

THE EFFECT OF NON THERMAL 900 MHZ GSM IRRADIATION ON HUMAN SPERMATOZOA

by

NADIA FALZONE

Submitted in fulfilment of the requirements for the degree

PHILOSOPHIAE DOCTOR (REPRODUCTIVE BIOLOGY)

in the

DEPARTMENT OF OBSTETRICS AND GYNAECOLOGY FACULTY OF HEALTH SCIENCES UNIVERSITY OF PRETORIA

Supervisor: Dr. C. Huyser

Co-Supervisors: Profs. D. R. Franken and D. Leszczynski

April 2007



To Paolo Thank you for your support in reaching this goal.

"I can do everything through Him who gives me strength"
Phil 4: 13.



DECLARATION BY CANDIDATE

"I hereby declare that the thesis su	ubmitted for the degree Philosophiae
•	ia, is my own original work and has
not previously been submitted	to any other institution of higher
education. I further declare that all	sources cited or quoted are indicated
and acknowledged by means of a co	omprehensive list of references".
Name in Block letters	Signature
Date:	



THE EFFECT OF NON-THERMAL 900 MHz MOBILE PHONE RADIATION ON HUMAN SPERMATOZOA

by

NADIA FALZONE

Submitted in fulfilment of the requirements for the degree

PHILOSOPHIAE DOCTOR

in the

FACULTY OF HEALTH SCIENCES DEPARTMENT OF OBSTETRICS AND GYNAECOLOGY UNIVERSITY OF PRETORIA

Supervisor: Dr. C. Huyser

Co-Supervisors: Profs. D. R. Franken and D. Leszczynski

April 2007



ABSTRACT

Several studies have highlighted the possibility that radio-frequency electromagnetic fields (RF-EMF) used in mobile phone technology could influence DNA integrity of male germ cells as well as sperm motility. Current knowledge concerning the influence of RF-EMF on male germ cells is extremely limited. In the present study the hypothesis that 900 MHz GSM radiation could induce the activation of stress response in human spermatozoa was investigated.

Ejaculated, density purified, human spermatozoa from donors were exposed to 900 MHz GSM mobile phone radiation at specific absorption rate (SAR) levels of 2.0 and 5.7 W/kg and examined at various time points post exposure. Sperm motility and morphology were evaluated by computer-aided sperm analysis (CASA). The ability of RF-EMF exposed sperm to undergo the acrosome reaction was evaluated by flow cytometry. Sperm binding to the zona pellucida of human oocytes was determined by the hemi-zona (HZA) assay. Apoptotic markers, phosphatidylserine (PS) externalization, change in mitochondrial membrane potential ($\Delta \psi m$), reactive oxygen species (ROS) generation, caspase activation and DNA fragmentation were analysed using flow cytometry. Heat shock protein (Hsp) 27 and 70 expression and activity were analyzed using specific antibodies with flow cytometry and Western blot methods. Stress fibre stabilization (F-actin polymerization) was visualized using fluorescent dye labelled phalloidin.

No effect was seen on kinematic parameters assessed at SAR 2.0 W/kg, however straight line velocity (VSL) and beat cross frequency (BCF) were significantly altered after exposure at SAR 5.7 W/kg. Sperm shr inkage (decrease in surface area) was observed at both exposure levels. RF-EMF did not influence exposed spermatozoa's ability to undergo the acrosome reaction. A significant decrease in sperm-zona binding was observed at both exposure levels. RF radiation did not have an effect on any apoptotic markers. ROS generation increased significantly with an increase in SAR (5.7 W/kg). RF-EMF did not induce a stress response in exposed sperm (no activation of Hsp70 and 27 activity).



These results cannot be ascribed to heating, as the temperature did not increase by more than 0.2 - 0.3°C during exposure. The decrease in sperm-zona binding is the result of an alternative non-stress inducible pathway. This study should be replicated at lower SAR levels that would simulate the radiation absorption from carrying the cell phone in a pocket close to the testes.

KEY WORDS

human spermatozoa, mobile phone radiation, sperm functionality, stress response, apoptosis.



TABLE OF CONTENTS

	Pag	e number
ACKN	OWLEDGEMENTS	X
STRUC	CTURE AND SCOPE OF THE THESIS	xi
OBJEC	CTIVES OF THE STUDY	xiii
SUMM	1ARY	xiv
PUBLI	ICATIONS	xvi
LIST C	OF ABBREVIATIONS	xviii
LIST O	OF FIGURES	XX
LIST O	OF TABLES	xxxi
SECT	TON A: LITERATURE SEARCH	
CHAP	TER 1: BIOLOGICAL EFFECTS OF MOBILE	PHONE
	RADIATION	
1.1	INTRODUCTION	
1.2	RADIO-FREQUENCY FIELDS FROM MOBILE PHONES –	
	PHYSICS AND DOSIMETRY	
1.2.1	Modulation	
	` 1	
	TDMA (Time Division Multiple Access)	
1.2.1.3	CDMA (Code Division Multiple Access)	4
1.2.2	Cellular Phone Technologies	5
1.2.3	Output from Mobile Phones	5
1.2.4	RF radiation dose and measurement	6
	(i) Micro-antennas:	7
	(ii) Miniature thermal probes:	7
	(iii) Numerical modelling:	8
1.2.5	Biological basis for limiting exposure to mobile phones	8



1.3	BIOPHYSICAL INTERACTION OF RF-EMF WITH	
	BIOLOGICAL SYSTEMS	10
1.3.1	Biophysical Mechanisms	10
1.4	BIOLOGICAL EFFECTS OF RADIO-FREQUENCY FIELDS	
	FROM MOBILE PHONES	13
1.4.1	Evidence of biological effects of RF-EMF fields	13
1.4.2	Health risks associated with genotoxic effects from RF-EMF	
	exposure	14
1.4.3	Health risks associated with the induction of apoptosis as a result	
	of RF-EMF exposure	15
1.4.4	Health risks associated with Gene/Protein expression as a result of	
	RF-EMF exposure	24
1.4.5	Health risks associated with effects on male germ cells from RF-	
	EMF exposure	31
1.5	STRESS RESPONSE AS A POSSIBLE PATHWAY FOR RF-	
	EMFEXPOSURE	34
1.6	REFERENCES	35
СНАРТ	TER 2: MOLECULAR BASIS FOR CELLULAR STR OCCURRENCE IN HUMAN SPERMATOZOA IMPLICATIONS FOR MALE FERTILITY	
2.1	INTRODUCTION - GENERAL ASPECTS OF CELLULAR	
	STRESS	50
2.2	BIOCHEMICAL CHARACTERIZATION OF APOPTOSIS	51
2.2.1	The effectors of apoptosis	51
2.2.2	Extrinsic regulation of Apoptosis	
2.2.3	Intrinsic regulation of Apoptosis	
2.2.3.1	The role of mitochondria in apoptosis	
2.2.3.2	The endoplasmic reticulum regulation of apoptosis	56



2.2.4	The role of kinases in the regulation of apoptotic signal	
	transduction5	57
2.3	THE HEAT SHOCK RESPONSE: HEAT SHOCK PROTEINS	57
2.3.1	Heat shock protein families	58
2.3.2	Induction and regulation of the heat shock response	58
2.3.3	Hsps as mediators of apoptosis	51
2.3.3.1	Hsp regulation of the intrinsic apoptotic pathway	52
2.3.3.2	Hsp regulation of the extrinsic apoptotic pathway	54
2.3.3.3	Hsp regulation of the MAPK activated apoptotic pathways	54
2.4	CELLULAR STRESS IN HUMAN SPERMATOZOA	56
2.4.1	Heat shock protein expression during spermatogenesis	57
2.4.2	Heat shock proteins: presence and functionality in spermatozoa	58
2.4.2.1	Role of heat shock proteins in fertilization	70
2.4.2.2	Apoptosis in spermatozoa	71
2.4.2.3	The Fas mediated pathway in sperm cell apoptosis	72
2.4.2.4	Apoptosis promotion by caspase activation in spermatozoa	72
2.4.2.5	Association of mitochondrial membrane potential with sperm	
	apoptosis	73
2.4.2.6	Externalisation of phosphatedylserine as an indication of apoptosis	
	in spermatozoa	74
2.4.2.7	DNA fragmentation in spermatozoa as a consequence of apoptosis	75
2.5	REFERENCES	17



SECTION B: THE EFFECT OF NON-THERMAL 900 MHz GSM MOBILE PHONE RADIATION ON HUMAN SPERMATOZOA

CHAPTER 3: CAPACITATION & OOCYTE BINDING

3.1	INTRODUCTION - MOLECULAR BASIS FOR	
	CAPACITATION IN HUMAN SPERMATOZOA	99
3.1.1	Hyperactivated Motility	100
3.1.2	The Human Acrosome Reaction	101
3.1.2.1	Signal transduction between the zona pellucida and the	
	spermatozoon	101
3.2	RF-EMF EXPOSURE SYSTEM AND EXPERIMENTAL	
	PROTOCOL	105
3.2.1	Experimental set-up	105
3.2.2	Collection of semen	106
3.2.3	Density gradient purification and preparation of spermatozoa	107
3.3	CAPACITATION: ASSESSMENT OF THE HUMAN	
	MOTILITY AND THE ACROSOME REACTION	108
3.3.1	Capacitation: Assessment of motility	108
3.3.1.1	Morphometric assessment	109
3.3.2	Capacitation: Assessment of the acrosomal status	110
3.3.2.1	Viability probes used in the acrosome reaction	111
	(i) 7-Amino Actinomycin D	112
	(ii) Propidium Iodide	112
	(iii) Comparison between 7-AAD and PI as viability probes used	
	in the acrosome reaction	113
3.3.2.2	Evaluation of the acrosome reaction by flowcytometry	113
(i)	Visual assessment of the acrosome reaction.	114
(ii)	Induction of the acrosome reaction by calcium ionophore	115
3.4	HEMI-ZONA ASSAY	115
3.4.1	Zona Pellucida Binding - Mechanism:	115



3.4.2	Source and preparation of human zonae pellucidae	116
3.4.3	Sperm-oocyte interaction	117
3.5	STATISTICAL ANALYSIS	118
3.6	RESULTS	119
3.6.1	Motility and Morphology: Computer aided sperm analysis	119
3.6.1.1	Percentage progressive motility	119
3.6.1.2	Velocity parameters	120
3.6.1.3	Motion parameters	123
3.6.1.4	Morphometric analysis	123
3.6.2	Acrosome Reaction	126
3.6.2.1	Comparisson between 7-AAD and PI as viability probes for flow	
	cytometry post fixation and permiabilisation	127
3.6.2.2	Visual assessment of the acrosome reaction	127
3.6.2.3	Assessment of the acrosome reaction by flowcytometry	128
(i)	Evaluation of 7-AAD staining	128
(ii)	Induction of the acrosome reaction	129
(iii)	Evaluation of the acrosome reaction post RF-EMF	132
3.6.3	Sperm-oocyte interaction - Hemizona Assay	132
3.7	DISCUSSION	134
3.8	REFERENCES	139
	TER 4: APOPTOSIS	
4.1	INTRODUCTION	
4.2	EXPERIMENTAL PROTOCOL	152
4.3	ASSESSMENT OF THE APOPTOTIC STATUS IN	
	SPERMATOZOA	154
4.3.1	Phosphatedylserine externalisation determined by the Annexin V	
	assay	
4.3.1.1	Annexin V-FITC staining protocol	
	Annexin V blocking by recombinant Annexin V	
4.3.1.3	Induction of apoptosis by staurosporine	156



4.3.2	Mitochondrial Membrane Potential	157
4.3.2.1	MitoTracker® Red CMXRos staining procedure:	157
4.3.2.2	Abolishment of Δψm by Carbamoylcyanide m-	
	chlorophenylhydrazone	157
4.3.3	Detection of superoxide	157
4.3.3.1	Detection of O ₂ with hydroethidine	158
4.3.3.2	Determination of leukocyte contamination in processed	
	spermatozoa	159
4.3.4	Caspase-3 Activation.	159
4.3.4.1	Active caspase-3 PE staining protocol	160
4.3.4.2	Caspase inhibition by CaspACE TM FITC-VAD-FMK	160
4.3.5	DNA Fragmentation.	161
4.3.5.1	Fixation protocol for APO-Direct TM samples	161
4.3.5.2	APO-Direct TM staining protocol	162
4.3.5.3	Induction of DNA fragmentation by DNase	162
4.4	STATISTICAL ANALYSIS	163
4.5	RESULTS	163
4.5 4.5.1	Phosphatedylserine externalisation determined by the Annexin V	163
	Phosphatedylserine externalisation determined by the Annexin V assay	163
4.5.1	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V	163 163
4.5.1.1 4.5.1.1 4.5.1.2	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V	163 163
4.5.1.1 4.5.1.1 4.5.1.2	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V Induction of apoptosis by staurosporine	163 163 164
4.5.1.1 4.5.1.1 4.5.1.2	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V Induction of apoptosis by staurosporine The effect of 900 MHz GSM irradiation on phosphatedylserine	163 164 165
4.5.1.1 4.5.1.1 4.5.1.2 4.5.1.3	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V Induction of apoptosis by staurosporine The effect of 900 MHz GSM irradiation on phosphatedylserine externalisation	163 164 165 170
4.5.1.1 4.5.1.2 4.5.1.3 4.5.2	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V Induction of apoptosis by staurosporine The effect of 900 MHz GSM irradiation on phosphatedylserine externalisation Mitochondrial Membrane Potential	163 164 165 170
4.5.1.1 4.5.1.2 4.5.1.3 4.5.2 4.5.3	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V Induction of apoptosis by staurosporine The effect of 900 MHz GSM irradiation on phosphatedylserine externalisation Mitochondrial Membrane Potential Detection of ROS	163 164 165 170 172
4.5.1.1 4.5.1.2 4.5.1.3 4.5.2 4.5.3 4.5.3.1	Phosphatedylserine externalisation determined by the Annexin V assay	163 164 165 170 173
4.5.1.1 4.5.1.2 4.5.1.3 4.5.2 4.5.3 4.5.3.1 4.5.3.2	Phosphatedylserine externalisation determined by the Annexin V assay	163 164 165 170 173 173
4.5.1.1 4.5.1.2 4.5.1.3 4.5.2 4.5.3 4.5.3.1 4.5.3.2 4.5.4	Phosphatedylserine externalisation determined by the Annexin V assay	163 164 165 170 173 173 175
4.5.1.1 4.5.1.2 4.5.1.3 4.5.2 4.5.3 4.5.3.1 4.5.3.2 4.5.4.4	Phosphatedylserine externalisation determined by the Annexin V assay Annexin V blocking by recombinant Annexin V Induction of apoptosis by staurosporine The effect of 900 MHz GSM irradiation on phosphatedylserine externalisation Mitochondrial Membrane Potential Detection of ROS Detection of leukocyte contamination in processed spermatozoa Detection of O2 with hydroethidine Caspase Activation Detection of active caspase-3	163 164 165 170 173 173 175 175



4.6	DISCUSSION	182
4.7	REFERENCES	190
СНАРТ	TER 5: HEAT SHOCK PROTEIN & STRESS FI	IBRE
5.1	INTRODUCTION	199
5.2	EXPERIMENTAL PROTOCOL	201
5.3	DETERMINATION OF HEAT SHOCK PROTEIN	
	EXPRESSION AND PHOSPHORYLATION AFTER 900MHz	
	GSM RADIATION	203
5.3.1	Flow cytometric analysis of Hsp70 expression and Hsp27	
	expression and phosphorylation	203
5.3.2	Western Blot Analysis of Hsps 110, 90, 75, 70, 60, 40 and 27	
	expression	204
5.3.2.1	Protein extraction	204
5.3.2.2	Electrophoresis	205
5.3.2.3	Blotting	205
5.3.2.4	Western blot analysis of Hsp70 and Hsp27 in EA.hy926 cells	205
5.4	PHYSIOLOGICAL EFFECTS OF HSP ACTIVATION	206
5.4.1	Detection of stress fibres in human spermatozoa	206
5.4.2	Detection of stress fibers in MCF-7 cells	207
5.5	STATISTICAL ANALYSIS	207
5.6	RESULTS	207
5.6.1	Flow cytometric analysis and visualisation of Hsp70 and 27	
	expression and phosphorylation after 900MHz GSM	207
5.6.1.1	Flow cytometric analysis of Hsp27 expression and	
	phosphorylation	207
5.6.1.2	Flow cytometric analysis of Hsp70 expression	208
5.6.2	Western blot analysis of Hsp110, 90, 70, 60, 40 and 27 expression	
	after 900MHz GSM	211



5.6.2.1	Western blot analysis of Hsp27	211
5.6.2.2	Western blot analysis of Hsp70	211
5.6.2.3	Western blot analysis of Hsp110, 90, 75, 60 and 40	214
5.6.3	Detection of stress fibres	217
5.6.3.1	F-actin polymerisation in RF-EMF exposed human spermatozoa	217
5.6.3.2	F-actin polymerisation in RF-EMF exposed MCF-7 cells	219
5.7	DISCUSSION	220
5.8	REFERENCES	224
SECT	ION C: CONCLUSIONS	
СНАР	TER 6: CONCLUSIONS AND RECOMMENDATIONS	
6.1	CONCLUSIONS AND RECOMMENDATIONS	233
6.2	DEFEDENCES	240
0.2	REFERENCES	240
SECT	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS	240
SECT	ION D: ANNEXURES	
SECT	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS	244
SECT ANNEX	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER	 244 244
SECT ANNEX A.1 A.1.1	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER Temperature control unit	 244 244 246
SECT ANNEX A.1 A.1.1 A.1.2	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER Temperature control unit	244 244 246
ANNEX A.1 A.1.1 A.1.2 A.1.3	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER Temperature control unit Signal generator GSM-Modulator	244 246 246 247
ANNEX A.1 A.1.1 A.1.2 A.1.3 A.1.4	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER Temperature control unit Signal generator GSM-Modulator RF-power amplifier	244 246 246 247
ANNEX A.1 A.1.1 A.1.2 A.1.3 A.1.4 A.1.5	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER Temperature control unit Signal generator GSM-Modulator RF-power amplifier. Circulator and coaxial termination.	244 246 246 247 247
ANNEX A.1 A.1.1 A.1.2 A.1.3 A.1.4 A.1.5 A.1.6	ION D: ANNEXURES XURE A: SAR SIMULATION RESULTS VERTICAL 900MHZ EXPOSURE CHAMBER	244 246 247 247 247



A.2.3	Ther	modynamic simulations249
A.3	MA(GNETIC FIELD MEASUREMENTS252
A.4	REF	ERENCES254
ANNEX	KURE	B: MACROSCOPIC AND MICROSCOPIC SPERM
		PARAMETERS
B.1	FVA	LUATION OF SPERMATOZOA255
B.1.1		en Parameters – Macroscopic and microscopic evaluation
B.1.1.1		oscopic analysis
D.1.1.1		•
	(i) (ii)	Appearance
	(iii)	
	, ,	Colour and odour
	(iv)	·
	(v)	Volume
D 1 1 2	(vi)	pH
B.1.1.2		oscopic analysis
	(i)	Sperm agglutination - SpermMar IgG test
	(ii)	Non-sperm cellular elements
	(iii)	Concentration - Neubauer haemocytometer
	(iv)	Motility - Differential count
	(v)	Morphology (Papanicolaou stain using Tygerberg strict
D 4	DEE	criteria)
B.2	KEF	ERENCES
ANNE	ZTIDE	C: DONOR SPERM PARAMETERS - RESULTS
PATATATE 2	LUKE	C. DUNUK SI EKWI I AKAMETEKS - KESULIS
C.1	MAG	CROSCOPIC SEMEN PARAMETERS266
C.2	MIC	ROSCOPIC SEMEN PARAMETERS266
C.3	REF	ERENCES269



ACKNOWLEDGEMENTS

My sincere gratitude and appreciation to the following persons and/or institutions for their contribution to this thesis:

- Dr. C. Huyser, Prof. D. Franken and Prof. D. Leszczynski, for your un-abating support, guidance and patience throughout this research project and the preparation of the manuscript.
- SABS for funding, and in particular, Dr. F. le Roux Fourie for his support in orchestrating the loan of the research equipment from Finland.
- The National Research Foundation for providing funding for this project.
- Prof. D. du Toit (Department of Biomedical Sciences, Tshwane University of Technology) for support and relief from duties.
- Prof. P. Becker (Biostatistics Unit, Medical Research Council), for his support and assistance in the statistical preparation of the data.
- Mr. T. Toivo and Prof. K. Jokela (NIR Laboratory, STUK, Helsinki, Finland) for the dosimetric calculations.
- Mrs. H. Tammio (Radiation Biology Laboratory, STUK, Helsinki, Finland) for Western blot analysis.
- Prof. P. Cilliers (Hermanus Magnetic Observatory) for the magnetic field evaluations.
- The Department of Obstetrics and Gynaecology for support, use of facilities and equipment.
- Mr. M. Buchwald for editing and technical support in the preparation of the manuscript.
- My family and friends for continued interest and support during this study.



STRUCTURE AND SCOPE OF THE THESIS

SECTION A: REVIEW OF LITERATURE

Chapter 1 provides an overview of the operating principles of mobile phones, in addition the biological mechanism/s of radio-frequency electromagnetic fields (RF-EMF) are discussed, and scientific evidence is presented of *in-vitro* biological effects. Based on evidence provided that RF-EMF could interact with biological systems by inducing a stress response, a review on current literature elucidating the stress response phenomenon was conducted. Chapter 2 therefore addresses the molecular basis for cellular stress in somatic cells. The occurrence of a stress pathway in human spermatozoa and the implications thereof for male fertility were considered.

SECTION B: THE EFFECT OF NON-THERMAL 900 MHz GSM MOBILE PHONE RADIATION ON HUMAN SPERMATOZOA

In **Chapters 3, 4** and **5** the effect of 900 MHz mobile phone radiation on human spermatozoa was evaluated at two specific absorption rates (SAR) of 2.0 and 5.7 W/kg. RF-EMF exposure of spermatozoa was firstly assessed using sperm specific assays (sperm capacitation and sperm-zona binding – **Chapter 3**). Based on these findings the hypothesis that a stress pathway as a result of RF-EMF insult is operational in human spermatozoa was tested. The induction of apoptosis (**Chapter 4**) and activation of heat shock proteins (**Chapter 5**) after RF-EMF were evaluated.



SECTION C: CONCLUSIONS

In **Chapter 6** a summary of the findings of this study are given, conclusions are drawn, and recommendations for future research are provided.

SECTION D: ANNEXURES

Annexure A addresses the characterisation of the vertical RF chamber used in this research. Dosimetric analysis results are also summarised. Dosimetric evaluations were conducted at STUK, Radiation and Nuclear Safety Authority, Helsinki, Finland **Annexure B** contains a summary of the macroscopic and microscopic sperm parameters used in the evaluation of the sperm donors.

Annexure C summarises the results of the macroscopic and microscopic sperm assessment of each donor.



OBJECTIVES OF THE STUDY

Human semen parameters could serve as valuable indicators of toxic and genotoxic effects of occupational and environmental factors. Furthermore, spermatozoa are terminally differentiated cells that are unable to repair DNA damage, which make sperm an extremely sensitive model to use in the investigation of the effect of environmental stressors such as RF-EMF.

To determine:

- 1. if and by which mechanism RF radiation from mobile phone emissions affect human spermatozoa and what implication these findings have on male fertility,
- 2. the effect of RF exposure on sperm capacitation and sperm-zona binding,
- 3. if a stress response is operational in spermatozoa as a result of RF-EMF, by investigating;
 - i. the induction of apoptosis,
 - ii. heat shock protein phosphorylation and expression,
- 4. the suitability of using human spermatozoa as a reproductive model to indicate effects/influences of mobile phone radiation.



SUMMARY

Several studies have highlighted the possibility that radio-frequency electromagnetic fields (RF-EMF) used in mobile phone technology could influence DNA integrity of male germ cells as well as sperm motility. Current knowledge concerning the influence of RF-EMF on male germ cells is extremely limited. The main objective of this research was directed at determining the effect of non-thermal 900 MHz GSM modulated RF-exposure on human sperm fecundity by assessing sperm specific functions, sperm functionality and induction of a stress response.

Ejaculated, density purified, human spermatozoa obtained from donors were exposed to RF-EMF at two specific absorption rate levels (SAR 2.0 and 5.7 W/kg) and examined at various time points post exposure. To determine the influence of RF exposure on sperm specific functions, sperm propensity for acrosomal exocytosis was assessed using a new technique developed to evaluate the acrosome reaction (AR) by flow cytometry. Sperm motility and morphometry were determined by computer aided sperm analysis (CASA) and sperm binding potential was evaluated by the hemi-zona assay (HZA). 900 MHz GSM exposure had no effect on the AR however some motility parameters (straight line velocity and beat cross frequency) were significantly altered. Sperm surface area and acrosomal region were also significantly reduced as a result of RF-EMF. The ability of RF-exposed sperm to bind the human oocyte evaluated with the HZA was significantly impeded.

Sperm functionality was assessed using flow cytometry; (i) the percentage of Annexin-V positive and, propidium iodide (PI) negative spermatozoa, (ii) the change in spermatozoa's mitochondrial membrane potential ($\Delta \psi m$), (iii) caspase activation, (iv) the percentage of TUNEL (terminal deoxynucleotidyl transferase-mediated dUTP nick end labelling) positive spermatozoa and (v) generation of reactive oxygen species (ROS) were examined. No effect on any of the examined parameters after 900 MHz GSM exposure at either SAR level was noted. These results show that mobile phone radiation does not induce apoptosis.



The ability of 900 MHz GSM radiation to induce a stress response was evaluated by heat shock protein (Hsp) activation. Hsp27 and Hsp70 expression as well as activity were analyzed using specific antibodies with flow cytometry and Western Blot methods. Stress fibre stabilisation (F-actin polymerization) was visualized using fluorescent dye labelled phalloidin. RF-EMF had no effect on Hsp27 expression and phosphorylation nor Hsp70 expression as determined by flow cytometry and Western blot analysis. Visual assessment of stress fibre stabilization after RF exposure in sperm cells did not show any increased F-actin accumulation in the cells. RF-EMF exposure did not induce an Hsp27- or Hsp70-dependent stress response in human spermatozoa.

The effect of RF-EMF on sperm-zona binding and cell shrinkage observed at exposures of 2.0 and 5.7 W/kg, seems to be the result of an alternative non-Hsp dependent mechanism. Additional studies investigating the effect of RF-EMF on sperm-zona binding should be conducted, specifically exploring the ligand-receptor effector systems involved in sperm-zona binding. It is also suggested that electron microscopy be used to investigate conformal and structural changes as a result of RF-EMF. Considering recent reports noting an effect on sperm motility at lower SAR levels than that employed in the present study, the effect of RF-EMF on human spermatozoa motility, using the expanded analysis criteria set in this study, should be replicated at lower SAR levels that would simulate the radiation absorption from carrying the cell phone in a pocket close to the testes.



PUBLICATIONS

Submitted for publication:

- 1) Falzone, N., Huyser, C., Fourie, F le R., Leszczynski, D., Franken, D.R. *In vitro* effect of 900 MHz GSM radiation on mitochondrial membrane potential and motility of human spermatozoa. *Bioelectromagnetics*., submitted February 2007. (Approved for publication April 2007).
- 2) Falzone, N., Huyser, C., Becker, P., Fourie, F le R., Leszczynski, D., Franken, D.R. The effect of non-thermal 900 MHz GSM mobile phone radiation on the acrosome reaction, head morphometry and zona binding of human spermatozoa. *Hum Reprod.*, submitted July 2007.

In preparation:

- Falzone, N., Huyser, C., Franken, D.R. Leszczynski, D. Lack of activation of Hsp27- and Hsp70-dependent stress response in human spermatozoa exposed to 900 MHz GSM radiation. *Bioelectromagnetics*., submittion September 2007.
- 2) Falzone, N., Huyser, C., Fourie, F le R., Cockeran, R., Leszczynski, D., Franken, D.R. *In vitro* exposure of human spermatozoa to 900 MHz GSM radiation: effect on apoptosis. *Rad Res.*, submittion November 2007.
- 3) Falzone, N., Huyser, C., Cockeran, R., Becker, P., Franken, D.R. Significance of somatic apoptosis markers in human spermatozoa.

Conference papers:

- Falzone, N., Huyser, C., Fourie, F le R., Franken, D.R., Leszczynski, D., 2005.
 Pilot study: Effects of 900 MHz GSM radiation on human sperm function.
 Annual Bioelectromagnetics Society meeting, Dublin, Ireland, June, 2005.
- 2) Falzone, N., Huyser, C., Fourie, F le R., Franken, D.R., Leszczynski, D., 2006. Effect of 900 mhz gsm radiation on apoptotic status and functionality of human spermatozoa. Annual Bioelectromagnetics Society meeting, Cancun, Mexico, June, 2006.



- 3) Falzone, N., Huyser, C., Fourie, F le R., Leszczynski, D., Franken, D.R., 2007. Effect of 900 MHz GSM radiation on human sperm functionality. IFFS conference, Durban, South Africa, May, 2007.
- 4) Falzone, N., Huyser, C., Fourie, F le R., Franken, D.R., Leszczynski, D., 2007. Lack of activation of Hsp27- and Hsp70-dependent stress response in human spermatozoa exposed to 900 MHz GSM radiation. Annual Bioelectromagnetics Society meeting, Kanazawa, Japan, June, 2007.



LIST OF ABBREVIATIONS

7-AAD - 7-amino-actinomycin D

? $\psi_{\rm m}$ - Change in the mitochondria membrane potential

aCp - Active caspase

AIF - Apoptosis inducing factor

ALH - Amplitude of lateral head displacement

AR - Acrosome reaction

ART - Artificial reproductive technologies

ATP - Adenosine triphosphate

BCF - Beat cross frequency

BSA - Bovine serum albumin

CASA - Computer aided sperm analysis

CDMA - Code Division Multiple Access

Cp - Caspase

CW - Continuous wave

DNA - Deoxyribonucleic acid

DISC - Death-inducing signalling complex

DPBS - Dulbecco's phosphate buffered saline

DMSO - Dimethyl sulfoxide

ELF - Extremely low frequencies

EM - Electromagnetic

EMF - Electromagnetic field

ER - Endoplasmic reticulum

ERK - Extracellular regulated kinases

FCM - Flow cytometry

FDMA - Frequency Division Multiple Access

FDTD - Finite Difference Time Domain

FITC - Fluorescein isothiocyanate

HSE - Heat shock element

HSF - Heat shock transcriptional factor

Hsc - Heat shock cognate protein



Hsp - Heat shock protein

HYPA - Hyper activated motility

HZA - Hemi zona assay

IVF - *In vitro* fertilization

kD - kilo Dalton

MAPK - Mitogen activated protein kinases

MMP - Mitochondrial membrane potential

MOMP - Mitochondrial outer membrane permiabilisation

PBS - Phosphate buffered saline

PCD - Programmed cell death

pCp - Pro-caspase

PBS - Phosphate buffered saline

PS - Phosphatidylserine

PI - Propidium iodide

PSA - Pisum sativum agglutinin

PTPC - Permeability pore complex

RF - Radio frequency

RF-EMF - Radio frequency electromagnetic field

ROS - Reactive oxygen species

SAR - Specific absorption rate

SD - Standard deviation

STR - Straightness

TDMA - Time Division Multiple Access

TUNEL - Terminal deoxynucleotidyl transferase-mediated dUTP nick end

labelling

VAP - Average path velocity

VCL - Curvilinear velocity

VSL - Straight line velocity

ZP - Zona pellucida



LIST OF FIGURES

Page number

Chapter 1
Figure 1.1 Triage of the Electromagnetic spectrum indicating the position
of mobile phone emissions (Adapted with permission from the EU
Commission, Health and Electromagnetic fields, 2005)
Chapter 2
Figure 2.1 The stress response: Exposure to damaging stimuli can trigger
a cellular stress response resulting either in recovery or activation of the
apoptotic program. Severe exposure could initiate cellular necrosis
Figure 2.2 Extrinsic Apoptotic Pathway (Danial and Korsmeyer, 2004 -
with permission). Binding of the various trimeric ligands to death receptors,
(A) TNF to TNRF-1, (B) FasL to Fas(APO-1/CD95) and (C)
APO2L/TRAIL to DR4/5 triggers the downstream assembly of the DISC 53
Figure 2.3 Intrinsic Apoptotic Pathway (Danial and Korsmeyer, 2004 –
with permission). BH3 protein activation lead to Bax, Bak activation
resulting either in the assembly of the apoptosome or ER induced apoptosis 55
Figure 2.4. Major transcription factors leading to the induction of Hsp
synthesis: (1) cytoplasmic complex of HSF-1 and Hsp90; (2) HSF-1
translocation to the nucleus; (3) intra-nuclear distribution of HSF-1; (4)
nuclear complex of HSF-1 and Hsp90; (5) retro translocation of HSF-1 to
the cytoplasm. (Adapted from Sõti et al., 2005).
Figure 2.5 Regulation of the intrinsic apoptotic pathway by Hsps (with
permission - Beere 2005); (A) inhibiting BAX translocation to the
mitochondrion, (B) suppression of AIF activity, (C) inhibiting the activation



of pro-caspase-9 by the apoptosome, (D) sequestering cytocrome-c release
as a result of MOMP, (E) inhibiting Akt/BAD activation of MOMP63
Figure 2.6 Regulation of the extrinsic apoptotic pathway by Hsps (with
permission - Beere, 2005). Hsps regulate both death receptor signalling
(Hsp27 - JNK and Bax pathway; Hsp70 - Bid pathway) and cell survival
pathways (Hsp90 - NF-κB pathway)65
Chapter 3
Figure 3.1. The human fertilisation process (Adapted from "fertilization."
Encyclopædia Britannica Online, 2007): (1) Chemoattraction, (2) specific
recognition – loose association, (3) acrosomal exocytosis, (4) penetration –
sperm-egg binding, (5) membrane fusion, (6) sperm invagination
Figure 3.2. Front view and set-up of RF-EMF exposure chamber. Two
glass Petri dishes are placed inside the chamber on top of a temperature
regulated waterbed. The RF-EMF signal is fed into the chamber placed
inside a CO ₂ incubator via a monopole type feed post
Figure 3.3 RF-EMF exposure protocol: Assessment of the acrosome
reaction and motility characteristics post RF EMF and control exposure
Figure 3.4 Metrix calculation of sperm morphometric parameters: (A) an
abnormal spermatozoon, (B) a normal spermatozoon
Figure 3.5 Flow cytometric assessment of 7-AAD and PI as viability
probes after fixation and permeabilization of human spermatozoa
Figure 3.6 The hemi-zona assay: (1) Oocytes from IVF were bisected and
kept at room temperature while (2) spermatozoa were exposed for 1 hour to
RF-EMF (SAR 2.0 and 5.7 W/kg). (3) After exposure hemi-zonae were
added to 50 ul spermatozoa droplets (0.5 x 10 ⁶ cells/ml). (4) Spermatozoa



assessment of binding	16
Figure 3.7 Velocity parameters comparing RF-EMF (dark-grey) exposed spermatozoa for SAR 2.0 W/kg (A) and 5.7 W/kg (B) with controls (light-grey) directly (T_1) , 3 (T_2) , and 24 (T_3) hours after a 1 hour 900 MHz GSM exposure (*p < 0.05).	22
Figure 3.8 Motion parameters comparing RF-EMF (dark grey) exposed spermatozoa for SAR 2.0 W/kg (A) and 5.7 W/kg (B) with controls (light grey) directly (T ₁), 3 (T ₂), and 24 (T ₃) hours after a 1 hour 900 MHz GSM exposure (*p<0.05).	24
Figure 3.9 (A) Linear regression of log (mean fluorescence) _{7-AAD} ($R^2 = 0.93$) and log (mean fluorescence) _{PI} ($R^2 = 0.83$). (B) Linear regression of log (%gated) _{7-AAD} fluorescence ($R^2 = 0.93$) and log (%gated) _{PI} fluorescence ($R^2 = 0.97$).	27
Figure 3.10 Fluorescent staining (PSA-FITC) of the human acrosome reaction: (A) Acrosomal cap intact, (B) Patchy fluorescence, (C) Equatorial staining and (D) Acrosome reacted.	28
Figure 3.11 7-AAD Viability staining - Flow cytometric histograms showing overlay plots of unstained sperm (red-brown), sperm labelled with 10μl 7-AAD (blue) and sperm treated with 2 μl of Triton x-100 (green) for 15 min. prior to 7-AAD staining.	29
Figure 3.12 PSA-FITC staining - Flow cytometric histograms showing overlay plots of capacitated sperm (24 h) induced to undergo the acrosome reaction by pre-incubation with A23187 (positive control-red-brown) and sperm treated with DMSO (negative control for calcium ionophore	



stimulation-green) prior to PSA-FITC staining. Fluorescence peaks note the
acrosome reacted (AR) and acrosome intact (AI) sperm populations
Figure 3.13 PSA-FITC staining - (A) Flow cytometric histograms showing
overlay plots of capacitated sperm, sperm induced to undergo the acrosome
reaction by pre-incubation with A23187 (positive control-red-brown); sperm
incubated for 3 hours (orange) and sperm incubated for 24 hours (green)
prior to PSA-FITC staining. Gating shows acrosome reacted (AR) and
acrosome intact (AI) spermatozoa. (B) Flowcytometric dot plots and
projections showing gating of PSA-FITC staining in 7-AAD (live cells)
only, region F denotes acrosome intact spermatozoa and region H acrosome
reacted sperm. 131
Figure 3.14 Comparison between RF-EMF and control sperm assessed by
FCM directly, 3 - and 24 - hours after a 1 hour 900 MHz GSM exposure
(SAR 2.0 W/kg) (A) % 7-AAD ⁺ staining spermatozoa, (B) % acrosome
intact live cells, (C) % acrosome reacted live
Figure 3.15 Box and whiskers plot (showing medians) of number of sperm
binding non-fertilized metaphase II oocytes (5 oocytes per donor, n = 10)
after an hour exposure to 900 MHz GSM radiation. RF-EMF exposure
caused a significant reduction in sperm bound to the hemi-zonae compared
to controls (*p = 0.02)
Figure 3.16 Correlation of morphology and number of spermatozoa
binding the hemi-zona after an hour exposure to 900 MHz GSM radiation.
Number of control spermatozoa is also shown for comparison
1
Chapter 4
Figure 4.1 RF-EMF exposure protocol: Assessment of the apoptotic status
post RF-EMF and control exposure
=



Figure 4.2 Flow cytometric histograms of total Annexin V before (red)
and after (blue) addition of recombinant Annexin V- demonstrating the
specificity of Annexin V staining
Figure 4.3 The percentage PI, Annexin V and Annexin V ⁺ /PI sperm after
a 2 hour exposure to 1 μM , 5 μM and 10 μM staurosporine (STS), each
datum represents the mean \pm SD of three determinations
Figure 4.4 Dot plot and fluorescence histograms of (A) Annexin V and
(B) PI staining of human spermatozoa
Figure 4.5 In A_1 (2.0 W/kg) and B_1 (5.7 W/kg) the total percentage
Annexin V staining is noted as a function of time. A_2 (2.0 W/kg) and B_2 (5.7
W/kg) depict the total percentage of non-viable cells (PI+) as a function of
time. A_3 (2.0 W/kg) and B_3 (5.7 W/kg) show the total percentage Annexin
V ⁺ viable cells (PΓ) as a function of time
Figure 4.6 In Figures A_1 (2.0 W/kg) and B_1 (5.7 W/kg) the total number
of apoptotic necrotic (Annexin V ⁺ PI ⁺) staining is noted as a function of
time. A_2 (2.0 W/kg) and B_2 (5.7 W/kg) depict the total percentage of dead
cells (Annexin V ⁻ PI ⁺) as a function of time
Figure 4.7 Cytofluorometric analysis of the depolarisation of the
mitochondrial membrane potential showing a frequency histogram of
processed spermatozoa (blue) stained with, MitoTracker® Red CMXRos
before treatment (green) with the mitochondrial membrane potential
abolisher mClCCP (red-brown)
Figure 4.8 The percentage of MitoTracker® Red CMXRos (150 nM)
staining in RF-EMF exposed sperm at a SAR of (A) 2.0 W/kg and (B) 5.7
W/kg compared to control samples determined as a function of time



Figure 4.9 Cytofluorometric dot plot showing (C) gated sperm
population, (D) lymphocytes, (E) monocytes and (F) granulocytes in (A) a
processed sperm population and (B) a processed sperm population spiked
with 2 x 10 ⁶ white blood cells.
Figure 4.10 Cytofluorometric analysis of O. production in human
Figure 4.10 Cytofluorometric analysis of O_2^- production in human
spermatozoa using hydroethidine (HE). Frequency histogram notes the increased production of ethidium (E ⁺) due to superoxide oxidation in
processed spermatozoa incubated in the presence of 2 x 10 ⁶ /ml white blood
cells (blue) compared to normal sperm (green)
Figure 4.11 The percentage of hydroethidine staining in sperm exposed to
RF-EMF at (A) SAR 2.0 W/kg and (B) SAR 5.7 W/kg compared to control
samples determined directly after exposure (T_1) , 3 hours after exposure (T_2)
and 24 hours after exposure (T ₃)
and 24 nours after exposure (13).
Figure 4.12 The percentage of cells staining positive for activated caspase-
3 determined for control and RF-EMF (SAR 2.0 W/kg) exposed cells
detected at T_1 (directly after exposure), T_2 (3 hours after exposure) and T_3
(24 hours after exposure)
Figure 4.13. Cytofluorometric analysis of the frequency histogram of
processed spermatozoa (red-brown) stained with, FITC-VAD-FMK directly
after RF-EMF exposure (blue), 2 hours after exposure (green) and 24 hours
after exposure (orange)
Figure 4.14 The percentage of cells staining positive for FITC-VAD-FMK
determined for control and RF-EMF (A) SAR 2.0 W/kg and (B) SAR 5.7
W/kg exposed cells detected at T_1 (directly after exposure), T_2 (3 hours after
exposure) and T ₃ (24 hours after exposure)



Figure 4.15. Cytofluorometric analysis of the frequency histogram of
processed unstained spermatozoa (red-brown), before TUNEL staining
(green) and induction of DNA damage using DNAse (blue)
Figure 4.16 The percentage of cells staining positive for TUNEL
determined for control and RF-EMF (A) SAR 2.0 W/kg and (B) SAR 5.7
W/kg exposed cells detected at T_1 (directly after exposure), T_2 (3 hours after
exposure) and T ₃ (24 hours after exposure)
Chapter 5
Figure 5.1 RF-EMF exposure protocol. Assessment of cellular stress in
human spermatozoa post RF-EMF exposure and heat shock at 43°C
Figure 5.2 (A) Flow cytometric analysis of Hsp27 phosphorylation (dark
grey) detected by anti-Hsp27P directly (T_1) , 3 (T_2) and 24 hours (T_3) after
an hour RF-EMF exposure at SAR 2.0 W/kg (n = 12). Baseline Hsp27
expression (white) as well as Hsp27 phosphorylation after a 1 hour heat
shock at 43°C (black and white) are given at time 1. Control (light grey)
samples were maintained at 37°C during the exposures. (B) Detection of
Hsp27P expression by immunofluorescence staining directly after exposure
(T ₁), in (I) control and (II) RF-exposed sperm, Hsp27P fluorescence was
mainly located in the neck area of the sperm
Figure 5.3 (A) Flow cytometric analysis of Hsp70 expression detected
directly (T ₁), 3 (T ₂) and 24 hours (T ₃) after an 1 hour RF-EMF exposure at
SAR 2.0 W/kg or heat shock at 43°C (n = 12). Control samples were
maintained at 37°C during the exposures. (B) Detection of Hsp70
immunofluorescence staining directly after exposure (T1), in (I) control and
(II) RF-exposed sperm, Hsp70 fluorescence was mainly located in the neck
area of the sperm. 210



rigure 5.4 Western Diot analysis of risp27: (A) Autoradiogram of 5D5-	
PAGE resolved proteins for donor 2 (I), donor 12 (II) and EA.hy926 cells	
(III). The position of the bands correspond to heat shock proteins of Mr	
27kDa, specific for Hsp27. (B) Densitometric analysis of Hsp27	
phosphorylation status in spermatozoa directly after a 1 hour exposure to	
RF-EMF at SAR 2.0 W/kg or 43°C. Control samples were maintained at	
37°C for the duration of the exposure. As a control Hsp27 expression in	
EA.hy926 cells (III) after heat shock at 43°C is also noted	212
Figure 5.5 Western blot analysis of Hsp70: (A) Autoradiogram of SDS-	
PAGE resolved proteins for donor 2 (D ₂ - I), donor 12 (D ₁₂ - II) and	
EA.hy926 cells (III). The position of the bands correspond to heat shock	
proteins of M ₁ 70kDa, specific for Hsp70. (B) Densitometric analysis of	
Hsp70 expression in spermatozoa directly after an hour exposure to RF-	
EMF at SAR 2 W/kg or 43°C. Control samples were maintained at 37°C for	
the duration of the exposure. As a control Hsp70 expression in EA.hy926	
cells (III) after heat shock at 43°C is also noted.	213
Figure 5.6 Western blot analysis of Hsps 110, 75 and 60: (A)	
Autoradiogram of SDS-PAGE resolved proteins for donor 2 (D2- I), and	
donor 12 (D12 - II). The position of the bands corresponds to heat shock	
proteins of Mr 110kDa, 75kDa and 60kDa specific for Hsp110, Hsp75 and	
Hsp60 respectively. (B) Densitometric analysis of Hsp110, Hsp75 and	
Hsp60 expression in spermatozoa directly after an hour exposure to RF-	
EMF at SAR 2 W/kg or 43°C. Control samples were maintained at 37°C for	
the duration of the exposure.	215
Figure 5.7 Western blot analysis of Hsps 90 and 40: (A) Autoradiogram	
of SDS-PAGE resolved proteins for donor 2 (D ₂ - I), and donor 12 (D ₁₂ - II).	
The position of the bands corresponds to heat shock proteins of M _r 90kDa,	
and 40kDa specific for Hsps 90 and 40. (B) Densitometric analysis of Hsps	
90 and 40 expression in spermatozoa directly after a 1 hour exposure to RF-	



EMF at SAR 2 W/kg of 45°C. Control samples were maintained at 37°C for	
the duration of the exposure.	216
Figure 5.8 Immunolocalization of F-actin. (A) AlexaFluor-labelled	
phalloidin stained spermatozoa predominantly in the acrosome (white	
arrow), post-acrosomal area (yellow arrow), as well as neck and principal	
tail-piece areas (red arrow). (B) Cellular response of spermatozoa exposed	
to RF-EMF, control cells were maintained at 37°C for the duration of the	
exposure. (C) Typical staining of AlexaFluor-labelled phalloidin	218
Chapter 6	
Figure 6.1 The processes of homologous sperm-zona binding and	
penetration.	237
Annexure A	
Figure A.1 (A) Front view of vertical exposure chamber showing the	
position of the petri-dishes with the hatch open. (B) Side view of the	
chamber with the water pump housing (I), water is circulated from the	
waterbed below the perti-dishes to the aluminium cooling plate (II) behind	
the chamber (thermal conducting tape attached to back of plate). The	
cooling plate is placed in contact with the inside wall of the incubator to	
allow for thermal exchange. (C) Signal generator (III), Amplifier (IV), GSM	
modulator (V), Coaxial terminator (VI), Power meter and sensor (VII), DC	
power supply (VIII).	245
Figure A.2 Schematic drawing of the irradiation chamber. The petri-dishes	
(diameter 54 mm) are placed in special 'cups' moulded into the epoxy	
laminate above the cooling water so that the medium (3 ml) is at same level	
than the cooling water. The water is covered with a 0.8 mm thick epoxy	• • •
laminate.	246
Figure A.3 Schematic diagram of temperature and electric field	
measurement set-up. In upper part is shown the RF-power generation and	



measurement part and below that the electric field and temperature	
measurement parts.	248
Figure A.4 Typical SAR measurements based on temperature increase using a Vitek type sensor. The SAR is evaluated from linearized temperature increase (dT) between the 1 to 8 seconds time (dt) after power	
on.	250
Figure A.5 Measured and simulated E-field in chamber. Measurements (blue diamonds) were made in air using the SPEAG ET3DV6 probe	251
Figure A.6 Simulated (XFDTD) relative SAR distribution shown in one petri-dish placed in the vertical chamber. The scale is given from 0 mW/g (black) to 15 mW/g (yellow). The inner diameter of Petri dish is 50 mm	251
Figure A.7 Temperature increase measurements. The thermistor was either placed in the middle of the field or in contact with the Petri dish wall	252
Figure A.8 Magnetometer measurement set-up. Placement of the magnetometer inside the CO ₂ incubator in positions 2 and 3 closely resembled that of the earth's magnetic field.	253
Annexure B Figure B.1 Diagram illustrating different semen assessment parameters, functional tests and bioassays in the evaluation of human spermatozoa.	255
(Adapted with permission from Oehninger <i>et al.</i> , 1991)	233
Figure C.1 Sperm parameters determined for each donor. Each end point	
represents the mean and standard deviation (SD) of at least 5 replicates. In	
all graphs WHO reference values are noted with dotted lines. (A) Total	
volume; (B) pH; (C1) Concentration pre-processing; (C2) Concentration	
post-processing; (D1) Morphology pre-processing; (D2) Morphology post-	
processing and (E) Forwards motility, of each semen sample.	267



LIST OF TABLES

Chapter 1	
Table 1.1	Summary of biophysical mechanisms of RF-EMF11
Table 1.2	Summary of IEGMP (2000) report on biological effects of
mobile phor	ne exposure in cellular systems
Table 1.3A	Genotoxic effects from mobile phone exposure
Table 1.3B	Effect of mobile phone exposure on Apoptosis
	Effect of mobile phone exposure on gene and protein
expression.	
Table 1.3D	Effect of mobile phone exposure on male germ cells
Chapter 2	
Table 2.1	Heat shock protein families, their expression and
functions	90
Table 2.2	Conditions that lead to the activation of heat shock proteins
(from Prohá	szka and Füst, 2004)61
Chapter 3	
Table 3.1	Percentage progressive motility after (A) RF-EMF (2.0 W/kg)
and (B) RF-	EMF (5.7 W/kg) exposure in exposed and control spermatozoa
determined	directly (T ₁), 2 h (T ₂) and 24 h (T ₃) after exposure
Table 3.2	Linear regression results of percentage rapid-, slow-, non-
progressive	and immotile spermatozoa after RF-EMF (2.0 and 5.7 W/kg)



exposure compared to control spermatozoa. Spermatozoa exposed at 2.0	
W/kg are also compared to sperm exposed at 5.7 W/kg	120
Table 3.3 Summary of morphometric results of sperm exposed to (A) 2.0	
W/kg and (B) 5.7 W/kg compared to control values	125
Table 3.4 Summary of p values of the linear regression analysis results of	
sperm morphology determined by CASA. RF exposed sperm were firstly	
compared to controls, morphometric parameters were compared directly	
(T_1) and 2 hours (T_2) after exposure and the number of normal and abnormal	
	105
forms were compared	125
Table 3.5 Linear regression analysis comparing sperm morphometric	
parameters of sperm exposed at 2.0 W/kg to sperm exposed at 5.7 W/kg are	
summarised as p values.	126
r	
Chapter 4	
Table 4.1 Summary of paired t-test results comparing viability (PI-	
fluorescence) and PS externalisation (Annexin V fluorescence) of RF-	
exposed spermatozoa at SAR 2.0 W/kg with SAR 5.7 W/kg	169
Table 4.2 Comparison between Annexin V^{+} PI^{-} , Annexin V^{+} PI^{+} and	
Annexin V PI staining cells as a progression of time.	169
Table 4.3 A Correlations between apoptotic biomarkers of RF-EMF (SAR	
2.0 W/kg) exposed spermatozoa evaluated directly (T_1) , 3 hours (T_2) and 24	
hours (T ₃) after exposure	190
nours (13) arter exposure	100
Table 4.3 B Correlations between apoptotic biomarkers of RF-EMF (SAR	
5.7 W/kg) exposed spermatozoa evaluated directly (T_1) , 3 hours (T_2) and 24	
hours (T ₃) after exposure	181



Chapter 6	
Table 6.1 Summary of effects of RF-EMF on sperm specific and	
functional assays in highly motile human spermatozoa	234
Annexure A	
Table A.1 SAR distribution results used in experimentation.	252
Annexure C	
Table C.1 Average sperm parameters: Volume, pH and motility averages	
\pm SD of the semen sample is noted. Concentration and morphology averages	
± SD are given both pre- and post- density gradient purification	268