

CHAPTER 1: INTRODUCTION.

1.1 General introduction

Extended-spectrum beta-lactamases (ESBLs) that are inhibited by clavulanic acid, belong mostly to class A of the Ambler classification scheme, conferring resistance to a wide variety of expanded-spectrum cephalosporins. To date however, a bewildering variety of enzymes have been described that can be classified as set out in Table 1-1. After being widely reported in *Enterobacteriaceae* isolates from the early 1980`s, ESBLs have been described in *Pseudomonas aeruginosa* only more recently (1, 2, 3). These enzymes described in *P. aeruginosa*, are either of the TEM- and SHV-types that are also well known in *Enterobacteriaceae*, or of the PER-type mostly originating from Turkey, or of the VEB-type from Southeast Asia and more recently, of the GES / IBC types originally reported from French Guinea, France, Greece and South Africa, respectively (4, 5, 6, 7). These five types of enzymes are remotely related, both from a genetic point of view and with respect to similarities in hydrolytic profiles. To date, CTX-M-type enzymes have not yet been described in *P. aeruginosa*.

Recent studies indicated that these enzymes may play an important role in the dissemination of antibiotic resistant bacterial isolates and may condition future choices of antibiotic regimens for treating life-threatening infections due to ESBL-producing *P. aeruginosa* (8, 9). Recent work further indicated the propensity of ESBL producing *P. aeruginosa* to establish long-term residence in the nosocomial environment, making re-infection an imminent danger (10). After the discovery of the novel ESBL, GES-2 from *P. aeruginosa*, in the Pretoria Academic Hospital (PAH) in May of 2000 (7), it was clear that class 1 integron borne ESBLs were established in the South African nosocomial setting (Discussed in detail in Chapter 3). The same *P. aeruginosa* strain subsequently caused a nosocomial outbreak in the PAH, exhibiting a 62.5% mortality

rate (11). The *P. aeruginosa* strain described during that outbreak exhibited a tendency to widely colonise and infect mostly debilitated patients, significantly increasing both their length of stay in the ICU and cost of treatment (11).

Table 1-1: Beta-lactamase classification schemes referred to in this study.

Bush group	Subgroup	Ambler molecular class	Characteristics
1		C	Mainly chromosomal located in Gram-negative bacteria but may be plasmid mediated. Confer resistance to beta-lactams (except carbapenems). Not inhibited by clavulanate.
2		A, D	Most enzymes inhibited by clavulanate (unless otherwise stated).
	2a	A	Penicillinases (narrow hydrolysis spectrum) conferring resistance to penicillins.
	2b	A	Broad-spectrum penicillinases (TEM-1, SHV-1) primarily from Gram-negative bacteria.
	2be	A	Extended-spectrum beta-lactamases conferring resistance to oxyimino-cephalosporins and monobactams.
	2br	A	Inhibitor resistant beta-lactamases (mostly TEM-types and to a lesser extent SHV derived enzymes).
	2c	A	Carbenicillinases.
	2d	D	Oxacillinases, modestly inhibited by clavulanate.
	2e	A	Cephalosporinases inhibited by clavulanate.
	2f	A	Serine active site carbapenemases, inhibited by clavulanate.
3	3a, b, c	B	Metallo-beta-lactamases conferring resistance to beta-lactams (except monobactams), not inhibited by clavulanic acid.
4			Miscellaneous unsequenced beta-lactamases that do not conform to other groups.

Data adapted from references 16 and 17.

Previous analysis of *bla*_{GES} genes suggested that these genetic structures did not primarily evolve from *P. aeruginosa* as the G+C content of *bla*_{GES-2} was 51.5%, a value which is not within the range of G+C content of *P. aeruginosa* genes (60.1 – 69.5%)

(7), this value corresponds with genes originating from the *Enterobacteriaceae*. This fact then raises the question about the origin of the GES-type genes and the possibility that they may have developed in *Enterobacteriaceae* strains including *Klebsiella pneumoniae* isolates, as was previously described in a GES-1 producing *K. pneumoniae* isolate originating from French Guinea (12). Recent developments in Japan with the finding of $bla_{\text{GES-3}}$ and $bla_{\text{GES-4}}$ (13, 14) in clinical isolates of *K. pneumoniae* strongly support this theory. The integron genetic support that these novel enzymes enjoy, not only confers resistance towards broad-spectrum beta-lactam antibiotics, but also towards unrelated classes of antibiotics such as aminoglycosides and sulphonamides as well as to several hospital disinfectants (15). Due to the composition of class 1 integrons and the simultaneous expression of all the gene cassettes comprising the integron structure (9), it is in theory possible that non-beta-lactam antibiotics may actually co-select ESBL producing *P. aeruginosa* and certain *Enterobacteriaceae*. This selection phenomenon may cause widespread nosocomial dissemination of bacterial strains harbouring these genetic structures, making treatment and control exceptionally difficult.

1.2 OBJECTIVES:

- To develop improved molecular screening and detection methods for the novel beta-lactamase GES-2 in *P. aeruginosa* isolates.
- To determine the genetic stability of bla_{GES} -type gene cassettes in class 1 integrons under antibiotic pressure in vitro.

1.3 HYPOTHESIS:

- Class 1 integron located bla_{GES} gene cassette stability is expected to change under selective antibiotic pressure.

1.4 REFERENCES

1. **Jarlier, V., M.-H. Nicolas, G. Fournier, and A. Philippon.** 1988. Extended broad-spectrum beta-lactamases conferring transferable resistance to newer beta-lactam agents in *Enterobacteriaceae*: hospital prevalence and susceptibility patterns. *Rev. Infect. Dis.* **10**:867-878.
2. **Medeiros, A. A.** 1997. Evolution and dissemination of beta-lactamases accelerated by generations of beta-lactam antibiotics. *Clin. Infect. Dis.* **24**:S19-S45.
3. **Nordmann, P., and M. Guibert.** 1998. Extended-spectrum beta-lactamases in *Pseudomonas aeruginosa*. *J. Antimicrob. Chemother.* **42**:128-131.
4. **Dubois, V., L. Poirel, C. Marie, C. Arpin, P. Nordmann, and C. Quentin.** 2002. Molecular characterization of a novel class 1 integron containing *bla_{GES-1}* and a fused product of *aac(3)-Ib/aac(6')-Ib'* gene cassettes in *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* **46**:638-645.
5. **Mavroidi, A., E. Tzelepi, A. Tsakris, V. Miriagou, D. Sofianou, and L. S. Tzouvelekis.** 2001. An integron-associated beta-lactamase (IBC-2) from *Pseudomonas aeruginosa* is a variant of the extended-spectrum beta-lactamase IBC-1. *J. Antimicrob. Chemother.* **48**:627-630.
6. **Naas, T., L. Poirel, A. Karim, and P. Nordmann.** 1999. Molecular characterization of In50, a class 1 integron encoding the gene for the extended-spectrum beta-lactamase VEB-1 in *Pseudomonas aeruginosa*. *FEMS Microbiol. Lett.* **176**:411-419.
7. **Poirel, L., G. F. Weldhagen, T. Naas, C. de Champs, M. G. Dove, and P. Nordmann.** 2001. GES-2, a class A beta-lactamase from *Pseudomonas*

- aeruginosa* with increased hydrolysis of imipenem. Antimicrob. Agents Chemother. **45**:2598-2603.
8. **Docquier J-D., F. Luzzaro, G. Amicosante, A. Toniolo, and G. M. Rossolini.** 2001. Multidrug-resistant *Pseudomonas aeruginosa* producing PER-1 extended-spectrum serine beta-lactamase and VIM-2 metallo- β -lactamase. Emerg. Infect. Dis. **7**: 910-911.
 9. **Girlich, D., T. Naas, A. Leelaporn, L. Poirel, M. Fennewald, and P. Nordmann.** 2002. Nosocomial spread of the integron-located *veb-I*-like cassette encoding an extended-spectrum beta-lactamase in *Pseudomonas aeruginosa* in Thailand. Clin. Infect. Dis. **34**:603-611.
 10. **Poirel L., E. Lebessi, M. Castro et al.** 2004. Nosocomial outbreak of extended-spectrum beta-lactamase SHV-5-producing isolates of *Pseudomonas aeruginosa* in Athens, Greece. Antimicrob. Agents Chemother. **48**: 2277 – 2279.
 11. **Poirel, L., G. F. Weldhagen, C. De Champs, and P. Nordmann.** 2002. A nosocomial outbreak of *Pseudomonas aeruginosa* isolates expressing the extended spectrum beta-lactamase GES-2 in South Africa. J. Antimicrob. Chemother. **49**:561-565.
 12. **Poirel, L., I. Le Thomas, T. Naas, A. Karim, and P. Nordmann** 2000. Biochemical sequence analyses of GES-1, a novel class A extended-spectrum beta-lactamase, and the class 1 integron In52 from *Klebsiella pneumoniae*. Antimicrob. Agents Chemother. **44**:622-632.
 13. **Wachino, J., Y. Doi, K. Yamane et al.** 2004. Nosocomial spread of ceftazidime resistant *Klebsiella pneumoniae* strains producing a novel class A

- beta-lactamase, GES-3, in a neonatal intensive care unit in Japan. *Antimicrob. Agents Chemother.* **48**: 1960 – 1967.
14. **Wachino, J., Y. Doi, K. Yamane et al.** 2004. Molecular characterization of a cephamycin-hydrolyzing and inhibitor resistant class A beta-lactamase, GES-4, possessing a single G170S substitution in the Ω -loop. *Antimicrob. Agents Chemother.* **48**: 2905 – 2910.
 15. **Weldhagen G. F., L. Poirel and P. Nordmann.** 2003. Ambler class A extended-spectrum beta-lactamases in *Pseudomonas aeruginosa*: Novel developments and clinical impact. *Antimicrob. Agents Chemother.* **47**: 2385 – 2392.
 16. **Essack S. Y.** 2004. Beta-lactamases – an overview. *South Afr J Epidemiol Infect.* **19 (3,4)**: 106 – 114.
 17. **Bush, K., G. A. Jacoby, and A. A. Medeiros.** 1995. A functional classification scheme for beta-lactamases and its correlation with molecular structure. *Antimicrob. Agents Chemother.* **39**:1211-1233.