

RESEARCH COMMUNICATION

SEROLOGICAL REACTIONS TO *LEPTOSPIRA* SPECIES IN BUFFALO (*SYNCERUS CAFFER*) FROM THE KRUGER NATIONAL PARK

J. G. MYBURGH<sup>(1)</sup>, R. G. BENGIS<sup>(2)</sup>, C. J. J. BESTER<sup>(1)</sup> and F. CHAPARRO<sup>(1)</sup>

ABSTRACT

MYBURGH, J. G., BENGIS, R. G., BESTER, C. J. J. & CHAPARRO, F., 1990. Serological reactions to *Leptospira* species in buffalo (*Syncerus caffer*) from the Kruger National Park. *Onderstepoort Journal of Veterinary Research*, 57, 281-282 (1990).

Four hundred and six serum samples from buffalo (*Syncerus caffer*) were tested for leptospirosis, using the microscopic agglutination test. Seven buffaloes (1.7 %) reacted positive and 27 (6.6 %) inconclusive. Reactions against *L. tarassovi* and *L. hardjo* were the most prevalent.

INTRODUCTION

*Leptospira* organisms occur in southern Africa, as demonstrated by serological surveys (Botes & Garifallou, 1967; Twigg, Sikes & Hughes, 1970; Feresu, 1987; Hunter, Flamand, Myburgh & Van der Merwe, 1988), and have been isolated by Herr, Riley, Nesor, Roux & De Lange, 1982; Herr & Winnen, 1983 and Te Brugge & Dreyer, 1985. The most prevalent serovars in cattle in southern Africa are *hardjo*, *pomona*, *tarassovi* and *mini* (Myburgh, unpublished data, 1989). Leptospiral infections in domestic animals are usually associated with abortions in southern Africa (Herr *et al.*, 1982; Te Brugge & Dreyer, 1985).

The role played by wild animals in the epidemiology of leptospirosis has not been fully investigated. In large areas of southern Africa, game is still abundant (Smithers, 1983) and could easily infect domestic animals, or vice versa (Faine, 1982; Wanyangu, Olubayo, Rositter & Waitkins, 1987).

That *Leptospira* organisms can survive in the environment is an important factor in the epidemiology, transmission and spread of the disease (Faine, 1982; Thierman, 1984). Waterholes or streams shared by game and cattle could be a source of *Leptospira* organisms (Faine, 1982).

It was decided to test buffalo from the Kruger National Park (KNP) to determine whether antibodies against *Leptospira* species occur in these animals. Buffaloes in the KNP are relatively isolated from domestic animals, and direct contact is unlikely (Bengis, personal communication, 1990). Positive serological evidence of leptospirosis in buffalo would be an indication that buffaloes are exposed to and stimulated by *Leptospira* organisms occurring in the KNP.

MATERIALS AND METHODS

Blood samples were collected during routine culling of buffalo. A total of 406 samples were collected, from different areas in the KNP over a year. These were centrifuged and the serum was separated. The serum samples were inactivated for 30 min at 58 °C, because the KNP is a foot-and-mouth control area. The sera were frozen (-12 °C) in small plastic tubes and stored until testing.

The sera were tested, using the microscopic agglutination micro-volume technique (Sulzer & Jones, 1978; Herr, Hunter & De Lange, 1987). The

following antigens were used: *canicola*, *copenhageni* (*icterohaemorrhagiae*), *grippotyphosa*, *hardjo*, *mini* (*swajizak*), *pomona*, *pyrogenes* and *tarassovi* (hyos).

Antigens were grown on liquid EMJH<sup>1</sup> medium and used between 4-14 days, when the growth of the leptospire exceeded  $2 \times 10^7$  organisms per ml (Sulzer & Jones 1978). The end-point titre was taken as the dilution, where 50 % of the organisms, as compared with the negative control, were either absent or visibly agglutinated and there was a greater degree of agglutination in the immediately preceding lower dilution. A titre of less than 160 was regarded as inconclusive and 160 and higher as positive.

RESULTS

Four hundred and six serum samples were tested and of these 7 (1.7 %) reacted with a titre of 1/160 or more and were considered positive. Twenty-seven (6.6 %) buffaloes reacted inconclusively.

The total number of reactions (positive and inconclusive) against serovars tested for are given in Fig. 1. Reactions against *L. tarassovi* and *L. hardjo* were the most prevalent.

DISCUSSION

The serology results give clear evidence of the existence of leptospirosis in buffalo from the KNP. Titres to *L. tarassovi* and *L. hardjo* are the most prevalent (Fig. 1) and the low titres recorded in the buffalo might point to carrier status.

The epidemiology of leptospirosis with regard to the role played by wild animals has not been fully investigated in southern Africa, but the occurrence of these serological reactions in buffalo indicate that they are being exposed to *Leptospira* organisms under natural conditions in an ecosystem relatively isolated (Bengis, personal communication, 1990) from direct contact with domestic animals. Fifty cattle from a farm adjacent to the KNP in the Phalaborwa area were also tested for the same serovars and reactions against *L. tarassovi* and *L. pomona* were the most prevalent (Myburgh, unpublished data, 1989).

Transmission of *Leptospira* organisms from domestic animals and wild animals other than buffalo is a possible explanation for the titres occurring in buffalo. Transmission by water, carrier animals or birds could play a very important role in the

<sup>(1)</sup> Veterinary Research Institute, Onderstepoort 0110  
<sup>(2)</sup> Directorate of Animal Health, P.O. Box 12, Skukuza 1350  
Received 6 July 1990—Editor

<sup>1</sup> Difco EMJH medium base + Difco EMJH enrichment. Difco Laboratories, Detroit, Michigan, USA



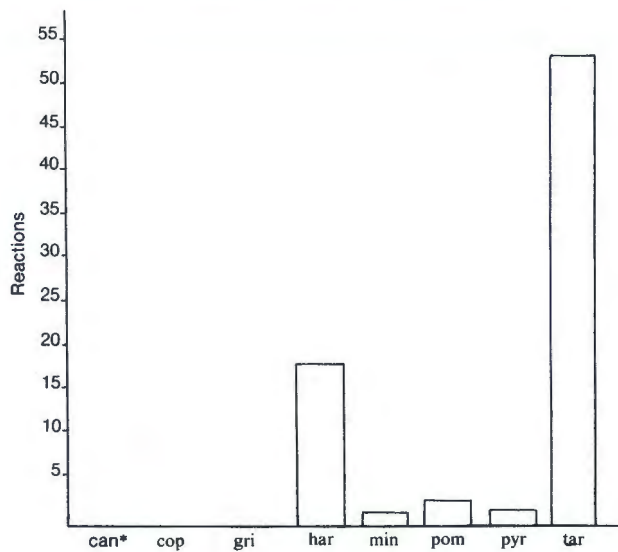


FIG. 1 Total number of reactions (positive and inconclusive) against all serovars tested for

- \* can = canicola
- cop = copenhageni
- gri = grippotyphosa
- har = hardjo
- min = mini
- pom = pomona
- pyr = pyrogenes
- tar = tarassovi

epidemiology of leptospirosis in buffaloes (Faine, 1982; Thierman, 1984).

The role of wet environments, aiding in the survival of *Leptospira* species, has been demonstrated and reviewed by various authors (Kirschner & Maguire, 1957; Gordon-Smith & Turner, 1961; Henry & Johnson, 1978). These serological reactions in buffalo could indicate that water-borne transmission of these organisms might be possible by streams and effluent water flowing into the park, from outside. Buffaloes are closely associated with water, and show wallowing and mud bathing behaviour (Smithers, 1983).

Shepherd & Leman (1958) were unable to isolate leptospire from wild rodents collected in various regions of the RSA, although these were collected mainly in drier parts of the country where leptospirosis is less prevalent. The role of rodents in the epidemiology of leptospirosis should be regarded as minor at this stage.

*Leptospira* organisms have been isolated from birds in other parts of the world (Van der Hoeden, 1964; Thierman, 1984). The role of birds, such as vultures, which frequently visit farms and cattle carcasses outside the KNP, could play a role in transmitting the disease, but data are not available.

The serological reactions in buffalo may therefore be the result of exposure to *Leptospira* organisms

excreted by buffaloes, other wild animals or domestic animals. Serological surveys and epidemiological studies in domestic and wild animals, as well as attempts to the isolate *Leptospira* organisms from these animals, should be conducted.

#### ACKNOWLEDGEMENTS

The authors wish to acknowledge the excellent cooperation and help of the personnel of the Directorate of Animal Health and National Parks Board.

#### REFERENCES

- BOTES, H. J. W. & GARIFALLOU, A., 1967. Leptospirosis: a brief review, general considerations and incidence in South Africa *Journal of the South African Veterinary Medical Association*, 38, 67-75.
- FAINE, S., 1982. Guidelines for the control of leptospirosis. Switzerland: World Health Organization.
- FERESU., SARA B., 1987. Serological survey of leptospiral antibodies in cattle in Zimbabwe. *Tropical Animal Health and Production*, 19, 209-214.
- GORDON-SMITH, C. E. & TURNER, L. H., 1961. The effect of pH on the survival of leptospire in water. *Bulletin of the World Health Organization*, 24, 35-43.
- HENRY, R. A. & JOHNSON, R. C., 1978. Distribution of the genus *Leptospira* in soil and water. *Applied and Environmental Microbiology*, 35, 492-499.
- HERR, S., RILEY, A. E., NESER, J. A., ROUX, D. & DE LANGE, J. F., 1982. *Leptospira interrogans* serovar pomona associated with abortion in cattle: isolation methods and laboratory animal histopathology. *Onderstepoort Journal of Veterinary Research*, 49, 57-62.
- HERR, S. & WINNEN, G. M., 1983. The first isolation of *Leptospira interrogans* serovar pomona from cattle in Botswana. *Journal of the South African Veterinary Association*, 54, 83-84.
- HERR, S., HUNTER, PAMELA & DE LANGE, J. F., 1987. Leptospirosis manual: a practical laboratory guide to the serology and isolation of *Leptospira*. Section of Reproduction, Veterinary Research Institute, Onderstepoort 0110 RSA.
- HUNTER, PAMELA, FLAMAND, J. R. B., MYBURGH, J. & VAN DER MERWE, SANETTE, M., 1988. Serological reactions to *Leptospira* species in game animals of northern Natal. *Onderstepoort Journal of Veterinary Research*, 55, 191-192.
- KIRSCHNER, L. & MAGUIRE, T., 1957. Survival of *Leptospira* outside their hosts. *New Zealand Medical Journal*, 56, 385-391.
- SHEPHERD, A. J. & LEMAN, P. A., 1958. Bacterial surveillance of South African rodents. *South African Journal of Science*, 81, 302-308.
- SMITHERS, REAY H. N., 1983. The mammals of the Southern African sub-region. Pretoria: University of Pretoria.
- SULZER, C. R. & JONES, W. L., 1978. Leptospirosis: methods in laboratory diagnosis. Revised edition. United States Department of Health, Education and Welfare, Center for Disease Control. Atlanta, Georgia.
- TE BRUGGE, LESLEY A. & DREYER, A., 1985. *Leptospira interrogans* serovar hardjo associated with bovine abortion in South Africa. *Onderstepoort Journal of Veterinary Research*, 52, 51-52.
- THIERMAN, A. B., 1984. Leptospirosis: Current developments and trends. *Journal of the American Veterinary Medical Association*, 184, 722-725.
- TWIGG, G. I., SIKES, SYLVIA K. & HUGHES, D. M., 1970. Evidence of leptospirosis in some large East African mammals. *East African Wildlife Journal*, 8, 197-198.
- VAN DER HOEDEN, J. (ed.), 1964. Zoonoses. Amsterdam: Elsevier.
- WANYANGU, S. W., OLUBAYO, R. O., ROSITTER, P. B. & WAITKINS, SHEENA A., 1987. The study of the ecology and prevalence of leptospirosis in large wild ruminants and domesticated bovines found in Kenya. *Israel Journal of Veterinary Medicine*, 43, 340-341.