NBD (Figure 4C). Coupled with accumulation of the NBD probe (**23**-NBD) in the DV, this data reiterates inhibition of hemozoin formation as a possible contributing mechanism of action for compound **23**.

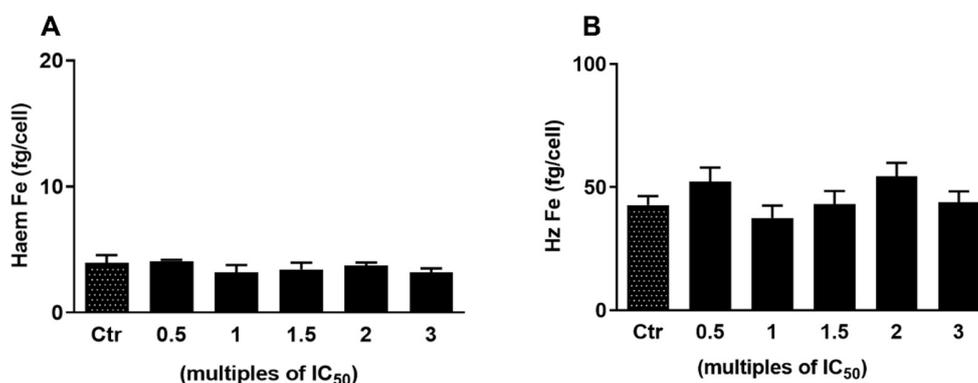
Aside from the digestive vacuole, the parasite possesses other organelles that equally play significant roles during the blood stage of the parasite's life cycle such as the mitochondrion and the endoplasmic reticulum (ER). These organelles have previously been shown to be clinically relevant drug targets.<sup>32</sup> Consequently, we examined the ER and the mitochondrion in conjunction with **23**-NBD. When incubated with MitoTracker Deep Red, we observed localized tube-like structures corresponding to the mitochondrion within the parasite. These structures

partially colocalized with regions of high signal intensity from the **23**-NBD (Figure 4D). This indicates some extent of colocalization between the mitochondrion and the compound. Also, significant colocalization was observed between the punctuate structure revealed by the ER Tracker red and the diffused signal from the **23**-NBD (Figure 4E). Making both organelles a potential site of action of compound **23**.

*Hemozoin formation Inhibition.* To augment the results from the SIM-SR microscopy which suggests inhibition of hemozoin formation as a plausible MoA of this class of compounds, and based on precedence from previous findings on AST and its analogues,<sup>7,11,12</sup> a selected number of active analogues across the SAR (*Pf*NF54-IC<sub>50</sub> < 0.100 μM) including **23** were evaluated for their inhibition of β-hematin (βH) formation (Supporting Information Table S4).<sup>33</sup> We conjured a 100 μM discriminatory IC<sub>50</sub> to identify strong inhibitors from this assay. Eight (8) of the fourteen (14) tested compounds blocked βH formation (Figure S3), while three (3) compounds (**8**, **10** and **21**) displayed activities comparable to standard βH formation inhibitors such as amodiaquine (IC<sub>50</sub>: 10 ± 2 μM) and chloroquine (IC<sub>50</sub>: 22 ± 2 μM). A weak but positive correlation ( $r^2 = 0.1356$ ) was established between inhibition of βH formation and *in vitro Pf* activity (Supporting Information Figure S4).

Furthermore, frontrunner **23** was subjected to the cellular heme speciation assay to delineate its dose-dependent effect on various haem species in the parasite and assess its ability to inhibit intracellular Hz formation in *Pf* parasites. Despite its high activity in the pyridine-based βH assay (Table S3), **23** did not produce a statistically significant concentration-dependent increase in the levels of free haem relative to that of Hz (Figure 5 and Supporting Information Table S4). This disconnect may in part be attributable to the limitations of *in vitro* βH assay in recapitulating the

complex physiology involving intracellular drug activity such as membrane permeation and accumulation in the digestive vacuole (DV).



**Figure 5:** Dose-dependent haem fractionation profiles of compound **23**. The amount of “free” haem Fe (A) and hemozoin (Hz) Fe (B) at increasing concentrations of the compounds.

## CONCLUSION

A novel set of 1,2,4-oxadiazole analogues of AST with high metabolic stability have been identified through metabolic stability-driven optimization of compound **1**. Various approaches including bioisosterism and lipophilicity-reduction driven modifications have been used, leading to the identification of **23**, which demonstrates high *in vivo* efficacy in the *P. berghei* mouse infection model of malaria. Additionally, analogues showing high early stage (EG, stage I/III) gametocytocidal activity, liver stage activity, and absence of cytotoxicity against the CHO and HepG2 cell lines have been identified. SAR from this work reaffirms the significance of the 4-amino piperidine moiety, as an essential pharmacophoric component in AST analogues for antiplasmodium activity. This work has revealed that with a 3-CH<sub>3</sub>/3-CF<sub>3</sub>/3-*tert*-butyl-1,2,4-oxadiazole moiety at the 4-position of the lateral phenyl group, the benzyl moiety at benzimidazole *N*-1 is not required for asexual-blood stage (ABS) antiplasmodium activity.

Removal of the benzyl moiety while retaining a 1,2,4-oxadiazole functionality in the lateral phenyl group produces high *Pf* activity, high *in vitro* metabolic stability, low lipophilicity, and low molecular weight which benefit solubility and selectivity over hERG. Although several analogues still display  $\beta$ -hematin inhibition, and significant amounts **23** accumulates in and around the parasite's DV, **23** does not induce any statistically significant effect on the levels of haem or Hz in *Pf*. It is worth noting that inhibiting the formation of Hz is only one of many processes in the DV that can be inhibited by **23** to lead to the parasite's death. We speculate that although inhibiting the formation of Hz might not be the primary MoA of **23**, the compound could be affecting other processes in the DV. Also, there is the possibility of a novel target at play in the other organelles associated with **23**. Compound **23** is the first direct analogue of AST to demonstrate multi-stage activity, high *in vivo* efficacy, with >1000-fold better hERG selectivity compared to AST, while retaining ideal drug-like properties. Further PK and hERG optimization into safer margins by utilizing other approaches not explored in this work as well as further studies into their MoA is warranted.

## EXPERIMENTAL SECTION

All commercially available chemicals were purchased from either Sigma-Aldrich (Germany) or Combi-Blocks (United States).  $^1\text{H}$  NMR (all intermediates and final compounds) and  $^{13}\text{C}$  NMR (for target compounds only) spectra were recorded on Bruker Spectrometer at 300, 400 or 600 megahertz (MHz). Melting points for all target compounds were determined using a Reichert-Jung Thermovar hot-stage microscope coupled to a Reichert-Jung Thermovar digital thermometer (20°C – 350°C range). Reaction monitoring using analytical thin-layer chromatography (TLC) was performed on aluminum-backed silica-gel 60 F<sub>254</sub> (70–230 mesh) plates with detection and visualization done using (a) UV lamp (254/366 nm), (b) iodine vapors,

or (c) ninhydrin spray reagent. Column chromatography was performed with Merck silica-gel 60 (70–230 mesh). Chemical shifts ( $\delta$ ) are reported in ppm downfield from trimethylsilane (TMS) as the internal standard. Coupling constants ( $J$ ) were recorded in Hertz (Hz). Purity of compounds was determined by an Agilent 1260 Infinity binary pump, Agilent 1260 Infinity diode array detector (DAD), Agilent 1290 Infinity column compartment, Agilent 1260 Infinity standard autosampler, and Agilent 6120 quadrupole (single) mass spectrometer, equipped with APCI and ESI multimode ionization source. All compounds tested for biological activity were confirmed to have  $\geq 95\%$  purity by HPLC. Solubility, biological assays, and any experimental data not shown below (i.e., NMR of compound intermediates) is fully supplied and detailed in the Supporting Information.

#### **General Procedure 1. Synthesis of *N*-hydroxyamidines (2a and 31a – e)**

To a solution of nitrile (1 equiv) in absolute EtOH (0.10 M) was added  $\text{NH}_2\text{OH}\cdot\text{HCl}$  (1.2 eq), followed by  $\text{Et}_3\text{N}$  (1.2 equiv) and 8-hydroxyquinolone (10 mol%). The resulting mixture was refluxed at 79 °C for 1.5 hour. After completion, the solvent was evaporated *in vacuo*, the residue was dissolved with  $\text{H}_2\text{O}$  (10 ml) and acidified to pH 3 using 10% HCl. The precipitate was filtered off and again washed with 10% HCl and dried.

**4-(2-bromoethyl)-*N*-hydroxybenzimidamide (2a).** Obtained from 4-(2-bromoethyl)benzotrile as a light green crystalline solid (0.57 g, 82%).  $^1\text{H}$  NMR (300 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.57 (s, 1H), 7.61 (d,  $J = 8.3$  Hz, 2H), 7.27 (d,  $J = 8.3$  Hz, 2H), 5.75 (s, 2H), 3.74 (t,  $J = 7.1$  Hz, 2H), 3.14 (t,  $J = 7.1$  Hz, 2H). LC-MS (APCI+/ESI): found  $m/z = 244.0, 245.0$  [ $\text{M}+\text{H}$ ]<sup>+</sup> (cal. for  $\text{C}_9\text{H}_{11}\text{BrN}_2\text{O}$ , 243.01, 244.01). HPLC Purity: 98%,  $t_R = 0.318$  min.

***N'*-hydroxyisobutyrimidamide (31a)**. Obtained from isobutyronitrile (0.250 g, 3.61 mmol) as a pale-yellow solid (0.34 g, 92%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.90 (s, 1H), 6.51 (s, 2H), 2.55 (hept, *J* = 5.9 Hz, 1H), 1.05 (d, *J* = 5.9 Hz, 6H).

**3-(4-(2-bromoethyl)phenyl)-1,2,4-oxadiazole (2b)**. A solution of **2a** (0.063 g, 0.26 mmol) in CH(EtO)<sub>3</sub> (1.5 mL) was charged with pyridine (1.5 ml) and catalytic amount of BF<sub>3</sub>·OEt<sub>2</sub> was stirred at 80 °C for 1 hour. After the completion, the solvent was removed *in vacuo*, and the product obtained by purification by flash column chromatography using 0 – 10% EtOAc/Hexanes as white crystalline solid (0.05 g, 51%). <sup>1</sup>H NMR (300 MHz, Methanol-*d*<sub>4</sub>) δ 9.24 (s, 1H), 8.04 (d, *J* = 8.3 Hz, 2H), 7.43 (d, *J* = 8.3 Hz, 2H), 3.81 (t, *J* = 7.1 Hz, 2H), 3.13 (t, *J* = 7.0 Hz, 2H). LC (APCI<sup>+</sup>/ESI): HPLC Purity: 98%, *t*<sub>R</sub> = 0.318 min.

**3-(4-(2-bromoethyl)phenyl)-5-methyl-1,2,4-oxadiazole (2c)**. A mixture of **2a** (0.10 g, 0.41 mmol) and acetyl chloride (350 μl, 0.49 mmol) was refluxed in THF at 65 °C for 30 min. At completion, excess acetyl chloride and THF were evaporated under reduced pressure and the reaction mixture quenched with H<sub>2</sub>O (5 ml). This mixture was extracted with DCM (3 × 15 ml). Combined organic extracts were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporating off solvent under vacuum to afford product as white crystalline solid (0.17 g, 80%). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 7.68 (d, *J* = 8.3 Hz, 2H), 7.38 (d, *J* = 8.3 Hz, 2H), 3.76 (t, *J* = 7.0 Hz, 2H), 3.19 (t, *J* = 7.0 Hz, 2H), 2.17 (s, 3H). LC (APCI<sup>+</sup>/ESI): HPLC Purity: 99%, *t*<sub>R</sub> = 0.318 min.

**3-(4-(2-bromoethyl)phenyl)-5-(trifluoromethyl)-1,2,4-oxadiazole (2d)**. A mixture of **2a** (0.10 g, 0.41 mmol) in 4 ml pyridine and trifluoroacetic acid anhydride (172 μl, 1.23 mmol) was stirred at room temperature (19 °C) for 20 min. The mixture was poured in ice H<sub>2</sub>O (10 ml) and extracted with ethyl acetate (3 × 15 ml). The combined organic extracts were washed with brine and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. Product obtained after flash column chromatography using 0

– 10% EtOAc/Hexanes as a pale-yellow oil (0.15 g, 76%). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 8.16 (d, *J* = 8.2, 2H), 7.49 (d, *J* = 8.2, 2H), 3.80 (t, *J* = 7.0 Hz, 2H), 3.24 (t, *J* = 7.0 Hz, 2H). HPLC Purity: 97%, *t*<sub>R</sub> = 3.406 min.

**General Procedure 2. Synthesis of Intermediates 3a – 3e.** To a solution of 4-(2-bromoethyl) benzoic acid (1 equiv) in DCM under ice, DMAP (0.1 equiv), EDCI (1.5 equiv.) and Et<sub>3</sub>N (1.5 were added, and the solution stirred at room temperature (21 °C) for 30 minutes. The appropriate amine (1.0 equiv) was then added dropwise, and the resulting solution stirred at room temperature for 2 hours. After completion, the reaction was diluted with another aliquot of initial amount of DCM and washed with saturated NaHCO<sub>3</sub> (×3), followed by H<sub>2</sub>O (×2), then brine (×1). The DCM phase was then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent evaporated *in vacuo*, products obtained after washing with Et<sub>2</sub>O.

**(S)-4-(2-bromoethyl)-N-(1-hydroxypropan-2-yl)benzamide (3a).** Obtained from (*S*)-2-aminopropan-1-ol (0.055g, 0.73 mmol) as a white solid (0.168 g, 80%). <sup>1</sup>H NMR (300 MHz, Methanol-*d*<sub>4</sub>) δ 7.89 (d, *J* = 8.2 Hz, 2H), 7.13 (d, *J* = 8.2 Hz, 2H), 3.71 (qt, *J* = 6.5, 6.2 Hz, 1H), 3.48 (t, *J* = 7.0 Hz, 2H), 3.12 (d, *J* = 6.2 Hz, 2H), 2.96 (t, *J* = 7.0 Hz, 2H), 1.65 (d, *J* = 6.5 Hz, 3H). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 286.0, 288.0 [M+H]<sup>+</sup> (cal. for C<sub>12</sub>H<sub>16</sub>BrNO<sub>2</sub>, 285.04, 287.03). HPLC Purity: 96%, *t*<sub>R</sub> = 2.593 min.

**Methyl 4-(2-bromoethyl) benzoate (3f).** To a solution of 4-(2-bromoethyl) benzoic acid (5 g, 21.8 mmol) in 4:1 Toluene/Methanol (60 ml) under ice, was added dropwise a solution of 2M of TMS-CHN<sub>2</sub> in hexanes until the yellow colour no longer dissipated (about 28 ml added in 25 minutes). The mixture was evaporated *in vacuo* to dryness, followed by purification *via* flash column chromatography using 0 – 20% EtOAc/Hexanes as eluent. Product obtained as a colourless/clear oil (5.15 g, 97%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.96 (d, *J* = 8.4 Hz, 2H),

7.20 (d,  $J = 8.4$  Hz, 2H), 3.85 (s, 3H), 3.54 (t,  $J = 4.4$  Hz, 2H), 3.14 (t,  $J = 4.4$  Hz, 2H). HPLC Purity: 99%,  $t_R = 2.955$  min.

**General Procedure 3: Synthesis of Intermediates 4a – 4c.** To a stirring suspension of amide **3a – 3c** (1 equiv) in DCM (5 ml), was added Et<sub>3</sub>N (2 equiv) followed by tosyl chloride (2 equiv). The mixture was stirred at 30 °C for 30 minutes. DCM was removed under vacuum followed by addition of EtOAc (15 ml). The mixture was washed with NaHCO<sub>3</sub> (2 × 10 ml) and brine (10 ml) then drying over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporating off solvent under vacuum. Crude solid products were purified by column chromatography using 0 – 25% EtOAc/hexanes gradient as an eluent.

**(S)-2-(4-(2-bromoethyl)phenyl)-4-methyl-4,5-dihydrooxazole (4a).** Obtained from **3a** (0.10 g, 0.35 mmol) as a white solid (0.066g, 70%). <sup>1</sup>HNMR (300 MHz, Methanol-*d*<sub>4</sub>) δ 8.01 (d,  $J = 8.2$  Hz, 2H), 7.41 (d,  $J = 8.2$  Hz, 2H), 4.11 (dd,  $J = 10.1, 7.9$  Hz, 1H), 3.96 (dd,  $J = 7.9, 6.2$  Hz, 1H), 3.67 (t,  $J = 7.1$  Hz, 2H), 3.33 (ddq,  $J = 10.1, 6.2, 6.0$  Hz, 1H), 3.24 (t,  $J = 7.1$  Hz, 2H), 1.95 (d,  $J = 6.0$  Hz, 3H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 268.0, 270.0$  [M+H]<sup>+</sup> (cal. for C<sub>12</sub>H<sub>14</sub>BrNO, 267.03, 267.02). HPLC Purity: 98%,  $t_R = 2.745$  min.

**1-Isopropyl-N-(piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (14c).**<sup>12</sup> White solid (0.336 g, 86%).  $R_f$ (10% MeOH/DCM), 0.23. <sup>1</sup>H NMR (400 MHz, Methanol-*d*<sub>4</sub>) δ 7.37 (dd,  $J = 7.7, 1.2$  Hz, 1H), 7.30 (dd,  $J = 8.0, 1.2$  Hz, 1H), 6.96 (ddd,  $J = 8.0, 7.2, 1.3$  Hz, 1H), 6.77 (ddd,  $J = 7.7, 7.2, 1.3$  Hz, 1H), 4.55 (hept,  $J = 6.8$  Hz, 1H), 4.20 (tt,  $J = 10.8, 4.0$  Hz, 1H), 3.98 – 3.91 (m, 2H), 3.05 – 2.96 (m, 2H), 2.11 – 2.03 (m, 2H), 1.59 (d,  $J = 6.8$  Hz, 6H), 1.55 – 1.47 (m, 2H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 259.2$  [M+H]<sup>+</sup> (cal. For C<sub>15</sub>H<sub>22</sub>N<sub>4</sub>, 258.18). Purity: 99%,  $t_R = 0.877$  min.

**General Procedure 5. N-alkylation of Benzimidazole.** A mixture of 2-chlorobenzimidazole (1 equiv), alkyl halide (1.2 equiv) and K<sub>2</sub>CO<sub>3</sub> (1.2 equiv) was stirred at room temperature (22 °C) in acetone for 2 hours. After completion, the solvent was removed *in vacuo*. H<sub>2</sub>O was added to the residue, followed by extracting with DCM (×3). Combined organic layers were washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent evaporated *in vacuo* to give crude residue, which was triturated with *n*-pentane to afford product.

**4-((2-chloro-1-benzo[d]imidazol-1-yl) methyl)benzonitrile (25).** Obtained from 2-chlorobenzimidazole (3.50 g, 22.9 mmol) and 4-(bromomethyl)benzonitrile (5.4 g, 27.6 mmol) as a white crystalline solid (6.00 g, 98%). R<sub>f</sub> (5% MeOH/DCM) 0.65. <sup>1</sup>H NMR (400 MHz, Methanol-*d*<sub>4</sub>) δ 7.68 (d, *J* = 8.4 Hz, 2H), 7.63 (dd, *J* = 6.0, 0.8 Hz, 1H), 7.39 (dd, *J* = 6.5, 1.0 Hz, 1H), 7.34 (d, *J* = 8.4 Hz, 2H), 7.31 – 7.27 (m, 2H), 5.59 (s, 2H). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 267.9, 269.9 [M+H]<sup>+</sup> (cal. for C<sub>15</sub>H<sub>10</sub>ClN<sub>3</sub>, 267.06, 269.05). Purity: 99%, *t*<sub>R</sub> = 2.505 min.

**General Procedure 6. Synthesis of 26a – c.** A mixture of **25** (1.0 equiv), an appropriate mono *N*-Boc protected cyclic diamine (1.5 equiv), and Et<sub>3</sub>N (2.0 equiv) in toluene was irradiated in a microwave reactor at 150 °C for 5 – 30 minutes. The residue was cooled and diluted with 10% MeOH/DCM. The mixture was washed with saturated NaHCO<sub>3</sub> solution (3 × 20 ml), combined organic layers were washed with brine (10 ml), and further dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The solvent was evaporated *in vacuo* to obtain a crude residue which was either triturated with *n*-pentane (or Et<sub>2</sub>O) or purified *via* flash column chromatography (3 – 4% MeOH/DCM) to afford the pure products.

***Tert*-butyl (S)-3-((1-(4-cyanobenzyl)-1H-benzo[d]imidazol-2-yl)amino)pyrrolidine-1-carboxylate (26a).** Obtained from **25** (0.300 g, 1.12 mmol) and *tert*-butyl (*S*)-3-aminopyrrolidine-1-carboxylate (293 μl, 1.68 mmol) as a light brown solid (0.304 g, 65%). R<sub>f</sub>

(10% MeOH/DCM), 0.50. <sup>1</sup>H NMR (600 MHz, Methanol-*d*<sub>4</sub>) δ 7.70 (d, *J* = 8.3 Hz, 2H), 7.49 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.38 (d, *J* = 8.3 Hz, 2H), 7.32 (ddd, *J* = 7.9, 7.0, 1.4 Hz, 1H), 7.28 (dd, *J* = 8.1, 1.2 Hz, 1H), 7.25 (ddd, *J* = 8.1, 7.0, 1.4 Hz, 1H), 5.62 (s, 2H), 4.70 (tt, *J* = 6.8, 4.2 Hz, 1H), 3.73 (dd, *J* = 12.9, 6.8 Hz, 1H), 3.55 – 3.47 (m, 2H), 3.40 (ddd, *J* = 11.5, 8.4, 5.7 Hz, 1H), 2.55 (ddd, *J* = 15.0, 8.4, 6.8 Hz, 1H), 2.29 (ddd, *J* = 11.5, 8.2, 4.1 Hz, 1H), 1.48 (s, 9H). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 418.2 [M+H]<sup>+</sup> (cal. for C<sub>24</sub>H<sub>27</sub>N<sub>5</sub>O<sub>2</sub>, 417.22). Purity: 99%, *t*<sub>R</sub> = 2.801 min.

**General Procedure 7. *N*-Boc deprotection.** Respective *N*-Boc protected amines were stirred in DCM and TFA (10 equiv) at 23 °C for 2 hours. Following completion, DCM and TFA were evaporated *in vacuo*, and the residue taken up 50% MeOH/DCM. This solution was stirred with Amberlyst<sup>®</sup> A-21 free base resin at room temperature (23 °C) until pH was neutral. The mixture was then filtered, and the filtrate evaporated *in vacuo* to afford free amines.

**(R)-4-((2-(Pyrrolidine-3-ylamino)-1H-benzo[*d*]imidazol-1-yl)methyl) benzonitrile (27a).**

Obtained from **26a** (0.280 g, 0.67 mmol) as a pale-yellow solid (0.181 g, 85%). *R*<sub>f</sub> (10% MeOH/DCM), 0.10. <sup>1</sup>H NMR (600 MHz, Methanol-*d*<sub>4</sub>) δ 7.73 (d, *J* = 8.4 Hz, 2H), 7.55 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.43 (d, *J* = 8.4 Hz, 2H), 7.38 (ddd, *J* = 8.0, 7.1, 1.5 Hz, 1H), 7.31 (dd, *J* = 8.2, 1.2 Hz, 1H), 7.29 (ddd, *J* = 8.2, 7.1, 1.4 Hz, 1H), 5.60 (s, 2H), 4.69 (tt, *J* = 6.7, 4.3 Hz, 1H), 3.79 (dd, *J* = 12.8, 6.7 Hz, 1H), 3.61 – 3.55 (m, 2H), 3.52 (ddd, *J* = 11.5, 8.2, 5.7 Hz, 1H), 2.56 (ddd, *J* = 14.7, 8.2, 6.7 Hz, 1H), 2.31 (ddd, *J* = 11.5, 8.2, 4.3 Hz, 1H). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 318.2 [M+H]<sup>+</sup> (cal. For C<sub>19</sub>H<sub>19</sub>N<sub>5</sub>, 317.16). Purity: 99%, *t*<sub>R</sub> = 2.499 min.

**Tert-butyl 4-isothiocyanatopiperidine-1-carboxylate (52).** To a solution of *tert*-butyl 4-aminopiperidine-1-carboxylate (8.00 g, 39.9 mmol) in DMF (50 ml) at 0 °C was added 1,1'-thiocarbonyldiimidazole (7.82 g, 43.9 mmol). The reaction mixture was allowed to rise to room

temperature (24 °C) and stirred for 20 hours at that temperature. The solvent was taken off *in vacuo*, the residue dissolved in EtOAc, and washed with H<sub>2</sub>O (3 × 50 ml). The solvent was removed *in vacuo*, the residue triturated with hexane, and filtered. The filtrate was treated with activated charcoal and filtered through Celite. Removal of solvent afforded product as a colourless oil (7.52 g, 78%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 4.11 – 3.98 (m, 2H), 3.69 (tt, *J* = 11.3, 4.1 Hz, 1H), 3.09 – 2.94 (m, 2H), 2.17 – 2.05 (m, 2H), 1.92 – 1.83 (m, 2H), 1.45 (s, 9H).

**General Procedure 8. Synthesis of 2-chlorobenzimidazoles 54a – c.** A mixture of commercially obtained benzene-1,2-diamines (1 equiv), 1,1'-Carbonyldiimidazole (1.5 equiv) and DMAP (1.0 mol%) was dissolved in dry THF (10 ml), and the resulting mixture stirred at 22 °C for 2 – 12 hr. After completion, the solvent was evaporated *in vacuo* and H<sub>2</sub>O (30 ml) added to the resulting residue. Benzimidazol-2-one products were obtained quantitatively, after filtering and subsequent oven drying. Crude benzimidazole-2-ones (1 equiv) were then treated with POCl<sub>3</sub> (5.0 equiv) in a seal tube and stirred at 110 °C for 12 h. The reaction mixture was then cooled to room temperature (22 °C), and excess POCl<sub>3</sub> evaporated under reduced pressure. The residue was taken up in EtOAc (20 ml) and neutralized using 15% NaOH under ice while stirring. After separation, the aqueous phase was further extracted with EtOAc (3 × 20 ml). The combined organic extracts were washed with brine solution (10 ml), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and solvent evaporated *in vacuo*. Pure products were obtained after flash chromatography using 20 – 50% EtOAc/Hexanes as eluent.

**2-chloro-5,6-difluoro-1H-benzo[d]imidazole (54a).** Obtained from 4,5-difluorobenzene-1,2-diamine (0.250g, 1.73 mmol) as a reddish solid (0.278 g, 84%). *R<sub>f</sub>*(5% MeOH/DCM), 0.76. <sup>1</sup>H NMR (300 MHz, Methanol-*d*<sub>4</sub>) δ 7.54 (m, 2H). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 189.0, 191.0 [M+H]<sup>+</sup> (cal. For C<sub>7</sub>H<sub>3</sub>ClF<sub>2</sub>N<sub>2</sub>, 188.00, 189.99). Purity: 98%, *t<sub>R</sub>* = 2.632 min.

**2-chloro-5,6-difluoro-1-methyl-1H-benzo[d]imidazole (55a).** Following the general procedure 2, obtained from **54a** (0.250 g, 1.32 mmol) and methyl iodide (99  $\mu$ l, 1.58 mmol) as a light green solid (0.212 g, 79%).  $R_f$ (5% MeOH/DCM), 0.93.  $^1\text{H}$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  7.42 (dd,  $J = 7.4, 5.3$  Hz, 1H), 7.27 (dd,  $J = 7.5, 5.5$  Hz, 1H), 3.83 (s, 3H). LC-MS (APCI $^+$ /ESI): found  $m/z = 203.0, 205.0$  [M+H] $^+$  (cal. for  $\text{C}_8\text{H}_5\text{ClF}_2\text{N}_2$ , 202.01, 204.01). Purity: 99%,  $t_R = 2.587$  min.

**General Procedure 9. Synthesis of 59a – c.** A mixture of **55a – c** (1.0 equiv), *N*-Boc-4-amino piperidine (1.5 equiv), and  $\text{Et}_3\text{N}$  (2.0 equiv) was stirred at 155  $^\circ\text{C}$  in a seal tube for 6 – 36 hours. After completion, the residue was cooled and diluted with 10% MeOH/DCM. The mixture was washed with saturated  $\text{NaHCO}_3$  solution ( $\times 3$ ), then brine, and dried over anhydrous  $\text{Na}_2\text{SO}_4$ . The solvent was evaporated *in vacuo* to obtain a crude residue which was either triturated with *n*-pentane (or diethyl ether) or purified *via* flash column chromatography to afford the pure products.

**Ethyl 4-((5,6-difluoro-1-methyl-1H-benzo[d]imidazol-2-yl) amino) piperidine-1-carboxylate (59a).** Obtained from **55a** (0.180 g, 0.88 mmol) and *tert*-butyl 4-aminopiperidine-1-carboxylate (0.265 g, 1.32 mmol), as a brown solid (0.184 g, 57%).  $R_f$ (5% MeOH/DCM), 0.78.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  7.40 (dd,  $J = 7.4, 5.3$  Hz, 1H), 7.32 (dd,  $J = 7.5, 5.5$  Hz, 1H), 6.69 (d,  $J = 7.6$  Hz, 1H), 4.51 – 4.42 (m, 2H), 3.91 (tt,  $J = 11.2, 4.0$  Hz, 1H), 3.52 (s, 3H), 3.09 – 2.98 (m, 2H), 2.15 – 2.06 (m, 2H), 1.59 – 1.51 (m, 2H), 1.39 (s, 9H). LC-MS (APCI $^+$ /ESI): found  $m/z = 367.2$  [M+H] $^+$  (cal. for  $\text{C}_{18}\text{H}_{24}\text{F}_2\text{N}_4\text{O}_2$ , 366.19). Purity: 98%,  $t_R = 2.579$  min.

**5,6-difluoro-1-methyl-N-(piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (60a).** Following general procedure 7, obtained from **59a** (0.160 g, 0.44 mmol) as a light brown solid (0.108 g, 93%).  $R_f$ (5% MeOH/DCM), 0.13.  $^1\text{H}$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  7.45 (dd,  $J = 7.4, 5.3$  Hz, 1H), 7.34 (dd,  $J = 7.5, 5.5$  Hz, 1H), 3.92 (tt,  $J = 11.2, 4.0$  Hz, 1H), 3.52 (s, 3H), 3.19 – 3.10 (m,

2H), 3.05 – 2.96 (m, 2H), 2.10 – 1.98 (m, 2H), 1.49 – 1.38 (m, 2H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 267.1$  [M+H]<sup>+</sup> (cal. for C<sub>13</sub>H<sub>16</sub>F<sub>2</sub>N<sub>4</sub>, 266.13). Purity: 96%,  $t_R = 0.822$  min.

**General Procedure 10. Synthesis of 58a – h.** A *o*-halo-nitrobenzene (1.0 equiv), an appropriate amine (1.2 equiv), and K<sub>2</sub>CO<sub>3</sub> (1.5 equiv) were mixed in MeCN (10 – 15 ml). Et<sub>3</sub>N (1.5 equiv) was added, and the resulting mixture stirred at 65 °C for 4 – 18 hr. After completion, the mixture was cooled to room temperature (18 °C), followed by the addition of H<sub>2</sub>O (15 ml) and EtOAc (30 ml). The mixture was separated, and the aqueous phase further extracted with EtOAc (2 × 15 ml). Combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and solvent evaporated *in vacuo*. Pure *o*-amino-nitrobenzene intermediates were obtained following recrystallization from ethanol and used in the next step.

To a solution of the crude *o*-amino-nitrobenzenes (1.0 equiv) in 1:1 MeOH/EtOAc (10 – 25 ml) was added 10% Pd/C (0.1 equiv), and the reaction mixture was stirred at 21 °C under a hydrogen (H<sub>2</sub>) atmosphere using a double padded balloon for 12 – 36 hours. After completion, the reaction mixture was filtered through a bed of Celite, and the filtrate concentrated *in vacuo* to afford products. If required, flash column chromatography using 20 – 70% EtOAc/Hexanes was performed.

**3-amino-4-(methylamino)benzonitrile (58a).** Obtained from 4-chloro-3-nitrobenzonitrile (0.500 g, 2.74 mmol) and methylamine (2M solution in THF, 1.64 ml, 3.29 mmol), as a dark brown solid (0.319 g, 79% over two steps).  $R_f$  (40% EtOAc/Hexane) 0.32. <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.58 (d,  $J = 1.6$  Hz, 1H), 7.28 (dd,  $J = 8.0, 1.6$  Hz, 1H), 6.21 (d,  $J = 8.0$  Hz, 1H), 5.02 (s, 2H), 2.71 (s, 3H). Purity: 99%,  $t_R = 0.153$  min.

**Tert-butyl (2-((2-aminophenyl)amino)ethyl)carbamate (58h).** Obtained from 1-fluoro-2-nitrobenzene (1.60 g, 5.70 mmol) and *tert*-butyl (2-aminoethyl)carbamate (1.10 g, 6.84 mmol) as

a dark brown solid (1.40 g, 98% over two steps).  $R_f$  (40% EtOAc/Hexane), 0.21.  $^1\text{H NMR}$  (300 MHz,  $\text{DMSO-}d_6$ )  $\delta$  6.88 (t,  $J = 5.7$  Hz, 1H), 6.55 (dd,  $J = 7.3, 1.6$  Hz, 1H), 6.52 – 6.37 (m, 3H), 4.39 (br-s, 3H), 3.21 – 3.11 (m, 2H), 3.11 – 3.00 (m, 2H), 1.40 (s, 9H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 252.1$   $[\text{M}+\text{H}]^+$  (cal. For  $\text{C}_{13}\text{H}_{21}\text{N}_3\text{O}_2$ , 251.16). Purity: 98%,  $t_R = 0.189$  min.

**General Procedure 11. Synthesis of 59d – k.** To a solution of **58a – h** (1 equiv) in MeCN (15 ml), isothiocyanate **52** (1.1 equiv) and  $\text{Et}_3\text{N}$  (1.2 equiv) were added, and the mixture refluxed at 85 °C for 1 hr. DCC (1.2 equiv) was then added, and the reaction mixture was further refluxed at 85 °C for 11 hr. After completion, MeCN was evaporated *in vacuo*, and the residue adsorbed on silica gel. Pure products were obtained after purification *via* flash chromatography using 10% MeOH/DCM as eluent.

**Tert-butyl 4-((5-cyano-1-methyl-1H-benzo[d]imidazol-2-yl) amino) piperidine-1-carboxylate (59d).** Obtained from **58d** (0.280 g, 1.90 mmol) and **52** (0.507 g, 2.10 mmol), as a light brown solid (0.573 g, 85%).  $R_f$  (5% MeOH/DCM), 0.53.  $^1\text{H NMR}$  (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  7.97 (d,  $J = 1.4$  Hz, 1H), 7.78 (d,  $J = 7.5$  Hz, 1H), 7.52 (dd,  $J = 7.5, 1.4$  Hz, 1H), 6.75 (d,  $J = 7.5$  Hz, 1H), 4.49 – 4.41 (m, 2H), 3.94 (tt,  $J = 10.8, 4.0$  Hz, 1H), 3.50 (s, 3H), 3.08 – 2.95 (m, 2H), 2.10 – 2.01 (m, 2H), 1.50 – 1.43 (m, 2H), 1.43 (s, 9H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 356.2$   $[\text{M}+\text{H}]^+$  (cal. For  $\text{C}_{19}\text{H}_{25}\text{N}_5\text{O}_2$ , 355.20). Purity: 99%,  $t_R = 2.397$  min.

**Tert-butyl 4-((1-(2-((tert-butoxycarbonyl)amino)ethyl)-1H-benzo[d]imidazol-2-yl)amino)piperidine-1-carboxylate (59k).** Obtained from **58h** (0.500 g, 2.00 mmol) and **52** (0.531 g, 2.18 mmol), as a white solid (0.780 g, 85%).  $R_f$  (7% MeOH/DCM), 0.38.  $^1\text{H NMR}$  (300 MHz,  $\text{DMSO-}d_6$ )  $\delta$  7.19 (dd,  $J = 7.9, 1.2$  Hz, 1H), 7.12 (dd,  $J = 8.0, 1.3$  Hz, 1H), 7.00 – 6.78 (m, 3H), 6.25 (d,  $J = 7.6$  Hz, 1H), 4.08 – 3.99 (m, 2H), 3.99 – 3.87 (m, 3H), 3.26 – 3.12 (m, 2H), 2.98 – 2.83 (m, 2H), 2.03 – 1.90 (m, 2H), 1.50 – 1.37 (m, 11H), 1.33 (s, 9H). LC-MS

(APCI<sup>+</sup>/ESI): found  $m/z = 460.3$  [M+H]<sup>+</sup> (cal. For C<sub>24</sub>H<sub>37</sub>N<sub>5</sub>O<sub>4</sub>, 459.28). Purity: 98%,  $t_R = 2.439$  min.

**1-methyl-2-(piperidin-4-ylamino)-1H-benzo[d]imidazole-5-carbonitrile (60d).** Following general procedure 7, obtained from **59d** (0.500 g, 1.41 mmol) as an off white solid (0.319 g, 89%).  $R_f$ (5% MeOH/DCM), 0.13. <sup>1</sup>H NMR (400 MHz, Methanol-*d*<sub>4</sub>)  $\delta$  7.97 (d,  $J = 1.4$  Hz, 1H), 7.78 (d,  $J = 7.5$  Hz, 1H), 7.52 (dd,  $J = 7.5, 1.4$  Hz, 1H), 3.94 (tt,  $J = 10.8, 4.0$  Hz, 1H), 3.50 (s, 3H), 3.21 – 3.13 (m, 2H), 3.08 – 2.99 (m, 2H), 2.09 – 1.98 (m, 2H), 1.52 – 1.39 (m, 2H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 256.2$  [M+H]<sup>+</sup> (cal. for C<sub>14</sub>H<sub>17</sub>N<sub>5</sub>, 255.15). Purity: 99%,  $t_R = 0.699$  min.

**1-(2-aminoethyl)-N-(piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (60k).** Following general procedure 7, obtained from **59k** (0.490 g, 10.7 mmol) as a white solid (0.235 g, 85%).  $R_f$ (10% MeOH/DCM), 0.09. <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.45 (br-s, 3H), 7.30 (dd,  $J = 7.1, 1.7$  Hz, 1H), 7.26 (dd,  $J = 7.1, 1.6$  Hz, 1H), 7.07 – 6.92 (m, 2H), 4.28 (t,  $J = 6.6$  Hz, 2H), 4.00 (tt,  $J = 10.3, 3.8$  Hz, 1H), 3.42 – 3.29 (m, 2H), 3.22 – 2.90 (m, 4H), 2.24 – 2.03 (m, 2H), 1.78 (dtd,  $J = 14.2, 10.9, 3.9$  Hz, 2H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 260.2$  [M+H]<sup>+</sup> (cal. for C<sub>14</sub>H<sub>21</sub>N<sub>5</sub>, 259.18). Purity: 98%,  $t_R = 0.186$  min.

**General Procedure 12. Synthesis of 6 – 13, 15 – 17, 23, 24, 28 – 30, 32, 61 – 68 & 70 – 72.** A solution of appropriate amine (1.0 equiv) and K<sub>2</sub>CO<sub>3</sub> (1.5 equiv) in MeCN was stirred under reflux at 80 °C for 30 minutes. An appropriate alkyl bromide (1.2 equiv) was added, and the mixture further stirred under reflux at 85 °C for 5 – 24 hours. After completion, MeCN was taken off under reduced pressure, the residue taken up in 10% MeOH/DCM and filtered. The filtrate was adsorbed on silica gel, after which column chromatography was performed using a 3 – 10% MeOH/DCM gradient as eluent, to afford final compounds.

**4-((2-((1-(4-(1,2,4-oxadiazol-3-yl) phenethyl) piperidin-4-yl) amino)-1-benzo[d]imidazol-1-yl) methyl) benzonitrile (6).** Obtained from **5** (0.080 g, 0.24 mmol) and **2b** (0.073 g, 0.29 mmol) as a pale-yellow solid (0.087 g, 72%); m.p.: 98 – 100 °C;  $R_f$ (10% MeOH/DCM), 0.59.  $^1\text{H}$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  7.65 (d,  $J = 8.7$  Hz, 2H), 7.63 (dd,  $J = 8.0$  Hz, 2H), 7.42 (d,  $J = 8.7$  Hz, 2H), 7.33 (dd,  $J = 7.9, 1.1$  Hz, 1H), 7.24 (d,  $J = 8.0$  Hz, 2H), 7.06 (ddd,  $J = 7.9, 7.2, 1.0$  Hz, 1H), 7.00 (dd,  $J = 8.0, 1.0$  Hz, 1H), 6.95 (ddd,  $J = 8.0, 7.2, 1.1$  Hz, 1H), 5.37 (s, 2H,  $\text{H}^5$ ), 3.84 (tt,  $J = 11.1, 4.2$  Hz, 1H), 3.06 – 3.01 (m, 2H), 2.94 – 2.87 (m, 2H), 2.70 – 2.66 (m, 2H), 2.36 – 2.30 (m, 2H), 2.12 – 2.08 (m, 2H), 1.67 – 1.58 (m, 2H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  154.10, 145.93, 142.30, 141.53, 140.68, 133.85, 132.25 (2C), 131.97 (2C), 129.47 (2C), 127.13 (2C), 121.34, 119.63, 118.44, 118.05, 114.82, 111.01, 109.67, 107.56, 58.97, 52.10 (2C), 49.83, 44.23, 32.67, 31.33 (2C). LC-MS (APCI $^+$ /ESI): found  $m/z = 504.2$  [M+H] $^+$  (cal. for  $\text{C}_{30}\text{H}_{29}\text{N}_7\text{O}$ , 503.24). Purity: 97%,  $t_R = 2.356$  min.

**4-((2-((1-(4-(5-methyl-1,2,4-oxadiazol-3-yl) phenethyl) piperidin-4-yl) amino)-1H-benzo[d]imidazol-1-yl) methyl) benzonitrile (7).** Obtained from **5** (0.080 g, 0.24 mmol) and **2c** (0.077 g, 0.29 mmol) as a pale-yellow solid (0.076 g, 61%); m.p.: 148 – 150 °C;  $R_f$ (10% MeOH/DCM) 0.48.  $^1\text{H}$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  8.02 (d,  $J = 7.9$  Hz, 2H), 7.70 (d,  $J = 8.0$  Hz, 2H), 7.47 (d,  $J = 7.9$  Hz, 2H), 7.41 (dd,  $J = 7.9, 0.9$  Hz, 1H), 7.30 (d,  $J = 8.0$  Hz, 2H), 7.17 (ddd,  $J = 7.9, 7.1, 1.0$  Hz, 1H), 7.12 (dd,  $J = 8.0, 1.0$  Hz, 1H), 7.08 (ddd,  $J = 8.0, 7.1, 1.0$  Hz, 1H), 5.46 (s, 2H), 4.04 (tt,  $J = 10.5, 4.1$  Hz, 1H), 3.75 – 3.63 (m, 2H), 3.44 – 3.38 (m, 2H), 3.26 – 3.18 (m, 2H), 3.18 – 3.12 (m, 2H), 2.65 (s, 3H), 2.40 – 2.31 (m, 2H), 1.99 – 1.89 (m, 2H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  177.47, 167.78, 152.31, 141.41, 139.89, 132.95, 132.38 (2C), 129.12 (2C), 127.43 (2C), 127.07 (2C), 125.74, 122.33, 121.13, 119.23, 117.95, 114.13,

111.28, 108.49, 57.10, 51.63 (2C), 44.63, 30.15, 29.19 (2C), 29.09, 10.64. LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 518.2$  [M+H]<sup>+</sup> (cal. for C<sub>31</sub>H<sub>31</sub>N<sub>7</sub>O, 517.26). Purity: 97%,  $t_R = 2.447$  min.

**N-(1-(4-(3-Methyl-1,2,4-oxadiazol-5-yl)phenethyl)piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (15)**. Obtained from **14a** (0.85 g, 3.93 mmol) and **3d** (1.35 g, 4.72 mmol) as a pale-yellow solid (1.18 g, 75%); m.p.: 171 – 173 °C;  $R_f$  (10% MeOH/DCM), 0.20. <sup>1</sup>H NMR (400 MHz, Methanol-*d*<sub>4</sub>) δ 8.04 (d,  $J = 8.3$  Hz, 2H), 7.46 (d,  $J = 8.3$  Hz, 2H), 7.20 (dd,  $J = 5.8, 3.2$  Hz, 2H), 6.98 (dd,  $J = 5.8, 3.2$  Hz, 2H), 3.69 (tt,  $J = 10.9, 4.3$  Hz, 1H), 3.10 – 3.03 (m, 2H), 2.97 – 2.91 (m, 2H), 2.75 – 2.68 (m, 2H), 2.42 (s, 3H), 2.40 – 2.31 (m, 2H), 2.15 – 2.08 (m, 2H), 1.71 – 1.59 (m, 2H). <sup>13</sup>C NMR (101 MHz, Methanol-*d*<sub>4</sub>) δ 176.08, 167.11, 154.29, 145.78, 137.16, 129.36 (2C), 127.75 (2C), 121.89, 120.10, 111.31, 59.21, 51.95 (2C), 49.33, 43.55, 32.72, 31.63 (2C), 9.99. LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 403.2$  [M+H]<sup>+</sup> (cal. For C<sub>23</sub>H<sub>26</sub>N<sub>6</sub>O, 402.22). Purity: 98%,  $t_R = 2.469$  min.

**N-(1-(4-(3-(trifluoromethyl)-1,2,4-oxadiazol-5-yl)phenethyl)piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (23)**. Obtained from **14a** (0.250 g, 1.16 mmol) and **3e** (0.474 g, 1.39 mmol) as a white crystalline solid (0.290 g, 55%). m.p.: 102 – 104 °C;  $R_f$  (10% MeOH/DCM), 0.38. <sup>1</sup>H NMR (600 MHz, Methanol-*d*<sub>4</sub>) δ 8.06 (d,  $J = 8.0$  Hz, 2H), 7.48 (d,  $J = 8.0$  Hz, 2H), 7.25 (dd,  $J = 5.8, 3.2$  Hz, 2H), 7.05 (dd,  $J = 5.9, 3.1$  Hz, 2H), 3.74 (tt,  $J = 10.5, 4.2$  Hz, 1H), 3.21 (dt,  $J = 12.3, 4.0$  Hz, 2H), 3.04 – 2.96 (m, 2H), 2.92 – 2.85 (m, 2H), 2.56 (td,  $J = 11.9, 2.7$  Hz, 2H), 2.21 – 2.13 (m, 2H), 1.80 – 1.69 (m, 2H). <sup>13</sup>C NMR (101 MHz, Methanol-*d*<sub>4</sub>) δ 169.05, 167.50, 153.15, 144.18, 135.53, 129.34 (2C), 127.50 (2C), 123.11, 120.87 (2C), 111.28 (2C), 58.70 (2C), 51.70, 49.10, 32.14, 30.94 (2C). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 457.2$  [M+H]<sup>+</sup> (cal. For C<sub>23</sub>H<sub>23</sub>F<sub>3</sub>N<sub>6</sub>O, 456.19). Purity: 98%,  $t_R = 2.246$  min.

**1-Methyl-N-(1-(4-(3-(trifluoromethyl)-1,2,4-oxadiazol-5-yl) phenethyl) piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (24)**. Obtained from **14b** (0.150 g, 0.65 mmol) and **3e** (0.266 g, 0.78 mmol) as a white solid (0.202 g, 66%). m.p.: 90 – 92 °C;  $R_f$ (10% MeOH/DCM), 0.35.  $^1\text{H}$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  8.12 (d,  $J = 8.3$  Hz, 2H), 7.56 (d,  $J = 8.3$  Hz, 2H), 7.51 – 7.45 (m, 2H), 7.40 – 7.33 (m, 2H), 4.09 – 3.99 (m, 1H), 3.87 – 3.79 (m, 2H), 3.72 (s, 3H), 3.52 – 3.46 (m, 2H), 3.27 – 3.20 (m, 4H), 2.44 – 2.36 (m, 2H), 2.24 – 2.12 (m, 2H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  168.88, 161.51, 149.24, 141.03, 131.40, 129.45 (2C), 128.97, 127.84 (2C), 124.01, 123.86, 123.70, 111.40, 109.61, 54.49 (2C), 42.38, 29.96, 28.28, 17.32, 15.90 (2C), 11.62. LC-MS (APCI $^+$ /ESI): found  $m/z = 471.2$  [M+H] $^+$  (cal. For  $\text{C}_{24}\text{H}_{25}\text{F}_3\text{N}_6\text{O}$ , 470.20). Purity: 99%,  $t_R = 2.375$  min.

**(R)-4-((2-((1-(4-(3-methyl-1,2,4-oxadiazol-5-yl)phenethyl) pyrrolidine-3-yl) amino)-1H-benzo[d]imidazol-1-yl) methyl) benzonitrile (28)**. Obtained from **27a** (0.070 g, 0.22 mmol) and **3d** (0.075 g, 0.26 mmol) as a pale-yellow solid (0.087 g, 78%); m.p.: 74 – 76 °C;  $R_f$ (10% MeOH/DCM), 0.37.  $^1\text{H}$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  8.02 (d,  $J = 8.3$  Hz, 2H), 7.66 (d,  $J = 8.4$  Hz, 2H), 7.45 (d,  $J = 8.3$  Hz, 2H), 7.33 (dd,  $J = 7.7, 1.1$  Hz, 1H), 7.25 (d,  $J = 8.4$  Hz, 2H), 7.06 (ddd,  $J = 7.7, 7.3, 1.0$  Hz, 1H), 7.01 (dd,  $J = 8.0, 1.3$  Hz, 1H), 6.96 (ddd,  $J = 8.0, 7.3, 1.1$  Hz, 1H), 5.36 (s, 2H), 4.49 (tt,  $J = 6.8, 4.3$  Hz, 1H), 2.99 – 2.89 (m, 4H), 2.86 – 2.74 (m, 3H), 2.59 (d,  $J = 6.8$  Hz, 1H), 2.46 – 2.34 (m, 4H), 1.85 – 1.79 (m, 1H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  175.43, 167.62, 154.06, 145.66, 142.19, 141.48, 133.92, 132.27 (2C), 129.32 (2C), 127.73 (2C), 127.13 (2C), 121.90, 121.34, 119.79, 118.01, 115.07, 111.06, 107.63, 60.32, 56.79, 52.70, 52.04, 44.32, 34.41, 31.40, 10.00. LC-MS (APCI $^+$ /ESI): found  $m/z = 504.2$  [M+H] $^+$  (cal. For  $\text{C}_{30}\text{H}_{29}\text{N}_7\text{O}$ , 503.24). Purity: 98%,  $t_R = 2.917$  min. Specific rotation,  $[\alpha]^{25}_D = -4.59^\circ$ .

**Methyl 4-(2-(4-((1-methyl-1H-benzo[d]imidazol-2-yl) amino) piperidin-1-yl) ethyl) benzoate (32).** Obtained from **14b** (5.00 g, 21.7 mmol) and **3f** (0.096 g, 6.37 mmol) as a cream white solid (5.78 g, 68%). M.p.: 148 – 149 °C;  $R_f$ (10% MeOH/DCM), 0.46.  $^1\text{H}$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  7.95 (d,  $J$  = 8.3 Hz, 2H), 7.37 (d,  $J$  = 8.3 Hz, 2H), 7.27 (dd,  $J$  = 7.4, 1.3 Hz, 1H), 7.13 (dd,  $J$  = 7.5, 1.6 Hz, 1H), 7.07 – 6.97 (m, 2H), 3.89 (s, 3H), 3.78 (tt,  $J$  = 11.2, 4.2 Hz, 1H), 3.52 (s, 3H), 3.15 – 3.06 (m, 2H), 2.96 – 2.88 (m, 2H), 2.72 – 2.65 (m, 2H), 2.32 (td,  $J$  = 12.1, 2.5 Hz, 2H), 2.17 – 2.09 (m, 2H), 1.69 (dtd,  $J$  = 12.2, 10.2, 3.8 Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz, Methanol- $d_4$ )  $\delta$  167.16, 154.29, 145.88, 141.29, 134.58, 129.35 (2C), 128.57 (2C), 127.95, 120.69, 119.23, 114.39, 106.92, 59.43, 52.31 (2C), 51.11, 49.87, 32.76, 31.59 (2C), 27.16. LC-MS (APCI<sup>+</sup>/ESI): found  $m/z$  = 393.2 [M+H]<sup>+</sup> (cal. For  $\text{C}_{23}\text{H}_{28}\text{N}_4\text{O}_2$ , 392.22). Purity: 98%,  $t_R$  = 0.422 min.

**1,5,6-trimethyl-N-(1-(4-(3-methyl-1,2,4-oxadiazol-5-yl) phenethyl) piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (61).** Obtained from **60c** (0.080 g, 0.31 mmol) and **3d** (0.108 g, 0.37 mmol) as a light brown solid (0.066 g, 48%). m.p.: 120 – 122 °C;  $R_f$ (10% MeOH/DCM), 0.33.  $^1\text{H}$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  8.01 (d,  $J$  = 7.8 Hz, 2H), 7.39 (s, 1H), 7.31 – 7.18 (m, 3H), 3.85 (tt,  $J$  = 11.0, 4.2 Hz, 1H), 3.51 (s, 3H), 3.19 – 3.12 (m, 2H), 2.99 – 2.93 (m, 2H), 2.76 – 2.69 (m, 2H), 2.43 (s, 3H), 2.37 – 2.30 (m, 2H), 2.25 (s, 3H), 2.19 (s, 3H), 2.06 – 1.92 (m, 2H), 1.69 – 1.62 (m, 2H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  176.22, 162.48, 141.73, 139.19, 137.65, 133.83, 132.01, 129.27, 128.76 (2C), 127.88 (2C), 119.19, 116.21, 108.54, 59.44, 56.56, 52.29 (2C), 34.92, 32.73, 31.66 (2C), 19.86, 19.72, 16.80. LC-MS (APCI<sup>+</sup>/ESI): found  $m/z$  = 445.3 [M+H]<sup>+</sup> (cal. for  $\text{C}_{26}\text{H}_{32}\text{N}_6\text{O}$ , 444.26). Purity: 98%,  $t_R$  = 2.301 min.

**Methyl 1-methyl-2-((1-(4-(3-methyl-1,2,4-oxadiazol-5-yl) phenethyl) piperidin-4-yl) amino)-1H-benzo[d]imidazole-5-carboxylate (68).** Obtained from **60h** (0.200 g, 0.69 mmol) and **3d** (0.238 g, 0.83 mmol) as a white solid (0.255 g, 78%). m.p.: 98 – 99 °C;  $R_f$ (10% MeOH/DCM),

0.20. <sup>1</sup>H NMR (400 MHz, Methanol-*d*<sub>4</sub>) δ 8.16 (d, *J* = 0.9 Hz, 1H), 8.02 (d, *J* = 7.9 Hz, 2H), 7.89 (dd, *J* = 7.8, 1.0 Hz, 1H), 7.58 (d, *J* = 7.8 Hz, 1H), 7.36 (d, *J* = 7.9 Hz, 2H), 3.89 (tt, *J* = 11.2, 4.5 Hz, 1H), 3.80 (s, 3H), 3.52 (s, 3H), 3.20 – 3.13 (m, 2H), 2.97 – 2.89 (m, 2H), 2.77 – 2.69 (m, 2H), 2.41 (s, 3H), 2.35 – 2.27 (m, 2H), 2.17 – 2.09 (m, 2H), 1.71 – 1.63 (m, 2H). <sup>13</sup>C NMR (151 MHz, Methanol-*d*<sub>4</sub>) δ 175.90, 167.32, 162.30, 141.55, 139.12, 137.87, 133.64, 128.36 (2C), 127.96 (2C), 125.68, 124.34, 119.35, 116.64, 112.44, 59.51, 56.72, 52.95 (2C), 52.22, 34.72, 32.99, 31.77 (2C), 16.85. LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 475.2 [M+H]<sup>+</sup> (cal. For C<sub>26</sub>H<sub>30</sub>N<sub>6</sub>O<sub>3</sub>, 474.24). Purity: 98%, *t*<sub>R</sub> = 2.245 min.

**General Procedure 13. Synthesis of Compounds 18 – 22.** A mixture of **15** (1.0 equiv), alkyl bromide (1.2 equiv) and K<sub>2</sub>CO<sub>3</sub> (1.5 equiv) in 5 ml DMF was stirred under nitrogen at 70 °C for 12 hours. After cooling to ambient temperature (23 °C), the mixture was diluted with EtOAc (25 ml). The resulting mixture was washed with H<sub>2</sub>O (3 × 30 ml), combined aqueous layers were then extracted with EtOAc (2 × 20 ml). Thereafter, combined EtOAc layers were further washed with 5% LiCl (2 × 10 ml), brine (15 ml), then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude product was triturated with Et<sub>2</sub>O to obtain products. Further purification was performed by flash chromatography (4 – 8% MeOH/DCM) if required.

**1-(Cyclohexyl methyl)-N-(1-(4-(3-methyl-1,2,4-oxadiazol-5-yl) phenethyl) piperidin-4-yl)-1H-benzo[d]imidazol-2-amine (18).** Obtained from **15** (0.080 g, 0.20 mmol) and (bromomethyl)cyclohexane (0.043 g, 0.24 mmol) as a pale-yellow solid (0.080 g, 80%); m.p.: 78 – 80 °C; *R*<sub>f</sub>(10% MeOH/DCM), 0.49. <sup>1</sup>H NMR (600 MHz, Methanol-*d*<sub>4</sub>) δ 8.03 (d, *J* = 8.2 Hz, 2H), 7.46 (d, *J* = 8.2 Hz, 2H), 7.28 (dd, *J* = 7.7, 1.1 Hz, 1H), 7.11 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.02 (ddd, *J* = 7.9, 7.2, 1.3 Hz, 1H), 6.98 (ddd, *J* = 7.7, 7.2, 1.3 Hz, 1H), 3.84 (d, *J* = 7.5 Hz, 2H), 3.81

(tt,  $J = 10.4, 4.0$  Hz, 1H), 3.14 – 3.08 (m, 2H), 2.97 – 2.92 (m, 2H), 2.75 – 2.68 (m, 2H), 2.42 (s, 3H), 2.36 – 2.31 (m, 2H), 2.14 – 2.08 (m, 2H), 1.91 – 1.84 (m, 1H), 1.76 – 1.64 (m, 6H), 1.63 – 1.57 (m, 2H), 1.24 – 1.15 (m, 2H), 1.11 – 1.03 (m, 2H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  176.35, 168.50, 154.86, 146.67, 141.92, 135.24, 130.27, 128.64, 125.33, 122.77, 121.54, 120.06, 115.27, 108.73, 60.14, 53.23 (2C), 50.89, 38.25, 33.62, 32.39 (2C), 31.14, 26.88 (2C), 26.36 (2C), 10.90. LC-MS (APCI $^+$ /ESI): found  $m/z = 499.3$  [M+H] $^+$  (cal. for  $\text{C}_{30}\text{H}_{38}\text{N}_6\text{O}$ , 498.31). Purity: 98%,  $t_{\text{R}} = 2.523$  min.

**General Procedure 14. Synthesis of 1,2,4-oxadiazoles (35 – 41) and Amides (43 – 51).** To a solution of **32** (4.00 g, 10.2 mmol) in MeOH (50 ml) was added 2M aqueous KOH (25 ml, 51 mmol). The reaction mixture was stirred at 79 °C temperature for 2 hr. After completion (monitored by TLC), MeOH was taken off *in vacuo* and the residue diluted with H<sub>2</sub>O. The solution was then acidified to pH 2 under ice with 3M HCl, and the precipitate was filtered and recrystallized in MeOH to afford carboxylic acid (**33**) product as an off-white solid quantitatively. A round bottomed flask containing **33** from the previous step (3.50 g, 9.26 mmol) was charged with thionyl chloride, SOCl<sub>2</sub> (15 ml), and the resulting mixture refluxed at 80 °C for 2 hr. After completion, excess thionyl chloride was evaporated *in vacuo*, and the residue was taken up in 30 ml toluene and evaporated *in vacuo* three (3) times to give acyl chloride **34** in quantitative yield.

An appropriate amine or amidoxime (**31**, 1.2 equiv) was added to a stirring solution containing acyl chloride **34** (0.060 g, 0.151 mmol) and Et<sub>3</sub>N (43  $\mu$ l, 0.303 mmol, 2.0 equiv) in 10 ml dry THF. The resulting mixture was stirred at room temperature (20 °C) for 6 – 10 h. After completion, THF was evaporated *in vacuo*, the residue purified *via* column chromatography to afford amides (**43 – 51**) and *O*-acylamidoximes. Each flask containing crude *O*-acyl amidoximes

was charged with  $K_2CO_3$  (0.042 g, 0.303 mmol, 2.0 equiv) and MeCN was added. The resulting solution was refluxed at 85 °C for 12 hr. Upon completion, the mixture was filtered, and the solvent was taken off *in vacuo*. 1,2,4-oxadiazoles (**35** – **41**) were obtained *via* column chromatography using 7 – 12% MeOH/DCM gradient as eluent.

**N-(1-(4-(3-isopropyl-1,2,4-oxadiazol-5-yl) phenethyl) piperidin-4-yl)-1-methyl-1H-benzo[d]imidazol-2-amine (35)**. Obtained from **31a** (0.019 g, 0.18 mmol) as a light brown solid (0.041 g, 61% over two steps). M.p.: 135 – 137 °C;  $R_f$  (10% MeOH/DCM), 0.42.  $^1H$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  8.06 (d,  $J = 8.2$  Hz, 2H), 7.53 (d,  $J = 8.2$  Hz, 2H), 7.33 (dd,  $J = 7.6, 1.2$  Hz, 1H), 7.11 (dd,  $J = 8.1, 1.2$  Hz, 1H), 7.03 (ddd,  $J = 7.6, 7.2, 1.2$  Hz, 1H), 6.97 (ddd,  $J = 8.1, 7.2, 1.2$  Hz, 1H), 3.80 (tt,  $J = 10.9, 4.3$  Hz, 1H), 3.52 (s, 3H), 3.21 (hept,  $J = 6.8$  Hz, 1H), 3.09 – 3.02 (m, 2H), 2.99 – 2.89 (m, 2H), 2.75 – 2.67 (m, 2H), 2.36 – 2.24 (m, 2H), 2.19 – 2.08 (m, 2H), 1.76 – 1.62 (m, 2H), 1.22 (d,  $J = 6.8$  Hz, 6H).  $^{13}C$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  175.44, 167.09, 155.33, 147.40, 141.09, 133.87, 130.69 (2C), 128.09 (2C), 121.88, 120.99, 120.08, 114.44, 107.28, 60.09, 54.11 (2C), 50.55, 39.85, 32.49, 31.89 (2C), 30.87, 28.76, 21.32 (2C). LC-MS (APCI $^+$ /ESI): found  $m/z = 445.2$   $[M+H]^+$  (cal. For  $C_{26}H_{32}N_6O$ , 444.26). Purity: 98%,  $t_R = 2.192$  min.

**N-(2-hydroxyethyl)-4-(2-(4-((1-methyl-1H-benzo[d]imidazol-2-yl) amino) piperidin-1-yl) ethyl) benzamide (43)**. Obtained from 2-aminoethanol (0.011 g, 0.18 mmol) as an off-white solid (0.052 g, 82%). m.p.: 214 – 216 °C;  $R_f$  (10% MeOH/DCM), 0.15.  $^1H$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  7.90 (d,  $J = 7.9$  Hz, 2H), 7.43 (d,  $J = 7.9$  Hz, 2H), 7.21 (dd,  $J = 8.0, 1.1$  Hz, 1H), 7.10 (dd,  $J = 7.8, 1.3$  Hz, 1H), 6.96 (ddd,  $J = 8.0, 7.5, 1.3$  Hz, 1H), 6.88 (ddd,  $J = 7.8, 7.5, 1.1$  Hz, 1H), 4.19 – 4.12 (dd,  $J = 13.8, 6.9$  Hz, 2H), 4.08 – 4.01 (m, 2H), 3.58 (tt,  $J = 10.8, 4.1$  Hz, 1H), 3.52 (s, 3H), 3.18 – 3.10 (m, 2H), 2.84 – 2.78 (m, 2H), 2.70 – 2.62 (m, 2H), 2.35 – 2.28 (m,

2H), 2.16 – 2.06 (m, 2H), 1.73 – 1.64 (m, 2H). <sup>13</sup>C NMR (151 MHz, Methanol-*d*<sub>4</sub>) δ 168.03, 142.87, 140.48, 138.03, 136.44, 130.28 (2C), 128.92 (2C), 123.00, 120.38, 119.33, 113.74, 105.81, 63.28, 62.44, 60.24, 52.48 (2C), 51.22, 42.87, 34.34, 30.19 (2C). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 422.2 [M+H]<sup>+</sup> (cal. for C<sub>24</sub>H<sub>31</sub>N<sub>5</sub>O<sub>2</sub>, 421.25). Purity: 99%, *t*<sub>R</sub> = 0.138 min.

**General Procedure 15: Hydrolysis of Esters, Synthesis of Carboxylic Acids 42 and 69.** To a solution of the ester in EtOH or MeOH was added 2M aqueous KOH (10 equiv). The reaction mixture was stirred at 80 °C temperature for 2 hours. After completion, EtOH or MeOH was taken off *in vacuo* and the residue diluted with H<sub>2</sub>O. The solution was then acidified to pH 2 under ice with 3N aqueous HCl, and the precipitate was filtered. The product was collected after recrystallization in EtOH.

**5-(4-(2-(4-((1-methyl-1H-benzo[d]imidazol-2-yl) amino) piperidin-1-yl) ethyl) phenyl)-1,2,4-oxadiazole-3-carboxylic acid (42).** Obtained from **41** (0.040 g, 0.084 mmol) in 5 ml EtOH, as an off white crystalline solid (0.036 g, 96%). m.p.: 194 – 196 °C; *R*<sub>f</sub>(10% MeOH/DCM), 0.05. <sup>1</sup>H NMR (600 MHz, Methanol-*d*<sub>4</sub>) δ 8.03 (d, *J* = 8.1 Hz, 2H), 7.46 (d, *J* = 8.1 Hz, 2H), 7.24 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.11 (dd, *J* = 8.0, 1.3 Hz, 1H), 6.97 – 6.83 (m, 2H), 3.82 (tt, *J* = 11.0, 4.4 Hz, 1H), 3.51 (s, 3H), 3.12 – 3.07 (m, 2H), 2.99 – 2.89 (m, 2H), 2.76 – 2.65 (m, 2H), 2.39 – 2.28 (m, 2H), 2.18 – 2.11 (m, 2H), 1.73 – 1.65 (m, 2H). <sup>13</sup>C NMR (151 MHz, Methanol-*d*<sub>4</sub>) δ 176.09, 172.25, 169.36, 147.87, 140.33, 138.47, 134.98, 129.83 (2C), 127.22 (2C), 122.39, 120.19, 119.15, 113.88, 105.22, 60.29, 53.10 (2C), 51.51, 35.87, 34.12, 30.40 (2C). LC-MS (APCI<sup>+</sup>/ESI): found *m/z* = 447.2 [M+H]<sup>+</sup> (cal. for C<sub>24</sub>H<sub>26</sub>N<sub>6</sub>O<sub>3</sub>, 446.21). Purity: 98%, *t*<sub>R</sub> = 0.135 min.

**1-methyl-2-((1-(4-(3-methyl-1,2,4-oxadiazol-5-yl) phenethyl) piperidin-4-yl) amino)-1H-benzo[d]imidazole-5-carboxylic acid (69).** Obtained from **68** (0.200 g, 0.42 mmol) in 10 ml

MeOH, as an off white solid (0.181 g, 94%). m.p.: 134 – 136 °C;  $R_f$ (10% MeOH/DCM), 0.08.  $^1\text{H}$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  8.11 (d,  $J$  = 1.3 Hz, 1H), 8.01 (d,  $J$  = 8.0 Hz, 2H), 7.89 (dd,  $J$  = 7.8, 1.3 Hz, 1H), 7.58 (d,  $J$  = 7.8 Hz, 1H), 7.36 (d,  $J$  = 8.0 Hz, 2H), 3.87 (tt,  $J$  = 10.7, 4.0 Hz, 1H), 3.52 (s, 3H), 3.20 – 3.12 (m, 2H), 2.97 – 2.90 (m, 2H), 2.78 – 2.69 (m, 2H), 2.44 (s, 3H), 2.36 – 2.27 (m, 2H), 2.18 – 2.09 (m, 2H), 1.71 – 1.64 (m, 2H).  $^{13}\text{C}$  NMR (151 MHz, Methanol- $d_4$ )  $\delta$  175.87, 167.22, 161.89, 141.49, 139.19, 137.77, 133.78, 128.44 (2C), 127.10 (2C), 125.86, 124.13, 119.45, 116.48, 112.58, 56.98, 52.88 (2C), 52.29, 34.69, 33.01, 31.65 (2C), 16.98. LC-MS (APCI<sup>+</sup>/ESI): found  $m/z$  = 461.2  $[\text{M}+\text{H}]^+$  (cal. for  $\text{C}_{25}\text{H}_{28}\text{N}_6\text{O}_3$ , 460.22). Purity: 96%,  $t_R$  = 0.135 min.

**7-nitro-N-(2-(2-((1-(4-(5-(trifluoromethyl)-1,2,4-oxadiazol-3-yl) phenethyl) piperidin-4-yl)amino)-1H-benzo[d]imidazol-1-yl)ethyl)benzo[c][1,2,5]oxadiazole 4-amine (23-NBD).** A mixture amine **71** (0.150 g, 0.30 mmol) and  $\text{NaHCO}_3$  (0.076 g, 0.90 mmol) in 5 ml  $\text{H}_2\text{O}$  was stirred at 65 °C for 10 min. A solution of 4-chloro-7-nitrobenzo[c][1,2,5]oxadiazole, NBD-Cl (0.060 g, 0.30 mmol) in MeCN (5 ml) was then added to the heated mixture dropwise *via* syringe. The resulting reaction mixture was stirred at 65 °C for 2 hr, during which a gradual colour change from yellow to dark brown was observed. After completion, the mixture was cooled and MeCN was removed in vacuo. The aqueous residue was then filtered and extracted with EtOAc (3 × 20 ml). Combined organic phases were washed with brine and dried over anhydrous  $\text{Na}_2\text{SO}_4$ . Column chromatography using 8 – 10% MeOH/DCM was performed to obtain the product as a blackish solid (0.145 g, 73%). m.p.: 53 – 54 °C.  $R_f$ (10% MeOH/DCM), 0.51.  $^1\text{H}$  NMR (600 MHz, DMSO- $d_6$ )  $\delta$  8.67 – 8.60 (br-s, 2H), 8.47 (d,  $J$  = 8.7 Hz, 1H), 8.21 (d,  $J$  = 8.7 Hz, 1H), 8.04 (d,  $J$  = 7.9 Hz, 1H), 7.56 (d,  $J$  = 7.9 Hz, 1H), 7.36 (d,  $J$  = 7.5 Hz, 1H), 7.29 – 7.18 (m, 1H), 7.15 – 7.08 (m, 1H), 7.03 – 7.95 (m, 1H), 6.47 (d,  $J$  = 6.2 Hz, 1H), 4.58 (m, 2H),

4.13 – 4.08 (m, 1H), 3.99 – 3.83 (m, 2H), 3.77 – 3.62 (m, 4H), 3.29 – 3.16 (m, 4H), 2.38 – 1.97 (m, 4H). LC-MS (APCI<sup>+</sup>/ESI): found  $m/z = 663.2$  [M+H]<sup>+</sup> (cal. for C<sub>31</sub>H<sub>29</sub>F<sub>3</sub>N<sub>10</sub>O<sub>4</sub>, 662.23). Purity: 98%,  $t_R = 2.658$  min.

**Aqueous Solubility.** Solubility was measured from amorphous solid forms of the compounds using the turbidimetric method. Following the dissolution of test compound in DMSO to make a 10 mM stock solution, a pre-dilution plate was prepared by taking from each stock solution and serially diluting in triplicate to yield concentrations from 0.25 mM to 10.0 mM on a 96 well plate. From each pre-dilution solution, secondary dilutions of the compounds in both DMSO and 0.01M pH 7.4 PBS were prepared in triplicate on a second 96-well plate. Wells in columns 1-6 would contain compound in DMSO, while those in columns 7 – 12 would contain samples in PBS at similar nominal concentrations as those in DMSO. The final volume of solvent in each assay plate was 200  $\mu$ l, prepared by pipetting 4  $\mu$ l each of solution from the pre-dilution plate to the corresponding well into both DMSO and PBS (both 196  $\mu$ L). This ensures that the final concentration of DMSO in the PBS aqueous buffer does not exceed 2% v/v. Similarly, a second secondary plate containing compound concentrations of 60, 100 and 120  $\mu$ M was also prepared. Different concentrations in DMSO were prepared as controls to determine false turbidimetric absorbance readings arising from the compounds in solution absorbing incident radiation at the test wavelength. Following preparation, the assay plate was covered and left to equilibrate for 2 h at 25 °C. Afterwards, UV-vis absorbance readings from the plate were measured at 620 nm using a SpectraMax 340PC<sup>384</sup> microplate reader. Plots of corrected absorbance against compound concentration were computed for a graphical representation of the data using MS Excel. Reserpine and hydrocortisone were used as positive and negative controls, respectively.

**In vitro Antiplasmodium Assay at UCT.** Compounds were tested using parasite lactate dehydrogenase assay as a marker for parasite survival. Briefly, the respective stock solutions of CQ diphosphate and test compounds were prepared to 2 mg.ml<sup>-1</sup> in distilled water (for CQ) and 100% DMSO for test compounds and then stored at -20 °C, and further dilutions were prepared on the day of the experiment. The cultures were synchronized in the ring stage using 15 ml of 5% (w/v) D-sorbitol in water. Synchronous cultures of *Pf*NF54 (CQ-S) and *Pf*K1 (MDR) in the late trophozoite stage were prepared to 2% parasitemia & 2% hematocrit. Compounds were tested at starting concentrations of 10,000 ng.ml<sup>-1</sup> (1000 ng.ml<sup>-1</sup> for CQ), which were then serially diluted 2-fold in complete medium to give 10 concentrations with a final volume of 200 µl in each well. Parasites were incubated in the presence of the compounds at 37 °C under hypoxic conditions (4% CO<sub>2</sub> and 3% O<sub>2</sub> in N<sub>2</sub>) for 72 h. After incubation, 100 µl of MalStat reagent and 15 µl of resuspended culture were combined, followed by addition of 25 µl of nitro blue tetrazolium chloride (NBT). The plates were kept in the dark for 10 min in order to fully develop, after which absorbance was measured at 620 nm on a microplate reader. Raw data was processed using GraphPad Prism 4.0 (La Jolla, California, USA) to analyze the dose-response.

**In vitro Antiplasmodium Assay at Swiss TPH.**

The testing was performed with the modified [<sup>3</sup>H]-hypoxanthine incorporation assay, as previously reported.<sup>34</sup>

**In vitro Cytotoxicity Assay.** Compounds were screened against Chinese Hamster Ovarian (CHO) mammalian cell lines, using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) assay.<sup>35</sup> Emetine was used as the reference standard. It was prepared to 2 mg/ml in distilled water, while the stock solutions of test compounds were prepared to 20 mg/ml in DMSO (100%), with the highest concentration of solvent to which the

cells were exposed having no measurable effect on the cell viability. The initial concentration of the compounds and control was 100  $\mu\text{g/ml}$ , which was serially diluted in complete medium with 10-fold dilutions to give 6 concentrations, the lowest being 0.001  $\mu\text{g/ml}$ . Plates were incubated for 48 h with 100  $\mu\text{l}$  of test compound and 100  $\mu\text{l}$  of cell suspension in each well and developed afterward by adding 25  $\mu\text{l}$  of sterile MTT (Thermo Fisher Scientific) to each well, followed by 4 h of incubation in the dark. The plates were then centrifuged, the medium aspirated, and 100  $\mu\text{l}$  of DMSO was added to dissolve crystals before reading the absorbance at 540 nm. Data were analyzed, and the sigmoidal dose–response was derived using GraphPad Prism version 4.0 software. All experiments were performed as three independent biological repeats, each with technical triplicates.

**In vitro Metabolic Stability Assay.** This assay was performed in duplicate using a 96-well microtiter plate. Test compounds (0.1  $\mu\text{M}$ ) were incubated at 37 °C in mouse and pooled human liver microsomes with a final protein concentration of 0.4  $\text{mg}\cdot\text{ml}^{-1}$ ; XenoTech, Lenexa, KS suspended in 0.1 M phosphate buffer at pH 7.4 for predetermined time points. This was in the presence and absence of cofactor-reduced nicotinamide adenine dinucleotide phosphate (NADPH, 1.0 mM). The reactions were quenched by adding ice-cold MeCN containing an internal standard (Carbamazepine, 0.0236  $\mu\text{g/ml}$ ). The samples were centrifuged, and the supernatant was analyzed via liquid chromatography-tandem mass spectrometry (LC–MS/MS) (Agilent Rapid Resolution HPLC, AB SCIEX 4500 MS). The relative loss of the parent compound with time was monitored, and plots were prepared for each compound of  $\text{Ln}\%$  remaining versus time to determine the first-order rate constant for compound depletion. This was used to calculate the degradation half-life and subsequently to predict the *in vitro* intrinsic clearance ( $\text{CL}_{\text{int}}$ ) and *in vitro* hepatic extraction ratio ( $E_{\text{H}}$ ).<sup>36</sup>

### ***In vivo* Antiplasmodium Assay at Swiss TPH.**

*In vivo* efficacy was assessed as previously described,<sup>37</sup> with the modification that mice (n = 3) were infected with a GFP-transfected *P. berghei* ANKA strain (donated by A. P. Waters and C. J. Janse, Leiden University, The Netherlands), and parasitemia was determined using standard flow cytometry techniques. The detection limit was 1 parasite in 1000 erythrocytes (that is, 0.1%). The activity was calculated as the difference between the mean percent parasitemia for the control and treated groups expressed as a percent relative to the control group. Compounds were dissolved or suspended in a vehicle consisting of 70% Tween-80 and 30% ethanol, followed by a 10-fold dilution in H<sub>2</sub>O and oral administration as four consecutive daily doses (4, 24, 48, and 72 h after infection). Blood samples for the quadruple-dose regimens were collected on day 4 (96 h after infection). The survival time in days was also recorded up to 30 days after infection. A compound was considered curative if the animal survived to day 30 after infection with no detectable parasites by slide reading.

*In vivo* studies conducted at the Swiss TPH, Basel were approved by the veterinary authorities of the Canton Basel-Stadt (Permit No. 1731 and 2303) based on Swiss Cantonal (Verordnung Veterinäramt Basel-Stadt) and National Regulations (The Swiss Animal Protection Law, Tierschutzgesetz).

**Fluorescence Live-cell Imaging.** A Nunc Lab-Tek II 8-well chamber slide (Thermo Fisher Scientific) with no. 1.5 cover glass was coated with a 150 µl poly-l-lysine solution for 10 min after which the excess solution was removed, and the chamber slide was air-dried. 5 µl of human erythrocytes infected with *P. falciparum* chloroquine-sensitive strain (NF54) were resuspended in 5 mL Ringer's solution (pH 7.5). 100 µl aliquot of the suspended cells was placed in each well of the chamber slide and incubated for 30 min to allow the cells to adhere to the glass chamber.

Excess Ringer's solution was removed alongside all non-adhering parasitized red blood cells. The cells were washed twice with Ringer's solution after which a new solution containing the appropriate concentration of drugs and organelle markers was added. SIM-SR microscopy was performed using a Elyra 7 with Lattice SIM2. A Plan-Apochromat 63×/1.40 Oil DIC M27 objective lens was used to keep laser transmission as low as possible to prevent phototoxicity to the cells. Images were captured and processed using Zeiss ZEN software (Carl Zeiss Microscopy GmbH).

## **ASSOCIATED CONTENT**

### **Supporting Information**

Information not included in this paper is available in the Supporting Information, which is available and free of charge on the ACS Publications website at DOI: (PDF).

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### **Notes**

The authors declare no competing material or financial interest whatsoever.

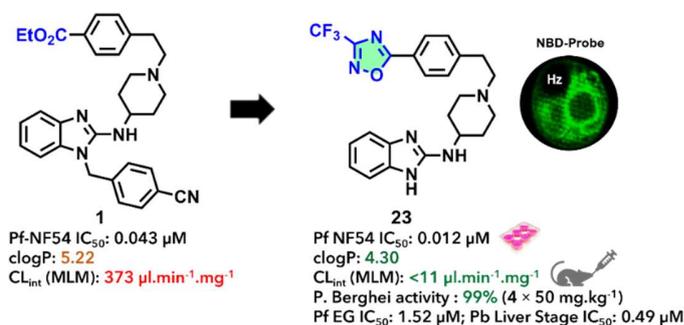
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## ABBREVIATIONS USED

AST, astemizole; DMAST, desmethylastemizole; hERG, human *ether-a-go-go*-related gene; EDCI, 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide; DMAP, 4-Dimethylaminopyridine; CDI, carbonyldiimidazole; DCC, N,N'-Dicyclohexylcarbodiimide; TCDI, 1,1'-Thiocarbonyldiimidazole; DMF, N,N-dimethylformamide; TFA, trifluoroacetic acid anhydride; PTSA, para-toluene sulfonic acid; *P. falciparum*, *Plasmodium falciparum*; *P. berghei*, *Plasmodium berghei*; SI, selectivity index; CQ, chloroquine; AQ, amodiaquine; CHO, Chinese hamster ovarian; ACT, artemisinin combination therapy; MSD, mean survival days; PK, pharmacokinetics; SAR, structure–activity relationship;  $I_{Kr}$ , potassium ion current;  $t_R$ , retention time;  $R_f$ , retardation factor; m.p, melting point;  $E_H$ , extraction ratio;  $IC_{50}$ , concentration of a drug that is required for 50% inhibition *in vitro*.

## Table of Contents Graphic:



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