

## **SUPPLEMENTARY MATERIAL**

### ***Ethics Statement***

All study participants provided written informed consent and participated in research or clinical studies at the host institute under ethically approved protocols. The studies and their approving institutes are: Australian site of the Breast Cancer Family Registry (BCFR-AU) - The University of Melbourne Health Sciences Human Ethics Sub-Committee; Northern California site of the Breast Cancer Family Registry (BCFR-NC) - Northern California Cancer Center Institutional Review Board; New York site of the Breast Cancer Family Registry (BCFR-NY) - Columbia University Medical Center Institutional Review Board; Ontario site of the Breast Cancer Family Registry (BCFR-ON) - Mount Sinai Hospital Research Ethics Board; Philadelphia site of the Breast Cancer Family Registry (BCFR-PA) - Institutional Review Board Fox Chase Cancer Center; Utah site of the Breast Cancer Family Registry (BCFR-UT) - Institutional Review Board University of Utah; Baltic Familial Breast and Ovarian Cancer Consortium (BFBOCC) - Centrālā medicīnas ētikas Komiteja; Lietuvos Bioetikos Komitetas; BRCA-gene mutations and breast cancer in South African women (BMBSA) - University of Pretoria and Pretoria Academic Hospitals Ethics Committee; Beckman Research Institute of the City of Hope (BRICOH) - City of Hope Medical Center Institutional Review Board; Copenhagen Breast Cancer Study (CBCS) - De Videnskabsetiske Komiteer i Region Hovedsladen; Spanish National Cancer Centre (CNIO) - Instituto de Salud Carlos III Comité de Bioética y Bienestar Animal; City of Hope Cancer Center (COH) - City of Hope Institutional Review Board; CONsorzio Studi ITaliani sui Tumori Ereditari Alla Mammella (CONSIT TEAM) - Comitato Etico Indipendente della Fondazione IRCCS "Istituto Nazionale dei Tumori"; National Centre for Scientific Research Demokritos (DEMOKRITOS) - Bioethics committee of NCSR "Demokritos", 240/EHΔ/11.3; National Centre for Scientific Research Demokritos (DEMOKRITOS) - Papageorgiou Hospital Ethics Committee; Dana Farber Cancer Institute (DFCI) - Dana Farber Cancer Institute Institutional Review Board; Deutsches Krebsforschungszentrum (DKFZ) - Ethik-Kommission des

Klinikums der Universität; Deutsches Krebsforschungszentrum (DKFZ) - Hospital Universitario de San Ignacio Comité de Investigaciones y Etica; Deutsches Krebsforschungszentrum (DKFZ) - Shaukat Khanum Memorial Cancer Hospital and Research Centre Institutional Review Board; Epidemiological study of BRCA1 and BRCA2 mutation carriers (EMBRACE) - Anglia & Oxford MREC; Fox Chase Cancer Center (FCCC) - Institutional Review Board Fox Chase Cancer Center; Fundación Pública Galega de Medicina Xenómica - Comité Autonómico de Etica da Investigación de Galicia; German Consortium of Hereditary Breast and Ovarian Cancer (GC-HBOC) - Ethik-Kommission der Medizinischen Fakultät der Universität zu Köln; Genetic Modifiers of cancer risk in BRCA1/2 mutation carriers (GEMO) - Comité consultatif sur le traitement de l'information en matière de recherche dans le domaine de la santé; Georgetown University (GEORGETOWN) - MedStar Research Institute - Georgetown University Oncology Institutional Review Board; Ghent University Hospital (G-FAST) - Universitair Ziekenhuis Gent - Ethics Committee; Hospital Clinico San Carlos (HCSC) - Comité Ético de Investigación Clínica Hospital Clínico San Carlos; Helsinki Breast Cancer Study (HEBCS) - Helsingin ja uudenmaan sairaanhoitopiiri (Helsinki University Central Hospital ethics committee); Hereditary Breast and Ovarian study Netherlands (HEBON) - Protocol Toetsingscommissie van het Nederlands Kanker Instituut/Antoni van Leeuwenhoek Ziekenhuis; Molecular Genetic Studies of Breast- and Ovarian Cancer in Hungary (HUNBOCS) - Institutional Review Board of the Hungarian National Institute of Oncology; University Hospital Vall d'Hebron (HVH) - The Hospital Universitario Vall d'Hebron Clinical Research Ethics Committee; Institut Català d'Oncologia (ICO) - Catalan Institute of Oncology Institutional Review Board; International Hereditary Cancer Centre (IHCC) - Komisji Bioetycznej Pomorskiej Akademii Medycznej (Pomeranian Medical University Bioethics Committee); Iceland Landspítali - University Hospital (ILUH) - Vísindasíðanefnd National Bioethics Committee; Interdisciplinary Health Research International Team Breast Cancer Susceptibility (INHERIT) - Comité d'éthique de la recherche du Centre Hospitalier Universitaire de Québec; Istituto Oncologico Veneto Hereditary Breast and Ovarian Cancer Study (IOVHBOCS) - Centro Oncologico Regionale

Azienda Ospedale Di Padova Comitato Etico; Portuguese Oncology Institute-Porto Breast Cancer Study - COMISSÃO DE ÉTICA PARA A SAÚDE (CES) ; Kathleen Cuningham Foundation Consortium for Research into Familial Breast Cancer (KCONFAB) - Queensland Institute of Medical Research - Human Research Ethics Committee; Kathleen Cuningham Foundation Consortium for Research into Familial Breast Cancer (KCONFAB) - Peter MacCallum Cancer Centre Ethics Committee; University of Kansas Medical Center(KUMC) - The University of Kansas Medical Center Human Research Protection Program; Mayo Clinic (MAYO) - Mayo Clinic Institutional Review Boards; McGill University (MCGILL) - McGill Faculty of Medicine Institutional Review Board; Modifier Study of Quantitative Effects on Disease (MOD-SQUAD) - Mayo Clinic Institutional Review Boards; Memorial Sloane Kettering Cancer Center (MSKCC) - Human Biospecimen Utilization Committee; Memorial Sloan Kettering Cancer Center (MSKCC) - Memorial Sloan-Kettering Cancer Center IRB; General Hospital Vienna (MUV) - Ethikkommission der Medizinischen Universität Wien; Women's College Research Institute Hereditary Breast and Ovarian Cancer Study - University of Toronto Health Sciences Review Ethics Board; National Cancer Institute (NCI) - NIH Ethics Office; National Israeli Cancer Control Center (NICCC) - Carmel Medical Center Institutional Review Board (Helsinki Committee); N.N. Petrov Institute of Oncology (NNPIO) - N.N. Petrov Institutional Ethical Committee; NorthShore University HealthSystem (NORTHSHORE) - Institutional Review Board of NorthShore University HealthSystem; NRG Oncology (NRG\_ONCOLOGY) - Cancer Prevention and Control Protocol Review Committee; Ontario Cancer Genetics Network (OCGN) - University Health Network Research Ethics Board; The Ohio State University Comprehensive Cancer Center (MACBRCA) - The Ohio State University Cancer Institutional Review Board; Odense University Hospital (OUH) - Den Videnskabsetiske Komité for Region Syddanmark; Pisa Breast Cancer Study (PBCS) - Azienda Ospedaliera Pisana Comitato Etico per lo studio del farmaco sull'uomo; Sheba Medical Centre - Chaim Sheba Medical Center IRB; Swedish Breast Cancer Study (SWE-BRCA) - Regionala Etikprövningsnämnden Stockholm; University of Chicago (UCHICAGO) - The University of Chicago Biological Sciences Division

Institutional Review Board (BSD IRB); University of California Los Angeles (UCLA) - UCLA Institutional Review Board (UCLA IRB); University of California San Francisco (UCSF) - Human Research Protection Program Institutional Review Board (IRB); UK and Gilda Radner Familial Ovarian Cancer Registries (UKGRFOCR) - Roswell Park Cancer Institute IRB; UK and Gilda Radner Familial Ovarian Cancer Registries (UKGRFOCR) - Cambridge Local Research Ethics Committee; University of Pennsylvania (UPENN) - University of Pennsylvania Institutional Review Board; Cancer Family Registry University of Pittsburgh (UPITT) - University of Pittsburgh Institutional Review Board; University of Texas MD Anderson Cancer Center (UTMDACC) - University of Texas MD Anderson Cancer Center Office of Protocol Research Institutional Review Board; Victorian Familial Cancer Trials Group (VFCTG) - Peter MacCallum Cancer Centre Ethics Committee; Women's Cancer Program at Cedars-Sinai Medical Center (WCP) - (Cedars-Sinai Medical Center) CSMC Institutional Review Board.

### ***Genotyping and SNP imputation***

Genotyping was performed on one of two bespoke SNP arrays. The majority of the samples were genotyped using the OncoArray (15,679 *BRCA1* and 10,981 *BRCA2* carriers)<sup>1-3</sup>. The OncoArray is a custom genotyping array comprising approximately 533,000 SNPs, including a GWAS backbone component tagging common SNPs across the genome which accounted for approximately half of the SNPs on the array. The remaining 3,256 (17.2%) *BRCA1* and 1,358 (11.0%) *BRCA2* pathogenic variant carriers were genotyped using the iCOGS array<sup>4,5</sup> which included approximately 210,000 SNPs selected primarily on the basis of evidence of association with breast, ovarian and prostate cancers.

A standard quality control (QC) process was applied for samples genotyped on both arrays, which included assessment of the SNP call rate, allele frequency, genotyping intensity clustering metrics, Hardy-Weinberg equilibrium and SNP concordance in duplicate samples<sup>2</sup>. Two-stage imputation was performed using SHAPEIT software<sup>6</sup> for phasing and IMPUTE2 software<sup>7</sup> for imputation using the 1000 Genomes Project (Phase 3) reference

panel.

SNPs were included in the PRS if they were adequately imputed in the CIMBA data. The imputation accuracy was assessed using the  $r^2$  statistic, based on the “info” statistic produced by the IMPUTE2 software ([https://mathgen.stats.ox.ac.uk/impute/impute\\_v2.html#info\\_metric\\_details](https://mathgen.stats.ox.ac.uk/impute/impute_v2.html#info_metric_details))<sup>7</sup>. This statistic takes values from 0 (complete uncertainty of imputed genotypes) to 1 (no uncertainty of imputed genotypes). The  $r^2$  values for the SNPs used in the current analyses are listed in Tables S1 and S2 and shown in Figures S1 and S2. The minimum  $r^2$  values among the 313 SNPs in the breast cancer PRS were 0.49 for samples genotyped on iCOGS and 0.90 for samples genotyped on OncoArray. For the 30 SNPs in the ovarian cancer PRS, the minimum  $r^2$  was 0.64 for iCOGS samples and 0.88 for OncoArray samples.

### **Principal components analysis**

To adjust for potential (intra-continental) population stratification in the OncoArray dataset, principal components analysis was performed using data from 33,661 uncorrelated SNPs (which included 2,318 SNPs specifically selected on informativeness for determining continental ancestry) with a MAF of at least 0.05 and maximum correlation of 0.1 in the OncoArray dataset, using purpose-written software (<http://ccge.medschl.cam.ac.uk/software/pccalc>). A similar approach was used for the iCOGS dataset.

### ***Breast cancer and epithelial ovarian cancer PRS***

PRSs were constructed as the weighted sum of alleles for 313 SNPs for breast cancer and 30 SNPs for epithelial ovarian cancer (EOC), thus the PRS for each participant,  $i$ , was calculated as:

$$PRS_i = \sum_{j=1}^N \beta_j g_{ij}$$

where  $g_{ij}$  is the genotype or imputed dosage for variant  $j$  observed for individual  $i$  and  $\beta_j$  is

weight for the  $j^{\text{th}}$  SNP.

The weights for the breast cancer PRS were the log Odds Ratio (log-OR) estimates of association used to construct the 313 SNP PRS based on data from the general population and reported by Mavaddat et al<sup>8</sup>. The weights used to construct the PRS for overall breast cancer (denoted as PRS<sub>BC</sub>), ER-negative breast cancer (PRS<sub>ER-</sub>) or ER-positive breast cancer (PRS<sub>ER+</sub>) are shown in Supplementary Table 1).

Two PRS for epithelial ovarian cancer (EOC) were constructed:

1. A PRS for invasive EOC (PRS<sub>EOC</sub>) based on 30 SNPs which were: (i) associated with EOC; or (ii) identified through pleiotropic GWAS of breast, EOC and prostate cancer<sup>3,9</sup> at genome-wide significance levels in the combined analyses of the three cancers, but also showed consistent associations with EOC in the Phelan et al<sup>3</sup>.
2. A PRS for high grade serous (HGS) ovarian cancer. As HGS is the predominant subtype observed in *BRCA1* and *BRCA2* pathogenic variant carriers<sup>10</sup> a 22 SNP high-grade serous EOC PRS (PRS<sub>HGS</sub>) was constructed. This PRS was restricted to SNPs that exhibited associations at genome-wide significance level ( $P < 5 \times 10^{-8}$ ) with any EOC histotype, was nominally associated ( $P < 0.05$ ) with HGS EOC, and the direction of the association for HGS EOC was consistent with the EOC association<sup>3</sup>.

The SNPs and the corresponding log-OR weights used in the PRS<sub>EOC</sub> and PRS<sub>HGS</sub> are shown in Supplementary Table 2.

### ***Calculating the theoretical PRS***

The theoretical PRS distribution under a multiplicative model was used in comparisons against the PRS percentile specific association estimates. For the theoretical PRS, the variance attributable to SNP  $i$  was given by:

$$V_i = (1 - p_i)^2 E_i^2 + 2p_i(1 - p_i)(\beta_i - E_i)^2 + p_i^2(2\beta - E_i)^2$$

where  $E_i$  is the expected value of  $\beta$ , given by:

$$E_i = 2p_i(1 - p_i)\beta_i + 2p_i^2\beta_i$$

where  $\beta_i$  is the per-allele log-OR and  $p_i$  is the allele frequency for SNP  $i$  and were obtained from the population-data used in the PRS construction for breast and ovarian cancer (Tables S1 and S2)<sup>3,8</sup>. The mean PRS is then given by:

$$\overline{PRS} = \sum_{i=1}^N E_i$$

and the theoretical PRS variance is given by:

$$V = \sum_{i=1}^N V_i$$

The allele frequencies were obtained from the 1000 Genomes Project European ancestry samples. The theoretical HRs at each percentile were calculated assuming the PRS is normally distributed with mean  $\overline{PRS}$  and variance  $V$  (i.e. the HRs were log-normally distributed).

### **Description of statistical models**

#### *Weighted cohort analysis*

The retrospective cohort association analyses were undertaken using weighted Cox regression models<sup>12</sup>. These analyses accounted for the non-random sampling of *BRCA1* and *BRCA2* carriers with respect to their disease (breast cancer and ovarian cancer) status. In such retrospective studies, affected carriers tend to be oversampled because *BRCA1* and *BRCA2* testing is targeted to affected individuals who may also be diagnosed at an early age. Therefore, the carriers in CIMBA retrospective study do not represent a true cohort of *BRCA1* and *BRCA2* carriers. We have previously shown that under these conditions, standard Cox regression analysis leads to biased estimates of the rate ratios<sup>12,13</sup>. To correct for this bias, we used the weighted cohort approach<sup>12,13</sup>. Briefly, this method involves assigning different weights to cancer cases and unaffected individuals which are age- and gene-specific, such that the weighted observed incidence rates are consistent with established incidence rates for carriers of pathogenic variants in *BRCA1* and *BRCA2*<sup>14</sup>. This

approach has been shown in simulation studies to yield unbiased estimates of association<sup>12,13</sup>. The weighted cohort analysis was carried out in R “survival” library command `coxph(model, robust=TRUE, weights=w)` where `w` represents the age specific weights.

### *Model comparisons*

Likelihood ratio tests (LRTs) were undertaken to determine whether the models which include interaction terms (age-varying PRS, PRS interaction with gene variant location and PRS interaction with gene variant class) fitted data better than the nested model that did not include the interaction term. Here we considered two models: (i) a model that includes the PRS interaction term, with a corresponding log-likelihood,  $L_I$  and the nested model without the interaction term with log-likelihood  $L_N$ . Hence, the LRT comparing these models has the form:

$$-2[L_N - L_I] \sim \chi_{\Delta d}^2$$

where  $\Delta d$  denotes degrees of freedom, given by the difference in number of parameters estimated between the two models.

### ***BRCA1 and BRCA2 Cohort Consortium (BBCC) prospective cohort***

The BBCC included data from the International *BRCA1/2* Carrier Cohort Study (IBCCS), Breast Cancer Family Registry (BCFR) and the Kathleen Cunningham Foundation Consortium for Research into Familial Breast Cancer (kConFab) IBCCS study participants were recruited between 1997 and 2011 from 18 European cancer genetics centres and Quebec, Canada. Most women were recruited through large national studies in the United Kingdom, Netherlands and France. All centres actively followed participants through questionnaires. Additionally, where possible, passive follow-up by pathology (Denmark and Netherlands), cancer and death registry linkage (Denmark, Netherlands, Sweden and the United Kingdom), and validation of self-reported cancer diagnoses and preventive surgeries



through medical records.

The BCFR is a family cohort recruited from six sites from Australia, Canada and the USA. The families were followed-up regularly by annual contact of probands and systematic 5-year follow-up of families that collected demographic and epidemiological data from all study participants.

The kConFab recruited pathogenic variant carriers from multi-case families that had been ascertained since 1997 by family cancer clinics in Australia and New Zealand. kConFab study participants were independent from BCFR participants from Australia. Study participants were systematically followed by the kConFab Follow-Up Study<sup>15</sup> using mailed questionnaires every three years, with self-reported cancers and prophylactic surgeries confirmed from medical records. BBCC follow-up ended in December 2013<sup>16</sup>.

### ***Association analysis in prospective cohorts***

To assess associations between the PRS and breast cancer risk, eligibility was restricted to female *BRCA1* and *BRCA2* carriers who at completion of the baseline questionnaire were free of any cancer diagnosis (excluding non-melanoma skin cancer) and had not undergone risk-reducing bilateral mastectomy. Study participants were followed from baseline until the first of: (i) age 80-years; (ii) death; (iii) completion of last follow-up questionnaire or last record linkage, whichever occurred last; (iv) risk-reducing bilateral mastectomy; or (v) diagnosis of any first cancer (apart from non-melanoma skin cancer). Participants diagnosed with a first breast cancer were considered affected.

To assess associations between the PRS and ovarian cancer risk, eligibility was restricted to women who had not been diagnosed with ovarian cancer and had not had RRSO at the time of baseline questionnaire completion. To maximise statistical power, carriers with a prior breast cancer or non-melanoma skin cancer diagnosis were retained for the prospective ovarian cancer analyses, but carriers with prior diagnoses of other cancers were excluded, in line with previous prospective studies of ovarian cancer risk for *BRCA1/2* pathogenic variant carriers<sup>16</sup>. Participants were followed from baseline until the first of: (i)

age 80-years; (ii) death; (iii) completion of last follow-up questionnaire or record linkage, whichever happened last; (iv) bilateral RRSO, or bilateral salpingectomy, or removal of both ovaries for any other reason; or (v) any cancer diagnosis (except breast or non-melanoma skin cancer). Carriers diagnosed with invasive ovarian, fallopian tube, or peritoneal cancer during the follow-up were considered affected.

Associations using the harmonised prospective cohorts were analysed using Cox regression, separately for *BRCA1* and *BRCA2* carriers. Statistical models were stratified by consortia (CIMBA or BBCC), birth cohort, country, and Ashkenazi Jewish ancestry, adjusted for family history of the appropriate cancer in first- and second-degree relatives. Robust variance estimates were calculated considering family membership.

### ***Calculating absolute cancer risks by PRS***

Breast cancer absolute risks were calculated by PRS category and also by PRS category in combination with variant locations in *BRCA1* and *BRCA2*, and in combination with family history of breast cancer (absence or presence of family history).

For these calculations we assumed external estimates of overall breast cancer incidence and breast cancer incidence estimates for different pathogenic variant locations and cancer family history status. The external estimates were obtained from previously published prospective penetrance studies in *BRCA1* and *BRCA2* pathogenic variant carriers<sup>16</sup>. For all analyses, to obtain the breast cancer incidences for each PRS percentile category, we constrained the breast cancer incidences over all PRS categories to agree with the prospectively estimated breast cancer incidence rates using the constraining approach described previously<sup>8,16-18</sup>. In this we assume that the breast cancer incidence for someone in PRS category  $c$  is given by  $\lambda_0(t)\exp(\beta_c)$  where  $\lambda_0(t)$  is the baseline incidence (for those in the baseline PRS category) which is unknown, and  $\beta_c$  is the corresponding log-HR of association with breast cancer risk for a carrier in category  $c$  relative to the baseline category. Given this constraining, it was previously shown<sup>16,17</sup> that  $\lambda_0(t)$  is given by:

$$\lambda_0(t) = i(t) \frac{\sum_c \tau_c S_c(t-1)}{\sum_c \tau_c \exp(\beta_c) S_c(t-1)}$$

where  $i(t)$  is the assumed external incidence (i.e. the average over all PRS effects),  $\tau_c$  is the proportion of carriers in PRS category  $c$  and  $S_c(t)$  is the probability of surviving the disease to age  $t$  in PRS category  $c$ .  $\lambda_0(t)$  can be calculated iteratively assuming  $S_c(0)=1$  over the ages  $t$ . Once  $\lambda_0(t)$  was calculated, the incidence for each PRS category is given by:  $\lambda_0(t)\exp(\beta_c)$ . This process was carried out assuming the external incidence estimates for overall breast cancer, incidences by pathogenic variant location or by family history separately<sup>16</sup>.

### **Calculating 10-year cancer risks**

The 10-year risk of developing breast or ovarian cancer at age  $t$  was calculated as the risk difference between ages  $(t+10)$  and  $t$ , conditional on not developing cancer up to age  $t$ .

Mathematically this can be written as:

$$R(t)_{10} = \frac{P(t+10) - P(t)}{1 - P(t)}$$

where  $R(t)_{10}$  is the 10-year risk and  $P(t)$  is the cumulative disease risk at age  $t$  and is calculated using the PRS specific incidences calculated in the previous section.

## **Supplementary Results**

### **Absolute risks by PRS, variant location and family history (results)**

Carriers of pathogenic variants in the non-central gene regions had greater risk of developing breast cancer (5th-95th PRS percentiles *BRCA1* 5' end 61%-88%, 3' end 60%-91%; *BRCA2* (narrow) 5' end 62%-87%, 3' end 60%-92%; *BRCA2* (wide) 5' end 67%-91%, 3' end 61%-93%) compared to carriers with variants in the central regions (*BRCA1* 49%-75%; *BRCA2* (narrow) 42%-73%; *BRCA2* (wide) 41%-71%) (Table S5; Figures S5-S7).

Carriers with a family history of breast cancer (at least one affected first or second degree relative) had larger absolute risks of developing breast cancer up to age 80-years (*BRCA1* 65%-88%; *BRCA2* 62%-85%) compared with carriers without a family history of breast cancer (*BRCA1* 46%-71%; *BRCA2* 62%-85%) (Table S5; Figures S8-S9).

### ***Detailed breast and ovarian cancer absolute risks by PRS percentiles (results)***

Table S6 illustrates the absolute risks by age 80-years of developing breast cancer and ovarian cancer for pathogenic variant carriers. These absolute risks are presented for the PRS deciles as well as the most extreme first (i.e. 1st and 99th) and fifth (i.e. 5th and 95th) PRS percentiles. The PRS<sub>ER</sub> is presented for *BRCA1* carriers with respect to their breast cancer risk, whilst the PRS<sub>BC</sub> is shown for *BRCA2* carrier breast cancer risk. The PRS<sub>HGS</sub> is presented for both *BRCA1* and *BRCA2* carriers with respect to ovarian cancer risks.

### **Statistical software (R and Stata) commands used for statistical analyses**

#### **R Cox regression**

```
library(survival)
coxph(Surv(CENSORING.AGE, CENSORING.STATUS) ~ strata(STRATA) +
cluster(FAMILY) + BIRTH.COHORT + PRINCIPAL.COMPONENTS +
NORMALISED.PRS, _robust = TRUE, weights = WEIGHTS, data=DATA)
```

#### **Stata age-varying PRS analysis**

```
stset CENSORINGAGE [pweight = WEIGHTS], id(ID) f(CENSORINGSTATUS)
xi: stcox NORMALISEDPRS i.BIRTHCOHORT PRINCIPALCOMPONENTS*, ///
strata(STRATA) cluster(FAMILY) tvc(NORMALISEDPRS)
// PRINCIPALCOMPONENTS* represents all principal components
```

#### **Stata: C-index for discrimination**

```
net from http://www.homepages.ucl.ac.uk/~rmjwiww/stata/epi
net install cindex
* Fit appropriate Cox model and obtain linear predictions
stcox ...
predict LINPRED, xb
set seed 25456
bootstrap c = r(C_adj_correct2), cluster(FAMILY) reps(1000): cindex
LINPRED, ///
strata(STRATA) adj(_IBIRTHCOHORT* PRINCIPALCOMPONENTS*)
// PRINCIPALCOMPONENTS* represents all principal components
// _IBIRTHCOHORT* represents all birth cohorts (created internally
from the "BIRTHCOHORT" variable by Stata after fitting the -stcox-
model)
```

## **SUPPLEMENTARY TABLE LEGENDS**

### **Table S1**

The 313 SNPs used to construct the breast cancer PRS<sup>8</sup>. The same set of 313 SNPs was used to construct the PRS<sub>ER-</sub> and PRS<sub>ER+</sub>. The ER-specific PRS used different SNP weights (log-ORs for ER-specific breast cancer) if they had a statistically significant different effect on ER-subtype from a population-based breast cancer case-only analysis.

### **Table S2**

The 30 SNPs used to construct the ovarian cancer PRS. The 22 SNPs used to form the high-grade serous ovarian cancer PRS are highlighted in grey. The high-grade serous specific PRS was limited to SNPs that showed genome-wide statistical significance ( $P < 5 \times 10^{-8}$ ) with any of the ten ovarian cancer subtypes, had concordant direction of effects between overall all invasive and high-grade serous disease, and exhibited nominal statistical significance ( $P < 0.05$ ) with high-grade serous ovarian cancer<sup>3</sup>. The “overall” and “high-grade serous” ovarian cancer (per-allele) effect sizes and P-values were taken from<sup>3</sup> and/or<sup>9</sup>.

### **Table S3**

Retrospective cohort characteristics for 18,935 *BRCA1* and 12,339 *BRCA2* carriers recruited by the CIMBA. Breast cancer and ovarian cancer refer to the first cancer diagnosis.

Censoring ages are reported in years. Pathogenic variant classes: I = unstable or no protein; II = stable mutant protein; III = consequence unknown. Pathogenic variant locations are in base pairs (bp) within the *BRCA1* and *BRCA2* genes. ER-status is oestrogen receptor status of the breast tumour. Cancer family history is reported for the relevant cancer from first and second degree relatives. “Unknown” family history = reported unknown cancer family history, “missing” family history = family history data not collected. IQR = interquartile range; SD = standard deviation.

### **Table S4**

Validation data summary statistics from prospective cohorts (CIMBA and BBCC). Validation data are presented for the breast cancer PRS and ovarian cancer PRS by disease status at censoring. The  $PRS_{ER-}$  is reported for *BRCA1* carriers, whilst the  $PRS_{ER+}$  is presented for *BRCA2* carriers with respect to the breast cancer data.  $PRS_{HGS}$  is shown for both *BRCA1* and *BRCA2* carriers for the ovarian cancer data. The median (IQR) age at start of follow-up, follow-up time and age at cancer diagnosis (years) are displayed. The mean and SD are shown for the appropriate PRS. IQR = interquartile range; N = sample size; SD = standard deviation.

#### Table S5

Assumed proportions and hazard ratios used to constrain the breast cancer incidences from the external BBCC prospective cohort study for breast cancer family history and gene variant location<sup>16</sup>. The absolute risks of breast cancer at the 5th, 50th and 95th percentiles of the PRS are shown (absolute risk curves are plotted in Figures S5-S9).

#### Table S6

Absolute breast cancer and ovarian cancer risks by age 80-years for *BRCA1* and *BRCA2* carriers for different PRS percentiles. The reported PRS percentiles are: (i)  $PRS_{ER-}$  for *BRCA1* carrier breast cancer; (ii)  $PRS_{BC}$  for *BRCA2* carrier breast cancer; (iii)  $PRS_{HGS}$  for *BRCA1* carrier ovarian cancer; and (iv)  $PRS_{HGS}$  for *BRCA2* carrier ovarian cancer.

## **SUPPLEMENTARY FIGURE LEGENDS**

### **Figure S1**

Histograms of imputation accuracy ( $r^2$  statistics) for the 313 breast cancer PRS SNPs. Imputations were performed separately for genotyping arrays (iCOGS or OncoArray) and separately for *BRCA1* and *BRCA2* carriers. All SNPs were well imputed ( $r^2 \geq 0.49$ ).

### **Figure S2**

Histograms of imputation accuracy ( $r^2$  statistics) for the 30 ovarian cancer PRS SNPs. Imputations were performed separately for genotyping arrays (iCOGS or OncoArray) and separately for *BRCA1* and *BRCA2* carriers. All SNPs were well imputed ( $r^2 \geq 0.64$ ).

### **Figure S3**

Forest plots of country specific PRS hazard ratios estimated using the CIMBA retrospective cohort. These models tested for heterogeneity in PRS effects across countries by fitting a PRS by country interaction term. The baseline country was assumed to be UK/Eire. Heterogeneity was assessed using a likelihood ratio test, comparing the model that included the interaction term to a nested model that did not include the interaction term. (A) PRS<sub>ER-</sub> was used for *BRCA1* carriers ( $P_{\text{het}}=0.26$ ). (B) PRS<sub>BC</sub> was used for *BRCA2* carriers ( $P_{\text{het}}=0.58$ ). (C) PRS<sub>HGS</sub> used for *BRCA1* carriers ( $P_{\text{het}}=0.08$ ). (D) PRS<sub>HGS</sub> used for *BRCA2* carriers ( $P_{\text{het}}=0.95$ ).

### **Figure S4**

Estimated 10-year risks of developing breast cancer and ovarian cancer by different PRS distribution percentiles.

### **Figure S5**

Predicted age-specific absolute risks of developing breast cancer by PRS<sub>ER-</sub> percentiles and by *BRCA1* variant location. Risks were calculated assuming the retrospective cohort HR

estimates (Table 2). (A) Predicted absolute risks of developing breast cancer for *BRCA1* carriers with a variant in the 5' to c.2281 region. (B) Predicted absolute risks of developing breast cancer for *BRCA1* carriers with a variant in the c.2282 to c.4071 region. (C) Predicted absolute risks of developing breast cancer for *BRCA1* carriers with a variant in the c.4072 to 3' region.

#### Figure S6

Predicted age-specific absolute risks of developing breast cancer by PRS<sub>BC</sub> percentiles and by *BRCA2* variant location (narrow definition). Risks were calculated assuming the retrospective cohort HR estimates (Table 2). (A) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with a variant in the 5' to c.3846 region. (B) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with a variant in the c.3847 to c.6275 region. (C) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with a variant in the c.6276 to 5' region.

#### Figure S7

Predicted age-specific absolute risks of developing breast cancer by PRS<sub>BC</sub> percentiles and by *BRCA2* variant location (wide definition). Risks were calculated assuming the retrospective cohort HR estimates (Table 2). (A) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with a variant in the 5' to c.2830 region. (B) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with a variant in the c.2831 to c.6402 region. (C) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with a variant in the c.6403 to 5' region.

#### Figure S8

*BRCA1* carriers: Predicted age-specific absolute risks of developing breast cancer by PRS<sub>ER</sub> percentiles and by family history (FH) of breast cancer. Risks were calculated assuming the retrospective cohort HR estimates (Table 2). (A) Predicted absolute risks of



developing breast cancer for *BRCA1* carriers with no family history of breast cancer. (B) Predicted absolute risks of developing breast cancer for *BRCA1* carriers with positive family history of breast cancer.

Figure S9

*BRCA2* carriers: Predicted age-specific absolute risks of developing breast cancer by PRS<sub>BC</sub> percentiles and by family history (FH) of breast cancer. Risks were calculated assuming the retrospective cohort HR estimates (Table 2). (A) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with no family history of breast cancer. (B) Predicted absolute risks of developing breast cancer for *BRCA2* carriers with positive family history of breast cancer.

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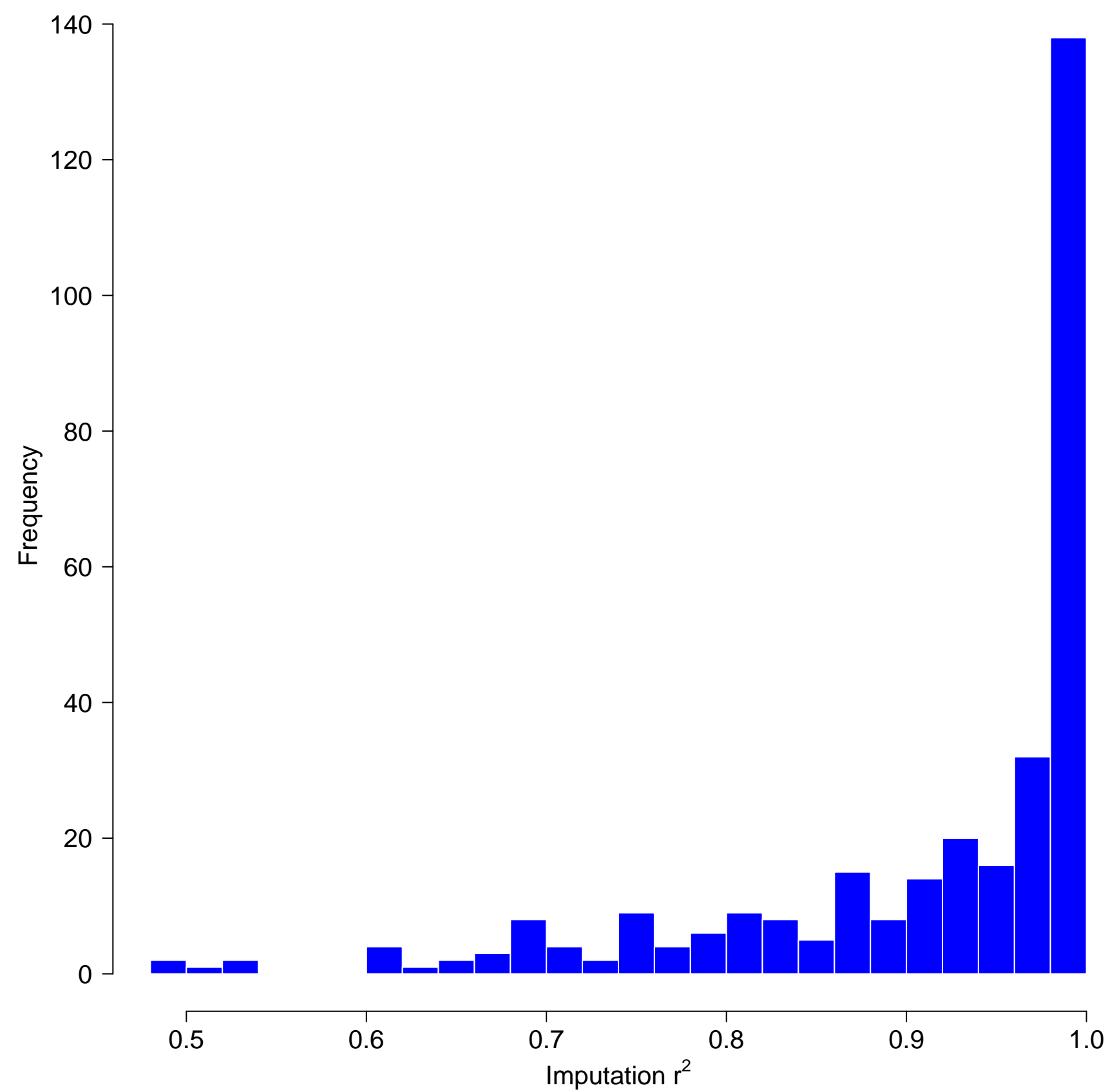
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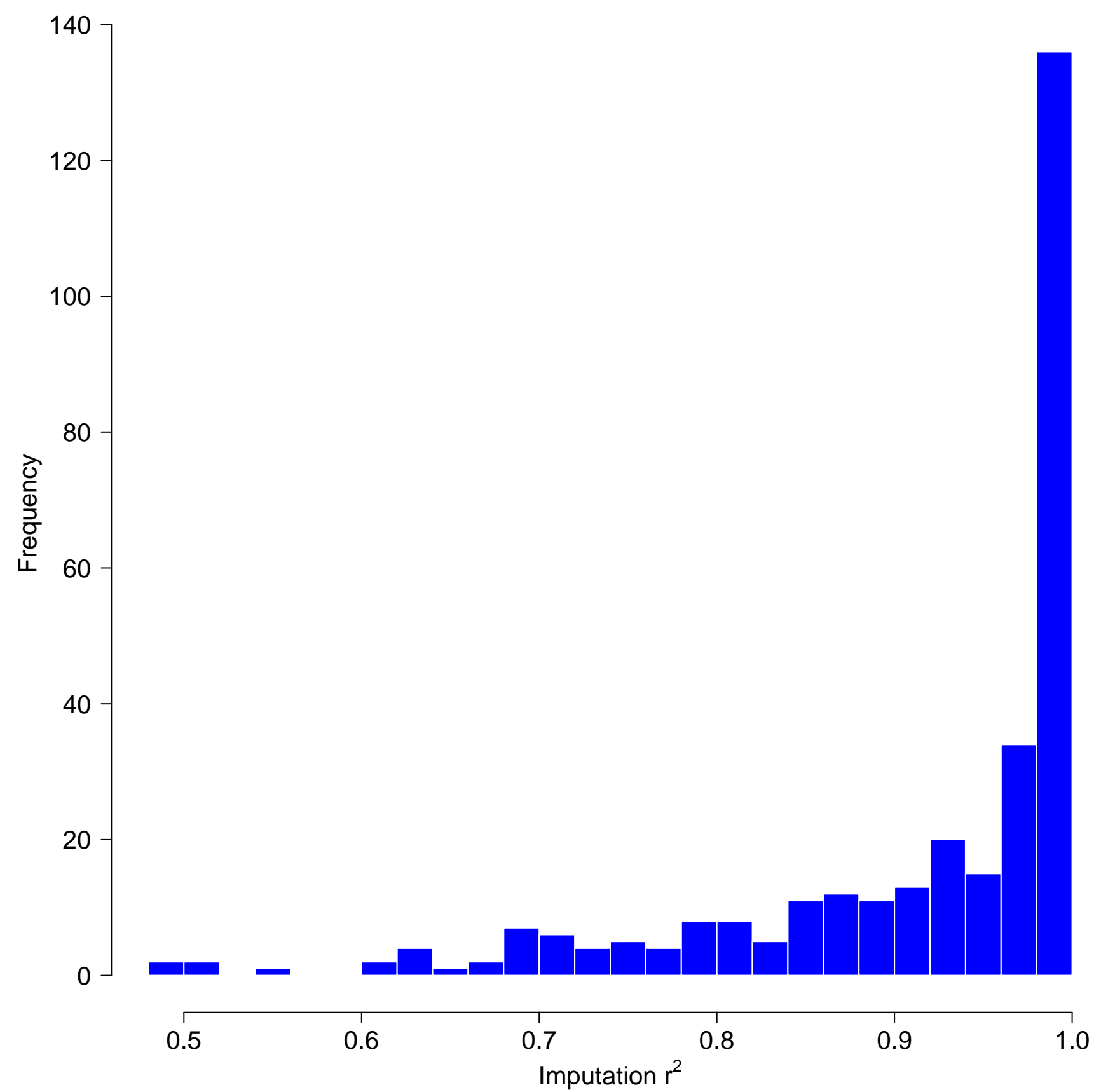
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Figure S1

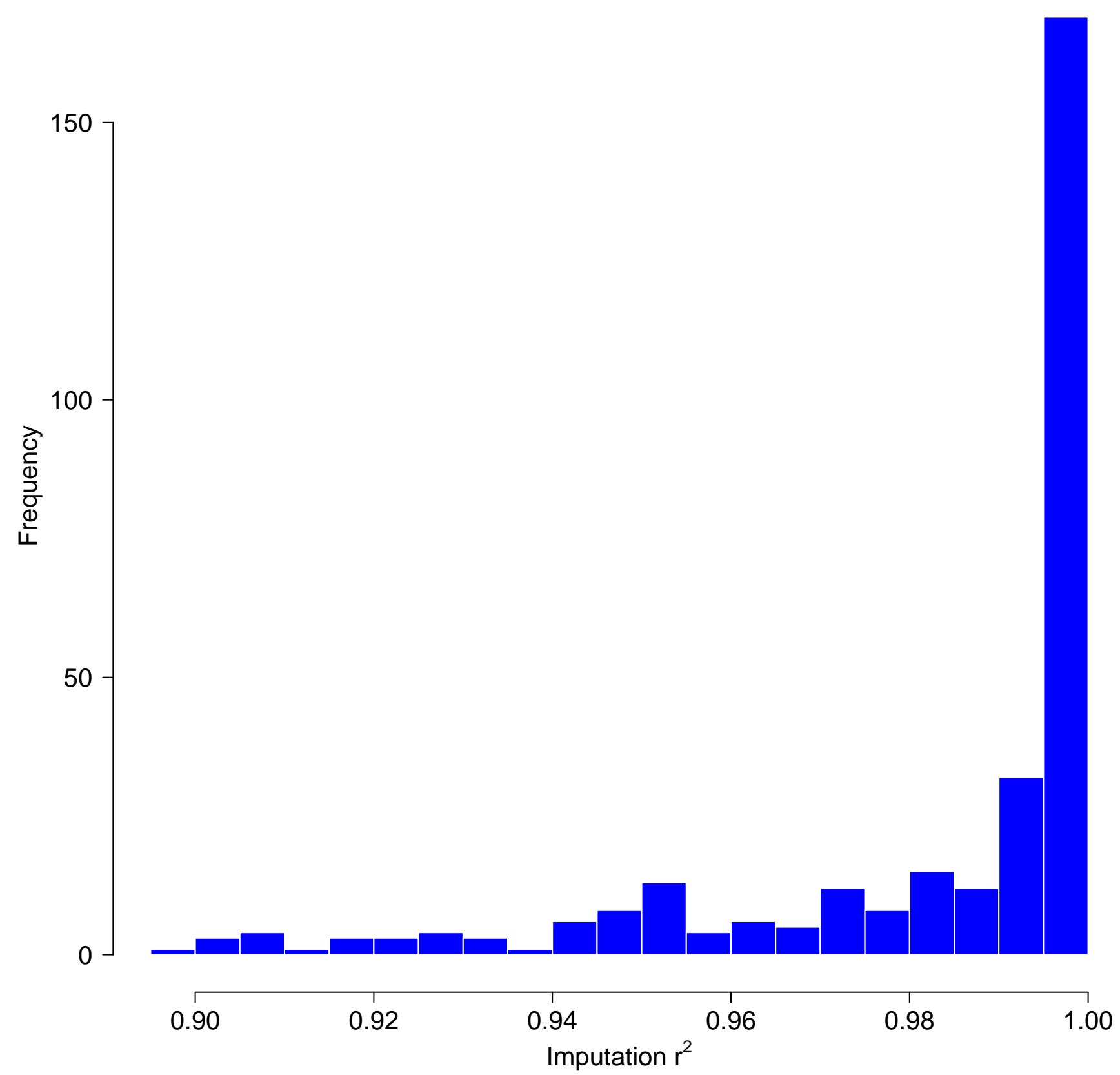
(A) iCOGS: BRCA1 carriers



(B) iCOGS: BRCA2 carriers



(C) OncoArray: BRCA1 carriers



(D) OncoArray: BRCA2 carriers

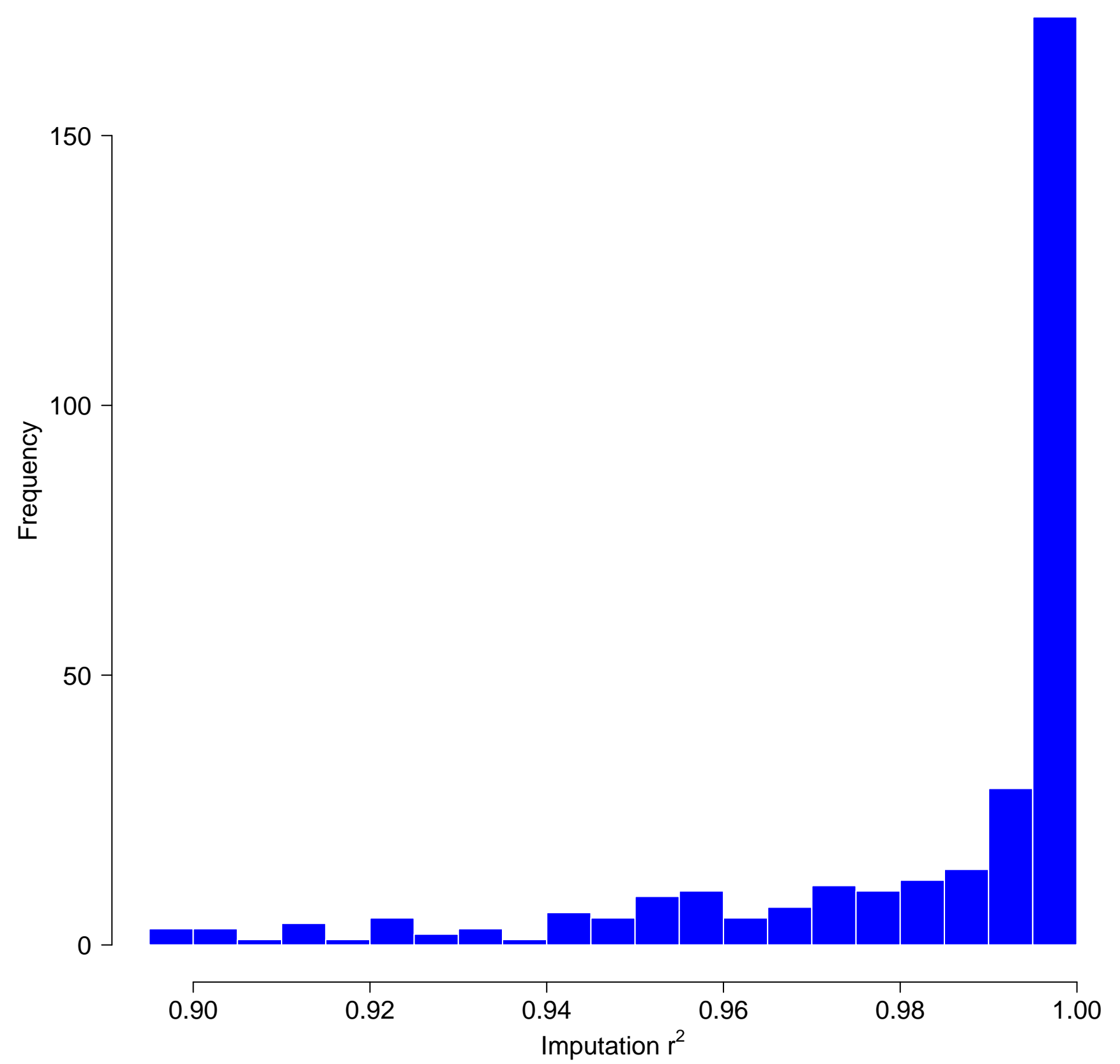
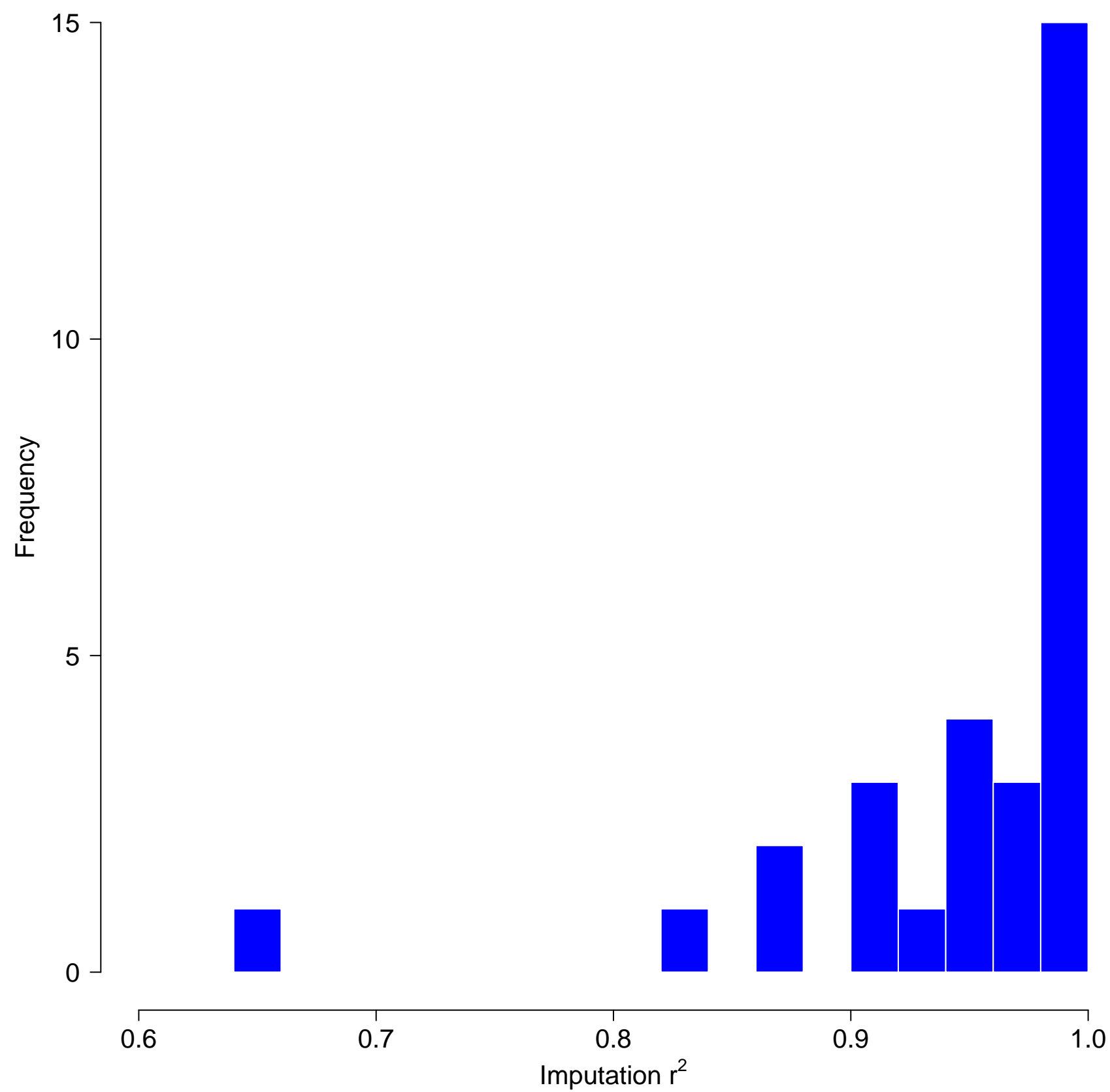
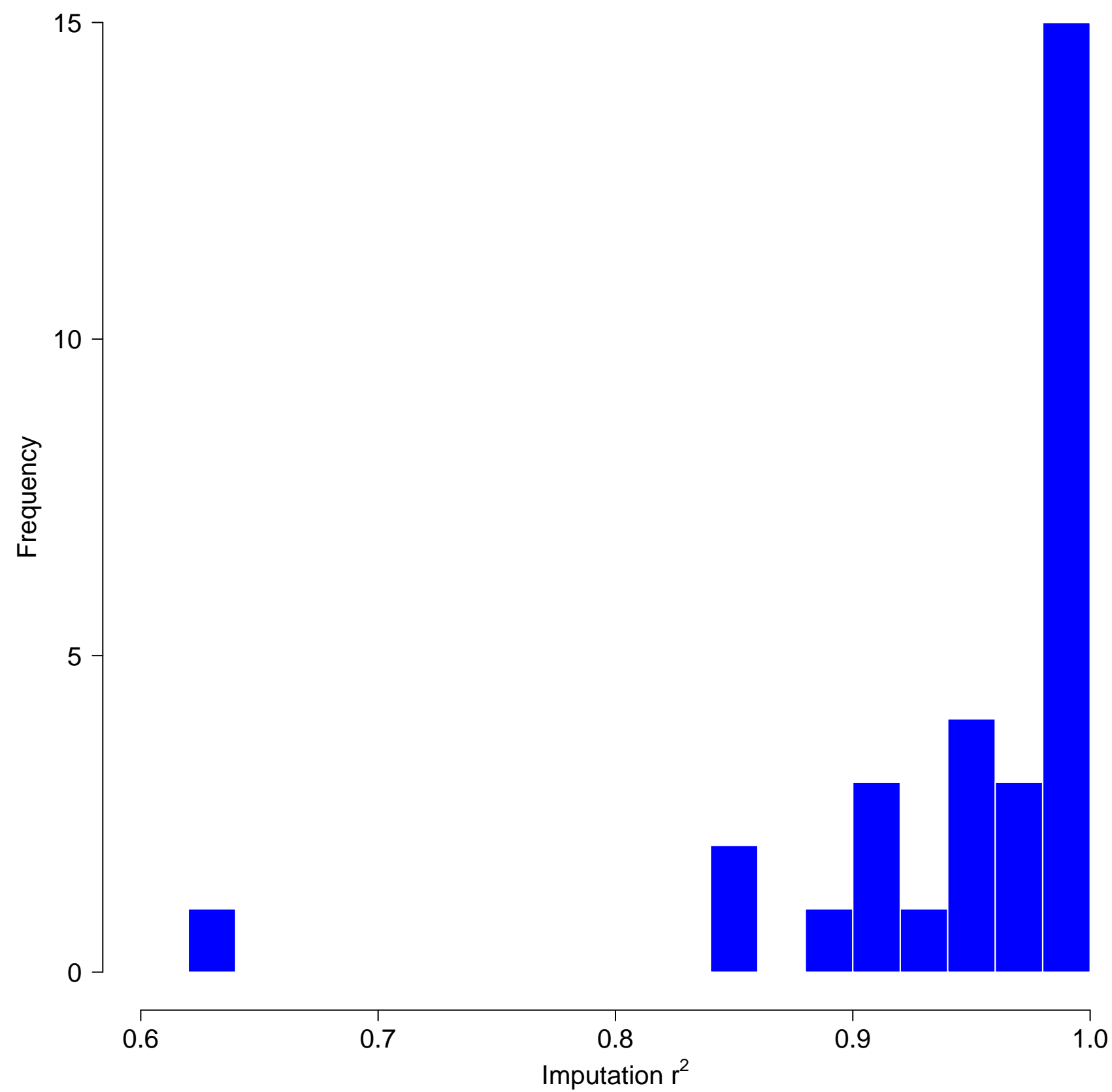


Figure S2

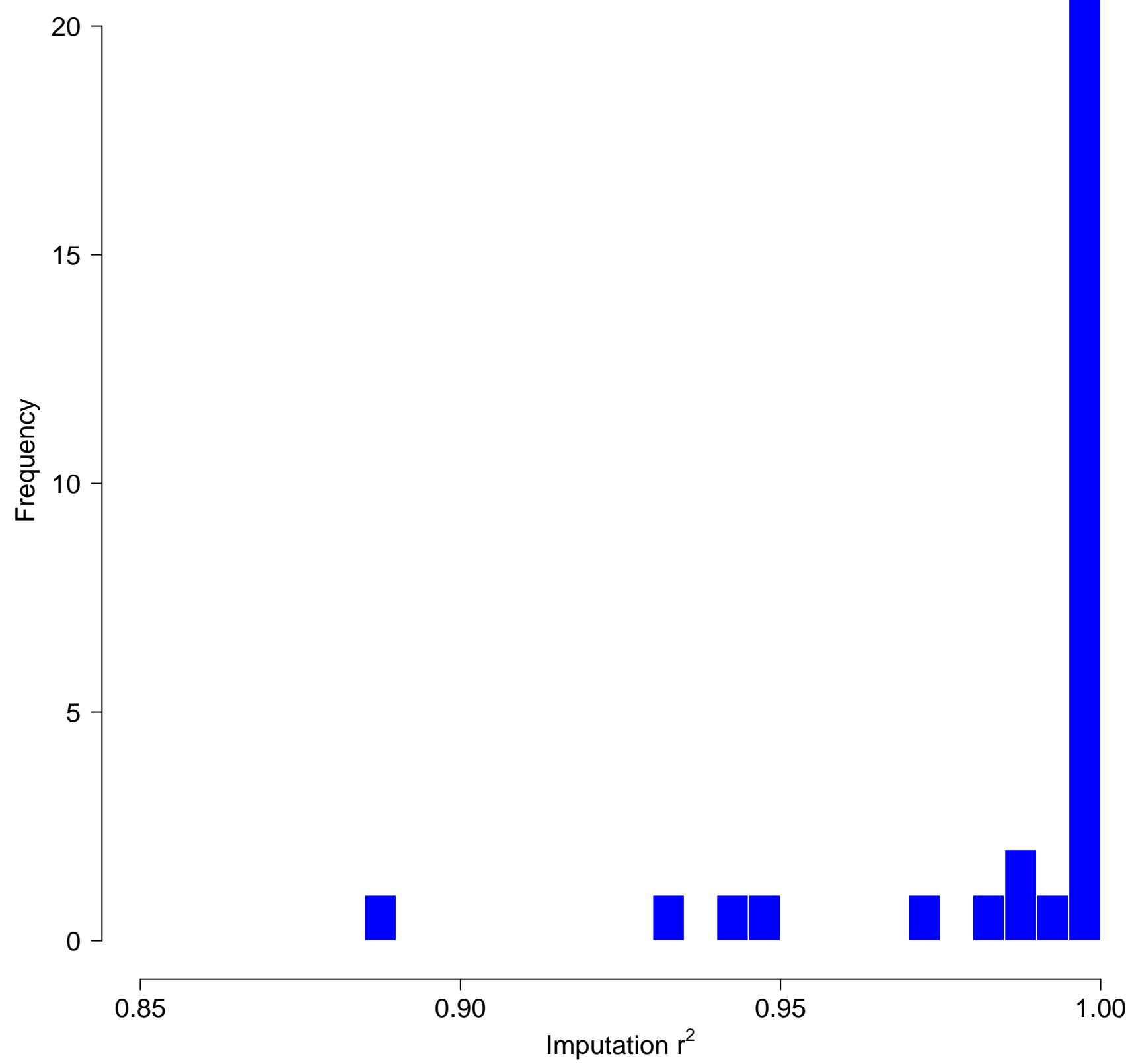
(A) iCOGS: BRCA1 carriers



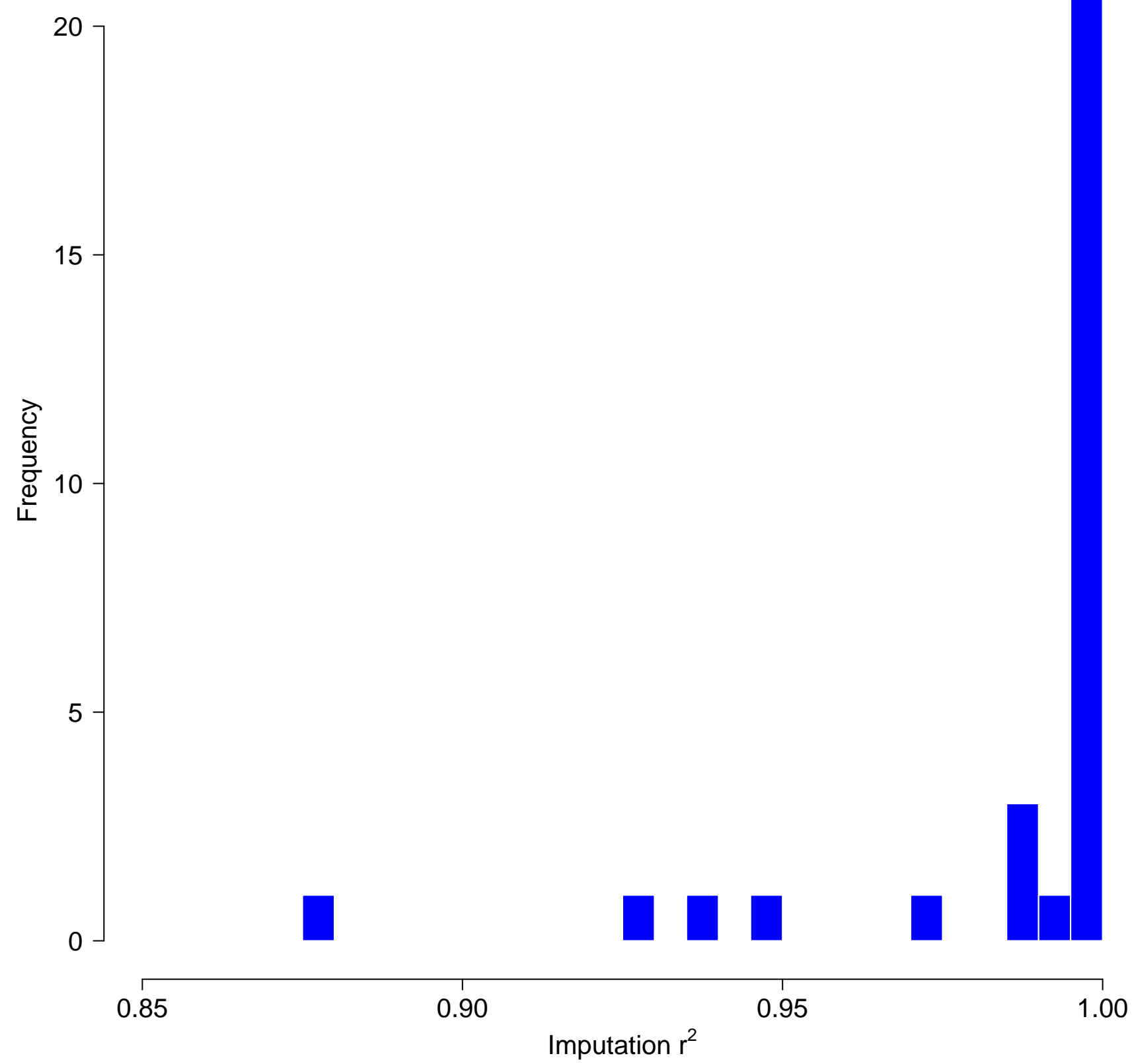
(B) iCOGS: BRCA2 carriers



(C) OncoArray: BRCA1 carriers

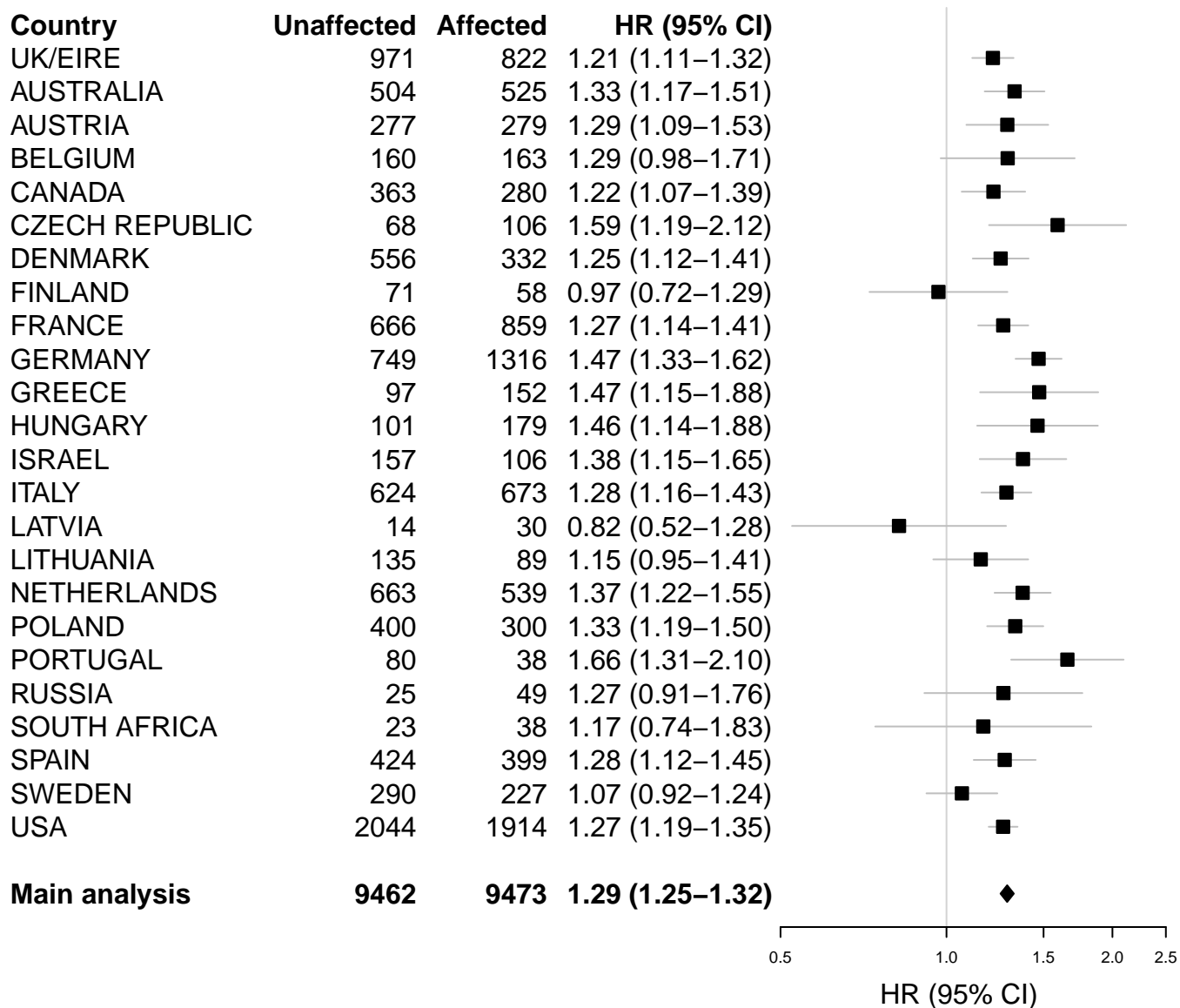


(D) OncoArray: BRCA2 carriers



# Figure S3

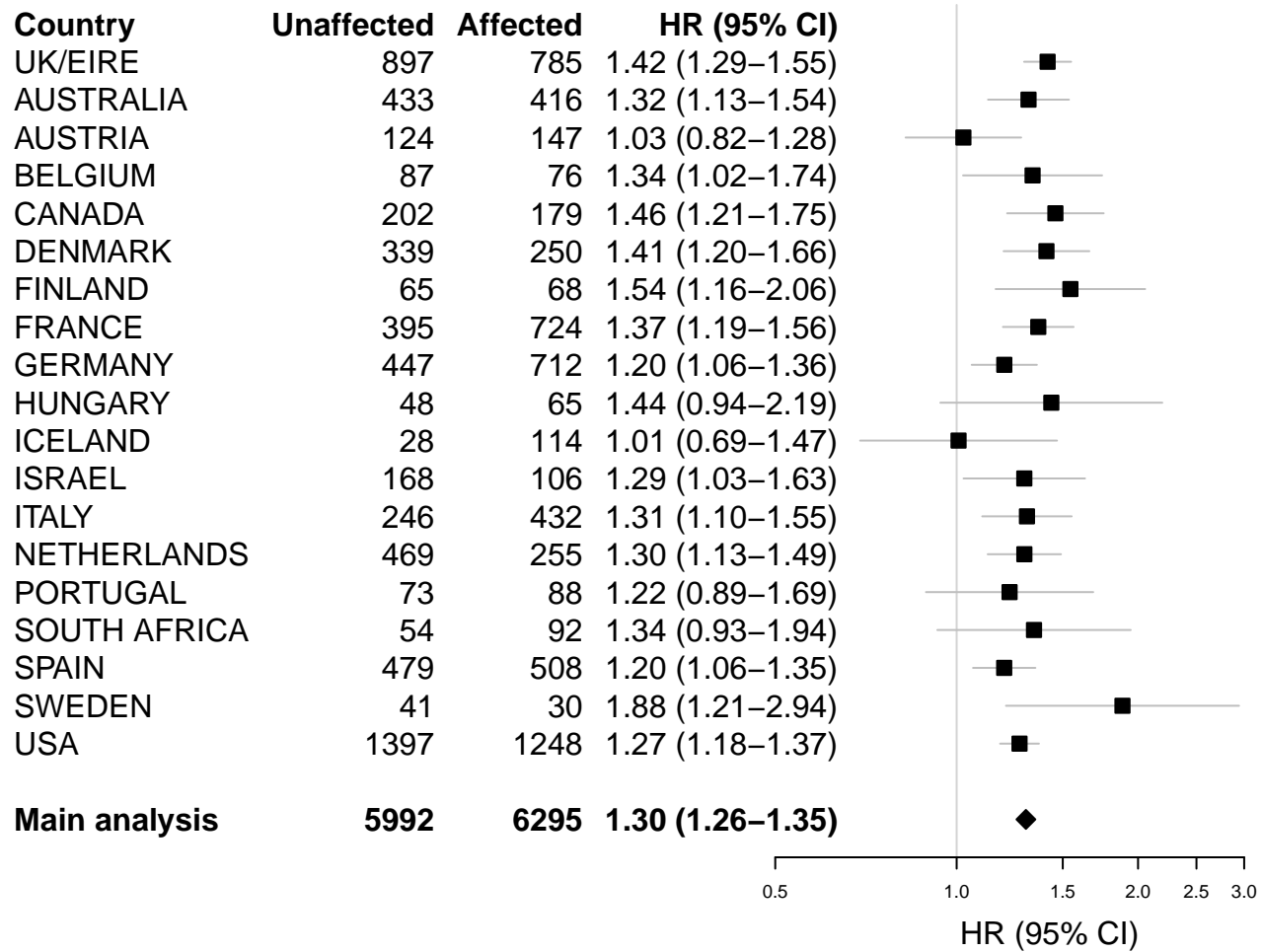
## (A) BRCA1 carriers: ER-negative PRS





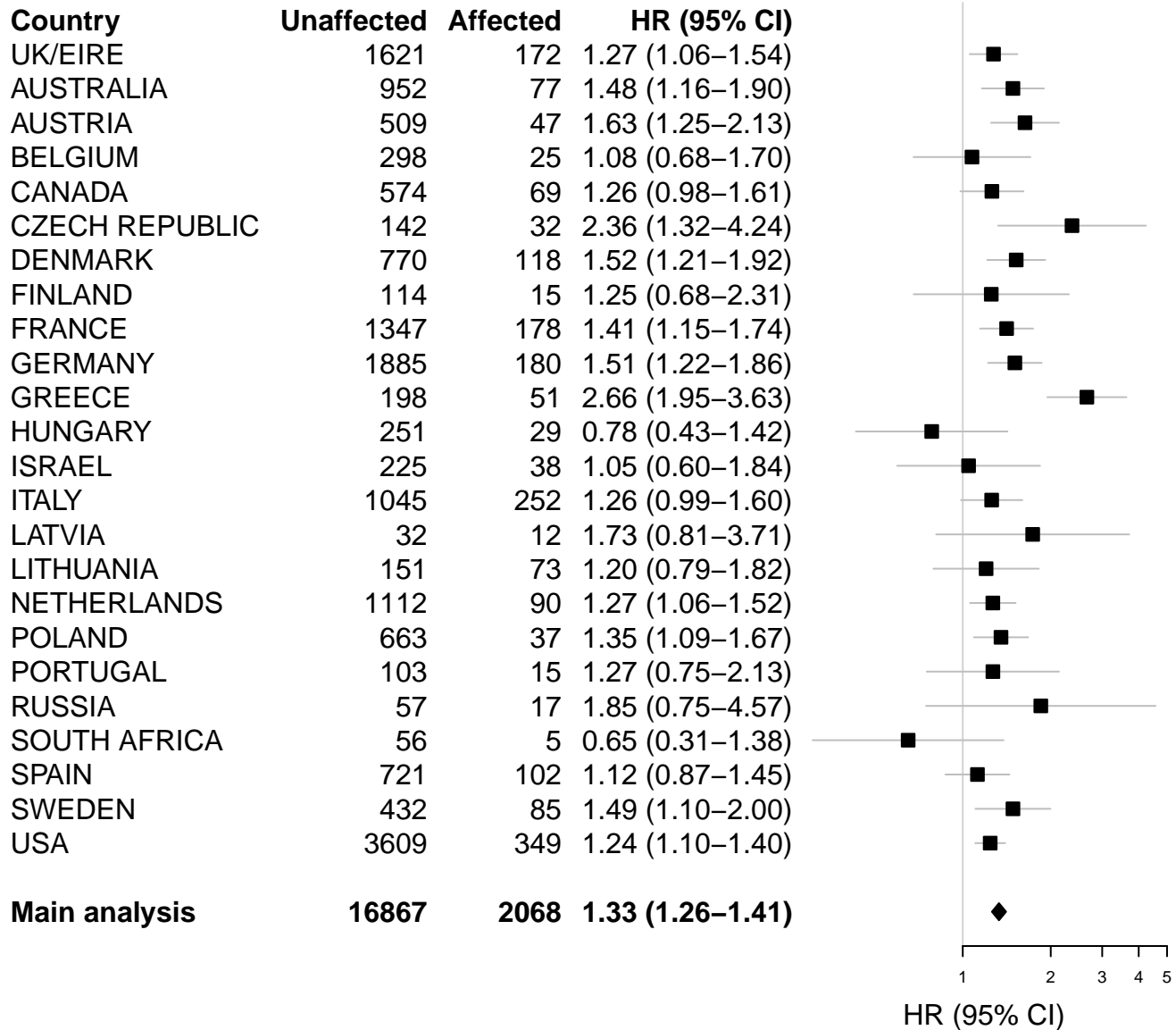
# Figure S3

## (B) BRCA2 carriers: Overall PRS



# Figure S3

## (C) BRCA1 carriers: HGS PRS



# Figure S3

## (D) BRCA2 carriers: HGS PRS

Country	Unaffected	Affected	HR (95% CI)
UK/EIRE	1579	103	1.63 (1.27–2.09)
AUSTRALIA	820	29	1.19 (0.89–1.60)
AUSTRIA	262	9	2.25 (1.30–3.87)
BELGIUM/NETHERLANDS	852	35	1.07 (0.77–1.48)
CANADA	355	26	1.52 (0.98–2.37)
DENMARK/ICELAND	687	44	1.47 (1.02–2.13)
FINLAND/LITHUANIA/RUSSIA	145	8	1.00 (0.36–2.78)
FRANCE	1053	66	1.64 (1.09–2.48)
GERMANY	1094	65	1.34 (0.92–1.95)
GREECE/ITALY	653	57	1.29 (0.80–2.07)
HUNGARY	105	8	2.39 (0.86–6.63)
ISRAEL	257	17	0.96 (0.56–1.66)
PORTUGAL/SPAIN	1087	61	1.31 (0.94–1.82)
SOUTH AFRICA	134	12	0.92 (0.14–6.07)
SWEDEN	61	10	1.70 (0.78–3.68)
USA	2477	168	1.52 (1.24–1.86)
<b>Main analysis</b>	<b>16867</b>	<b>2068</b>	<b>1.42 (1.28–1.57)</b>

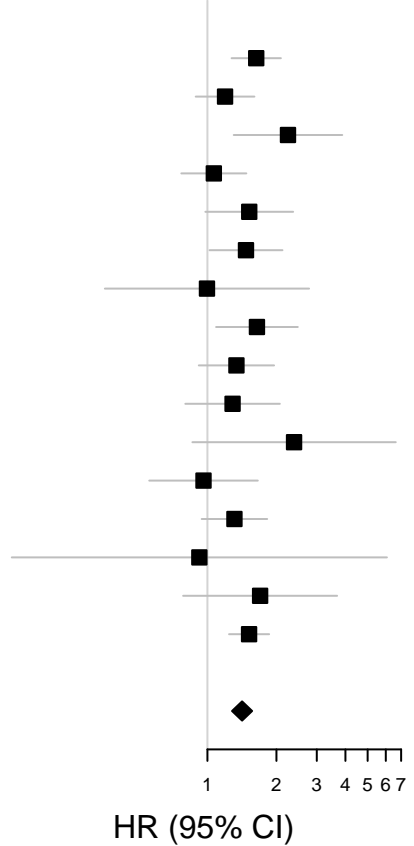
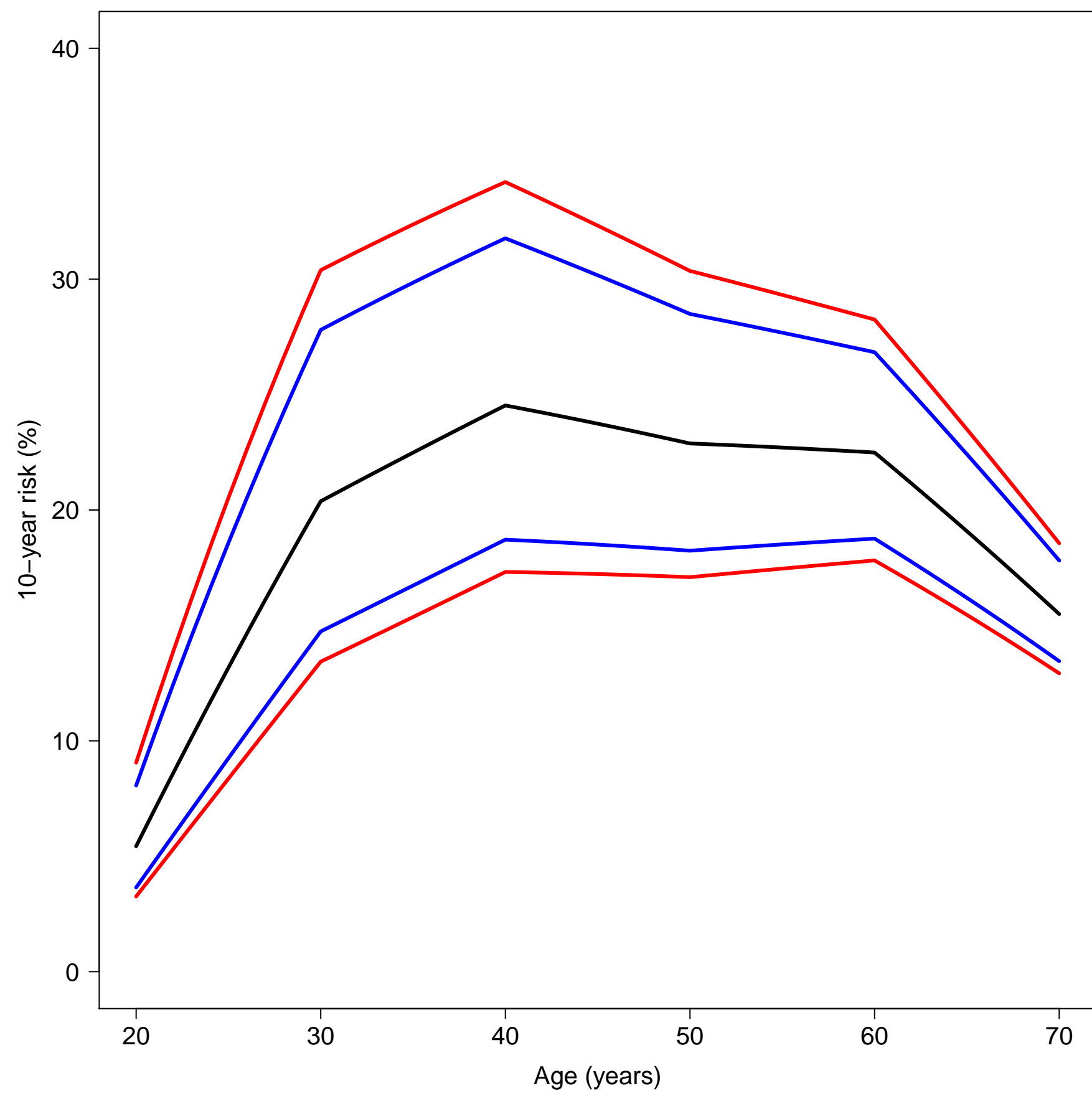
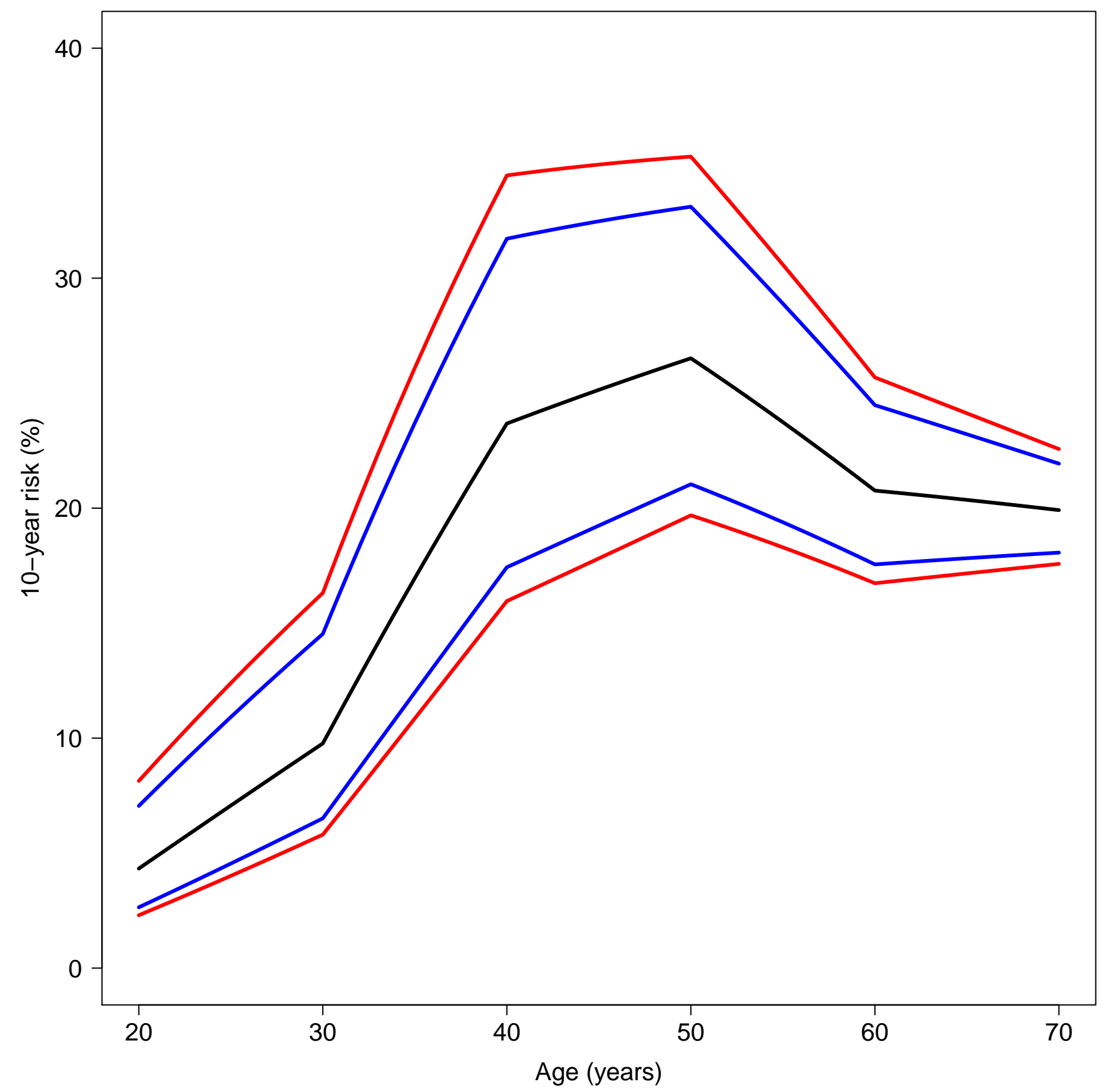


Figure S4

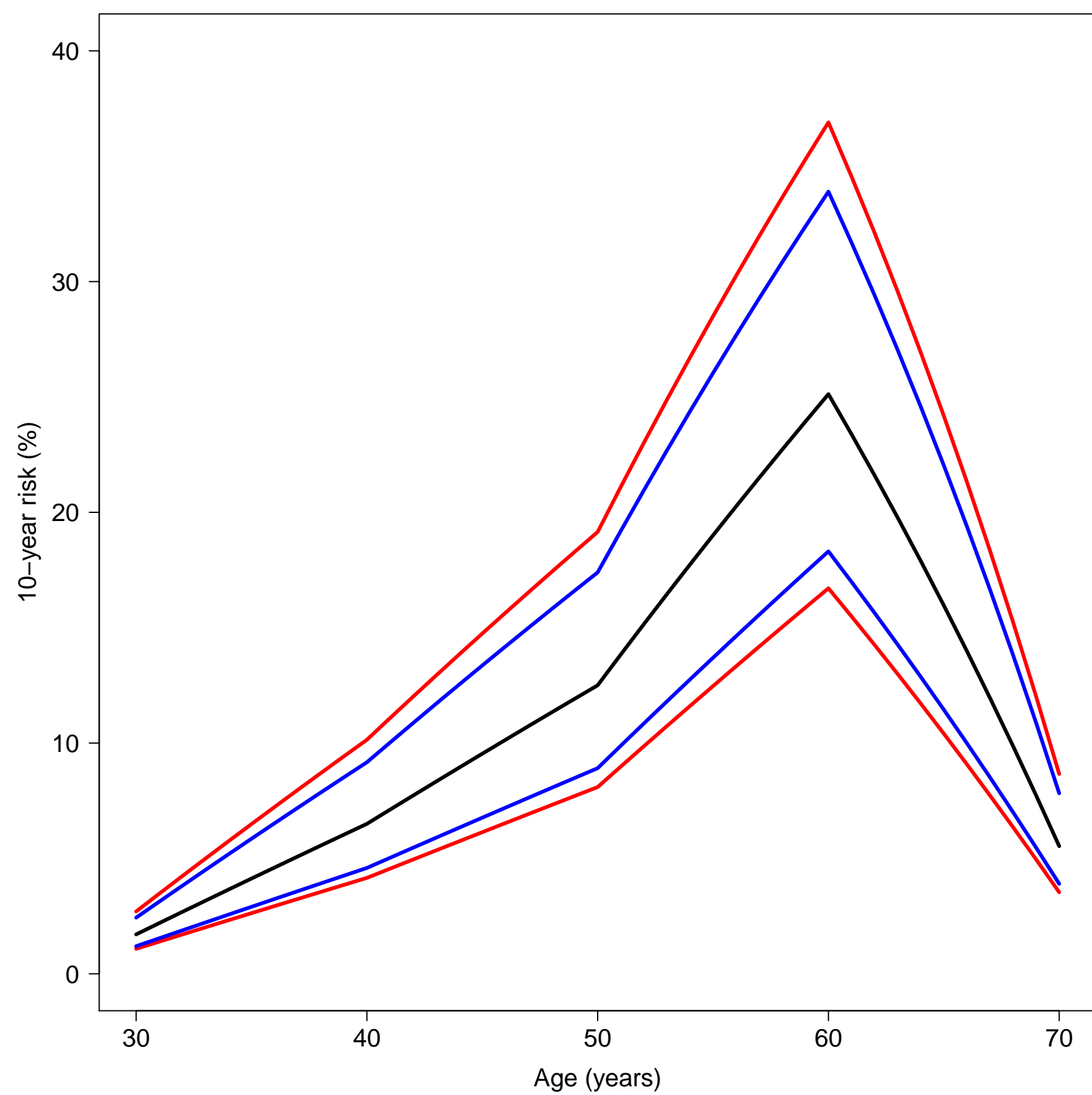
(A) BRCA1 carriers: ER-negative PRS



(B) BRCA2 carriers: Overall PRS



(C) BRCA1 carriers: HGS PRS



(D) BRCA2 carriers: HGS PRS

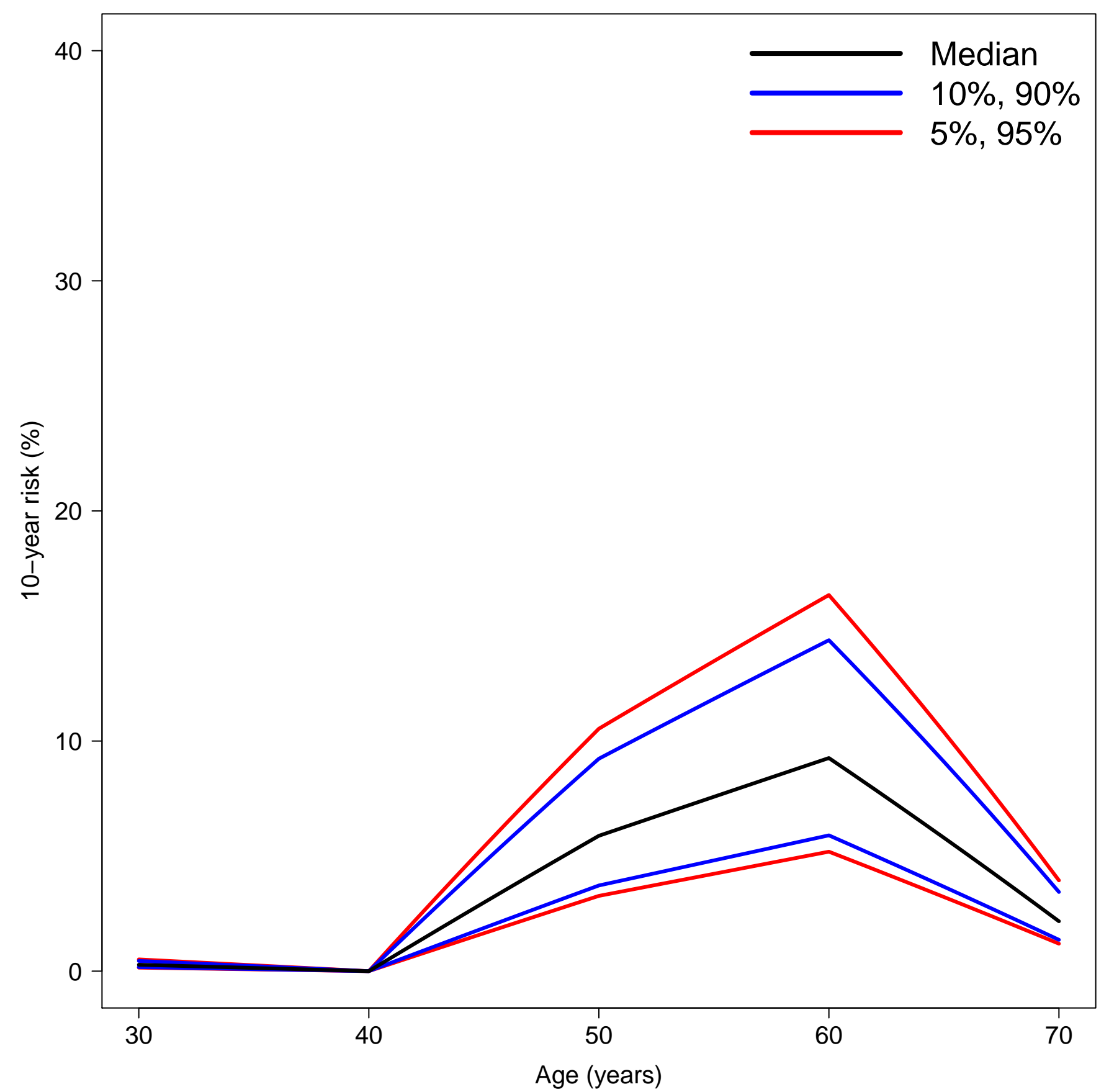
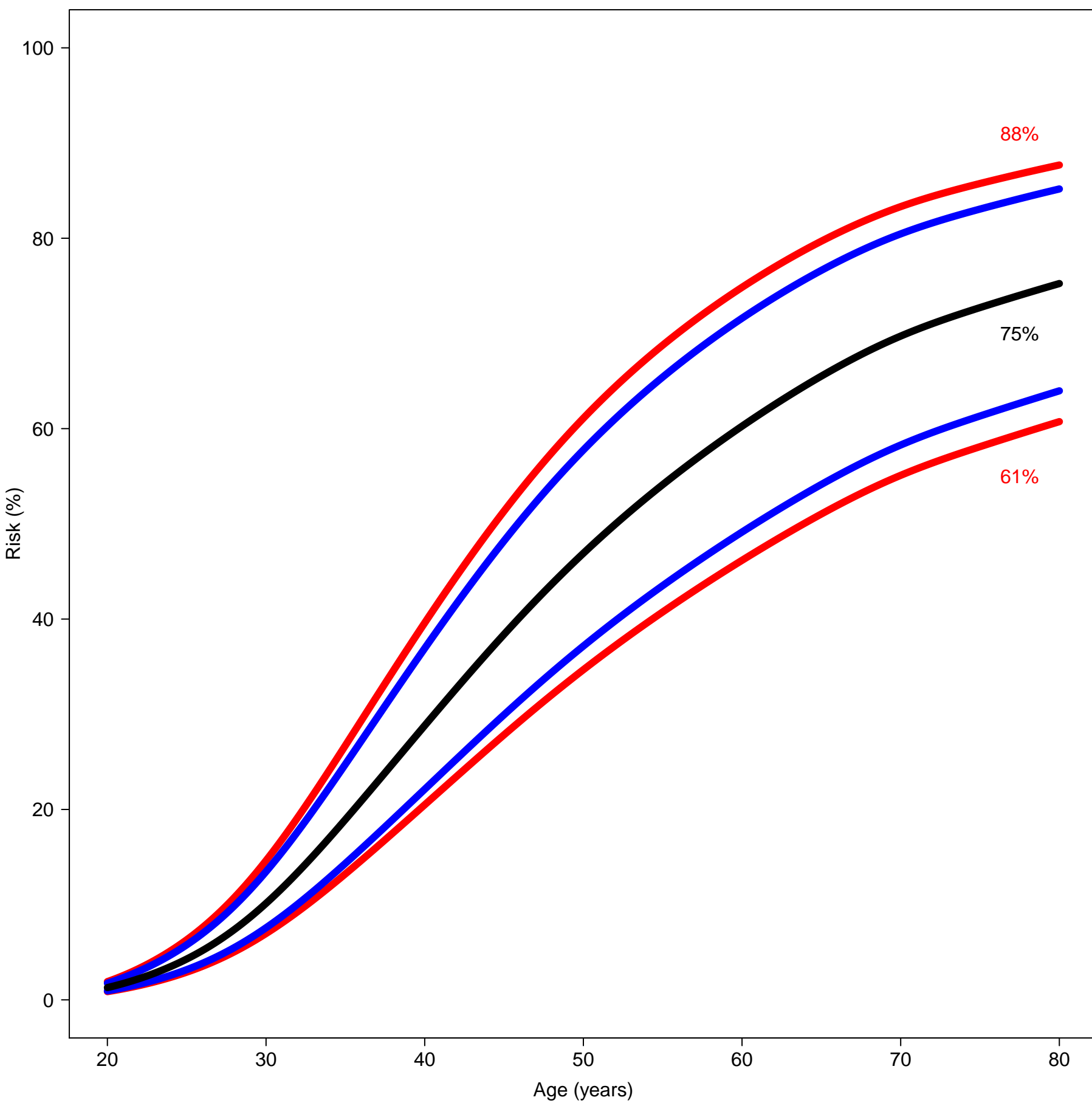
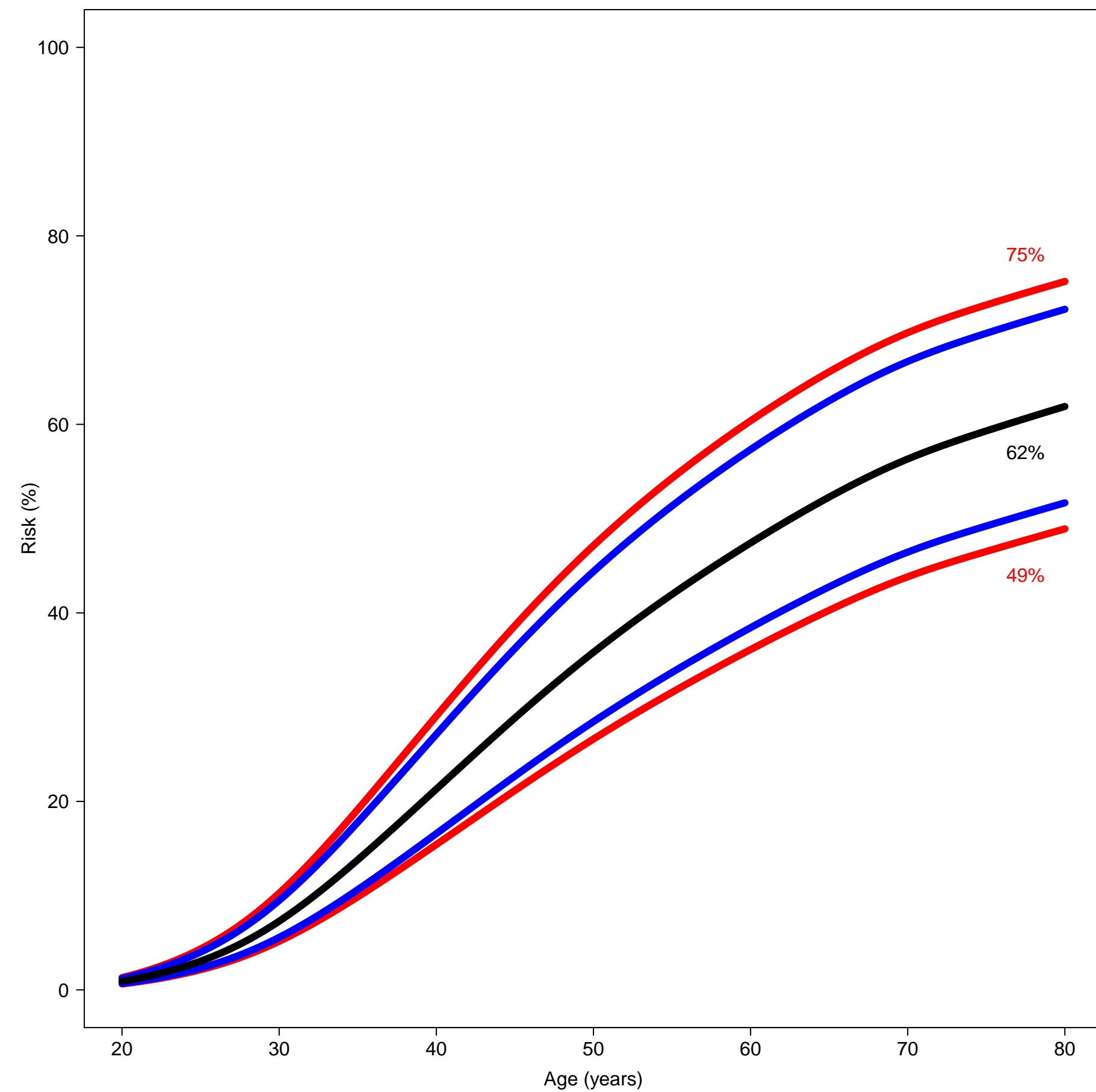


Figure S5

(A) 5' to c.2281



(B) c.2282 to c.4071



(C) c.4072 to 3'

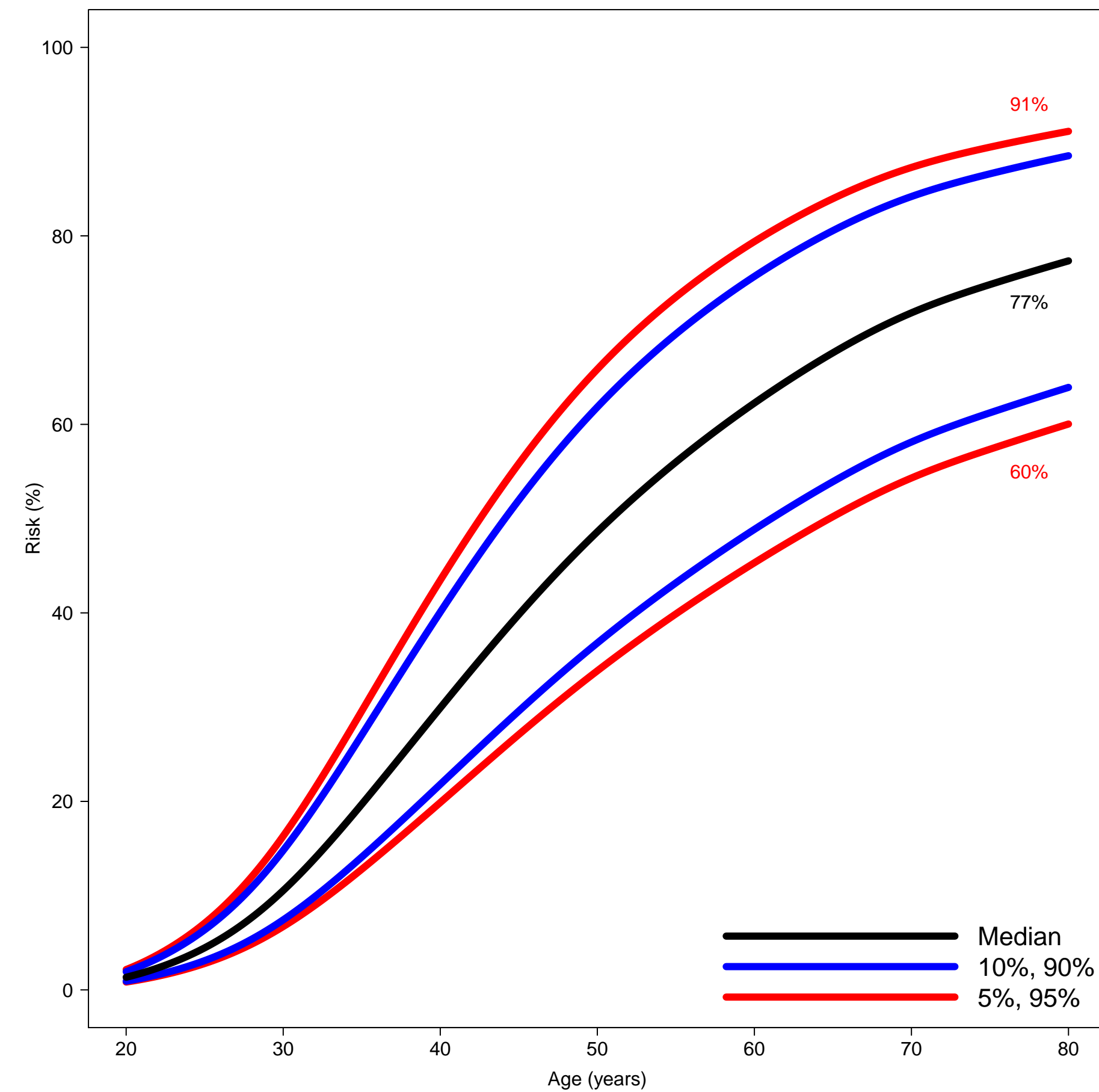
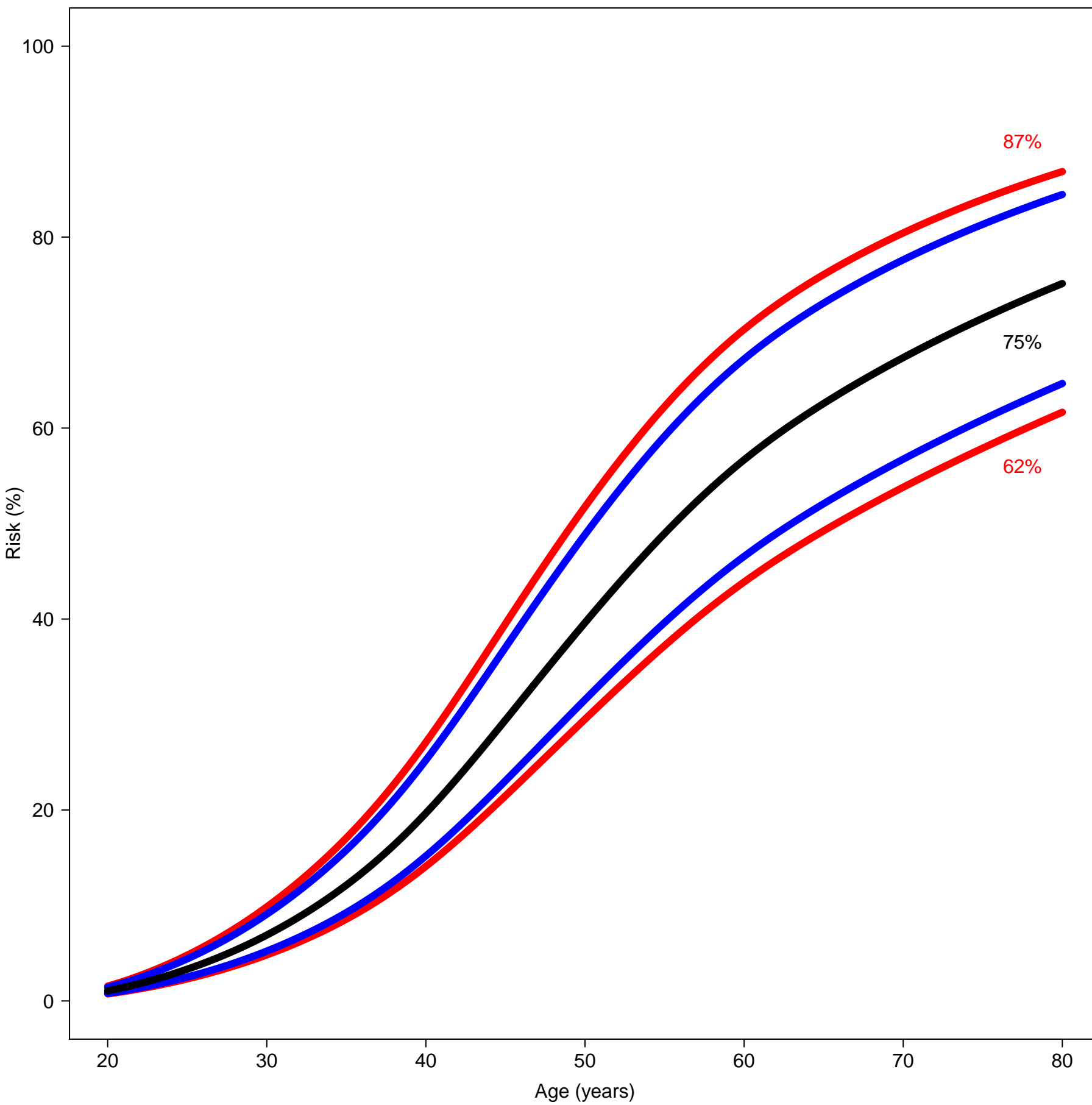
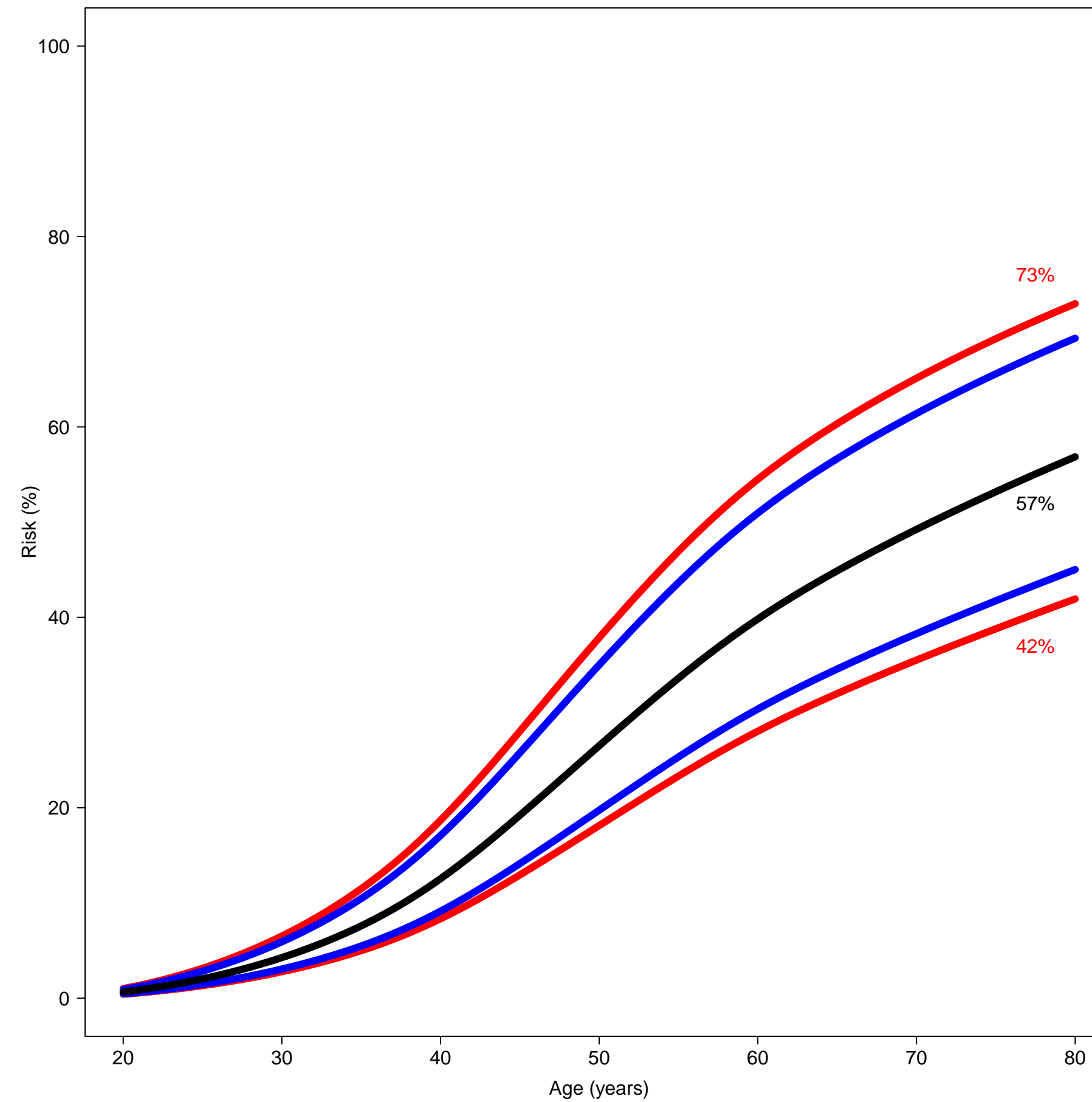


Figure S6

(A) 5' to c.3846



(B) c.3847 to c.6275



(C) c.6276 to 3'

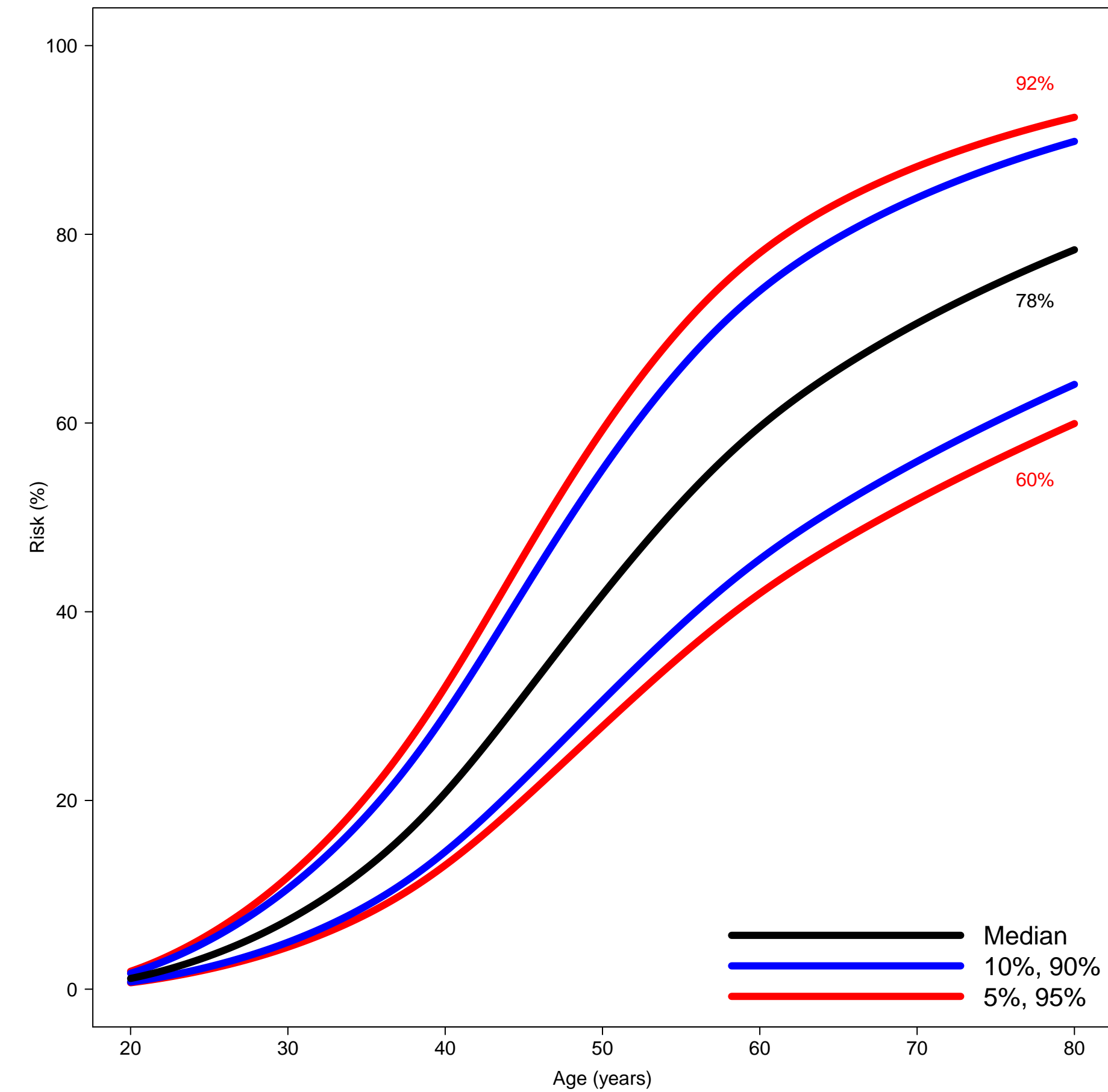
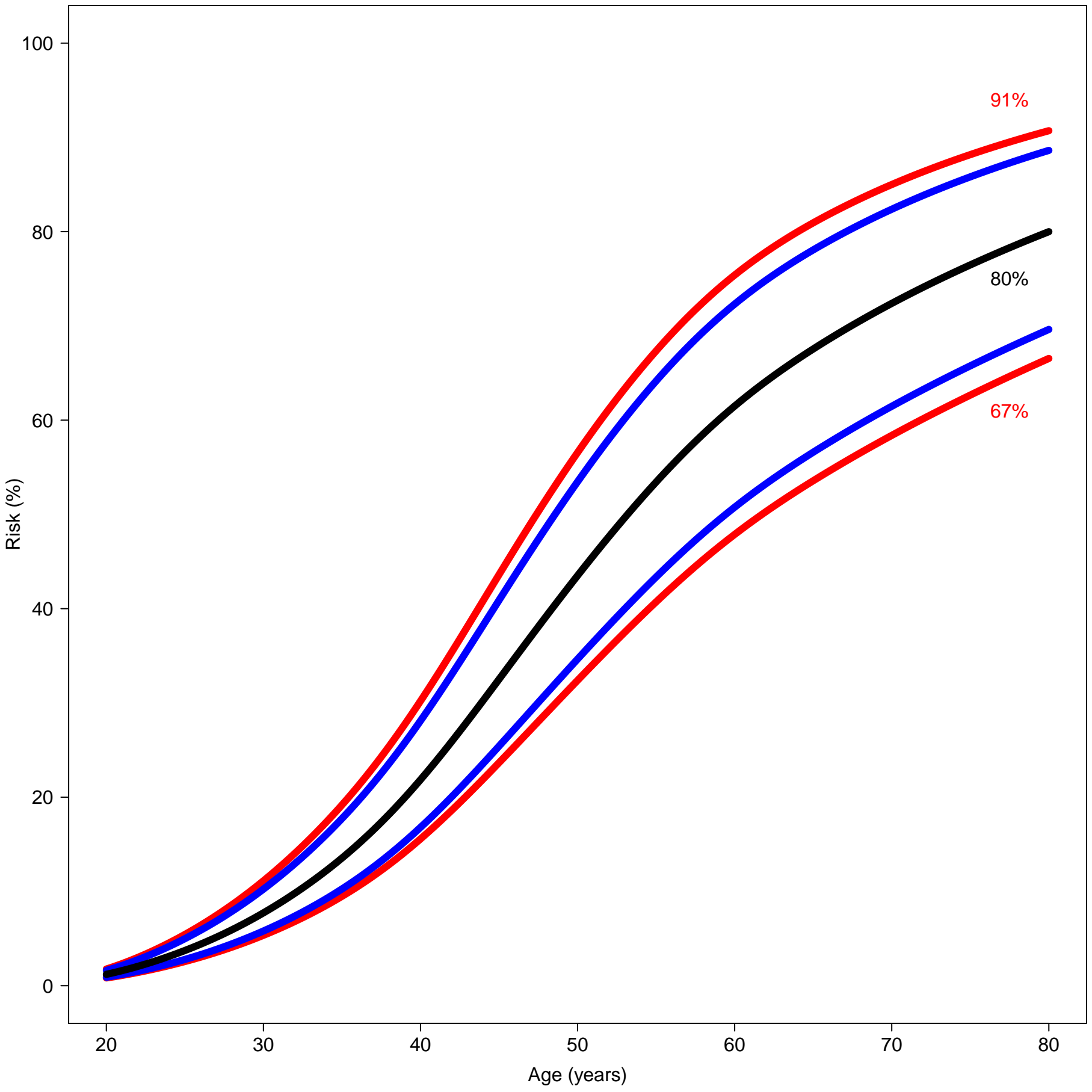
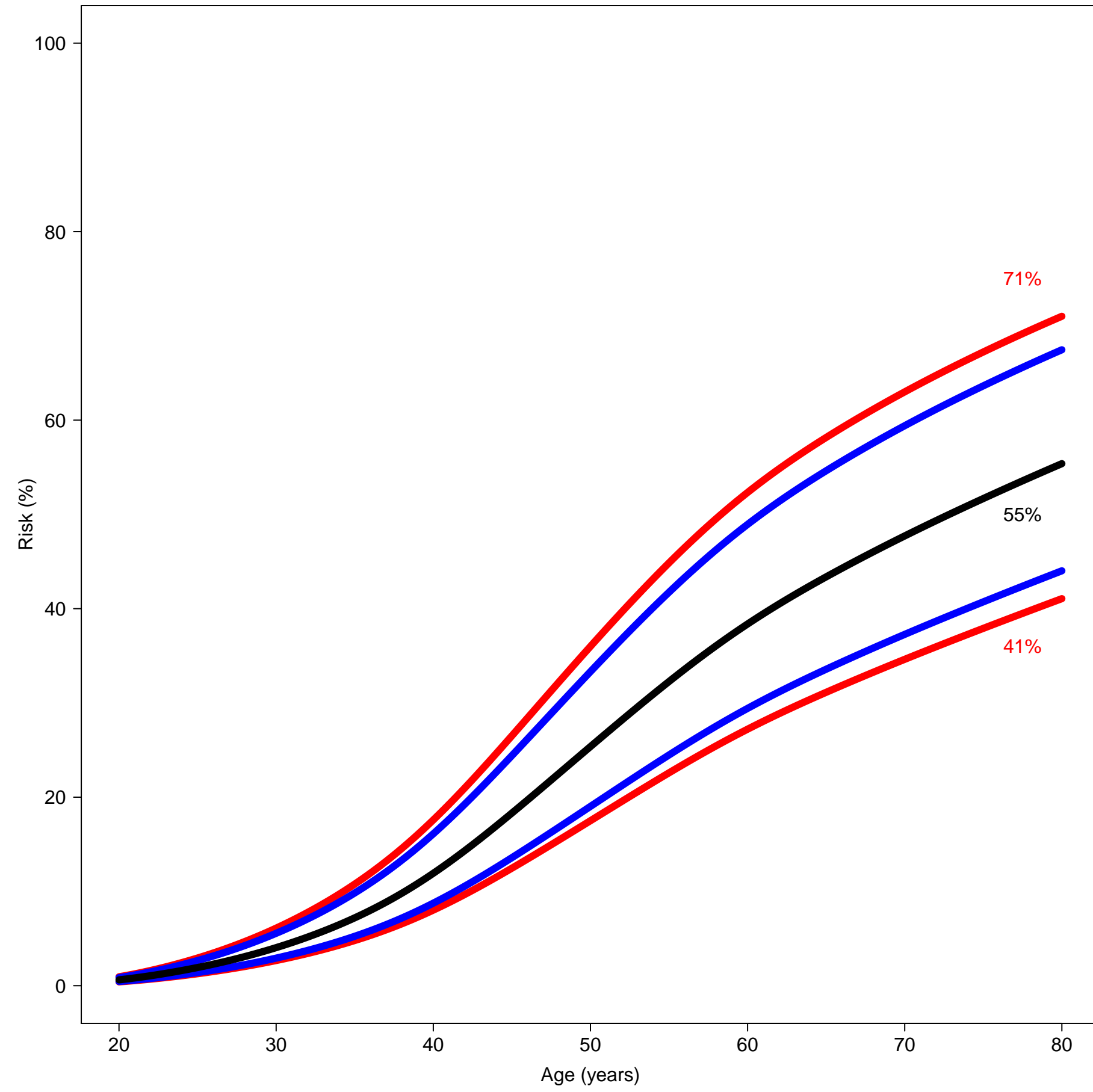


Figure S7

(A) 5' to c.2830



(B) c.2831 to c.6402



(C) c.6403 to 3'

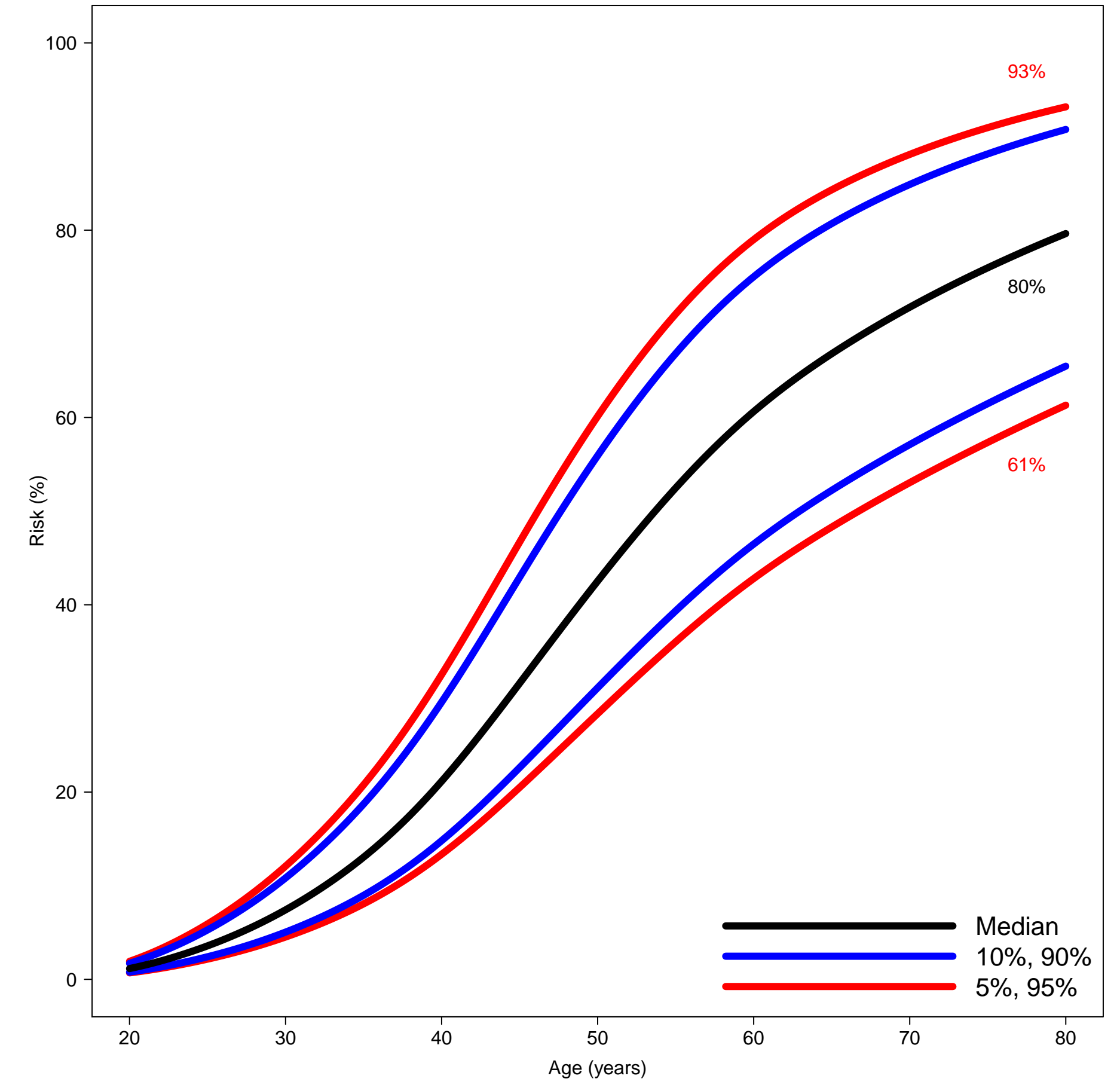
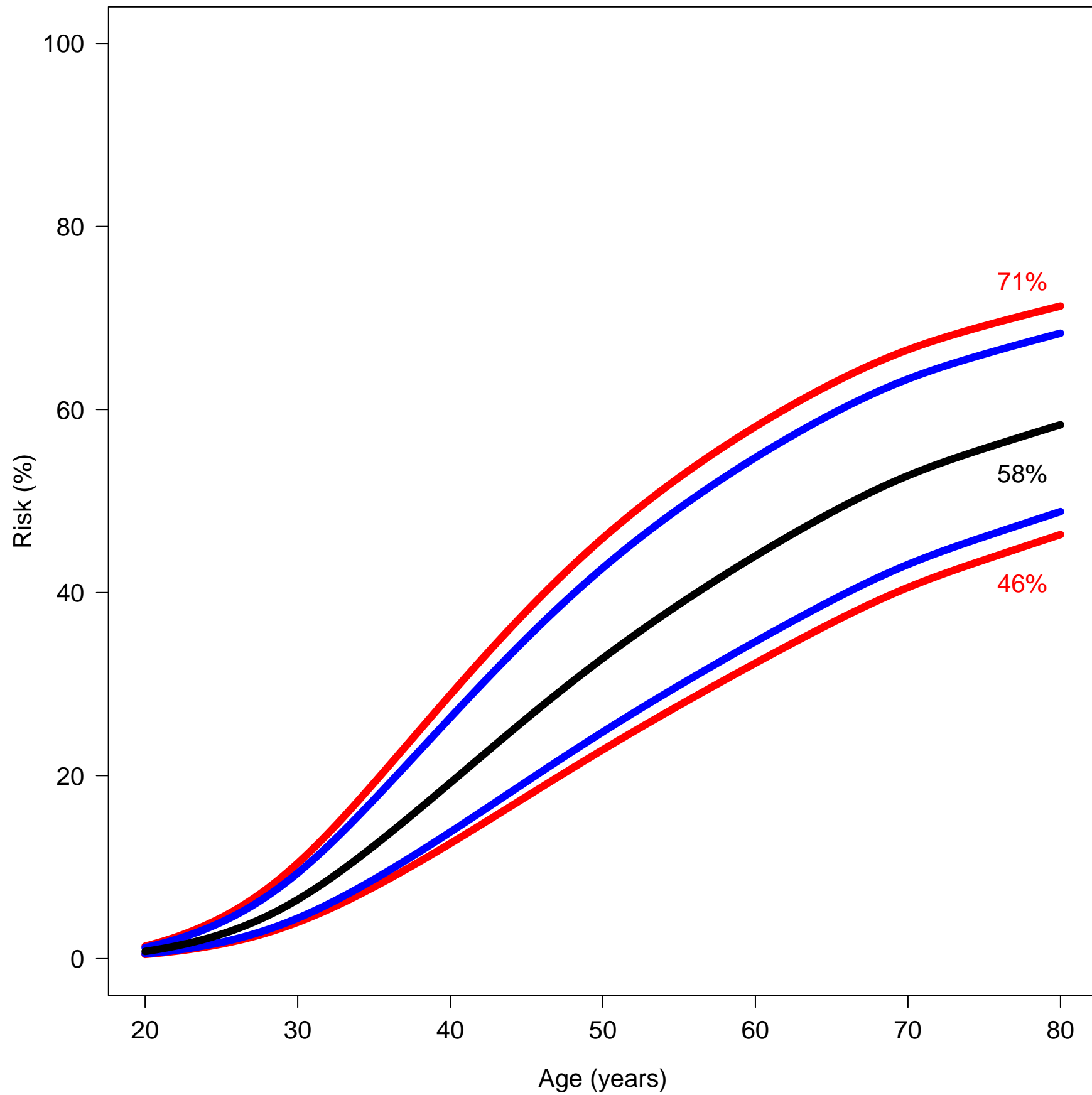


Figure S8

(A) FH-negative



(B) FH-positive

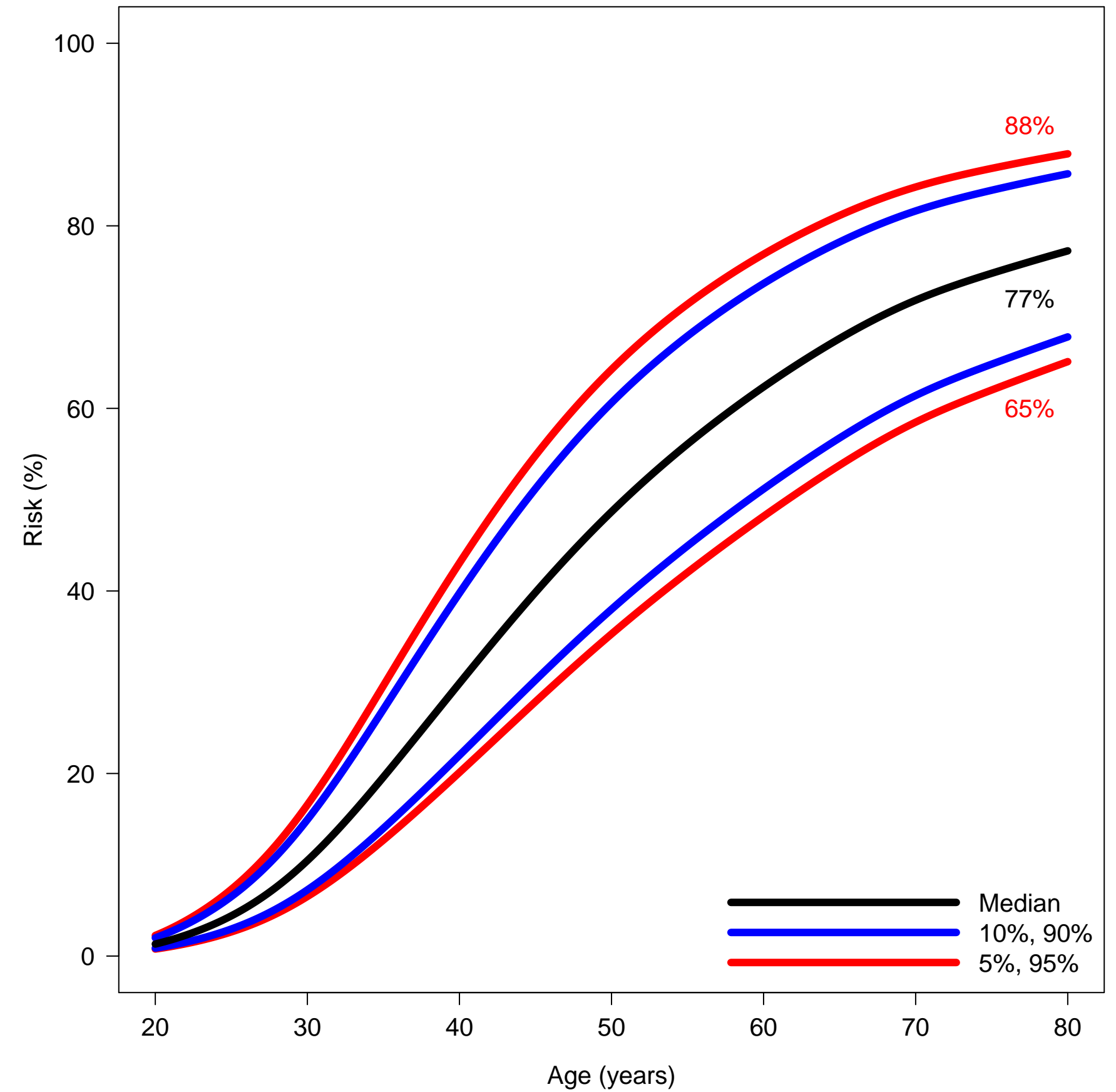
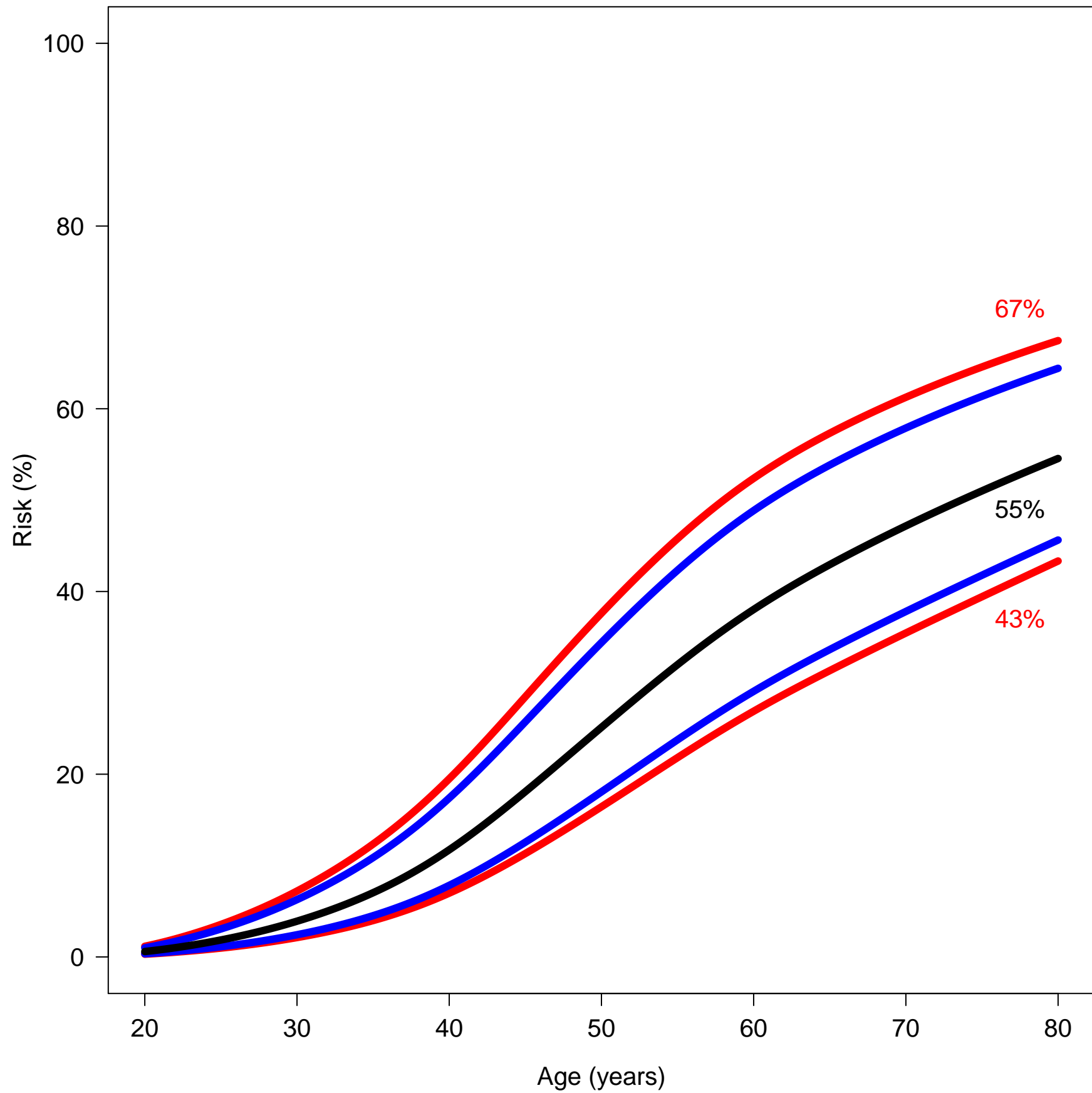




Figure S9

(A) FH-negative



(B) FH-positive

