

***BRCA1* and *BRCA2* pathogenic sequence variants in women of African origin or ancestry**

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Abstract

BRCA1 and *BRCA2* (*BRCA1/2*) pathogenic sequence variants (PSVs) confer elevated risks of multiple cancers. However, most *BRCA1/2* PSVs reports focus on European ancestry individuals. Knowledge of the PSV distribution in African descent individuals is poorly understood. We undertook a systematic review of the published literature and publicly available databases reporting *BRCA1/2* PSVs also accessed the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA) database to identify African or African descent individuals. Using these data, we inferred which of the *BRCA* PSVs were likely to be of African continental origin. Of the 43,817 *BRCA1/2* PSV carriers in the CIMBA database, 469 (1%) were of African descent. Additional African descent individuals were identified in public databases ($n = 291$) and the literature ($n = 601$). We identified 164 unique *BRCA1* and 173 unique *BRCA2* PSVs in individuals of African ancestry. Of these, 83 *BRCA1* and 91 *BRCA2* PSVs are of likely or possible African origin. We observed numerous differences in the distribution of PSV type and function in African origin versus non-African origin PSVs.

Research in populations of African ancestry with *BRCA1/2* PSVs is needed to provide the information needed for clinical management and decision-making in African descent individuals worldwide.

KEYWORDS; African ancestry, *BRCA1*, *BRCA2*, mutation, pathogenic sequence variant

INTRODUCTION

Many women benefit from genetic testing information that can guide their knowledge of cancer risks and help them to make informed prevention and medical management decisions if they have inherited pathogenic sequence variants (PSVs) in *BRCA1* (GenBank reference sequence: NG_005905.2) or *BRCA2* (GenBank reference sequence NG_012772.3; *BRCA1/2*). Knowledge of genetic risk is valuable because it can lead to the use of risk-reducing salpingo-oophorectomy or bilateral mastectomy, which significantly decrease breast and/or ovarian cancer risk and mortality *BRCA1/2* PSV carriers (Domchek et al., 2006; Domchek et al., 2010; Domchek and Rebbeck, 2007; Domchek, Stopfer, and Rebbeck, 2006; Rebbeck, Kauff, and Domchek, 2008; T. R. Rebbeck et al., 2005; Rebbeck, Kauff, and Domchek, 2009). However, the vast majority of information about cancer risk and risk reduction has to date been conducted in Caucasian and Ashkenazi Jewish women (Rebbeck et al., 2018).

Because there has been limited capture of *BRCA1/2* PSVs in more diverse race/ethnicity groups, we anticipate that a large number of PSVs have yet to be reported. Similarly, there is very limited information about cancer risks and the effect of prevention strategies in non-White/non-Jewish women. Therefore, a better understanding of PSVs and cancer risks in underrepresented populations is a critical unmet need in the *BRCA1/2* field.

The *BRCA1/2* International Diversity by Geography and Ethnicity (BRIDGE) Study has been developed to provide information about *BRCA1/2* PSVs in populations who are currently underrepresented in research. We characterize PSVs in self-reported African ancestry (SRAA) individuals identifying commonly reported PSVs that may have had their origins on the African continent and identify potential African founder PSVs and PSV hot spots.

2 METHODS

The individuals included in this analysis were inferred to have African ancestry based on self-report or inference based on place of birth or residence. We included Africans from sub-Saharan Africa and Black South Africans. We also included those reporting African origins from North America, South America, the Caribbean, Europe, or Asia. We excluded White (Afrikaners) and Asian South Africans, mixed-race (colored) South Africans, and individuals who self-reported to be of other race/ethnicity groups whose ancestral origins were from outside Africa (e.g., Asian and Middle Eastern) and North Africans (Morocco, Tunisia, Algeria, Libya, and Egypt). In addition, this report only includes females with PSVs. In some cases, the basis of the race/ethnicity determination was not stated in the original research, and the assumption was made that the source of race/ethnicity information was by self-report or determined by the research team. None of the information about race or ethnicity was based on ancestral genomic information.

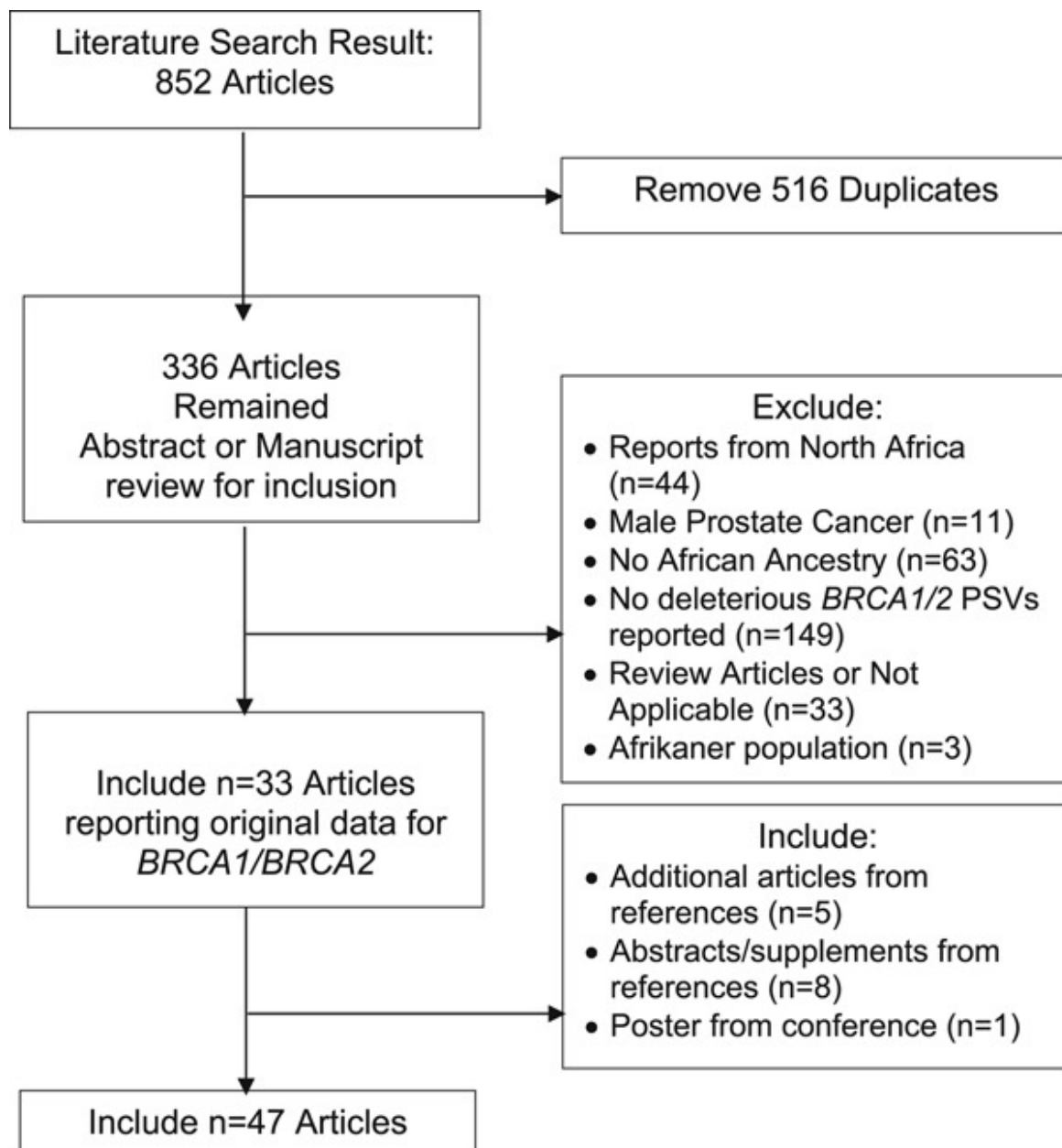


Figure 1. Summary of literature review used in this report for *BRCA1* (GenBank reference sequence: NG_005905.2) or *BRCA2* (GenBank reference sequence NG_012772.3)

To characterize the distribution of PSVs in SRAA individuals, we evaluated a series of data sources (Figure 1). First, we undertook a systematic literature review of published studies reporting a deleterious *BRCA1/2* PSV in an individual of SRAA, including sub-Saharan Africa, Caribbean, and African American. To identify reports of *BRCA1/2* PSVs in SRAA individuals, we used the following keywords to search the medical literature: *BRCA1* or *BRCA2* and Africa or African (includes African American), or the Caribbean. Each paper was reviewed to determine if it described an independent study done in SRAA individuals and reported the presence of a disease-associated PSV in *BRCA1/2*. PSVs reported in other genes were excluded. Reports of PSV testing for *BRCA1/2* that reported no disease-associated PSVs were not included in this report. Reports of variants of unknown significance were also excluded. All studies were included regardless of sampling design (e.g., family studies,

population screening, genetic testing in clinical case series, etc.). In addition, a small number of papers or abstracts were identified from the citation list of the papers that were identified from the public literature search.

Second, we obtained data from the CIMBA consortium (Chenevix-Trench et al., 2007) using the coded ethnicity variable for African American and countries in Sub-Saharan Africa, which included reports from Nigeria and South Africa only. White South Africans, Afrikaners, and mixed-race (colored) South Africans were excluded.

Third, we used publicly available data from the BIC (<https://research-nhgri-nih.gov.uplib.idm.oclc.org/bic/>) database. A copy of the BIC database was downloaded on February 1, 2019 to identify reported *BRCA1/2* PSV carriers. The ethnicity and nationality variables were searched for any notation of African descent individuals, African, African American, or the Caribbean.

Lastly, the NIH public database ClinVar (<https://www.ncbi.nlm.nih.gov.uplib.idm.oclc.org/clinvar/>) was used as a check for PSV nomenclature and confirmation of the pathogenic status of each variant. CIMBA and BIC, among other sources, submitted their PSVs to ClinVar. The majority of sources did not contribute race/ethnicity information to ClinVar, so this database was determined not to be a useful data source for this report. Most PSVs reported here have been deposited into the ClinVar database.

PSVs were categorized according to type and function, including large deletion (DL), large duplication (DP), frameshift (FS), in-frame deletion (IFD), missense (MS), nonsense (NS), splice (SP), and individuals who carried multiple PSV types (including those listed above); No RNA, premature termination codon (PTC), nonsense-mediated decay (NMD), NMD with or without reinitiation; and mutation class (1, 2, or 3).

To establish the continental origin of *BRCA1/2* PSVs that were reported in SRAA individuals, we identified PSVs reported in regions in which non-African admixture was likely (i.e., North America, Caribbean, South America, South Africa) and where non-African admixture was less likely (i.e., Sub-Saharan Africa excluding South Africa). Table S1 presents PSVs reported only in Sub-Saharan Africa and PSVs reported only in the Americas. In addition, we evaluated whether any of these PSVs were reported in non-African ancestry groups (e.g., Europeans or European Americans). We judged a PSV to be likely African origin if the PSV has only been reported in SRAA and never in a non-SRAA individual, and of possibly African origin if reported greater than 50% of the time in SRAA individuals. This inference was strengthened by the observation of a PSV reported in Africa. We judged the PSV to be likely non-African origin if it reported less than 50% of the time in SRAA individuals.

Collection of original data was undertaken under approved research and/or clinical human subjects protocols at each contributing center. Retrospective anonymized data analysis for the present research was undertaken under human subjects approvals at the Dana-Farber Cancer Institute.

3 RESULTS

3.1 African ancestry individuals with *BRCA1/2* pathogenic sequence variants

The results of our literature search are shown in Figure 1. When more than one paper from a research group was found, we included the largest or most recent in the series. Studies that searched for but did not find any *BRCA1/2* PSVs in SRAA women were excluded. Note that only 11 published studies were found that included male *BRCA1/2* PSV carriers, but no additional *BRCA1/2* PSVs were identified from these reports. Forty-eight studies were included in the present report: 16 studies of Sub-Saharan Africans (Awadelkarim et al., 2007; Biunno et al., 2014; Diez et al., 2011; Elimam et al., 2017; Fackenthal et al., 2005; Fackenthal et al., 2012; Francies et al., 2015; Gao et al., 2000; Luyeye Mvila et al., 2014; Stoppa-Lyonnet et al., 1997; van der Merwe et al., 2012; Zhang et al., 2009; Zhang et al., 2012; Zhang, Fackenthal, Huo, Zheng, & Olopade, 2010; Zheng et al., 2018; Zoure et al., 2018), 27 of African Americas (Arena et al., 1996; Arena et al., 1997; Arena et al., 1998; Castilla et al., 1994; Churpek et al., 2015; Dangel et al., 1999; Futreal et al., 1994; Ganguly, Dhulipala, Godmilow, & Ganguly, 1998; Q. Gao, Neuhausen, Cummings, Luce, & Olopade, 1997; Q. Gao, Sveen, Cummings, & Olopde, 1998; Q. Gao et al., 2000; Gayol et al., 1999; Haffty et al., 2009; Hall et al., 2009; John et al., 2007; Kanaan et al., 2003; Kedar-Barnes et al., 2000; Lynce et al., 2015; Martin et al., 2009; Miki et al., 1994; Nanda et al., 2005; Olopade et al., 2003; Pal et al., 2008; Pal et al., 2015; Pal, Permeth-Wey, Holtje, & Sutphen, 2004; Panguluri et al., 1999; Shen et al., 2000; Sutphen & Ferlita, 1999; Whitfield-Broome, Dunston, & Brody, 1999), and three of Afro-Caribbean populations (Akbari et al., 2014; Donenberg et al., 2011; Donenberg et al., 2016) and one study reporting SRAA of unspecified geography (Hall et al., 2009). From these papers, we identified 414 *BRCA1* and 187 *BRCA2* PSVs, and 108 unique *BRCA1* and 103 unique *BRCA2* PSVs.

The BIC database included 15,311 total submissions of *BRCA1* PSVs, of which 8,564 were reported to be pathogenic. Of these, 206 (2.4%) were identified as SRAA. The BIC database included 14,914 total submissions of *BRCA2* PSVs, of which 4,516 were deleterious. Of these, 85 (1.9%) identified as SRAA.

The CIMBA database consisted of 43,817 *BRCA1/2* female PSV carriers. Of these, 382 (0.9%) were identified as African Americans and 269 (0.6%) were ascertained in Africa. Thirty-two of these carriers were from Nigeria and 237 were from South Africa. After excluding White South Africans, Afrikaaners, or mixed-race (Colored) South Africans, 11 Black South Africans were included. In total, 435 (1%) women of African descent were found in the CIMBA database. Of these 273 were *BRCA1* and 162 were *BRCA2*. In addition, 34 African American carriers from the University of Illinois at Chicago, 14 *BRCA1*, and 20 *BRCA2*, not previously reported to CIMBA, were also included in the CIMBA dataset after consultation with the University of Illinois center.

3.2 *BRCA1/2* PSVs in African ancestry individuals

Table s1 presents the complete list of all SRAA PSVs identified in this study and relevant characteristics according to PSV designation, type, and function as well as where and in what populations they have been reported. From our three primary data sources, a *BRCA1*

PSV in SRAA was reported 909 times: 404 (44.3%) in African Americans, 170 (18.6%) in Africans, 135 (14.8%) in the Caribbean, and 200 (21.9%) in SRAA of unknown or unreported geographic origin. From all data sources, we identified 164 unique *BRCA1* PSVs. For *BRCA2*, a PSV in a woman of SRAA was reported 454 times, 262 (57.7%) in African Americans, 114 (25.1%) in Africans, 35 (7.7%) in the Caribbean, and 43 (9.5%) in SRAA of unknown origin. Of these, we identified 173 unique *BRCA2* PSVs. We were not able to determine if an individual was included more than once in these data sources, so these figures do not represent the actual frequency of PSVs expected in SRAA.

The most commonly identified PSVs in *BRCA1* and *BRCA2* are presented in Table 1. Most of these PSVs have been reported in multiple geographical locations, and in particular, most have been reported in both the New and Old World. A small number of PSVs to date ($n = 39$) have been reported only in Africans residing in Africa.

In order to identify PSVs that were most likely to be of African origin, we characterized PSVs according to the number of times they were reported in individuals of African and non-African descent (Table 2). We characterized those being reported only in SRAA populations and/or only in Africa as being of likely African origin ($n = 115$), those reported >50–99% as possibly African origin ($n = 59$), those 25–50% as probably non-African ($n = 35$), and those reported <25% of the time in SRAA individuals as likely non-African origin ($n = 128$). For *BRCA1*, 50 (30.5%) PSVs are likely to be of African origin, 33 (20.1%) possibly of African origin, 19 (11.6%) probably non-African, and 62 (37.8%) likely of non-African origin. For *BRCA2*, 65 (37.6%) PSVs are likely to be of African origin, 26 (15.0%) possibly of African origin, 16 (9.2%) probably non-African, and 66 (38.2%) likely of non-African origin. The list of these PSVs and their corresponding designations is presented in Table S1. Thus, about half of PSVs identified in SRAA can be inferred to be of likely or possibly African origin.

Table 2. Determination of continent of origin for PSVs in *BRCA1* (GenBank reference sequence: NG_005905.2) or *BRCA2* (GenBank reference sequence NG_012772.3) Identified in SRAA individuals, including likely African (100%), possibly African (50–99%), unlikely African (25–49%), and likely non-African (<25%)

Determination	<i>BRCA1</i>		<i>BRCA2</i>		Total
	<i>n</i>	%	<i>n</i>	%	
Likely African	50	30.5%	65	37.6%	115
Possibly African	33	20.1%	26	15.0%	59
Unlikely African	19	11.6%	16	9.2%	35
Likely non-African	62	37.8%	66	38.7%	128
Total	164	100%	173	100%	337

Abbreviations: PSV, pathogenic sequence variant; SRAA, self-reported African ancestry.

Figures 2 and 3 display the frequency and distribution of likely or possible African origin PSVs across the span of each gene. For *BRCA1*, two common PSVs (c.5324T>G and c.4357+1G>A) were observed in multiple populations of African descent. Among those identified as unlikely to be of African origin include the c.68_69del Jewish founder PSV, as well as PSVs that have been reported as common or founder PSVs in European populations (e.g., c.211A>G in Spain and c.5251C>T in Greece (Janavičius, 2010) or in the CIMBA database (e.g., c.1504_1509del in Asians in Australia and c.5177_5180del in French Caucasians). Notably, the c.68_69del PSV was found in an African population in Burkina Faso

Table 1. Ten most common pathogenic sequence variants in *BRCA1* (GenBank reference sequence: NG_005905.2) or *BRCA2* (GenBank reference sequence NG_012772.3) in self-reported African ancestry (SRAA) individuals

Gene	PSV	Total PSV in SRAA	All reported PSVs	% SRAA PSVs of total PSVs reported	Countries in which PSV has been reported in SRAA individuals	Continent on which PSV has been reported in non-SRAA individuals
<i>BRCA1</i>	c.5324T>G	103	125	82%	Bahamas, Nigeria, Trinidad, USA	North America, Europe
	c.815_824dup	100	155	65%	Senegal, Bahamas, Ivory Coast, France, USA, Spain, France	North America, Europe, South America
	c.4357+1G>A	73	88	83%	Bahamas, USA	North America, Europe
	c.5177_5180del	54	142	38%	USA, Canada, Trinidad, Nigeria	North America, Europe
	c.4986+6T>C	35	45	78%	Bahamas, USA, Sudan	North America, Europe, South America
	c.68_69del	34	4834	1%	Bahamas	North America, Europe, South America, Australia, Asia
	c.303T>G	26	34	76%	Nigeria	North America, Europe
	c.190T>G	19	30	63%	USA	North America, Europe
	c.5467+1G>A	16	25	64%	USA	North America, Europe, Asia
	c.5251C>T	15	188	8%	USA	North America, Europe, South America, Australia, Asia
	c.1310_1313del	26	192	14%	Bahamas, Nigeria, Trinidad, USA	North America, Europe, Australia, Asia
	c.9382C>T	23	205	11%	Trinidad, USA	North America, Europe, Caribbean, Australia, Asia
	c.2808_2811del	22	646	3%	Nigeria, Brazil, USA	North America, Europe, South America, Australia, Asia
c.5616_5620del	18	23	78%	USA	North America, Europe	
<i>BRCA2</i>	c.658_659del	12	217	6%	USA	North America, Europe, Caribbean, Australia, Asia
	c.4552del	11	12	92%	USA	North America
	c.9253dup	10	69	14%	USA	North America, Europe, Asia
	c.5771_5774del	9	18	50%	South Africa	North America, Europe
	c.2402_2412del	7*	7	100%	Nigeria	
	c.2957_2958insG	7*	8	88%	USA, Nigeria	North America
	c.6405_6409del	7*	131	5%	USA, Brazil, Nigeria	North America, Europe, South America, Australia, Asia

(Zoure et al., 2018) as well as many other populations in Europe, the Middle East, and North Africa. In *BRCA2*, c.5616_5620del was observed in multiple African descent populations. As with *BRCA1*, the most common PSVs that were inferred as unlikely to be of African origin have been reported as common or founder PSVs in European populations (e.g., c.2808_2811del in Spain and c.1310_1313del in Denmark, or in the CIMBA database (e.g., c.658_699del in Germany and c.9382C>T globally, including Japan and in the CIMBA database in Europe; Arai et al., 2018; Janavičius, 2010).

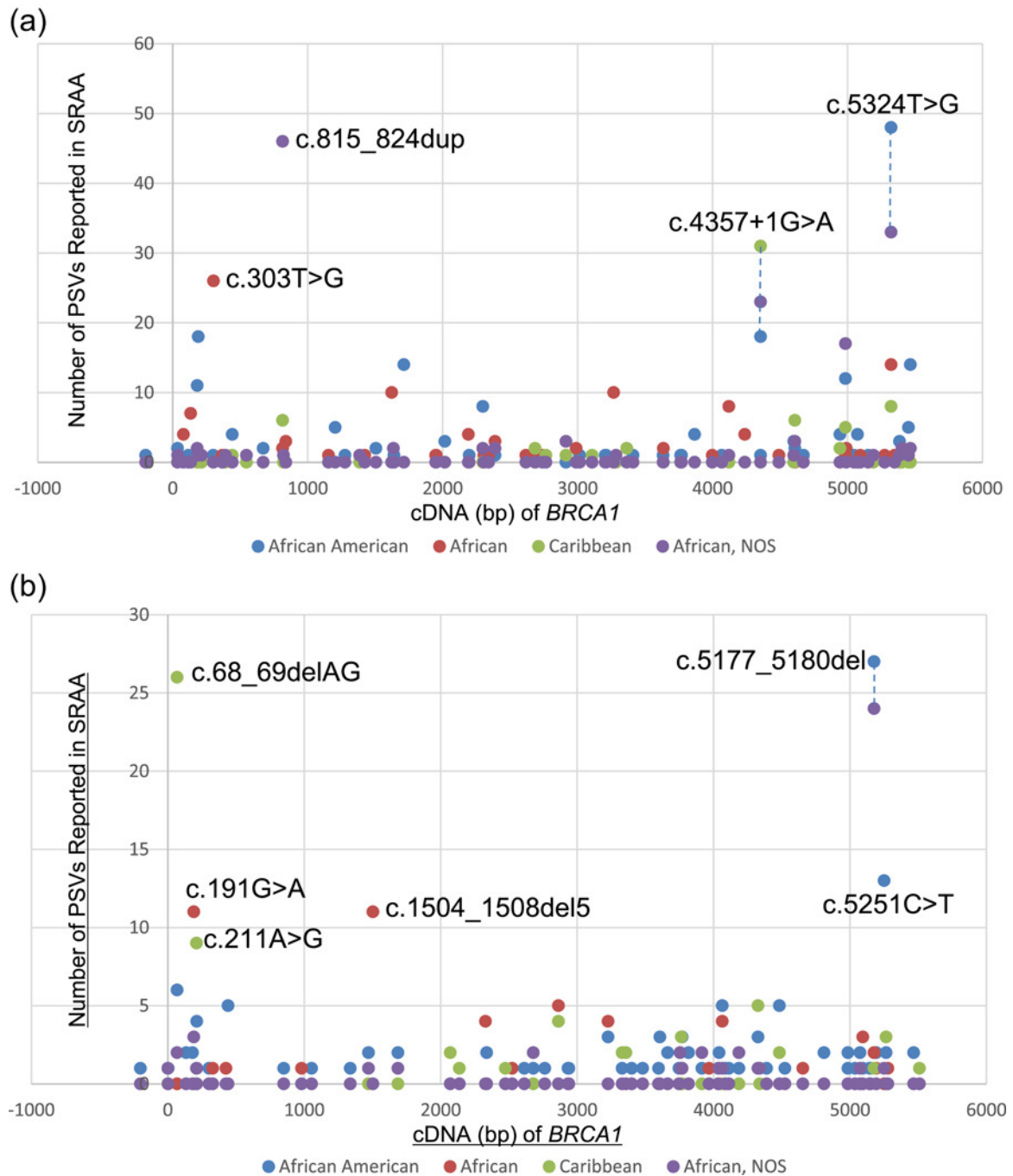


Figure 2. *BRCA1* (GenBank reference sequence: NG_005905.2) pathogenic sequence variants (PSVs) by determination of African origin and self-identification. (a) Likely or possible African origin and (b) unlikely African origin

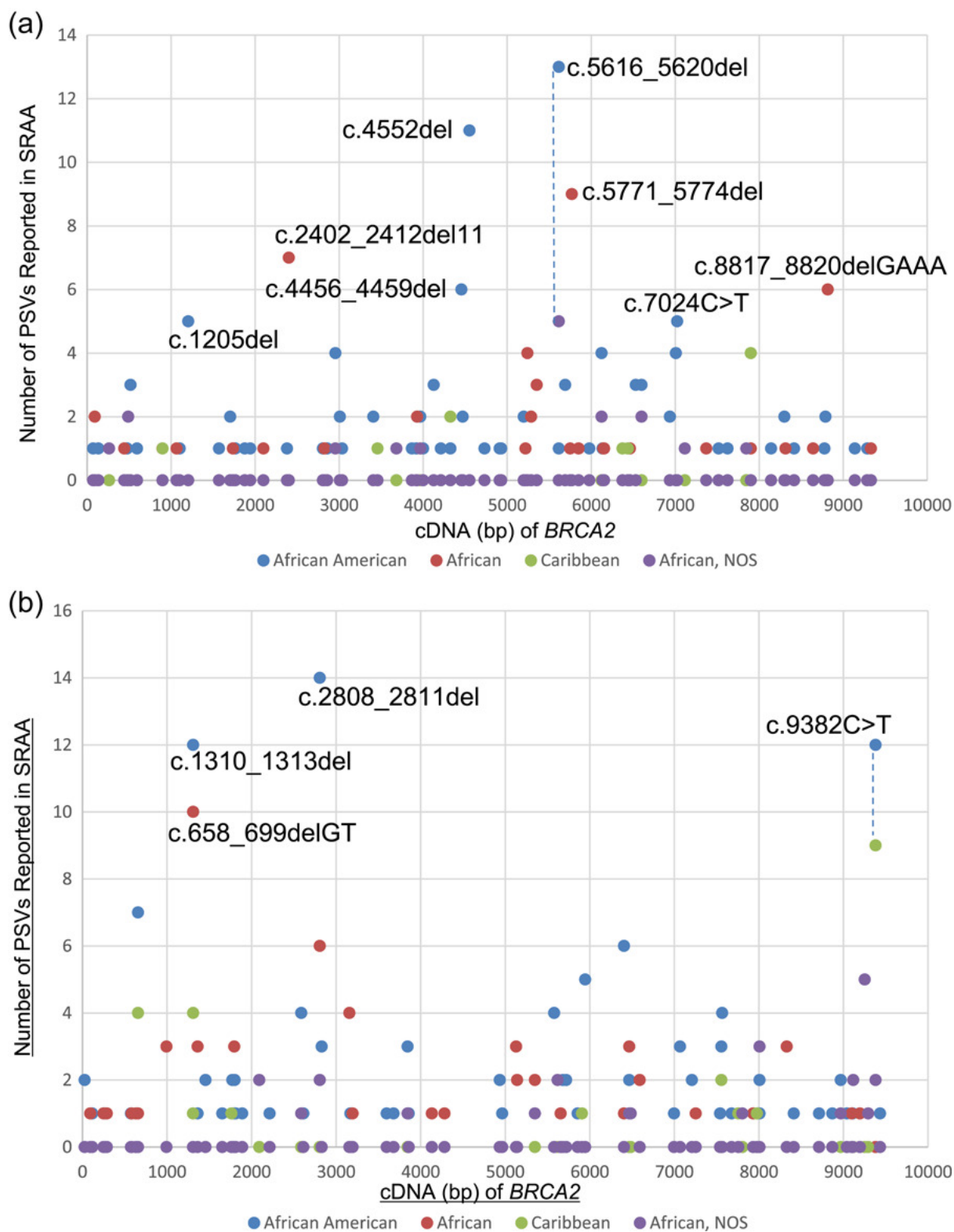


Figure 3. *BRCA2* (GenBank reference sequence NG_012772.3) PSVs by determination of African origin and self-identification. (a) Likely or possible African origin and (b) unlikely African origin. PSV, pathogenic sequence variant

Table 3. Characteristics of *BRCA1* (GenBank reference sequence: NG_005905.2) PSVs in the CIMBA Database (T. R. Rebbeck et al., 2018), by unique PSV, compared to likely or possible PSVs of African origin

		<i>n</i> = 1,650 CIMBA PSVs (All ethnicities)		<i>n</i> = 164 likely or possibly African PSVs		
		<i>n</i>	%	<i>n</i>	%	<i>p</i>-value*
	Large deletion (DL)	130	7.9	10	6.1	.749
	Large duplication (DP)	27	1.6	2	1.2	>.999
	Frameshift (FS)	948	57.5	66	40.2	<.0001
PSV type	In-frame deletion (IFD)	1	<0.1	4	2.4	<.0001
	Missense (MS)	46	2.8	9	5.5	.078
	Nonsense (NS)	313	19.0	40	24.4	.081
	Splice (SP)	166	10.1	20	12.2	.259
	Multiple types (including those listed above)/Unknown	20	1.1	13	7.9	<.0001
PSV effect	No RNA	21	1.3	5	3.0	.055
	Premature termination codon (PTC)	1,331	81.0	95	57.9	<.0001
	Unknown/Other	298	18.0	64	39.0	<.0001
	Nonsense-mediated decay (NMD)**	1,213	73.9	84	51.2	<.0001
	No NMD	58	3.5	2	1.2	.229
	Reinitiation	4	0.2	2	1.2	.080
	NMD/Reinitiation	60	3.7	8	4.9	.492
	Unknown/Other	294	17.8	68	41.5	<.0001
PSV class	1	1,298	78.6	99	60.6	.756
	2	112	6.8	10	6.1	
	3	240	14.6	19	11.6	
	Unknown			36	22.0	

Abbreviations: PSV, pathogenic sequence variant; SRAA, self-reported African ancestry.

* *p*-values from Fisher's exact test reflect the comparison of all ethnicities versus likely or possible African PSVs, for the specific mutation group versus all other mutations.

** References (Anczukow et al., 2008; Buisson, Anczukow, Zetoune, Ware, & Mazoyer, 2006; Mikaelssdottir, Valgeirsdottir, Eyfjord, & Rafnar, 2004; Perrin-Vidoz, Sinilnikova, Stoppa-Lyonnet, Lenoir, & Mazoyer, 2002; Ware et al., 2006).

Table 4. Characteristics of *BRCA2* (GenBank reference sequence NG_012772.3) PSVs in the CIMBA database (T. R. Rebbeck et al., 2018), by unique PSV, compared to likely and possible PSVs of African origin

	<i>n</i> = 1,731 CIMBA PSVs (All ethnicities)		<i>n</i> = 173 likely or possibly African PSVs		
	<i>n</i>	%	<i>n</i>	%	<i>p</i> -value*
PSV type					
Large deletion (DL)	34	1.9	1	0.01	.003
Large duplication (DP)	11	0.6	0	0.0	NE
Frameshift (FS)	1,141	65.9	106	61.3	<.0001
In-frame deletion (IFD)	2	0.1	0	0.0	NE
Missense (MS)	13	0.8	1	0.01	NE
Nonsense (NS)	380	22.0	47	27.2	.617
Splice (SP)	131	7.6	8	4.6	.062
Multiple types (including those listed above)	19	1.1	10	5.8	<.0001
PSV effect					
No RNA	6	0.3	0	0.0	NE
Premature termination codon (PTC)	1,542	89.0	132	76.3	<.0001
Unknown/Other	183	10.6	41	23.7	<.0001
Nonsense-mediated decay (NMD)**	1,523	88.0	132	76.3	<.0001
No NMD	16	0.9	0	0	NE
Reinitiation	0	0.0	0	0	NE
NMD/Reinitiation	0	0.0	0	0	NE
Unknown/Other	187	10.7	41	23.7	<.0001
PSV class					
1	1,529	88.3	131	75.7	<.0001
2	36	2.1	0	0	
3	167	9.6	6	3.5	
Unknown			36	20.8	

Abbreviations: NE, not estimable; PSV, pathogenic sequence variant; SRAA, self-reported African ancestry.

* *p*-values from Fisher's exact test reflect the comparison of all ethnicities versus likely or possible African PSVs, for the specific mutation group versus all other mutations.

** References (Anczukow et al., 2008; Buisson et al., 2006; Mikaelsdottir et al., 2004; Perrin-Vidoz et al., 2002; Ware et al., 2006).

Using the SRAA CIMBA PSV carriers, we were also able to characterize the distribution of PSVs by type and effect (Tables 3 and 4). FS, PTC, and NMD were less common in SRAA than in CIMBA overall for both *BRCA1* and *BRCA2*. IFD was more common in *BRCA1* SRAA (but this included very few PSVs), and PSV class was different between SRAA and non-SRAA in *BRCA2*. For *BRCA1*, large deletions and duplications were not significantly different in frequency in CIMBA SRAA versus the previously reported CIMBA distribution (T. R. Rebbeck et al., 2018). There was also no significant difference for splice variants, PSV class, or lack of NMD with reinitiation, and marginally significant differences with missense PSVs, nonsense PSVs, no RNA, or reinitiation. Other PSV types or functions included frameshift, in-frame deletions, PSVs of multiple types, premature termination codon, and NMD. There were also significant differences in the number of PSVs with unknown PSV effect. There were approximately twice as many SRAA PSVs of an unknown effect than in other groups.

For *BRCA2*, there were significant differences between SRAA and the published CIMBA PSV distribution for large deletions, frameshift, in-frame deletions, missense, and multiple PSV types. There were also statistically significant differences for all PSV effects including premature termination codon PSVs, NMD, and PSV class. Again, the proportion of PSVs of unknown effect was significantly higher in SRAA than CIMBA as a whole.

4 DISCUSSION

Despite the availability of substantial information regarding cancer risks in *BRCA1/2* PSV carriers, the amount of information available to women of African ancestry is extremely limited. The worldwide CIMBA database containing over 30,000 families with *BRCA1/2* PSVs includes less than 1% of individuals of African ancestry (T. R. Rebbeck et al., 2016). This very limited capture of PSVs in women of African ancestry presents a number of important limitations for the clinical implementation and decision-making for African ancestry women (Olopade et al., 2003; Oluwagbemiga, Oluwole, & Kayode, 2012).

We summarize all reported PSVs found in SRAA individuals and infer which of these PSVs may be of African origin. Approximately half of PSVs reported in SRAA are likely or possibly of African origin. A number of individuals who are SRAA have inherited PSVs of European origin. These PSVs were observed primarily in individuals in North America or the Caribbean and likely reflect recent European admixture. However, the c.68_69del PSV was also commonly observed in all SRAA populations. Haplotype analyses suggest that this PSV arose no more than 1500 years ago, suggesting that it was transmitted from the middle east to Africa, and did not arise there before the original migration of individuals out of Africa (Laitman et al., 2013). However, this PSV has been reported in native Africans (Zoure et al., 2018) and has arisen in multiple populations independently, so it is not known whether this PSV is a new PSV at a locus that demonstrates high mutation rates, or if it is the common Jewish founder PSV, which has been observed in Middle Eastern or North African populations (Laitman et al., 2013; Slaoui et al., 2014; Zoure et al., 2018).

We also report that the distribution of PSV type or effect differs for some PSV groups between SRAA and the CIMBA population as a whole. It is well established that African women, as well as African American women, have an earlier average age of diagnosis and a higher proportion of hormone receptor negative and triple negative tumors than women of

other ethnicities (American Cancer Society, 2016; Vanderpuye et al., 2017). It is not clear how much of this phenotypic pattern in African descent women could be explained by PSVs in *BRCA1/2*, but it is possible that, at least in Africa, a substantial proportion of breast cancer may be hereditary in nature. To the degree this is true, the observation in this report that the distribution of PSV type or effect may be different in African descent populations compared with other populations could have clinical implications for assessing cancer risk in women of SRAA. It is also possible that the penetrance associated with mutations in SRAA is different than that of non-SRAA populations. However, the currently available data do not allow us to evaluate the relationship of PSVs in *BRCA1/2*, as well as specific types of PSVs, in cancer susceptibility. Reports in the literature do not provide consistent information about individual women and their cancer status that would be required to undertake association studies. Such associations could be undertaken in the CIMBA study, but the sample size of SRAA in the CIMBA dataset remains underpowered for these analyses. In addition, the relatively larger proportion of PSVs that could not be characterized with respect to their effect compared with all PSV carriers suggests that additional research is required to understand the phenotypic consequences of *BRCA1/2* PSVs in SRAA.

We have also made initial ad hoc inferences about the likelihood that each PSV reported in SRAA is of African origin. We used a value of 50% of PSVs being observed in SRAA to suggest that the PSV was of African origin. However, given that the number of individuals tested for *BRCA1/2* is much higher in non-Africans, it is possible this number is too conservative. If a PSV exists in both African women and non-African women regardless of percentage, the PSV may have arisen prior to the initiation of migration out of Africa 200,000–300,000 years ago. Mutations found only in SRAA women may represent new mutations that arose in Africa (i.e., since the migration out of Africa) or arose in populations that did not migrate out of Africa. In the future, the estimation of haplotype backgrounds and PSV age will be important to further characterize the origin of PSV with respect to African versus non-African origin.

It is clear from our findings that there is a great need for increased participation among currently underserved populations in *BRCA1/2* research. Large-scale genomic studies of *BRCA1/2* in Africa have yet to be undertaken, although initiatives such as H3Africa (<https://h3africa.org/initiative>) may provide relevant data in the future if *BRCA1/2* testing is undertaken by participating groups. Participation of these groups in *BRCA1/2* research will provide a more comprehensive evaluation of PSVs in non-European descent populations and improve risk assessment and an understanding of risk modifiers. It will be critical to evaluate in these groups the complete PSV spectrum, common/founder PSVs (that may aid in developing efficient genetic testing panels) and understand the origins of these PSVs through population/evolutionary genetics research. Additional research will be required to understand cancer risk and risk modifiers so that cancer prevention and treatment can be optimized in these populations.

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REFERENCES

Akbari, M., Donenberg, T., Lunn, J., Curling, D., Turnquest, T., Krill-Jackson, E., ... Hurley, J. (2014). The spectrum of BRCA1 and BRCA2 mutations in breast cancer patients in the Bahamas. *Clinical Genetics*, 85(1), 64– 67.

<https://doi-org.uplib.idm.oclc.org/10.1111/cge.12132>

American Cancer Society (2016). Cancer Facts & Figures for African Americans 2016-2018. Retrieved from Atlanta, GA.

Anczukow, O., Ware, M. D., Buisson, M., Zetoune, A. B., Stoppa-Lyonnet, D., Sinilnikova, O. M., & Mazoyer, S. (2008). Does the nonsense-mediated mRNA decay mechanism prevent the synthesis of truncated BRCA1, CHK2, and p53 proteins? *Human Mutation*, 29(1), 65– 73.

<https://doi-org.uplib.idm.oclc.org/10.1002/humu.20590>

Arai, M., Yokoyama, S., Watanabe, C., Yoshida, R., Kita, M., Okawa, M., & Nakamura, S. (2018). Genetic and clinical characteristics in Japanese hereditary breast and ovarian cancer: First report after establishment of HBOC registration system in Japan. *Journal of Human Genetics*, 63(4), 447– 457. <https://doi-org.uplib.idm.oclc.org/10.1038/s10038-017-0355-1>

Arena, J., Baumbach, L., Smith, S., Gayol, L., Perera, E., & Lubs, H. (1998). BRCA1 mutation analysis in 20 at-risk African-American families supports a low frequency of germ-line mutation. *American Journal of Human Genetics* (A62), 325.

Arena, J., Smith, S., Plewinska, M., Gayol, L., Perera, E., Murphy, P., & Lubs, H. (1996). BRCA1 mutations in African-American women. *American Journal of Human Genetics* (A34), 169.

Arena, J., Smith, S., Vincek, V., Gayol, L., Villegas, F., Perera, E., & Lubs, H. (1997). A BRCA1 founder mutation in African Americans. *American Journal of Human Genetics*, 61(Suppl), A14.

Awadelkarim, K. D., Aceto, G., Veschi, S., Elhaj, A., Morgano, A., Mohamedani, A. A., ... Mariani-Costantini, R. (2007). BRCA1 and BRCA2 status in a Central Sudanese series of

breast cancer patients: Interactions with genetic, ethnic and reproductive factors. *Breast Cancer Research and Treatment*, 102(2), 189– 199.
<https://doi-org.uplib.idm.oclc.org/10.1007/s10549-006-9303-z>

Biunno, I., Aceto, G., Awadelkarim, K. D., Morgano, A., Elhaj, A., Eltayeb, E. A., ... Mariani-Costantini, R. (2014). BRCA1 point mutations in premenopausal breast cancer patients from Central Sudan. *Familial cancer*, 13(3), 437– 444.
<https://doi-org.uplib.idm.oclc.org/10.1007/s10689-014-9717-4>

Buisson, M., Anczukow, O., Zetoune, A. B., Ware, M. D., & Mazoyer, S. (2006). The 185delAG mutation (c.68_69delAG) in the BRCA1 gene triggers translation reinitiation at a downstream AUG codon. *Human Mutation*, 27(10), 1024– 1029.
<https://doi-org.uplib.idm.oclc.org/10.1002/humu.20384>

Castilla, L. H., Couch, F. J., Erdos, M. R., Hoskins, K. F., Calzone, K., Garber, J. E., ... Weber, B. L. (1994). Mutations in the BRCA1 gene in families with early-onset breast and ovarian cancer. *Nature Genetics*, 8(4), 387– 391.
<https://doi-org.uplib.idm.oclc.org/10.1038/ng1294-387>

Chenevix-Trench, G., Milne, R. L., Antoniou, A. C., Couch, F. J., Easton, D. F., Goldgar, D. E., & CIMBA (2007). An international initiative to identify genetic modifiers of cancer risk in BRCA1 and BRCA2 mutation carriers: The Consortium of Investigators of Modifiers of BRCA1 and BRCA2 (CIMBA). *Breast Cancer Research*, 9(2), 104.
<https://doi-org.uplib.idm.oclc.org/10.1186/bcr1670>

Churpek, J. E., Walsh, T., Zheng, Y., Moton, Z., Thornton, A. M., Lee, M. K., ... Olopade, O. I. (2015). Inherited predisposition to breast cancer among African American women. *Breast Cancer Research and Treatment*, 149(1), 31– 39.
<https://doi-org.uplib.idm.oclc.org/10.1007/s10549-014-3195-0>

Dangel, J., Wagner-Costalas, J., Bove, B., Vanderveer, L., Itzen, M., Daly, M., & Godwin, A. K. (1999). Novel germline BRCA1 mutation (155del4) in an African American with early-onset breast cancer. *Human Mutation*, 14(6), 545– 545.

Diez, O., Pelegrí, A., Gadea, N., Gutiérrez-Enríquez, S., Masas, M., Tenés, A., ... Graña, B. (2011). Novel BRCA1 deleterious mutation (c.1949_1950delTA) in a woman of Senegalese descent with triple-negative early-onset breast cancer. *Oncology Letters*, 2(6), 1287– 1289.
<https://doi-org.uplib.idm.oclc.org/10.3892/ol.2011.390>

Domchek, S. M., Friebel, T. M., Neuhausen, S. L., Wagner, T., Evans, G., Isaacs, C., ... Rebbeck, T. R. (2006). Mortality after bilateral salpingo-oophorectomy in BRCA1 and BRCA2 mutation carriers: A prospective cohort study. *The Lancet Oncology*, 7(3), 223– 229.
[https://doi-org.uplib.idm.oclc.org/S1470-2045\(06\)70585-X](https://doi-org.uplib.idm.oclc.org/S1470-2045(06)70585-X) [pii] 10.1016/S1470-2045(06)70585-X

Domchek, S. M., Friebel, T. M., Singer, C. F., Evans, D. G., Lynch, H. T., Isaacs, C., ... Rebbeck, T. R. (2010). Association of risk-reducing surgery in BRCA1 or BRCA2 mutation carriers with cancer risk and mortality. *JAMA*, 304(9), 967– 975.
<https://doi-org.uplib.idm.oclc.org/10.1001/jama.2010.1237>

Domchek, S. M., & Rebbeck, T. R. (2007). Prophylactic oophorectomy in women at increased cancer risk. *Current Opinion in Obstetrics and Gynecology*, 19(1), 27– 30.
<https://doi-org.uplib.idm.oclc.org/10.1097/GCO.0b013e32801195da>

Domchek, S. M., Stopfer, J. E., & Rebbeck, T. R. (2006). Bilateral risk-reducing oophorectomy in BRCA1 and BRCA2 mutation carriers. *Journal of the National Comprehensive Cancer Network: JNCCN*, 4(2), 177– 182.

Donenberg, T., Ahmed, H., Royer, R., Zhang, S., Narod, S. A., George, S., ... Hurley, J. (2016). A survey of BRCA1, BRCA2, and PALB2 mutations in women with breast cancer in Trinidad and Tobago. *Breast Cancer Research and Treatment*, 159(1), 131– 138.
<https://doi-org.uplib.idm.oclc.org/10.1007/s10549-016-3870-4>

Donenberg, T., Lunn, J., Curling, D., Turnquest, T., Krill-Jackson, E., Royer, R., ... Hurley, J. (2011). A high prevalence of BRCA1 mutations among breast cancer patients from the Bahamas. *Breast Cancer Research and Treatment*, 125(2), 591– 596.
<https://doi-org.uplib.idm.oclc.org/10.1007/s10549-010-1156-9>

Elimam, A. A., Aabdein, M. E. M. M., Eldeen, M. E. M., Altayb, H. N., Taha, M. A., Nimir, M. N., ... Hassan, M. A. S. (2017). Monoallelic characteristic-bearing heterozygous L1053X in BRCA2 gene among Sudanese women with breast cancer. *BMC Medical Genetics*, 18(1), 85.
<https://doi-org.uplib.idm.oclc.org/10.1186/s12881-017-0448-x>

Fackenthal, J. D., Sveen, L., Gao, Q., Kohlmeir, E. K., Adebamowo, C., Ogundiran, T. O., ... Olopade, O. I. (2005). Complete allelic analysis of BRCA1 and BRCA2 variants in young Nigerian breast cancer patients. *Journal of Medical Genetics*, 42(3), 276– 281.

Fackenthal, J. D., Zhang, J., Zhang, B., Zheng, Y., Hagos, F., Burrill, D. R., ... Olopade, O. I. (2012). High prevalence of BRCA1 and BRCA2 mutations in unselected Nigerian breast cancer patients. *International Journal of Cancer*, 131(5), 1114– 1123.
<https://doi-org.uplib.idm.oclc.org/10.1002/ijc.27326>

Francies, F. Z., Wainstein, T., DeLeeneer, K., Cairns, A., Murdoch, M., Nietz, S., ... Claes, K. B. (2015). BRCA1, BRCA2 and PALB2 mutations and CHEK2 c.1100delC in different South African ethnic groups diagnosed with premenopausal and/or triple negative breast cancer. *BMC Cancer*, 15, 912. <https://doi-org.uplib.idm.oclc.org/10.1186/s12885-015-1913-6>

Futreal, P., Liu, Q., Shattuck-Eidens, D., Cochran, C., Harshman, K., Tavtigian, S., ... Miki, Y. (1994). BRCA1 mutations in primary breast and ovarian carcinomas. *Science*, 266(5182), 120– 122.

Ganguly, T., Dhulipala, R., Godmilow, L., & Ganguly, A. (1998). High throughput fluorescence-based conformation-sensitive gel electrophoresis (F-CSGE) identifies six unique BRCA2 mutations and an overall low incidence of BRCA2 mutations in high-risk BRCA1-negative breast cancer families. *Human Genetics*, 102(5), 549– 556.

Gao, Q., Neuhausen, S., Cummings, S., Luce, M., & Olopade, O. I. (1997). Recurrent germline BRCA1 mutations in extended African American families with early-onset breast cancer. *American Journal of Human Genetics*, 60(5), 1233– 1236.

Gao, Q., Sveen, L., Cummings, S., & Olopade, O. (1998). Contribution of recurrent BRCA1 (B1) and BRCA2 (B2) mutations to breast cancer in African Americans (AA) women. *Proceedings of the American Association for Cancer Research*, 39, 475– 491.

Gao, Q., Tomlinson, G., Das, S., Cummings, S., Sveen, L., Fackenthal, J., ... Olopade, O. I. (2000). Prevalence of BRCA1 and BRCA2 mutations among clinic-based African American families with breast cancer. *Human Genetics*, 107(2), 186– 191.

Gayol, L., Scholl, T., Basterrechea, H., Pfeifer, I., Davies, J., Perera, E., & Baumbach, L. (1999). BRCA1 mutation analysis in at-risk African-American families: Results and implications. *American Journal of Human Genetics*, 65(4), A127.

Haffty, B. G., Choi, D. H., Goyal, S., Silber, A., Ranieri, K., Matloff, E., ... Moran, M. S. (2009). Breast cancer in young women (YBC): Prevalence of BRCA1/2 mutations and risk of secondary malignancies across diverse racial groups. *Annals of Oncology*, 20(10), 1653– 1659. <https://doi-org.uplib.idm.oclc.org/10.1093/annonc/mdp051>

Hall, M. J., Reid, J. E., Burbidge, L. A., Pruss, D., Deffenbaugh, A. M., Frye, C., ... Noll, W. W. (2009). BRCA1 and BRCA2 mutations in women of different ethnicities undergoing testing for hereditary breast-ovarian cancer. *Cancer*, 115(10), 2222– 2233. <https://doi-org.uplib.idm.oclc.org/10.1002/cncr.24200>

Janavičius, R. (2010). Founder BRCA1/2 mutations in the Europe: Implications for hereditary breast-ovarian cancer prevention and control. *The EPMA Journal*, 1(3), 397– 412. <https://doi-org.uplib.idm.oclc.org/10.1007/s13167-010-0037-y>

John, E. M., Miron, A., Gong, G., Phipps, A. I., Felberg, A., Li, F. P., ... Whittemore, A. S. (2007). Prevalence of pathogenic BRCA1 mutation carriers in 5 US racial/ethnic groups. *Journal of the American Medical Association*, 298(24), 2869– 2876.

Kanaan, Y., Kpenu, E., Utey, K., Adams-Campbell, L., Dunston, G. M., Brody, L. C., & Broome, C. (2003). Inherited BRCA2 mutations in African Americans with breast and/or ovarian cancer: A study of familial and early onset cases. *Human Genetics*, 113(5), 452– 460.

Kedar-Barnes, I., Devilee, P., Meijers-Heijboer, H., Klijn, J., Plon, S., 2000, I. B., ... A484., S. (2000). Intronic BRCA1 mutations in two highly affected kindreds. *American Journal of Human Genetics*, 67(99), A484.

- Laitman, Y., Feng, B. J., Zamir, I. M., Weitzel, J. N., Duncan, P., Port, D., ... Friedman, E. (2013). Haplotype analysis of the 185delAG BRCA1 mutation in ethnically diverse populations. *European Journal of Human Genetics*, 21(2), 212– 216. <https://doi-org.uplib.idm.oclc.org/10.1038/ejhg.2012.124>
- Luyeye Mvila, G., Postema, S., Marchal, G., VanLimbergen, E., Verdonck, F., Matthijs, G., ... VanOngeval, C. (2014). From the set-up of a screening program of breast cancer patients to the identification of the first BRCA mutation in the DR Congo. *BMC Public Health*, 14, 759. <https://doi-org.uplib.idm.oclc.org/10.1186/1471-2458-14-759>
- Lynce, F., Smith, K. L., Stein, J., DeMarco, T., Wang, Y., Wang, H., ... Isaacs, C. (2015). Deleterious BRCA1/2 mutations in an urban population of Black women. *Breast Cancer Research and Treatment*, 153(1), 201– 209. <https://doi-org.uplib.idm.oclc.org/10.1007/s10549-015-3527-8>
- Martin, S. E., Sausen, M., Joseph, A., Biggs, D. D., Kingham, B. F., & Martin, E. S. (2009). BRCA1 E1644X: A deleterious mutation in an African American individual with early onset breast cancer. *Breast Cancer Research and Treatment*, 113(2), 393– 395. <https://doi-org.uplib.idm.oclc.org/10.1007/s10549-008-9928-1>
- van derMerwe, N. C., Hamel, N., Schneider, S. R., Apffelstaedt, J. P., Wijnen, J. T., & Foulkes, W. D. (2012). A founder BRCA2 mutation in non-Afrikaner breast cancer patients of the Western Cape of South Africa. *Clinical Genetics*, 81(2), 179– 184. <https://doi-org.uplib.idm.oclc.org/10.1111/j.1399-0004.2010.01617.x>
- Mikaelsdottir, E. K., Valgeirsdottir, S., Eyfjord, J. E., & Rafnar, T. (2004). The Icelandic founder mutation BRCA2 999del5: Analysis of expression. *Breast Cancer Research*, 6(4), R284– R290. <https://doi-org.uplib.idm.oclc.org/10.1186/bcr785>
- Miki, Y., Swensen, J., Shattuck-Eidens, D., Futreal, P. A., Harshman, K., Tavtigian, S., ... Mark, H. (1994). A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1. *Science*, 266(5182), 66– 71.
- Nanda, R., Schumm, L. P., Cummings, S., Fackenthal, J. D., Sveen, L., Ademuyiwa, F., ... Olopade, O. I. (2005). Genetic testing in an ethnically diverse cohort of high-risk women: A comparative analysis of BRCA1 and BRCA2 mutations in American families of European and African ancestry. *Journal of the American Medical Association*, 294(15), 1925– 1933.
- Olopade, O. I., Fackenthal, J. D., Dunston, G., Tainsky, M. A., Collins, F., & Whitfield-Broome, C. (2003). Breast cancer genetics in African Americans. *Cancer*, 97(1 Suppl), 236– 245.
- Oluwagbemiga, L. A., Oluwole, A., & Kayode, A. A. (2012). Seventeen years after BRCA1: What is the BRCA mutation status of the breast cancer patients in Africa? – a systematic review. *SpringerPlus*, 1(1), 83. <https://doi-org.uplib.idm.oclc.org/10.1186/2193-1801-1-83>

Pal, T., Bonner, D., Cragun, D., Monteiro, A. N., Phelan, C., Servais, L., ... Vadaparampil, S. T. (2015). A high frequency of BRCA mutations in young black women with breast cancer residing in Florida. *Cancer*, 121(23), 4173– 4180.
<https://doi-org.uplib.idm.oclc.org/10.1002/cncr.29645>

Pal, T., Permut-Wey, J., Holtje, T., & Sutphen, R. (2004). BRCA1 and BRCA2 mutations in a study of African American breast cancer patients. *Cancer Epidemiology, Biomarkers and Prevention*, 13(11 Pt 1), 1794– 1799.

Pal, T., Vadaparampil, S., Betts, J., Miree, C., Li, S., & Narod, S. A. (2008). BRCA1/2 in high-risk African American women with breast cancer: Providing genetic testing through various recruitment strategies. *Genetic Testing*, 12(3), 401– 407.
<https://doi-org.uplib.idm.oclc.org/10.1089/gte.2007.0108>

Panguluri, R. C., Brody, L. C., Modali, R., Utey, K., Adams-Campbell, L., Day, A. A., ... Dunston, G. M. (1999). BRCA1 mutations in African Americans. *Human Genetics*, 105(1-2), 28– 31.

Perrin-Vidoz, L., Sinilnikova, O. M., Stoppa-Lyonnet, D., Lenoir, G. M., & Mazoyer, S. (2002). The nonsense-mediated mRNA decay pathway triggers degradation of most BRCA1 mRNAs bearing premature termination codons. *Human Molecular Genetics*, 11(23), 2805– 2814.

Rebbeck, T., Kauff, N., & Domchek, S. (2008). Meta-analysis of risk reduction estimates associated with risk-reducing salpingo-oophorectomy in BRCA1 or BRCA2 mutation carriers. *Journal of the National Cancer Institute*, 101(2), 80– 87.

Rebbeck, T. R., Friebel, T., Wagner, T., Lynch, H. T., Garber, J. E., Daly, M. B., ... Weber, B. L. The PROSE Study Group (2005). Effect of short-term hormone replacement therapy on breast cancer risk reduction after bilateral prophylactic oophorectomy in BRCA1 and BRCA2 mutation carriers. *Journal of Clinical Oncology*, 23(31), 7804– 7810.
<https://doi-org.uplib.idm.oclc.org/JCO.2004.00.8151>

Rebbeck, T. R., Friebel, T. M., Friedman, E., Hamann, U., Huo, D., Kwong, A., ... Nathanson, K. L. (2018). Mutational spectrum in a worldwide study of 29,700 families with BRCA1 or BRCA2 mutations. *Human Mutation*, 39(5), 593– 620.
<https://doi-org.uplib.idm.oclc.org/10.1002/humu.23406>

Rebbeck, T. R., Friebel, T. M., Mitra, N., Wan, F., Chen, S., Andrulis, I. L., ... Ramus, S. J. (2016). Inheritance of deleterious mutations at both BRCA1 and BRCA2 in an international sample of 32,295 women. *Breast Cancer Research*, 18(1), 112.
<https://doi-org.uplib.idm.oclc.org/10.1186/s13058-016-0768-3>

Rebbeck, T. R., Kauff, N. D., & Domchek, S. M. (2009). Meta-analysis of risk reduction estimates associated with risk-reducing salpingo-oophorectomy in BRCA1 or BRCA2 mutation carriers. *Journal of the National Cancer Institute*, 101(2), 80– 87.
<https://doi-org.uplib.idm.oclc.org/10.1093/jnci/djn442>

- Shen, D., Wu, Y., Subbarao, M., Bhat, H., Chillar, R., & Vadgama, J. V. (2000). Mutation analysis of BRCA1 gene in African-American patients with breast cancer. *Journal of the National Medical Association*, 92(1), 29– 35.
- Slaoui, M., Razine, R., Ibrahimi, A., Attaleb, M., Mzibri, M. E., & Amrani, M. (2014). Breast cancer in Morocco: A literature review. *Asian Pacific Journal of Cancer Prevention: APJCP*, 15(3), 1067– 1074.
- Stoppa-Lyonnet, D., Laurent-Puig, P., Essioux, L., Pages, S., Ithier, G., Ligot, L., ... Thomas, G. (1997). BRCA1 sequence variations in 160 individuals referred to a breast/ovarian family cancer clinic. Institut Curie Breast Cancer Group. *American Journal of Human Genetics*, 60(5), 1021– 1030.
- Sutphen, R., & Ferlita, T. (1999). Inherited breast/ovarian cancer in African-American. *American Journal of Human Genetics*, 65(Suppl), A325.
- Vanderpuye, V., Grover, S., Hammad, N., PoojaPrabhakar, Simonds, H., Olopade, F., & Stefan, D. C. (2017). An update on the management of breast cancer in Africa. *Infectious Agents and Cancer*, 12, 13. <https://doi-org.uplib.idm.oclc.org/10.1186/s13027-017-0124-y>
- Ware, M. D., DeSilva, D., Sinilnikova, O. M., Stoppa-Lyonnet, D., Tavtigian, S. V., & Mazoyer, S. (2006). Does nonsense-mediated mRNA decay explain the ovarian cancer cluster region of the BRCA2 gene? *Oncogene*, 25(2), 323– 328. <https://doi-org.uplib.idm.oclc.org/10.1038/sj.onc.1209033>
- Whitfield-Broome, C., Dunston, G., & Brody, L. (1999). BRCA2 mutations in African Americans. *Proceedings of the American Association of Cancer Research*, 40, 269.
- Zhang, B., Fackenthal, J. D., Niu, Q., Huo, D., Sveen, W. E., DeMarco, T., ... Olopade, O. I. (2009). Evidence for an ancient BRCA1 mutation in breast cancer patients of Yoruban ancestry. *Familial cancer*, 8(1), 15– 22.
- Zhang, J., Fackenthal, J. D., Huo, D., Zheng, Y., & Olopade, O. I. (2010). Searching for large genomic rearrangements of the BRCA1 gene in a Nigerian population. *Breast Cancer Research and Treatment*, 124(2), 573– 577.
- Zhang, J., Fackenthal, J. D., Zheng, Y., Huo, D., Hou, N., Niu, Q., ... Olopade, O. I. (2012). Recurrent BRCA1 and BRCA2 mutations in breast cancer patients of African ancestry. *Breast Cancer Research and Treatment*, 134(2), 889– 894. <https://doi-org.uplib.idm.oclc.org/10.1007/s10549-012-2136-z>
- Zheng, Y., Walsh, T., Gulsuner, S., Casadei, S., Lee, M. K., Ogundiran, T. O., ... Olopade, O. I. (2018). Inherited breast cancer in Nigerian Women. *Journal of Clinical Oncology*, 36, 2820– 2825. <https://doi-org.uplib.idm.oclc.org/10.1200/JCO.2018.78.3977>

Zoure, A. A., Slaoui, M., Bambara, H. A., Sawadogo, A. Y., Compaoré, T. R., Ouédraogo, N. L. M., ... Bakri, Y. (2018). BRCA1 c.68_69delAG (exon2), c.181T>G (exon5), c.798_799delTT and 943ins10 (exon11) mutations in Burkina Faso. *Journal of Public Health in Africa*, 9(1), 663. <https://doi-org.uplib.idm.oclc.org/10.4081/jphia.2018.663>