

CHARACTERIZATION OF COMPOUNDS FROM CURTISIA DENTATA (CORNACEAE) ACTIVE AGAINST CANDIDA ALBICANS

LESHWENI JEREMIA SHAI

THESIS SUBMITTED TO THE DEAPRTMENT OF PARACLINICAL SCIENCES, FACULTY OF VETERINARY SCIENCES, UNIVERSITY OF PRETORIA, FOR THE FULFILMENT OF THE DEGREE DOCTOR OF PHILOSOPHY

SUPERVISOR: DR. LYNDY J. MCGAW (PhD)

CO-SUPERVISOR: PROF. JACOBUS N. ELOFF (PhD)

OCTOBER 2007

DECLARATION

I declare that the thesis hereby submitted to the University of Pretoria for the degree Philosophiae Doctor has not previously been submitted by me for a degree at this or any other university, that it is my own work in design and in execution, and that all material contained herein has been duly acknowledged.

Mr. L.J. Shai				
Prof. J.N. Eloff (Promoter)				
Dr. L.J. McGaw (Co-promoter)				



ACKNOWLEDGEMENTS

'Moreko ga itekole, ebile motho ke motho ka bangwe batho (Sepedi proverb). In short, no man is an island'. THIS PROJECT IS DEDICATED TO MY FAMILY (My wife Grace, my daughters Pontsho and Bonnie, I love you. This is for you).

I want to thank the following:

- GOD ALMIGHTY for love and life he has shown throughout my life. Without HIS plants, HIS chemicals, HIS gift of love and HIS gift of life this work would not have been a reality.
- 2. Dr. L.J. McGaw (I called you Lyndy), for the expert supervision and guidance throughout the project. Your understanding and most importantly, your friendship enabled successful completion of this research project.
- 3. Prof. J.N. Eloff for expertise and knowledge in Phytochemistry and Biochemistry without which this work would not have been possible.
- 4. Dr. P. Masoko for assistance with antifungal work and encouragement to carry on working hard.
- 5. Dr. E. Bizimenyera for help with anthelminthic activity assays. I enjoyed the laughs we had whilst working.
- 6. Drs. L. Mdee and M. Aderogba, your expertise and willingness to assist with structure elucidation at no charge have been amazing. I thank GOD for you.
- 7. My colleagues in the Phytomedicine Program (Thomas, Tsholofelo, Bellonah, Victor, Phineas, Havanna, Elsie, Moraba, Snow, Theresa, Jerry Angeh and his wife Irene and those I forgot to mention), the contributions you have made are truly not insignificant in my eyes.
- 8. My mother, Caroline Malema and my uncle, Thiniel stood by me even though they thought there was no need for more studying.
- 9. My in-laws, Mpho, Ntebe, Mampe, Mme, ausi Maruti and all the uncles and cousins for giving a new meaning to the word 'family'.
- 10. My colleagues at University of Limpopo, Medunsa campus for being good role models.
- My colleagues at the National Department of Health's Medicines Control Council for making me realise I needed to study hard not to fail.

LIST OF ABBREVIATIONS

Af Aspergillus fumigatus

AMB Amphotericin B

BEA Benzene, ethyl acetate, ammonia (90:10:1)

Ca Candida albicans

ACN acetone

CD Curtisia dentata

CEF Chloroform: ethyl acetate: formic acid (5:4:1)

Cn Cryptococcus neoformans

CZ Cussonia zuluensis
DCM dichloromethane

DE Dichloromethane: ethyl acetate (4:1)

DEPT Distortionless enhancement by polarization transfer

DMSO Dimethylsulfoxide
Ec Escherichia coli

Ef Enterococcus faecalis

EMW Ethyl acetate: methanol: water (40:5.4:4)

IPUF Indigenous Plant Use Forum

KA Kigelia africana
Mc Microsporum canis

MIC Minimal inhibitory concentration

MS Mass spectrometry

NMR Nuclear magnetic resonance
Pa Pseudomonas aeruginosa

Retardation factor

Sa Staphylococcus aureus
Ss Sporothrix schenckii

TE Trichilia emetica

TLC Thin layer chromatography
TP Terminalia phanerophlebia

TS Terminalia sambesiaca

UPPP University of Pretoria's Phytomedicine Programme

UV Ultraviolet VR Vepris reflexa



PAPERS PREPARED FROM THIS THESIS

LJ Shai, LJ McGaw, P Masoko and JN Eloff. Evaluation of seven South African plant species with activity against *Candida albicans* (Manuscript).

LJ Shai, LJ McGaw, MA Aderogba, LK Mdee and JN Eloff, Antimicrobial Activity of Four Pentacyclic Triterpenoids from *Curtisia dentata* (Manuscript)

L.J. Shai, E.S. Bizimenyera, L.J. McGaw and J.N. Eloff Lupeol, ursolic acid, betulinic acid and extracts of *Curtisia dentata* inhibit motility of *Trichostrongylus colubriformis, Haemonchus contortus* and *Caenorhabditis elegans* (Manuscript)

L.J. Shai, L.J. McGaw, J.N. Eloff. Extracts of the leaves and twigs of *Curtisia dentata* are more active against *Candida albicans* than the stem bark extract (Manuscript)

CONFERENCE PRESENTATIONS

2005

Paper: L.J. Shai, L.J. McGaw, M. Aderogba and J.N. Eloff. Anti-*Candida* activity of *Curtisia dentata* extracts. **Indigenous Plant Use Forum (IPUF), Rhodes University, Grahamstown (South Africa).**

2006

Paper: L.J. Shai, L.J. McGaw, L.K. Mdee, M. Aderogba and J.N. Eloff. Antifungal constituents from *Curtisia dentata* leaves. **Indigenous Plant Use forum (IPUF)**, **University of Botswana, Gaborone (Botswana)**.

Poster: L.J. Shai, L.J. McGaw, L.K. Mdee, M. Aderogba and J.N. Eloff. Antibacterial triterpenes isolated from *Curtisia dentata*. **27**th **African Health Sciences Congress (AHSC), Durban, South Africa.**

Poster: L.J. Shai., E. Bizimenyera, L.J. McGaw and J.N. Eloff. *Curtisia dentata* extracts, betulinic acid, lupeol and ursolic acid have anthelminthic activity against



Trichostrongylus colubriformis and Hemonchus contortus in vitro. . 27th African Health Sciences Congress (AHSC), Durban, South Africa.

2007

Paper: Shai L.J., McGaw L.J., Picard J. and Eloff J.N. *In vivo* wound healing activity of *Curtisia dentata* extracts and isolated compounds. **Indigenous Plant Use forum** (IPUF), University of Johannesburg, South Africa.



TABLE OF CONTENTS

DECLARATION	i
ACKNOWLEDGEMENTS	ii
LIST OF ABBREVIATIONS	iii
PAPERS PREPARED FROM THIS THESIS	iv
CONFERENCES	lv
ABSTRACT	xvi
CHAPTER 1	1
INTRODUCTION	1
1.1. THE IMPACT OF INFECTIOUS DISEASES	1
1.2. CANDIDOSIS	2
1.2.1. Predisposing factors to candidosis	3
1.2.2. Immunodeficiency and candidosis	3
1.2.3. Pathogenesis	4
1.2.3.1. Adhesion	4
1.2.3.2. Epithelium invasion	4
1.2.4. Treatment	5
1.2.4.1. Polyenes	6
1.2.4.2. 5-Flucytosine	7
1.2.4.3. Azoles	7
1.3. MEDICINAL PLANTS	8
1.3.1. Overview	8
1.3.2. Isolation of Compounds from Medicinal Plant Species	11
1.3.3. Some compounds isolated from plants	13
1.3.3.1. Phenolic compounds	13
1.3.3.2. Terpenoids	13
1.3.3.3. Essential oils	14
1.3.4. Ethnobotany and Drug Discovery	14
1.3.5. Determination of Biological Activities of Medicinal Plants	15
1.4. PLANTS USED IN THE STUDY	17
1.4.1. Curtisia dentata (Burm. f) C.A. Sm	18
1.4.2. Trichilia emetica Vahl	19
1.4.3. Kigelia africana (Lam.) Benth	21
1.4.4. Cussonia zuluensis Strey	22
1.4.5. Terminalia phanerophlebia Engl. & Diels	23



1.4.6.	Terminalia sambesiaca Engl. & Diels	24
1.4.7.	Vepris reflexa I. Verd	25
1.5.	SUMMARY AND PROBLEM STATEMENTS	26
1.6.	HYPOTHESIS	27
1.7.	AIM OF THE STUDY	27
1.8.	OBJECTIVES OF THE STUDY	27
СНАР	PTER 2	29
GENE	RAL MATERIALS AND METHODS	
2.1.	REVIEW OF GENERAL METHODS	29
2.1.1.	Selection of plant species	29
2.1.2.	Extraction	29
2.2.	PROCESSING OF PLANT MATERIAL	30
2.2.1.	Plant Collection and storage	30
2.2.2.	Preliminary extraction for screening	30
2.2.3.	Serial Exhaustive Extraction	30
2.2.4.	Thin Layer Chromatography (TLC)	31
2.3.	BIOLOGICAL ACTIVITY METHODS	31
2.3.1.	Fungal cultures	31
2.3.2.	Bacterial cultures	32
2.3.3.	Bioautography Procedure	32
2.3.4.	Minimal Inhibitory Concentration Determination	32
2.3.5.	Cytotoxicity	33
2.4.	BIOASSAY-GUIDED FRACTIONATION	34
2.5.	STRUCTURE ELUCIDATION	34
СНАР	PTER 3	
PREL	IMINARY SCREENING	36
3.1.	MATERIALS AND METHODS	37
3.1.1.	Extraction	37
3.1.2.	Test Organisms	38
3.1.3.	Bioautography of extracts	38
3.1.4.	Minimal Inhibitory Concentration Determination	38
3.1.5.	Cytotoxicity and selectivity index	38
3.2.	RESULTS	39
3.2.1.	Mass extracted from leaves	39
3.2.2.	Antifungal activity of extracts of the leaves	39



3.2.2.1	. Bioautography	39
3.2.2.2	2. Minimal Inhibitory Concentrations against fungi	41
3.2.3.	Antibacterial activity of extracts of the leaves	49
3.2.3.1	. Bioautography	49
3.2.3.2	2. MIC values against bacteria	50
3.2.4.	Cytotoxicity of acetone and dichloromethane extracts of <i>C. dentata</i>	54
3.3.	Comparison of antifungal and antibacterial activity of stem bark and leave	es of
	Curtisia dentata	56
3.3.1.	TLC fingerprints	56
3.3.2.	Comparative bioautography	56
3.3.3.	Minimal inhibitory concentrations	58
3.4.	DISCUSSION	59
3.5.	CONCLUSIONS	62
CHAP	TER 4	
ISOLA	TION OF ACTIVE COMPOUNDS FROM <i>C. dentata</i> (Burm.f) C.A. Sm.	64
4.1.	INTRODUCTION	64
4.1.1.	Compounds isolated from Cornaceae family	64
4.2.	MATERIALS AND METHODS	65
4.2.1.	General Methods	65
4.2.1.1	. Plant Part Collection	65
4.2.1.2	Serial exhaustive fractionation	65
4.2.2.	Overview of approach followed	65
4.2.2.1	. Selection of stationary phase	65
4.2.2.2	2. Group separation (column I)	66
4.2.2.3	Combination of fractions from column I (DCM extract)	67
4.2.2.4	Isolation of compound CI (column II)	67
4.2.2.5	i. Isolation of compound CII and CIII (column III)	68
4.2.2.6	i. Isolation of compound CIV (column IV)	68
4.2.2.7	. Isolation of compound HI (column V)	68
4.3.	RESULTS	69
4.3.1.	Serial exhaustive extraction	69
4.3.2.	Activity of serial exhaustive extraction samples	70
4.3.2.1	. Anti-Candida activity	70
4.3.2.2	2. Antibacterial activity	71
4.3.3.	Isolation of compounds	72
4.4.	DISCUSSION	78



4.4.1.	Serial Exhaustive Extraction of <i>C. dentata</i> Leaves	78
4.4.2.	Isolation of compounds	79
4.5.	CONCLUSIONS	79
CHAP	TER 5	
STRU	CTURE ELUCIDATION	80
5.1.	INTRODUCTION	80
5.2.	MATERIALS AND METHODS	80
5.2.1.	Thin layer chromatography	81
5.2.2.	¹³ C and ¹ H NMR spectroscopic analysis	81
5.2.3.	Mass Spectrometry	81
5.3.	RESULTS AND DISCUSSION	81
5.3.1.	Thin Layer Chromatographic Analysis of Isolated Compounds	81
5.3.2.	Characterization of compound CI	82
5.3.3.	Characterization of compound CII	83
5.3.4.	Compound CIII	84
5.3.5.	Compound CIV	85
5.3.6.	Estimation of betulinic acid content of C. dentata leaves	87
CHAP	TER 6	
ANTIN	MICROBIAL ACTIVITY OF ISOLATED COMPOUNDS	93
6.1.	INTRODUCTION	93
6.2.	MATERIALS AND METHODS	94
6.2.1.	Bioautography	94
6.2.2.	Minimal Inhibitory Concentration (MIC) Determination	95
6.2.3.	Cytotoxicity	95
6.3.	RESULTS	95
6.3.1.	Antifungal activity	95
6.3.2.	Antibacterial Activity	97
6.3.3.	Cytotoxicity	98
6.4.	DISCUSSION	103
6.4.1.	Lupeol and betulinic acid	102
6.4.2.	Ursolic acid and Hydroxyursolic acid	104
6.4.3.	Cytotoxicity	104
6.5.	CONCLUSIONS	105



CHAPTER 7

8.3. RESULTS

0 11/\(\)		
IN VIV	O ANTIFUNGAL AND WOUND HEALING ACTIVITY OF CURTISIA DEN	ATATI
EXTR	ACTS, LUPEOL AND BETULINIC ACID	
7.1.	BACKGROUND	107
7.2.	MATERIALS AND METHODS	109
7.2.1.	Rats	110
7.2.2.	Housing and feeding conditions	110
7.2.3.	Preparation of animals	110
7.2.4.	Wound creation	110
7.2.5.	Infection with C. albicans	111
7.2.6.	Preparation of extracts	111
7.2.7.	Observations	112
7.2.8.	Evaluation of lesions	117
7.2.9.	Recording of data	117
7.2.10	. Statistical analysis	117
7.2.11	. Pathological and histopathological studies	117
7.3.	RESULTS	118
7.4.	DISCUSSION AND CONCLUSIONS	132
7.4.1.	Weight loss and temperature	132
7.4.2.	Lesion sizes	133
7.4.3.	Exudate and erythema	133
7.4.4.	Presence of infection on skin and internal organs	134
СНАР	TER 8	
ANTIH	ELMINTIC ACTIVITY OF CURTISIA DENTATA EXTRACTS, LUPEOL,	
URSO	LIC ACID AND BETULINIC ACID	
8.1.	INTRODUCTION	135
8.2.	MATERIALS AND METHODS	136
8.2.1.	Plant extracts and compounds	136
8.2.2.	Anthelminthic activity	136
8.2.2.1	. Recovery and preparation of eggs	136
8.2.2.2	2. Egg hatch inhibition assay	137
8.2.2.3	Anthelminthic activity against L3 stage larvae	137
8.2.2.4	Anthelminthic activity against developing larvae	137
8.2.2.5	5. C. elegans	138
8.2.2.6	6. Calculation of LC ₅₀ values	138

139



8.3.1.	Egg hatching	139
8.3.2.	Inhibition of motility of adult parasitic nematodes	141
8.3.3.	Inhibition of motility of free-living nematodes	145
8.4.	DISCUSSION AND COCLUSION	148
CHAP	TER 9	
GENE	RAL DISCUSSIONS AND CONCLUSIONS	
9.1.	ANTIFUNGAL AND ANTIBACTERIAL ACTIVITY OF SELECTED PLAN	١T
	SPECIES	150
9.2. SELECTION OF THE MOST PROMISING TREE SPECIES FOR ISOLATION		ATION
	OF ACTIVE COMPOUNDS	151
9.3.	ISOLATED COMPOUNDS	151
9.4.	ANTHELMINTHIC ACTIVITY	152
9.5.	CYTOTOXICITY	153
9.6.	IN VIVO STUDIES	153
9.7.	CONCLUSIONS	154
CHAP	TER 10	156
REFE	RENCE	156
APPE	NDIX	177



LIST OF FIGURES

Figure 1.1:	Structures of some of the antibiotics used to treat fungal infections in	
	humans and animals.	6
Figure 1.3:	The leaves and stem of Curtisia dentata (photographed at the National	
	Botanical Garden, Mpumalanga, South Africa in June 2007)	19
Figure 1.4:	Trichilia emetica flowers, leaves and fruits (van Wyk et al., 1997).	21
Figure 1.5:	Kigelia africana tree, flowers and fruit (van Wyk et al., 1997).	22
Figure 1.6:	Terminalia phanerophlebia leaves. Picture taken at the Lowveld National	
	Botanical Garden in Nelspruit, Mpumalanga, S Africa, in October 2006.	24
Figure 2.1:	Flow diagram illustrating the layout of the methods and order of events in	the
	study	35
Figure 3.1:	Bar charts showing the mass extracted (mg/g of dry material) from leaves	s of
	different plant species.	40
Figure 3.2:	TLC plates sprayed with vanillin-sulphuric acid (A) and bioautograms sho	wing
	growth inhibitory activity (B) of acetone extracts of seven selected plant	
	species against Candida albicans.	42
Figure 3.3:	TLC fingerprints (A) and anti-Candida activity (B) of DCM extracts of seven	en
	different plant species.	43
Figure 3.4:	TLC bioautograms showing antifungal activity of acetone extracts of leav	es of
	different species against C. neoformans (CN) and S. schenckii (Ss).	44
Figure 3.5:	Combined average MIC values of each plant extract against all the funga	ıl
	species.	47
Figure 3.6:	Combined average total activity values of each plant extract against all the	ne
	fungal species.	47
Figure 3.7:	Sensitivity of fungal test organisms to all plant extracts (MIC values).	48
Figure 3.8:	Sensitivity of fungal test organisms to all plant extracts (total activity value	es).
		48
Figure 3.9:	TLC bioautograms showing antibacterial activity against four test bacteria	al
	suspensions.	49
Figure 3.10:	Combined average MIC values of each plant extract against all the bacte	rial
	species.	52
Figure 3.11:	Combined average total activity values of each plant extract against all the	ie
	bacterial species.	53
Figure 3.12:	Sensitivity of bacterial test organisms to all plant extracts.	53
Figure 3.13:	Sensitivity of bacterial test organisms to all plant extracts (total activity	
	values).	54



Figure 3.14:	Cytotoxicity and acetone extract $LC_{50} = 0.02440967$ mg/ml	55
Figure 3.15:	Cytotoxicity of the DCM extract ($LC_{50} = 0.0066110474 \text{ mg/ml}$)	55
Figure 3.16:	Comparison of the chemical components present in leaves (L) and stem	bark
	(S) of Curtisia dentata.	57
Figure 3.17:	Comparison of the antibacterial chemical components present in leaves	(L)
	and stem bark (S) of Curtisia dentata.	57
Isolation Sch	eme: Schematic diagram showing the isolation of active compounds from	om
	Curtisia dentata leaves using column chromatography with silica gel 60 a	as
	the stationary phase.	66
Figure 4.1:	Amount extracted from Curtisia dentata leaves (837 g) using solvents of	
	varying polarities.	69
Figure 4.2:	TLC plates showing growth inhibition activity of Curtisia dentata leaf extra	acts
	against Candida albicans.	70
Figure 4.3:	TLC bioautograms (BEA as eluent) showing activity of hezane (H),	
	dichloromethane (D), acetone (AC) and methanol (M) extracts of Curtisia	3
	dentata against P. aeruginosa, E. coli, Enterococcus faecalis and S. auro	eus.
		72
Figure 4.4:	TLC chromatograms of serially extracted C. dentata leaves developed in	
	EMW, CEF and BEA.	72
Figure 4.5:	Fractions collected from separation of dichloromethane extract of Curtisi	а
	dentata leaves (fraction 1-22).	73
Figure 4.6:	Fractions collected from separation of dichloromethane extract of C. den	tata
	leaves (fraction 1-24 [A] and 23-33 [B]).	74
Figure 4.7:	TLC bioautograms (A) (CEF) showing activity of fractions A-F against C.	
	albicans, and vanillin-sprayed TLC plates developed in CEF (B) and BEA	4 (C)
		75
Figure 4.8:	TLC chromatograms showing compound CI analysed side-by-side with	
	fraction B from column I.	76
Figure 4.9:	TLC chromatogram showing separation of CIII from CII in column III.	76
Figure 4.10:	Fractions 1 – 36 collected from separation of hexane extract of <i>C. denta</i>	ta 77
Figure 5.1:	TLC analysis of the four isolated compounds (CI, CII, CIII and CIV) and	
-	dichloromethane crude (Cr).	82
Figure 5.2:	The structure of CI (lupeol) isolated from the leaves of <i>Curtisia dentata</i>	83
Figure 5.3:	The structure of CII (betulinic acid) isolated from the leaves of <i>Curtisia</i>	
-	dentata	84



Figure 5.4:	The structure of CIII (ursolic acid) isolated from the leaves of C.dentata	
		86
Figure 5.5:	Retro-Diels-Alders fragmentation patterns of ursolic acid (CIII) isolated from	m the
	leaves of C. dentata	86
Figure 5.6:	The structure of CIV isolated from the leaves of C. dentata	87
Figure 5.7:	Estimation of the betulinic acid content in dichloromethane and acetone	
	extracts of C. dentata leaves.	88
Figure 5.8:	Estimation of betulinic acid content in the acetone and methanol extracts	of
	Curtisia dentata leaves.	88
Figure 6.1:	TLC bioautograms showing activity of three isolated compounds against	
	Candida albicans	96
Figure 6.2:	Average MIC values of each compound against all the fungal test organis	ms.
		98
Figure 6.3:	Sensitivity of each tested fungal species to all the isolated compounds.	99
Figure 6.4:	Antibacterial activity of isolated compounds against three bacterial test	
	organisms.	100
Figure 6.5:	Averages of MIC values of each compound against all bacterial test organ	nisms.
		101
Figure 6.6:	Average MIC values against each bacterial test organisms by all the isola	ted
	compounds.	101
Figure 6.7:	Cytotoxicity of lupeol (LC ₅₀ = 89.4688722 ug/ml) against Vero cells	102
Figure 6.8:	Cytotoxicity Betulinic acid ($LC_{50} = 10.93846914 \text{ ug/ml}$) against Vero cells	102
Figure 7.1:	A diagrammatic representation of wound positions on the back of each ra	t.113
Figure 7.2:	The diagrammatic representation of surgical wound creation and dressing	g of
	excisional wounds on rats.	114
Figure 7.3.	Representation of changing of wound dressing and treatment with varying	3
	products.	115
Figure 7.4:	Preparation of tissue specimens for histopathological examinations.	116
Figure 7.5:	Average weights of the rats on different treatment days.	121
Figure 7.6.	Average temperature of rats up until 11 days of treatment.	122
Figure 7.7:	Average sizes of lesions treated with different extracts of <i>C. dentata</i>	123
Figure 7.8:	Average sizes of lesions treated with different compounds isolated from C	
	dentata.	124
Figure 7.9:	Exudate release from lesions treated with different <i>C. dentata</i> -derived	
	compounds	126



Figure 7.10:	Exudate release from lesions treated with different C. dentata-derived ex	xtracts
	(acetone and water extracts).	126
Figure 7.11:	Erythema (arbitrary values) in wounds treated with compounds isolated	from C.
	dentata:	127
Figure 7.12:	Erythema (arbitrary values) in wounds treated with extracts (acetone ar	nd water
	extracts) derived from C. dentata:	128
Figure 7.13:	Crust formation in wounds treated with various concentrations of com-	pounds
	during specified days of treatment.	129
Figure 7.14:	Crust formation in wounds treated with various concentrations of com-	pounds
	during specified days of treatment.	130
Figure 7.15:	Histopathology for granulomatous dermatitis.	132
Figure 8.1:	An example of a graph drawn to estimate the LC ₅₀ values of compound	nds and
	bulk fractions ahainst parasitic and free-living nematodes.	139
Figure 8.2:	Percentage of dead Trichostrongylus colubriformis larvae after 48 hours	of
	incubation in the presence of varying concentrations of the acetone and	
	dichloromethane extracts of C. dentata leaves.	140
Figure 8.3:	Percentage of dead Hemonchus contortus larvae after 48 hours of incub	ation
	in the presence of varying concentrations of the acetone and dichlorome	ethane
	extracts of C. dentata leaves.	140
Figure 8.4:	Percentage of dead T. colubriformis larvae after 48 hours of incubation in	n the
	presence of varying concentrations of the lupeol and betulinic acid.	141
Figure 8.5:	Percentage of dead H. contortus larvae after 48 hours of incubation in t	he
	presence of varying concentrations of the lupeol and betulinic acid.	143
Figure 8.6:	Percentage of motile larvae (mixture of H. contortus and T. colubriformis	s)
	treated with isolated compounds after five days of incubation.	144
Figure 8.7:	Percentage of motile larvae (mixture of H. contortus and T. colubriformis	s)
	treated with acetone and dichloromethane extracts of C. dentata after five	e days
	of incubation.	144
Figure 8.8:	Inhibition of motility of free-living nematode, C. elegans with compounds	;
	isolated from Curtisia dentata leaves.	145
Figure 8.9:	Inhibition of motility of free-living nematode, C. elegans with acetone and	d
	dichloromethane of Curtisia dentata leaves.	146
Figure 8.10:	Inhibition of motility of free-living nematode, C. elegans with isolated	
	compounds from leaves of Curtisia dentata.	147
Figure 8.11:	Inhibition of motility of free-living nematode, C. elegans with the acetone	and
	dichloromethane extracts of the leaves of Curtisia dentata.	147



ABSTRACT

The main aim of the study was to isolate compounds active against *Candida albicans* from the most active species from a pool of several trees. Seven tree species with good antifungal activity were selected from the Phytomedicine Programme database. The selected plant species investigated were screened for growth inhibitory activity against *Candida albicans* using bioautography and serial microplate dilution methods. These tree species were: *Cussonia zuluensis*, *Vepris reflexa*, *Curtisia dentata*, *Trichilia emetica*, *Terminalia phanerophlebia*, *Terminalia sambesiaca* and *Kigelia africana*. Using the serial microplate dilution method for the determination of minimal inhibitory concentrations, *Terminalia phanerophlebia* and *T. sambesiaca* were active against *Candida albicans* with MIC values as low 0.02 mg/ml. The acetone and dichloromethane extracts of all plant leaves were active against *C. albicans* with MICs varying from 0.02-2.5 mg/ml. Based on bioautography, the acetone extract of the leaves of *Curtisia dentata* had more active (5) compounds against *C. albicans* than any of the tree species investigated.

The dichloromethane, acetone and hexane extracts of the seven tree species were further screened for antifungal activity using other fungal test organisms. The fungal species used were Aspergillus fumigatus, Microsporum canis, Sporothrix schenckii and Cryptococcus neoformans. Extracts of Curtisia dentata, Terminalia sambesiaca and Terminalia phanerophlebia had the highest activities against these fungal test organisms with minimal inhibitory concentration (MIC) values as low as 0.02 mg/ml. Cussonia zuluensis was the least active with high MIC values (>250 µg/ml in some cases) and the lowest number (1) of active chemical components on bioautograms. The highest number of active compounds (5) against C. albicans on bioautograms was observed in the acetone extracts of *C. dentata*. The plant species were further investigated for presence of antibacterial compounds, using Escherichia coli, Staphylococcus aureus, Enterococcus faecalis and Pseudomonas aeruginosa as test bacterial organisms. Compounds with similar R_f values in the acetone extract of C. dentata were active against both bacterial and fungal test organisms, suggesting that the growth inhibitory activity of C. dentata extracts was non-selective. C. dentata was chosen for isolation of compounds due to 1) the highest number of active compounds on bioautogram against C. albicans, 2) the MIC values (0.12-0.6 mg/ml) against C. albicans.



Acetone extracts of the leaves, stem bark and twigs of *Curtisia dentata* were compared for antibacterial and antifungal activity using the serial microplate dilution and bioautography methods in order to select the plant part to isolate compounds from. The TLC fingerprints of the twigs and leaves were largely similar. A non-polar compound and two medium polarity compounds, present in the leaves and twigs, were missing in the stem bark extract. Bioautography indicated that the leaves contained more antibacterial and antifungal compounds than the stem bark extracts. Extracts of the leaves were 5-fold more active than the stem bark extracts against *Candida albicans*, with total activities of 1072 and 190 ml/g, respectively. Against bacterial test organisms extracts of the leaves, stem bark and twigs resulted in comparable activities. These findings encourage the interchangeable usage of the stem bark, leaves and twigs of this plant, which may lead to sustainable harvesting of the species. This approach may conserve this and other threatened or endangered plant species.

The leaves of *Curtisia dentata* (Cornaceae) were serially extracted with solvents of varying polarities, starting with hexane, then dichloromethane, followed by acetone with methanol completing the fractionation. The dichloromethane (DCM) and acetone bulk fractions of *Curtisia dentata* contained the highest number of active compounds and resulted in low MIC values. The hexane and the methanol bulk fractions were the least active. In the hexane bulk fraction, bioautography revealed the presence of one active compound. The DCM bulk fraction showed cytotoxicity against Vero cells similar to the positive control, berberine with an LC50 value of 10 μ g/ml. The acetone and dichloromethane fractions resulted in total activity values of 3312 and 4240 ml, respectively. However, these fractions were cytotoxic to the Vero cells with LC50 values of 24.4 μ g/ml for acetone fraction and 6.6 μ g/ml for the dichloromethane fraction. The cytotoxicity data may serve to discourage the use of these extracts to treat candidosis. However, preparations of these fractions may be used topically on wounds to combat infections. The application of these extracts on rat wound model did not result in any observable pathologies.

The DCM and acetone bulk fractions each contained 4 compounds active against *Candida albicans*. Only the dichloromethane extract was fractionated as these extracts contained almost similar active compounds. Column chromatography using silica as the stationary phase afforded four compounds from the DCM extract. These compounds were identified using nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry (MS) as lupeol (CI), betulinic acid (CII), ursolic acid (CIII) and

hydroxyl-ursolic acid (**CIV**). These compounds have been isolated from several plant species and have been to be found active against several pathogens including the human immunodeficiency virus (HIV). This is the first report of the isolation of these compounds from *Curtisia dentata*. The antibacterial activity of these compounds have been reported. The anti-*Candida* activity of ursolic oleanolic and ursolic acid has been reported with MIC values exceeding 128 µg/ml (Hiriuchi *et al.*, 2007). However, the anti-*Candida* activity of betulinic acid and lupeol has not been reported.

The four isolated compounds were tested for activity against several fungal (*Candida albicans*, *C. spicata*, *C. guillermondi*, *Aspergillus fumigatus*, *Sporothrix shenckii*, *Cryptococcus neoformans* and *microsporaum canis*) and bacterial (*Escherichia coli*, *Staphylococcus aureus*, *Enterococcus faecalis* and *Pseudomonas aeruginosa*) species. Ursolic acid and hydroxyursolic acid were the most active with MIC values. Hydroxyursolic acid resulted in an MIC value as low as 8 μg/ml against *M. canis*. *A. fumigatus* was the most resistant microorganism while *M. canis* and *S. schenckii* were the most sensitive. *C. albicans* was moderately sensitive to the compounds with MIC values ranging from 16 μg/ml for betulinic acid to over 250 μg/ml for lupeol.

Compounds isolated in sufficient quantities, namely, lupeol and betulinic acid, were investigated for cytotoxicity against Vero cells. It appeared that lupeol was less toxic than betulinic acid, with LC_{50} values of 89.5 and 10.9 μ g/ml, respectively. The cytotoxicity of betulinic acid was comparable to that induced by the positive control, berberine with an LC_{50} of 10 μ g/ml.

Lupeol was the least active of the isolated compounds. Betulinic acid and lupeol, together with the water and acetone extracts were tested in an *in vivo* rat model to determine antifungal and wound healing activities. The rats were immunocompromised prior to the surgical and treatment procedures. Treatments with any of the formulations did not affect wound healing activity. The rate of wound healing was comparable to both the positive (amphotericin B) and negative (cream only) controls. It was however difficult to judge and score antifungal activity. The model developed to evaluate skin infections will have to be improved to allow for testing for anti-*Candida* activity *in vivo*.

Some antifungal compounds, such as azoles, are known to also have anthelminthic activity. The isolated compounds, which had antifungal activity, were tested for anthelminthic activity against both parasitic and free-living nematodes. Furthermore, other publications demonstrated that betulinic acid had anthelminthic activity against



C. elegans. Lupeol, ursolic acid and betulinic acid, together with the DCM and acetone extracts were investigated for anthelminthic activity against both free living and parasitic nematodes. The acetone and dichloromethane extracts were active against all nematodes to concentrations as low as 160 μg/ml. Betulinic acid and lupeol were active against the parasitic nematodes at high concentrations of 1000 and 200 μg/ml. All compounds were active against the free-living Caenorhabditis elegans with concentrations as low as 8 μg/ml. Betulinic acid was less active than lupeol and ursolic acid against C. elegans. The acetone and dichloromethane extracts were also active against C. elegans with a concentration of 0.31 mg/ml resulting in almost 80% inhibition of larval motility. It would appear that the anthelminthic activity against both parasitic and free-living nematodes occurred at high concentrations of the compounds or extracts. Extracts of various medicinal plant species may provide the solutions to illhealth of small ruminants caused by parasitic nematodes in poor communities of southern Africa.

The extracts of *Curtisia dentata* and isolated compounds have anti-*Candida* activity *in vitro*. Their usage is hampered by associated toxicity. The cytotoxicity of the compounds and extracts was only demonstrated with Vero cells (monkey line). Experiments with several human cell lines may indicate the safety of these compound and extracts when used as treatment against *Candida* infections. No toxic effects were noted when extracts and isolated compounds were tested in an animal experiment indicating that extracts may be safe in a topical application. The extract from 1 g of leaf material can be diluted to more than a litre and still inhibit the growth of *C. albicans*.