





Genome Sequences of Bacillus sporothermodurans Strains Isolated from Ultra-High-Temperature Milk

Rodney Owusu-Darko, a Mushal Allam, b D Sílvia D. de Oliveira, c Carlos A. S. Ferreira, c Sunita Grover, d Senzo Mtshali, b Arshad Ismail, Bashmi H. Mallappa, Frederick Tabit, Delna M. Buysa

ABSTRACT Here, we report the draft genome sequences of 3 Bacillus sporothermodurans strains isolated from ultra-high-temperature milk products in South Africa and Brazil and the type strain MB 581 (DSM 10599). The genomes will provide valuable information on the molecular dynamics of heat resistance in B. sporothermodurans.

acillus sporothermodurans is a thermoresistant Gram-positive bacterium that can produce highly heat-resistant endospores (HRS) capable of surviving ultra-hightemperature (UHT) heat treatments (1, 2). First detected in UHT milk (3), it has subsequently been isolated from other dairy products, including UHT cream, chocolate milk, evaporated milk, and reconstituted milk (4). Furthermore, B. sporothermodurans has been isolated from non-dairy-based foods, including Indian curry (5), as well as from marine sources (6). After heat processing, the surviving spores may germinate and grow in the stored milk (1). The spores of B. sporothermodurans grow at low levels ($\approx 10^5$ CFU/ml) and do not affect the pH of the milk (2); as a result, its presence may go unnoticed. However, there are reports of B. sporothermodurans strains isolated in Brazil causing significant proteolytic activity leading to UHT milk spoilage (7). If spoilage does occur, though, there may be slight changes in color, off flavors, and the destabilization of casein micelles (2). Consequently, the main concerns to the dairy industry are milk quality, nonsterility of milk products, and biofilm formation in milk processing equipment.

B. sporothermodurans strains SAD and SA01 were isolated from UHT milk produced in South Africa, and B. sporothermodurans strain BR12 was isolated from UHT milk from Brazil. The type strain B. sporothermodurans DSM 10599 was obtained from Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures GmbH (DSMZ). The enumeration and isolation of strains SAD and SA01 were undertaken as previously described (8), with the substitution of plate count agar for brain heart infusion (BHI) agar (Oxoid, UK). Single colonies of overnight fresh cultures of all four strains of B. sporothermodurans were inoculated into BHI broth (Oxoid) and incubated at 37°C for 72 hours. Genomic DNA was extracted using the ZR bacterial DNA miniprep kit (Zymo Research, USA) and quantified using the Qubit instrument and doublestranded DNA (dsDNA) broad-range (BR) assay kit (Life Technologies, USA). Multiplexed paired-end libraries were prepared using the Nextera XT DNA sample preparation kit (Illumina, USA). Genome sequencing was carried out on an Illumina MiSeq system. The paired-end reads (2 imes 300 bp) were checked for quality and trimmed and de novo assembled using the CLC genomics workbench version 9

Citation Owusu-Darko R, Allam M, de Oliveira SD, Ferreira CAS, Grover S, Mtshali S, Ismail A, Mallappa RH, Tabit F, Buys EM. 2019. Genome sequences of Bacillus sporothermodurans strains isolated from ultra-high-temperature milk. Microbiol Resour Announc 8:e00145-19. https://doi.org/10.1128/MRA.00145-19.

Editor David A. Baltrus, University of Arizona Copyright © 2019 Owusu-Darko et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Elna M. Buys, elna.buys@up.ac.za.

Received 13 February 2019 Accepted 6 May 2019 Published 30 May 2019

^aDepartment of Consumer and Food Sciences, University of Pretoria, Pretoria, South Africa

^bNational Institute for Communicable Diseases, Sandringham, South Africa

cSchool of Sciences, Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre, Brazil

^dDairy Microbiology Division, Molecular Biology Unit, National Dairy Research Institute, Karnal, India

^eDepartment of Life and Consumer Sciences, University of South Africa, Pretoria, South Africa

TABLE 1 Summary report of the de novo assembly of four B. sporothermodurans strains

	GenBank	SRA	Total no.	Genome	No. of coding	Coverage	No. of	N ₅₀
Organism	accession no.	accession no.	of reads	size (bp)	sequences	(×)	contigs	(kpb)
B. sporothermodurans DSM 10599	NAZD00000000	SRR8741694	3,570,064	3,783,858	4,257	226	527	15,402
B. sporothermodurans SAD	NAZB00000000	SRR8732968	2,523,238	3,857,089	4,111	175	110	114,649
B. sporothermodurans SA01	NAZA00000000	SRR8732969	1,858,252	3,414,010	3,768	146	290	22,386
B. sporothermodurans BR12	NAZC00000000	SRR8741693	3,421,418	3,974,872	4,558	193	805	9,377

(Qiagen, Netherlands), with all low-quality (Q, <20) data filtered out. The resultant contigs were submitted to GenBank, where gene annotation was implemented using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (9). The annotation was further uploaded to Rapid Annotation using Subsystem Technology (RAST) for subsystems-based annotation (10–12).

The four assembled genomes had an average G+C content of 38.6%. Overall, genomes of *B. sporothermodurans* contain heat shock and hyperosmotic proteins (including DnaJ, GrpE, and GroEL) that will have an influence on the heat resistance and, consequently, the processing dynamics of food products. Additionally, all four strains sequenced contain the biofilm matrix protein component TasA and its homologs, which have been shown to be the major biofilm matrix component (13), especially in *Bacillus subtilis*. Ultimately, the whole-genome sequence of *B. sporothermodurans* will help improve our understanding of the heat resistance of this bacterium with the view of improving milk quality.

Data availability. The genome sequences of all four strains of *B. sporothermodurans* are publicly available at NCBI GenBank under the BioProject accession number PRJNA379529. Raw and trimmed sequencing reads have been deposited in the NCBI SRA under the study accession number SRP188520. The GenBank and SRA accession numbers are listed in Table 1. This announcement represents the first version of all four genomes.

ACKNOWLEDGMENTS

R.O.-D. acknowledges support from the Department of Science and Technology (DST), National Research Foundation (NRF), South Africa, in a form of scholarship disbursed through the Institute for Food, Nutrition and Well-being, Faculty of Natural and Agricultural Sciences, University of Pretoria, South Africa. We also acknowledge the HESA/IBSA Research Cooperation Program.

We have no potential conflicts of interest to disclose.

REFERENCES

- Huemer IA, Klijn N, Vogelsang HWJ, Langeveld L. 1998. Thermal death kinetics of spores of Bacillus sporothermodurans isolated from UHT milk. Int Dairy J 8:851–855. https://doi.org/10.1016/S0958-6946(98) 00129-0.
- Klijn N, Herman L, Langeveld L, Vaerewijck M, Wagendorp AA, Huemer I, Weerkamp AH. 1997. Genotypical and phenotypical characterization of Bacillus sporothermodurans strains, surviving UHT sterilisation. Int Dairy J 7:421–428. https://doi.org/10.1016/S0958-6946(97)00029-0.
- Pettersson B, Lembke F, Hammer P, Stackebrandt E, Priest FG. 1996. Bacillus sporothemodurans, a new species producing highly heat-resistant endospores. Int J Syst Bacteriol 46:759–764. https://doi.org/10.1099/00207713-46-3-759.
- Herman L, Heyndrickx M. 2000. The presence of intragenically located REP-like elements in Bacillus sporothermodurans is sufficient for REP-PCR typing. Res Microbiol 151:255–261. https://doi.org/10.1016/S0923 -2508(00)00146-7.
- Krawczyk AO, de Jong A, Holsappel S, Eijlander RT, van Heel A, Berendsen EM, Wells-Bennik MHJ, Kuipers OP. 2016. Genome sequences of 12 sporeforming bacillus species, comprising Bacillus coagulans, Bacillus licheniformis, Bacillus amyloliquefaciens, Bacillus sporothermodurans, and Bacillus vallismortis, isolated from foods. Genome Announc 4:e00103-16. https://doi .org/10.1128/genomeA.00103-16.

- Ki J-S, Zhang W, Qian P-Y. 2009. Discovery of marine Bacillus species by 16S rRNA and rpoB comparisons and their usefulness for species identification. J Microbiol Methods 77:48–57. https://doi.org/10.1016/j .mimet.2009.01.003.
- Pinto CLO, Souza LV, Meloni VAS, Batista CS, Silva R, Martins EMF, Cruz AG, Martins ML. 2018. Microbiological quality of Brazilian UHT milk: identification and spoilage potential of spore-forming bacteria. Int J Dairy Technol 71:20–26. https://doi.org/10.1111/1471-0307.12339.
- Scheldeman P, Pil A, Herman L, De Vos P, Heyndrickx M. 2005. Incidence and diversity of potentially highly heat-resistant spores isolated at dairy farms. Appl Environ Microbiol 71:1480–1494. https://doi.org/10.1128/ AEM.71.3.1480-1494.2005.
- Tatusova T, Dicuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. https://doi.org/10.1093/nar/gkt1226.
- 11. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K,

Volume 8 Issue 22 e00145-19

- Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- 12. Disz T, Akhter S, Cuevas D, Olson R, Overbeek R, Vonstein V, Stevens R,
- Edwards R. a. 2010. Accessing the SEED genome databases via Web services API: tools for programmers. BMC Bioinformatics 11:319. https://doi.org/10.1186/1471-2105-11-319.
- Branda SS, Chu F, Kearns DB, Losick R, Kolter R. 2006. A major protein component of the Bacillus subtilis biofilm matrix. Mol Microbiol 59: 1229–1238. https://doi.org/10.1111/j.1365-2958.2005.05020.x.

Volume 8 Issue 22 e00145-19 mra.asm.org **3**