

Supplementary material

T-Cell Phenotypes and Systemic Cytokine Profiles of People Living with HIV Admitted to Hospital with COVID-19

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Supplementary Table 1: Univariate comparison of T-cell profiles of people with and without HIV at admission with COVID-19, as well as controls

T-cell population	COVID+PLWOH (n=117)	Control PLWOH (n=10)	COVID+PLWH (n=36)	Control PLWH (n=19)	1 vs 2	1 vs 3	3 vs 4	2 vs 4	Overall p-value
	Group 1	Group 2	Group 3	Group 4					
CD4+	66.51 (54.66 - 76.47)	73.00 (65.88-79.63)	48.96 (17.23 - 58.12)	42.56 (22.23-54.93)	0.110	<0.001	0.336	<0.001	<0.001
CD8+	27.77 (18.91-37.08)	22.92 (15.18-28.04)	47.86 (36.02-73.5)	53.13 (39.30-70.11)	0.151	<0.001	0.404	<0.001	<0.001
DP	0.84 (0.47-1.73)	0.50 (0.40-0.92)	0.44 (0.23-0.76)	0.55 (0.19-0.81)	0.132	<0.001	0.461	0.195	<0.001
DN	3.28 (2.19-4.78)	3.70 (2.74-5.39)	4.58 (2.48-6.28)	5.47 (3.66-7.44)	0.223	0.015	0.083	0.076	0.004
CD8+ PD-1+	35.09 (21.11-44.27)	27.66 (19.59-36.61)	37.76 (27.89-51.04)	34.96 (30.86-48.30)	0.197	0.032	0.375	0.082	0.146
CD4+ CM	53.45 (44.96-65.76)	53.81 (44.70-58.86)	50.66 (37.52-58.72)	51.96 (44.45-59.22)	0.470	0.025	0.132	0.221	0.252
CD4+ EM	10.97 (6.93-17.2)	7.76 (5.46-9.35)	17.88 (9.61-31.90)	23.36 (13.96-31.79)	0.071	0.002	0.109	<0.001	<0.001
CD4+ EM2	0.51 (0.22-0.74)	0.31 (0.12-0.60)	0.34 (0-0.54)	0.82 (0.27-1.81)	0.212	0.018	0.003	0.050	0.033
CD4+ CM PD-1+	37.66 (29.59-47.76)	33.72 (24.64-37.91)	57.86 (51.18-73.30)	53.00 (45.61-73.03)	0.051	<0.001	0.229	<0.001	<0.001
CD8+ EM1	46.64 (27.69-60.5)	64.74 (59.49-70.49)	32.66 (24.06-42.73)	38.64 (19.21-46.76)	0.010	<0.001	0.268	0.001	0.002
CD8+ EM2	13.42 (9.63-20.24)	11.55 (7.28-14.80)	23.94 (15.35-35.40)	18.13 (8.69-43.22)	0.131	<0.001	0.090	0.016	<0.001
CD8+ EM1 PD-1+	35.97 (21.72-49.78)	40.86 (37.80-59.26)	25.37 (21.10-35.43)	19.10 (10.99-35.10)	0.085	0.012	0.131	0.001	0.001
CD8+ EM2 PD-1+	9.72 (4.84-14.42)	6.72 (4.51-9.93)	18.3 (12.24-29.91)	11.00 (3.41-33.25)	0.113	<0.001	0.006	0.052	<0.001

CD8+ EM3 PD-1+	9.18 (4.45-21.02)	5.55 (2.66-11.13)	13.89 (9.49-24.84)	17.82 (6.55-21.28)	0.0713	0.008	0.317	0.020	0.073
CD8+ TEMRA ECD57+	52.4 (37.11-62.61)	40.29 (21.49-53.80)	44.07 (25.91-52.95)	50.95 (31.87-61.15)	0.035	0.006	0.146	0.143	0.036
CD8+ TEMRA pE1	5.43 (3.04-10.21)	12.14 (4.42-17.38)	4.37 (2.62-7.01)	5.55 (2.83-13.12)	0.062	0.027	0.115	0.086	0.071
CD8+ CM PD-1+	45.64 (36.05-56.02)	51.26 (42.11-54.99)	62.22 (55.68-68.67)	55.34 (44.71-65.50)	0.325	<0.001	0.069	0.109	<0.001

The Kruskal-Wallis test with post hoc Dunn test was used to compare continuous variables between groups. Results are presented as median and interquartile range (IQR).

Abbreviations: Central memory (CM), double -negative (DN), double -positive (DP), end stage effector (E), effector memory (EM), people living without HIV (PLWOH), people living with HIV (PLWH), programmed cell death protein 1 (PD-1), pre-effector 1 (pE1), terminally differentiated T-cells re-expressing CD45RA (TEMRA).

Supplementary Table S2: Univariate comparison of T-regulatory cell cytokines of people with and without HIV at admission with COVID-19, as well as controls.

Treg marker	COVID+PLWOH (n=102)	Control PLWOH (n=8)	COVID+PLWH (n=35)	Control PLWH (n=8)					Overall p-value
(pg/mL)	Group 1	Group 2	Group 3	Group 4	1 vs 2	1 vs 3	3 vs 4	2 vs 4	
IL-2	6.33 (2.48-19.12)	0.25 (0.25-2.67)	7.88 (2.48-15.52)	0.26 (0.25-8.26)	0.003	0.489	0.025	0.307	0.008
IL-10	3.37 (1.11-3.37)	1.11 (1.11-1.11)	3.37 (1.11-3.37)	1.11 (1.11-1.11)	0.011	0.368	0.009	0.492	0.013
IL-12p40	125.45 (87.66-180.04)	144.33 (57.21-134.65)	129.14 (77.92-178.12)	112.48 (82.38-136.36)	0.164	0.343	0.298	0.443	0.682
IL-19	72.54 (13.22-132.94)	3.78 (3.78-3.79)	75.87 (13.22-104.94)	38.17 (3.78-132.35)	<0.001	0.386	0.196	0.055	0.010
IL-20	35.52 (15.92-63.05)	22.37 (4.24-28.76)	41.37 (6.91-56.9)	17.54 (4.24-38.22)	0.027	0.483	0.041	0.484	0.073
IL-22	2.69 (2.02-2.69)	2.69 (2.69-2.69)	2.69 (2.02-2.69)	2.69 (2.69-2.69)	0.189	0.384	0.165	0.500	0.631
IL-26	358.6 (100.27-358.6)	100.27 (100.27-100.27)	358.6 (100.27-358.66)	100.27 (100.27-100.27)	0.004	0.392	<0.001	0.245	<0.001
IL-28A	26.56 (13.91-48.3)	9.12 (2.17-12.93)	28.81 (5.82-49.74)	13.69 (10.65-31.07)	0.004	0.488	0.114	0.162	0.037
IL-29	29.64 (10.85-52.67)	46.36 (11.94-57.74)	33.08 (6.99-52.67)	19 (7.89-38.71)	0.234	0.464	0.270	0.164	0.809
IL-35	97.6 (13.78-180.85)	126.10 (43.84-188.46)	116.51 (50.82-199.85)	95.2 (31.15-162.48)	0.296	0.157	0.328	0.364	0.757
IL-27p28	16.01 (5.05-16.01)	5.05 (5.05-18.98)	16.01 (16.01-16.01)	5.05 (5.05-8.48)	0.062	0.072	0.002	0.303	0.013

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range) in picogram (pg)/mL.

Abbreviations: people living without HIV (PLWOH), interleukin (IL), people living with HIV (PLWH), T-regulatory cell (Treg)

Supplementary Table S3: Univariate comparison of pro- and anti-inflammatory cytokines of people with and without HIV at admission with COVID-19, as well as controls.

Cytokine	COVID+PLWOH (n=137)	Control PLWOH (n=9)	COVID+PLWH (n=37)	Control PLWH (n=9)	1 vs 2	1 vs 3	3 vs 4	2 vs 4	Overall p-value
	Group 1	Group 2	Group 3	Group 4					
TGF-β1 (ng/mL)	8.08 (5.73-11.85)	5.02 (3.10-5.73)	7.78 (5.20-11.57)	3.24 (3.16-7.12)	0.007	0.479	0.005	0.020	0.004
IL-1β (pg/mL)	1.95 (1.46-2.89)	1.22 (0.98-1.71)	1.95 (1.46-2.66)	0.72 (0.59-0.72)	0.009	0.406	<0.001	0.039	<0.001
IL-1Ra (pg/mL)	812.8 (631.27-1081.29)	237.73 (214.08-407.80)	894.53 (725.59-1133.51)	259.97 (214.08-554.28)	<0.001	0.104	<0.001	0.010	<0.001
IL-4 (pg/mL)	4.14 (3.07-5.49)	2.83 (2.07-3.79)	4.54 (3.9-5.49)	2.7 (2.32-3.19)	0.016	0.105	<0.001	0.080	0.002
IL-6 (pg/mL)	4.09 (1.54-8.93)	0.49 (0.30-0.54)	6.38 (2.29-13.18)	0.49 (0.49-0.77)	<0.001	0.102	<0.001	0.010	<0.001
IL-7 (pg/mL)	45.82 (28.96-64.84)	10.79 (6.88-27.05)	41.79 (71.89-27.05)	17.74 (10.79-38.34)	<0.001	0.445	0.004	0.128	<0.001
IL-8 (pg/mL)	17.5 (10.73-23.53)	3.58 (2.8-6.47)	19.69 (15.32-28.23)	4.62 (3.84-6.47)	<0.001	0.114	<0.001	0.011	<0.001
IL-13 (pg/mL)	3.61 (2.18-5.4)	1.43 (0.91-1.55)	3.2 (1.76-5.01)	0.91 (0.47-1.76)	<0.001	0.267	<0.001	0.054	<0.001
IL-15 (pg/mL)	18.02 (18.02-18.02)	18.02 (18.02-18.02)	18.02 (18.02-18.02)	18.02 (18.02-18.02)	0.227	0.032	0.247	0.268	0.226
IL-17 (pg/mL)	22.61 (17.36-32.32)	12.58 (10.37-19.62)	24.85 (18.74-34.51)	15.22 (12.14-19.62)	0.009	0.223	0.007	0.172	0.008
Eotaxin (pg/mL)	22.81 (15.84-31.11)	33.93 (26.19-42.79)	27.01 (20.02-35.66)	32.93 (23.68-33.75)	0.010	0.025	0.414	0.334	0.027
IFN-γ (pg/mL)	15.81	6.75	16.93	4.99	<0.001	0.186	<0.001	0.090	<0.001

	(10.7-23.01)	(4.54-10.67)	(12.37-26.35)	(4.1-9.37)					
	952.33	160.02	680.22	261.01	<0.001	0.207	0.024	0.308	<0.001
IP-10 (pg/mL)	(421.09-2087.96)	(129.86-208.83)	(415.47-1442.47)	(213.06-763.50)					
	83.87	55.49	90.85	50.67	<0.001	0.100	0.005	0.403	<0.001
TNF- α (pg/mL)	(68.6-102.39)	(50.67-68.60)	(73.41-118.4)	(47.67-81.53)					

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range) in picograms (pg)/mL with the exception of TGF- β 1 which is expressed in nanograms (ng)/mL.

Abbreviations: people living without HIV (PLWOH), interleukin (IL), IL-1 receptor antagonist (IL-1Ra), interferon gamma (IFN- γ), interferon gamma-induced protein 10 (IP-10), people living with HIV (PLWH), transforming growth factor- β 1 (TGF- β 1), tumor necrosis factor- α (TNF- α).

Supplementary Table S4: Univariate comparison of the cytokine profiles of people living with HIV according to CD4 count at admission with COVID-19, as well as with their respective controls.

Cytokine	COVID CD4 ≥200 (n=19)	Control CD4 ≥200 (n=4)	COVID CD4 <200 (n=17)	Control CD4 <200 (n=5)	1 vs 2	1 vs 3	3 vs 4	2 vs 4	Overall p-value
	Group 1	Group 2	Group 3	Group 4					
TGF-β1 (ng/mL)	7.98 (5.20-17.06)	3.91 (3.16-8.42)	7.35 (5.08-11.57)	3.24 (2.70-7.12)	0.046	0.289	0.032	0.380	0.074
IL-1β (pg/mL)	1.95 (1.35-3.69)	0.66 (0.39-0.85)	2.03 (1.71-2.43)	0.72 (0.59-0.72)	0.002	0.443	0.022	0.227	0.007
IL-1Ra (pg/mL)	894.53 (655.64-1133.51)	237.03 (153.06-407.13)	952.91 (747.98-1175.93)	301.18 (214.08-580.65)	0.005	0.298	0.004	0.360	0.003
IL-2 (pg/mL)	9 (4.23-10.99)	1.63 (1.26-1.63)	5.47 (1.63-10.30)	1.63 (1.63-1.71)	0.001	0.141	0.077	0.191	0.008
IL-4 (pg/mL)	4.54 (3.9-6.47)	2.95 (2.39-3.31)	4.70 (3.98-5.04)	2.45 (2.32-2.70)	0.011	0.430	0.035	0.343	0.037
IL-6 (pg/mL)	4.32 (1.25-7.28)	0.49 (0.28-0.63)	13.18 (5.39-72.30)	0.49 (0.49-1.01)	0.029	0.009	0.001	0.343	0.001
IL-7 (pg/mL)	50.12 (35.61-96.45)	16.06 (12.59-28.04)	35.53 (24.05-58.82)	17.74 (4.7-50.12)	0.004	0.059	0.136	0.278	0.017
IL-8 (pg/mL)	18.59 (14.45-20.24)	5.15 (4.1-6.21)	23.81 (15.32-31.61)	4.62 (1.78-7.53)	0.015	0.091	0.001	0.465	0.002
IL-13 (pg/mL)	3.61 (2.59-7.56)	0.80 (0.47-1.39)	3.05 (1.55-4.61)	0.91 (0.47-1.76)	0.004	0.122	0.026	0.439	0.006
IL-15 (pg/mL)	18.02 (18.02-109.12)	18.02 (18.02-18.02)	18.02 (18.02-18.02)	18.02 (18.02-18.02)	0.106	0.124	0.353	0.232	0.521
IL-17 (pg/mL)	26.64 (16.1-35.38)	15.88 (9.26-20.50)	23.36 (18.74-34.51)	15.22 (13.46-18.74)	0.021	0.361	0.097	0.308	0.119
Eotaxin (pg/mL)	26.38	33.34	30.25	25.01	0.125	0.188	0.217	0.137	0.566

	(19.45-33.39)	(28.31-39.61)	(21.89-37.84)	(15.05-33.57)					
	16.23	4.50	20.48	4.99	0.026	0.134	0.007	0.389	0.014
IFN- γ (pg/mL)	(8.66-29.31)	(1.37-12.15)	(14.53-26.35)	(4.54-9.37)					
	554.9	349.50	854.15	261.01	0.099	0.035	0.034	0.285	0.046
IP-10 (pg/mL)	(338.98-981.23)	(181.59-629.52)	(672.38-1701.47)	(252.58-922.75)					
	84.60	52.46	93.16	50.67	0.019	0.378	0.115	0.174	0.111
TNF- α (pg/mL)	(73.41-124.78)	(47.03-69.71)	(76.85-116.12)	(48.25-114.98)					

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range) in picograms (pg)/mL with the exception of TGF- β 1 which is expressed in nanograms (ng)/mL.

Abbreviations: Interleukin (IL), IL-1 receptor antagonist (IL-1Ra), interferon gamma (IFN- γ), interferon gamma-induced protein 10 (IP-10), transforming growth factor- β 1 (TGF- β 1), tumor necrosis factor- α (TNF- α).

Supplementary Table S5: Univariate comparison of the regulatory cytokine profiles of people living with HIV according to CD4 count at admission with COVID-19, as well as with their respective controls.

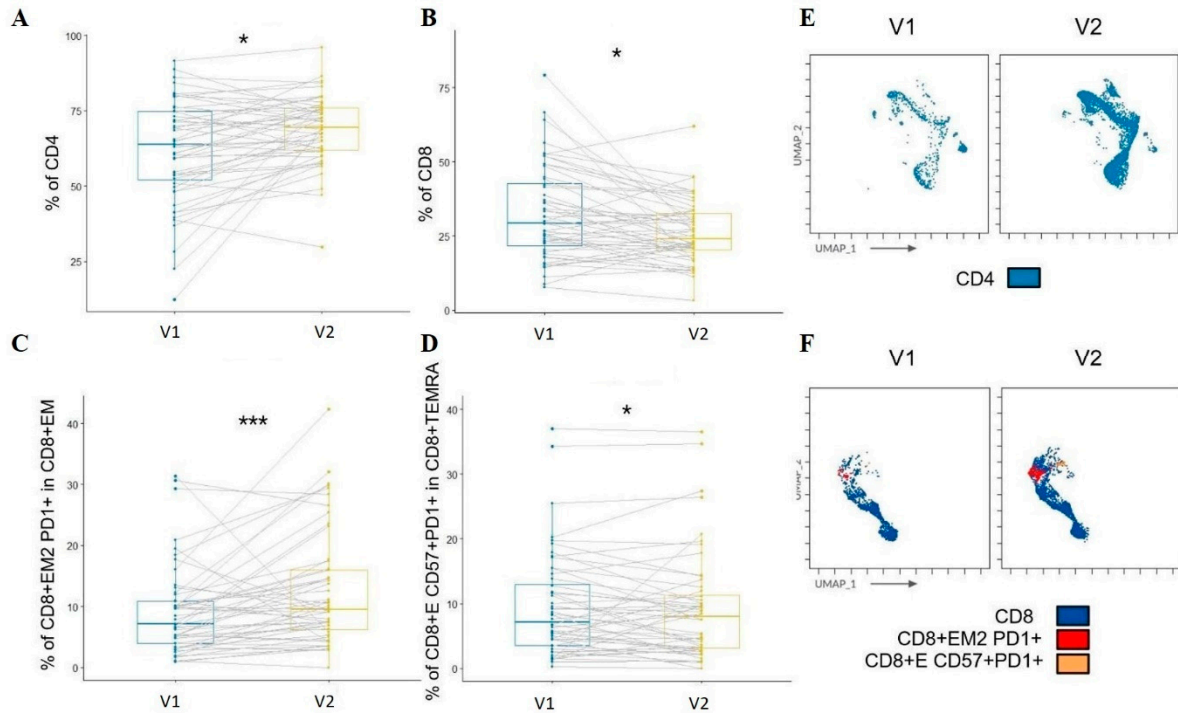
Treg marker (pg/mL)	COVID CD4 ≥200 (n=19)	Control CD4 ≥200 (n=4)	COVID CD4 <200 (n=17)	Control CD4 <200 (n=5)	1 vs 2	1 vs 3	3 vs 4	2 vs 4	p-value
	Group 1	Group 2	Group 3	Group 4					
IL-2	6.99 (2.48-24.14)	6.47 (0.25-13.77)	7.88 (2.48-12.85)	0.26 (0.25-2.05)	0.207	0.421	0.033	0.180	0.217
IL-10	3.37 (1.11-3.37)	1.11 (1.11-2.58)	3.37 (2.24-5.52)	1.11 (1.11-1.11)	0.191	0.118	0.005	0.215	0.048
IL-12p40	143.81 (87.66-182.42)	136.36 (97.41-172.80)	91.86 (52.33-156.37)	97.45 (68.83-112.48)	0.489	0.084	0.318	0.143	0.367
IL-19	85.32 (13.22-104.94)	59.21 (3.78-136.46)	80.20 (13.22-109.27)	38.17 (3.79-111.30)	0.418	0.446	0.262	0.392	0.930
IL-20	42.93 (22.38-63.63)	30.21 (10.95-49.15)	39.46 (6.91-53.40)	10.84 (2.51-25.55)	0.219	0.187	0.048	0.127	0.161
IL-22	2.69 (2.02-32.3)	2.69 (2.69-2.69)	2.03 (2.02-2.69)	2.69 (2.69-2.69)	0.360	0.071	0.105	0.500	0.337
IL-26	358.60 (100.27-358.60)	100.27 (100.27-100.27)	358.60 (229.44-358.60)	100.27 (100.27-100.27)	0.011	0.323	0.005	0.500	0.008
IL-28A	33.32 (15.21-53.55)	19.74 (12.17-31.07)	27.12 (4.16-43.85)	12.17 (6.04-25.78)	0.226	0.251	0.222	0.365	0.623
IL-29	50.19 (27.02-57.47)	28.09 (7.89-58.85)	19.00 (7.00-33.08)	19.00 (7.89-29.62)	0.275	0.016	0.426	0.235	0.140
IL-35	124.21 (50.82-199.85)	126.97 (43.84-196.55)	106.35 (56.28-174.00)	82.51 (31.14-130.71)	0.482	0.348	0.226	0.227	0.790
IL-27p28	16.01 (16.01-26.98)	8.48 (5.05-34.40)	16.01 (16.01-16.01)	5.05 (5.05-5.05)	0.100	0.351	0.005	0.113	0.028

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range) in picograms (pg)/mL.

Abbreviations: Interleukin (IL), T-regulatory cell (Treg)

Differences in the T-cell profiles of people with COVID-19 without HIV between V1 and V2

Differences were found between bloods taken at visit 1 (V1) and visit 2 (V2) during hospitalization for COVID-19 (Supplementary Table S6 and Figure S1). At the V2 timepoint, the percentages of CD4+ T-cells increased ($p=0.019$) and CD8+ T-cells decreased ($p=0.025$). The increase was seen within the CD8+ effector memory 2 (EM2: CCR7-CD45RA-CD28-CD27+) ($p=0.001$) subset, specifically CD8+ EM2 T-cells expressing PD-1 ($p<0.001$). A difference was also found between V1 and V2 for the TEMRA subset. There was an increase in the percentage of CD8+ end-stage effector (E: CCR7-CD45RA+CD28-CD27-) T-cells co-expressing CD57 and PD-1 ($p=0.019$).



Supplementary Figure S1: Differences in the T-cell phenotypes of people with COVID-19 without HIV at hospital admission (V1) and 5 ± 3.6 days after admission (V2). **A:** The percentage of CD4+ T-cells increased from V1 to V2. **B:** The percentage of CD8+ T-cells decreased from V1 to V2. **C:** The percentage of CD8+ EM2 expressing PD-1 increased from V1 to V2. **D:** Boxplots show an increase in the percentage of CD8+ E co-expressing CD57 and PD-1 from V1 to V2. **E:** A representative UMAPs showing an increase of CD4+ T-cells between V1 and V2 in people without HIV admitted with COVID-19. **F:** Representative UMAP plots show the increases/decreases in the respective CD8+ T-cell populations between V1 and V2 in people without HIV admitted with COVID-19.

p-value: * = < 0.05 , ** = < 0.01 .

Abbreviations: End-stage effectors (E), effector memory (EM), programmed cell death protein 1 (PD-1), visit 1 (V1), visit 2 (V2)

Supplementary Table S6: Differences in T-cell populations between V1 and V2 in people without HIV hospitalized with COVID-19.

T-cell population	COVID-19+ PLWOH V1 (n=52)	COVID-19+PLWOH V2 (n=52)	p-value
CD4+	63.85 (51.74-74.73)	69.59 (61.93-75.98)	0.019
CD8+	29.32 (21.27-43.22)	24.02 (19.92-32.93)	0.025
CD4+ EM4 PD-1+	2.41 (1.23-4.90)	4.10 (2.16-5.55)	0.080

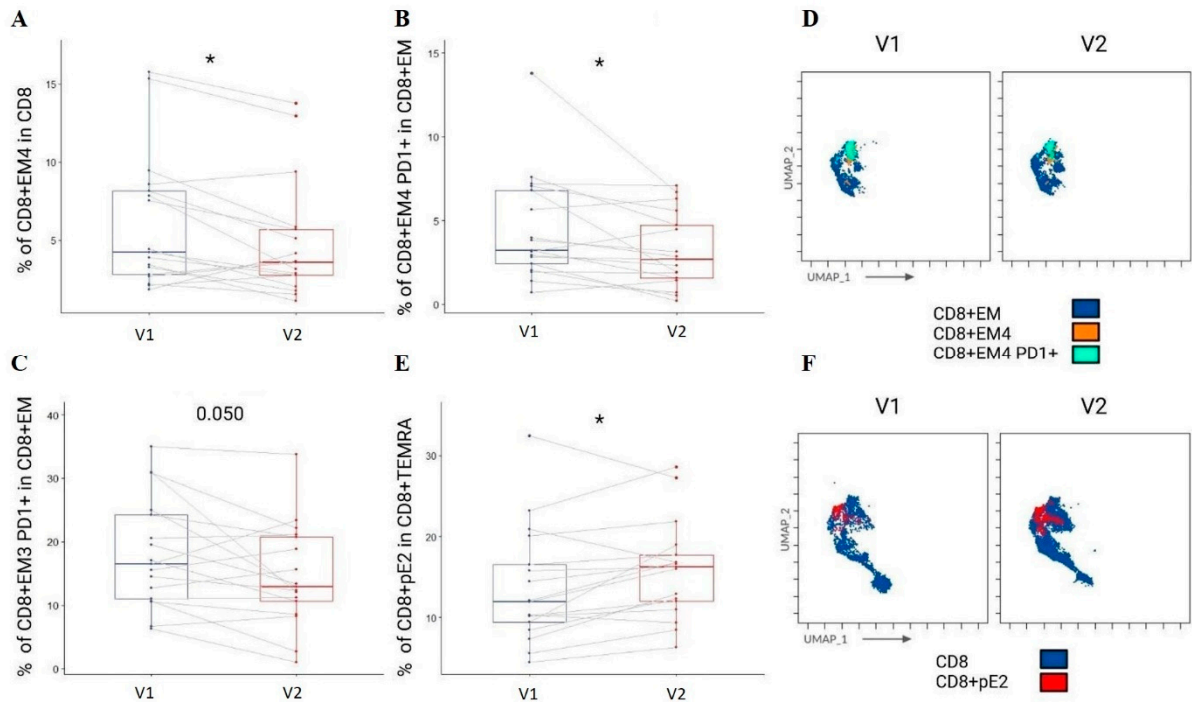
CD8+ EM2 CD57+PD-1+	3.27 (1.50-6.65)	4.47 (2.37-7.57)	0.019
CD8+ EM4 CD57+ PD-1+	0.45 (0.13-1.14)	0.23 (0.04-0.64)	0.089
CD8+ EM2 PD-1+	7.11 (3.98-10.92)	9.53 (6.09-16.08)	<0.001
CD8+ EM2	13.13 (8.19-17.82)	14.86 (10.66-24.760)	0.001
CD4:CD8	2.18 (1.22-3.49)	3 (1.89-3.70)	0.077

A Wilcoxon signed rank test was performed. All variables are shown as median (interquartile range).

Abbreviations: Cluster of differentiation (CD), effector memory (EM), programmed cell death protein 1 (PD-1).

Differences in the T-cell phenotypes of PLWH between V1 and V2

When running a univariate analysis, PLWH admitted with COVID-19 had higher percentages of CD8+ pre-effector 2 (pE2:CCR7-CD45RA+CD27+CD28+) at V2 (8 ± 1.45 days after admission) when compared to V1 (p-value=0.023). At the V2 timepoint, the percentage of CD8+ EM4 (p-value=0.015) subset, specifically activated CD8+ EM4 (p-value=0.011) T-cells decreased in PLWH. The percentage of activated CD8+ EM3 T-cells decreased, albeit not significantly (p-value=0.050). Supplementary Table S7 and Figure 2 show the differences found between hospital admission and 2.5 ± 3.37 days after admission in PLWH with COVID-19.



Supplementary Figure S2: Differences in the T-cell phenotypes in PLWH with COVID-19 at hospital admission (V1) and 2.5 ± 3.37 days after admission (V2). **A-C:** Boxplots showing a decrease in the percentages of CD8+ EM4, CD8+ EM4 expressing PD-1 and CD8+ EM3 expressing PD1 at the V2 timepoint. **D:** Representative UMAP plots show the decreases in the respective CD8+ EM T-cell populations between V1 and V2 in PLWH admitted with COVID-19. **E:** The percentage of CD8+ pE2 cells increased from V1 to V2. **F:** Representative UMAP plots show the increase in the CD8+ pre-Effector 2 TEMRA population between V1 and V2 in PLWH admitted with COVID-19.

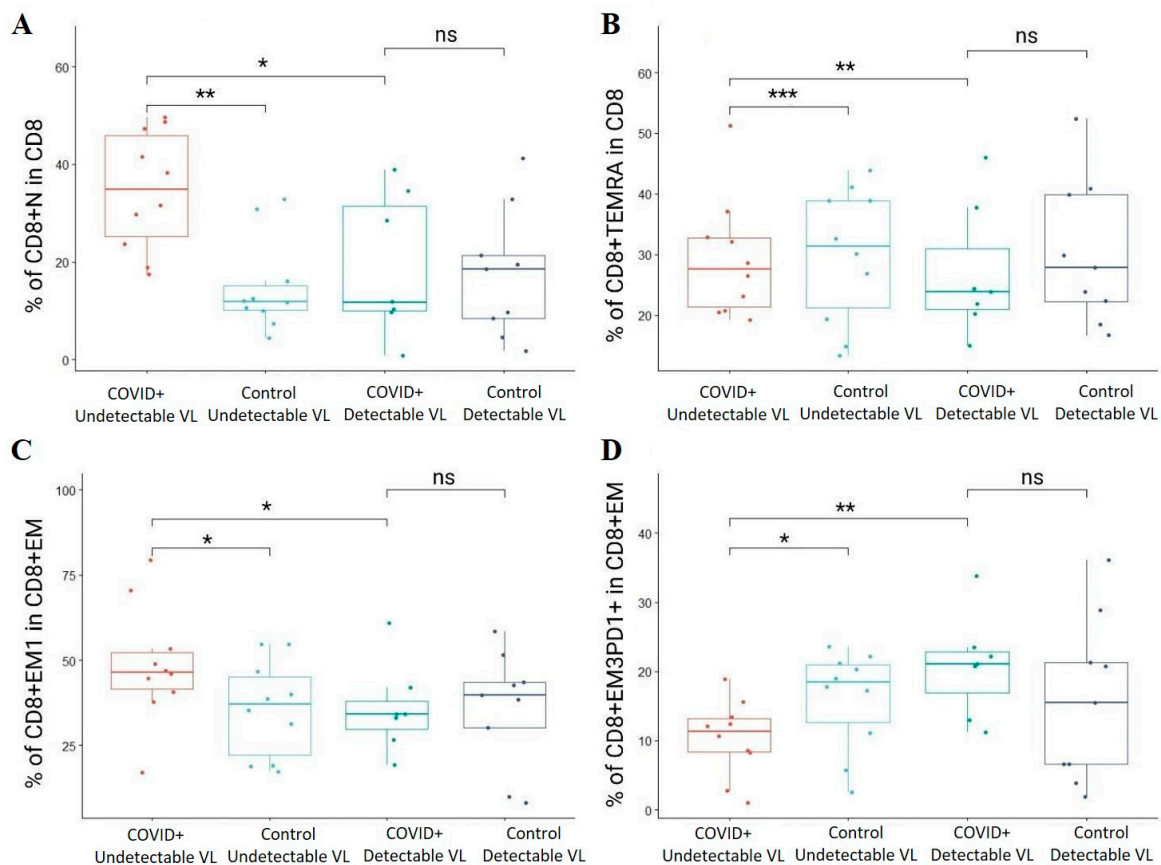
p-value: * = < 0.05.

Abbreviations: Effector memory (EM), pre-effector (pE), programmed cell death protein 1 (PD1), terminally differentiated T-cells re-expressing CD45RA (TEMRA), visit 1 (V1), visit 2 (V2)

Of the 17 PLWH with a V1 and V2 sample, just under half (8/17- 47.06%) had a CD4+ T-cell count of <200 cells/mm³ (median 120; IQR: 111-135). There was no significant difference in terms of age (47 ± 14.37 vs 48 ± 8.60 , p-value=0.736) and sex (p-value=0.402) between PLWH with a CD4+ T-cell count <200 cells/mm³ or >200

cells/mm³. No significant differences were present in terms of co-morbidities in PLWH with a CD4+ T-cell count <200 cells/mm³: Hypertension (3/8, p-value=0.858), diabetes (2/8, p-value=0.707), cardiovascular disease (1/8, p-value=0.600), pulmonary disease (0/8, p-value=0.331) and tuberculosis (2/8, p-value=0.110). PLWH with a CD4+ T-cell count <200 cells/mm³ were not hospitalized significantly longer than those patients with a higher CD4+ T-cell count [10 ± 4 days vs 8 ± 5 days, p-value=0.313]. The percentage of CD8+ T-cells was significantly higher at V2 in the group with a lower CD4+ T-cell count (p=0.034). The COVID-19 groups were compared to healthy PLWH with a CD4+ T-cell count of above or below 200 cells/mm³. Both PLWH COVID-19 groups had significantly higher percentages of activated CD4+ (CD4+ T-cell count <200 cells/mm³ vs control p-value=0.008, CD4+ T-cell count >200 cells/mm³ vs control p-value=0.002) and CD8+ (CD4+ T-cell count <200 cells/mm³ vs control p-value=0.008, CD4+ T-cell count >200 cells/mm³ vs control p-value<0.001) EM1 subsets when compared to controls. Supplementary Table S7 shows the comparison of the COVID-19 groups with each other.

Next, we analyzed PLWH with a detectable and undetectable viral load (VL). Ten (10/17-58.82%) PLWH admitted with COVID-19 had an undetectable VL (<20 copies/mL) and seven (7/10-41.18%) had a detectable VL. PLWH with a detectable VL were younger (37 ± 12 vs 52 ± 8 years of age, p-value=0.051). In terms of co-morbidities, patients with a detectable VL had significantly lower incidences of diabetes (0/7 vs 5/10, p-value=0.026) and higher incidences of heart disease (4/7 vs 0/10, p-value=0.023). At V2, PLWH with a detectable VL had higher percentages of CD8+ T-cells (p-value=0.023) and lower percentages of CD4+ T-cells (p-value=0.023). Of the CD8+ T-cell subsets, PLWH with detectable VL had significantly lower CD8+ naïve (p-value=0.015), TEMRA (p-value=0.004) and CD8+ EM1 (p-value=0.039) subsets. This group also had significantly higher percentages of activated CD8+ EM3 T-cells (p-value=0.005), specifically CD8+ EM3 T-cells co expressing PD-1 and CD57 (p-value=0.044).



Supplementary Figure S3: Boxplots depicting the differences between PLWH admitted to hospital with COVID-19 with a detectable and undetectable HIV VL, as well as with their respective SARS-CoV-2 negative controls at V2. **A:** PLWH with a detectable VL admitted with COVID-19 had significantly lower CD8+ naïve cells when compared with those that have an undetectable VL. PLWH with COVID-19 had a higher percentage of CD8+ naïve T-cells when compared to PLWH with an undetectable VL without COVID-19. **B:** PLWH with a detectable

VL had lower percentages of CD8+ TEMRA cells when compared to PLWH with an undetectable VL. PLWH with COVID-19 with an undetectable VL had lower percentages of CD8+ TEMRA T-cells when compared to PLWH with an undetectable VL without COVID-19. **C:** PLWH with a detectable VL had lower percentages of CD8+ EM1 T-cells than those with a detectable VL at V2. PLWH with COVID-19 with an undetectable VL had higher percentages of CD8+ EM1 T-cells when compared to PLWH with an undetectable VL without COVID-19. **D:** PLWH with a detectable VL had higher percentages of CD8+EM3 T-cells expressing PD-1 than those with an undetectable VL at V2. PLWH with COVID-19 with an undetectable VL had lower percentages of CD8+ EM3 T-cells expressing PD-1 when compared to PLWH with an undetectable VL without COVID-19.

p-value: * = < 0.05, ** = <0.01, *** = <0.001.

Abbreviations: Effector memory (EM), naïve (N), programmed cell death protein 1 (PD-1), terminally differentiated T-cells re-expressing CD45RA (TEMRA), viral load (VL)

When comparing the VL for each COVID-19 group with their respective controls without COVID-19, PLWH admitted with COVID-19 had lower percentages of CD4+ CM PD-1+ (COVID-19 + detectable VL vs detectable VL control p-value<0.001, COVID-19 + undetectable VL vs undetectable VL control p-value=0.002), CD4+ EM4 (COVID-19 + detectable VL vs detectable VL control p-value=0.009, COVID-19 + undetectable VL vs undetectable VL control p-value=0.002), CD4+ EM4 PD-1+ (COVID-19 + detectable VL vs detectable VL control p-value<0.001, COVID-19 + undetectable VL vs undetectable VL control p-value=0.002), CD4+ TEMRA (COVID-19 + detectable VL vs detectable VL control p-value=0.030, COVID-19 + undetectable VL vs undetectable VL control p-value=0.022) CD8+ N PD-1+ (COVID-19 + detectable VL vs detectable VL control p-value=0.020, COVID-19 + undetectable VL vs undetectable VL control p-value=0.016), CD8+ CM (COVID-19 + detectable VL vs detectable VL control p-value=0.024, COVID-19 + undetectable VL vs undetectable VL control p-value=0.034) T-cells. On the other hand, PLWH admitted with COVID-19 had significantly higher activated CD8+ EM1 (COVID-19 + detectable VL vs detectable VL control p-value=0.008, COVID-19 + undetectable VL vs undetectable VL control p-value<0.001) and CD8+ EM2 CD57+ T-cells.

Supplementary Table S7: Differences in the T-cell phenotypes between V1 and V2 for PLWH admitted to hospital with COVID-19

T-cell population	COVID+ PLWH V1 (n=17)	COVID+ PLWH V2 (n=17)	p-value
CD8+ EM	24.48 (16.43-35.95)	28.04 (25.72-40.84)	0.050
CD8+ E	77.98 (72.48-86.22)	76.61 (68.75-83.64)	0.093
CD8+ pE2	11.21 (9.43-16.51)	16.12 (10.97-17.72)	0.023
CD8+ EM4 PD1+	3.19 (2.41-6.81)	2.78 (1.58-4.71)	0.011
CD8+ EM3 PD1+	16 (10.68-24.22)	12.65 (8.53-20.77)	0.050
CD8+ EM2	15.79 (11.79-20.51)	16.83 (12.89-25.38)	0.050
CD8+ EM4	4.04 (2.79-8.15)	3.61 (2.76-5.66)	0.015

A Wilcoxon signed rank test was performed. All variables are shown as median (interquartile range).

Abbreviations: Effector memory (EM), pre-effector (pE), programmed cell death protein 1 (PD1).

Supplementary Table S8: Comparisons of each COVID-19 group between controls, as well as between PLWH admitted with COVID-19 with a CD4 count of above and below 200 cells/mm³ at the V2 timepoint after performing a Kruskal-Wallis test with post hoc Dunn's test.

T-cell population	CD4 >200 (n=9) Group 1	Control CD4 >200 (n=11) Group 2	CD4<200 (n=8) Group 3	Control CD4<200 (n=7) Group 4	1 vs 2	3 vs 4	1 vs 3	2 vs 4	p-value
DN	4.17 (3.15-5.87)	4.37 (2.73-7.44)	3.67 (2.86-4.78)	6.17 (5.41-9.02)	0.360	0.010	0.343	0.010	0.106
CD4	58.65 (49.47-62.10)	42.56 (26.12-54.93)	44.95 (23.81-53.74)	46.02 (14.85-64.11)	0.053	0.390	0.050	0.390	0.303
CD8	36.79 (31.75-43.60)	53.13 (39.30-64.98)	50.82 (42.11-70.74)	47.65 (28.00-70.52)	0.065	0.272	0.038	0.272	0.030
CD4+ TEMRA	0.12 (0.02-0.47)	0.55 (0.19-1.51)	0.02 (0-0.84)	0.55 (0.26-1.39)	0.044	0.024	0.355	0.024	0.073
CD4+ EM	11.97 (11.44-18.33)	23.80 (15.70-38.41)	15.40 (13.33-44.35)	13.57 (12.28-26.00)	0.003	0.281	0.085	0.281	0.041
CD4+ N	32.84 (26.61-36.70)	16.57 (6.34-33.17)	24.28 (9.09-32.69)	31.41 (19.04-40.83)	0.024	0.186	0.091	0.186	0.190
CD4+ CM PD1+	33.19 (25.75-39.73)	48.53 (46.08-74.17)	31.86 (29.26-35.85)	53.01 (46.08-74.18)	0.006	0.006	0.409	0.006	0.005
CD4+ EM4	13.08 (7.27-22.02)	28.75 (20.58-35.16)	12.23 (6.59-16.67)	38.73 (30.79-49.61)	0.014	<0.001	0.347	0.001	0.002
CD4+ EM1 PD1+	45.27 (29.43-48.86)	10.02 (7.00-34.37)	41.78 (29.43-53.96)	23.47 (8.31-26.39)	0.002	0.008	0.456	0.008	0.002
CD4+ EM4 PD1+	5.75 (2.68-8.44)	14.00 (9.79-17.21)	4.50 (3.84-5.70)	26.39 (17.65-30.09)	0.008	<0.001	0.288	<0.001	<0.001
CD8+ N	34.49 (17.44-41.61)	12.07 (9.73-18.61)	26.05 (14.26-33.97)	19.50 (8.38-32.89)	0.006	0.299	0.160	0.299	0.085
CD8+ TEMRA	23.83 (21.86-32.10)	38.86 (29.89-40.91)	26.49 (19.89-39.41)	18.52 (14.83-27.80)	0.042	0.098	0.362	0.097	0.059
CD8+ EM	26.54 (25.72-31.88)	38.44 (32.26-43.60)	37.38 (26.36-44.28)	38.74 (25.47-56.08)	0.021	0.322	0.105	0.322	0.190
CD8+ CM	7.24	10.83	5.15	13.84	0.122	0.007	0.098	0.007	0.039

	(5.04-14.86)	(8.08-12.69)	(3.31-8.75)	(5.90-17.05)					
CD8 N PD1+	7.00	20.07	5.33	17.77	0.034	0.010	0.320	0.010	0.031
	(4.67-15.16)	(7.60-25.39)	(2.70-8.98)	(10.48-30.58)					
CD8+ pE1	9.03	4.49	4.62	5.55	0.045	0.406	0.064	0.406	0.318
	(6.01-16.58)	(2.44-9.32)	(2.95-9.24)	(3.27-13.12)					
CD8+ pE1 PD1+	7.62	2.46	2.65	3.03	0.048	0.236	0.086	0.236	0.344
	(2.59-9.71)	(1.44-8.71)	(1.74-6.95)	(2.74-9.83)					
CD8+ E CD57+ PD1+	9.74	12.51	5.93	17.40	0.297	0.011	0.152	0.011	0.135
	(4.75-14.12)	(6.07-18.63)	(4.61-7.39)	(7.11-24.86)					
CD8+ EM2 CD57+	5.95	2.50	5.87	1.93	0.032	0.043	0.313	0.043	0.081
	(4.02-10.89)	(2.26-5.80)	(4.70-6.77)	(1.26-5.81)					
CD8+ EM3 CD57+ PD1+	9.82	8.03	7.36	16.31	0.333	0.072	0.261	0.072	0.474
	(5.81-15.74)	(4.75-14.08)	(5.21-10.13)	(3.50-22.65)					
CD8+ EM4 CD57+ PD1+	0.99	0.60	0.39	1.81	0.285	0.008	0.109	0.008	0.098
	(0.23-2.34)	(0.25-1.21)	(0.22-0.74)	(0.65-3.09)					
CD8+ EM4 PD1+	2.70	2.15	2.40	3.19	0.180	0.089	0.267	0.089	0.362
	(1.58-4.71)	(1.24-3.44)	(1.20-4.36)	(2.18-8.34)					
CD8+ EM3 PD1+	18.87	17.82	11.61	20.79	0.420	0.098	0.152	0.098	0.599
	(13.37-21.10)	(6.57-21.28)	(9.56-12.65)	(5.73-28.81)					
CD8+ EM1 PD1+	35.22	1.44	34.98	2.89	<0.001	0.003	0.452	0.003	<0.001
	(29.75-41.05)	(0.84-2.55)	(26.51-45.80)	(1.40-10.64)					
CD8+ EM1	44.65	31.38	39.18	42.76	0.031	0.396	0.279	0.396	0.259
	(34.21-48.89)	(18.82-40.09)	(30.34-53.43)	(38.40-54.63)					
CD8+ EM4	4.13	5.97	3.35	7.34	0.327	0.029	0.138	0.029	0.255
	(2.77-9.39)	(3.20-7.83)	(2.25-4.38)	(3.42-15.96)					
CD8+ CM PD1+	61.82	52.27	55.83	59.43	0.199	0.263	0.356	0.263	0.707
	(51.76-63.76)	(43.45-65.50)	(46.05-62.46)	(51.97-66.54)					

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range) Abbreviation: Central memory (CM), double-positive (DP), end-stage effector (E), effector memory (EM), naïve (N), pre-effector (pE), programmed cell death protein 1 (PD1), terminally differentiated T-cells re-expressing CD45RA (TEMRA).

Supplementary Table S9: Comparison of the COVID-19 groups by VL with each other, as well as their respective controls, at the V2 timepoint after performing a Kruskal-Wallis test with post hoc Dunn test.

T-cell population	Undetectable VL COVID (n=10) Group 1	Undetectable VL Control (n=10) Group 2	Detectable VL COVID(n=7) Group 3	Detectable VL Control (n=9) Group 4	1 vs 2	3 vs 4	1 vs 3	2 vs 4	p-value
CD4+	56.80 (49.47-60.30)	47.55 (26.30-64.19)	33.64 (13.08-53.87)	26.12 (12.83-46.02)	0.175	0.329	0.023	0.044	0.042
CD8+	39.47 (35.68-44.62)	48.80 (29.73-70.11)	59.66 (40.13-79.57)	63.88 (47.65-64.98)	0.181	0.475	0.023	0.092	0.089
CD4+ TEMRA	0.06 (0-0.74)	0.60 (0.19-1.51)	0.10 (0-0.47)	0.36 (0.29-1.39)	0.022	0.030	0.490	0.447	0.052
CD4+ EM	13.66 (9.94-15.59)	18.27 (13.96-27.33)	18.74 (11.83-69.97)	26.00 (17.93-31.79)	0.034	0.246	0.067	0.266	0.103
CD4+ CM PD1+	29.51 (25.75-33.81)	48.34 (38.90-59.15)	35.02 (29.89-49.80)	73.03 (47.67-74.17)	0.003	0.014	0.060	0.071	<0.001
CD4+EM4	12.00 (6.41-18.73)	30.74 (23.46-36.25)	13.08 (7.27-23.98)	28.75 (21.22-46.86)	0.002	0.009	0.473	0.453	0.003
CD4+ EM1 PD1+	46.99 (31.43-56.19)	9.16 (7.00-10.97)	33.33 (25.53-48.86)	24.74 (11.17-34.37)	<0.001	0.126	0.174	0.062	<0.001
CD4+ EM4 PD1+	4.53 (2.68-7.84)	14.58 (10.63-17.65)	4.55 (3.58-9.18)	20.94 (17.21-27.21)	0.002	<0.001	0.362	0.159	<0.001
CD8+ N	34.91 (23.67-47.36)	11.88 (9.98-16.12)	11.79 (9.69-34.49)	18.61 (8.38-21.35)	0.002	0.437	0.015	0.359	0.015
CD8+ TEMRA	27.56 (20.77-32.56)	31.34 (19.42-38.91)	23.83 (20.22-37.69)	27.80 (22.29-39.82)	<0.001	0.221	0.004	0.469	0.007
CD8+ CM	5.69 (4.25-13.31)	11.28 (8.08-15.19)	5.95 (3.33-11.79)	12.69 (10.69-16.12)	0.034	0.024	0.377	0.474	0.064
CD8+ N PD1+	6.50 (1.97-11.97)	15.58 (8.90-22.71)	5.83 (4.84-24.74)	18.24 (11.42-30.89)	0.016	0.020	0.295	0.229	0.018
CD8+ E CD57+ PD1+	6.63 (5.60-12.94)	16.40 (12.51-22.40)	5.33 (3.86-9.74)	7.11 (3.71-17.40)	0.036	0.324	0.370	0.054	0.160

CD8+ EM2 CD57+	5.43 (4.02-6.40)	2.41 (2.23-2.86)	6.49 (4.83-10.89)	3.80 (1.26-5.81)	0.020	0.046	0.442	0.379	0.068
CD8+ EM3 CD57+ PD1+	7.35 (5.02-9.82)	11.37 (3.49-16.20)	11.43 (5.81-19.37)	8.03 (4.75-18.36)	0.149	0.206	0.044	0.465	0.392
CD8+ EM4 PD1+	2.29 (0.71-3.16)	2.19 (1.87-3.19)	4.48 (1.94-6.68)	2.81 (1.24-7.26)	0.424	0.311	0.064	0.184	0.373
CD8+ EM3 PD1+	11.31 (8.27-13.37)	18.39 (11.09-21.14)	21.10 (12.95-23.43)	15.44 (6.55-21.28)	0.047	0.091	0.005	0.357	0.077
CD8+ EM2 PD1+	10.50 (9.35-12.06)	4.39 (2.99-22.86)	21.28 (12.46-28.06)	19.07 (5.67-33.25)	0.305	0.325	0.078	0.065	0.202
CD8+ EM1 PD1+	37.12 (34.97-42.26)	1.23 (0.84-1.84)	29.75 (22.86-37.40)	1.23 (0.84-1.84)	<0.001	0.008	0.219	0.152	<0.001
CD8+ EM1	46.39 (40.58-53.28)	37.00 (19.21-46.76)	34.12 (26.57-41.97)	39.69 (30.15-43.48)	0.047	0.363	0.039	0.451	0.229

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range).

Abbreviation: Central memory (CM), double-positive (DP), end-stage effector (E), effector memory (EM), naïve (N), pre-effector (pE), programmed cell death protein 1 (PD1), terminally differentiated T-cells re-expressing CD45RA (TEMRA).

Supplementary Table S10: Visit 2 differences in T-cell profiles between PLWH and those without HIV hospitalized with COVID-19 after performing a Kruskal-Wallis test with post hoc Dunn test.

T-cell population	COVID+PLWOH (n=52) Group 1	Control PLWOH(n=10) Group 2	COVID+PLWH (n=17) Group 3	Control PLWH (n=19) Group 4	1 vs 2	1 vs 3	3 vs 4	2 vs 4	p-value
DN	3.03 (2.06-4.29)	3.70 (2.74-5.39)	3.88 (2.77-5.25)	5.47 (3.66-7.44)	0.163	0.039	0.071	0.050	0.003
CD4+	69.59 (61.93-75.98)	73.01 (65.88-79.63)	52.72 (34.54-58.65)	42.56 (22.23-54.93)	0.292	<0.001	0.248	<0.001	<0.001
CD8+	24.02 (19.92-32.93)	22.92 (15.18-28.04)	43.11 (36.79-59.66)	53.13 (39.30-70.11)	0.296	<0.001	0.352	<0.001	<0.001
CD4+ TEMRA	0.05 (0.01-0.85)	0.15 (0.06-0.45)	0.068 (0-0.47)	0.55 (0.26-1.51)	0.262	0.341	0.003	0.065	0.013
CD4+ EM	11.82 (8.19-15.95)	7.76 (5.46-9.35)	14.44 (11.83-20.92)	23.36 (13.96-31.79)	0.034	0.0572	0.050	<0.001	<0.001
CD4+ CM	57.33 (50.69-63.76)	53.81 (44.70-58.86)	51.27 (47.25-56.57)	51.96 (44.45-59.22)	0.242	0.015	0.318	0.298	0.108
CD4+ CM PD1+	20.26 (15.02-26.38)	33.72 (24.64-37.91)	33.26 (28.11-39.73)	53.01 (45.61-73.03)	0.020	<0.001	0.007	0.003	<0.001
CD4+ EM PD1+	77.56 (70.79-83.57)	61.07 (50.99-70.59)	80.84 (76.81-85.14)	76.18 (65.12-82.58)	<0.001	0.114	0.054	0.008	0.002
CD4+ EM4	9.84 (7.13-14.04)	27.50 (26.18-33.72)	11.54 (6.78-18.73)	30.69 (21.22-38.94)	<0.001	0.200	<0.001	0.442	<0.001
CD4+ EM1	56.29 (43.49-68.43)	67.06 (47.22-73.00)	56.24 (38.82-65.97)	45.69 (20.73-54.26)	0.188	0.247	0.071	0.006	0.035
CD4+ EM3	19.49 (12.40-36.19)	2.25 (0.30-16.60)	18.26 (13.13-30.72)	16.79 (4.72-42.73)	0.002	0.348	0.368	0.021	0.033
CD4+ EM1 PD1+	41.62 (32.39-51.87)	23.75 (14.49-26.19)	44.50 (29.43-53.11)	10.97 (8.12-29.97)	<0.001	0.493	<0.001	0.397	<0.001
CD4+ EM4 PD1+	4.10 (2.16-5.55)	16.00 (12.97-20.17)	4.50 (3.41-7.84)	17.21 (10.63-26.39)	<0.001	0.147	<0.001	0.466	<0.001
CD8+ N	21.30	20.17	29.06	12.07	0.361	0.154	0.007	0.045	0.076

	(11.65-35.00)	(15.61-44.65)	(11.79-38.89)	(8.38-21.35)					
CD8+ EM	24.46	25.98	28.04	38.74	0.406	0.122	0.045	0.006	0.007
	(16.12-36.67)	(14.65-34.88)	(25.72-40.84)	(30.90-54.10)					
CD8+ CM	8.07	13.39	5.83	11.74	0.013	0.147	0.003	0.346	0.006
	(5.68-11.77)	(7.47-20.88)	(4.25-11.79)	(8.08-16.12)					
CD8+ N PD1+	6.40	8.41	6.33	18.24	0.380	0.458	0.001	0.008	0.001
	(2.86-12.43)	(3.84-11.43)	(2.57-9.00)	(9.32-30.58)					
CD8+ E	70.60	47.69	76.61	69.63	0.061	0.193	0.168	0.122	0.273
	(51.40-84.19)	(25.93-77.88)	(68.75-83.64)	(56.62-79.34)					
CD8+ pE2	17.59	28.34	16.12	19.13	0.066	0.255	0.069	0.296	0.213
	(10.04-26.09)	(16.15-34.67)	(10.97-17.72)	(11.34-33.11)					
CD8+ E CD57+	54.31	40.29	48.58	50.95	0.045	0.311	0.377	0.191	0.364
	(35.04-68.20)	(21.49-53.80)	(40.19-60.90)	(31.87-61.15)					
CD8+ E CD57+ PD1+	7.97	9.58	6.40	14.11	0.272	0.463	0.066	0.207	0.245
	(3.09-11.55)	(4.29-12.53)	(4.75-9.74)	(6.07-21.26)					
CD8+ EM2 CD57+	7.68	3.63	6.14	2.41	0.001	0.094	0.018	0.464	<0.001
	(4.87-11.77)	(2.23-5.25)	(4.49-7.42)	(1.93-5.80)					
CD8+ EM2 CD57+ PD1+	4.47	2.48	3.90	3.28	0.029	0.258	0.243	0.271	0.174
	(2.37-7.57)	(1.73-4.86)	(2.60-5.46)	(1.63-5.35)					
CD8+ EM3 CD57+ PD1+	4.63	4.35	7.36	9.30	0.381	0.084	0.479	0.097	0.265
	(1.89-13.91)	(1.85-9.49)	(5.02-11.23)	(3.49-16.31)					
CD8+ EM4 CD57+ PD1+	0.23	0.41	0.46	0.65	0.450	0.016	0.221	0.019	0.006
	(0.04-0.64)	(0.14-0.46)	(0.23-1.38)	(0.34-1.81)					
CD8+ EM4 PD1+	1.26	4.15	2.78	2.23	<0.001	0.003	0.460	0.021	<0.001
	(0.58-2.10)	(3.25-6.38)	(1.58-4.71)	(1.24-3.73)					
CD8+ EM3 PD1+	6.83	5.55	12.65	17.82	0.221	0.033	0.446	0.017	0.040
	(3.42-16.97)	(2.66-11.13)	(8.53-20.77)	(6.55-21.28)					
CD8+ EM2 PD1+	9.53	6.72	11.86	11.00	0.064	0.072	0.145	0.069	0.136
	(6.09-16.08)	(4.51-9.93)	(9.35-21.28)	(3.41-33.25)					
CD8+ EM1 PD1+	39.03	3.44	35.12	1.64	<0.001	0.272	<0.001	0.218	<0.001
	(29.08-50.17)	(2.64-5.13)	(29.75-42.26)	(0.86-3.29)					

CD8+ EM1	49.51 (38.25-64.36)	64.74 (59.49-70.49)	43.31 (34.12-53.28)	38.64 (19.21-46.76)	0.064	0.067	0.136	<0.001	0.002
CD8+ EM2	14.86 (10.66-24.760)	11.55 (7.28-14.80)	16.83 (12.89-25.38)	18.13 (8.69-43.22)	0.051	0.152	0.488	0.014	0.110
CD8+ EM4	4.08 (2.73-5.21)	7.68 (5.68-9.35)	3.61 (2.76-5.66)	5.97 (3.20-9.00)	0.001	0.373	0.075	0.041	0.009
CD8+ EM3	21.18 (8.69-40.23)	12.08 (5.38-23.72)	28.95 (22.64-39.28)	27.70 (12.99-47.45)	0.098	0.073	0.382	0.027	0.118
CD8+ CM PD1+	45.65 (38.63-54.42)	51.26 (42.11-54.99)	58.60 (46.87-65.42)	55.34 (44.71-65.50)	0.326	0.005	0.398	0.114	0.022

The Kruskal-Wallis test with post hoc Dunn's test was used to compare continuous variables between groups. All variables are shown as median (interquartile range)

Abbreviation: Central memory (CM), double-positive (DP), end-stage effector (E), effector memory (EM), naïve (N), pre-effector (pE), programmed cell death protein 1 (PD1), terminally differentiated T-cells re-expressing CD45RA (TEMRA).

S2.1 Extra flow cytometry information

S2.1.1 Panel design: Fluorescent antibodies used and compensation

Supplementary Table S11: Fluorescent reagents used in the T-cell panel.

Characteristic	Analyte	Detector	Reporter	Manufact.	Clone	Cat#
T helper	CD4	Anti-CD4	APC	Beckman Coulter	13B8.2	B53328
T cytotoxic	CD8	Anti-CD8	AF-700	Beckman Coulter	B9.11	B53328
T lineage marker	CD3	Anti-CD3	APC-750	Beckman Coulter	UCHT-1	B53328
Common leukocyte antigen	CD45RO	Anti-CD45RO	Krome Orange	Beckman Coulter	J33	B53328
Common leukocyte antigen	CD45RA	Anti-CD45RA	FITC	Beckman Coulter	2H4	B53328
T-cell homing marker	CD197 (CCR7)	Anti-CD197	PE	Beckman Coulter	G043H7	B53328
Co-stimulatory ligand to CD80 and CD86	CD28	Anti-CD28	ECD	Beckman Coulter	CD28.2	B53328
Co-stimulatory molecule	CD27	Anti-CD27	PC7	Beckman Coulter	1A4.CD27	B53328
T-cell exhaustion marker	CD57	Anti-CD57	Pacific Blue	Beckman Coulter	NC1	B53328
T-cell activation marker	CD279 (PD-1)	Anti-CD279	PC5.5	Beckman Coulter	PD1.3.5	B53328

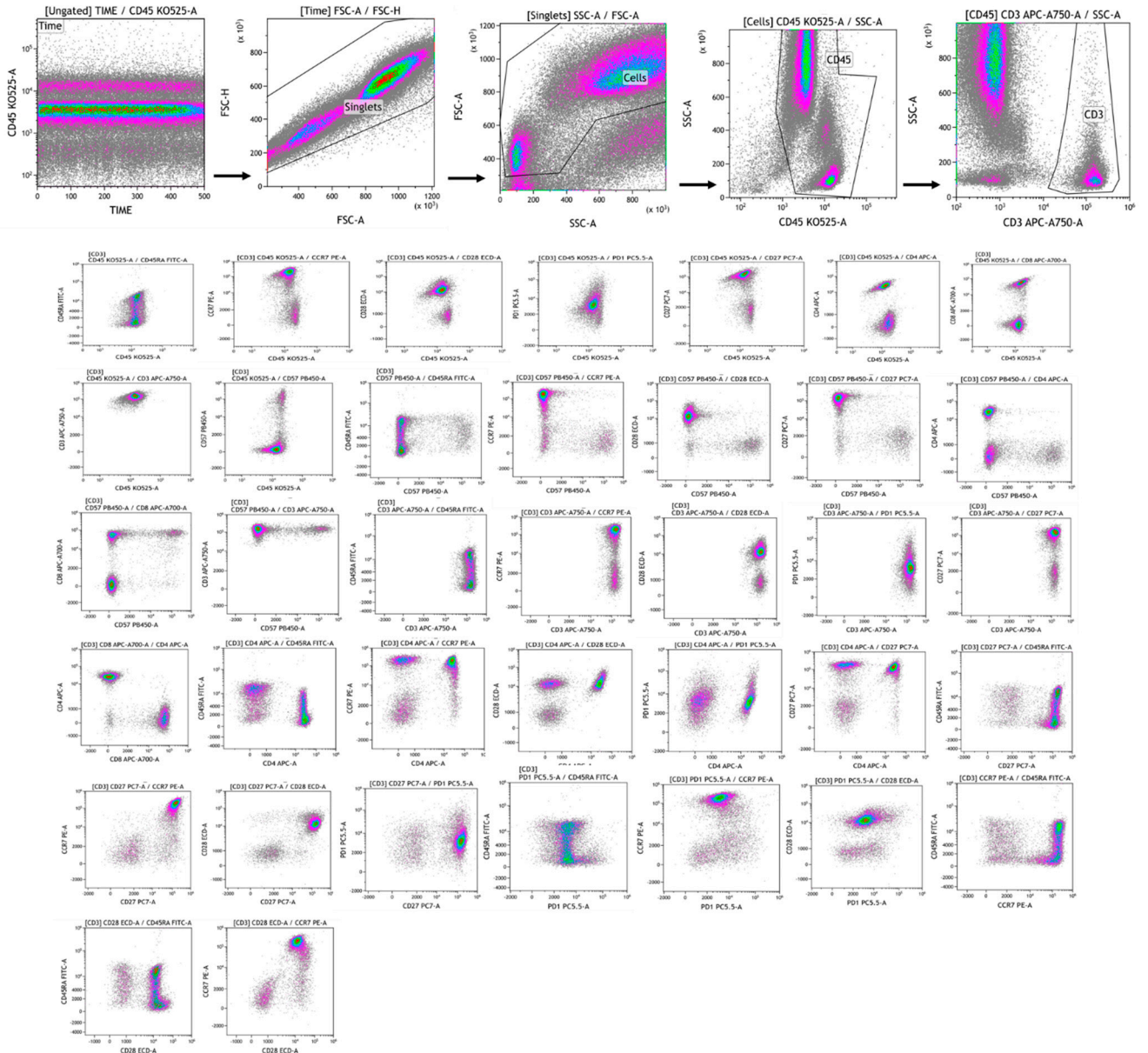
Abbreviations: Alexa fluor (AF), Allophycocyanin (APC), C-C chemokine receptor type 7 (CCR7), cluster of differentiation (CD), Phycoerythrin-Texas red conjugate (ECD), Fluorescein isothiocyanate (FITC), fluorescence channel (FL), Phycoerythrin cyanine (PC), Programmed cell death protein 1 (PD-1), Phycoerythrin (PE).

Compensation set up (Supplementary Table S12) was performed by utilizing the compensation kit provided as part of the Duraclone kit by staining whole blood according to the manufacturer's instructions. bi-exponential plots of scaling and compensation can be seen in Supplementary Figure S4.

Supplementary Table S12: Compensation tubes have been created as follows.

Optical detector	FL9	FL10	FL1	FL2	FL3	FL4	FL5	FL6	FL7	FL8
Reporter	Pacific Blue	Krome Orange	FITC	PE	ECD	PC5.5	PC7	APC	AF-700	AF750
Tube #1	+Ab	-	-	-	-	-	-	-	-	-
Tube #2	-	+Ab	-	-	-	-	-	-	-	-
Tube #3	-	-	+Ab	-	-	-	-	-	-	-
Tube #4	-	-	-	+Ab	-	-	-	-	-	-
Tube #5	-	-	-	-	+Ab	-	-	-	-	-
Tube #6	-	-	-	-	-	+Ab	-	-	-	-
Tube #7	-	-	-	-	-	-	+Ab	-	-	-
Tube #8	-	-	-	-	-	-	-	+Ab	-	-
Tube #9	-	-	-	-	-	-	-	-	+Ab	-
Tube #10	-	-	-	-	-	-	-	-	-	+Ab

Abbreviations: antibody (Ab), Alexa fluor (AF), Allophycocyanin (APC), Phycoerythrin-Texas red conjugate (ECD), Fluorescein isothiocyanate (FITC), fluorescence channel (FL), Phycoerythrin cyanine (PC), Phycoerythrin (PE).



Supplementary Figure S4: Example of data clean-up gating strategy and compensation for T-cells. Each axis was changed to logicle to ensure that all of the data can be seen on the plot. Each marker was plotted against one another to visualize the compensation before uploading to Cytobank.

S2.1.2 Cytobank analysis

S2.1.2.1 PeacoQC was used on all the FCS files uploaded to Cytobank

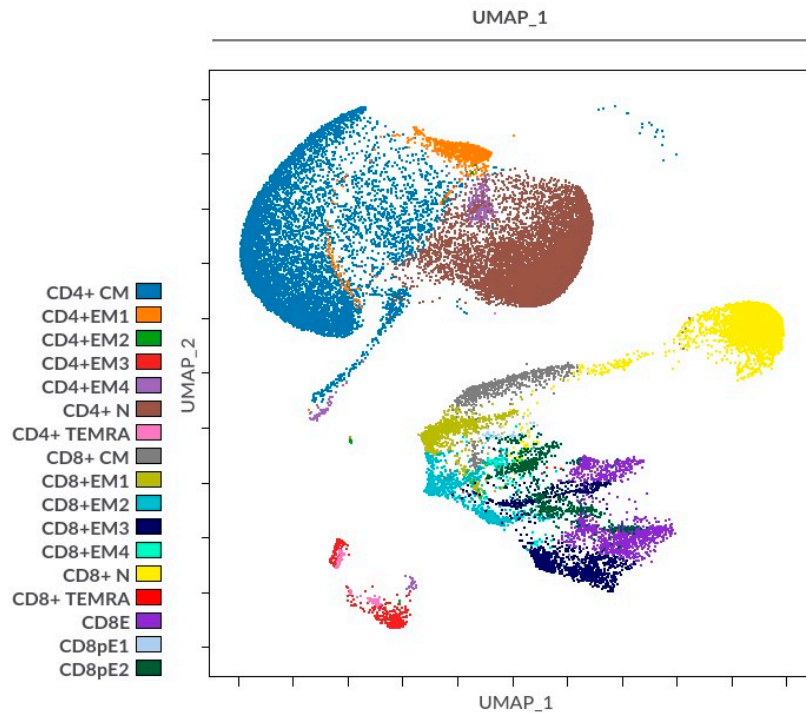
After uploading the files onto the Cytobank platform an additional quality check was performed by running the PeacoQC algorithm. PeacoQC identifies and removes any irregularities such as upregulated or downregulated signal during the acquisition of the data and creates a new file which it labels as a “Cleaned file”.

S2.1.2.2 UMAP was used to visualize and gate the data in Cytobank

On the “Cleaned files” generated by the PeacoQC algorithm, the lymphocyte population was generated by plotting SSC vs CD45RO (Supplementary Figure S6A). Next by plotting SSC vs CD3, the T-cell population was gated by gating on the CD3+ population. The CD3+ population was selected as the input parameter for the UMAP algorithm. UMAP settings included:

- Channels to be included: CCR7-PE, CD27-PC7, CD28-ECD, CD4-APC, CD8 APC-A700, CD45RA, CD57-Pacific Blue and PD1-PC5.5

- Events to be sampled: All events in CD3 gate (total of 3 580 511 across all files)
- Advanced settings included: Number of neighbors:20, minimum distance between clusters:0,01 and we selected to collapse outliers
- The data was normalized and the internal file compensation was applied



Supplementary Figure S5: Example of UMAP output generated of T-cell populations identified.

S2.1.2.3 CITRUS

CITRUS was used to identify significant biological differences between PLWH and those without HIV admitted with COVID-19.

After performing the UMAP analysis CITRUS was run as downstream analysis. Two CITRUS experiments were setup, one in which the input gate was CD4 and one in which the input gate was CD8. CITRUS performs statistical analysis thus the groups and files to be compared need to be specified. PLWH and people without HIV admitted with COVID-19 were matched for sex and age within STATA 17 and the exact matched files were selected to be compared with one another in the CITRUS experiment. Thus 36 files from both groups were selected (PLWH and people without HIV) and labelled “HIV+” and “HIV-”. Clustering channels chosen include: CCR7-PE, CD27-PC7, CD28-ECD, CD45RA, CD57-Pacific Blue and PD1-PC5.5. Internal file compensation was used (refer to above mentioned compensation matrix Supplementary Figure S4). Association model specified: Significance Analysis of Microarrays (SAM) – Correlative. Cluster characterization was set to abundance. Equal event sampling was selected.

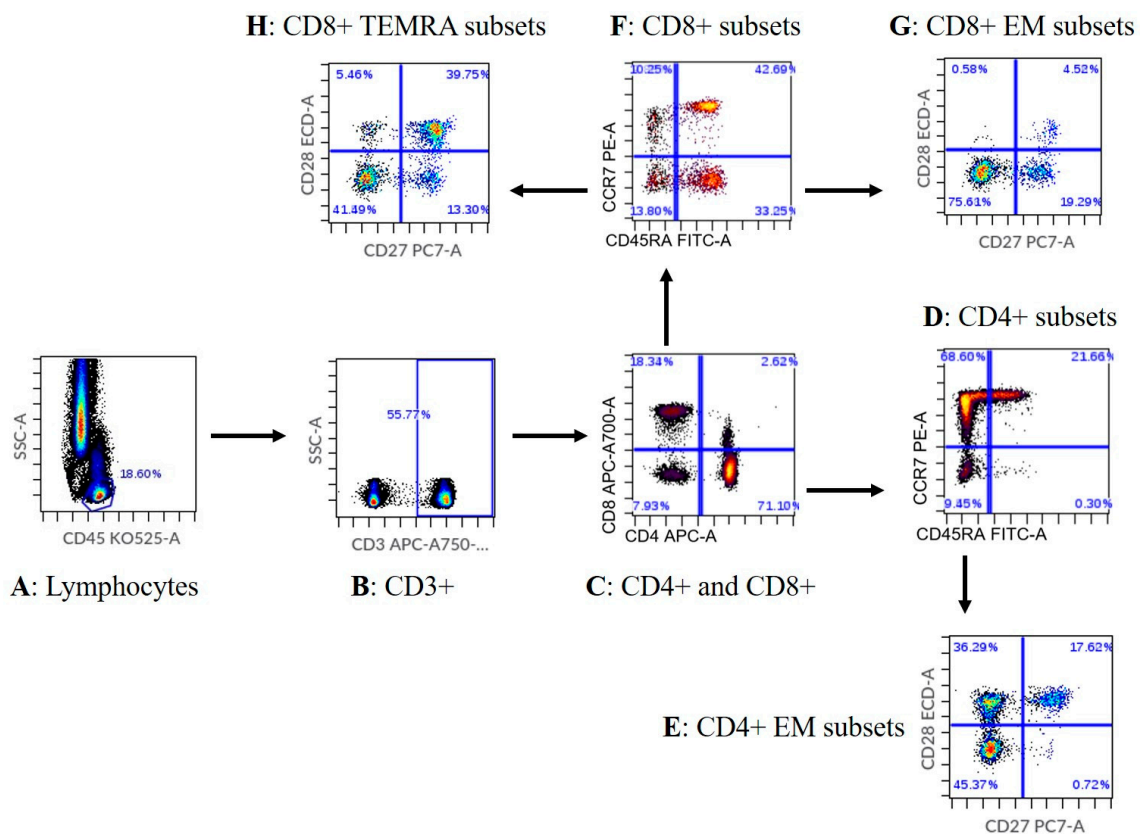
S2.1.2.4 Gate Description T-cell panel

The gating strategy involves the following gates:

- SSC vs CD45KO- lymphocyte gate to define the lymphocytes (Supplementary Figure S6A).
- SSC vs CD3- leukocyte gate applied- gate CD3 to define the T-cells (Supplementary Figure S6B).
- CD8 vs CD4- CD3 gate applied- create a quadrant gate to differentiate between CD4, CD8, DP and DN T-cells. (Supplementary Figure S6C).
- CD4 gate applied:
- CCR7 vs CD45RA to define the different CD4 memory subsets: naïve (CD4 N), central memory (CD4 CM), effector memory (CD4 EM) and terminal effector cells re-expressing CD45RA (CD4 TEMRA). (Supplementary Figure S6D).

-CD4 EM gate applied:
 -CD28 vs CD27 to define the different effector memory subsets: EM1 (CD4 EM1), EM2 (CD4 EM2), EM3 (CD4 EM3), EM4 (CD4 EM4). (Supplementary Figure S6E)
 - CD8 gate applied- CCR7 vs CD45RA to define the different CD8 memory subsets: naïve (CD8 N), central memory (CD8 CM), effector memory (CD8 EM) and terminal effector cells re-expressing CD45RA (CD8 TEMRA). (Supplementary Figure S6F).
 -CD8 EM gate applied:
 -CD28 vs CD27 to define the different effector memory subsets: EM1 (CD8 EM1), EM2 (CD8 EM2), EM3 (CD8 EM3), EM4 (CD8 EM4). (Supplementary Figure S6G)
 -CD8 TEMRA gate applied
 -CD28 vs CD27 to define the different TEMRA subsets pre-effector 1 (CD8 pE1), pre-effector 2 (CD8 pE2), end-stage effectors (CD8 E). (Supplementary Figure S6H)
 - The following gates were drawn on the UMAP1 vs UMAP2 plot by using the Z-channel coloring: CD4 CM PD1+, CD4 EM1 PD1+, CD4 EM2 PD1+, CD4 EM3 PD1+, CD4 EM4 PD1+, CD4 EM3 PD1+CD57+, CD8 CM PD1+, CD8 EM1 PD1+, CD8 EM2 PD1+, CD8 EM3 PD1+, CD8 EM1 CD57+, CD8 EM2 CD57+, CD8 EM3 CD57+, CD8 EM4 CD57+, CD8 TEMRA pE1 PD1+, CD8 TEMRA E PD1+, CD8 TEMRA E PD1+CD57+. (Supplementary Figure S6).

S2.1.2.5 Gate Boundaries



Supplementary Figure S6: Gating strategy followed to gate T-cell populations. A: SSC vs CD45KO-lymphocyte gate to identify the lymphocyte population. B: SSC vs CD3- leukocyte gate applied- gate CD3 to describe the T-cell population. C: CD8 vs CD4- CD3 gate applied- a quadrant gate was created to differentiate between CD4, CD8, DP and DN T-cells. D: CCR7 vs CD45RA to gate the different CD4 memory subsets Naïve (CD4 N), central memory (CD4 CM), effector memory (CD4 EM) and terminal effector cells re-expressing CD45RA (CD4 TEMRA). E: CD28 vs CD27 to define the different effector memory subsets EM1 (CD4 EM1), EM2 (CD4 EM2), EM3 (CD4 EM3), EM4 (CD4 EM4). F: CCR7 vs CD45RA to define the different CD8 memory subsets CD8 N, CM, EM and TEMRA. G: CD28 vs CD27 to define the different effector memory subsets EM1,

EM2 , EM3 , EM4. H: CD28 vs CD27 to define the different TEMRA subsets pre-effector 1 (pE1), pre-effector 2 (pE2), end-stage effectors (E).