

Fig. S1. Growth kinetics of all virus strains in BSR cells over 72 h p.i.. Titres of virus released, cell associated virus and total virus yield indicated. Tables representing the TCID50 titre values and log conversions of the titres included. Titrations were done in quadruplicate for all biological repeats at all three time points.

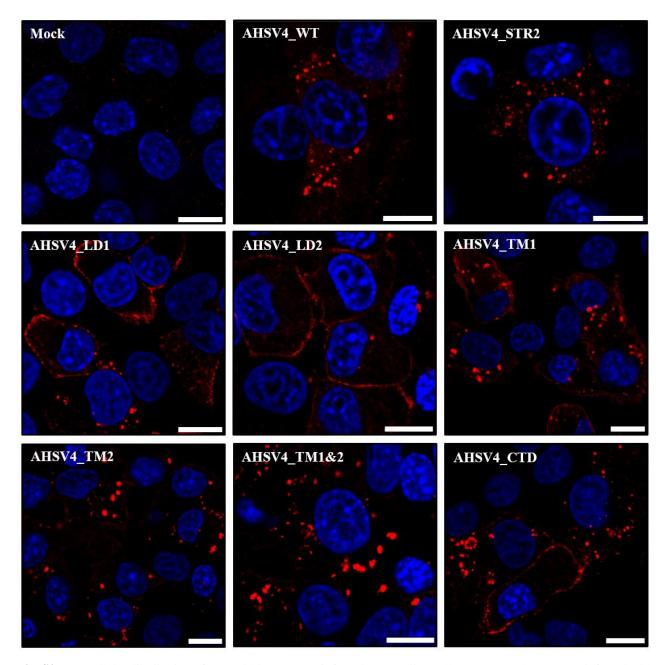


Fig. S2. Intracellular distribution of VP5 within AHSV-4 infected mammalian cells. BSR monolayers were infected with wild-type or mutant AHSV-4 virus strains and fixed at 24 h p.i. Mock infected cells were included as control. VP5 was detected with an anti-VP5 antibody (mouse) and Alexa FluorTM 594 conjugated goat anti-mouse secondary antibody (red). Nuclei were stained with DAPI (blue). Scale bars represent $10~\mu m$.

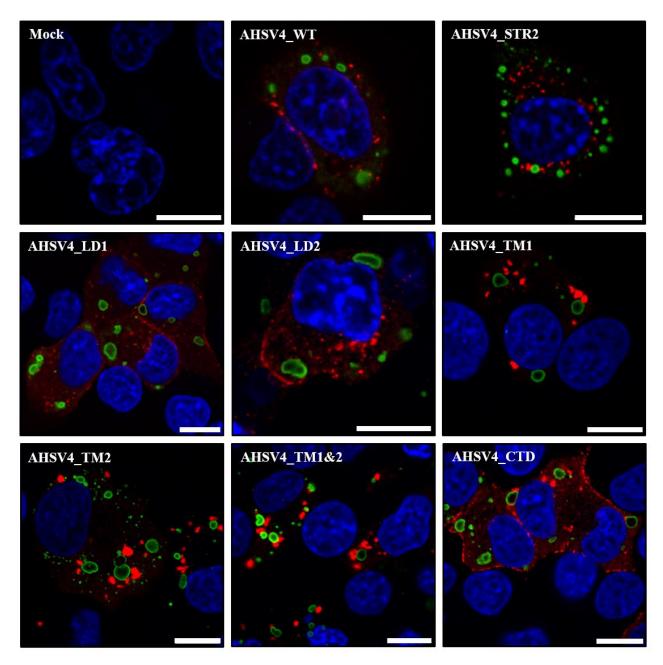


Fig. S3. Intracellular localisation of VP2 and NS2 within AHSV-4 infected mammalian cells. BSR monolayers were infected with wild-type or mutant AHSV-4 virus strains and fixed at 24 h p.i. Mock infected cells were included as a control. NS2 was detected with an anti-NS2 antibody (rabbit) and Alexa FluorTM 488 conjugated goat anti-rabbit secondary antibody (green). VP2 was detected with an anti-VP2 antibody (mouse) and Alexa FluorTM 594 conjugated goat anti-mouse secondary antibody (red). Nuclei were stained with DAPI (blue). Scale bars represent 10 μm.

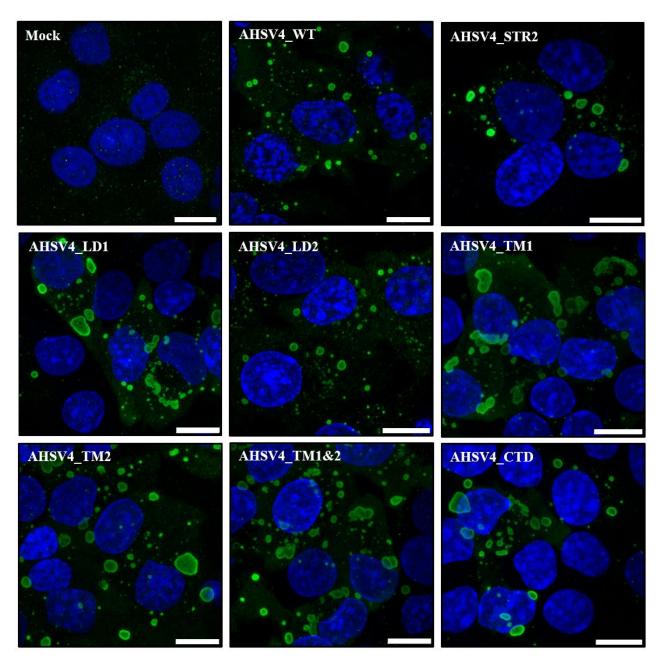


Fig. S4. Representative MIPs of the intracellular distribution of VIBs within AHSV-4 infected mammalian cells. BSR monolayers were infected with wild-type or mutant AHSV-4 virus strains and fixed at 24 h p.i. Mock infected cells were included as control. NS2 was detected with an anti-NS2 antibody (rabbit) and Alexa FluorTM 488 conjugated goat antirabbit secondary antibody (green). Nuclei were stained with DAPI (blue). Scale bars represent 10 μm.

Table 1: List of primers used for QuickChange® II modifications of Seg-10						
Primer	Primer sequence with mismatches underlined (5'-3')	Amino acid change	Length (nucleotides [nt])			
STR 1 Forward	${\tt GTTAAAATTATCCCTTGTC} \underline{{\tt GC}} {\tt GAATCTAGCTACAATCGCC}$	M to A	40			
STR 1 Reverse	${\tt GGCGATTGTAGCTAGATTC} \underline{{\tt GC}} {\tt GACAAGGGATAATTTTAAC}$		40			
STR 2 Forward	CAATCGCCAAGAATTATAGC <u>GC</u> GCATAATGGAGAGTCGGG	M to A	40			
STR 2 Reverse	$CCCGACTCTCCATTATGC\underline{GC}GCTATAATTCTTGGCGATTG$		40			
LD1 Forward	CACCACCATACAATTTCG <u>G</u> A <u>GC</u> TGCTCCGACGTTTTCTC	A.C. 4. C.A.	39			
LD1 Reverse	GAGAAAACGTCGGAGCA <u>GC</u> T <u>C</u> CGAAATTGTATGGTGGTG	- AS to GA	39			
LD2 Forward	GATCGTCCCTTATGTG <u>G</u> CA <u>G</u> CACCA <u>GG</u> CAATTTCGCAAGTGCTCC	PPPY to AAPG	45			
LD2 Reverse	GGAGCACTTGCGAAATTG <u>CC</u> TGGTG <u>C</u> TG <u>C</u> CACATAAGGGACGATC		45			

Table 2: g	Table 2: gBlock® gene fragments for introduction of specific mutations to Seg-10					
	gBlock [®] gene fragment sequence with nucleotide modifications underlined (5'-3')	Amino acid change	Length (nt)			
TM 1	GCAACAATGCGTCGAAAGAAATCGGCATTAAAAAATAATGATCTTTATTAGT GGATGC <u>CGAACGCGAGATAAA</u> TCGATGGTTGGGGGGATTGAGTATCGTTGA CGACGAAATATTAAGAGATTATAAGAACAACG	VTLAT to RTRDK	133			
TM 2	GTATCGTTGACGACGAAATATTAAGAGATTATAAGAACAACGATTGGTTAA TGAAGACTATACATGGGCTGAATTTGTTATGTACTACA <u>AAGCGCGACGTG</u> G CGGGTAAGATTTCCGATAAAATGCAAGAGGAGATTTCACGGACTAAACGT GACATTGCGAAAAGAGAGTCTTACGTGTCAGCGGCGAGTATGTCGTGGAG TGGAG	VLLA to KRDV	207			
TM 1&2	GCAACAATGCGTCGAAAGAAATCGGCATTAAAAATAATGATCTTTATTAGT GGATGC <u>CGAACGCGAGATAAA</u> TCGATGGTTGGGGGGATTGAGTATCGTTGA CGACGAAATATTAAGAGATTATAAGAACAACGATTGGTTAATGAAGACTA TACATGGGCTGAATTTGTTATGTACTACA <u>AAGCGCGACGTG</u> GCGGGTAAGA TTTCCGATAAAATGCAAGAGGAGATTTCACGGACTAAACGTGACATTGCGA AAAGAGAGTCTTACGTGTCAGCGGCGAGTATGTCGTGGAGTGGAG	VTLAT to RTRDK VLLA to KRDV	298			
CTD	TGTTATGTACTACAGTTTTGTTAGCGGCGGGTAAGATTTCCGATAAAATGC AAGAGGAGATTTCACGGACTAAACGTGACATTGCGAAAAGAGAGTCTTAC GTGTCAGCGGCGAGT <u>TAG</u> TATGACCTCCACGAGCGGAAAATCCATCGTGTT GGATG	Deletion of 19 aa at C-terminal end	157			

Table 3: List of primers used for pJAD.S10 linearisation					
Primer	Primer sequence (5'-3')	Length (nt)			
TM 1 Forward	ATTATAAGAACAACGATTGGTTAATGAAGACTATAC	36			
TM 1 Reverse	TCGACGCATTGTTGCTAACTCCATCTTTAGG	31			
TM 2 Forward	TGTCGTGGAGTGGAGATACTGAGATG	26			
TM 2 Reverse	CGTCGTCAACGATACTCAATCC	22			
CTD Forward	CCATCGTGTTGGATGGATGG	20			
CTD Reverse	CTGTAGTACATAACAAATTCAGCCCATGTATAGTC	35			