

Development of clonal propagation protocols for *Uapaca kirkiana* and *Pappea capensis*, two southern African trees with economic potential

By

Simon Alfred Mng'omba

Submitted in partial fulfilment of the requirements for the Doctor of Philosophy degree (Horticulture)

Department of Plant Production and Soil Science in the Faculty of Natural and Agricultural Sciences University of Pretoria Pretoria

Supervisor : Prof. E. S. du Toit

Co-supervisors : Dr F. K. Akinnifesi

: Dr H. M. Venter

June 2007



Declaration

I hereby declare that the thesis I am subm	itting for the Doctor of Philosophy degree
(Horticulture) at the University of Pretoria is	my own work and has not been submitted for
a degree at any other institution.	
Simon Alfred Mng'omba	Date



Acknowledgements

Thanks and Glory to the Almighty God for guiding me through thick and thin. God was always my buckler in this endeavour (Psalms 23: 1-6)

Many people have contributed a lot towards the completion of my studies but my apologies if I do not mention everyone involved. I wish to express my profound gratitude to my supervisors: Prof. E.S. du Toit (major supervisor), Dr F.K. Akinnifesi and Dr H.M. Venter. Their contributions towards completion of my studies were tremendous, remarkable and constructive. Thanks to Dr T. Regnier and Dr W. du Plooy for their valuable contributions on phenol analysis. Prof. C. F. Reinhardt deserves a mention for his timely intervention and encouragement when I was struggling at the beginning of my studies. Thanks to Prof. P. J. Robbertse for his valuable inputs.

Thanks to Annemarie Liebenberg, Louis van der Merwe, Ronnie Gilfillan and Rilette Kiggen for their technical support during execution of various experiments. Thanks to the staff members of SADC-ICRAF Malawi for their incessant help and I cherish the wise guidance of Dr F.K. Akinnifesi throughout my studies. Finally, I would like to acknowledge the financial support from the Federal Ministry of Economic Cooperation (BMZ/GTZ) Germany through the World Agroforestry Centre for Southern Africa Regional Programme. I also acknowledge the partial financial support from Agro-Forest-Bio Energy Association.

To my dear wife, Moureen and my son, Wellington, the road to education was rough and hopefully the fruits will be sweet.



Table of Contents

		P	age
Dec	laratio	on	ii
Ack	nowle	edgements	iii
Tab	le of C	Contents	iv
List	of Tal	bles	xi
List	of Fig	gures	xiv
List	of Ab	breviationsx	xiii
Abs	tract		xxiv
GE l	NERA	AL INTRODUCTION	1
CH	APTE	ER 1: LITERATURE REVIEW	7
1.1	Uapa	ca kirkiana fruit trees	7
	1.1.1	Botany and ecological distribution	7
	1.1.2	Importance and commercial potential	8
	1.1.3	Production and cultivation	9
1.2	Jacke	et plum (Pappea capensis) tree species	
	1.2.1	Botany and ecological distribution	
	1.2.2	1	
	1.2.3	Production and cultivation	. 11
1.3	Tree	domestication process	. 12
1.4	Propa	agation methods	12
	1.3.1	Sexual propagation	12
	1.3.2	Vegetative propagation	. 13
	1.3.3	Culture contamination	. 15



1.5	Myco	rrhizae	16
1.6	Effect	ts of phenolics on graft compatibility	16
	1.6.1	Methods of separation for phenolic compounds	18
1.7	Sumn	nary	19
СН	APTE	R 2: IN VITRO PROPAGATION OF MATURE	
UA.	PACA	KIRKIANA Müell Arg. TREES	24
2.1	Abstr	act	24
2.2	Intro	duction	25
2.3	Mate	rials and methods	26
	2.3.1	Plant material	26
	2.3.2	Site description	27
	2.3.3	Efficacy of sodium hypochlorite and calcium hypochlorite	
		on field collected explants	28
	2.3.4	Efficacy of mercuric chloride on shoot explants from grafted trees	28
	2.3.5	Effect of medium supplements on contamination of leaf explants	29
	2.3.6	Shoot multiplication	29
	2.3.7	Root regeneration	29
	2.3.8	Culture conditions	30
	2.3.7	Statistical analysis	30
2.4	Resul	ts and discussion	30
	2.4.1	Efficacy of sodium hypochlorite and calcium hypochlorite	
		on field collected explants	30
	2.4.2	Efficacy of mercuric chloride on explants	
		excised from grafted trees	32
	2.4.3	Effect of medium supplements on decontamination	
		of leaf explants	32
	2.4.4	Effect of explant age on <i>in vitro</i> phenol production	33



	2.4.5	Shoot multiplication	33
	2.4.6	Rooting of micro-shoots	36
2.5	Concl	lusion	38
СН	APTE	R 3: HISTOLOGICAL EVALUATION OF EARLY GRAFT	
CO	MPAT	TIBILITY OF SCION/STOCK COMBINATIONS IN UAPACA	
KIF	RKIAN	A Müell Arg. TREE PROVENANCES	43
3.1.	Abstr	act	43
3.2	Intro	duction	44
3.3	Mater	rials and methods	45
	3.3.1	Field study	45
	3.3.2	Histological and anatomical studies	46
	3.3.3	Statistical analyses	47
3.4	Resul	ts and discussion	47
	3.4.1	Field study	47
	3.4.2	Histological and anatomical studies	49
3.5	Concl	lusion	55
СН	APTE	R 4: EARLY RECOGNITION OF GRAFT COMPATIBILITY	
IN	UAPA (CA Müell Arg. FRUIT TREES	64
4.1	Abstr	ract	64
4.2	Intro	duction	65
4.3	Mater	rials and methods	66
	4.3.1	Plant material	66
	4.3.2	Decontamination and callus induction	67
	4.3.3	Callus co-culture and fixation	68



	4.3.4	Statistical analyses	69
4.4	Resul	ts and discussion	69
	4.4.1	Decontamination of explants	69
	4.4.2	Callus induction.	70
	4.4.3	Callus union interface between <i>Uapaca kirkiana</i> clones	71
	4.4.4	Callus union interface within <i>Uapaca kirkiana</i> provenances	72
	4.4.5	Callus union between Uapaca kirkiana and Uapaca nitida	74
	4.4.6	Callus union between <i>Uapaca kirkiana</i> and <i>Jatropha curcas</i>	74
4.5	Concl	usion	75
		R 5: DIAGNOSIS OF PHENOLIC COMPOUNDS ATED IN GRAFT INCOMPATIBILITY IN <i>UAPACA</i>	
		A Müell Arg. FRUIT TREES	85
5.1	Abstr	act	85
5.2	Intro	duction	86
5.3	Matei	rials and methods	87
	5.3.1	Plant materials	87
	5.3.2	Phenol extraction	88
	5.3.3	Quantification of total soluble phenol	88
	5.3.4	Fluorescence microscopy	89
	5.3.5	Phenol analysis using RP-HPLC	90
5.4	Resul	ts and discussion	91
	5.4.1	Diameters of scion, stock and graft union	91
	5.4.2	Total soluble phenol quantity in the wood	92
	5.4.3	Total soluble phenols quantity in the bark	93
	5.4.4	Cell wall bound phenols.	94
	5.4.5	Fluorescence microscopy	95
	5.4.6	Phenol analysis using RP-HPLC	97



5.5	Conc	lusion	99
CII			
		R 6: PRE-TREATMENT METHODS FOR IMPROVING PLUM (<i>PAPPEA CAPENSIS</i>) SEED GERMINATION	115
U			
6.1	Abstr	act	115
6.2	Intro	duction	116
6.3	Mate	rials and methods	117
	6.3.1	Site description	117
	6.3.2	Effects of temperature and light on seed germination	118
	6.3.3	Germination of floating and submerged seeds	118
	6.3.4	Seed decontamination	119
	6.3.5	Incubation condition	119
	6.3.6	Statistical analysis	119
6.4	Resul	Its and discussion	120
	6.4.1	Effects of temperature and light on seed germination	120
	6.4.2	Germination of floating and submerged seeds	121
	6.4.3	Seed decontamination	122
6.5	Conc	lusion	123
		R 7: IN VITRO PROPAGATION OF JACKET PLUM	
(PA	PPEA	CAPENSIS) TREE SPECIES	128
7.1	Abstr	act	128
7.2	Intro	duction	129
7.3	Mate	rials and methods	130
	7.3.1	Stem cuttings	130
	7.3.2	Pilot micro-propagation experiment	131
	7.3.3	Shoot multiplication	132



	7.3.4	Root regeneration	134
	7.3.5	Repeated exposure of micro-cuttings to IBA	134
	7.3.6	Statistical analysis	134
7.4	Resul	ts and discussion	134
	7.4.1	Stem cuttings	134
	7.4.2	Pilot micro-propagation experiment	135
	7.4.3	Shoot multiplication	135
	7.4.4	Root regeneration	135
	7.4.5	Repeated exposure of micro-cuttings to IBA	139
	7.4.6	Acclimatization	139
7.5	Concl	usion	139
		R 8: JACKET PLUM (<i>PAPPEA CAPENSIS</i>) EGENERATION THROUGH INDIRECT	
		C EMBRYOGENESIS	145
SO	MATIO		
SO:	MATI(C EMBRYOGENESIS	145
8.1 8.2	MATIO Abstr	C EMBRYOGENESISact	145
8.1 8.2	MATIO Abstr	C EMBRYOGENESIS act duction	145 146 147
8.1 8.2	MATIO Abstra Introd Mater	C EMBRYOGENESIS act luction rials and methods	145 146 147
8.1 8.2	Abstraction Mater 8.3.1	C EMBRYOGENESIS act luction rials and methods. Plant material	145 146 147 148
8.1 8.2	Abstraction Mater 8.3.1 8.3.2	c EMBRYOGENESIS act duction rials and methods Plant material Callus induction	145 146 147 148
8.1 8.2	Abstraction Mater 8.3.1 8.3.2 8.3.3	C EMBRYOGENESIS act duction rials and methods Plant material Callus induction Induction of somatic embryos	145 146 147 148 148
8.1 8.2	MATIC Abstraction Introd Mater 8.3.1 8.3.2 8.3.3 8.3.4	C EMBRYOGENESIS act duction rials and methods Plant material Callus induction Induction of somatic embryos Germination of somatic embryos	145 146 147 148 148 149
8.1 8.2 8.3	MATIC Abstraction Mater 8.3.1 8.3.2 8.3.3 8.3.4 8.3.5 8.3.6	act duction rials and methods Plant material Callus induction Induction of somatic embryos Germination of somatic embryos Culture conditions	145 147 147 148 149 150
8.1 8.2 8.3	MATIC Abstraction Mater 8.3.1 8.3.2 8.3.3 8.3.4 8.3.5 8.3.6	c EMBRYOGENESIS act duction rials and methods Plant material Callus induction Induction of somatic embryos Germination of somatic embryos Culture conditions Statistical analysis	145 146 147 148 149 150 150
8.1 8.2 8.3	MATION Abstraction Mater 8.3.1 8.3.2 8.3.3 8.3.4 8.3.5 8.3.6 Result	ct EMBRYOGENESIS act luction rials and methods Plant material Callus induction Induction of somatic embryos Germination of somatic embryos Culture conditions Statistical analysis ts and discussion	145146147148149150150



8.5 Conclusion	
CHAPTER 9: GENERAL DISCUSSION	
AND CONCLUSIONS	161
9.1 Propagation of woody perennial trees	161
9.2 Suggestions for future investigations	
SUMMARY	172
REFERENCES	175



List of Tables

Table	les	Page
2.1	Uapaca kirkiana leaf culture percentage asepsis explanted on	
	Murashige and Skoog (MS) medium supplemented with	
	benzylaminopurine (BAP),α-naphthaleneacetic acid (NAA),	
	thidiazuron (TDZ) and indole-3-butyric acid (IBA). There were	
	no significant differences (P≤0.05) amongst treatments	39
2.2	Uapaca kirkiana shoot multiplication on three quarter strength	
	Murashige and Skoog (MS) medium supplemented with thidiazuron	
	(TDZ), Benzylaminopurine (BAP), kinetin (Kin), indole-3-butyric acid	
	(IBA), α-naphthaleneacetic acid (NAA) or casein hydrolysate (CH).	
	Means are calculated with standard errors	40
3.1	Tree identification (ID) of <i>Uapaca kirkiana</i> stocks and scions	
	from different districts and locations in the landscape (natural	
	forest or cultivated field) in Malawi	56
3.2	Average scion, stock, and graft union diameters and bark thicknesses	
	of young Uapaca kirkiana fruit trees (one-year old after grafting).	
	Measurements taken approximately 5 mm below and above the graft	
	union and means are calculated with standard errors ($N = 40$)	57
3.3	Mean scores of <i>Uapaca kirkiana</i> graft combinations with respect	
	to absence or presence of visible line in the bark and wood, callus	
	proliferation and phenol accumulation	58
4.1	Callus co-cultures of <i>Uapaca kirkiana</i> , <i>Uapaca nitida</i> and	
	Jatropha curcas	77



4.2	Percentage contamination of young and old Uapaca kirkiana
	leaves explanted on different Murashige and Skoog medium
	supplemented with benzylaminopurine (BAP), indole-3-butyric acid
	(IBA), α-naphthaleneacetic acid (NAA), 2, 4-dichlorophenoxyacetic
	acid (2, 4-D) and thidiazuron (TDZ). Means are calculated with
	standard deviations
4.3	Callus induction of <i>Uapaca kirkiana</i> explants on Murashige and Skoog (MS) medium supplemented with benzylaminopurine (BAP),
	indole-3-butyric acid (IBA), α-naphthaleneacetic acid (NAA),
	2,4-dichlorophenoxyacetic acid (2,4-D) and thidiazuron (TDZ).
	Means are calculated with standard errors
5.1	Tree identification (ID) of <i>Uapaca kirkiana</i> graft combinations
	collected at Makoka Research Station in Malawi
5.2	Total soluble phenol quantity (mg gallic acid equivalent per
	g of dry weight, DW) in the wood of ten Uapaca kirkiana trees
	measured above, below and at the union
5.3	Total soluble phenol quantity (mg gallic acid equivalent per g
	of dry weight, DW) in the bark of <i>Uapaca kirkiana</i> grafted trees measured
	above, below and at the unions
5.4	Total soluble phenol quantity (mg gallic acid equivalent per g
	of dry weight, DW) at different positions of Uapaca kirkiana graft unions
	in the wood and bark. Data calculated with standard errors 103
5.5	Table of fluorescence colours and the associated phenol groups
	(Regnier & Macheix, 1996; Du Plooy, 2006)
6.1	Effect of temperature (°C) and darkness (D) on root length (cm)
	and shoot length (cm) of jacket plum (Pappea capensis) seedlings 124



7.1	Mean number and condition of jacket plum (Pappea capensis)	
	micro-shoots produced on Murashige and Skoog (MS) medium	
	supplemented with benzylaminopurine (BAP), kinetin (Kin),	
	α-naphthaleneacetic acid (NAA) and gibberellic acid (GA ₃)	139
8.1	Callus induction (%) from leaf and cotyledon sections of jacket plum	
	(Pappea capensis) after four weeks on Murashige and Skoog (MS)	
	medium supplemented with benzylaminopurine (BAP), thidiazuron	
	(TDZ), kinetin (Kin), α-naphthaleneacetic acid (NAA),	
	dichlorophenoxyacetic acid (2,4-D) and indole-3-butyric acid (IBA).	
	Means with the same letters within a column are not significantly	
	different at P≤0.05	155
8.2	Pappea capensis somatic embryos induced on different Murashige	
	and Skoog (MS) medium supplements from juvenile and mature tissues	
	after 12 weeks. Data were recorded with standard error of means	156
8.3	Effect of different Murashige and Skoog (MS) medium supplements	
	on jacket plum (Pappea capensis) somatic embryos after 10 weeks	157



List of Figures

Figu	ire	Page
1.1	Uapaca kirkiana (A) tree in fruit; (B) grafted tree growing at	
	Makoka Station in Malawi (two years old after grafting);	
	(C) a tree with heavy fruit load; (D) fruits	20
1.2	Map of southern Africa showing the distribution of	
	Miombo woodlands (shaded areas) (adapted from White, 1983)	21
1.3	Jacket plum (Pappea capensis) (A) adult tree; (B) fruits	
	and a few shattered pods	21
1.4	A schematic diagram for domestication of wild tree species	22
1.5	Formation of cinnamic acid derivatives from phenylalanine	
	and tyrosine	23
1.6	Structure of (A) para- coumaric acid and	
	(B) ferulic acid (Liu et al., 2006b)	23
2.1	Uapaca kirkiana explants on three quarter strength Murashige and	
	Skoog medium supplemented with (A) 0.1 mg l ⁻¹ benzylaminopurine	
	(BAP), 0.04 mg 1^{-1} α -naphthaleneacetic acid (NAA) and 0.3 mg 1^{-1}	
	casein hydrolysate (CH); (B) callusing on 0.2 mg l ⁻¹ thidiazuron	
	(TDZ) and 0.3 mg l ⁻¹ CH; (C) old shoot explant not responding to	
	three quarter strength MS medium supplemented with a combination	
	of 0.05 mg l ⁻¹ TDZ and 0.3 mg l ⁻¹ CH after three weeks	41
2.2	Uapaca kirkiana root regenerated on half strength Murashige and	
	Skoog medium supplemented with 2.5 mg l ⁻¹ indole-3-butyric acid	
	(IBA). Arrow shows base callusing of a plantlet	42



2.3	Six months old <i>Uapaca kirkiana</i> plant growing in a pot
3.1	Morphology of <i>Uapaca kirkiana</i> graft unions (A) HardwickMW26
	(scion) on ElsoniMW22 (stock) and (B) NkhumbaMW1 (scion) on
	MigowiMW61 (stock)
3.2	Growth irregularities at the graft unions of Uapaca kirkiana fruit trees
	(A) a groove across the union; (B) a small scion on an overgrown stock;
	(C) a constriction at the union (pen points at the union)
3.3	Longitudinal sections and external view of the graft unions of Uapaca
	kirkiana trees showing (A) poor rate of callusing at the union interface;
	(B) good amount of callus formation at the union. (Arrows show necrotic
	tissues for the internal sections and differences in callus proliferation for
	the external sections, $X = \text{dead bark area of the incompatible graft}$
	combination)
3.4	Uapaca kirkiana scion/stock combinations showing (A) a visible line
	between the scion (NazombeMW84) and stock (NkhumbaMW57);
	(B) a faint line between HardwickMW26 and ElsoniMW22 (X = death
	of bark and vascular tissues, arrows show visible union lines)
3.5	Two-dimensional correspondence analysis showing distribution and
	association of different Uapaca kirkiana graft combinations with respect
	to union line in the bark and wood, presence of phenols and callus
	callus proliferation at the union (U = unknown stock, λ = inertias)
3.6	Anatomical observations of callus tissues very prolific below the union
	area (A) numerous unfilled (lacuna) areas; (B) a few unfilled areas; (C)
	absence of unfilled areas (arrows indicate callus tissues and the stains
	at the union interface are phenols, $St = stock$, $Sc = scion$, $bar = 5 \mu m$)



3.7	Deposits at <i>Uapaca kirkiana</i> scion/stock union (A) incompatible partner
	(NkhumbaMW1/MigowiMW61) with high levels of deposits; (B) partially
	compatible union with high levels of deposits above and at the union;
	(C) callus cells breaking up deposits from below the union area. (Arrows
	indicate deposits, $St = stock$, $Sc = scion$, $bar = 5 \mu m$)
3.8	Sections below the union of a compatible HardwickMW26/
	ElsoniMW22 Uapaca kirkiana combination (A) invisible union line;
	(B) HardwickMW26 and ElsoniMW22 graft union section; (C) a faint
	union line (X = pocket/line of deposits, bar = 5 μ m)
4.1	Callus union interfaces within <i>Uapaca kirkiana</i> clones (A) MW32/32
	clonal homograft; (B) MW49/49 inter-specific homograft. The brown
	stains on the cells are due to deposits (polyphenols) and arrows show
	faint union line for MW49/49 inter-specific homograft
4.2	Sections of callus union interfaces of (A) Jatropha curcas heterograft;
	(B) Uapaca nitida heterograft (arrows indicate the union interfaces).
	No phenolic compound staining was present on J. curcas cells, but it
	was present on <i>U. nitida</i> cells
4.3	Callus union interfaces of <i>Uapaca kirkiana</i> tree provenances (A)
	Dedza/Mpwapwa; (B) Phalombe/Dedza; (C) Phalombe/Murewa;
	(D) Dedza/Chipata (Arrows show the union line)
4.4	Callus union interface of a <i>Uapaca kirkiana</i> tree provenance (A) Dedza
	(juvenile) and Choma (mature) callus union in a polythene ring; (B)
	same callus combination showing callus degeneration at the union
	(JC = callus from juvenile planting materials, MC = callus from mature
	planting materials). Arrows indicate union area between calli from
	mature and juvenile stock plants (bar = $10 \mu m$)



4.5	Callus co-cultures of <i>Uapaca kirkiana</i> tree provenances in thermal
	labile rings (A) Phalombe/Nyamakwaar; (B) Dedza/Chipata (jc = callus
	from juvenile tissues, mc = callus from mature tissues). Arrows show
	bead-like projections
4.6	Callus union interface sections between: (A) Uapaca kirkiana (Uk)
	And Uapaca nitida (Un); (B) Uapaca kirkiana (Uk) and Jatropha
	curcas (Jc) showing phenol accumulation on the U. kirkiana cells.
	Arrows show callus union
4.7	Distribution and association of <i>Uapaca kirkiana</i> tree clone and
	provenance, U. nitida and Jatropha curcas callus combinations
	with respect to phenol, union line and cell necrosis attributes from
	correspondence analysis output (PP = MW84/84, DD = MW32/32,
	NK = MW49/49, $PAN = MW26/57$, $NN = U$. nitida heterograft,
	$JJ = J. \ curcas \ heterograft, \ NKP = MW57/26, \ PN = U. \ kirkiana/nitida,$
	DJ = <i>U. kirkiana</i> (Dedza)/ <i>J. curcas</i> , PAM = Phalombe/Murewa, PAC =
	Phalombe/Chipata, PAD = Phalombe/Dedza, NKM = MW57/Murewa,
	NKC = Phalombe/Chipata, NKN = Phalombe/Nyamakwaar, DMP =
	Dedza/Mpwapwa, DMU = Dedza/Murewa, DCH = Dedza/Chipata,
	DEC = Dedza/Choma) (λ = inertias)
5.1	Stem segments dissected from three-year old <i>Uapaca kirkiana</i>
	Grafted tree (1 and 2 = segments towards the stock, a & b = segments
	Towards the scion)
5.2	Diameters of scions, stocks and graft unions of Uapaca kirkiana
	trees (three years old after grafting) pooled together and measured
	approximately 5 mm above and below the union area



5.3	Morphological growth structure of <i>Uapaca kirkiana</i> wood at the union	
	(A) wart-like projections at the union; (B) lacuna or holes (thin arrows)	
	at the wood pith; (C) a swollen graft union (thick arrows) with some	
	projections (Rs = rootstock and Sc = scion)	106
5.4	Morphology of MW32 Uapaca kirkiana scions on two different	
	stocks (A) MW32/32 clonal homograft without wood discoloration	
	or holes; (B) MW32/28 heterograft with lacuna in wood	106
5.5	Cell wall bound phenol quantity (mg gallic acid equivalent	
	per g of dry weight, DW) in the bark measured above, below and at	
	the union (bars with the same letters within a column are not	
	significantly different at $P \le 0.05$)	107
5.6	Cell wall bound phenol quantity (mg gallic acid equivalent	
	per g of dry weight, DW) in the wood measured above, below and	
	at the union (bars with the same letters within a column are	
	not significantly different at $P \le 0.05$)	107
5.7	MW32/28 <i>Uapaca kirkiana</i> sections (above, below and at the union)	
	viewed under a fluorescence microscope using (ABC) UV light and	
	(DEF) Vanillin-HCl and white light	108
5.8	MW7/10 <i>Uapaca kirkiana</i> sections (above, below and at the union)	
	viewed under fluorescence microscope using (ABC) UV light and	
	(DEF) Vanillin-HCl and white light	109
5.9	MW26/26 Uapaca kirkiana sections (above, below and at the graft	
	union) viewed under a fluorescence microscope using (ABC)	
	UV light and (DEF) Vanillin-HCl and white light	110



5.10	MW12/12 Uapaca kirkiana sections (above the union, at the union and
	below the union) viewed under fluorescence microscope using (ABC)
	UV light and (DEF) Vanillin-HCl and white light (arrows show isolated
	white spots)
5.11	Chromatograms of cell wall bound phenols extracted from MW12/12
	Uapaca kirkiana in the bark (A) above the union and (B) below the union 112
5.12	Chromatograms of cell wall bound phenols extracted from the bark of
	MW26/26 Uapaca kirkiana combination (A) above the union and
	(B) below the graft union
5.13	Chromatograms of cell wall bound phenols extracted from the bark
	of MW7/10 Uapaca kirkiana combination (A) above the union and
	(B) below the graft union
5.14	Chromatograms of cell wall bound phenols extracted from
	MW12/12 Uapaca kirkiana in the wood (A) above the union
	and (B) below the union
5.15	Chromatograms of cell wall bound phenol extracts from the wood
	of MW26/26 Uapaca kirkiana combination (A) above the union and
	(B) below the graft union
5.16	Chromatograms of cell wall bound phenol extracts from the wood
	of MW7/10 Uapaca kirkiana combination (A) above the union and
	(B) below the union
6.1	Percentage seed germination of jacket plum (<i>Pappea capensis</i>)
	exposed to different temperatures (°C), darkness (D) and light (L)
	four weeks after sowing
	2



6.2	Jacket plum (Pappea capensis) seed/seedlings exposed to 24 hours
	of darkness and at (A) 12 °C; (B) 20 °C and (C) 25 °C temperatures
	(note that vigorous seedling growth was obtained at 25° C)
6.3	Jacket plum (Pappea capensis) seed germination (%) separated by
	water floatation method into floating, submerged (sunk), and a mixture
	of submerged and floating seeds
6.4	Figure 6.4 Percentage asepsis of jacket plum (Pappea capensis)
	seeds to 70% ethanol, 0.1% HgCl ₂ or 1.75% NaOCl pre-treatments.
	Bars with the different letters are significantly different at P≤0.05 126
6.5	Percentage germination of jacket plum (Pappea capensis) seeds to
	70% ethanol, 0.1% HgCl ₂ or 1.75% NaOCl pre-treatments.
	Bars with the same letters are not significantly different at P≤0.05
6.6	Jacket plum (Pappea capensis) seedlings in test tubes (A) normal
	seedling six days old; (B) abnormal seedling four weeks old
	(note brown culture medium in test tube B)
7. 1	Callus formation and bud break percentage of jacket plum (<i>Pappea</i>
	capensis) micro-shoots explanted on Murashige and Skoog medium
	supplemented with different concentrations (mg l ⁻¹) and combinations
	of benzylaminopurine (BAP), indole-3-acetic acid (IAA), indole-3-butyric
	acid (IBA), Kinetin (Kin), α-naphthaleneacetic acid (NAA) or thidiazuron
	(TDZ) four weeks after explanting
7.2	Jacket plum (Pappea capensis) micro-shoots on Murashige and
	Skoog medium (A) a high amount of callus formation at the bases
	of micro-shoots on 0.1 mg l ⁻¹ benzylaminopurine and 0.05 mg l ⁻¹
	α-naphthaleneacetic acid; (B) micro-shoot with a necrotic shoot tip
	(arrow shows shoot tip death) and new micro-shoots being produced



7.3	Jacket plum (<i>Pappea capensis</i>) shoot multiplication on Murashige	
	and Skoog medium supplemented with 2.0 mg l ⁻¹ benzylaminopurine	
	(BAP) after two weeks: (A) three healthy micro-cuttings; (B) single	
	shoot produced after subculturing the ortet explant	142
7.4	Percentage rooting of jacket plum (Pappea capensis) explanted on half	
	strength Murashige and Skoog medium supplemented with different	
	concentrations (mg l ⁻¹) and combination of indole-3-butyric acid (IBA),	
	α-naphthaleneacetic acid (NAA) and indole-3-acetic acid (IAA) three	
	weeks after explanting	143
7.5	Mean number of roots regenerated on jacket plum (Pappea capensis)	
	micro-cuttings rooted on different concentrations (mg l ⁻¹) and	
	combinations of indole-3-butyric acid (IBA), α-naphthaleneacetic	
	acid (NAA) and indole-acetic acid (IAA)	143
7.6	Rooted jacket plum (Pappea capensis) micro-cuttings on half strength	
	Murashige and Skoog medium supplemented with (A) 0.5 mg l ⁻¹	
	indole-3-butyric acid (IBA) and 0.5 mg l^{1} α -naphthaleneacetic acid	
	(NAA) after three weeks; (B) 0.5 mg l^{-1} IBA; (C) 0.5 mg l^{-1} IBA and	
	50 mg l ⁻¹ thiamine HCl. (Arrow shows high amount of callus formation	
	at the base of plantlet)	144
7.7	Jacket plum (Pappea capensis) plantlet rooted on half strength	
	Murashige and Skoog (MS) medium supplemented with 0.5 mg l ⁻¹	
	indole-3-butyric acid when explanted for the second time on the	
	same media (three weeks old)	144
8.1	Jacket plum (<i>Pappea capensis</i>) calli induced on three quarter Murashige	
	and Skoog medium with 0.05 mg 1 ⁻¹ thidiazuron and 0.3 mg 1 ⁻¹ casein	
	hydrolysate (A) chlorophyllous and embryogenic calli after three weeks;	
	(B) calli after four weeks	158



8.2	Jacket plum (Pappea capensis) somatic embryos on three quarter
	Murashige and Skoog medium with 0.05 mg l ⁻¹ thidiazuron and
	0.3 mg l ⁻¹ casein hydrolysate globular stage; (B) open cotyledonary
	somatic embryo; (C) closed-up polycotyledonary somatic embryo.
	(A long arrow shows aclosed-up embryo tip while a short arrow
	shows an open tip)
8.3	Jacket plum (Pappea capensis) embryos regenerated through
	Indirect embryogenesis (A) cluster of somatic embryos on Murashige
	and Skoog medium with 0.05 mg l ⁻¹ thidiazuron and 0.3 mg l ⁻¹ casein
	hydrolysate (CH); (B) embryo germination on 0.2 mg l ⁻¹ benzylaminopurine
	and 0.3 mg l ⁻¹ CH five days old, after being separated from a cluster of
	embryos. Arrows indicate roots
8.4	Jacket plum (Pappea capensis) plantlets regenerated through
	Embryogenesis after eight weeks on three quarter strength
	Murashige and Skoog (MS) medium with either (A) 0.2 mg l ⁻¹
	benzylaminopurine (BAP) and 0.3 mg l ⁻¹ casein hydrolysate (CH)
	or (B) 0.05 mg l ⁻¹ thidiazuron and 0.3 mg l ⁻¹ CH and later on with
	0.2 mg l ⁻¹ BAP and 0.3 mg l ⁻¹ CH
9.1	A schematic diagram for grafted (splice method) <i>Uapaca</i>
	kirkiana tree species
9.2	A schematic diagram for in vitro propagation of Uapaca
	kirkiana tree species
9.3	A schematic diagram for propagation methods of
	Pappea capensis tree species



List of Abbreviations

BAP benzylaminopurine
CH casein hydrolysate
IAA indole-3-acetic acid

IBA indole-3-butyric acid

DW dry weight

GA₃ gibberellic acid

Kin kinetin

MS Murashige and Skoog
NAA naphthaleneacetic acid

PAR photosynthetically active radiation

RP-HPLC reverse phase high performance liquid chromatograph

psi pounds per square inch

 $TDZ \hspace{1cm} thidiazuron \\ t_R \hspace{1cm} retention time \\ UV \hspace{1cm} ultra \ violet$

2,4-D dichlorophenoxyacetic acid



Development of clonal propagation protocols for *Uapaca kirkiana* and *Pappea capensis*, two southern African trees with economic potential

By

Simon Alfred Mng'omba

Supervisor : Prof. E. S. du Toit

Co-supervisors : Dr F. K. Akinnifesi

: Dr H. M. Venter

Department : Plant Production and Soil Science

Degree : PhD (Horticulture)

Abstract

Experiments were carried out with the objectives of developing propagation protocols for *Uapaca kirkiana* and *Pappea capensis* tree species of southern Africa, and evaluating the graft compatibility within *U. kirkiana* tree clones, provenances and species. Reverse phase high performance liquid chromatography (RP-HPLC), Folin-Ciocalteau reagent, fluorescence microscopy and callus fusion methodologies were used to diagnose graft compatibility. Results indicated that *U. kirkiana* culture asepsis was achieved with 0.1% w/v mercuric chloride (HgCl₂) and using pre-conditioned grafted trees. Sodium hypochlorite (NaOCl) improved *P. capensis* seed asepsis and germination, and discarding



floating seeds improved germination. Murashige and Skoog (MS) medium with 2.0 mg 1⁻¹ benzylaminopurine (BAP) and 0.3 mg 1⁻¹ casein hydrolysate (CH) was superior in shoot multiplication and 0.5 mg 1⁻¹ indole-3-butyric acid (IBA) for rooting of *P. capensis* microshoots. For somatic embryogenesis, three quarter strength MS medium with 0.05 mg 1⁻¹ thidiazuron (TDZ) and 0.3 mg 1⁻¹ CH, or 0.2 mg 1⁻¹ BAP with 0.3 mg 1⁻¹ CH, were effective in germination of *P. capensis* somatic embryos.

For *U. kirkiana* lateral shoot explants, shoot multiplication was superior on three quarter strength MS medium with 0.1 mg Γ^1 BAP and 0.3 mg Γ^1 CH. Rooting of micro-cuttings (36%) was achieved on ½ MS with 2.5 mg Γ^1 IBA. RP-HPLC, fluorescence microscopy and callus fusion studies showed that phenolic compounds play a major role in *U. kirkiana* graft incompatibility. Less graft compatible combinations showed an increase in phenol deposits above the union and graft incompatibility was more pronounced above the union than below the union. Proliferation of parenchymatous tissues was better below the union than above the union. Fluorescence microscopy showed presence of flavonoids and polymers above the union of less graft compatible combinations. The chromatograms showed that ferulic acid was abundant and responsible for wood discolouration. The chromatograms also isolated ρara -coumaric acids which were predominant above the union of the less compatible combinations. Therefore, ρara -coumaric acids, flavonoids and polymers were implicated in graft incompatibility of *U. kirkiana* trees.

Key words: decontaminants, embryogenesis, graft compatibility, Miombo woodland, organogenesis, phenolics, rejuvenation, seed germination



GENERAL INTRODUCTION

Trees contribute significantly to the livelihood of people in many different ways. They are sources of food, timber, poles and medicines and are grown for ornamental purposes such as for shade and protection. Wild trees have become potential commercial crops because they have multiple uses and imminent value. Edible fruits and oils are sources of food and income, and hence becoming important to many countries. Fruit trees can be utilized in different forms, for example *Uapaca kirkiana* Müell Arg. fruits can be eaten raw and made into refreshing drinks and a variety of wines (Kwesiga & Mwanza, 1995). The fruits have, therefore, become important to people in rural communities, especially in the arid and semi-arid countries of Africa where production of staple food has declined due to erratic rainfall, infertile soils and lack of farm inputs (Akinnifesi, Simons & Kwesiga, 2000a). Consequently, many households are food insecure and suffer from chronic malnutrition, of which children are the most vulnerable group (Akinnifesi et al., 2000b). Jacket plum (Pappea capensis L.) is another important wild tree which is rich in seed oil (74%) and this oil is usually used to make soap and for oiling guns (Venter & Venter, 1996). This oil is also potential source of bio-diesel (Le Roux, 2004). The tree also produces edible fruits which are made into jelly, vinegar and beverages (Venter & Venter, 1996).

People in rural communities can generate substantial income from wild fruit trees. For example, *U. kirkiana* fruits are gathered from the wild to be sold. They also serve as food reserves during seasonal food shortages (Maghembe & Seyani, 1992; Akinnifesi *et al.*, 2006). *U. kirkiana* trees grow in symbiotic association with mushrooms, which is also a cheap but rich source of nutrients (Maghembe, Simons, Kwesiga & Rarieya, 1998). In



many rural areas, women and children collect edible mushrooms for food and extra income (Okafor & Lamb, 1992; Saka, Mwendo-Phiri & Akinnifesi, 2002; Akinnifesi *et al.*, 2004). Moreover, fresh fruits are good sources of vitamins (e.g. vitamin C). *P. capensis* seed oil can be a viable source of income to the rural communities as it is a potential source of biodiesel fuel.

Food shortage in arid and semi-arid countries is prevalent but this problem can be reduced through food diversification. Many countries of southern Africa are hit by human immunodeficiency virus (HIV) / acquired immunodeficiency syndrome (AIDS) pandemic, and hence poor nutrition aggravates the plight of HIV/AIDS infected people. Therefore, fruits as nutritional and dietary supplements are vital and would fill in the food shortage gaps as well as being a source of income. The HIV/AIDS pandemic has a serious impact on agriculture as there is a low labour force, and hence loss in productivity (White & Robinson, 2000). Fruit trees are perennial and do not require much labour once they are established unlike annual crops where labour is required annually (Le Roux, 2004).

Incorporating potential wild fruit tree species onto farmland or managing fruit tree species in their natural habitats is important, especially tree species in demand (Simons, 1997). This will reduce continuous harvesting from the wild and allow these trees to be managed as renewable resources. However, incorporating wild fruit trees onto farmland would demand developing reliable propagation methods and protocols that maintain desirable traits of the high valued wild fruit tree species (Akinnifesi *et al.*, 2000b).



Major challenges to the rural masses of southern Africa include poverty, poor health and food insecurity and yet the region has some important wild tree species that can provide alternative sources of income and food (Akinnifesi *et al.*, 2000b). Availability of markets for *U. kirkiana* and other wild fruits in countries of southern Africa indicates the need for domestication and subsequent commercial production of these wild fruit trees. Many wild tree seeds have valuable oil which can be an alternative source of diesel fuel such as *P. capensis*. These trees can be produced locally and create income-generating activity for the rural communities. Currently, there is lack of knowledge and a shortage of improved planting stocks. Furthermore, cultivation and domestication processes are hampered (Akinnifesi *et al.*, 2000b). Rapid propagation would speed up cultivation and commercialisation of potential fruits and reduce food insecurity and poverty prevailing in many rural communities of southern Africa. Therefore, efforts are needed to provide reliable planting materials and adequate information on management of wild trees.

Grafted fruit trees address the fruiting precocity problem and enable the capture of proven superior fruit traits. However, scion/stock incompatibility in certain grafted fruit trees is a major constraint that may cause high losses in planting stock and established trees (Errea, Felipe, Treutter & Feucht, 1994b). Simons (1987) estimated that about half a million grafted peach trees died in southeast USA due to scion/stock incompatibility. Therefore, selection of compatible scion/stock combinations is important to ensure successful orchard establishment and productivity. Such selection of compatible scions and stocks has been neglected previously in many orchards and do not exist in domestication of wild fruit trees. It is against this background that the present study focused on development of propagation



protocols for *U. kirkiana* and *P. capensis* tree species. The general objectives for the study are:

- To develop propagation protocols that enable rapid and mass production of *Uapaca kirkiana* and *Pappea capensis* planting materials
- 2. To evaluate the graft compatibility within *Uapaca kirkiana* tree clones, provenances and species

In order to test these general objectives, the following hypotheses were developed:

- Pappea capensis and Uapaca kirkiana tree species are amenable to different propagation methods to achieve mass production of planting materials
- 2. Graft compatibility exists within *Uapaca kirkiana* tree clones, provenances and species

Approach:

Topics are organised into chapter format. A general synopsis of the entire thesis and the two indigenous fruit tree species studied are presented in the general introduction. The literature review (Chapter 1) provides the available information on *P. capensis* and *U. kirkiana* tree species. These include existing propagation methods, graft compatibility and the influence of polyphenols on *U. kirkiana* scion/stock combinations.



Major constraints to the micro-propagation of mature stock plants are rejuvenation and decontamination of plants associated with endophytic or cryptic microbes. Chapter 2 presents methods used to decontaminate *U. kirkiana* explants excised from grafted trees and this is followed by micro-propagation techniques employed (Chapter 3).

There are many unresolved questions on the main cause of graft incompatibility and methodologies to recognize graft incompatibility at an early stage are very limited. Secondary metabolites (phenols) occurring in vascular plants have been implicated in graft incompatibility. The amounts of phenols in plant organs vary with age, developmental state and growth conditions (Muofhe & Dakora, 1999). This provides an opportunity to time the grafts, and hence avoid accumulation of phenols at the graft unions. Chapters 4-6 aim at establishing the role of phenols on graft compatibility and potential methods to identify early scion/stock incompatibility in *U. kirkiana* trees. Histological studies, *in vitro* callus fusion, HPLC, fluorescence microscopy and Folin-Ciocalteau reagent procedures were used to improve our understanding of the role that phenols play in graft compatibility.

P. capensis is an unknown tree crop and is unexploited in terms of its commercial potential. No scientific research has yet been published on P. capensis propagation. Therefore, Chapters 7 - 8 focus on different propagation methods of P. capensis. Available literature documents that P. capensis seed germination is erratic and seedling growth is very slow (Venter & Venter, 1996). Organogenesis and somatic embryogenesis have been achieved in a few tree crops but many are still recalcitrant to in vitro propagation. No research has yet



been conducted in this field. The aim is, therefore, to improve germination and achieve mass multiplication of different plant sections taken from *P. capensis*.

Chapter 9 provides a general discussion of all the chapters and some general information related to the present study. Literature citation is found at the end of Chapter 9.

From this background information, it is clear that some wild fruit trees need special attention as they contribute significantly to food and income sources for rural community dwellers. Therefore, research is required on propagation techniques of the above mentioned valuable tree species. The results of research presented in various chapters provide the needed information to be utilised for the ultimate aim of improving productivity of the two wild fruit trees of southern Africa.