

Development and applications of a novel, thermoresponsive scaffold for three-dimensional cell culture

by

Claire Louise Rossouw

Submitted in partial fulfilment of the requirements for the degree *Philosophiae Doctor*

In the Faculty of Natural and Agricultural Sciences

Department of Biochemistry

University of Pretoria

Pretoria

South Africa

SUPERVISOR:

Prof. Lyn-Marie Birkholtz

Department of Biochemistry, University of Pretoria, South Africa

CO-SUPERVISORS:

Dr. Dalu Mancama

Molecular and Biomedical Technologies, Biosciences, CSIR, South Africa

Prof. Heinrich Hoppe

Department of Biochemistry, Microbiology and Biotechnology, Rhodes University, South Africa

Dr. Sean Moolman

Licensing & Ventures, CSIR, South Africa

I, Claire Louise Rossouw, declare that the thesis, which I hereby submit for the degree
Philosophiae Doctor at the University of Pretoria, is my own work and has not previously been
submitted by me for a degree at this or any other tertiary institution.
SIGNATURE:
DATE:

I understand what plagiarism is and am aware of the University's policy in this regard.

I declare that this thesis is my own original work. Where other people's work has been used (either from printed source, internet or any other source), this has been properly acknowledged and referenced in accordance with departmental regulations.

I have not used work previously produced by another student or any other person to hand in as my own.

I have not allowed and will not allow anyone to copy my work with the intention of passing it off as his or her own work.

Student's signature

Date



The completion of this dissertation was made possible by the following people whom I would like to thank:

My supervisors, Prof. Lyn-Marie Birkholtz, Dr Dalu Mancama, Prof. Heinrich Hoppe and Dr Sean Moolman for your guidance, support and encouragement. It was a privilege to work with you!

Avashnee Chetty, for stimulating discussions and for supplying the 3D scaffolds.

Kobus van Wyk, for technical assistance and help with the bioreactor experimental work.

Dr Jetsumon Sattabongkot (AFRIMS), for allowing me to train in your laboratory and for the long distance help via e-mail.

Megan Dowler (WRAIR), for your willingness to share your vast gametocyte culturing knowledge with me.

The CSIR and the 3R foundation in Switzerland for funding.

My fellow students, past and present: Dr Walter Campos and Dayaneethie Coopusamy for your help in tissue culture; Dr Tharina van Brummelen and Dr John Becker for teaching me to culture malaria parasites and Lia Rotherham for your friendship and encouragement.

My husband, Johann Rossouw, for all the weekends spent in the lab so I could feed my parasites, your unconditional love and unwavering support.

My parents, Gavin and Jane Munro and my family. I am so thankful for all the love, support and encouragement you have given me all my life.



SUMMARY

Although conventional two-dimensional (2D) cell culture is convenient for routine work, researchers are turning to three-dimensional (3D) cell culture for more accurate, physiologically representative information on the way their cells behave and respond to stimuli. Cells can now be routinely cultured in the many commercially available 3D formats.

In this study, we developed non-woven scaffolds for 3D cell culture and enhanced cell function. By making use of methods that measure the behaviour of liver cells in the 3D system we were able to demonstrate, compared to standard 2D systems, significantly higher expression of key liver enzymes involved in drug metabolism and albumin production (specifically cytochrome P450). Cell proliferation on the various scaffolds was comparable to that of a commercially available hydrogel 3D cell culture system, AlgimatrixTM.

When culturing cells in 3D, the means by which cells are harvested or extracted from the 3D scaffold for downstream applications is more challenging than in 2D. For this reason, many of the 3D scaffolds currently manufactured are either bio-degradable or require the use of salts to dissolve the scaffold which may negatively impact on the cells they contain. By grafting the non-woven scaffolds with the thermoresponsive polymer, poly(*N*-isopropylacrylamide) (PNIPAAm), we demonstrated that cells growing on the scaffolds are able to be released from the scaffold in a 3D conformation, non-enzymatically, through temperature changes. Selected thermoresponsive non-woven fabrics were also tested in an automated cell culture device for cell proliferation and thermally induced harvesting.

One of the applications of a 3D cell culturing system would be in exploration of the many diseases plaguing mankind, in particular malaria which is still responsible for severe disease and mortality, especially in Africa. Most available antimalarials are designed to target the pathogenic blood stages in humans and to address the constant threat of drug resistance. However, to meet the objective of malaria eradication, medicines that block parasite transmission also need to be developed. Molecules that efficiently target the parasite stages in the liver would prevent pathogenesis, symptoms and transmission. Equipped with the knowledge that the infectious sporozoites traverse several hepatocytes prior to cell infection, it may be physiologically limiting to culture the exo-erythrocytic stage *in vitro* in a 2D cell culture system where the hepatocytes are in an unnatural flat conformation, distinctly different to their *in vivo* counterparts. Moreover, monolayer cell cultures lose their tissue-related functions rapidly, greatly impairing the predictive power of such assays. Thus, the second aim of this thesis was



to establish if hepatocytes that have been cultured on 3D non-woven scaffolds improve *in vitro* sporozoite invasion compared to conventional 2D systems. Sporozoite invasion was detected in the conventional 2D monolayers using a TaqMan® assay but not in the hepatocytes growing in 3D. Future studies beyond the scope of this thesis will include modifications to the 3D scaffold to attempt achieving superior sporozoite invasion in this model system.

TABLE OF CONTENTS

Submission Declaration	ii
Plagiarism Statement	iii
Acknowledgements	iv
Summary	v
Table of Contents	vii
List of Figures	X
List of Tables	xiii
Abbreviations	xiv
Appendix A	xv
CHAPTER 1	1
Three dimensional cell culture systems and their applications: A review	1
1.1. 2D vs 3D: Why do we need 3D?	2
1.2. 3D Scaffolds	5
1.2.1. Materials of choice	5
1.2.2. Bulk modifications	9
1.2.2. Surface modifications	10
1.3. Applications of in vitro 3D cell culture	11
1.3.1. In vitro anti-cancer drug screens: Tumour tissue models	12
1.3.2. In vitro toxicology models	13
1.3.3. In vitro host-pathogen interactions	14
1.4. 3D cell culture: Future directions	15
1.5. Objectives	17
CHAPTER 2	19
Hepatocyte proliferation and thermally induced cell detachment on non-woven F	PP, PET and
nylon three-dimensional scaffolds	
2.1. Introduction	19
2.1.1. Scaffold selection	21
2.1.2. Cell culture automation	
2.1.3. Objectives	
2.2. Materials and Methods	
2.2.1. Scaffold fabrication	24
Z.Z. I. OCATIOIO IADITICATION	∠4



	2.2.2.	Grafting methods	24
	2.2.3.	Cell-scaffold interaction	25
	2.2.4.	Cell viability and proliferation	.27
	2.2.4.1.	AlamarBlue® assay	.27
	2.2.4.2.	DNA quantification using Hoechst 33258	.28
	2.2.5.	Imaging cell-scaffold-interaction	.29
	2.2.6.	Hepatocyte metabolic activity measurement	.29
	2.2.6.1.	Bradford standard curve	.29
	2.2.6.2.	Albumin assay	30
	2.2.7.	Cytochrome P450 mRNA expression	30
	2.2.7.1.	RNA extractions	.31
	2.2.7.2.	cDNA synthesis and qRT-PCR	.32
	2.2.8.	Thermal release of cells from the various scaffolds	.32
	2.2.9.	Automated cell culture and thermal cell release	.33
2	.3. Res	ults	36
	2.3.1.	Grafting methods	.36
	2.3.2.	Cell viability and proliferation	.37
	2.3.3.	Fluorescence microscopy and albumin quantification	40
	2.3.4.	Cytochrome P450 mRNA expression	43
	2.3.5.	Thermal release	46
	2.3.6.	Automated cell culture device	.51
	2.3.6.1.	Prototype	.52
	2.3.6.2.	Final system design and testing	54
2	.4. Disc	cussion	.58
CHAP	TER 3		63
App	lication o	f A 3D scaffold: A sporozoite-hepatocyte model	63
		oduction	
J.			
	3.1.1.	Malaria	
	3.1.2.	Sporozoite hepatocyte invasion	
2	3.1.3.	In vitro culturing of hepatocytes for malaria sporozoite invasion	
3	.2. Mat	erials and Methods	. 00
	3.2.1.	In vitro cultivation of asexual P. falciparum cultures	.68
	3.2.2.	In vitro cultivation of sexual stage P. falciparum parasites (gametocyte	70
	3.2.2.1.	n)	
	J.Z.Z. I.	Candle jai memod	. , 0



3.2.2.2	2. Flask method	71
3.2.2.3	3. Monitoring of exflagellation	71
3.2.3.	Mosquito rearing	72
3.2.4.	Mosquito feeding and dissections	72
3.2.4.	1. Mosquito preparation	72
3.2.4.2	Mosquito mid-gut dissections and sporozoite isolation from salivary g	lands 73
3.2.5.	Populating 3D scaffolds and 2D wells with hepatocytes	75
3.2.6.	Seeding sporozoites into 2D hepatocytes and 3D scaffolds	76
3.2.7.	gDNA isolation	77
3.2.8.	TaqMan® assay	77
3.3. R	esults	80
3.3.1.	Gametocyte production and exflagellation	80
3.3.2.	Sporozoite invasion in 2D and 3D	88
3.3.2.	1. Mosquito feeding and dissections	88
3.3.2.2	2. Sporozoite invasion of HC04 cells	89
3.3.2.3	3. Quantification of sporozoite invasion in HC04 cells	90
3.4. Dis	cussion	93
CHAPTER 4		101
Concluding	g Discussion	101
Reference	s	106
Appendix A	Α	116



LIST OF FIGURES

Figure 1.1. 3D cell culture publications per year (1977- June 2012)2
Figure 1.2. 2D and 3D cell culture morphology3
Figure 1.3. Bridging the gap between 2D cell culture and natural tissue4
Figure 1.4. The three criteria used to design 3D scaffolds5
Figure 1.5. Polymer scaffolds for 3D cell culture6
Figure 1.6. Selected <i>in vitro</i> applications of 3D cell culture11
Figure 2.1. Poly(<i>N</i> -isopropyl acrylamide)20
Figure 2.2. Temperature-responsive culture dishes21
Figure 2.3. The prototype automated cell culture device
Figure 2.4. AlamarBlue® assay standard curves for HC04 and HepG2 hepatocytes37
Figure 2.5. Cell proliferation on poly(propylene) (PP), poly(ethylene terephthalate) (PET) and nylon non-woven scaffolds, determined by the AB assay
Figure 2.6. Hoechst 33258 standard curve using crude cell lysates
Figure 2.7. Cell proliferation of hepatocytes growing on poly(propylene) (PP), poly(ethylene terephthalate) (PET) and nylon non-woven scaffolds, determined by DNA staining40
Figure 2.8. Representative fluorescence micrographs of HC04 (top panel) and HepG2 (bottom panel) hepatocytes growing on the PP-g-PNIPAAm-B scaffold
Figure 2.9. Bradford assay and albumin standard curves



3D non-woven control scaffold (pore size 200 μm)43
Figure 2.11: Relative CYP gene expression
Figure 2.12. Thermal release of HC04 hepatocytes from grafted and control scaffolds 10 dpi
Figure 2.13. Thermal release of HC04 and HepG2 hepatocytes
Figure 2.14. Additional controls used to assess if omitting any stages of the grafting process have any effect on thermal release
Figure 2.15. Effect of non-woven scaffolds' pore size on thermal release50
Figure 2.16. The perfusion circuit layout and system components of the proposed automated cell culture system
Figure 2.17. HC04 hepatocytes growing on the three non-woven scaffolds housed in the bioreactor
Figure 2.18. The final automated cell culture system56
Figure 2.19. HC04 hepatocytes released from grafted (PP- <i>g</i> -PNIPAAm-B1) and control (PP-Cont) non-woven scaffolds 10 dpi in the final automated system
Figure 3.1. The life cycle of <i>P. falciparum</i> parasites in the human host and the <i>Anopheles</i> mosquito vector
Figure 3.2. Mosquito mid-gut dissections74
Figure 3.3. Mosquito salivary gland dissections
Figure 3.4. A qualitative comparison of the different gametocyte producing methods using three parasite strains after 10 days in culture



Figure 3.5. A comparison of culture health and gametocyte production for between	the candle
jar and flask method and the three parasite strains	83
Figure 3.6. Gametocyte staging and sexing	85
Figure 3.7. <i>P. falciparum</i> gametocytogenesis	86
Figure 3.8. Mature microgamete exflagellation	88
Figure 3.9. Mosquito mid-gut and salivary gland dissections	89
Figure 3.10. HC04 hepatocytes populating the non-woven scaffolds	90
Figure 3.11. TaqMan® expression data	91
Figure 4.1. An illustration of the "human-on-a-chip" concept	104



LIST OF TABLES

Table 2.1. Non-woven scaffolds used for this study	25
Table 2.2. A summary of successful methods used to achieve a grafted layer of PNIPAAm or he surfaces of poly(propylene) (PP), poly(ethylene terephthalate) (PET) and nylon non-wovelenges	en
المادة المادة المادة Fable 2.3. Primer sequences for cytochrome qRT-PCR	43
Table 2.4. Hepatocyte cell release from scaffolds PP-g-PNIPAAm-B1, PP-g-PNIPAAm-B2 a PP-g-PNIPAAm-B3 via trypsin treatment or thermal release	
Table 2.5. Experimental parameters assessed using the prototype automated cell cultudevice	
Table 3.1. Mosquitoes fed for malaria infection studies	72
Table 3.2. The ring:trophozoite ratio	87



ABBREVIATIONS

AB - AlamarBlue®

APS - Ammonium Persulphate

ATR-FTIR - Attenuated Total Reflection-Fourier Transform Infrared Spectroscopy (ATR-FTIR),

DMEM - Dulbecco's Modified Eagle Medium

DPI - Days Post Inoculation

DSC - Differential Scanning Calorimetry

ECM - Extracellular Matrix

EDTA - Ethylenediaminetetraacetic acid

EEF - Exoerythrocytic Form

FCS - Foetal Calf Serum

FDA - Fluorescein Diacetate

HPI - Hours Post Inoculation

HSPG - Heparin Sulphate Proteoglycan

LCST - Lower Critical Solution Temperature

MCTS - Multicellular Tumour Spheroid

MFP - Median Flow Pore

PBS - Phosphate Buffered Saline

PCL - Poly(ε-caprolactone)

PEG - Poly(ethylene glycol)

PET - Poly(ethylene terephthalate)

PNIPAAm - Poly(*N*-isopropylacrylamide)

PP - Poly(propylene)

SEM - Scanning Electron Microscopy

SFRP - Solution Free-Radical Polymerisation

XPS - X-Ray Photoelectron Spectroscopy



APPENDIX A

Table A1. TaqMan® expression data1	16
Table A2. TaqMan® expression data for increased primer and probe concentrations1	17
Table A3. TaqMan® expression data from invaded cells growing in 2D and nylon scaffolds1	118