

The effect of preserving liver tissue in formalin on the concentration of trace minerals in the liver

by

Cornel Smith

BSc. (Agric) (Animal Science) (UP)

Submitted in partial fulfilment of the requirements

for the degree

Magister Scientiae (Agriculturae) in Animal Nutrition

in the

Department of Animal and Wildlife Sciences

Faculty of Natural - and Agricultural Sciences

University of Pretoria Pretoria

Promotor : Prof J.B.J. van Ryssen

November 2002



I declare that this thesis for the degree Magister Scientiae (Agriculturae) at the University of Pretoria has not been submitted by me for a degree at any other university.



Acknowledgements

First, I would like to give thanks to my Heavenly Father. "Commit thy way unto the Lord; trust also in him; and he shall bring it to pass." Ps 37: 5

I wish to thank the following people without whom the successful completion of this study would not have been possible:

Prof J.B.J. van Ryssen, my promoter from the Department of Animal and Wildlife Sciences at the University of Pretoria, for his guidance, interest, help and support which led to the successful completion of this study.

Mr E.B. Spreeth and *Ms G. Smit* from the Department of Animal and Wildlife Science at the University of Pretoria for their assistance and advice with the laboratory procedures.

Mr R.J. Coertze and Prof H. Groeneveld for their assistance and advice concerning the statistical analysis of the data.

My parents, the Lotz's, the Nell's and especially Lisa Riback, Hester Hill and Annelize van der Baan, for all their love, support, understanding and encouragement during my studies.



Table of contents

Ackn	owledments	3
List c	of Figures	7
List c	of Tables	9
List c	of Abbreviations	12
Defin	ition of terms	14
Abstr	act	15
Same	vatting	17
Introd	luction	19
Chap	ter 1: Literature study	21
1.1	Metabolism of trace elements	21
1.1.1	Absorption	21
1.1.2	Distribution of trace elements between tissues an	d organs
		21
1.1.3	Excretion	22
1.1.4	Animal reserves	22
1.2	The three important principals of biochemical	diagnosis:
	relationship with intake, time and function	23
1.2.1	Relationship with trace element intake	23
1.2.2	Relationship to time	25
1.2.3	Relationship to function	25
1.3	Interpretation of biochemical criteria of trace e	element status
		26
1.3.1	Copper	26



1.3.2	Cobalt	27
1.3.3	Selenium	29
1.3.4	Zinc	30
1.3.5	Manganese	30
1.3.6	Biochemical values used to assess the trace element	nutrition of sheep
		32
1.4	Trace element dispersion within a liver	33
1.5	Formalin preservation	39
1.6	Prolonged storage	49
1.7	Use of glass and plastic bottles for storage	55
1.8	Blenders used for homogenisation	57
1.9	Trace element analysis by atomic absorption spec	ctrophotometry
		57
1.10	Moisture content of fresh and formalinised livers	58
1.10.1	Dispersion of moisture within a fresh liver	58
1.10.2	2 Dispersion of moisture within a formalin-preserved live	er
		59
1.11	Evaluation of ethanol based fixatives as a substite	ute for formalin
		60
Chap	ter 2: Materials and methods	61
2.1	Introduction	61
2.2	Experimental material	61
2.3	Preparation	61
2.4	Laboratory analysis	63
2.5	Statistical analysis	66
-	ter 3: Results	67
3.1 M	anganese, Zn, Co, Cu and Se concentrations in live	er samples
		67
	Manganese	68
3.1.2		69
	Cobalt	70
	Selenium	71
3.1.5	Copper	72



the manganese, En, ee, ou and be concentrations in for	maim
	73
3.2.1 Manganese	74
3.2.2 Zinc	75
3.2.3 Selenium	75
3.2.4 Cobalt	76
3.2.5 Copper	76
3.3 Dry matter % in formalinised and fresh liver	77
Chapter 4: Discussion	78
Chapter 5: Conclusion	83
Recommendations	85
Chapter 6: References	86
Chapter 7: Appendix	93

3.2 Manganese, Zn, Co, Cu and Se concentrations in formalin



List of Figures

- Figure 1.2.1Schematic representation of the relationship between direct and
indirect biochemical markers of trace element status in blood or
tissues and the intake of the element at a fixed time23
- Figure 1.2.2Schematic representation of the relationship between direct and
biochemical markers of trace element status in blood or tissues
and the duration of a dietary deficiency24
- Figure 1.4.1 Diagram of the area of the liver from which biopsies were taken33
- Figure 1.4.2 Distribution of copper throughout the liver of an eight week oldpig35
- Figure 1.4.3 Distribution of copper throughout the liver in relation to the positions of the caudal, dorsal and ventral lobes for: new-born lambs (a) and (b), four-tooth ewes (c) and (d), and an aged ewe (e) 35
- Figure 1.4.4 Diagram of a sheep liver showing the sites from which sampleswere taken for analysis38
- Figure 1.6.1 Mean Pb concentrations of frozen and formalin-fixed livers andkidneys of 8 raccoons administered oral Pb acetate53
- Figure 3.1.1 Mean manganese concentration (mg/kg DM) in livers preservedin formalin for different periods of time68



Figure 3.1.2	gure 3.1.2 Mean zinc concentration (mg/kg DM) in livers preserved in		
	formalin for different periods of time	69	
Figure 3.1.3	Mean cobalt concentration (mg/kg DM) in livers preser		
	formalin for different periods of time	70	
Figure 3.1.4	Mean selenium concentration (ng/g DM) in livers prese	erved in	
	formalin for different periods of time	71	
	Moon connection (maller DM) in livers areas	un co d in	
Figure 3.1.5	Mean copper concentration (mg/kg DM) in livers prese		
	formalin for different periods of time	72	
Figure 3.2.1	Mean manganese concentration (mg/l) in formalin after	er one	
	month, three months and six months of preservation o		
	formalin	74	
Figure 3.2.2	Mean zinc concentration (mg/l) in formalin after one m	onth,	
	three months and six months of preservation of liver in	n formalin	
		75	
Figure 3.2.3	Mean selenium concentration (ng/ml) in formalin after	one	
	month, three months and six months of preservation o		
	formalin		
		75	
Figure 3.2.4	Mean cobalt concentration (mg/l) in formalin after one	month	
· · · · · · · · · · · · · · · · · · ·	three months and six months of preservation of liver in		
		76	
Figure 3.2.5	Mean copper concentration (mg/l) in formalin after one		
-	three months and six months of preservation of liver in		
	·		

.....



List of Tables

Table 1.1.1Distribution of Cu, Fe, Mn, Mo, Se, and Zn between various
organs and tissues of a 40 kg sheep with a 3 kg fleece22

 Table 1.3.1 Biochemical values used to assess the trace element nutrition of sheep

32 Table 1.4.1 The deviation of iron values in different parts of the liver

 Table 1.4.2
 Copper concentrations in samples taken at different depths from top surface

Table 1.4.3The mean concentrations of copper, vitamin B_{12} and zinc in sixlivers of sheep

39

 Table 1.5.1
 Comparison of fresh and formalin preserved livers

Table 1.5.2Mean (s.e.) concentrations of selenium in fresh, frozen and
formalin-fixed porcine liver 0 – 28 days after collection41

 Table 1.5.3
 Liver copper analysis on goats with neurological and other diseases

42

34

37

40



Table 1.5.4	Effects of fixation of liver tissue in formalin solution on	weight
	and copper analysis	42
Table 1.5.5	The influence of the storage of liver tissue for 24 h in b	ouffer
	•	44
Table 1 5 6	Influence of storage in different solutions on the iron co	ontent of
		45
	liver (normal) tissue	40
		1 1 - 8
Table 1.5.7	Influence of storage in different solutions on the iron c	
	liver (Secondary Hemochromatosis) tissue	45
Table 1.5.8	Mean Pb concentration in human lung	46
Table 1.5.9	Mean Pb concentration in human vertebra	48
Table 1.6.1	Comparison of mineral levels after six weeks and six r	nonths
		49
Table 1.6.2	The mean results \pm s.d. as founded by Bratton <i>et al.</i> (*	1984)
		50
Table 1.6.3	Zn, Fe and Cu concentrations in frozen and formalin-f	ixed
	kidney and liver	51
		JI.
	Tion of concentrations is former and formalis five	d liver and
1 able 1.6.4	Tissue lead concentrations in frozen and formalin-fixe	
	kidney of eight raccoons administered oral lead aceta	
		53
Table 1.7.1	Trace element content of materials that are frequently	used for
	construction of animal accommodation and for sample	e collection
		55
Table 1.10.1	Dispersion of moisture within a fresh liver	58



Table 1.10.2 Dispersion of moisture within a formalin-preserved liver59Table 1.10.3 Moisture content of livers of various storage periods

Table 2.1Type analysis of 40 % formaldehyde solution (AC-grade)62

- Table 3.1.1Mean (±s.e.) for Mn, Zn, Co, Cu (mg/kg) and Se (ng/g)
concentration in livers after preservation in formalin over
different time periods (DM basis)67
- Table 3.2.1Determination of Se concentration in formalin that was wet
ashed and formalin that was centrifuged from the same sample73
- Table 3.2.2Mean (±s.e.) for manganese, zinc, cobalt, copper (mg/l) and
selenium (ng/ml) concentration in formalin after preservation of
livers over different time periods74
- Table 3.3.1Mean (±s.e.) for dry matter % in fresh and formalinised liver over
different time periods76
- Table 4.1Mineral concentrations in sheep liver (mg/kg DM)79
- Table 7.1Volume of formalin per mass of liver94
- Table 7.2Volume (ml) of formalin before and after the trial98



List of Abbreviations

Cu	copper
Zn	zinc
Mn	manganese
Se	selenium
Со	cobalt
S.e.	standard error
s.d.	standard deviation
mg/kg	milligrams/kg
mg	milligram
kg	kilogram
mg/l	milligrams/litre
I	litre
ng/g	nanograms/kilogram
ng	nanograms
g	gram
ng/ml	nanogram/millilitre

12



۱	il		li	I	i	tı	•	Э
	1	nil	nill	nilli	nillil	nillili	hillilitr	hillilitre

- MMA methylmalonic acid
- AAS atomic absorption spectrophotometry
- GPX glutathione peroxidase
- VLG viral leukoencephalomyelitis of goats



Definition of terms

- Deficient: Levels at which clinical or pathological signs of deficiency should be apparent (Underwood & Suttle, 1999).
- Marginal: Levels at which subclinical effects may prevail, such as reduced immune response, or reduced growth rate (Underwood & Suttle, 1999).
- Adequate: Levels sufficient for optimum functioning of all body mechanisms with a small margin of reserve to counteract commonly encountered antagonistic conditions (Underwood & Suttle, 1999).
- High: Levels well above normal but not necessarily toxic (Underwood & Suttle, 1999).
- Toxic: Levels at which subclinical, clinical or pathological signs of toxicity would be expected to occur (Underwood & Suttle, 1999).
- Normal: Used where deficiencies are unknown, indicates normal background levels (Underwood & Suttle, 1999).

_ ...



Abstract

The effect of preserving liver tissue in formalin on the concentration of trace minerals in the liver

by

Cornel Smith

Supervisor:	Prof J.B.J. van Ryssen
Department:	Animal and Wildlife Sciences,
	Faculty of Nature and Agricultural Sciences,
	University of Pretoria
Degree:	Magister Scientiae (Agriculturae) (Animal Nutrition)

The concentrations of trace minerals (Mn, Cu, Co, Zn and Se) were examined after formalin preservation of 31 sheep and 5 impala livers over differing storing periods (one month, three months and six months). Under field conditions liver samples are often preserved in formalin until micro mineral analysis in the laboratory.

Analyses of trace minerals, expressed on dry basis, were done using atomic absorption spectrophotometry after wet ashing of the liver. After one month of preserving of livers in formalin, the Mn concentration was significantly (P<0.05) lower than the concentration in fresh liver. It was found that at three months of preservation of livers in formalin the Zn and Co concentration were significantly (P<0.05) higher and the Mn and Cu concentration were significantly (P<0.05) lower compared to the concentration in fresh liver.



The Mn, Co and Cu concentrations were significantly (P<0.05) lower after six months of storage in formalin compared to fresh liver. The difference in the concentration between fresh liver and liver stored in formalin was small. It would not have any effect on the interpretation of the relative mineral concentration.

Mineral determinations using atomic absorption spectrophotometry was also done on the formalin in which the liver was preserved. There was a significant (P<0.05) increase in the mineral concentrations from pure formalin to formalin in which liver was stored for all the time periods. This was probably due to leaching of the minerals.

An additional investigation was also done to determine if there was a difference between the dry matter % of fresh (90.73 % DM) and formalinised (92.1% DM) liver. The dry matter % increased significantly (P<0.05) from fresh liver to liver that was preserved in formalin for all the time periods.



Samevatting

Die invloed van die preservering van lewer weefsel in formalien op die konsentrasie van mikro-minerale in die lewer

deur

Cornel Smith

Studieleier:	Prof. J.B.J. van Ryssen
Departement:	Vee- en Wildkunde,
	Fakulteit van Natuur- en landbouwetenskappe,
	Universiteit van Pretoria
Graad:	Magister Scientiae (Agriculturae) (Voedingkunde)

Die invloed van die preservering van 31 skaap en 5 impala lewers in formalien oor verskillende periodes (een maand, drie maande en ses maande) op mikromineraal konsentrasies (Mn, Cu, Co Zn and Se) in die lewer, is ondersoek. Onder veldtoestande word lewer monsters algemeen gepreserveer in formalien tot wanneer dit in die laboratorium geanaliseer kan word.

Die mineraal bepalings, uitgedruk op 'n droë basis, is gedoen met behulp van atoom-absorpsie spektrofotometrie nadat nat verassing op die lewer gedoen is.



Die Mn konsentrasie was betekenisvol (P<0.05) laer in die lewers wat gepreserveer is in formalien vir een maand in vergelyking met die Mn konsentrasie in vars lewers. Na drie maande van preservering van lewers in formalien was die Zn en Co konsentrasies betekenisvol (P<0.05) hoër en die Mn en Cu konsentrasies betekenisvol laer as die konsentrasies in vars lewer. Die Mn, Co en Cu konsentrasies was betekenisvol (P<0.05) laer na ses maande van preservering van lewer in formalien. Die verskil in die konsentrasie tussen vars lewer en lewer wat in formalien gepreserveer is, was klein. Dit sal geen invloed hê op die interpretasie (of die mineraalvlak toksies is en of daar 'n tekort is) van die betrokke mineraal konsentrasie nie.

Mineraal bepalings op die formalien waarin die lewer gepreserveer was, is ook gedoen deur middel van atoom-absorpsie spektrofotometrie. Daar was 'n betekenisvolle (P<0.05) toename in al die minerale konsentrasies vanaf suiwer formalien na formalien waarin lewer gepreserveer is vir al die tydperke. Dit was moontlik a.g.v. loging van die minerale.

'n Verdere ondersoek is ook gedoen om te bepaal of daar 'n verskil was tussen die droë materiaal % van vars lewer (90.37 % DM) en lewer wat gepreserveer was in formalien (92.1 % DM). Die droë materiaal % het betekenisvol toegeneem vanaf vars lewer na lewer wat in formalien gepreserveer is, vir al die tydperke.



Introduction

'Most of the trace minerals can be measured accurately some of the time. Some of the trace minerals can be measured accurately most of the time. Most of the trace minerals are not measured accurately most of the time.' (Mertz, 1987).

The liver has been analysed for trace minerals more often than any other internal organ, mainly since variations in dietary uptake are more readily reflected in the liver, which acts as the main storage organ for some of the minerals (Theron *et al.*, 1973).

It has received special attention as a sample source, because the liver is the body's metabolic centre, and most minerals are integral portions of metalloenzymes, which serve as catalysts for metabolic processes (Boyazoglu, 1976). Hepatic concentrations of trace minerals are commonly used to estimate trace mineral storage pools because dietary intake is rarely available and nutrient interactions affect availability or retention (Thomas *et al.*, 1994).

The liver is the main storage organ for copper and its copper concentration has been found to vary enormously (Widdowson & Dickerson, 1964). Low levels of the copper in the livers of cattle are the result of a primary deficiency, when the diet is inadequate, or a secondary (conditioned) deficiency, when the dietary intake is sufficient, but the utilization of the copper is impeded for example, by the interaction of molybdenum and sulphate (Ehret *et al.*, 1975).

Manganese is not concentrated in any particular organ or tissue. The concentrations in the liver are, however, higher than most other tissues and can be raised or lowered with varying manganese intake. The manganese storage capacity of the liver is limited when compared with the remarkable capacity of this organ to accumulate iron and copper (Widdowson & Dickerson, 1964; Ehret *et al.*, 1975).



The capacity of the animal to store zinc in any of its organs other than bones, is extremely limited so that animals do not normally carry large reserves of zinc. However, as a zinc deficiency develops, there is usually, but not invariable, a small decline in the concentration of zinc in the liver and certain other tissues (Underwood, 1966). High dietary levels of zinc give rise to large increases in bovine liver zinc concentrations (Ehret *et al.*, 1975).

Ehret *et al.* (1975) investigated the effect of formalin storage, over different time periods, on the copper, iron, manganese, zinc and magnesium concentrations in bovine livers. Theron *et al.* (1974) found no statistically significant differences for Cu, Fe, Mn, Zn and Mg concentrations after 22 days of formalin preservation.

The concentrations of Cu, Fe and Mg were also not affected by storage for six months in formalin, but statistically significant differences were detected in the concentrations of manganese and zinc after six months (Theron *et al.*, 1974).

Sullivan *et al.* (1993) found that tissue concentrations of Se and Cu remains unchanged by formalin-fixation and that the analysis of formalin-fixed tissues for diagnostic purposes can be recommended for Se and Cu.

Only a few studies have been done on this subject and under field conditions liver tissues are often preserved in formalin before trace mineral analysis in the laboratory. The aim of this investigation was to determine if the copper, cobalt, manganese, zinc and especially selenium concentrations in fresh and formalinised liver, from the same liver sample, differ significantly and if they are giving the same interpretation.