

Chapter 1

Theoretical background and aim of the study

The etiology and pathogenesis of renal diseases vary widely yet as chronic renal failure progresses, the manifestations in different patients are remarkably similar. These similarities make it possible to formulate common principles of pathophysiology that apply to the greater majority of patients, irrespective of the underlying pathogenic (infections, immunological, vascular or other) mechanism responsible for renal function destruction. (1) Intracellular calcium dyshomeostasis would appear to be one such common principle of the pathophysiology of chronic renal failure. This disturbance in intracellular calcium homeostasis involves many different cell types in the chronic renal failure patient. It is therefore possible that the abnormal intracellular calcium homeostasis, indicated in many different cell types in the chronic renal failure patient, may involve the body as a whole. Intracellular calcium homeostatic mechanisms are complex, and a large number of pathological factors are known to cause intracellular calcium dyshomeostasis. Many of these factors generally known to alter intracellular calcium status are indeed found in the chronic renal failure patient.

The present study comprises the investigation of intracellular calcium homeostasis and possible factors influencing intracellular calcium homeostasis in the chronic renal failure patient on maintenance haemodialysis treatment (MHT). The first introductory chapter deals with the theoretical background to the study and includes discussions on intracellular calcium signal transduction, intracellular calcium dyshomeostasis, possible manifestations of intracellular calcium



dyshomeostasis, status of intracellular calcium homeostasis in the chronic renal failure patient on maintenance haemodialysis treatment, and possible factors contributing to intracellular calcium dyshomeostasis in these patients. The aim of this study is presented in conclusion.

1) Intracellular calcium signal transduction

Ionised calcium (Ca2+) is the most common signal transduction factor in cells ranging from bacteria to specialised neurons. (2) Unlike many of the other signal transduction elements, Ca2+ is required throughout the life of a typical cell since Ca²⁺ regulates many diverse cellular processes ranging from gametogenesis, fertilisation, cell proliferation, and specialised control functions including muscle contraction, exocytosis, energy metabolism, chemotaxis, and synaptic plasticity during memory and learning. (3) The vast quantity of the Ca2+ sensitive effectors responsible for the regulation of these processes are located in the cytosol. (4) It is therefore the free ionised Ca2+ fraction in the cytosol that is the most critical with regard to the regulation of these intracellular events. Unlike any other signal transduction element, Ca2+ is not synthesised or subsequently degraded during the signal transduction process. In contrast Ca2+ is moved and/or released and subsequently sequestered for every signal transduction cycle. In order for Ca2+ to be available for signal transduction functions, the cell is endowed with at least two mechanisms to increase Ca2+ in the cytosol, firstly by releasing Ca²⁺ from the intracellular storage sites and secondly by moving Ca2+ from the extracellular medium into the cytosolic space where the effectors are functionally distributed. (5) The intracellular storage sites include Ca2+ bound to the plasma membrane (anionic sites consisting of either phospholipids or membrane proteins), Ca2+ sequestered within intracellular organelles such as the endoplasmic reticulum, and Ca2+ both free and bound within the cytosol. (6) The various mechanisms capable of generating a rise in cytosolic free Ca2+ are



all passive processes since the cytosolic free Ca2+ is lower than the Ca2+ concentration in any of the calcium pools surrounding the cytosolic space. It is therefore not necessary to expend any energy to move calcium down its concentration gradient into the cytosol upon agonist activation. (3, 7) In contrast with these passive processes capable of generating a rise in the cytosolic free Ca2+, the processes necessary to reduce the surplus Ca2+ at the end of the signal transduction cycle require the expenditure of energy. These energy expensive processes responsible for rapidly decreasing cytosolic free Ca²⁺ include the Ca²⁺-ATPase in the plasma membrane and the Ca2+--ATPase in the The activation of these endo/sarcoplasmic reticulum or calciosome. calcium pumps result in the transfer of Ca2+ from the cytosol to the extracellular medium and the sequestration of Ca2+ into the intracellular calcium storage organelles. The subsequent decrease in cytosolic free Ca²⁺ results in termination of the calcium response. (2, 8)

2) Intracellular calcium dyshomeostasis

It is imperative that the cytosolic free Ca²⁺ levels be maintained within normal physiological limits, since unlike any other signal transduction element, calcium dyshomeostasis is an intracellular trigger for cellular dysfunction and damage. (9) The derangements in intracellular calcium homeostasis, resulting in cellular dysfunction and damage, include the intracellular redistribution of calcium pools during the important early stages of cellular damage, followed by the influx of Ca²⁺ from extracellular to intracellular spaces. Many pathogenic factors, **(6)** resulting in cellular dysfunction and damage, affect cellular calcium regulation because their initial effect impair one or more of the following; energy metabolism, plasma membrane integrity, calcium translocation systems or calcium signalling. The ensuing (10)intracellular calcium dyshomeostasis aggravates the initial impairment of various cellular functions, due to the inappropriate activation of



various calcium sensitive enzymes. These calcium sensitive enzymes include various enzymes regulating the intracellular calcium signalling process and a whole array of degradative enzymes: such as phospholipases, proteases and endonucleases. Intracellular targets for these degradative enzymes include the plasma membrane, mitochondria, the endoplasmic reticulum and the cytoskeleton. calcium activated degradative processes result in plasma membrane bleb formation. nuclear membrane bleb formation. deterioration of mitochondrial structure and well function. as as chromatin condensation. (11) Since an inappropriate elevation in intracellular free calcium results in the activation of so many degradative processes, the cell is furnished with several calcium translocation systems to carefully regulate intracellular calcium activity. The function of the Ca2+-ATPase pump, as explained above, is to rapidly obviate the surplus calcium at the end of a transduction cycle. (8) In addition, these calcium pumps also play an important role in decreasing intracellular free Ca2+ under conditions of relative intracellular Ca2+ overload. These pumps are well suited for their roles since they display dynamic, low capacity enzyme kinetics. A rise in cytosolic free calcium results in immediate activation of these pumps and the rapid extrusion out of the cell, and sequestration into intracellular organelles of calcium. In the face of a prolonged elevation in intracellular free calcium, a second high capacity, slow to become activated, calcium extrusion pump plays a more important role namely the Na⁺/Ca²⁺-exchanger. This pump extrudes Ca²⁺ ions to the outside of the cell in exchange for Na⁺ ions. The sodium concentration gradient - high sodium concentration on the outside of the cell and low sodium concentration on the inside - provides the energy needed for Na⁺/Ca²⁺-exchanger activities. This sodium gradient is maintained by the Na⁺/K⁺-ATPase. (7) During prolonged intracellular free calcium elevation the roles of the Na⁺/Ca²⁺-exchanger and in combination the buffering of calcium by the mitochondria become more important. (11)



In fact, the mitochondrium becomes the single most important membrane system in the cell to act as a long-term calcium buffer and to handle large amounts of calcium whenever the need arises. (11) All the pumps mentioned play a significant role in maintaining calcium homeostasis. Plasma membrane integrity, on the other hand, fulfills a dual function in the maintenance of intracellular calcium homeostasis. An intact plasma membrane, impermeable to calcium ions forms a barrier to unwanted calcium inflow into the cell. Plasma membrane phospholipid and fatty acid composition are furthermore important in supporting the structure and function of the calcium homeostasis pumps. (12) Impairment of any of the mechanisms mentioned may lead to intracellular calcium dyshomeostasis.

3) Manifestations of intracellular calcium dyshomeostasis

A rise in intracellular free calcium can potentially activate a number of calcium sensitive enzymes that could culminate in cell dysfunction and damage. This cellular dysfunction and damage, resulting from the activation of calcium sensitive enzymes, could subsequently result in further aggravation of the initial rise in intracellular free calcium.

• As mentioned mitochondria become the single most important membrane system in the cell to act as long-term calcium buffers, but with the subsequent increase in mitochondrial calcium content functional and structural changes are triggered. The prolonged uptake of calcium by the mitochondria results in a reduction of cellular ATP content, since mitochondrial calcium transport takes precedence over oxidative phosphorylation. Calcium uptake occurs by an electrophoretic uniport that involves transport of calcium ions across the mitochondrial membrane without direct coupling to the movement of another ion. This net inward movement of positive charges, associated with due calcium flux, causes depolarisation of the mitochondrial membrane, which is immediately compensated for by an increase in the respiratory



activity followed by an increase in H⁺ extrusion. Thus mitochondrial H⁺ efflux coupled to electron transfer along the respiratory chain is the active energy-linked process driving the electrophoretic calcium uptake. This utilisation of the electrochemical gradient across the mitochondrial membrane for calcium uptake makes the electrochemical gradient unavailable for ATP production. (6) Furthermore, calcium uptake is followed by the activation of a mitochondrial associated phospholipase Phospholipase A₂ hydrolyses ester bonds of membrane A,. phospholipids, specifically phosphatidylcholines containing the fatty acid arachidonic acid in the sn₂ position. Activation of this enzyme may therefore be responsible for the dramatic ultrastructural changes in the crystal membranes and mitochondrial swelling with a prolonged elevation in intracellular calcium. (9 - 11) The structural and functional changes of mitochondria upon prolonged elevation in intracellular calcium, most notably result in the further decrease of ATP production. Many of the enzymes responsible for decreasing intracellular free calcium are energy-dependent, therefore decreased ATP production Therefore. could possibly result in decreased calcium extrusion. mitochondrial dysfunction following the initial elevation in intracellular free calcium could result in the further progressive aggravation of intracellular calcium dyshomeostasis.

• An elevation in intracellular free calcium can possibly result in the activation of calpain, a neutral protease. Calpain activation results in degradation of cytoskeletal proteins such as actin and tubulin, and cytoskeletal and plasma membrane organisational proteins that are responsible for anchoring cytoskeletal elements to the plasma membrane. This results in alterations in the microfilament network and subsequently plasma membrane blebbing. Many of these organisational proteins restrict different membrane proteins to specific regions of the plasma membrane. Degradation of these organisational proteins



necessary for the restriction of membrane proteins results in the subsequent loss of cell polarisation and cell dysfunction. (9 - 11, 13)

An inappropriate elevation in intracellular free calcium results in the unwanted production of oxygen radicals by phagocytic cells. Phagocytic cells such as neutrophils possess a H₂O₂/O₂-generating NADPH oxidase. NADPH oxidase activation is triggered by the influx of calcium resulting in the production of oxygen radicals. Therefore, an inappropriate elevation in intracellular free calcium can result in the unwanted activation of the NADPH oxidase and the excessive production of oxygen radicals. In addition, oxidative stress, a condition in which the pro-oxidant: anti-oxidant ratio favours the affect intracellular calcium homeostasis is known to Disruption of many of the intracellular calcium mechanisms. homeostasis mechanisms due to an increased production of oxygen radicals, can potentially result in the elevation of basal intracellular free calcium. (15, 16) This disruption of intracellular calcium homeostasis mechanisms is the result of the reaction of oxygen radicals with cellular The superoxide anion is one of the oxygen radicals constituents. produced by the NADPH oxidase. The superoxide anion is not particularly reactive but is capable of diffusing over relatively large distances in the cell where, in the presence of iron or copper, a metalcatalysed Haber-Weiss reaction can occur with the formation of the highly reactive hydroxyl radical. (9) This hydroxyl free radical rapidly reacts with cellular constituents in its immediate vicinity and is the key radical species in causing cellular damage. With regard to intracellular calcium homeostasis the reaction with membrane lipids (to cause nonenzymatic lipid peroxidation), and the reaction with the sulphydryl groups contained in cellular proteins appear to be of particular importance. Lipid peroxidation can result in the loss of membrane integrity and a further influx of calcium into the cytosol due to the breakdown of the membrane barrier between the cytosol and the high



Additionally, a change in calcium containing pools. (9, 15, 17) membrane phospholipid and fatty acid composition as a result of lipid peroxidation may cause an alteration in the lipid microdomain of the Ca²⁺-ATPase and the other enzymes responsible for lowering of intracellular free Ca2+. The lipid microdomain surrounding an integral membrane protein determines the activity of the protein. This is mainly due to the support of the three-dimensional structure of the protein. Therefore, an optimal lipid environment is an important factor that can influence membrane protein function such that a change in the microdomain of the calcium pumps can result in the suppression of calcium extrusion. (12, 18-23) Furthermore, the calcium pumps responsible for decreasing calcium with the unwanted accumulation of calcium in the cytosol are highly sensitive to oxygen radicals. unchecked production of oxygen radicals can possibly react with protein sulphydryl groups. The sulphydryl groups form part of the active center of the calcium pump, oxidation of these groups result in the suppression of calcium extrusion. Oxidative stress can also result in (16, 24)disruption of microfilament organisation. Thiol oxidation of actin Microfilament molecules perturbs microfilament organisation. organisation determines the size and distribution of the intracellular calcium stores, through the interaction with intracellular calcium store membranes. A change in the distribution of intracellular calcium stores can affect intracellular calcium signalling processes. During conditions of oxidative stress, the loss of both reduced pyridine nucleotides and protein thiols inhibits mitochondrial calcium sequestration resulting in the loss of this vital defence mechanism against elevated calcium while at the same time releasing calcium into the cytosol. (9) Lastly, one of the important properties of the calcium extrusion pump is its stimulation by calmodulin, which is due to a direct interaction of calmodulin with the pump. (25) Calmodulin binds four calcium ions, and this binding of calcium is accompanied by a substantial



conformational change whereupon the protein assumes an active form. The active form of calmodulin is now capable of interaction with the calcium pump. These high-affinity calcium-binding sites on the surface of calmodulin include methionine residues. Oxidation of these residues, due to the reaction of oxygen radicals with the sulphydryl group results in the alteration of calcium binding properties with the apparent loss of high-affinity calcium binding sites. The loss of the high-affinity calcium binding sites results in decreased activation of calmodulin and diminished capability of the up-regulation of calcium-pump activity. (26) This can result in decreased calcium-pump activity and the subsequent rise in intracellular free Ca²⁺.

An elevation in intracellular free calcium results in the activation of a cytosolic phospholipase A₂ (PLA₂). PLA₂ hydrolyses specific ester bonds of a specific membrane phospholipid; phosphatidylcholine with arachidonic acid in the sn₂ position. The activation of PLA₂ as a result of an elevation in intracellular free calcium upon agonist stimulation is a normal physiological response. It is when an unwanted elevation in intracellular free calcium occurs that PLA2 activation can result in substantial membrane damage. PLA₂ activation consequently has been considered a candidate for causing cellular damage, since the loss of membrane integrity could result in an unwanted influx of calcium from the extracellular medium. (9, 27) Membrane integrity play an important role in the maintaining of cell viability, therefore an increase in membrane breakdown will result in an increase in membrane repair mechanisms. Acyltransferase is one of the enzymes responsible for membrane phospholipid and fatty acid resynthesis. For the adequate functioning of this enzyme ATP is required. One of the possible results of an elevation in intracellular free calcium is the reduced production of ATP. An unwanted elevation in intracellular free Ca2+ could therefore result in the perturbation of membrane repair due to the inadequate ATP production for acyltransferase activity.



To ensure adequate signalling responses cells have evolved an ingenious mechanism of signalling based on presenting calcium as brief spikes organised as oscillating regenerative waves. (28) It is generally accepted today that calcium spiking is an important signalling mechanism and that various patterns of intracellular calcium oscillations are of physiological significance. (5) These kinds of responses are both spatially and temporally regulated and are obtained when cells are concentrations of agonists representing with stimulated This is in contrast with the calcium **(5)** physiological situation. response obtained when cells are stimulated with high concentrations of agonist, i.e., a sustained elevation in intracellular free calcium and the Calcium appears to be the subsequent return to baseline levels. transduction element in these responses since receptors and enzymes displaying various degrees of sensitivity to the surrounding calcium concentration regulates the generation of these calcium patterns. Therefore, perturbation in intracellular free Ca2+ could result in the unwanted activation of these receptors and enzymes resulting in inappropriate calcium signalling responses. Following, is an account of these receptors and enzymes sensitive to calcium and the possible influence of an unwanted rise in intracellular free Ca2+ on calcium Agonist-receptor interaction stimulates these signalling processes. intracellular calcium oscillations. Initially, agonist-receptor interaction stimulates the activity of a phosphatidyl inositol specific phospholipase Activation of this enzyme leads to the hydrolysis of phosphatidyl inositol 4,5-bisphosphate and formation of downstream effectors, water soluble 1,4,5-trisphosphate (InsP₃) and lipid soluble diacygliserol (DAG). The initial PI-PLC isoform activated as a result of receptor stimulation involves the activation of a specific PI-PLC isoform that is coupled to the receptor via a G-protein. This PI-PLC isoform appears to be largely insensitive to calcium. However, during the later phase of the response a second PI-PLC isoform, a calcium-dependent



isoform is activated. It is hypothesised that the activation of this calcium dependent PI-PLC isoform by an increase in intracellular free calcium, plays a role in the generation of calcium oscillations. inappropriate elevation in intracellular free calcium, therefore can result in perturbation of these intracellular calcium oscillations due to the unwanted activation of the calcium-dependent PI-PLC. It is the primary action of InsP₃ to liberate calcium ions from intracellular organelles, mostly calciosomes (the equivalent of the calcium pool in the sarcoplasmic reticulum of skeletal muscle) into the cytosol. (29, 30) InsP₃ releases calcium from these stores by binding to the InsP₃-sensitive calcium channels in the membranes of these organelles. activated channel is composed of four non-covalently bound identical subunits. The InsP₃ binding domain resides at the extreme N-terminal region, whilst the C-terminal portion contains six putative membranespanning domains that make up, or at least contain the calcium channel. The region between the InsP₃-binding domain and the putative calciumchannel constitutes the so-called 'coupling' domain, some 1400-1500 amino acids in length, which lies in the cytoplasm. This so-called 'coupling' domain constitutes the site for modulatory actions. (5) This domain contains a site for calcium binding. There are at least four different types of InsP₃ receptors derived from three or four distinct Between these different types of InsP₃ receptors there exist a strong homology in the membrane-spanning domains but large divergencies are indicated in the cytoplasmic modulatory domains. It is hypothesised that since the InsP₃-receptor contains a site for calcium binding in the modulatory domain and in the light of the fact that there exists receptor subtypes containing large divergencies in these modulatory domains, that the functional distribution of these InsP₃receptor subtypes play a significant role in the regulation of these spatially and temporally organised calcium patterns. Indeed there are many indications that the InsP₃-receptor plays the central role as



possible regulator of these complex intracellular calcium signalling There are indications that an inappropriate elevation in patterns. intracellular free calcium may affect these InsP₃-receptor functions. This can be the result of one of two factors. An elevation in intracellular free calcium results in cytoskeletal disorganisation followed by a change in the structure of the calcium stores. The structure of these calcium stores can be controlled by the integrity of the cytoskeleton since the internal membranes are associated with the elements of the cytoskeleton and it has been shown that cytoskeletal disruption leads to fragmentation of the InsP₃-sensitive calcium stores. The intracellular calcium signalling patterns are spatially organised and these spatially organised calcium patterns are possibly determined by the functional distribution of InsP₃receptor subtypes. Therefore the structure of the InsP₃-sensitive calcium store is an important determinant for the generation of these spatially organised calcium signalling patterns. The following calcium signalling processes are determined by the distribution of InsP₃-receptor subtypes: Upon agonist activation the intracellular calcium signal is initiated at a discrete subcellular locus adjacent to a small region of the plasma membrane. This intracellular calcium wave initiation domain does not simply result from a polarised distribution of agonist receptors but indications are that there may be specialised subcellular regions for intracellular calcium release containing InsP₃-receptor subtypes that are more sensitive to InsP₃. (31) Various intracellular calcium responses do not involve the entire cell and are therefore localised within a specific To avoid the undesirable activation of calcium-dependent processes elsewhere in the cell, calcium signalling patterns are produced locally which are confined to the relevant region. The production of such local calcium signalling patterns are largely determined by the spatial arrangement of the InsP₃-sensitive stores. (31) In cells displaying chemotaxis the polarisation of the cell changes with every new There are indications of a calcium gradient within these movement.



cells that underlies the activities of calcium-dependent processes that favour the polarisation and directed locomotion of these cells. The gradient changes continually as the polarisation of the cell changes. The direction of the new calcium gradient within cells is established by the reorganisation of the InsP₃-sensitive stores. An elevation in (31)intracellular free calcium not only results in the disorganisation of the calcium stores and the subsequent altered distribution of the InsP₃receptors, but the free calcium concentration can also modulate the The free calcium degree of InsP₃-evoked channel opening. concentration is an important factor in determining the degree of InsP₃gated channel opening. These InsP₃-gated calcium channels display a bell-shaped calcium response curve. In the physiological calcium concentration range (0.1 - 1 µM) small changes in the calcium concentration can have dramatic effects on the probability of channel opening. When the intracellular free calcium concentration is increased from 0.1 to about 0.3 µM there is a marked increase in the open-state probability, but when the calcium concentration is further increased to 1 μM a marked fall in the degree of InsP₃-evoked channel opening is observed. Therefore an increase in intracellular free calcium concentration to above 0.3 µM can result in an inhibition of InsP₃evoked calcium release. (30)InsP₃ liberation not only results in releasing of calcium from intracellular calcium stores as explained in the previous section, but also activates calcium entry pathways in the plasma membrane. It is most probably not InsP₃ per se that activates the calcium channels in the plasma membrane, but a product of the enzymatic modification of InsP₃ namely inositol-1.3.4.5 tetrakisphosphate (InsP₄). The plasma membrane InsP₃-receptor has a lower affinity for InsP₃ but a higher affinity for InsP₄ when compared to the well-known endoplasmic reticulum InsP₃-receptor, therefore providing for such a role for InsP₄. InsP₄ is formed as a result of the



phosphorylation of InsP₃ by InsP₃-kinase. These two processes, i.e., the release of calcium from the intracellular calcium storage organelles and the influx of calcium from the extracellular medium, are highly The InsP₃-kinase coordinated and possibly regulated by calcium. responsible for the formation of InsP₄ is calcium activated, therefore InsP₃ liberation results in the releasing of intracellular calcium and subsequently the activation of InsP3-kinase. The subsequent formed InsP4 triggers the influx of calcium from the extracellular medium. An unwanted increase in intracellular free Ca2+ could result in the perturbation of the interaction between these processes. (30) It is not only the calcium release and calcium influx pathways that are regulated by intracellular free calcium, but the pathways resulting in the termination of intracellular calcium signalling responses are also These processes; calcium influenced by intracellular free calcium. release, calcium influx and calcium efflux are highly coordinated. It was indicated that pulsatile calcium extrusion occurred synchronously with the cytosolic calcium spikes. The activity of the Ca2+-ATPase mostly regulates the calcium extrusion and calcium sequestration at the end of the response. In the calcium range 0.1 – 0.3 μM , where InsP₃-gated channel opening is increasing very steeply with a rise in intracellular free calcium, calcium extrusion is moderately activated. In the calcium range $0.3 - 1 \mu M$ where InsP₃-gated calcium channel opening decreases with increasing free intracellular calcium, calcium extrusion rises Intracellular sharply as intracellular free calcium increases. (30)calcium per se plays an important role in the coordination of these processes. Three processes activated by the increase in intracellular free calcium results in the extrusion of calcium by the Ca2+-ATPase. An increase in intracellular free calcium results in calmodulin activation. In the absence of calmodulin only a small increase in the calcium transport rate is indicated with the increase in intracellular free calcium. But in the presence of calmodulin, a small increase in intracellular free calcium



results in a dramatic increase in calcium pumping activity. calcium-calmodulin-dependent activation of the Ca2+-ATPase calcium pump increases the capacity or Vmax of the pump 15-20 fold and the efficiency two-fold, i.e., lowers the Km for calcium. The calciumcalmodulin complex binds to the Ca²⁺-ATPase resulting in an increase in pump activity and the more rapid extrusion of calcium. The enzyme protein kinase C (PKC) also plays a significant role in the activation of the Ca2+-ATPase. PKC is activated as a consequence of both the initial rise in the diacylglyserol (DAG) content of the plasma membrane and the transient rise in intracellular free calcium. Upon activation PKC moves from the cytosol to the plasma membrane. When PKC associates with the plasma membrane, it undergoes a conformational change to become a calcium-sensitive, plasma membrane associated protein kinase. The activity of this form of the kinase is regulated by the rate of calciumcycling across the plasma membrane. (29) PKC activated by calcium leads to phosphorylation of the Ca2+-ATPase pump, resulting in a 4 - 6 fold increase in pump activity. A third process possibly contributing to the coordination of calcium oscillations is the calcium-mediated activation of PLA₂. PLA₂ activation leads to the release of arachidonic Arachidonic acid possibly performs a dual function in the acid. coordination of intracellular calcium signalling patterns, i.e., arachidonic acid can inhibit InsP₃-evoked calcium mobilisation and can at the same time activate calcium extrusion. (31) In conclusion, many calcium sensitive receptors and enzymes are responsible for the generation of the In order to assure adequate complex calcium signalling patterns. signalling responses, these various processes are highly coordinated. Since the receptors and enzymes responsible for the generation of the calcium signalling patterns are calcium sensitive, calcium seems to be the modulator of its own signalling cycle. Therefore, perturbation in intracellular calcium could culminate in disturbed calcium signalling responses.



4) Intracellular calcium status in chronic renal failure

It was Massry who suggested that chronic renal failure is a state of calcium toxicity. (32) Soft tissue calcification is indeed a common finding in chronic renal failure (CRF), with an increase in calcium content known to occur in the cornea, skin, blood vessels, brain, peripheral nerves, heart, lungs, pancreas, liver, epididymal fat and testis of patients as well as animals with chronic renal failure. Intracellular free calcium would appear to reflect this increased tissue content of calcium. An elevation in basal intracellular free calcium in various cell types in CRF is documented, indicating a potential association between this increase in the calcium content of tissues with an elevation in basal intracellular free calcium. A summary of published results of basal intracellular free calcium in humans and rats with chronic renal failure can be seen in Table 1. The cell types in which the levels have already been determined include brain synaptosomes, pancreatic islets, cardiac myocytes, thymocytes, B cells, T cells, neutrophils, adipocytes, and platelets. (33-37, 40-42, 44-46, 48, 50-51)

4.1) Pathological factors associated with CRF possibly contributing to the derangements in intracellular calcium homeostasis

There are a multitude of possible factors contributing to the elevation in basal intracellular free calcium in the CRF patient including secondary hyperparathyroidism, oxidative stress, deranged phospholipid and fatty acid metabolism and reduced ATP concentrations.

4.1.1) Secondary hyperparathyroidism

It is hypothesised that an elevation in PTH levels in these patients could be the main cause for the increase in basal intracellular free calcium in the various cell types. (33, 44)



Table 1. Intracellular free Ca $^{2+}$ in various cell types in humans and rats with CRF

| Cell type | Model | CRF [Ca ²⁺]i | Normal [Ca²+]i | Fluorescent calcium indicator | Reference |
|--------------|-----------------|-----------------------------|-------------------|-------------------------------------|-----------|
| Neutrophils | HD patients | 73±3.6nM | 42±0.09nM | Fura-2 | 34 |
| Neutrophils | HD patients | 92nM | 60nM | Fura-2 | 35 |
| Neutrophils | HD patients | 60±1.2nM | 46±1.2nM | Fura-2 | 36 |
| Neutrophils | HD patients | 79±5.1nM | 52±3.4nM | Fura-2 | 37 |
| Neutrophils | HD patients | 68±1.1nM | 57±1.1nM | Fura-2 | 38 |
| | CRF rats | 133±3.1nM | 104±1.3nM | | |
| Neutrophils | CRF rats | 149±2.7nM | 108±2.4nM | Fura-2 | 39 |
| Platelets | CRF patients | 117±33nM | 47±14nM | Fura-2 | 40 |
| Platelets | HD patients | 137±8.86nM | | Fura-2 | 41 |



Table 1. (continue) Intracellular free Ca $^{2+}$ in various cell types in humans and rats with CRF

| Cell type | Model | CRF [Ca²+]i | Normal [Ca ²⁺]i | Fluorescent calcium indicator | Reference |
|-------------------------|-----------------|----------------|--------------------------------|-------------------------------------|------------|
| Platelets | CRF patients | 138±16nM | 83±7nM | Fura-2 | 42 |
| Platelets | CRF patients | 173±44nM | 150±30nM | Indo-1 | 43 |
| T-cells | HD patients | 87nM | 55nM | Fura-2 | 44 |
| B-cells | HD patients | 115±5.2nM | 86±4.4nM | Fura-2 | 4 5 |
| Brain synaptosomes | CRF rats | 437±18nM | 345±9nM | Fura-2 | 46 |
| Brain synaptosomes | CRF rats | 445±10nM | 353±7.5nM | Fura-2 | 47 |
| Islets of Langerhans | CRF rats | 252±7.4nM | 137±4.5nM | Fura-2 | 48 |
| Heart cells | CRF rats | 101±3.2nM | 56±1.6nM | Fura-2 | 49 |



Indeed, this hypothesis finds much credit, since prevention of the elevation in PTH levels by parathyroidectomy or vitamin D therapy, or by the prevention of the action of PTH by calcium channel blocker administration, resulted in the normalisation of basal intracellular free calcium levels. (35, 37, 48, 51-53) It is generally accepted that patients with CRF have secondary hyperparathyroidism and elevated blood levels (45) PTH tightly regulates plasma calcium and phosphate concentrations by modulating the activity of specific cells in the bone and kidney. (54, 55) In the normal functional kidney, PTH is known to increase calcium reabsorption and to stimulate phosphate excretion. PTH also activates the renal synthesis of the active vitamin D metabolite, which, in turn is able to stimulate or inhibit intestinal absorption of calcium and phosphate and is able to increase or decrease osteoclast mediated bone resorption depending on the concentration of vitamin D. (56) These actions of PTH are classical hormonal functions. It is with an abnormal elevation in PTH levels that PTH targets many other cell types. It has been shown that PTH can cause a sharp increase in the basal intracellular free calcium content of pancreatic islets cells, thymocytes (57), cardiac myocytes, hepatic cells, adipocytes, kidney cells, and osteoblasts. This effect of PTH is receptor-mediated. PTH activates the voltage-dependent calcium channels. This PTH-mediated calcium influx can be blunted or prevented by calcium channel blockers such as verapamil or nifedipine. The influence of PTH on nonconventional targets is supported by recent data which demonstrated that the mRNA for the PTH/PTHrP receptor is present in non-classical targets for PTH such as heart, brain, spleen, aorta, ileum, skeletal muscle, lung and testis. (33, 54) In addition to the experimental evidence for the central role of PTH in the elevation of intracellular free calcium in different cell types, indications suggest deranged cellular functions in association with an elevation in PTH. These include studies indicating deranged cellular functions in combination with



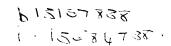
either an elevation in PTH or an elevation in PTH accompanied by an elevation in intracellular free calcium. Intracellular free calcium plays a paramount role in the regulation of a multitude of cellular functions. Intracellular calcium dyshomeostasis in CRF may thus be a major contributor to abnormal cellular function in uraemia.

4.1.2) Oxidative stress

Chronic renal failure patients receiving maintenance haemodialysis treatment (MHT) are continuously exposed to oxidative stress. Oxidative stress is a state of increased pro-oxidant levels and/or a deficit in the anti-oxidant capacity. In the MHT patient both an increase in prooxidant production and a decrease in anti-oxidant mechanisms are indicated. The increase in pro-oxidant production is mainly due to the relentless activation of polymorphonuclear leucocytes (PMNL) during haemodialysis. During extracorporeal circulation activation of PMNL's results in increased free oxygen radical production. (58) This activation of PMNL's is the result of the exposure of the blood constituents to the dialyser membrane. Various pathways are activated resulting in PMNL activation. These pathways include both complement-dependent and complement-independent PMNL activation. Complement-(59)dependent activation leads to the production and binding of C5a and C5a des Arg to specific PMNL receptors inducing aggregation, adherence to endothelial cells, releasing of oxygen radicals, degranulation, arachidonic acid metabolism, and enhanced phagocytotic capacity. (60) Complement-independent activation of PMNL's is the result of either direct stimulation of PMNL's due to interaction with the dialyser membrane or binding of monocyte-derived cytokines to their respective PMNL receptors. Cytokine exposure primes the respiratory burst to generate elevated levels of oxidants. (9, 61) There are indications that the activation of PMNL's during extracorporeal circulation involves an increase in intracellular free calcium. (59, 60, 62, 63) The process



responsible for the reduction of molecular oxygen to superoxide anion is catalysed by a multicomponent NADPH-oxidase and this process is activated and regulated by intracellular free calcium. It is therefore anticipated that an increase in intracellular free calcium will result in an increase in oxygen radical production. (64) Anti-oxidant mechanisms include various enzymes responsible for the reduction of the oxygen radicals to water. These anti-oxidant enzymes include superoxide dismutase, glutathione peroxidase and catalase. Superoxide dismutase results in the dismutation of the superoxide anion (O₂) to hydrogen peroxide (H₂O₂). In the following reactions the H₂O₂ formed is reduced to H₂O by either glutathione peroxidase or catalase. This subsequent reduction of H₂O₂ prevents the potential formation of hydroxyl radicals. If H₂O₂ accumulates due to insufficient glutathione peroxidase and catalase activity, hydroxyl radicals may be formed by the heavy metal catalysed Fenton reaction. During the glutathione peroxidase catalysed reduction of hydrogen peroxides, glutathione (GSH) serves as an electron donor, and the glutathione disulphide (GSSG) formed in the reaction is subsequently reduced back to GSH by glutathione reductase at the expense of nicotinamide adenine dinucleotide phosphate (NADPH). (9) NADPH is produced by normal activity of the hexose monophosphate shunt. (65)In MHT patients, a metabolic block of the pentose phosphate shunt has been described that impairs the reduction of oxidised glutathione due to the reduced NADPH formation. (65, 66) Most of the enzymes responsible for the reduction of oxygen radicals to H₂O contain certain trace elements at their active sites. Trace element levels are altered in MHT patients. The dialysis procedure further could appreciably alter the body's supply of trace elements by removing some, whereas contaminating trace elements in the dialysis fluid could be transferred to the patient. A reduction in the trace elements necessary for anti-oxidant enzyme activity can result in reduced anti-oxidant activity and therefore an increase in oxidative stress. The concentrations





of Zn and Se for instance are reduced in the MHT patient. These trace elements are necessary for normal activity of the anti-oxidant enzymes; superoxide dismutase and glutathione peroxidase. (67-69) Other trace elements catalyse the production of hydroxyl radicals from hydrogen peroxide, via the heavy metal catalysed Fenton reaction. An increase in these trace elements would therefore result in an elevation in oxidative stress. (68) Under conditions of oxidative stress, when the cell must cope with large amounts of H₂O₂, the role of glutathione oxidation exceeds the slower rate of GSSG reduction by glutathione reductase, and GSSG accumulates. To avoid the detrimental effects of increased intracellular levels of GSSG (e.g. formation of mixed disulphides with protein thiols) the cell actively secretes GSSG, which can lead to depletion of the intracellular glutathione pool. (9) In addition vitamins A, E and C are powerful anti-oxidants. Vitamin E and t-carotene are lipid-soluble anti-oxidants preventing radical mediated breakdown of poly-unsaturated fatty acids and vitamin C acts as a reductant in the cell, both regenerating oxidised vitamin E and preventing oxidation of cytosolic thiols. (9, 65, 66) In the CRF/MHT patient an increase in oxidative stress can result in the overconsumption of available antioxidant vitamins.

The increase in the oxidative stress in the CRF patient receiving maintenance haemodialysis due to both an increase in pro-oxidant production and a decrease in anti-oxidant activity, can potentially result in intracellular calcium dyshomeostasis. Unwanted oxygen radical production can result in the oxidation of membrane polyunsaturated fatty acids. Oxidation of membrane polyunsaturated fatty acids can subsequently result in an increase in the permeability for calcium ions causing an unwanted influx of calcium ions. (58, 65, 70) Furthermore, an unwanted oxidation of membrane polyunsaturated fatty acids can result in the derangement of membrane fatty acid composition possibly



due to an increase in the saturated fatty acid/unsaturated fatty acid ratio. The membrane fatty acid composition is an important factor regulating membrane-associated protein function. It is well known that various membrane-associated enzymes are highly sensitive to the membrane micro-environment. The Ca²⁺-ATPase calcium pump responsible for lowering intracellular free calcium is one such membrane-associated enzyme that is very sensitive to membrane fatty acid composition. (71-73) An increase in oxidative stress will furthermore result in the oxidation of cellular proteins. Many of the membrane-associated proteins are rich in SH-groups which are targets for oxygen radical action. The Ca²⁺-ATPase is a thiol-rich protein at the cytosol-membrane surface. (74) Therefore, an increase in the oxidative stress may result in oxidation of the SH-groups of the pump and suppression of the pump's activity. (75)

4.1.3) Inhibition of the calcium pump

A circulating inhibitor of the calcium pump is indicated in the plasma of CRF patients. This inhibitor is dialysable, smaller than 3000 molecular weight, heat-stable, and said to be protease resistant. (76, 77) It is hypothesised that this inhibitor of the calcium pump may be one of the factors contributing to the elevation in intracellular free calcium in various cells in the CRF patient.

4.1.4) Inhibition of the Na⁺-K⁺-ATPase

There are indications of an elevation in plasma levels of natriuretic factor in CRF. Natriuretic factor is a potent inhibitor of the Na⁺-K⁺-ATPase. Na⁺-K⁺-ATPase plays an important role in the maintenance of intracellular calcium homeostasis. Na⁺-K⁺-ATPase maintains the sodium gradient across cellular membranes, a high sodium concentration extracellularly and a low sodium concentration on the inside. This sodium gradient provides the energy necessary for the



extrusion of calcium via the Na⁺-Ca²⁺-exchanger. A decrease in the activity of the Na⁺-K⁺-ATPase may ultimately result in an elevation in intracellular free calcium. (78)

4.2) Results of an elevation in basal intracellular free calcium in chronic renal failure

4.2.1) Deranged membrane phospholipid metabolism

It is indicated that the content of phospholipids in brain synaptosomes and specifically phosphatidyl inositol (PI), phosphatidyl serine (PS), and phosphatidyl ethanolamine (PE) are significantly reduced in CRF. This reduction in the phospholipid content is attributed to the increase in basal intracellular free calcium caused by the chronic exposure to PTH. (79) Phospholipids are important components of the cell membrane and play an important role in maintaining membrane fluidity and membrane protein function. (80) Changes in phospholipid composition of cell membranes may therefore affect the activity of proteins and enzymes present in the membranes. (81, 82) The various enzymes responsible for lowering of intracellular free calcium, Ca2+-ATPase, Na+/Ca2+exchanger and the Na⁺/K⁺-ATPase are extremely sensitive to membrane phospholipid composition. (83, 84)Indeed, the changes in synaptosomal phospholipid composition are associated with changes in Na⁺/K⁺-ATPase activity. This change in the activity of the enzymes responsible for lowering of intracellular free calcium may thus possibly aggravate the initial increase in intracellular free calcium. exposure to PTH has been shown to inhibit the activity of many cellular enzymes. It is therefore possible that chronic exposure to excess PTH may also inhibit the activity of enzymes involved in the synthesis of phospholipids. (79, 85)

4.2.2) Deranged membrane fatty acid synthesis



It is indicated in the CRF patient that intracellular free calcium accumulation results in inhibition of various cellular enzymes. One of the enzymes possibly inhibited by an elevation in intracellular free calcium is carnitine palmitoyl transferase (CPT). (79) Adequate CPT activity is important in the maintaining of a favourable acyl CoA/free CoA ratio. CPT maintains a favourable acyl CoA/free CoA ratio by forming acylcarnitine from carnitine and acyl CoA. Membrane fatty acid turnover depends on both long chain acyl CoA synthetase and lysophospholipid acyl CoA transferase (LAT). The former generates acyl CoA from free fatty acids and CoA, while the latter reacylates lysophospholipids by using acyl CoA as substrate. When the rate of formation of acyl CoA becomes different from that of its utilisation for reacylation, acyl CoA and the acyl CoA/free CoA ratio become altered, leading to disturbances in membrane fatty acid turnover. In the CRF patient reduced activity of CPT is indicated. (85)Under such circumstances, the lack of buffering activity of CPT alters the acyl CoA/free CoA ratio, and eventually the availability of fatty acid moieties for reacylation are impeded. Additionally, the relatively high concentrations of free CoA may further act as an inhibitor of the enzyme LAT. This possible reduced activities of CPT and LAT may result in the inadequate repairing of membrane fatty acids damaged by free radicals and the loss of membrane integrity. (85)

4.2.3) Decreased ATP production

An elevation in basal intracellular free calcium inhibits mitochondrial oxidation, resulting in the decrease of basal and stimulated ATP production. (53, 78)

5) Aim of this study

Over the past decade a significant amount of work has been done on intracellular calcium in chronic renal failure. It should however be



remembered that the term intracellular calcium is used relatively nonspecifically, as can be seen from the various publications on the subject where the expression intracellular calcium is used to denote anything from total cellular calcium to diffusible calcium content, depending on the methods available in specific laboratories. The complete picture of the intracellular calcium status, including a comparison between the intracellular content of the various calcium compartments, the characteristics of the transmembrane calcium movement, the possible causes which may contribute to the assumed intracellular calcium disturbances in CRF, as well as the contribution of such disturbances to the pathogenesis of chronic renal failure still need to be clarified. At this point in time it can, with a relative amount of certainty, be assumed that both the total intracellular calcium content and the intracellular free Ca2+ are increased in end stage chronic renal failure patients. This despite a small number of contradictory reports. Very little is, however, known about the movement across the cell membranes as well as about the intercompartmental shifts. The parathyroid hormone disturbance in chronic renal failure is at present seen as the major contributor to the deranged calcium status, but the evidence for such an assumption is, however tempting, still far from conclusive.

The aim of this study was to contribute to the knowledge on intracellular calcium in chronic renal failure by

a) Examining the intraneutrophilic free cytosolic Ca²⁺ content in end stage chronic renal failure patients on maintenance haemodialysis treatment. The aim is not to evaluate the influence of renal failure *per se* on calcium metabolism, but rather to determine the status of intracellular calcium in the maintenance haemodialysis patient as it presents as a result of renal failure, dialysis treatment and medication. The rationale is that firstly the disturbed cellular functions and other



effects of the disturbed calcium content are functions of the existing calcium status and secondly that the application of stringent exclusion criteria would eliminate too many patients from the already small available group.

- b) Examining the characteristics of the transmembrane movement of calcium, i.e., the calcium fluxes.
- c) Investigating the intracellular distribution of calcium in the neutrophils of the patients by means of electron microscopy.
- d) Exploring the possibility of a link between the alterations in intracellular calcium status and factors known to influence the calcium status. The factors investigated in this study include the lipid composition of the membrane, the oxidative status as reflected by anti-oxidant vitamin levels, the levels of parathyroid hormone, and ionised serum calcium.

In order to establish conditions inducive to accurate research, the techniques for calcium determinations, which did not previously exist in the current laboratory, had to be developed and evaluated. This constituted a rather large part of the project.

The rest of the chapters of this dissertation are presented in the following sequence

Chapter 2: Evaluation of the techniques for the determination of intracellular free calcium, transmembrane calcium fluxes and intracellular distribution of calcium

Chapter 3: Materials and methods

Chapter 4: Results

Chapter 5: Discussion and conclusions



References

- 1) Stein JH, editor. Nephrology. New York: Grune & Stratton, Inc; 1980.
- 2) Clapham DE. Calcium signaling. Cell 1995 Jan 27; 80: 259-268.
- 3) Berridge MJ. Inositol trisphosphate and calcium signalling.
 Nature 1993 Jan 28; 361: 315-325.
- 4) Heizmann CW, Hunziker W. Intracellular calcium-binding proteins: more sites than insights. TIBS 1991 Mar; 16: 98-103.
- 5) Shuttleworth TJ. Intracellular Ca²⁺ signalling in secretory cells.

 The Journal of Experimental Biology 1997; 200: 303-314.
- 6) Humes HD. Role of calcium in the pathogenesis of acute renal failure. American Journal of Physiology 1986; 250: F579-F589.
- Pozzan T, Rizzuto R, Volpe P, Meldolesi J. Molecular and cellular physiology of intracellular calcium stores. Physiological Reviews 1994 Jul; 74(3): 595-636.
- 8) Carafoli E. The Ca²⁺ Pump of the Plasma Membrane. The Journal of Biological Chemistry 1992 Feb 5; 267(4): 2115-2118.
- Duncan CJ, editor. Calcium, Oxygen Radicals and Cellular Damage. Cambridge, Great Britain: Cambridge University Press; 1991.
- Farber JL. The Role of Calcium in Cell Death. Life Sciences 1981;29: 1289-1295.
- 11) Trump BF, Berezesky IK. Calcium-Mediated Cell Injury and Cell Death. FASEB 1995; 219-228.
- 12) Matthews PLJ, Bartlett E, Ananthanarayanan VS. Reconstitution of Rabbit Sarcoplasmic Reticulum Calcium ATPase in a Series of Phosphatidylcholines Containing a Saturated and an Unsaturated Chain: Suggestion of an Optimal Lipid Environment. Biochemistry and Cell Biology 1993; 71: 381-389.



- 13) Edelstein CL, Ling H, Schrier RW. The Nature of Renal Cell Injury. Kidney International 1997; 51: 1341-1351.
- 14) Hallett MB, Davies EV, Campbell AK. Oxidase Activation in Individual Neutrophils is Dependent on the Onset and Magnitude of the Ca²⁺ Signal. Cell Calcium 1990; 11: 655-663.
- 15) Swann JD, Smith MW, Phelps PC, Maki A, Berezesky IK, Trump BF. Oxidative Injury Induces Influx-Dependent Changes in Intracellular Calcium Homeostasis. Toxicologic Pathology 1991; 19(2): 128-137.
- Shalev O, Lavi V, Hebbel RP, Eaton JW. Erythrocyte (Ca²⁺+Mg²⁺)-ATPase Activity: Increased Sensitivity to Oxidative Stress in Glucose-6-Phosphate Dehydrogenase Deficiency. American Journal of Hematology 1985; 19: 131-136.
- 17) Kimura M, Maeda K, Hayashi S. Cytosolic Calcium Increases in Coronary Endothelial Cells after H₂O₂ Exposure and the Inhibitory Effect of U78517F. British Journal of Pharmacology 1992; 107: 488-493.
- Palmeira CM, Santos MS, Carvalho AP, Oliveira CR. Membrane Lipid Peroxidation Induces Changes in •-[3H]Aminobutyric Acid Transport and Calcium Uptake by Synaptosomes. Brain Research 1993; 609: 117-123.
- 19) Dinis TCP, Almeida LM, Madeira VMC. Lipid Peroxidation in Sarcoplasmic Reticulum Membranes: Effect on Functional and Biophysical Properties. Archives of Biochemistry and Biophysics 1993 Mar; 301(2): 256-264.
- 20) Wu L-L, Liu M-S. Heart Sarcolemmal Ca²⁺ Transport in Endotoxin Shock: I. Impairment of ATP-dependent Ca²⁺ Transport. Molecular and Cellular Biochemistry 1992; 112: 125-133.



- 21) Cardoso CM, De Meis L. Modulation by Fatty Acids of Ca²⁺ Fluxes in Sarcoplasmic-Reticulum Vesicles. Biochemical Journal 1993; 296: 49-52.
- 22) Starling AP, East JM, Lee AG. Separate Effects of Long-Chain Phosphatidylcholine on Dephosphorylation of the Ca²⁺-ATPase and on Ca²⁺ Binding. Biochemical Journal 1996; 318: 185-788.
- 23) Starling AP, East JM, Lee AG. Effects of Phosphatidylcholine Fatty Acyl Chain Length on Calcium Binding and Other Functions of the (Ca²⁺+Mg²⁺)-ATPase. Biochemistry 1993; 32: 1593-1600.
- 24) Suzuki YJ, Ford GD. Inhibition of Ca²⁺-ATPase of Vascular Smooth Muscle Sarcoplasmic Reticulum by Reactive Oxygen Intermediates. American Journal of Physiology 1991; 261: H568-H574.
- Verma AK, Filoteo AG, Stanford DR, Wieben ED, Penniston JT.
 Complete Primary Structure of a Human Plasma Membrane Ca²⁺
 Pump. The Journal of Biological Chemistry 1988 Oct 5; 263(28): 14152-14159.
- Walsh M, Stevens FC. Chemical Modification Studies on the Ca²⁺-Dependent Protein Modulator: The Role of Methionine Residues in the Activation of Cyclic Nucleotide Phosphodiesterase. Biochemistry 1978; 17(19): 3924-3930.
- 27) Sun FF, Fleming WE, Taylor BM. Degradation of Membrane Phospholipids in the Cultured Human Astroglial Cell Line UC-11MG during ATP Depletion. Biochemical Pharmacology 1993; 45(5): 1149-1155.
- 28) Berridge MJ. Elementary and Global Aspects of Calcium Signalling. The Journal of Experimental Biology 1997; 200: 315-319.
- Rasmussen H. The Complexities of Intracellular Ca²⁺ Signalling.
 Biological Chemistry 1990 Mar; 371: 191-206.



- 30) Tepikin AV, Peterson OH. Mechanisms of Cellular Calcium Oscillations in Secretory Cells. Biochimica et Biophysica Acta 1992; 1137: 197-207.
- 31) Bock GR, Ackrill K, editors. Calcium waves, Gradients and Oscillations. England: John Wiley & Sons Ltd; 1995.
- 32) Massry SG, Fadda GZ. Chronic Renal Failure is a State of Cellular Calcium Toxicity. American Journal of Kidney Disease 1993 Jan; 21(1): 81-86.
- 33) Massry SG, Smogorzewski M. Mechanisms through which Parathyroid Hormone mediates its Deleterious Effects on Organ Function in Uremia. Seminars in Nephrology 1994 May; 14(3): 219-231.
- 34) Alexiewicz JM, Smogorzewski M, Gill SK, Akmal M, Massry SG.

 Time Course of the Effect of Nifedipine Therapy and Its
 Discontinuation on [Ca²⁺]_i and Phagocytosis of
 Polymorphonuclear Leukocytes from Hemodialysis Patients.

 American Journal of Nephrology 1997; 17: 12-16.
- 35) Haag-Weber M, Mai B, Hörl WH. Normalization of Enhanced Neutrophil Cytosolic Free Calcium of Hemodialysis Patients by 1,25-Dihydroxyvitamin D_3 or Calcium Channel Blocker. American Journal of Nephrology 1993; 13: 467-472.
- 36) Alexiewicz JM, Smogorzewski M, Fadda GZ, Massry SG.
 Impaired Phagocytosis in Dialysis Patients: Studies on
 Mechanisms. American Journal of Nephrology 1991; 11: 102-111.
- 37) Hörl WH, Haag-Weber M, Mai B, Massry SG. Verapamil Reverses Abnormal [Ca²⁺]_i and Carbohydrate Metabolism of PMNL of Dialysis Patients. Kidney International 1995; 47: 1741-1745.
- 38) Kiersztejn M, Smogorzewski M, Thanakitcharu P, Fadda GZ, Massry SG. Decreased O₂ Consumption by PMNL from Humans and Rats with CRF: Role of Secondary Hyperparathyroidism. Kidney International 1992; 42: 602-609.



- 39) Chervu I, Kiersztejn M, Alexiewicz JM, Fadda GZ, Smogozewski M, Massry SG. Impaired Phagocytosis in Chronic Renal Failure is mediated by Secondary Hyperparathyroidism. Kidney International 1992; 41: 1501-1505.
- 40) Moosa A, Greaves M, Brown CB, MacNeil S. Elevated Platelet-Free Calcium in Uremia. British Journal of Haematology 1990; 74: 300-305.
- 41) Lu K-C, Shieh S-D, Lin S-H, Chyr S-H, Lin Y-F, Diang L-K, et al. Hyperparathyrodism, Glucose Tolerance and Platelet Intracellular Free Calcium in Chronic Renal Failure. Quarterly Journal of Medicine 1994; 87: 359-365.
- 42) Raine AEG, Bedford L, Simpson AWM, Ashley CC, Brown R, Woodhead JS, et al. Hyperparathyroidism, Platelet Intracellular Free Calcium and Hypertension in Chronic Renal Failure. Kidney International 1993; 43: 700-705.
- Ware JA, Clark BA, Smith M, Salzman EW. Abnormalities of Cytoplasmic Ca²⁺ in Platelets from Patients with Uremia. Blood 1989 Jan; 73(1): 172-176.
- 44) Alexiewicz JM, Gaciong Z, Klinger M, Linker-Israeli M, Pitts TO, Massry SG. Evidence of Impaired T Cell Function in Hemodialysis Patients: Potential Role for Secondary Hyperparathyroidism. American Journal of Nephrology 1990; 10: 495-501.
- 45) Gaciong Z, Alexiewicz JM, Linker-Israeli M, Shulman IA, Pitts TO, Massry SG. Inhibition of Immunoglobulin Production by Parathyroid Hormone. Implications in Chronic Renal Failure. Kidney International 1991; 40: 96-106.
- 46) Smogorzewski M, Koureta P, Fadda GZ, Perna AF, Massry SG. Chronic Parathyroid Hormone Excess In Vivo Increase Resting Levels of Cytosolic Calcium in Brain Synaptosomes: Studies in



- the Presence and Absence of Chronic Renal Failure. Journal of the American Society of Nephrology 1991; 1: 1162-1168.
- 47) Hajjar SM, Smogorzewski M, Zayed MA, Fadda GZ, Massry SG. Effect of Chronic Renal Failure on Ca²⁺ ATPase of Brain Synaptosomes. Journal of the American Society of Nephrology 1991; 2: 1115-1121.
- 48) Fadda GZ, Hajjar SM, Perna AF, Zhou XJ, Lipson LG, Massry SG.
 On the Mechanism of Impaired Insulin Secretion in Chronic
 Renal Failure. Journal of Clinical Investigation 1991; 87: 255-261.
- 49) Smogorzewski M, Tian J, Massry SG. Down-Regulation of PTH-PTHrP Receptor of Heart in CRF: Role of [Ca²⁺]_i. Kidney International 1995; 47: 1182-1186.
- 50) Perna AF, Fadda GZ, Zhou X-J, Massry SG. Mechanisms of Impaired Insulin Secretion after Chronic Excess of Parathyroid Hormone. American Journal of Physiology 1990; 59: F210-F216.
- Ni Z, Smogorzewski M, Massry SG. Elevated Cytosolic Calcium of Adipocytes in Chronic Renal Failure. Kidney International 1995; 47: 1624-1629.
- 52) Haag-Weber M, Hörl WH. Effect of Calcium Channel Blockers on Intracellular Calcium Accumulation. Nephrology, Dialysis, Transplantation Supplement 1994; 3: 24-27.
- 53) Levi E, Fadda GZ, Thanakitcharu P, Massry SG. Chronology of Cellular Events Leading to Derangements in Function of Pancreatic Islets in Chronic Renal Failure. Journal of the American Society of Nephrology 1992; 3: 1139-1146.
- 54) Bro S, Olgaard K. Effects of Excess PTH on Nonclassical Target Organs. American Journal of Kidney Diseases 1997Nov; 30(5): 606-620.
- 55) Shurtz-Swirski R, Shkolnik T, Shasha SM. Parathyroid Hormone and the Cellular Immune System. Nephron 1995; 70: 21-24.



- Ureña P, Kubrusly M, Mannstadt M, Hruby M, Tringh Trang Tan M-M, Silve C, et al. The Renal PTH/PTHrP Receptor is Down-Regulated in Rats with Chronic Renal Failure. Kidney International 1994; 45: 605-611.
- 57) Stojceva-Taneva O, Fadda GZ, Smogorzewski M, Massry SG. Parathyroid Hormone Increases Cytosolic Calcium of Thymocytes. Nephron 1993; 64: 592-599.
- 58) Luciak M, Trznadel K. Free Oxygen Species Metabolism during Hemodialysis with Different Membranes. Nephrology, Dialysis, Transplantation Supplement 1991; 3: 66-70.
- 59) Hänsch GM, Karnaoukhova S, Chang SH, Rus H, Nicolescu F, Deppisch R, et al. Activation of Human Neutrophils after Contact with Cellulose-Based Haemodialysis Membranes: Intracellular Calcium Signalling in Single Cells. Nephrology, Dialysis, Transplantation 1996; 11: 2453-2460.
- 60) Haag-Weber M, Mai B, Deppisch R, Göhl H, Hörl WH. Studies of Biocompatibility of Different Dialyzer Membranes: Role of Complement System, Intracellular Calcium and Inositol-Trisphosphate. Clinical Nephrology 1994; 41(4): 245-251.
- Buoncristiani U, Galli F, Rovidati S, Albertini MC, Campus G, Canestrari F. Oxidative Damage during Hemodialysis Using A Vitamin-E-Modified Dialysis Membrane: A Preliminary Characterization. Nephron 1997; 77: 57-61.
- 62) Haag-Weber M, Schollmeyer P, Hörl WH. Granulocyte Activation during Haemodialysis in the Absence of Complement Activation: Inhibition by Calcium Channel Blockers. European Journal of Clinical Investigation 1988; 18: 380-385.
- 63) Haag-Weber M, Mai B, Hörl WH. Effect of Hemodialysis on Intracellular Calcium in Human Polymorphonuclear Neutrophils. Mineral and Electrolyte Metabolism 1992; 18: 151-155.



- Porter CJ, Burden RP, Morgan AG, Daniels I, Fletcher J. Impaired Bacterial Killing and Hydrogen Peroxide Production by Polymorphonuclear Neutrophils in End-Stage Renal Failure. Nephron 1997; 77: 479-481.
- 65) Taccone-Gallucci M, Lubrano R, Clerico A, Meloni C, Morosetti M, Meschini L, et al. Administration of GSH has no Influence on the RBC Membrane: Oxidative Damage to Patients on Hemodialysis. ASAIO Journal 1992; 38: 855-857.
- Taccone-Gallucci M, Lubrano R, Trapasso E, Clerico A, Latorre P, Meloni C, et al. Oxidative Damage to RBC Membranes and Pentose Phosphate Shunt Activity in Hemodialysis Patients after Suspension of Erythropoietin Treatment. ASAIO Journal 1994; 40: M663-M666.
- 67) Richard MJ, Ducros V, Forêt M, et al. Reversal of Selenium and Zinc Deficiencies in Chronic Hemodialysis Patients by Intravenous Sodium Selenite and Zinc Gluconate Supplementation. Biological Trace Element Research 1993; 39: 149-159.
- 68) Lin TH, Chen JG, Liaw JM, Juang JG. Trace Elements and Lipid Peroxidation in Uremic Patients on Hemodialysis. Biological Trace Element Research 1996; 51: 277-283.
- 69) Richard MJ, Arnaud J, Jurkovitz C, Hachache T, Meftahi H, Laporte F, et al. Trace Elements and Lipid Peroxidation Abnormalities in Patients with Chronic Renal Failure. Nephron 1991; 57: 10-15.
- 70) Nagase S, Aoyagi K, Hirayama A, Gotoh M, Ueda A, Tomida C, et al. Decreased Serum Antioxidant Activity of Hemodialysis Patients Demonstrated by Methylguanidine Synthesis and Microsomal Lipid Peroxidation. Nephron 1996; 74: 555-560.
- 71) Davis FB, Davis PJ, Blas SD, Schoenl M. Action of Long-Chain Fatty Acids In Vitro on Ca²⁺-Stimutable, Mg²⁺-Dependent ATPase



- Activity in Human Red Cell Membranes. Biochemical Journal 1987; 248: 511-516.
- 72) Taffet GE, Pham TT, Bick DLM, Entman ML, Pownall HJ, Bick RJ. The Calcium Uptake of the Rat Heart Sarcoplasmic Reticulum is altered by Dietary Lipid. The Journal of Membrane Biology 1993; 131: 35-42.
- 73) Starling AP, East JM, Lee AG. Effects of Phospholipid Fatty Acyl Chain Length on Phosphorylation and Dephosphorylation of the Ca²⁺-ATPase. Biochemical Journal 1995; 310: 875-879.
- 74) Shalev O. Decreased Erythrocyte (Ca²⁺ + Mg²⁺)-ATPase Activity in Hemodialyzed Uremic Patients. Renal Failure 1991; 13(1):27-30.
- 75) Takahashi H, Yamaguchi M. Activating Effect of Regucalcin on (Ca²⁺-Mg²⁺)-ATPase in Rat Liver Plasma Membranes: Relation to Sulfhydryl Group. Molecular and Cellular Biochemistry 1994; 136: 71-76.
- 76) Lindner A, Vanholder R, De Smet R. HPLC Fractions of Human Uremic Plasma Inhibit the RBC Membrane Calcium Pump. Kidney International 1997; 51: 1042-1052.
- 77) Zidek W, Rustemeyer T, Schlüter W, Karas M, Kisters K, Graefe U. Isolation of an Ultrafilterable Ca²⁺-ATPase Inhibitor from the Plasma of Uraemic Patients. Clinical Science 1992; 82: 659-665.
- 78) Briggs WA, Sillìx DH, Mahajan S, McDonald F. Leukocyte Metabolism and Function in Uremia. Kidney International 1983; 24(Suppl 16): S93-S96.
- 79) Islam A, Smogorzewski M, Massry SG. Effect of Chronic Renal Failure and Parathyroid Hormone on Phospholipid Content of Brain Synaptosomes. American Journal of Physiology 1989; 256: F705-F710.



- 80) Loh HH, Law PY. The Role of Membrane Lipids in Receptor Mechanisms. Annual Reviews of Pharmacology and Toxicology 1980; 20: 201-234.
- Okamoto H, Kawaguchi H, Sano H, Kageyama K, Kudo T, Koyama T, et al. Microdynamics of the Phospholipid Bilayer in Cardiomyopathic Hamster Heart Cell Membrane. Journal of Molecular and Cellular Cardiology 1994; 26: 211-218.
- 82) Gavrilova NJ, Petkova DH. Role of Rat Liver Plasma Membrane Phospholipids in Regulation of Protein Kinase Activities. Journal of Lipid Mediators in Cell Signalling 1995; 11: 241-252.
- 83) Liu M-S, Wu L-L. Heart Sarcolemmal Ca²⁺ Transport in Endotoxin Shock: II. Mechanism of impairment in ATP-dependent Ca²⁺ Transport. Molecular and Cellular Biochemistry 1992; 112: 135-142.
- 84) Tu YP, Xu H, Yang FY. Transmembrane Ca²⁺ Gradient-Mediated Change of Fluidity in the Inner Layer of Phospholipids Modulates Ca²⁺-ATPase of Sarcoplasmic Reticulum. Biochemistry and Molecular Biology International 1994 Jun; 33(3): 597-605.
- 85) De los Reyes B, Perez-García R, Liras A, Arenas J. Reduced Carnitine Palmitoyl Transferase Activity and Altered Acyl-Trafficking in Red Blood Cells from Hemodialysis Patients. Biochimica et Biophysica Acta 1996, 1315: 37-39.