

Antioxidant properties and cellular protective effects of selected African green leafy vegetables

by

Nangula Paulina Mavhungu

Submitted in partial fulfilment of the requirements for the degree

PhD Nutrition

in the

Centre for Nutrition

Faculty of Natural and Agricultural Sciences

University of Pretoria

Pretoria

South Africa

14 November 2011



DECLARATION

I hereby declare that the thesis which I herewith submit at the University of Pretoria for the award of PhD degree (Nutrition) is my research and has not been submitted by me for a degree at any other university or institution of higher learning.

Nangula Paulina Mavhungu

14 November 2011



"perseverance, character; and character, hope." $\,$

Romans 5:4 (NIV)



ABSTRACT

Antioxidant properties and cellular protective effects of selected African green leafy vegetables

By

Nangula Paulina Mavhungu

Supervisor: Prof. A. Oelofse

Co-Supervisors: Prof. M.J. Bester

Dr. K.G. Duodu

Centre: Nutrition

Degree: PhD (Nutrition)

Phenolic compounds in African green leafy vegetables (GLVs) may have a significant impact on human health. However, there is little information on the phenolic composition, antioxidant properties, as well as biological and cellular protective effects of these vegetables. The effects of boiling and extraction solvent on these compounds and on their antioxidant properties are also unknown.

Phenolic content, antioxidant activity and cellular protective effects of four African GLVs in comparison with spinach, an exotic GLV, was determined. African GLVs had appreciable levels of total phenolics and antioxidant activity and in higher quantities compared to spinach. Boiling decreased the antioxidant content and activity of these vegetables and 75% acetone was more effective in extracting antioxidants from the GLVs compared to water. GLVs with high levels of phenolics also contained higher levels of antioxidant activity, suggesting that phenolics are likely to have contributed to radical scavenging activity of these vegetable extracts, even though the degree of scavenging varied in each extract of the vegetable species.



The flavonoid compositions of raw and boiled African GLVs and spinach were determined using high-performance liquid chromatography. Epicatechin and rutin were the most dominant flavonoids found in both water and 75% acetone extracts. Among water extracts, pumpkin contained higher concentrations of detected flavonoids, while among the acetone extracts, cowpea exhibited higher concentrations. The effect of boiling was dependent on the type of vegetable and the specific flavonoids. There were no major differences observed between the type of flavonoids detected in extracts of African GLVs and those in spinach. However, similar to the results of total phenolics and antioxidant activity, the 75% acetone extracts of African GLVs also exhibited higher amounts of flavonoids than spinach.

The protective effects of GLVs against oxidative haemolysis were dependent on the type of vegetable species. Boiling had variable effects depending on the species. The highest level of protection of erythrocytes against oxidative damage was offered by amaranth extracts, while extracts of raw jute mallow contributed to the damage of erythrocytes. The highest antioxidant protection activity against oxidative damage in plasmid DNA was offered by extracts of jute mallow and lowest by spinach.

For the cell viability assays, GLVs were evaluated to determine their cytotoxicity levels and functional role in oxidative damage. The results of the long-term cell viability (i.e. MTT, NR and CV) assays indicated no cytotoxicity, while the short-term cell viability (i.e. DCF) assay indicated that all extracts of raw GLVs were significantly (p < 0.05) cytotoxic to SC-1 fibroblast and human adenocarcinoma colon cancer (Caco-2) cells than extracts of cooked samples, and the levels of toxicity in the extracts of spinach was higher than in African GLVs. These results indicate that there was an initial cytotoxic effect as extracts of raw GLVs were added to the cells. However, after about 72 h, the cells recovered from the initial shock and started proliferating as usual. In the presence of peroxyl radicals, extracts of African GLVs exhibited higher protective effects against oxidative damage in both types of cell cultures than extracts of spinach. These results indicate that these protective effects could be attributed to the presence of phenolics and antioxidant properties of these extracts.



Although boiling reduced the antioxidant content and activity of African GLVs, the levels remained higher than in spinach. Boiling also decreased the cytotoxicity and cell damage caused by extracts of raw GLVs samples. African GLVs are consumed after boiling, and therefore the observed cytotoxicities might not be experienced in practical terms. African GLVs have therefore a potential to reduce the risk and development of diseases associated with oxidative stress in communities that consume these vegetables.



ACKNOWLEDGEMENTS

There is an African proverb which says, "Wisdom is like a baobab tree. One individual cannot embrace it". This wise saying reminds us that no matter how great the achievement, no individual may claim to have all the wisdom there is. I would like, therefore, to thank the following persons and institutions for their valuable contributions and assistance towards the successful completion of this research

Prof. A. Oelofse, Supervisor and Director at the Centre for Nutrition, University of Pretoria, for believing in me and for his constant inspiration that funds for my studies would be available even when I knew that resources were limited. With his support, every mountain suddenly became a hill. The guidance and assistance he rendered to me throughout this research study was invaluable; My co-supervisors, Dr. K.G. Duodu, senior lecturer at the Department of Food Science, University of Pretoria and Prof. M.J. Bester, professor at the Department of Anatomy, University of Pretoria, made the mission of climbing the hills both possible and achievable. Dr Duodu's expertise, intellectual guidance, insight and much appreciated review of my work have taught me to think out-of-the box and challenge my own ideas. With him I often felt academically challenged and yet highly empowered. Prof. Bester's expertise, scholarly guidance, patience, open door policy and regular support have taught me the immense value of having an academic mentor. I was lucky to have her in my team of supervisors; I will always be indebted to Mrs. C. Bowles for her regular and efficient administrative support.

Ms J. Serem, a post graduate student in the Cell Biology laboratory, for her motivation and for carrying out part of the cell culture work. With her, the journey was never lonely. Dr A.D. Cromarty and his colleagues in the Department of Pharmacology, University of Pretoria, for making their facilities available to me.

Dr N. Luruli, for kindly providing me with computer software programmes and other support I needed during this study. He was always the trouble-shooter outside the university.



The Faculty for the Future Programme of the Schlumberger Foundation, the Organization for Women in Science for the Developing World and the Water Research Commission for financial assistance provided throughout this research.

Mr W. Jansen van Rensberg for his advice with regards to African green leafy vegetables and and for providing me with the photographs of the vegetables used in this study; the Agricultural Research Council (ARC), Roodeplaat, for kindly providing the African green leafy vegetable samples.

My fellow post-graduate students, who I cannot all mention by names, for assistance and continuous encouragement throughout this research; Former colleagues at the Department of Food Science and Technology, University of Namibia, for continuous support.

My husband Khaukanani, for his unfaltering love and support, and for always having faith in me. His constant bragging that I was the only member of our family without a doctoral qualification served as a motivation in good and challenging times; My daughter Masindi, for her unconditional love, even when she never understood why in some weekends I would choose my studies over taking her to Magnolia Park. Her constant assurance that she was saving plenty of coins for my graduation party motivated me to work even harder at finishing this research; My parents and siblings, for their unfailing love and making me believe that we are a family of achievers; and last but not least,

God, for providing me the wisdom and perseverance to complete this study.



TABLE OF CONTENTS

DECLARATION	i
ABSTRACT	iii
ACKNOWLEDGEMENTS	vi
TABLE OF CONTENTS	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
GLOSSARY	xvi
CHAPTER 1: INTRODUCTION	1
1.1 Statement of the Problem	1
1.2 Literature Review	4
1.2.1 African green leafy vegetables	4
1.2.2 Nutritional composition of African GLVs	6
1.2.2.1 Proximate composition of African GLVs	8
1.2.2.2 Micronutrient content of African GLVs	9
1.2.3 Chemistry of plant phenolics	11
1.2.3.1 Phenolic acids	12
1.2.3.2 Flavonoids	13
1.2.3.3 Tannins	14
1.2.4 Phenolic compounds present in GLVs	16
1.2.5 Antioxidant properties of GLVs phenolics	16
1.2.6 Antioxidant mechanisms and structure-activity relationship of plant phenolics	18
1.2.7 Free radicals and oxidative stress	19
1.2.8 Evidence for health-promoting effects of fruits and vegetables	22
1.2.9 Health-promoting effects of some flavonoid-rich foods	23



1.2.10 Health-promoting effects of plant phenolics with particular reference to 0	GLVs 24
1.2.11 Dietary intake of flavonoids	25
1.2.12 Bioavailability of phenolics	26
1.2.13 Effects of cooking on phenolic content and antioxidant activity	27
1.2.14 Analytical methodology for the determination of antioxidant content and	activity28
1.2.14.1 Determination of total polyphenol and flavonoid content	30
1.2.14.2 Measurement of antioxidant activity	30
1.2.14.3 Biological and cellular assays	32
1.2.15 Conclusions	35
1.3 Hypotheses	35
1.4 Objectives	37
CHAPTER 2: RESEARCH	39
2.1 Raw and cooked African green leafy vegetables have greater antioxidant content	t and
activity than spinach	40
2.1.1 Abstract	40
2.1.2 Introduction	40
2.1.3 Materials and Methods	42
2.1.3.1 Green leafy vegetable samples and their preparation	42
2.1.3.2 Crude plant extracts	42
2.1.3.3 Analyses	43
2.1.3.3.1 Total phenolics	43
2.1.3.3.2 Total flavonoids	43
2.1.3.3.3 Antioxidant activity	43
2.1.3.4 Statistical analysis	45
2.1.4 Results and Discussion	45
2.1.4.1 Antioxidant content and activity of African GLVs compared to s	pinach 45
2.1.4.2 Effect of boiling on antioxidant content and activity of GLVs	48
2.1.4.3 Effect of extraction solvent	52
2.1.5 Conclusion	55



2.1.6 References	55
2.2 Comparative determination of flavonoids of African green leafy vegeta	ables and spinach by
high-performance liquid chromatography	61
2.2.1 Abstract	61
2.2.2 Introduction	61
2.2.3 Materials and Methods	62
2.2.3.1 Preparation of GLV samples and crude plant extracts	62
2.2.3.2 Reversed-phase HPLC analysis	62
2.2.3.3 Statistical analyses	63
2.2.4 Results and Discussion	64
2.2.4.1 Levels of flavonoids in raw GLVs	64
2.2.4.2 Effect of boiling on flavonoid contents of GLVs	67
2.2.4.3 Effect of extraction solvent	71
2.2.5 Conclusion	72
2.2.6 References	72
2.3 Protective effects of African green leafy vegetables against AAPH-indu	uced oxidative
damage	77
2.3.1 Abstract	77
2.3.2 Introduction	77
2.3.3 Materials and Methods	79
2.3.3.1 Green leafy vegetable samples and the preparation of co	rude plant extracts
	79
2.3.3.2 Analyses	79
2.3.3.2.1 Biological assays	79
2.3.3.2.2 In-vitro cellular assays	80
2.3.3.3 Statistical Analysis	83
2.3.4 Results and Discussion	83
2.3.4.1 Biological assays	83
2.3.4.1.1 Protection of erythrocytes by African GLVs	against oxidative
damage	83



2.3.4.1.2 Protection of plasmid DNA by African GLVs aga	inst oxidative
damage	85
2.3.4.2 In-vitro cellular assays	88
2.3.4.2.1 Cell viability assays	88
2.3.4.2.2 In vitro cellular antioxidant properties: Compariso	on of total,
intra- and extracellular protection assays	95
2.3.4.3 Correlation coefficients between different assays	100
2.3.5 Conclusion.	101
2.3.6 References	102
CHAPTER 3: GENERAL DISCUSSION	110
3.1 Methodologies	110
3.2 Research Findings	117
3.3 African GLVs may reduce chronic diseases of lifestyle	128
CHAPTER 4: CONCLUSIONS AND RECOMMENDATIONS	135
REFERENCES	138
APPENDIX	170



LIST OF TABLES

Table 1.2.1	Proximate composition of some African green leafy vegetables (values
	per 100 g edible portion, fresh weight (fw) basis)
Table 1.2.2	Vitamin and mineral content of African green leafy vegetables (values
	per 100 g edible portion, fw basis)
Table 1.2.3	Functional groups of phenolic acids
Table 1.2.4	Values of phenolic composition and antioxidant activity reported in
	African GLVs and spinach
Table 1.2.5	Flavonol and flavone contents of exotic vegetables
Table 1.2.6	Assays for determination of total phenolic and flavonoid contents
Table 1.2.7	Commonly used antioxidant assays
Table 1.2.8	Biological and cellular assays used to measure antioxidant effects
Table 2.1.1	Total phenolic content (TPC), total flavonoid content (TFC) and total
	antioxidant activity of water and 75% acetone extracts of raw African
	green leafy vegetables (GLVs) compared to spinach
Table 2.1.2	Effect of boiling on total phenolic content (TPC), total flavonoid content
	(TFC) and total antioxidant activity of water extracts of green leafy
	Vegetables (GLVs)
Table 2.1.3	Effect of boiling on total phenolic content (TPC), total flavonoid content
	(TFC) and total antioxidant activity of 75% acetone extracts of green
	leafy vegetables (GLVs)
Table 2.1.4	Correlation coefficients (r) between water and 75% acetone extracts for
	each assay per green leafy vegetable
Table 2.1.5	Correlation coefficients (r) between TPC, TFC, ABTS, DPPH and
	ORAC for water and 75% acetone extracts.
Table 2.2.1	Effect of boiling on levels of flavonoids (mg/g, dry weight) in water
	extracts of selected green leafy vegetables (GLVs)
Table 2.2.2	Effect of boiling on levels of flavonoids (mg/g, dry weight) in aqueous
	acetone extracts of selected green leafy vegetables (GLVs)



Table 2.3.1	Correlation coefficients (r) between different antioxidant assays	101
Table 3.1	Summary of the effect of boiling (for 30 min) on antioxidant activity of	
	African green leafy vegetables as found in this study	118
Table 3.2	Estimated flavonoid presence in different body compartments	132



LIST OF FIGURES

Figure 1.2.1	Photographs of African green leafy vegetables (a) Amaranthus cruentus
	L., (b) Corchorus olitorius L., (c) Cucurbita maxima Duchesne, and (d)
	Vigna unguiculata (L.) Walp
Figure 1.2.2	Chemical structures of common phenolic acids
Figure 1.2.3	Chemical structures of the flavonoid family
Figure 1.2.4	Types of tannins. 15
Figure 1.2.5	Structural groups responsible for radical scavenging
Figure 2.2.1	HPLC chromatograms of (a) standards and water extracts of (b) raw
	and (c) boiled amaranth
Figure 2.2.2	HPLC chromatograms of (a) standards and aqueous acetone extracts of
	(b) raw and (c) boiled amaranth
Figure 2.3.1	Protection of green leafy vegetable extracts against AAPH-induced
	damage on erythrocytes
Figure 2.3.2	Effect of green leafy vegetable extracts on oxidatively damaged pBR
	322 plasmid DNA
Figure 2.3.3	Protection of green leafy vegetable extracts against AAPH-induced
	damage on pBR 322 plasmid DNA
Figure 2.3.4	Effect of green leafy vegetable extracts on the proliferation of SC-1
	fibroblast cells as determined with (a) MTT, (b) neutral red, and (c)
	crystal violet assays
Figure 2.3.5	Effect of green leafy vegetable extracts on the viability of SC-1
	fibroblast cells, as determined with dichlorofluorescein assay
Figure 2.3.6	Effect of green leafy vegetable extracts on the viability of Caco-2 cells,
	as determined with dichlorofluorescein assay
Figure 2.3.7	Percentage damage of SC-1 fibroblast cells due to (a) treatment with
	both green leafy vegetable extracts and AAPH, and (b) AAPH only, as
	determined with the dichlorofluorescein assay
Figure 2.3.8	Percentage damage of Caco-2 cells due to (a) treatment with both green



	leafy vegetable extracts and AAPH, and (b) AAPH only, as determined	
	with the dichlorofluorescein assay	94
Figure 2.3.9	Percentage (a) total, (b) intra- and (c) extracellular protection of	
	green leafy vegetable extracts against AAPH-induced oxidative damage	
	on SC-1 fibroblast cells, as determined with the dichlorofluorescein	
	assay	96-97
Figure 2.3.10	Percentage (a) total, (b) intra- and (c) extracellular protection of green	
	leafy vegetable extracts against AAPH-induced oxidative damage on	
	Caco-2 cells, as determined with the dichlorofluorescein assay	97-98
Figure 3.1	Changes in the overall antioxidant activity due to different and	
	simultaneous events in a vegetable matrix subjected to heating	119
Figure 3.2	Chemical structures of the flavonoids detected in green leafy vegetable	
	extracts	122
Figure 3.3	Sequence of reactions involved in the lipid oxidation chain process in	
	the absence or presence of a flavonoid (FOH) acting as antioxidant	125
Figure 3.4	A schematic diagram illustrating the process involved in health-	
	promoting effects of African green leafy vegetables	130



GLOSSARY

AAPH 2,2'-azobis(2-amidinopropane) dihydrochloride

ABTS 2,2'-Azinobis-(3-ethyl-benzothiazoline-6-sulfonic acid) diammonium salt

ADME absorption, distribution, metabolism and excretion

ANOVA analysis of variance

approx approximately

ARC Agricultural Research Council

AUC area under the fluorescence curve

°C degree Celsius

C carbon

Caco-2 human adenocarcinoma colon cancer

CDL chronic diseases of lifestyle

CH₃O methoxyl

CHD coronary heart diseases

CO₂ carbon dioxide

Cu copper

CV crystal violet

CVD cardiovascular diseases

DCF dichlorofluorescein
DCFH dichlorofluorescin

DCFH-DA dichlorofluorescein diacetate

DMEM Dulbecco's modified eagle medium
DMPD *N,N*-dimethyl-*p*-phenylelendiamine

DMSO dimethyl sulfoxide

DNA deoxyribonucleic acid

DPPH 2,2-diphenyl-2-picrylhydrazyl

dw dry weight

EDTA ethylenediaminetetracetric acid

EGCG epigallocatechin gallate

EtBr ethidium bromide



FAO Food and Agriculture Organization of the United Nations

F-C Folin Ciocalteu
FCS Fetal calf serum

Fe iron
Fig Figure

fw fresh weight

FRAP ferric reducing antioxidant power

g gram

GAE gallic acid equivalents

GC/MS gas chromatography combined with mass spectrometry

GLVs green leafy vegetables
GSH glutathione peroxidase

H hydrogen h hour

H₂O₂ hydrogen peroxide

HAT hydrogen atom transfer

HCl hydrochloric acid

HIV/AIDS human immunovirus / acquired immune deficiency syndrome

HOBr hypobromous acid HOCl hypochlorous acid

HOO hydroperoxyl

HORAC hydroxyl radicals averting capacity

HPLC high performance liquid chromatography

HPLC/DAD/MS high performance liquid chromatography equipped with a diode array

detector and mass spectrophotometer

i.e. that is

kcal kilo calories

KCl potassium chloride

kg kilogram kJ kilo joules

K₂S₂O₈ potassium peroxodisulfate

L litre

LDH lactate dehydrogenase LDL low density lipoprotein

LPH lactase phloridzin hydrogenase

LSD least significant difference

M molar

mg milli gram
ml milli litre
min minutes
mM milli molar

MTT 3,(4,5-dimethyl thiazol-2-yl)2,5-diphenyl tetrazolium bromide

n number of flavonoid units

NaCl sodium chloride

Na₂EDTA ethylene diamine tetra acetic acid disodium salt dehydrate

NaH₂PO₄ sodium phosphate monobasic

Na₂HPO₄ di-sodium hydrogen orthophosphate dehydrate

n.d. not determinedn.d.a. no data available

nm nano mitre
NO nitric oxide

NO₂ nitrogen dioxide

NR neutral red

O oxygen

 O_2^- superoxide

OD optical density
OH hydroxyl ion

OH hydroxyl

ONOO peroxynitrite anion

ORAC oxygen radical absorption capacity

pBR plasmid Boliver and Rodrigues

PBS phosphate buffer solution

PCL photochemiluminescence

pH potential hydrogen RE retinol equivalents

RO alkoxyl ROO peroxyl

ROS reactive oxygen species

SACN Scientific Advisory Committee on Nutrition

SD standard deviation

SET single electron transfer
SEM standard error of means
SOD superoxide dismutase

sp. species

TAA total antioxidant activity

TE Trolox equivalents

TEAC Trolox equivalent antioxidant capacity

TFC total flavonoid content

TIFF tagged image file format

TPC total phenolic content

TRAP total radical-trapping antioxidant parameter

USA United States of America

USDA United States Department of Agriculture

UV ultra violet

WHO World Health Organization

 $\begin{array}{ll} \mu g & \text{micro gram} \\ \mu l & \text{micro litre} \\ \mu M & \text{micro molar} \\ \mu mol & \text{micro moles} \end{array}$