

**Development of an analytical method to measure  
17 $\beta$ -estradiol metabolite concentrations in MCF-7  
and MCF-12A cell lines.**

**by**

**Hermia van Zyl (9700538)**

**Submitted in fulfilment of part of the requirements for the degree of  
Master of Science (Physiology) in the Faculty of Health Sciences**

**University of Pretoria**

**Study leaders: Dr M-L Lottering**

**Dr A Joubert**

**Dr JB Laurens**

**November 2004**

**This project is dedicated to my Lord God: Father, Son and Holy Spirit.  
For without His mercy, grace and continuous encouragement and  
strengthening, I would not have been able to complete what I have  
started**

**Numbers 23:19**

God is not a man, that He should tell or act a lie, neither the son of man, that He should feel repentance or compunction [for what He has promised]. Has He said and shall He not do it? Or has He spoken and shall He not make it good?

**Hebrews 10: 35-36**

Do not, therefore, fling away your fearless confidence, for it carries a great and glorious compensation of reward. For you have need of steadfast patience and endurance, so that you may perform and fully accomplish the will of God, and thus receive and carry away [and enjoy to the full] what is promised.

**Philippians 2: 14**

Do all things without grumbling and faultfinding and complaining [against God] and questioning and doubting [among yourselves],

**Philippians 4:13**

I have strength for all things in Christ who empowers me [I am ready for anything and equal to anything through Him Who infuses inner strength into me; I am self-sufficient in Christ's sufficiency].

**1 Corinthians 10:13**

... But God is faithful [to His word and His compassionate nature], and He [can be trusted] not to let you be tempted and tried and assayed beyond your ability and strength of resistance and power to endure, but with the temptation He will [always] also provide the way out (the means of escape to a landing place), that you may be capable and strong and powerful to bear up under it patiently.

## Acknowledgements

I would like to thank the following:

- My Lord God: Father, Son and Holy Spirit, who are my source of strength and hope.
- My mother and father, for their encouragement and belief in me.
- My brothers, for their interest in my work and their love displayed in my life.
- My dear friend Hanri, for coping with my mood swings and frustration and still remaining a true friend.
- My dear friend Vera, for her support from afar.
- Dr Mona-Liza Lottering, for her excellent leadership, and continual support.
- Dr Annie Joubert, for her emotional support and leadership.
- Dr Tim Laurens, for his patience with my lack of knowledge and for showing me what it is to be a researcher.
- Dr Ilse Ker, for her friendship and for listening when I needed to blow off some steam.
- Dr Becker from the Medical Research Council for his help with the statistical planning of the physiological side of this study.
- The Department of Chemical Pathology at the University of Pretoria, for making their labs and instrumentation available to me, and for accepting me as one of their own.
- The Department of Physiology at the University of Pretoria, for making their labs and instruments available to me, and supporting me throughout this project.
- Marie Griffiths, who lighted the candle of interest and love for this incredible subject of Physiology.
- Angelique Elliott, for checking my grammar.

## Abstract

Breast cancer is one of the most common cancers affecting women. It remains the leading cause of death in American women from 30 – 70 years of age and approximately 10% of the women living in western countries will develop breast cancer during their life time. Estrogens are a family of female hormones involved in the reproductive function of the human body. Estradiol is the most abundant estrogen in premenopausal woman. Initially it was thought that estradiol itself was responsible for tumourigenesis, but it has since been discovered that the catechol metabolites of estradiol and other estrogens cause carcinogenesis. 4-Hydroxyestradiol (4-HE<sub>2</sub>) is a potent cell proliferating estrogen whereas 2-methoxyestradiol (2-ME<sub>2</sub>) is a potent inhibitor of cell proliferation through the activation of apoptosis. 2-Hydroxyestradiol (2-HE<sub>2</sub>) also causes increased cell proliferation but it is not as potent as 4-HE<sub>2</sub> and it is *O*-methylated rapidly to 2-ME<sub>2</sub>. Catechol metabolites of estradiol are also involved in producing reactive oxygen species through redox cycling. The reactive oxygen species cause DNA damage and mutations to occur which can lead to carcinogenesis. A significant ratio to consider as a biomarker for breast cancer risk therefore is the 4-HE<sub>2</sub>/2-ME<sub>2</sub> ratio. In this study, an analytical method was developed to measure the concentration levels of E<sub>2</sub>, 2-ME<sub>2</sub>, 2-HE<sub>2</sub> and 4-HE<sub>2</sub> in cell culture medium. The analytical method made use of gas chromatography-mass spectrometry (GCMS) analysis, since the expected physiological concentrations of these metabolites were very low. Various extraction and derivatisation techniques were applied during the development of the method. The final method made use of protein precipitation with concentrated hydrochloric acid, liquid-liquid extraction using diethyl

ether and derivatisation with trimethylsilylimidazole (TMSI). During the validation of this method, it was found that the method did not produce accurate measurements and that it could only be used to determine trends. Since the precise *in vitro* concentration levels of the metabolites were still unknown, it was decided to proceed with experiments using this method, to provide preliminary results with which further course of action could be planned. Equal numbers ( $1 \times 10^6$  cells/flask) of MCF-7 and MCF-12A cells were provided with 11 ml medium containing  $E_2/2\text{-HE}_2/4\text{-HE}_2/2\text{-ME}_2$  ( $10^{-6}$  M) and medium containing  $E_2/2\text{-HE}_2/4\text{-HE}_2/2\text{-ME}_2$  ( $10^{-8}$  M). Of each metabolite and each concentration two flasks were prepared. Each flask represented a specific time interval. At the appropriate time 10 ml of the medium was extracted. The time intervals used for each experiment were 0 hours, 8 hours and 24 hours for incubation with  $E_2$ , 0 hours, 1 hour and 8 hours for incubation with  $2\text{-ME}_2$ , 0 hours, 10 minutes and 1 hour for incubation with  $2\text{-HE}_2$  and  $4\text{-HE}_2$ . The time intervals used were according to the expected rate of metabolism. Each experiment was repeated three times. Differences in the metabolism of breast tumour cells and normal cells were found and the concentration of the metabolites present in the cell incubation medium had an influence on the metabolism of the cells. The need to investigate the intracellular concentrations of the metabolites has also been accentuated through the results obtained.

## Opsomming

Borskanker is een van die mees algemene vorme van kanker wat by vrouens voorkom. Dit bly die vernaamste oorsaak van dood onder vrouens tussen 30 en 70 jaar en ongeveer 10% van die vrouens in westerse lande sal borskanker ontwikkel gedurende hulle leeftyd. Estrogene is ‘n familie van vroulike hormone wat betrokke is by die voortplantingsfunksie van die menslike liggaam. estradiol is die volloppste estrogeen in premenopousale vrouens. Oorspronklik is daar gedink dat estradiol self verantwoordelik was vir die tumorgenese maar intussen is vasgestel dat die katesjolmetaboliete van estradiol karsinogenese veroorsaak. 4-Hidroksie estradiol (4-HE<sub>2</sub>) is ‘n sterk selprolifererende estrogeen terwyl 2-metoksie estradiol (2-ME<sub>2</sub>) ‘n sterk inhibitor is van selproliferasie deur die aktivering van apoptose. 2-Hidroksie estradiol (2-HE<sub>2</sub>) veroorsaak ook verhoogde selproliferasie, maar tot ‘n mindere mate as 4-HE<sub>2</sub> en dit word vinnig ge-*O*-metileer na 2-ME<sub>2</sub>. Die katesjolmetaboliete van estradiol is ook betrokke by die vorming van reaktiewe suurstofspesies deur die redokskringloop. Die reaktiewe suurstofspesies veroorsaak DNS skade en mutasies wat kan lei tot karsinogenese. ‘n Vername verhouding om ingedagte te hou as ‘n biomarker vir borskankerrisiko is die 4-HE<sub>2</sub>/2-ME<sub>2</sub> verhouding. In hierdie studie is ‘n analitiese metode ontwikkel om die konsentrasievlekke van E<sub>2</sub>, 2-ME<sub>2</sub>, 2-HE<sub>2</sub>, en 4-HE<sub>2</sub> in selkultuurmedium te meet. Die metode maak gebruik van gaschromatografie-massaspektrometrie (GC-MS) analyse omdat die verwagte fisiologiese konsentrasies van hierdie metaboliete baie laag is. Verskeie ekstraksie- en derivatiseringstegnieke is beproef tydens die ontwikkeling van die metode. Die finale metode het gebruik gemaak

van proteïenpresipitasie met gekonsentreerde soutsuur, vloeistof-vloeistof ekstraksie met diëtieletter en derivatisering met trimetilsilielimidazool (TMSI). Gedurende die validering van die metode is daar gevind dat die metode nie akkurate metings produseer nie en dat dit slegs gebruik kan word vir die bepaling van nygings. Omdat die presiese *in vitro* konsentrasievlekke van die metaboliete nog nie bekend is nie, is daar besluit om voort te gaan met eksperimente en gebruik te maak van hierdie analitiese metode om resultate te produseer waarmee die rigting vorentoe kan beplan word. Gelyke hoeveelhede ( $1 \times 10^6$  selle/fles) van MCF-7 en MCF-12A selle was voorsien van 11 ml medium wat E<sub>2</sub>/2-HE<sub>2</sub>/4-HE<sub>2</sub>/2-ME<sub>2</sub> ( $10^{-6}$  M) en medium wat E<sub>2</sub>/2-HE<sub>2</sub>/4-HE<sub>2</sub>/2-ME<sub>2</sub> ( $10^{-8}$  M) bevat. Vir elke metaboliet en elke konsentrasie is twee flesse voorberei. Elke fles het ‘n spesifieke tydsverloop verteenwoordig. Na afloop van die korrekte tydsinterval is 10 ml van die medium afgetrek. Die betrokke tydsintervalle was 0, 8 en 24 uur vir E<sub>2</sub>, 0, 1 en 8 uur vir 2-ME<sub>2</sub>, 0, 10 minute en 1uur vir 2-HE<sub>2</sub> en 4-HE<sub>2</sub>. Die tydsintervalle was bepaal volgens die verwagte tempo van metabolisme. Elke eksperiment was drie keer herhaal. Verskille in die metabolisme van borskankerselle in vergelyking met normale selle is gevind en die konsentrasies van die metaboliete teenwoordig in die selinkubasiemedium het ‘n invloed op die metabolisme van die selle. Die behoefte aan verdere navorsing in hierdie gebied het ook na vore gekom uit die verkrygde resultate.

# Table of Contents

<b>Abstract.....</b>	<b>i</b>
<b>Opsomming.....</b>	<b>iii</b>
<b>Table of Contents .....</b>	<b>v</b>
<b>List of Figures.....</b>	<b>viii</b>
<b>List of Tables .....</b>	<b>x</b>
<b>Abbreviations .....</b>	<b>xi</b>
<b>1 Introduction.....</b>	<b>1</b>
<b>1.1 Breast Cancer.....</b>	<b>3</b>
1.1.1 Etiology of Breast Cancer .....	3
1.1.1.1 Estrogen-related breast cancer .....	5
1.1.2 Estrogen Metabolism and Breast Cancer .....	9
1.1.2.1 Aromatisation of androgens .....	9
1.1.2.2 Hydroxylation of estrogens .....	10
1.1.2.2.1 2-Hydroxylation of estrogens .....	11
1.1.2.2.2 4-Hydroxylation of estrogens .....	12
1.1.2.3 16 $\alpha$ -Hydroxylation of estrogens .....	17
1.1.2.4 O-Methylation of estrogens.....	18
1.1.2.5 Conjugation of estrogens.....	22
<b>1.2 Problem Statement.....</b>	<b>23</b>
<b>1.3 Analytical Procedures.....</b>	<b>24</b>
<b>1.3.1 Gas chromatography (GC).....</b>	<b>24</b>
1.3.1.1 The Inlet System .....	25
1.3.1.1.1 Split injection .....	25
1.3.1.1.2 Splitless injection .....	25
1.3.1.2 The column .....	26
1.3.1.2.1 Stationary phases.....	28
1.3.1.2.2 Variables affecting column performance.....	28
1.3.1.2.3 Mobile phase .....	30
1.3.1.2.4 Temperature .....	30
1.3.1.3 Detectors .....	31
1.3.1.3.1 Flame Ionisation Detector (FID) .....	31
1.3.2 The GC-MS interface.....	31
<b>1.3.3 Mass Spectrometry (MS).....</b>	<b>32</b>
1.3.3.1 The ion source .....	32
1.3.3.2 The mass analyser .....	33
1.3.3.3 The ion detection system.....	34
1.3.3.4 Scanning modes .....	34
1.3.3.5 Ion chromatograms.....	35
1.3.3.6 Quantitation.....	35
1.3.3.7 Method of quantitation.....	36
1.3.4 Analytical method validation.....	37
<b>1.3.5 Sample preparation .....</b>	<b>40</b>
1.3.5.1 Protein precipitation.....	40
1.3.5.2 Extraction techniques .....	40
1.3.5.2.1 Solid phase extraction.....	40
1.3.5.2.2 Liquid/liquid extraction.....	41
1.3.5.3 Derivatisation .....	41
<b>2 Experimental Procedures.....</b>	<b>42</b>
<b>2.1 Development of Analytical Method.....</b>	<b>42</b>

<b>2.1.1</b>	<b>Materials .....</b>	<b>42</b>
<b>2.1.2</b>	<b>Methods.....</b>	<b>43</b>
2.1.2.1	Preparation of solvents .....	43
2.1.2.2	Steroid extraction .....	44
2.1.2.2.1	Liquid-liquid extraction with diethyl ether .....	44
2.1.2.2.2	Solid phase extraction with Varian C18 columns .....	44
2.1.2.2.3	Extraction with ethyl acetate after freeze drying .....	45
2.1.2.3	Derivatisation .....	45
2.1.2.3.1	BSTFA (N,O-Bis(trimethylsilyl)trifluoroacetamide).....	45
2.1.2.3.2	MTBSTFA (N-tert-Butyldimethylsilyl-N-methyltrifluoroacetamide) .....	45
2.1.2.3.3	TMSI (N-Trimethylsilylimidazole) .....	46
2.1.2.4	Deactivation of glassware .....	46
2.1.2.5	Protein precipitation .....	46
<b>2.1.3</b>	<b>Instrumentation .....</b>	<b>47</b>
<b>2.1.4</b>	<b>Procedures .....</b>	<b>47</b>
2.1.4.1	GC-FID parameter settings .....	47
2.1.4.2	GC-MS column parameter settings .....	47
2.1.4.2.1	J&W DB-1 Column.....	48
2.1.4.2.2	J&W DB-5MS column.....	48
2.1.4.2.3	Zebron ZB-5 Column .....	49
2.1.4.3	Sample gathering.....	50
2.1.4.4	Statistics .....	50
<b>2.2</b>	<b><i>In vitro Application of Analytical Method .....</i></b>	<b>50</b>
2.2.1	Aim .....	50
2.2.2	Materials .....	51
2.2.3	Methods.....	52
2.2.3.1	Culturing of cell lines.....	52
2.2.3.2	Determination of estrogen levels in cell culture medium.....	53
2.2.4	Statistics .....	54
<b>3</b>	<b>Results and Discussion.....</b>	<b>55</b>
<b>3.1</b>	<b>Method Development.....</b>	<b>55</b>
3.1.1	Elution order and retention times (BSTFA).....	55
3.1.2	Derivative stability (BSTFA).....	57
3.1.3	Elution order and retention time determination (MTBSTFA) .....	58
3.1.4	Reproducibility (MTBSTFA) .....	60
3.1.5	Elution order (TMSI) .....	62
3.1.6	Reproducibility (TMSI) .....	62
3.1.7	Optimisation of steroid extraction from physiological matrix.....	63
3.1.8	Calibration.....	65
3.1.9	Characterisation of controls by Standard Addition.....	76
<b>3.2</b>	<b>Application of Analytical Method .....</b>	<b>78</b>
3.2.1	Results.....	78
3.2.1.1	Estrogen levels in cell culture medium after incubation with E <sub>2</sub> .....	78
3.2.1.2	Estrogen levels in cell culture medium after incubation with 2-ME <sub>2</sub> .....	79
3.2.1.3	Estrogen levels in cell culture medium after incubation with 2-HE <sub>2</sub> .....	85
3.2.1.4	Estrogen levels in cell culture medium after incubation with 4-HE <sub>2</sub> .....	85
3.2.2	Discussion .....	86
3.2.2.1	Estrogen levels in cell culture medium after incubation with E <sub>2</sub> .....	87
3.2.2.2	Estrogen levels in cell culture medium after incubation with 2-ME <sub>2</sub> .....	89
3.2.2.3	Estrogen levels in cell culture medium after incubation with 2-HE <sub>2</sub> .....	90
3.2.2.4	Estrogen levels in cell culture medium after incubation with 4-HE <sub>2</sub> .....	91
<b>4</b>	<b>Conclusion.....</b>	<b>93</b>

<b>5</b>	<b>Mass spectra of TMS derivatives .....</b>	<b>96</b>
5.1	Mass spectrum of TMS derivative of E <sub>2</sub> .....	96
5.2	Mass spectrum of TMS derivative of 2-ME <sub>2</sub> .....	97
5.3	Mass spectrum of TMS derivative of 2-HE <sub>2</sub> .....	98
5.4	Mass spectrum of TMS derivative of 4-HE <sub>2</sub> .....	99
5.5	Mass spectrum of TMS derivative of 4HE <sub>2</sub> -5d.....	100
<b>6</b>	<b>References .....</b>	<b>101</b>

# List of Figures

Figure 1: Summary of the metabolism of estradiol under discussion.....	7
Figure 2: Activating and deactivating pathways of estradiol metabolism and the formation of DNA adducts. ....	15
Figure 3: The effects of estradiol 2-hydroxylation with subsequent <i>O</i> -methylation on cancer.....	20
Figure 4: Schematic representation of the Quadropole MS system .....	34
Figure 5: An example of a calibration curve used in internal standard quantitation .....	37
Figure 6: Derivatisation reaction: estradiol and trimethylsilylimidazole (TMSI). The arrows indicate the steps of the reaction .....	41
Figure 7: The results of the BSTFA derivative stability test .....	58
Figure 8: Elution order of MTBSTFA derivatised metabolites (5 $\beta$ -C = 5 $\beta$ -cholestane).....	59
Figure 9: Relative response of samples analysed for determination of CV (MTBSTFA derivatised).....	61
Figure 10: Relative response of samples analysed for determination of CV (TMSI).....	63
Figure 11: Diagram showing the effect of serum and the extraction procedure on relative peak area (DEE = diethyl ether) .....	64
Figure 12: Estradiol calibration curve on the GC-MS (5-1000ng/ml).....	66
Figure 13: 2-Methoxyestradiol calibration curve on the GC-MS (5-1000ng/ml).....	66
Figure 14: 2-Hydroxyestradiol calibration curve on the GC-MS (5-1000ng/ml) .....	67
Figure 15: 4-Hydroxyestradiol calibration curve on the GC-MS (5-1000ng/ml) .....	67
Figure 16: Extracted Ion Chromatogram (EIC) (A), TIC (B), and mass spectrum of peak at 8.16 min (C) of medium in which cells had grown. (416 = E <sub>2</sub> , 446 = 2-ME <sub>2</sub> , 504 = 2-HE <sub>2</sub> & 4-HE <sub>2</sub> , 357 = 5 $\beta$ -cholestane) .....	70
Figure 17: The Extracted Ion Chromatogram (EIC) of the ion used to identify E <sub>2</sub> (416) after 8 hours of exposure of cells to E <sub>2</sub> .....	71
Figure 18: The EIC of the ion used to identify 2-ME <sub>2</sub> (446) after 8 hours of exposure of cells to E <sub>2</sub> .....	71
Figure 19: The EIC of the ion used to identify 2-HE <sub>2</sub> and 4-HE <sub>2</sub> (504) after 8 hours exposure of cells to E <sub>2</sub> . .....	72
Figure 20: The EIC of the ion used to identify 4-HE <sub>2</sub> -d5 (509) 8 hours exposure of cells to E2.....	72
Figure 21: Estradiol calibration over the 40-1000 pg/ml range.....	73
Figure 22: 2-Methoxy estradiol calibration over the 40-1000 pg/ml range.....	73
Figure 23: 2-Hydroxy estradiol calibration over the 40-1000 pg/ml range.....	73
Figure 24: 4-Hydroxy estradiol calibration over the 40-1000 pg/ml range.....	74
Figure 25: 4-Hydroxyestradiol shows a non-linear relationship with the internal standard at a concentration higher than 100 ng/ml .....	75
Figure 26: The Natural isotope contribution of 4-HE <sub>2</sub> to the internal standard with increased 4-HE <sub>2</sub> concentration. (The two black dashed lines indicate mean $\pm 2\sigma$ ) .....	76
Figure 27: E <sub>2</sub> (A), 2-ME <sub>2</sub> (B), 2-H E <sub>2</sub> (C), 4-HE <sub>2</sub> (D) remaining in the medium of MCF-7 cells and MCF-12A cells after 0, 8, and 24 hours of incubation. Medium concentration is presented as a percentage of the initial concentration of the metabolite the celss were exposed to .....	80
Figure 28: Concentration of metabolites in medium of MCF-7 cells exposed to 1 x 10 <sup>-8</sup> M E <sub>2</sub> (A), 1 x 10 <sup>-6</sup> M E <sub>2</sub> (B), and in medium of MCF-12A cells exposed to 1 x 10 <sup>-8</sup> M E <sub>2</sub> (C), as well as 1 x 10 <sup>-6</sup> M E <sub>2</sub> (D) measured at different intervals including 0, 8 and 24 hours.....	81
Figure 29: Concentration of metabolites in medium of MCF-7 cells exposed to 1 x 10 <sup>-8</sup> M 2-ME <sub>2</sub> (A), 1 x 10 <sup>-6</sup> M 2-ME <sub>2</sub> (B), and in medium of MCF-12A cells exposed to 1 x 10 <sup>-8</sup> M 2-ME <sub>2</sub> (C), as well as 1 x 10 <sup>-6</sup> M 2-ME <sub>2</sub> (D) measured at different intervals including 0, 1 and 8 hours.....	82
Figure 30: Concentration of metabolites in medium of MCF-7 cells exposed to 1 x 10 <sup>-8</sup> M 2-HE <sub>2</sub> (A), 1 x 10 <sup>-6</sup> M 2-HE <sub>2</sub> (B), and in medium of MCF-12A cells exposed to 1 x 10 <sup>-8</sup> M 2-HE <sub>2</sub> (C), as well as 1 x 10 <sup>-6</sup> M 2-HE <sub>2</sub> (D) measured at different intervals including 0, 10 minutes and 1hour.....	83
Figure 31: Concentration of metabolites in medium of MCF-7 cells exposed to 1 x 10 <sup>-8</sup> M 4-HE <sub>2</sub> (A), 1 x 10 <sup>-6</sup> M 4-HE <sub>2</sub> (B), and in medium of MCF-12A cells exposed to 1 x 10 <sup>-8</sup> M 4-HE <sub>2</sub> (C), as well as 1 x 10 <sup>-6</sup> M 4-HE <sub>2</sub> (D) measured at different intervals including 0, 10 minutes and 1 hours.....	84
Figure 32: Mass spectrum of TMS derivative of E <sub>2</sub> .....	96

Figure 33: Mass spectrum of TMS derivative of 2-ME <sub>2</sub> .	97
Figure 34: Mass spectrum of TMS derivative of 2-HE <sub>2</sub> .	98
Figure 35: Mass spectrum of TMS derivative of 4-HE <sub>2</sub> .	99
Figure 36: Mass spectrum of TMS derivative of 4-HE <sub>2</sub> -5d.	100

# List of Tables

Table 1: Summary of the established risk and protective factors for breast cancer.....	6
Table 2: Summary of the E <sub>2</sub> concentrations in various tissue of pre- and postmenopausal woman.....	9
Table 3: Molecular masses of the silyl derivatised metabolites.....	56
Table 4: Structures of all the compounds applicable to this study.....	56
Table 5: Structures of the TMS (trimethylsilyl) derivatives formed by BSTFA derivatisation.....	57
Table 6: Molecular masses of MTBSTFA derivatised metabolites.....	59
Table 7: Structures of the t-butyldimethylsilyl (t-BDMS) derivatives formed by MTBSTFA derivatisation .....	60
Table 8: Coefficient of variance for the different metabolites using MTBSTFA.....	61
Table 9: CV of different metabolites using TMSI.....	62
Table 10: CV (concentration) of metabolites in calibration curve (0.5-10µg/ml) .....	65
Table 11: CV (concentration) of metabolites in calibration curve (5-1000ng/ml) .....	66
Table 12: CV(concentration) of metabolites in calibration curve on GC-MS (5-1000ng/ml).....	67
Table 13: Summary of the linear statistics of the calibration curves.....	74
Table 14: Statistical characterisation of the two controls .....	77

## Abbreviations

•OH	hydroxyl radical
16α-HE <sub>1</sub>	16α-hydroxyestrone
16α-HE <sub>2</sub>	16α-hydroxyestradiol
17β-HSD	17β-hydroxysteroid dehydrogenase
2-HE <sub>2</sub>	2-hydroxyestradiol
2-ME <sub>2</sub>	2-methoxyestradiol
4-HE <sub>2</sub>	4-hydroxyestradiol
4-HE <sub>2</sub> -d <sub>5</sub>	deuterated 4-hydroxyestradiol
5β-C	5β-cholestane
B	longitudinal diffusional spreading
BSTFA	N,O-bis(trimethylsilyl)trifluoroacetamide
C <sub>4</sub>	carbon atom 4
CE	catechol estrogens
C <sub>m</sub>	resistance to mass transfer in the mobile phase
COMT	catechol <i>O</i> -methyl transferase
C <sub>s</sub>	resistance to mass transfer in the stationary phase
CV	coefficient of variation
CYP450	cytochrome P450
DEE	diethylether
DMSO	dimethylsulphoxide
DNA	deoxyribonucleic acid

E <sub>1</sub>	estrone
E <sub>2</sub>	estradiol
E <sub>3</sub>	estriol
ECD	electron capture detection
EI	electron impact
EIC	extracted ion chromatogram
FID	flame ionisation detection
FSH	follicle-stimulating hormone
GC	gas chromatography
GC-FID	gas chromatography with flame ionisation detection
GC-MS	gas chromatography – mass spectrometry
GSH-S-transferase	glutathione S-transferase
H	theoretical plate height
HCl	hydrochloric acid
HPLC	high pressure liquid chromatography
i.d.	internal diameter
kPa	kilopascal
LH	luteinising hormone
MCF-12A	Michigan Cancer Foundation cell line 12A
MCF-7	Michigan Cancer Foundation cell line 7
MEME	minimum essential medium eagle
MS	mass spectrometer
MTBSTFA	N-tert-butyldimethylsilyl-N-methyltrifluoroacetamide

N	number of theoretical plates
NADH	nicotinamide adenine dinucleotide
NADPH	nicotinamide adenine dinucleotide phosphate
O <sub>2</sub> <sup>•</sup>	superoxide radical
PBS	phosphate buffered saline
ppb	parts per billion
ppm	parts per million
ppt	parts per trillion
RIA	radioimmuno assay
RNA	ribonucleic acid
ROS	radical oxygen species
R <sub>s</sub>	resolution
SCOT	support coated open tubular
SIM	single ion monitoring
t-BDMS	t-butyldimethylsilyl
TCDD	2,3,7,8-tetrachlorodibenzo- <i>p</i> -dioxin
TDLU	terminal ductal lobular units
TIC	total ion chromatogram
TMS	trimethylsilyl
TMSI	trimethylsilylimidazole
WCOT	wall coated open tubular