Pharmacogenetics of CYP2B6, CYP2A6 and UGT2B7 in HIV treatment in African populations: focus on efavirenz and nevirapine

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Abstract
The CYP450 and UGT enzymes are involved in phase I and phase II metabolism of the majority of clinically prescribed drugs, including the non-nucleoside reverse transcriptase inhibitors, efavirenz and nevirapine, used in the treatment of HIV/AIDS. Variations in the activity of these enzymes due to gene polymorphisms can affect an individual’s drug response or may lead to adverse drug reactions. There is an inter-ethnic distribution in the frequency of these polymorphisms, with African populations exhibiting higher genetic diversity compared to other populations. African specific alleles with clinical relevance have also emerged. Given the high prevalence of HIV/AIDS in sub-Saharan Africa, understanding the frequency of pharmacogenetically relevant alleles in populations of African origin, and their impact on efavirenz and nevirapine metabolism, is becoming increasingly critical. This review aims to investigate ethnic variation of CYP2B6, CYP2A6 and UGT2B7, and to understand the pharmacogenetic relevance when comparing frequencies in African populations to other populations worldwide.

Introduction
Drug metabolism and pharmacogenomics
Pharmacogenomics and pharmacogenetics take into consideration an individual’s genotype during drug development and drug prescription, in order to personalize therapy. Commonly, pharmacogenetics refers to how variation in a single gene influences an individual’s response to a single drug, while pharmacogenomics refers to how all of the genes in the genome can collectively influence responses to drugs (https://www.pharmgkb.org/page/faqs [last accessed 27 April 2014]). The overall aim of pharmacogenomics is to improve health care by increasing the number of responders while decreasing the number of adverse drug reactions. Genetic variation is investigated for drug transporters, drug metabolizing enzymes as well as drug targets. This review will focus on drug metabolizing enzymes.

The most important class of the drug metabolizing enzymes is the cytochrome P450 (CYP450) family, which is involved in phase I metabolism of various endogenous and exogenous compounds, including the majority of clinically prescribed drugs (Warnich et al., 2011). CYP450 enzymes are heme oxygenases, found mainly in the liver. They are involved in converting liposoluble compounds into more hydrosoluble compounds, most commonly through the addition of a hydroxyl or other hydrophilic group, in order for the compound to be effectively eliminated by the kidneys. The major families involved in phase I biotransformation reactions of drugs are CYP1, CYP2 and CYP3.

High nucleotide diversity has been reported for the CYP450 genes. Single nucleotide polymorphisms (SNPs) are defined as a variation in the nucleotide base at a single position in a DNA sequence. When a SNP is present within a gene and occurs in at least 1% of the population, the different representation of that gene is then known as an allele. Over 400 alleles have been assigned to date across the CYP1, CYP2 and CYP3 subfamilies (http://www.cypalleles.ki.se/ [last accessed 28 Oct 2014]). Polymorphisms in genes encoding the drug metabolizing enzymes may affect the activity of the gene products (Ingelman-Sundberg et al., 2007). The consequence of these variations on enzyme activity and substrate metabolism are shown in Figure 1. SNPs and insertions and deletions of DNA sequences (indels), causing frame shift mutations or premature stop codons, can result in either no functional enzyme being formed, unstable enzymes leading to reduced metabolism or even increased enzyme activity. Certain mutations may also lead to amino acid changes that change substrate specificity. Copy number variation, such as multiple copies of a functional allele, can lead to higher enzyme levels and increased metabolism.

It is therefore evident that these polymorphisms may lead to wide variations in catalytic activities (Alessandrini et al., 2013). Individuals can be classified into four phenotypic
groups: ultrarapid metabolizers (UM), generally with more than two active genes, or SNPs that result in induced expression and increased activity; extensive metabolizers (EM), carrying two functional genes; intermediate metabolizers (IM), characterized by genetic variants that result in reduced enzymatic activity; and poor metabolizers (PM), who lack functional enzymes due to defective or deleted genes (Ingelman-Sundberg et al., 2007). Figure 2 shows the general frequency of each phenotype across the population, with the majority of the population falling into the EM group. Population-based dosing results in standard dosing levels for a given drug being set at a level considered to be appropriate for the majority of the population, despite the differences in phenotype present. Population-based dosing therefore results in PM individuals having higher plasma concentrations of drugs than necessary leading to adverse drug reactions and toxicity. In contrast, the absence of a drug response may be seen in UM individuals. The opposite is true for pro-drugs which need to be activated in order to exert their biological activity.

Another important family of drug metabolizing enzymes is the uridine-diphosphate-glucuronosyltransferases (UGT) involved in phase II drug metabolism. The UGT enzymes are involved in conjugating a glucuronic acid group to the functional group of a specific substrate (Guillemette, 2003), resulting in a more hydrophilic and therefore more easily excretable molecule. The UGT enzymes are also known to be polymorphic, with certain variants having pharmacogenetic relevance (Guillemette, 2003). Similar to the CYP450 enzymes, genetic variations can be in the form of SNPs, indels and multiple copies of functional and non-functional alleles, and can occur in the regulatory and coding sequences, introns and the 5' and 3'-untranslated regions. These variations can significantly alter the glucuronidation capacity of the individual carrying a variant, resulting in either ineffective drug levels due to rapid metabolism, or toxicity due to slow metabolism (Court, 2010). The UGT enzymes include more than 26 genes, 18 of which code for functional proteins. They are grouped into two families, UGT1 and UGT2, which are further subdivided into subfamilies UGT1A, UGT2A and UGT2B. Inter-individual variation in the UGT enzymes has also been observed, with UGT1A showing the highest and UGT2B7 the lowest variability (Court, 2010).

**Pharmacogenetics in Africa**

The majority of pharmacogenetic studies to date have been performed on Caucasian and Asian populations. Pharmacogenetic profiles based on these studies are then extrapolated for use in other populations, despite the fact that variant frequencies can differ markedly between different populations (Ikediobi et al., 2011). Dandara et al. (2014) provide a review on the importance of CYP450 pharmacogenetics in African populations and the implications for public health. The authors highlight that a large number of ethnic groups need to be characterized to capture the full
extent of existing genetic diversity. Moreover, by excluding African populations from drug research, unexpected adverse drug reactions may occur when these populations, which have novel variations, are exposed to drugs for the first time.

Polymorphisms exhibit inter-individual as well as inter-population differences. When investigating genetic variation within and between population groups for the CYP2B6 and UGT2B7 genes, \( \pm 90\% \) of the genetic variation was observed among individuals within a population, i.e. due to inter-individual differences. The remaining 10% of genetic variation occurs between different populations or ancestral groups, i.e. inter-population differences (Li et al., 2012). Li et al. (2012) also compared the extent of genetic diversity in the CYP2B6 and UGT2B7 enzymes across a variety of ancestral backgrounds and concluded that genetic variation among individuals within the same population is greater for UGT2B7 than for the highly polymorphic CYP2B6 enzyme. However, genetic diversity among individuals in different population groups is higher for CYP2B6 than UGT2B7. These findings indicate that the impact of ancestry on genetic diversity may be greater for CYP2B6 than for UGT2B7. A clear example of this inter-population difference can be seen in a study investigating the extent of CYP2B6 genetic variability in populations of African and Asian origin. The study found a large number of polymorphisms and a low frequency of the CYP2B6*1 wild-type allele in only 44.3 and 39% of African-Americans and Ghanaians, respectively, compared to the significantly lower number of polymorphisms and higher frequency of the *1 wild-type allele in 68 and 76.1% of Japanese and Koreans, respectively (Klein et al., 2005). It was concluded that Africans exhibit a higher degree of genetic variation than Asian populations with respect to this gene.

Intra-ethnic genetic diversity within Africa has also been reported. The Bantu-speaking populations of Africa make up the majority of sub-Saharan African people. They originated from the Niger-Congo regions, and expanded throughout eastern and southern parts of Africa. The older Khoisan populations based in the South-Western parts of Africa make up the remaining African population. In South Africa, there is also a Cape mixed ancestry population, which originated from the sub-Saharan African Bantu-speakers, with influences from both Europe and Asia (de Wit et al., 2010). When two Bantu-speaking populations from Cameroon and South Africa were compared, they were found to be genetically similar with regard to CYP2A6 and CYP2B6. However, there were some statistically significant differences between the genotype frequencies seen in the two populations with respect to CYP1A2, CYP3A4 and CYP3A5 (Swart et al., 2012). The difference in genotype frequencies demonstrates that African populations show intra-ethnic genetic diversity that needs to be characterized appropriately, and that even linguistically related Bantu-speaking populations may not be genetically homogenous. Allele frequencies between these two Bantu-speaking populations were compared to other populations, including other African, Asian and Caucasian populations. In general, significant differences were seen in allele frequencies between individuals of African origin compared to Asian and Caucasian populations, but statistically significant differences were also reported between the South African population and another Bantu-speaking population from Yoruba in Nigeria with respect to CYP2A6. It is therefore evident that it is important to conduct pharmacogenetic studies across a wide range of ethnic backgrounds on different continents. Unfortunately, there have been limited reports detailing pharmacogenetically relevant genes in Africa, even though African populations show higher levels of genetic diversity compared to other populations (Tishkoff et al., 2009).

Recent reports also suggest that the existence of population specific alleles may be clinically relevant. The CYP2B6*18 allele, characterized by the 983T>C SNP which results in decreased protein expression (Wang et al., 2006), is thought to be an African specific allele. Only individuals of African ancestry and not those of Asian or European ancestry have been reported to carry the allele (Li et al., 2012; Mehlotra et al., 2007). In a study by Radloff et al. (2013), eight novel functionally uncharacterized non-synonymous variants were found in the CYP2B6 gene in a Rwandese population, and it was demonstrated that four of these variants resulted in complete or almost complete loss of function. Three novel SNPs in the CYP2B6 gene were found in a Zimbabwean population (341T>C, 444G>T and 1158A>G), and two novel SNPs in a Ugandan population (856C>T and 1459C>A; Jamshidi et al., 2010). These findings demonstrate the potentially large number of African specific alleles that might exist.

Discordance between genotype and phenotype exists in certain African populations, and has been well documented for the CYP450 enzymes (Alessandrini et al., 2013). For example, African individuals that appear to have wild-type genotypes have been reported to show a PM phenotype. This discordance may be due to a lack of comprehensive genotyping or as a result of population specific alleles that have not yet been identified. Most of the research that has been done to date on pharmacogenetically relevant genes has focused on a subset of well-known and well-described SNPs. As a result, individuals who do not carry at least one of the SNPs from this subset are characterized as having a wild-type genotype. However, the previously mentioned studies demonstrate the need to explore potentially novel variants that may be functionally relevant, particularly in poorly investigated African populations and may help to explain genotype-phenotype correlations more clearly. All possible variants in the complete gene(s) of interest should therefore be sought using next generation sequencing as opposed to genotyping for specific SNPs. In silico analyses will then allow for the determination of possible functional implications of novel variations, and thereby identify variants that may be functionally relevant and require additional in vitro and/or in vivo investigations.

Africa is severely affected by both communicable and non-communicable diseases. Although the continent accounts for only 15.5% of the global population, it carries approximately 25% of the global disease burden (Murray et al., 2013). Adverse drug reactions further complicate the situation due to resulting non-compliance, adding to morbidity, mortality and increasing medical costs across the continent. Pharmacogenetic studies can therefore add value by helping to understand the degree of genetic variability on the
continent, as well as by defining populations that may be at a higher risk for developing either toxicity or a reduced response to certain drugs, with the goal of improved healthcare in Africa (Alessandrini & Pepper, 2014).

When assessing the number of African countries for which frequency data of clinically relevant CYP450 alleles has been reported, it is clear that there is a lack of data, with several regions for which no pharmacogenetic data exists. A discrepancy between the disease burden in a specific region and the pharmacogenetically relevant genes studied in relation to those diseases is also apparent (Alessandrini & Pepper, 2014). Alessandrini & Pepper (2014) define priority pharmacogenetics as the CYP450 genes that should be investigated as a priority given both the disease burden and the extent of CYP450 data currently available in a given region. In southern Africa, HIV/AIDS and malaria rank as the top two diseases contributing to the health burden; however, the majority of CYP450 data that is available for this region relates to genes encoding enzymes that metabolize drugs required for the treatment of cardiovascular/circulatory disease, diabetes and major depressive disorders. The discordance between the health burden and available CYP450 reports clearly points to the fact that the genes associated with the metabolism of drugs used in the treatment of HIV/AIDS should be studied as a top priority in the southern Africa region.

Role of pharmacogenetics in HIV therapy

In Africa, HIV/AIDS constitutes a major disease burden, with sub-Saharan Africa being more affected than any other region in the world (Joint United Nations Programme on HIV/AIDS, 2011). Highly active antiretroviral therapy (HAART) is widely used in the treatment of HIV/AIDS; however treatment is compromised by the development of drug resistance, a high degree of variability in drug response and high rates of adverse drug reactions. While there are many factors which may influence a person’s response to HAART, genetic polymorphisms affecting the activity of key drug metabolizing enzymes is likely to be an important source of inter-patient variability. The CYP450 and UGT enzymes play a major role in phase I and phase II metabolism of drugs, respectively, and thus polymorphisms in the CYP450 and/or UGT genes can result in inter-patient variability in antiretroviral drug efficacy and toxicity. In a study by Kwara et al. (2009b), it was concluded that pharmacogenomic testing used for the estimation of the appropriate dose of the antiretroviral drug efavirenz, should incorporate both oxidative (CYP450) as well as glucuronidation (UGT) pathways.

Pharmacogenetics and HIV therapy

Treatment of HIV

Antiretroviral drugs target various elements of the HIV replication cycle, and include nucleoside/nucleotide reverse transcriptase inhibitors (NRTI), non-nucleoside reverse transcriptase inhibitors (NNRTI), protease inhibitors (PI) and entry inhibitors. Standard antiretroviral treatment usually includes a combination of drugs targeting these different elements to effectively control HIV infection and reduce resistance. Combinations usually include two NRTIs as well as one NNRTI or PI (Department of Health (DOH), Republic of South Africa, 2013). The CYP450 enzymes do not play a central role in the metabolism of NRTIs, for which hepatic glucuronidation is believed to be the predominant metabolic pathway (Mahungu et al., 2009). The CYP450 enzymes are however involved in the metabolism of NNRTIs, including efavirenz and nevirapine, protease inhibitors, the CCR5 coreceptor antagonist maraviroc and the integrase inhibitor elvitegravir (Michaud et al., 2012).

Efavirenz is included in first line antiretroviral regimens for all adults and adolescents in South Africa, and is also included in the treatment of patients co-infected with tuberculosis (TB) [Department of Health (DOH), Republic of South Africa, 2013]. Efavirenz blocks viral replication by binding to reverse transcriptase thereby altering the function of the enzyme and rendering it incapable of converting viral RNA to DNA (Ward et al., 2003). Both the CYP450 as well as UGT enzymes play a role in the metabolism of efavirenz (Figure 3). Efavirenz is mainly metabolized by CYP2B6 into 8-hydroxyefavirenz and to a lesser extent by CYP2A6 into 7-hydroxyefavirenz, with CYP3A4/5 and CYP1A2 playing a minor role in this step (Ward et al., 2003; Whirl-Carrillo et al., 2012). N-Glucuronide-efavirenz is formed when efavirenz undergoes conjugation by UGT2B7 (Bélangier et al., 2009). It has been proposed however that CYP2A6 and UGT2B7 only play a significant role in the efavirenz pathway when CYP2B6 is impaired (di Iulio et al., 2009). Significant inter-individual variability in efavirenz plasma concentrations has been reported, with increased plasma levels leading to adverse side effects including neurotoxicity.

Figure 3. Primary efavirenz metabolism by CYP450 and UGT enzymes. Efavirenz is mainly metabolized by CYP2B6 into 8-hydroxyefavirenz, with CYP2A6, CYP3A4/5 and CYP1A2 playing a minor role in this step. EFV can also be hydroxylated to 7-hydroxyefavirenz by CYP2A6, although efavirenz 8-hydroxylation is the major route of clearance. N-glucuronide-efavirenz is formed when efavirenz undergoes conjugation by UGT2B7. Copyright to PharmGKB. Adapted with permission from PharmGKB and Stanford University (Whirl-Carrillo et al., 2012). Original pathway can be found at http://www.pharmgkb.org/pathway/PA166123135.
In cases where persistent neurotoxicity is seen, nevirapine or a protease inhibitor is substituted. In South Africa, nevirapine is also the preferred treatment for infants of HIV infected women to prevent mother to child transmission (Department of Health (DOH) Repulic of South Africa, 2013). Nevirapine is also still extensively used in other African regions due to its efficacy and affordability (Ciccacci et al., 2010). Decreased metabolism of nevirapine results in increased plasma levels leading to adverse side effects including hepatotoxicity (Haas et al., 2006). Nevirapine is mainly metabolized by CYP2B6 into 3- and 8-hydroxynevirapine and CYP3A4 into 2- and 12-hydroxynevirapine. Nevirapine also undergoes glucuronide conjugation of the hydroxyl metabolites by the UGT enzymes (Riska et al., 1999; Whirl-Carrillo et al., 2012; Figure 4).

Drug–drug interactions may also be a source of variation that can contribute to inter-individual responses as well as adverse drug reactions. This is particularly important in the present context as antiretroviral therapy constitutes a combination of drugs as previously described. Given the high incidence of TB infection in HIV-positive individuals, drug–drug interactions between anti-TB drugs such as rifampicin and antiretroviral drugs such as efavirenz should also be considered, particularly since rifampicin is known to induce CYP2B6 activity.

The review by Dandara et al. (2014) provides a summary of efavirenz metabolism and CYP2B6 pharmacogenetics in African populations, specifically the effect of the CYP2B6 516G>T variant. However, other CYP2B6 polymorphisms may also play a role in altered enzyme activity, as may polymorphisms in CYP2A6 and UGT2B7. Frequencies of all reported polymorphisms in the CYP2A6, CYP2B6 and UGT2B7 genes associated with altered efavirenz and nevirapine plasma concentrations have therefore been reviewed.

**CYP2B6, CYP2A6 and UGT2B7**

**CYP2B6**

The CYP2B6 subfamily of enzymes is known to be highly polymorphic, with 38 allelic variants reported to date (http://www.cypalleles.ki.se/cyp2b6.htm [last accessed 28 October 2014]). Various polymorphisms have been shown to be clinically relevant resulting in increased or decreased metabolism of target drugs. Table 1 provides a summary of the frequency of variants in the CYP2B6 gene in African populations, and the resulting effect on efavirenz and nevirapine metabolism.

**CYP2B6*6 (516G>T and 785A>G)**

Allele frequencies in African populations. The CYP2B6*6 haplotype is characterized by the presence of two non-synonymous variants 516G>T and 785A>G (Thorn et al., 2010). Strong linkage disequilibrium between 516G>T and 785A>G can be seen in many populations, including Africans, Caucasians, Asians and Hispanics (Li et al., 2012; Maimbo et al., 2012; Mehlotra et al., 2007; Swart et al., 2012, 2013). Substantial inter-population differences in the frequency of the 516G>T and 785A>G SNPs have been reported, with higher frequencies seen in African populations. The frequency of the 516G>T SNP differs significantly between African-Americans and European-Americans (38 and 21.9%, respectively, $p=0.005$). Furthermore, 20% of African-Americans have been shown to be homozygous for the 516G>T polymorphism, compared to only 3.4% of European-Americans (Haas et al., 2004). In other studies, the *6 allele was found to be present in 33–35% of African-Americans (Klein et al., 2005; Mehlotra et al., 2007).

The *6 haplotype has been reported at high frequencies in Africa. Mehlotra et al. (2007) reported a frequency of 41.5% in a West African population including Ghana, Guinea, Ivory Coast, Sierra Leone and Senegal. Klein et al. (2005) also reported a high frequency of the *6 haplotype in Ghanaians (46.9%), and Sarfo et al. (2014) reported a frequency of 48% for the 516G>T SNP in another Ghanaian population. In an analysis of the 1000 genomes database, the 516G>T and 785A>G SNPs were found in 35–38% and 22–35% of populations of African ancestry, respectively, including Luhya (Kenya), Yoruba (Nigeria) and those of African ancestry in the southwestern USA (Li et al., 2012). The 516G>T variant was observed at a frequency of 20–32% in a Xhosa population, and at 23–30% in a Cape mixed ancestry population from South Africa (Ikediobi et al., 2011;
Table 1. Frequency of CYP2B6 variants and associated phenotype in populations of African origin.

<table>
<thead>
<tr>
<th>Population</th>
<th>516G&gt;T Frequency</th>
<th>Phenotype</th>
<th>785A&gt;G Frequency</th>
<th>Phenotype</th>
<th>983T&gt;C Frequency</th>
<th>Phenotype</th>
<th>1459C&gt;T Frequency</th>
<th>Phenotype</th>
<th>References</th>
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<td>South Africa</td>
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<tr>
<td>Xhosa</td>
<td>20–32% (6.25%)</td>
<td>No significant association with changes in CD4-cell count</td>
<td>32%</td>
<td>No significant association with changes in CD4-cell count</td>
<td>17%</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Ikediobi et al. (2011), Parathyras et al. (2009)</td>
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<tr>
<td>Cape Mixed Ancestry</td>
<td>23–30% (5.71%)</td>
<td>–</td>
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<td>–</td>
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<tr>
<td>Bantu</td>
<td>36% (12.5–18.6%)</td>
<td>Increased efavirenz plasma concentration</td>
<td>–</td>
<td>–</td>
<td>5.1–12.7% (1.7%)</td>
<td>Increased efavirenz plasma concentration</td>
<td>1.4–3.1%</td>
<td>–</td>
<td>Swart et al. (2013)</td>
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<td>Black</td>
<td>43% (23%)</td>
<td>Increased efavirenz plasma concentration. Adverse side effects. No association with immune response (CD4 count)</td>
<td>–</td>
<td>–</td>
<td>–</td>
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<td>Gounden et al. (2010)</td>
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<td>Other African</td>
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<tr>
<td>West Africa</td>
<td>41.5%*</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>4.7%</td>
<td>–</td>
<td>–</td>
<td>Mehlotra et al. (2007)</td>
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<tr>
<td>Ghana</td>
<td>48.8%</td>
<td>–</td>
<td>47.5%</td>
<td>–</td>
<td>6.6%</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Klein et al. (2005), Sarfo et al. (2014)</td>
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<tr>
<td>Mozambique</td>
<td>41–49% (17–23%)</td>
<td>No association with nevirapine-induced hepatotoxicity</td>
<td>43–45%</td>
<td>4%</td>
<td>Increased efavirenz plasma concentration</td>
<td>–</td>
<td>–</td>
<td>Ciccacci et al. (2010)</td>
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<tr>
<td>Botswana</td>
<td>36.6% (17.8%)*</td>
<td>–</td>
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<td>–</td>
<td>–</td>
<td>Gross et al. (2008)</td>
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<tr>
<td>Malawi</td>
<td>31%</td>
<td>Increased nevirapine exposure</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Brown et al. (2012)</td>
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<tr>
<td>Zimbabwe</td>
<td>48–49%*</td>
<td>Increased efavirenz plasma concentration</td>
<td>–</td>
<td>–</td>
<td>Increased efavirenz plasma concentration</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Jamshidi et al. (2010), Nyakatira et al. (2008)</td>
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<tr>
<td>Uganda</td>
<td>29–35.6%</td>
<td>Increased efavirenz plasma concentration. Increased nevirapine concentration</td>
<td>32–36.4%</td>
<td>9%</td>
<td>Heterozygous patients had a fourfold higher efavirenz plasma concentration</td>
<td>–</td>
<td>–</td>
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<td>Maimbo et al. (2012)</td>
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<td></td>
<td>42%</td>
<td>Significantly associated with efavirenz plasma concentration</td>
<td>42%</td>
<td>–</td>
<td>5.4–10.4%</td>
<td>Increased efavirenz plasma concentration</td>
<td>–</td>
<td>–</td>
<td>Jamshidi et al. (2010), Penzak et al. (2007), Mukonzo et al. (2009)</td>
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<tr>
<td>Location</td>
<td>Percentage</td>
<td>Significant predictor of plasma and intracellular efavirenz concentration</td>
<td>Frequency</td>
<td>Other notes</td>
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<tr>
<td>Tanzania</td>
<td>41.8% (18.6%)</td>
<td>Significant predictor of plasma and intracellular efavirenz concentration</td>
<td>6.9%b</td>
<td>Ngaimisi et al. (2013), Wang et al. (2006)</td>
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<tr>
<td>Ethiopia</td>
<td>31.4% (8.7%)</td>
<td>Significant predictor of plasma and intracellular efavirenz concentration</td>
<td>3%</td>
<td>Ngaimisi et al. (2013)</td>
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<tr>
<td>Cameroon</td>
<td>37%</td>
<td>22.50%</td>
<td>1%</td>
<td>Swart et al. (2012)</td>
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<tr>
<td>1000 Genomes and HapMap</td>
<td>35–38%</td>
<td>22–35%</td>
<td>4–12%</td>
<td>Li et al. (2012)</td>
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<tr>
<td>Yoruba (Nigeria) HapMap</td>
<td>45%</td>
<td></td>
<td>4%</td>
<td>Ikediobi et al. (2011)</td>
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<tr>
<td>Luyha (Kenya) HapMap</td>
<td>31%</td>
<td></td>
<td>7%</td>
<td>Ikediobi et al. (2011)</td>
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<tr>
<td>Maasai (Kenya) HapMap</td>
<td>37%</td>
<td></td>
<td>2%</td>
<td>Ikediobi et al. (2011)</td>
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<tr>
<td>African–American</td>
<td>38% (20%)</td>
<td>Efavirenz plasma concentration three times higher for 516TT genotype. Central nervous system side effects after one week</td>
<td>1%</td>
<td>Haas et al. (2004)</td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td></td>
<td>33–35%a</td>
<td>29.80%</td>
<td>4.4–7.5%</td>
<td>Mehlotra et al. (2007), Klein et al. (2005)</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

Percentages in brackets indicate homozygous individuals. ‘–’ no data present in the published study.

Frequency relates to the *6 haplotype.

Frequency relates to the *16 haplotype.

West African populations include Ghana, Guinea, Ivory Coast, Sierra Leone and Senegal.

1000 genomes African Ancestry populations include Luhya (Kenya), Yoruba (Nigeria) and African ancestry from southwest USA.
A total of 6.25% of individuals in the Xhosa population were found to be homozygous for the 516G>T SNP, as were 5.71% of the Cape mixed ancestry population (Parathyras et al., 2009). The 516G>T variant was reported at a frequency of 45, 31 and 37% in the Yoruba (Nigeria), Luhya (Kenya) and Maasai (Kenya) HapMap populations, respectively (Ikediobi et al., 2011). Within a Bantu-speaking South African population, the CYP2B6*6 allele was found in 36% of individuals, amongst whom 18.6% of HIV/AIDS patients and 12.5% of HIV-negative individuals were found to be homozygous for this allele (Swart et al., 2013). In another HIV-positive South African population, the allele frequency of the 516G>T SNP was 43%, with 23% being homozygous and 41% heterozygous (Gounden et al., 2010). In a population from Mozambique, the 516G>T SNP was found in 41–49% of individuals, with 17–23% of individuals being homozygous (Ciccacci et al., 2010). The 785A>G SNP was found in 43–45% of these individuals, with 20% having a homozygous genotype. The CYP2B6*6 haplotype is also common in Botswana. In a study involving 101 HIV-positive Batswana, the CYP2B6*6 allele was present in 36.6% individuals, and the homozygous haplotype was present in 17.8% of individuals (Gross et al., 2008). A high allele frequency of 49% for the *6 allele was seen in HIV patients in Zimbabwe (Nyakutira et al., 2008). The CYP2B6*6 allele frequency also differed significantly between a Zimbabwean and a Ugandan population, being 48 and 17% (p < 0.004), respectively (Jamshidi et al., 2010). In another Ugandan population, the 516G>T and 785A>G SNPs were found in 35.6 and 36.4% of the population, respectively (Mukonzo et al., 2009). Ngaimisi et al. (2013) found the frequency of the 516G>T SNP to be significantly higher in Tanzanians (41.8%) than Ethiopians (31.4%), with 18.6% compared to only 8.7% homozygous individuals in the Tanzanian and Ethiopian populations, respectively.

Effect of variants on efavirenz and nevirapine plasma concentration. The CYP2B6*6 haplotype is associated with decreased enzyme activity, with the 516G>T SNP being largely responsible. Individuals with either a *1/*6 or *6/*6 haplotype have significantly decreased CYP2B6 protein expression, and a decrease in efavirenz metabolism (Desta et al., 2007). Individuals within an adult AIDS clinical trials group with a 516TT genotype had a three-fold higher efavirenz plasma exposure (defined as the 24-h area under the curve) than individuals with the 516GG wild-type genotype (Haas et al., 2004). Efavirenz plasma clearance in this same group was 23 and 54% lower in individuals with the 516GT and 516TT genotype respectively when compared to the wild-type 516GG genotype. The 516G>T genotype was also associated with central nervous system side effects after 1 week of efavirenz medication, although central nervous system side effects were not significant in later weeks. Elens et al. (2010) found a clear linear relationship between efavirenz plasma concentrations and the number of 516G>T mutated alleles. The 516TT genotype was significantly associated with increased efavirenz plasma concentrations in a Ghananian population, with the 516G>T SNP accounting for as much as 45% of the variability using a linear regression analysis (Kwara et al., 2009b). Patients with the homozygous 516TT genotype in an HIV-positive South African population had significantly higher efavirenz concentrations than those with the GG or GT genotype, with 516TT patients experiencing more severe adverse side effects (Gounden et al., 2010). The CYP2B6*6 allele was found to be a significant predictor of both plasma and intracellular efavirenz concentrations in both Tanzanian and Ethiopian populations (Ngaimisi et al., 2013). An increase in efavirenz concentration is associated with an increase in the number of loss of function alleles, namely, CYP2B6 *6, *18, *20 and *27, in both Zimbabwean and Ugandan populations, with the number of either the 516G>T or 983T>C alleles having the greatest impact on increasing efavirenz concentrations (Jamshidi et al., 2010). Mukonzo et al. (2009) reported a 21% lower oral clearance of efavirenz in another Ugandan population for individuals homozygous for the *6 allele.

In a Bantu-speaking South African HIV-positive population, 35% had plasma efavirenz levels above the therapeutic range of 4 µg/ml, which is likely to be associated with toxicity (Swart et al., 2013). Ninety-two percent of these patients with high plasma efavirenz levels were carriers of either the CYP2B6*6/*6 or CYP2B6*1/*6 genotypes. The homozygous CYP2B6*6 genotype was associated with 46% sensitivity and 97% specificity in predicting plasma efavirenz levels above 4 µg/ml. The homozygous CYP2B6*6 genotype also had an 88% positive predictive value for efavirenz plasma levels above 4 µg/ml. Therefore, given that the study found between 12.5 and 18.6% of the population to be homozygous for the *6 allele, it can be estimated that between 10 and 16% of Bantu-speaking South African HIV/AIDS patients will have plasma efavirenz levels of above 4 µg/ml.

The 516G>T SNP has also been shown to affect nevirapine plasma concentrations. Penzak et al. (2007) reported a significant correlation between the 516G>T SNP and nevirapine concentration in an HIV-positive Ugandan population. Brown et al. (2012) reported the 516G>T SNP to be significantly correlated with increased nevirapine exposure in a Malawian population. However, no significant association was found between the 516G>T SNP and nevirapine-induced hepatotoxicity in a Mozambiquan population (Ciccacci et al., 2010). More studies involving nevirapine plasma concentrations and CYP2B6 polymorphisms are thus needed to clarify these differences in observations.

Effect of variants on immune response. When assessing the impact of polymorphisms on immune function using CD4 cell recovery as an indicator of response to HAART, no significant association was found between the 516G>T and 785A>G SNPs and CD4 counts in South African Xhosa and Mixed-Ancestry HIV-positive patients (Parathyras et al., 2009). Similarly, no significant correlation was found between genotype and follow up viral loads following efavirenz treatment in a South African HIV-positive population (Gounden et al., 2010). These results suggest that although these variants increase plasma concentrations of efavirenz or nevirapine leading to toxicity as previously described, immune function is unlikely to be affected.
Allele frequencies in African populations. The 983T>C SNP is the defining SNP of the *18 allele, but also occurs in the *16 haplotype with the 785A>G SNP (Thorn et al., 2010). In a study investigating populations in the 1000 genomes database, the CYP2B6 983T>C SNP was reported to be highly African specific, with the CYP2B6*18 allele having been found only in populations of African ancestry (4–12% frequency) and in a Puerto Rican population. No other population including European Caucasian, Asian or Mexican was found to carry the CYP2B6*18 allele (Li et al., 2012).

Inter-ethnic variation between African populations is evident for this SNP. In an analysis of pharmacogenetic traits in two distinct HIV-positive South African populations, the CYP2B6*18 variant allele was shown to occur at a frequency of 17% in a Xhosa population, and 9% in a Cape mixed ancestry population, but differed considerably when compared to the reported frequencies of 4, 7 and 2% in the Yoruba (Nigeria), Luhy (Kenya) and Maasai (Kenya) HapMap populations, respectively (Ikediobi et al., 2011). Within another Bantu-speaking South African population, the CYP2B6*18 allele was found in 5.1% of HIV-negative participants and in 12.7% of HIV/AIDS patients, with 1.7% of the HIV/AIDS patients being homozygous for this allele (Swart et al., 2013). The 983T>C SNP was reported in 4.7% in west Africa (including Ghana, Guinea, Ivory Coast, Sierra Leone and Senegal), in 7.5% of African-Americans and in 1.1% of Hispanic-Americans, but was not found in Papua New Guinea, Caucasian-American or Asian-American populations (Mehlotra et al., 2007). Sarfo et al. (2014) also reported a frequency of 4% in another Ghanaian population. The 983T>C SNP was reported in 9% of an HIV-positive Zimbabwean population (Maimbo et al., 2012), and in 5–7% of individuals in Mozambique, with no individuals being of a homozygous genotype (Ciccacci et al., 2010).

The *16 haplotype (983T>C and 785A>G) is seen less frequently. Only 1% of Nigerians were found to carry the CYP2B6*16 allele, whereas this haplotype was neither seen in any other population in the 1000 genomes database, including other African, Caucasian or Asian populations (Li et al., 2012), nor it seen in a Zimbabwean population (Maimbo et al., 2012). Neither was there any indication of the *16 haplotype in West Africa and African-American populations (Mehlotra et al., 2007). However, it was reported in 6.9% of a Tanzanian population, and 4.1% in a Turkish population, where strong linkage between 983T>C and 785A>G was shown (Wang et al., 2006).

Effect of variants on efavirenz and nevirapine plasma concentration. The CYP2B6 983T>C SNP has been associated with increased efavirenz and nevirapine plasma concentrations and adverse side effects. When investigating nevirapine-induced hepatotoxicity, the 983T>C SNP was significantly correlated with higher alanine aminotransaminase (ALT) values in a Mozambique population (Ciccacci et al., 2010). Increased levels of this transaminase are associated with liver damage and hepatotoxicity. In a similar investigation using efavirenz, subjects carrying the *16 allele had a three-fold higher efavirenz concentration compared to other genotypes (Wang et al., 2006). Similarly, Sarfo et al. (2014) and Elens et al. (2010) found significantly elevated efavirenz concentrations in individuals carrying the 983T>C SNP. An in vitro analysis has shown the 983T>C variant results in decreased protein expression, with a protein level of only 15–30% of that of the wild-type CYP2B6*1 allele (Wang et al., 2006). Lower protein levels result in a poor metabolizer phenotype and therefore higher drug plasma concentrations leading to adverse drug reactions.

CYP2B6 785A>G

The CYP2B6 785A>G variant is found alone as the CYP2B6*4 allele, but is also found as part of other haplotypes including CYP2B6*6 (with 516G>T) and CYP2B6*16 (with 983T>C) as described previously. In the absence of other variants, the 785A>G SNP has been reported to be associated with higher levels of protein expression and increased enzyme activity. In an in vitro analysis, the 785A>G SNP alone resulted in a two-fold increase in protein expression compared to the wild-type allele (Wang et al., 2006). In combination with the 983T>C SNP, the 785A>G SNP slightly, but not significantly, resulted in increased expression compared to constructs carrying only the 983T>C SNP. The increase in expression may indicate that the 785A>G SNP partially overrides the inhibitory effect of the 983T>C SNP. The same may be true for the CYP2B6*6 allele, where the 785A>G SNP may partially override the inhibitory effect of the 516G>T SNP.

Other CYP2B6 haplotypes

Differences have been reported in allele frequencies between different ethnic populations in other CYP2B6 variants. Allele frequencies differed significantly between African-Americans and European-Americans for 1459C>T, which characterizes the *5 allele (1 and 11.8%, respectively, p < 0.0001; Haas et al., 2004), indicating that the frequency of this variant may be lower in African populations. In this study, there was no significant association between the 1459C>T SNP and efavirenz related central nervous system side effects. Desta et al. (2007) reported that a decrease in the CYP2B6 protein in samples with *1/*5 and *5/*6 genotypes, but likewise this did not result in a significant reduction in efavirenz metabolism. Within a Bantu-speaking South African population, the 1459C>T SNP was found in 3.1% of HIV-negative participants, and in 1.4% of HIV/AIDS patients (Swart et al., 2013). In this same population, another reduced activity polymorphism 136A>G, which characterizes the *11 allele, was found in 11.9% of HIV-negative participants and in 15.9% of HIV/AIDS patients. A total of 1.9% and 4.7% of HIV-negative and HIV/AIDS patients, respectively, were homozygous for this SNP. The *11 allele was found in 13.6% of a Ugandan population, with homozygous individuals having a 20% lower apparent oral clearance of efavirenz (Mukonzo et al., 2009). The haplotypes *17, *19, *20 and *21 may also be considered to be African specific, as they have only been reported in populations of African origin; these haplotypes were absent in both the Asian and Caucasian populations studied (Klein et al., 2005). Other SNPs that have been associated
with nevirapine-induced hepatotoxicity include 646-17C>T, 646-159G>A and 6986A>G (Ciccacci et al., 2010).

Based on the data reviewed above, there is therefore strong evidence that the 516G>C and 983T>C SNPs that characterize the *6 and *18 haplotypes have the greatest effect on efavirenz and nevirapine plasma concentrations, and an argument can be made for decreasing dosage in individuals carrying these variants. Gatanga et al. (2007) showed that reducing the dose of efavirenz from 600 mg to either 400 mg or 200 mg in individuals carrying the *6/*6 genotype, successfully decreased efavirenz plasma concentrations and central nervous system side effects, yet successfully suppressed viral load. Such findings present an argument for the utility of pharmacogenetic testing of individuals to personalize dosage. However, due to the limited number of genotype–phenotype reports in African populations, further studies are needed in these populations to determine the true correlation between the genotype of an individual and the resulting change in drug plasma levels. It will also be necessary to determine the true effect of other CYP2B6 variants on efavirenz and nevirapine plasma concentrations.

**CYP2A6**

**Allele frequencies in African populations.** CYP2A6 exhibits a high number of polymorphisms, with 45 alleles having been reported to date (http://www.cypalleles.ki.se/cyp2a6.htm [last accessed 28 October 2014]). These result in a wide range of catalytic activities, with certain variants having been shown to be of pharmacogenetic relevance. Although on the African continent CYP2A6 variants have been studied to a lesser extent than variants for CYP2B6, certain alleles have nonetheless been shown to exhibit inter-ethnic variability. Schoedel et al. (2004) reported significant ethnic variation in allele frequencies between the five ethnic groups studied (African North American, Canadian Native Indian, Caucasian, Chinese and Japanese). The *1B, *4, *7, *8 and *9 allele frequencies were significantly different between ethnic groups, with African-North Americans having lower allele frequencies than European groups (Schoedel et al., 2004). The *1B allele was observed in 11.9% of Ghanaians, while the *4 whole gene deletion and the *9 allele were present in 1.9% and 5.7% of Ghanaians, respectively (Gyamfi et al., 2005). The *1B allele was present in 11.9% of Ghanaian patients (1.9%), none of the subjects studied in a Zimbabwean population possessed this allele (Kwara et al., 2009a). However, the *17 allele has to date not been reported in either Caucasian or Asian populations. Within a Bantu-speaking South African population, the 1093G>A reduced activity polymorphism (present in the *17 haplotype) was found in 15.6% of HIV-negative participants, and in 13.6% of HIV/AIDS patients, with 1.7% of the HIV/AIDS patients being homozygous for this SNP (Swart et al., 2013). While the *4 allele was present in the Ghanaian population (1.9%), the *17 allele has also been reported at a low frequency of 1.2% in Caucasian populations; however, it is common in Asian populations with a frequency of 24% in a Japanese population (Schoedel et al., 2004).

**Effect of variants on efavirenz and nevirapine plasma concentration.** Certain CYP2A6 variants have been reported to affect both efavirenz as well as nevirapine plasma concentrations. Table 2 summarizes the frequency and effect on phenotype of the CYP2A6 *9 and *17 haplotypes in African populations. Di Iulio et al. (2009) reported an association between decreased and loss of function CYP2A6 alleles and efavirenz metabolite concentrations. In a Ghanaian population, the *9 and/or *17 alleles were significantly associated with altered efavirenz plasma concentrations, and accounted for 9% of the inter-individual variability as determined by linear regression analysis (Kwara et al., 2009b). Individuals with the *9 and/or *17 alleles had a 32% higher efavirenz concentration based on the regression model. Another study involving Ghanaian patients also concluded that the −48 G>T SNP, characterizing the *9 allele, was significantly associated with elevated efavirenz concentrations (p < 0.0001; Sarfo et al., 2014). However, the association of these SNPs with efavirenz concentration has not been reported in other studies. Of the SNPs studied by Elens et al. (2010), no significant association was seen either the *9b or *17 alleles and efavirenz concentrations. Similarly, no association was found between CYP2A6 allele frequencies.

<table>
<thead>
<tr>
<th>Population</th>
<th>−48G&gt;T (*9)</th>
<th>1093G&gt;A (*17)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Frequency</td>
<td>Phenotype</td>
</tr>
<tr>
<td>African–American</td>
<td>7.1%</td>
<td>−</td>
</tr>
<tr>
<td>Ghanaian</td>
<td>5.7%</td>
<td>Increased efavirenz plasma concentration</td>
</tr>
<tr>
<td>Bantu (South African)</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td>Zimbabwean</td>
<td>7.0%</td>
<td>No association with efavirenz plasma concentration</td>
</tr>
</tbody>
</table>

The −48G>T SNP characterizes the *9 haplotype and the 1093G>A SNP characterizes the *17 haplotype. “−” indicates no data present in the published study.
and efavirenz plasma concentrations in a Zimbabwean population (Maimbo et al., 2012). Therefore, while the *9 and *17 haplotypes may be variants for which a possible phenotypic effect exists, similarly to the CYP2B6 variants, further investigations are needed to clarify these relationships, particularly in African populations.

**UGT2B7**

**Allele frequencies in African populations.** The UGT2B7 enzyme is known to be polymorphic, with three non-synonymous SNPs, as well as several synonymous, intronic and promoter SNPs (Guillemette, 2003). Four alleles (*1–*4) with sub-variants, have to date been described for UGT2B7 (http://www.pharmacogenomics.pha.ulaval.ca/files/content/sites/pharmacogenomics/files/Nomenclature/UGT2B/UGT2B7.htm [last accessed 28 October 2014]). When investigating worldwide variation in UGT2B7, certain significant differences were observed across different ethnic backgrounds (Li et al., 2012). The *2a allele was found to be significantly higher in European-Americans (49%), compared to African-Americans (32%), West Africans (19%) and individuals from Papua New Guinea (28%). The *1m allele was found to be significantly higher in Hispanic-Americans (37%) than any other North American population, but was comparable to the West African population (34%). The *3 allele was found at low frequencies in Hispanic-American and Asian-American populations, but not in any of the other populations examined. Allele *1a was highly prevalent in all populations (32–45%), while allele *1b had a low prevalence across all populations (1–9%). The *4 allele was not seen in any of the populations studied; however, it has previously been reported in a Japanese population (Saeki et al., 2004). Ngaimisi et al. (2013) found the frequency of the UGT2B7–327G>A SNP, which occurs in certain *2 alleles, to be significantly different in a Tanzanian population (29.3%) compared to an Ethiopian population (48.3%). The *1c and *2 alleles are present in 15 and 23% of Ghanaians, respectively (Sarfo et al., 2014).

**Effect of enzyme activity on efavirenz and nevirapine plasma concentration.** UGT2B7 haplotypes that have been reported to have an effect on efavirenz concentration include the *1 and *2 haplotypes. In a Ghanaian population, the *1a and *2 alleles were significantly associated with altered efavirenz plasma concentrations, with the *1a allele accounting for 10% of the inter-individual variability seen (Kwara et al., 2009b). Individuals with the *1a allele had on average a 41% higher efavirenz plasma concentration. However, other studies have not found such an association. In another study involving Ghanaian patients, no association was found between the *1c and *2 alleles and efavirenz concentration. Elens et al. (2010) as well as Bélanger et al. (2009) also concluded that the *1c and *2 alleles were not associated with variations in efavirenz plasma concentrations. Likewise, no association was found between UGT2B7 allele frequencies and efavirenz plasma concentrations in a Zimbabwean population (Maimbo et al., 2012). Ngaimisi et al. (2013) also found no association between the UGT2B7–327G>A SNP and efavirenz plasma concentrations. Opposing observations may be due to limited sample sizes and/or differing methodologies used and further highlight the need for additional analysis. In addition, since UGT2B7 only plays a minor role in the metabolism of efavirenz, evaluating UGT2B7 variants in conjunction with variants in CYP2B6 and CYP2A6 becomes important. The effect of UGT2B7 variants on efavirenz and nevirapine plasma concentrations in individuals with impaired CYP2B6 and/or CYP2A6 enzymes should likewise be further investigated.

Therefore, we know that the UGT2B7 enzyme plays a role in efavirenz metabolism, yet it is unclear as to what the clinical relevance of specific variants on enzyme activity might be. The *1a and *2 haplotypes may represent functionally relevant alleles. It is evident that inter-ethnic differences exist in the frequency of these variants; however, the frequency of these variants in African populations is largely unknown. The number of reports detailing UGT2B7 polymorphisms in Africa and the phenotypic effect of these variants are scarce, which further highlights the need for in-depth investigations in these populations.

**Conclusion**

Genetic polymorphisms in drug metabolizing enzymes, including CYP2B6, CYP2A6 and UGT2B7, have been shown to have potential clinical relevance with frequencies of variants exhibiting population as well as intra-population differences. In certain instances, these polymorphisms have been shown to affect plasma concentrations of the antiretroviral drugs efavirenz and nevirapine, leading to neurotoxicity and hepatotoxicity, respectively. Given the high prevalence of HIV/AIDS across Africa and in particular in sub-Saharan Africa, and the wide-spread use of both efavirenz and nevirapine in the treatment of this disease, a strong argument can be made for pharmacogenetic testing in affected populations to personalize dosage and thereby achieve optimal plasma concentrations. Pharmacogenetic research is particularly relevant in the context of the treatment of HIV on the African continent given that (a) Africans exhibit a greater degree of genetic diversity than other populations, (b) frequencies of clinically relevant polymorphisms differ between African populations and other ethnic groups and (c) the existence of African specific alleles.

There is a significant amount of evidence showing that the CYP2B6*6 and the *18 alleles lead to reduced rates of efavirenz and nevirapine metabolism. As a result, individuals carrying these alleles are more prone to adverse side effects. The prevalence of CYP2B6*6 is higher in African populations than in Caucasians, while *18 is considered to be an African specific allele. Some studies have shown that CYP2A6*9 and *17 are associated with higher efavirenz concentrations while others have not been able to confirm these results. Given the importance of CYP2A6 in efavirenz metabolism, particularly in cases where the CYP2B6 enzyme is not fully functional, and the lack of CYP2A6 investigations in Africa, there is an urgent need for additional studies on the African continent. Ethnic differences also exist in polymorphism frequencies for the UGT2B7 enzymes; however, studies in African populations are very limited. It remains unclear whether an association between efavirenz concentrations and UGT2B7 polymorphisms exist, with the *1a and *2 alleles...
demonstrating a possible association. A more extensive analysis is thus required. The relationship between concomitant variants in the CYP2B6, CYP2A6 and the UGT2B7 enzymes in the same individual and the subsequent preferred mode of metabolism may also be particularly interesting, given the critical role of accessory pathways in efavirenz metabolism in individuals with CYP2B6 reduced function alleles. A well-controlled and well-designed genotype–phenotype study of these three enzymes in African populations is therefore extremely important.

It is essential that future research must lead to a clear understanding of the correlation between genotype and phenotype in African populations, allowing appropriate conclusions to be made regarding drug dosage levels based on the presence of genetic variants. Ideally, where there is a known pharmacogenetic effect for a certain treatment, patients should be tested for the relevant polymorphisms before starting treatment, and the appropriate dose adjustment made from the outset. Pharmacogenetic research should therefore be used to inform decisions made by health authorities, with the overall aim of reducing adverse drug reactions, increasing the number of responders and improving healthcare on the African continent.

Declaration of interest

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References


