# Haematopoietic stem cell transplantation in South Africa: Current limitations and future perspectives

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The growing need for haematopoietic stem cell transplantation (HSCT) is reflected in the increasing number of transplants performed globally each year. HSCT provides life-changing and potentially curative therapy for a range of pathologies including haematological malignancies; other indications include certain congenital and acquired disorders of the haematopoietic system, autoimmune conditions and hereditary diseases. The primary goals of HSCT are either to replace haematopoietic stem and progenitor cells (HSPC) following myeloablative chemotherapy or to cure the original pathology with allogeneic HSPCs. Success depends on optimal outcomes at various stages of the procedure including mobilisation of marrow stem/progenitor cells for harvesting from the patient or donor, long-term and sustainable engraftment of these cells in the recipient, and prevention of graft-versus-host disease in the case of allogeneic HSCT. Challenges in South Africa include high cost, limited infrastructure and lack of appropriately trained staff, as well as limitations in securing suitable haematopoietic stem cell donors. This review aims to provide an overview of HSCT and some of the challenges that are faced in the South African context.

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Allogeneic haematopoietic stem cell transplantation (HSCT) involves the transfusion of donor haematopoietic stem and progenitor cells (HSPC) procured from bone marrow, peripheral blood or the umbilical cord into a suitably primed recipient. Autologous haematopoietic stem cell transplantation, on the other hand, denotes the collection of the patient's own stem cells prior to high-dose myeloablative chemotherapy, which are subsequently re-infused as a stem cell rescue to facilitate recovery of the marrow function. HSCT was pioneered by E D Thomas in 1957.[1] At that time, this procedure provided the first evidence that bone marrow had the capacity to regenerate the haematopoietic system and effectively treat or alleviate disorders of the same origin. Since 1957, HSCT has formed part of the treatment regimen of more than one million patients. [2] The European Group of Blood and Marrow Transplantation (EBMT) annually conducts a survey on HSCT activity in order to observe and monitor trends in Europe and affiliated countries. Data collected by these surveys provide evidence on the growing need for this procedure, with the number of HSCTs performed annually increasing from 4 234 in 1990 to over 40 000 in 2017.[3] There are no statistics currently available for the number of transplants performed in South Africa (SA), nor data reporting the need for this treatment or patient burden, and thus the deduction of an increased demand can be made on observed global trends. Schonfeld et al.[4] reported that 14 662 patients were diagnosed with haematological malignancies between 2000 and 2006, being responsible for 6% of the new cancer diagnoses and 8% of the cancer deaths reported in SA. These data were collected from the National Cancer Registry, which has certain caveats in that there may be underreporting of these cancers owing to diagnostic and reporting challenges in rural and lower socio-economic communities. A regional study conducted

by Oelofse and Truter<sup>[5]</sup> found that haematological malignancies were not uncommon in Eastern Cape Province, and the incidence was comparable to some European populations. Additionally, certain black and Afrikaner population groups of SA are at greater risk of inheriting Fanconi's anaemia owing to the genetic founder effect of the mutant Fanconi-associated genes.[6]

Indications for the procedure have expanded to encompass treatment of not only haematological and non-haematological malignancies, but also congenital and other acquired disorders of the haematopoietic system, as well as autoimmune and hereditary diseases (Table 1).[7]

Based on data collected from the World Health Organization (WHO)-defined global regions, Gratwohl et al.[2] reported that 58% of all HSCTs performed were autologous whilst 42% were allogeneic. Furthermore, the highest number of HSCTs were performed in Europe, followed by the Americas, Southeast Asia and the Western Pacific. The lowest number of HSCTs were performed in Eastern Mediterranean countries and Africa. Additionally, HSCTs were not carried out in countries with populations of less than 300 000 or with a gross national income less than USD1 260 per capita. [2] These findings were attributed to the high costs associated with the procedure, limiting the availability of this treatment option in developing nations. These facts raise pertinent ethical questions regarding distributive justice, encompassing the limited access of lower socio-economic populations in sub-Saharan Africa to this treatment modality. This procedure is life-changing as it (i) counters the toxic effects of treatment regimens that are used to reduce tumour load; (ii) lengthens the time to relapse; or (iii) may potentially cure disorders (such as allogeneic HSCT for aplastic anaemia or acute myeloid leukaemia (AML)) that may otherwise prove fatal.

Disorder	Autologous HSCT	Allogeneic HSCT
Haematological malignancies	Acute myeloid leukaemia	Acute myeloid leukaemia and myelodysplastic syndrome
	Multiple myeloma	Primary myelofibrosis
	Primary amyloidosis	Chronic myeloid leukaemia
	Acute lymphoblastic lymphoma	Acute lymphoblastic leukaemia
	Hodgkin's lymphoma	Hodgkin's lymphoma
	Non-Hodgkin's lymphoma	Non-Hodgkin's lymphoma
Other haematological disorders		Aplastic anaemia
		Paroxysmal nocturnal haemoglobinuria
		Severe combined immunodeficiency
Autoimmune diseases	Multiple sclerosis	
	Systemic lupus erythematosis	
	Crohn's disease	
	Rheumatoid arthritis	
Hereditary disorders		Fanconi anaemia
		Sickle cell anaemia
		Thalassaemia major
		Wiskott-Aldrich syndrome
		Diamond-Blackfan anaemia
HSCT = haematopoietic stem cell transplantation.		

In a single-centre study, Remberger et al.[8] reported a 70% 3-year overall survival rate in patients who received HSCT for a range of pathologies. Svenberg et al. [9] reported an overall survival rate of 82% for malignant and 96% for non-malignant disorders in paediatric patients 5 years after receiving the HSCT. As technology and skill have improved in the HSCT field, mortality rates have dropped from 82% between 1992 and 2000 to 46% by the end of 2009 as a composite for all eligible pathologies.[8]

Therapeutic success of the procedure depends on successful mobilisation of HSPCs from the marrow (either the patient's marrow or a suitable donor), long-term and sustainable engraftment of the transplanted cells, and the prevention of graft-versus-host disease in the case of allogeneic HSCT.

### Haematopoietic stem and progenitor cells

Embryonic haematopoiesis begins in the yolk sac and continues in the aorta-gonad-mesonephros region as well as at multiple other locations throughout gestation. [10] Adult haematopoiesis occurs predominantly in the bone marrow and may include extramedullary locations such as the liver and spleen.[11] HSPCs are multipotent cells responsible for the production and maintenance of all blood cell lineages.[12] These self-renewing cells firstly differentiate into common myeloid (CMP) and common lymphoid (CLP) progenitor cells (Fig. 1).[10] The adult HSPC niche (micro-environment that protects and sustains HSPC development, mobilisation and homing) is located in bone marrow along endosteal surfaces of trabecular bone in close proximity to endothelial cells. The latter allows for gaseous diffusion and nutrient movement between the intravascular compartment and the niche.[13]

# Haematopoietic stem cell transplantation

The essential component of HSCT is the HSPCs which can be collected directly from bone marrow, peripheral blood through apheresis or umbilical cord blood. [14] Guidelines set out to identify and enumerate HSPCs were consolidated in 1996 by the International Society of Hematotherapy and Graft Engineering (ISHAGE), [15]

recently renamed the International Society for Cell and Gene Therapy (ISCT). These guidelines remain the gold standard. HSPCs are identified and enumerated based on expression of the CD34 surface protein and the dimly expressed common leukocyte antigen CD45.[15] HSPCs are also characterised by the absence of lineagespecific markers.

The HSCT procedure may be divided into several stages, as illustrated in Fig. 2.[16] The fundamental principle behind HSCT in haematopoietic maligancy is the administration of HSPCs after myeloablative treatment, with the aim of having the donor stem cells home to their niche in the bone marrow to regenerate the haematopoietic system (stem cell rescue).

# Disease-specific selection of patients for transplant and eligibility for transplant

HSCT is indicated as a primary treatment modality in patients diagnosed with various pathologies, or as second-line therapy when conventional protocols have failed or are ineffective (Table 1). Patient suitability is determined by several physiological and pathological factors (Table 2). These include cardiac, hepatic, pulmonary and renal function evaluations, viral and other infection screens, and the determination of the comorbidity index score. [17] Failure to meet these criteria may be seen as relative contraindications, as they may be superseded by expert opinion based on advances in medication, supportive care and technology.

HSCT is subject to identification of a suitable source for the acquisition of stem cells. In the case of autologous HSCT, a patient's stem cells may not adequately mobilise into the peripheral circulation, yielding inadequate numbers to proceed with the protocol. Allogeneic HSCT may not be feasible if there are no available donors. The latter constitutes a particular challenge in the South African setting, and is highlighted later in this article.

## **Mobilisation**

To facilitate harvesting, HSPCs are released from their marrow niche into the peripheral circulation by mobilisation. Mobilisation occurs when neutrophils, activated by granulocyte colony-stimulating

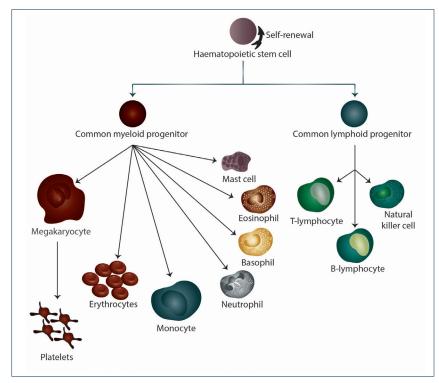


Fig. 1. Schematic representation of haematopoiesis. Haematopoiesis is the process during which the various blood cells are produced and regenerated. Multipotent haematopoietic stem cells have the ability to self-regenerate and subsequently differentiate into two distinct cell lineages, myeloid and lymphoid. The common myeloid progenitor differentiates into platelet-forming megakaryocytes, erythrocytes, monocytes, neutrophils, eosinophils, basophils and mast cells. The common lymphoid progenitor differentiates into T- and B-lymphocytes and natural killer cells).



Fig. 2. The sequence of haematopoietic stem cell transplantation (HSCT) in haematopoietic malignancies. Once haematopoietic stem cell transplantation is clinically indicated, stem cells are mobilised from their marrow niche using medical mobilising agents in either the patient (autologous) or a suitable donor (allogenic). Haematopoietic stem and progenitor cells are harvested, most commonly by apheresis. The conditioning phase encompasses treatment with high-dose chemotherapeutic agents and/or total body irradiation with the aim of achieving myeloablation. This is followed by infusion of the donor stem cells. The aplastic phase in which patients do not have a functional haematopietic system lasts between 10 and 20 days during which they are nursed in protective isolation until engraftment is achieved. Thus the desired outcome is a patient with a fully functioning haematopoietic system without the initial pathology).

factors (G-CSF), release proteolytic enzymes such as elastase and cathepsin G. These enzymes cleave the terminal amino sequence of stromal cell-derived factor-1 (SDF-1) attached to the C-X-C chemokine receptor type 4 (CXCR-4) on HSPCs which tether the stem cells to their marrow niche.[18] G-CSF is the agent most frequently used for mobilisation as a single agent in allogeneic donors. Autologous stem cell collection requires that the patient concurrently receives a chemotherapeutic agent such as cyclophosphamide, as this augments the number of mobilised CD34+ cells.[19] Cyclophosphamide causes the release of proteases which cleave key adhesion molecules such as CXCR-4.[19] CD34+ counts within the peripheral circulation are determined 3 hours prior to the first apheresis procedure. This is done either approximately 5 days after G-CSF for allogenic donors, or 14 days postmobilisation for patients who have been treated with G-CSF in combination with chemotherapy.[20] Mobilisation agents may also be used in combination with CXCR-4 antagonists such as plerixafor (mozobil), if first-line regimens fail.[20] A minimum threshold in the pre-harvest blood of 2.5  $\times$ 

106 × CD34+ cells/kg predicts a satisfactory harvest yield.[21]

# **Factors influencing** autologous haematopoietic stem and progenitor cell mobilisation

Factors which have been linked to poor HSPC mobilisation in autologous stem cell donors include advanced age, prior radiation and malignant infiltration. Exposure to purine analogues (fludarabine), alkylating agents (melphalan) and immunomodulators (lenalidomide) has also been associated with a negative prediction of mobilisation.[22] 'Proven' poor mobilisers are characterised by circulating CD34+ cells peaking at less than 20 CD34+ cells/µL up to 6 days after mobilisation with G-CSF alone, up to 20 days after G-CSF in combination with chemotherapy, or if there is a yield of less than 2.0 × 106 CD34+cells/kg collected with 3 apheresis attempts.[23] 'Predicted' poor mobilisers have a history of previously failed collection attempts, have had prior treatment with chemotherapeutic or radiation regimens that affect HSPC mobilisation, and if any two of the following criteria are met: advanced disease treated with more than 2 lines of chemotherapy, refractory disease, extensive bone marrow infiltration, marrow cellularity less than 30% at the time of mobilisation, or age >65 years (Table 3).[23]

## Factors influencing allogeneic donor suitability

Eligibility of donors largely depends on matching human leukocyte antigen (HLA) alleles, which are heterogeneous between individuals and races. Donors for allogeneic transplants may be sourced either from consenting family members (HLA-matched related donors if they have identical HLA alleles, or haploidentical if they have halfmatched HLA genotypes) or from unrelated HLA-matched donors (non-familial). Finding a suitable donor remains a challenge for many patients. The odds of finding an HLA-matched related donor is approximately 1 in 4 and this depends on family size and structure. This drops significantly (as low as 1 in 100 000) when the search extends to HLA-matched unrelated donors. The HLA genes are located on chromosome 6 and are some of the most polymorphic genes in the human genome, resulting in large variability of expression.[24] Pertinently, the SA population is genetically highly diverse in respect to HLA alleles, particularly in people of African descent.[25] Although certain HLA

Parameter	Recommendation	
General parameters		
Maximum age	75 years	
Karnofsky performance index	≥60 - 70	
Left ventricular ejection fraction	≥45%	
Heart rhythm	No uncontrolled arrhythmias	
Secondary malignancy	None evident	
Pregnancy tests	Negative	
Uncontrolled infections	Not evident	
Lung function		
FEV <sub>1</sub> /FVC ratio	≥60%	
Diffusion capacity	≥50%	
Liver function		
Serum bilirubin	≤34 μmol/L	
ALT/AST	≤2 × normal	
Viral hepatitis screen		
Negative	Preferred	
Positive	Not an absolute contraindication, but patients with a positive screen may	
	risk viral reactivation following a HSCT	
HIV screen		
Negative	Preferred	
Positive	HIV-positive patients with a CD4 count $>100/\mu L$	
	A low to undetectable viral load, <10 000 copies/mL	
HCT-CI	Recommended score of 0 - 2	
FEV $_{_{\parallel}}$ = forced expiratory volume in 1 second; FVC = forced vital capacities comorbidity index. Adapted from Hamadani <i>et al.</i> <sup>[17]</sup>	$city; ALT = alanine\ aminotransferase;\ AST = aspartate\ aminotransferase;\ HCT-CI = haematopoietic\ cell\ transplantation-like the contraction of the contraction $	

Proven poor mobilisers	Predicted poor mobilisers
Circulating CD34 <sup>+</sup> cell peak <20/μL for up to 6 days after mobilisation with G-CSF alone	Previous failed collection attempts
Circulating CD34 <sup>+</sup> cell peak <20/μL up to 20 days after G-CSF in combination with chemotherapy	Prior treatment with chemotherapeutic or radiation regimens that affect HSPC mobilisation
A yield $<$ 2.0 $\times$ 10 $^6$ CD34 $^+$ cells/kg after 3 apheresis collections	Two of the following criteria:  • advanced disease treated with >2 lines of chemotherapy  • refractory disease  • extensive bone marrow infiltration  • bone marrow cellularity <30% at the time of mobilisation  • age >65 years
G-CSF = granulocyte colony-stimulating factors; HSPC = haematopoietic stem and progenitor cel	ls.

alleles may be associated more frequently with particular population groups, it is the expression of uncommon population-specific alleles that further decreases the chances of identifying suitable donors, even in international registries. [26]

There are additional challenges in the South African context to providing successful HSCT to all patients in need of the treatment. HSCTs are predominantly performed in the private medical sector, which limits the access to this treatment modality to patients on higher-end medical insurance (17% of the SA population<sup>[27]</sup>) or to those who have the financial means. The high cost involved in accessing international donors is therefore a limitation, owing to the same reason.<sup>[28]</sup> The past few years have seen an increase in government-funded hospitals providing HSCT, but there is at present only one academic centre with a fully-fledged HSCT programme for allogeneic (including haploidentical) transplantation. Van Eyssen *et al.*<sup>[28]</sup> assessed the outcomes of 48 state-funded children

who received allogeneic HSCTs at the Groote Schuur Hospital/University of Cape Town Private Academic Hospital transplant unit. The authors aimed to evaluate the clinical outcomes of allogeneic HSCT in order to motivate future allocation of the substantial resources required to sustain such a treatment programme in SA. The overall survival in children treated for non-oncological diseases was 91% (median follow-up of 3.94 years) and 57% (median follow-up of 1.86 years) for children treated for haematological malignancies. These observations were comparable to outcomes reported in highincome counterpart facilities, [28] and justify allocating the country's limited resources to additional state-funded programmes.

Towards the aim of facilitating more equitable access to this treatment option, the South African Bone Marrow Registry (SABMR), <sup>[29]</sup> an internationally recognised non-profit organisation endeavouring to find suitable donors for critically ill children and adults in SA, had as of 2017 recruited more than 73 000 registered

donors. The SABMR provides services to patients both locally and internationally. Their collaboration with the World Marrow Donor Association (WMDA) provides further access for patients to a database of over 32 million international donors. The SABMR is coordinating a drive to sign on more local donors in order to meet the complex requirements of SA's genetically highly diverse population more comprehensively. [26] The Sunflower Fund[30] is an independent not-for-profit organisation founded as a donor recruitment centre and registry. It is actively involved in the education of the population to raise awareness regarding the need for South African donors, and assists in acquiring funds for patients unable to afford the costs of the HSCT. It has created collaborations with international stem cell/bone marrow registries, including those of other African countries, such as Ghana.

In the past decade, the use of haploidentical donors has become a well-recognised and successful practice should a sibling or matched unrelated donor not be unavailable.<sup>[31]</sup> For African and mixed-race patients, a matched unrelated donor is particularly difficult to source on any donor registry. This is due to ethnic diversity and unique genetic determinants making the likelihood of an international donor match unlikely. Additionally, there is a dearth of local volunteer donors in these population groups. The pool of suitable donors may be further restricted owing to the high prevalence of the human immunodeficiency virus (HIV) in the SA population (13.1% according to StatsSA).<sup>[32]</sup> A haploidentical donor from consenting first- or second-degree relatives is available immediately, and is considerably less costly to procure.

# Conditioning

Preparative or conditioning regimens in haematological malignancies aim to achieve three goals: (i) to reduce tumour load (treat the underlying condition); (ii) to facilitate sufficient immunoablation to prepare a niche for new marrow formation; and (iii) to suppress immune function sufficiently to prevent graft rejection and graftversus-host disease. Conditioning is specifically tailored to each pathology and is administered either prior to or after mobilisation as well as post HSPC collection. Regimens involve combinations of multiple chemotherapeutic agents, total body irradiation (TBI), corticosteroids and other disease-specific pharmaceuticals. Conventional conditioning regimens range from myeloablative to non-myeloablative, based on the dose of chemotherapy and/or TBI.[33] The latter encompass myeloablative and antineoplastic alkylating agents such as melphalan, busulphan and cyclophosphamide, as monotherapy or in various combinations. For example, busulphancontaining agents or fludarabine (a purine antimetabolite) may be used in combination with cyclophosphamide to reduce cyclophosphamidespecific cellular toxicity.[34,35] The development of reduced-intensity conditioning (RIC) regimens with diminished adverse chemotoxicity while promoting immunomodulation has improved certain patient outcomes, particularly in the elderly. [36,37] The choice to employ a specific regimen is based on the balance between immunosuppressive and myelosuppressive properties and the desired outcome in the management of the specific pathology. For instance, a combination of cyclophosphamide and 12 Gy TBI used in myeloablative regimens is highly myeloablative and immunosuppressive. Reduction of the TBI to 5.5 Gy would be classified as a reduced intensity regimen. Fludarabine, cyclophosphamide and 2 Gy TBI may be used in non-myeloablative regimens for their immunosuppressive effects. [38] Myeloablative protocols have increased toxicity profiles and rely less on the graft-versus-malignancy (GVM) effect to eliminate residual disease. The reduced-intensity and non-myeloablative regimens are less toxic and rely more on the GVM effect to eradicate residual disease while reducing graft-versus-host interactions.[38]

# **Engraftment**

An infusion of 2  $\times$  106 harvested CD34+ cells/kg is considered the minimum number of transplanted HSPCs needed to reliably ensure sustained engraftment. Engraftment success is monitored by neutrophil and platelet counts measured daily post transplant. Successful neutrophil engraftment is defined as an absolute neutrophil count of more than  $0.5 \times 10^9$  neutrophils/L for 3 consecutive days. Platelet engraftment success is attained when the non-transfusion-related platelet count is more than  $20 \times 10^9$  platelets/L for 3 consecutive days. Onsecutive days. Consensus in the literature considers the number of viable CD34+ cells transplanted to be the primary indicator of engraftment success or failure, with higher numbers being associated with a more favourable prognosis.

#### Graft-versus-host disease

Graft-versus-host disease (GVHD) is defined as a syndrome in which immunocompetent donor immune cells react to and destroy host tissues in immunocompromised recipients. [40] Lazaryan et al.[41] investigated potential risk factors and found that mobilised peripheral blood as a graft source was associated with a higher incidence of GVHD. Advanced recipient age was associated with a higher incidence of acute and chronic GVHD. Furthermore, conditioning regimen intensity appears to have an influence, with non-myeloablative conditioning regimens increasing the risk for acute GVHD.[41] GVHD is initiated by tissue damage induced by the chemotherapeutic regimen prior to the transplant, and is intensified as T cell activation and clonal proliferation orchestrate cytokines, chemokines and direct cellular toxicity in an exaggerated imflammatory reaction. [42] Damage-associated molecular patterns (DAMPs) lead to the activation of host antigen presenting cells (APCs). The activated APCs subsequently present host antigens to the donor's T cells, resulting in the release of pro-inflammatory cytokines that include tumour necrosis factor-α, γ-interferon and interleukin (IL)-1 and IL-12, which exacerbate tissue damage. [43]

Acute GVHD presents during the treatment phase with involvement of the skin, liver and gastrointestinal tract. Diverse clinical presentations include exfoliative and/or ulcerative dermatitis, liver dysfunction, and severe vomiting and diarrhoea. The mechanisms involved in the pathophysiology of chronic GVHD remain unclear. Cells and cytokines associated with chronic GVHD include CD4+ T cells, CD8+ T cells and IL-1/6/10. Chronic GVHD may develop without a history of, or clinically diagnosed, acute GVHD. Onset of chronic GVHD occurs roughly 100 days post transplantation, and involves the skin, mucous membranes including the vulva and/or vagina and mouth, liver, kidneys, heart and eyes. Symptoms and signs may include erythema, dermal and subcutaneous sclerosis, oedema, cytopenia, nephrotic syndrome, liver dysfunction, exocrine pancreatic insufficiency and cardiomyopathy.

First-line treatment for acute GVHD includes corticosteroids either as monotherapy or in combination with other agents such as cyclosporine, with the starting dose being dependent on the clinical severity. [47] Second-line treatments for steroid-refractory chronic GVHD may include extracorporeal photopheresis (ECP) or immune modulators such as methotrexate, rituximab and mycophenolate mofetil. [47] Serotherapy encompasses antibodies that target T cells and other immune cells that would theoretically modulate GVHD intensity. Agents such as antithymocyte globulin and alemtuzumab, a monoclonal antibody to CD52, have shown promise in prophylaxis and/or treatment of this complication, but absence of unequivocal long-term benefits, a lack of consensus on treatment regimens and their own unwanted effects render these treatments of little benefit

in steroid-refractory GVHD currently.[47] Alemtuzumab effectively reduces the frequency and severity of GVHD owing to its capacity to deplete competent donor T cells, and can be used as GVHD prophylaxis. [48] Most SA centres employ this GVHD preventive agent. Immune-suppressing drugs such as tacrolimus, sirolimus, cyclophosphamide, methotrexate and mycophenolate mofetil may also be used in prophylaxis regimens.

A desirable phenomenon known as the GVM effect, may occur as donor T cells recognise and destroy residual malignant cells within the recipient. GVM is observed in allogeneic HSCT.[49] A potential strategy to utilise this effect therapeutically may be through the use of T-regulatory cells (Tregs; also known as T-suppressor cells). Tregs develop in the thymus and function as important immune modulators of self-tolerance.<sup>[50]</sup> Patients infused with Tregs have a lower risk of developing GVHD. [51,52] Infusions of mesenchymal stromal cells, found in various tissues including the bone marrow, have also been used successfully to reduce the occurrence of GVHD.[53,54] These treatments are still experimental and more research is needed to define their physiological mechanisms and respective actions.

# **Transplant-related infections**

Transplantation-related microbial infections are a leading cause of morbidity and mortality post HSCT. Infections may be bacterial, viral and/or fungal and may vary between patients. Allogeneic HSCT patients should ideally undergo a pre-transplantation assessment of IgG antibodies to syphylis, toxoplasmosis and specific viral diseases to identify the potential risk of disease re-activation. Syphilis, tuberculosis, Toxoplasma gondii and hepatitis B and C viruses may persist for life after primary infection and can be reactivated post HSCT.[55] Viral infection post HSCT is commonly caused by cytomegalovirus, and less commonly by human herpes virus-6, parainfluenza virus, varicella zoster virus, respiratory syncytial virus and herpes simplex virus. Viral prophylaxis post HSCT may include aciclovir, valacyclovir or valganciclovir. Of these, aciclovir is used most frequently.<sup>[56]</sup> Invasive fungal infections include Candida, Aspergillus, Mucorales and Pneumocystis jirovecii. Fluconazole, voriconazole or posaconazole may be prescribed as prophylaxis. The prophylaxis for *P. jirovecii* includes trimethoprim-sulfamethoxazole, dapsone or atovaquone.<sup>[56]</sup> HIV is not a contraindication for HSCT and antiretroviral therapy may be continued post-transplantation if the toxcitity permits.  $^{\hbox{\scriptsize [55]}}$  Bacteraemias are caused by Gram-positive bacteria (such as coagulase-negative Staphylococci, Enterococci) and Gramnegative organisms such as Pseudomonas aeruginosa. [56] Antibiotic prophylaxis, as with ciprofloxacin, has been demonstrated to effectively reduce the incidence of Gram-negative sepsis, provided the microbiological resistance profile is favourable.<sup>[55]</sup>

# New frontiers in allogeneic transplantation

The emergence of RIC regimens has moved the upper age limit of myeloablative transplantation well past the 55-year cut-off formerly used for myeloablative regimens. Likewise, the barrier to performing haploidentical transplantation has been removed by utilising posttransplant cyclophosphamide to prevent graft failure and graftversus-host disease.[57] Two South African challenges in providing HSCTs are the high incidence of HIV in potential donors, as well as the genetic diversity of the population, which reduces the probability of finding a matched unrelated donor in local or international registries. Over the past 3 years, haploidentical HSCT has been successfully implemented in South African centres for patients with no recourse to fully matched sibling or volunteer donors.

#### Conclusion

With the increasing need for HSCT, it remains a challenge to establish and adequately staff HSCT programmes, particularly in the state healthcare setting in SA. This leads to implicit rationing of healthcare, whereby only patients in proximity to, or referred to HSCT centres, have access to this life-changing medical procedure. With improved treatment algorithms and increased survival rates following HSCT, there will be an additional need for infrastructure and skills required for long-term follow-up programmes post HSCT.[58] It is essential that all these factors are considered in policy making and planning to ensure ethical and equitable allocation of resources. Key objectives to achieve public-private parity of HSCT services include an increase in the number of clinical haematologist training posts and government support for establishment of fully resourced HSCT centres in every province. This task will also include raising the profile of HSCT in SA by advocacy based on haematological treatment successes as well as the creation of sustainable pathways to care. These endeavours will be advanced by the establishment of a national reporting body to determine the number of HSCTs performed in SA on a yearly basis. The data, however, would not provide an accurate representation on the definitive need of our population, as they would be biased toward patients with access to existing stem cell services and would be affected by demographic and financial constraints. Challenges in the diagnosis and reporting of pathologies, particularly in rural and resource-poor communities, will need to be addressed. To mitigate the population-specific genetic variability present in HLA-genes, recruitment of donors especially from African and mixed-race population groups is encouraged by the SABMR. However, the prevalence of HIV, and the high migration rate in SA make acquiring volunteer donors challenging. Haploidentical transplantation is a necessary tool that has to be implemented by every HSCT centre in SA to ensure equitable and affordable access to HSCT for all patients, but especially for the ethnic subpopulations where other donor pathways are limited. Favourable outcomes have been observed with haploidentical transplantation, and this may offer a suitable alternative in the absence of fully matched suitable donors,. Not-for-profit organisations such as the Sunflower Fund aim to recruit 100 000 donors of African ancestry by 2030 as part of the Stem Cell Registry Alliance (SCRA), and increase education and awareness in our population.

The benefits of HSCT are undeniable as it remains one of the only curative options for a variety of haematological pathologies, thereby decreasing morbidity and mortality in affected patients. In the coming years, we need to examine and improve upon the local systems in place regarding HSCT so that we can address the lack of access to HSCT in the majority of the population. The ultimate aim would be to establish a sustainable and cost-effective model to allow all South Africans in need of this intervention to have access to the required facilities.

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- 1. Thomas ED, Lochte Jr HL, Lu WC, et al. Intravenous infusion of bone marrow in patients receiving radiation and chemotherapy. N Engl J Med 1957;257(11):491-496. https://doi.org/10.1056/nejm195709122571102
- Gratwohl A, Pasquini MC, Aljurf M, et al. One million haemopoietic stem-cell transplants: A retrospective observational study. Lancet Haematol 2015;2(3):91-100. https://doi.org/10.3410 /f.726040527.793530721
- 3. Passweg JR, Baldomero H, Basak GW, et al. The EBMT activity survey report 2017: A focus on allogeneic HCT for nonmalignant indications and on the use of non-HCT cell therapies. Bone Marrow Transplant 2019. https://doi.org/10.1038/s41409-019-0465-9
- 4. Schonfeld SJ, Erdmann F, Wiggill T, et al. Hematologic malignancies in South Africa 2000 2006: Analysis of data reported to the National Cancer Registry. Cancer Med 2016;5(4):728-738. https:// doi.org/10.1002/cam4.597
- 5. Oelofse D, Truter I. Incidence of haematological malignancies, Eastern Cape Province; South Africa, 2004-2013. Cancer Epidemiol 2018;53:166-171. https://doi.org/10.1016/j.canep.2018.01.016
- 6. Feben C, Wainstein T, Kromberg J, Essop F, Krause A. Fanconi anaemia in South Africa: Past, present and future. S Afr Med J 2018;108(5):393-398. https://doi.org/10.7196/samj.2018.v108i5.13004
  7. Sureda A, Bader P, Cesaro S, et al. Indications for allo- and auto-SCT for haematological of the state of
- atological diseases, solid tumours and immune disorders: Current practice in Europe, 2015. Bone Marrow Transplant 2015;50(8):1037-1056. https://doi.org/10.1038/bmt.2015.6
- 8. Remberger M, Ackefors M, Berglund S, et al. Improved survival after allogeneic hematopoietic stem cell transplantation in recent years. A single-center study. Biol Blood Marrow Transplant
- 2011;17(11):1688-1697. https://doi.org/10.1016/j.bbmt.2011.05.001
  9. Svenberg P, Remberger M, Uzunel M, et al. Improved overall survival for pediatric patients undergoing allogeneic hematopoietic stem cell transplantation – a comparison of the last two decades. Pediatr Transplant 2016;20(5):667-674. https://doi.org/10.1111/petr.12723
- Seita J, Weissman IL. Hematopoietic stem cell: Self-renewal versus differentiation. Wiley Interdiscip Rev SystBiol Med 2010;2(6):640-653. https://doi.org/10.1002/wsbm.86
- 11. Ho MSH, Medcalf RL, Livesey SA, et al. The dynamics of adult haematopoiesis in the bone and bone arrow environment. Br J Haematol 2015;170(4):472-486. https://doi.org/10.1111/bjh.13445
- Weiskopf K, Schnorr PJ, Pang WW, et al. Myeloid cell origins, differentiation, and clinical implications. Microbiol Spectr 2016;4(5). https://doi.org/10.1128/microbiolspec.mchd-0031-2016
- 13. Zhang J, Niu C, Ye L, et al. Identification of the haematopoietic stem cell niche and control of the niche size. Nature 2003;425(6960):836-841. https://doi.org/10.1038/r
- Copelan EA. Hematopoietic stem-cell transplantation. N Engl J Med 2006;354(17):1813-1826. https://doi.org/10.1056/nejmra052638
- 15. Sutherland DR, Anderson L, Keeney M, et al. The ISHAGE guidelines for CD34+ cell determination by
- flow cytometry. J Hematother 1996;5(3):213-226. https://doi.org/10.1089/scd.1.1996.5.213
  16. Saccardi R, Gualandi F. Hematopoietic stem cell transplantation procedures. Autoimmunity 2008;41(8):570-576. https://doi.org/10.1080/08916930802197776
- Hamadani M, Craig M, Awan FT, et al. How we approach patient evaluation for hematopoietic stem cell transplantation. Bone Marrow Transplant 2010;45(8):1259-1268. https://doi.org/10.1038/bmt.2010.94
- 18. Kopp H-G, Avecilla ST, Hooper AT, et al. The bone marrow vascular niche: home of HSC differentiation and mobilization. Physiology 2005;20(5):349-356. https://doi.org/10.1152/physiol.00025.2005
- 19. Tuchman SA, Bacon WA, Huang L-W, et al. Cyclophosphamide-based hematopoietic stem cell mobilization before autologous stem cell transplantation in newly diagnosed multiple myeloma. J Clin Apher 2015;30(3):176-182. https://doi.org/10.1002/jca.21360
- Yuan S, Wang S. How do we mobilize and collect autologous peripheral blood stem cells? Transfusion 2017;57(1):13-23. https://doi.org/10.1111/trf.13868
- 21. Tiwari AK, Pandey P, Subbaraman H, et al. Autologous peripheral blood stem cell harvest: Collection efficiency and factors affecting it. Asian J Transfus Sci 2016;10(1):93-97. https://doi.org/10.4103/0973-
- 22. Tekgündüz E, Arat M, Göker H, et al. Autologous hematopoietic progenitor cell mobilization and collection in adult patients presenting with multiple myeloma and lymphoma: A position-statement from the Turkish Society of Apheresis (TSA). Trans Apher Sci 2017;56(6):845-849. https://doi. ci.2017.11.025
- 23. Mohty M, Hübel K, Kröger N, et al. Autologous haematopoietic stem cell mobilisation in multiple nyeloma and lymphoma patients: A position statement from the European Group for Blood Marrow Transplantation. Bone Marrow Transplant 2014;49(7):865-872. https://doi.org/10.1038/
- 24. The South African Bone Marrow Registry. The Development of Bone Marrow Transplantation in South Africa. http://www.sabmr.co.za/wp-content/uploads/2009/07/P-Ruff.pdf (accessed 20 November 2018).
- 25. Mellet J, Gray CM, Pepper MS. HLA typing: Conventional techniques v. next-generation sequencing. S Afr Med J 2015;106(1):88-91. https://doi.org/10.7196/samj.2016.v106i1.9571
- Tshabalala M, Mellet J, Pepper MS. Human leukocyte antigen diversity: A Southern African perspective. J Immunol Res 2015;2015:1-11. https://doi.org/10.1155/2015/746151
- 27. Statistics South Africa. General Household Survey 2017. http://www.statssa.gov.za/publications/ P0318/P03182017.pdf (accessed 20 November 2018).
- 28. Van Eyssen A, Hendricks M, Davidson A, et al. Single-centre experience of allogeneic haemopoietic stem cell transplant in paediatric patients in Cape Town, South Africa. S Afr Med J 2017;107(3):232-238. https://doi.org/10.7196/samj.2017.v107i3.11313
- 29. The South African Bone Marrow Registry. The South African Bone Marrow Registry 2017 Annual https://sabmr.co.za/newsite/wp-content/uploads/2018/11/AR-2017-FINAL-16-Oct\_.pdf (accessed 20 November 2018).

- The Sunflower Fund. https://www.sunflowerfund.org/ (accessed 13 March 2019).
- 31. Bashey A, Zhang X, Sizemore CA, et al. T-cell-replete HLA-haploidentical haematopoietic transplantation for haematologic malignancies using post-transplantation cyclophosphamide results in outcomes equivalent to those of contemporaneous HLA-matched related and unrelated donor transplantation. J Clin Oncol 2013;31:1310-1316. https://doi.org/10.1200/jco.2012.44.3523
- 32. Statistics South Africa. Midyear Population Estimates 2018. https://www.statssa.gov.za/publications/ P0302/P03022018.pdf (accessed 13 March 2019).
- 33. Gyurkocza B, Sandmaier BM. Conditioning regimens for hematopoietic cell transplantation: One size does not fit all. Blood 2014;124(3):344. https://doi.org/10.1182/blood-2014-02-514778
- 34. Pasquini MC, Le-Rademacher J, Zhu X, et al. Intravenous busulfan-based myeloablative conditioning regimens prior to hematopoietic cell transplantation for hematologic malignancies. Biol Blood Marrow Transplant 2016;22(8):1424-1430. https://doi.org/10.1016/j.bbmt.2016.04.013
  35. Katsanis E, Sapp LN, Pelayo-Katsanis L, et al. Alternative donor hematopoietic cell transplantation
- conditioned with myeloablative busulfan, fludarabine, and melphalan is well tolerated and effective against high-risk myeloid malignancies. J Pediatr Hematol Oncol 2016;38(8):e315-e318. https://doi. org/10.1097/mph.0000000000000621
- 36. Gay F. Engelhardt M. Terpos E, et al. From transplant to novel cellular therapies in multiple myeloma: European Myeloma Network guidelines and future perspectives. Haematologica 2018;103(2):197-211. https://doi.org/10.3324/haematol.2017.174573

  37. William BM, de Lima M. Advances in conditioning regimens for older adults undergoing allogeneic
- stem cell transplantation to treat hematologic malignancies. Drugs Aging 2013;30(6):373-381. https:// doi.org/10.1007/s40266-013-0076-x
- 38. Baron F, Storb R. Allogeneic hematopoietic cell transplantation following nonmyeloablative conditioning as treatment for hematologic malignancies and inherited blood disorders. Mol Ther 2006;13(1):26-41. https://doi.org/10.1016/j.ymthe.2005.09.011
- D'Rozario J, Parisotto R, Stapleton J, et al. Pre infusion, post thaw CD34+ peripheral blood stem cell enumeration as a predictor of haematopoietic engraftment in autologous haematopoietic cell  $transplantation.\ Trans\ Apher\ Sci\ 2014; 50(3): 443-450.\ https://doi.org/10.1016/j.transci.2014.02.021$
- Billingham RE. The biology of graft-versus-host reactions. Harvey Lect 1966;62:21-78.
- 41. Lazaryan A, Weisdorf DJ, DeFor T, et al. Risk factors for acute and chronic graft-versus-host disease after allogeneic hematopoietic cell transplantation with umbilical cord blood and matched sibling donors. Biol Blood Marrow Transplant 2016;22(1):134-140. https://doi.org/10.1016/j.bbmt.2015.09.008
- 42. Magenau J, Runaas L, Reddy P. Advances in understanding the pathogenesis of graft-versus-host disease. Br J Haematol 2016;173(2):190-205. https://doi.org/10.1111/bjh.13959
- 43. Holtan SG, Pasquini M, Weisdorf DJ. Acute graft-versus-host disease: A bench-to-bedside update. Blood 2014;124(3):363-373. https://doi.org/10.1182/blood-2014-01-514786
- 44. Hymes SR, Alousi AM, Cowen EW. Graft-versus-host disease: Part I. Pathogenesis and clinical manifestations of graft-versus-host disease. J Am Acad Dermatol 2012;66(4):515. https://doi. org/10.1016/j.jaad.2011.11.960
- 45. Holtick U. Albrecht M. Chemnitz IM, et al. Bone marrow versus peripheral blood allogeneic haematopoietic stem cell transplantation for haematological malignancies in adults. Cochrane Database Syst Rev 2014;(4). https://doi.org/10.1002/14651858.cd010189.pub2
- 46. Schultz K. Pathophysiology of chronic graft versus host disease. Chronic Graft-Versus-Host Disease: Interdisciplinary Management 2009;1:17-30. https://doi.org/10.1017/cbo9780511576751.004
- 47. Garnett C, Apperley JF, Pavlů J. Treatment and management of graft-versus-host disease: Improving response and s org/10.1177/2040620713489842 and survival. Ther Adv Hematol 2013;4(6):366-378. https://doi.
- Novitzky N, Thomas V, du Toit C. Prevention of graft vs. host disease with alemtuzumab 'in the bag' decreases early toxicity of stem cell transplantation and in multiple myeloma is associated with improved long-term outcome. Cytotherapy 2008;10(1):45-53. https://doi.org/10.1080/1465324070173277
- Gratwohl A, Sureda A, Cornelissen J, et al. Alloreactivity: The janus-face of hematopoietic stem cell transplantation. Leukemia 2017; 31(8):1752-1759. https://doi.org/10.1038/leu.2017.79
- Edinger M, Hoffmann P, Ermann J, et al. CD4+CD25+ regulatory T cells preserve graft-versus-tumor activity while inhibiting graft-versus-host disease after bone marrow transplantation. Nat Med 2003;9(9):1144-1150. https://doi.org/10.1038/nm915
  51. Brunstein CG, Miller JS, McKenna DH, et al. Umbilical cord blood-derived T regulatory cells to
- prevent GVHD: Kinetics, toxicity profile and clinical effect. Blood 2015;127(8):1044-1051. https://doi. org/10.3410/f.725934129.793546241
- 52. Di Ianni M, Falzetti F, Carotti A, et al. Tregs prevent GVHD and promote in HLA-haploidentical transplantation. Blood 2011;117(14):3921-3928. https://doi.org/10.1182/
- 53. Introna M, Lucchini G, Dander E, et al. Treatment of graft versus host disease with mesenchymal stromal cells: A phase I study on 40 adult and pediatric patients. Biol Blood Marrow Transplant 2014;20(3):375-381. https://doi.org/10.1016/j.bbmt.2013.11.033
- 54. Peng Y, Chen X, Liu Q, et al. Mesenchymal stromal cells infusions improve refractory chronic graft versus host disease through an increase of CD5+ regulatory B cells producing interleukin 10. Leukemia 2015;29(3):636-646. https://doi.org/10.1038/leu.2014.225
- 55. Ullmann AJ, Schmidt-Hieber M, Bertz H, et al. Infectious diseases in allogeneic haematopoietic stem cell transplantation: Prevention and prophylaxis strategy guidelines 2016. Ann Hematol 2016;95(9):1435-1455. https://doi.org/10.1007/s00277-016-2711-1
- Schuster MG, Cleveland AA, Dubberke ER, et al. Infections in hematopoietic cell transplant recipients: Results from the organ transplant infection project, a multicenter, prospective, cohort study. Open Forum Infect Dis 2017;4(2):ofx050. https://doi.org/10.1093/ofid/ofx050
- 57. Pagliardini P, Harbi S, Furts S, et al. Post-transplantation cyclophosphamide-based haploidentical versus Atg-based unrelated donor allogeneic stem cell transplantation for patients younger than 60 years with hematological malignancies: A single-center experience of 209 patients. Bone Marrow Transplant 2018. https://doi.org/10.1038/s41409-018-0387-y
- 58. Gajewski JL, LeMaistre CF, Silver SM, et al. Impending challenges in the hematopoietic stem cell transplantation physician workforce. Biol Blood Marrow Transplant 2009;15(12):1493-1501. https:// doi.org/10.1016/j.bbmt.2009.08.022