

# CHARACTERISATION OF ASTROVIRUSES FROM SELECTED CLINICAL AND ENVIRONMENTAL SETTINGS

by

SANDRAMA NADAN

submitted in partial fulfilment of the requirements for the degree

MAGISTER SCIENTIAE

MSc (Medical Virology)

in the
Faculty of Health Sciences
University of Pretoria
Pretoria, South Africa

April 2002



### **ACKNOWLEDGEMENTS**

I wish to express my sincere appreciation to the following:

Prof WOK Grabow, Head of the Department of Medical Virology, for the opportunity to do this research in his department and for his Interest and guidance throughout this investigation;

My supervisor, Prof MB Taylor, for teaching me what science really is, Thank you, and co-supervisor, Prof DO Matson, for his keen interest and advice;

Profs JE Walter and DK Mitchell, Center for Pediatric Research, Eastern Virginia Medical School, Norfolk, Virginia, USA, for their interest, provision of valuable control material and phylogenetic analyses on the South African isolates;

Dr DW Cubitt, Great Ormond Street Hospital for Sick Children, London, for supplying reference isolates;

Personnel from the Diagnostic Services of the Department of Medical Virology, University of Pretoria for assisting with the collection of clinical specimens and for performing selected assays;

Prof EJ van Rensburg and the research students and staff, Department of Human Genetics, University of Pretoria, for their invaluable advice; and

The Poliomyelitis Research Foundation and National Research Foundation for post-graduate bursaries;

This study was supported, in part, by grants from the Water Research Commission and the National Research Foundation of South Africa.



This dissertation is dedicated to my parents and family for their continued encouragement and support.



# CHARACTERISATION OF ASTROVIRUSES FROM SELECTED CLINICAL AND ENVIRONMENTAL SETTINGS

by

#### SANDRAMA NADAN

SUPERVISOR: PROF MB TAYLOR

CO-SUPERVISOR: PROF DO MATSON

DEPARTMENT : MEDICAL VIROLOGY

DEGREE: MAGISTER SCIENTIAE (MEDICAL VIROLOGY)

#### SUMMARY

Astroviruses (AstVs), classified within the family *Astroviridae*, include both animal and human pathogens. Human AstVs (HAstVs) include eight serotypes and after rotaviruses, are the most common pathogen in childhood viral diarrhoea. With the impending licensure of a rotavirus vaccine, the significance of HAstVs in diarrhoeal disease needs to be reassessed. HAstV infection has been reported in all age groups, with the young, elderly and immunocompromised at greatest risk. Transmission occurs via the faecal-oral route and the occurrence of AstVs in water sources has been documented. The significance to humans of AstVs in environmental sources has not been quantified.

The aim of this study was to optimise and apply molecular techniques for the detection and characterisation of AstVs in human stool specimens and water samples. The HAstV serotypes primarily responsible for gastroenteritis in this region of South Africa (SA) could then be established. Animal stools would also be screened for AstVs to obtain a SA reference strain for further characterisation and comparative studies. Nucleotide sequences



of the clinical and environmental isolates could be compared with each other and to AstVs found outside SA. These data would also provide information on the role of water as a source of human infection and of the source of faecal contamination of surface waters.

Human stool specimens, water concentrates and cell culture derivatives of these, were screened for HAstVs by enzyme immunoassay and type-common reverse transcriptase polymerase chain reactions (RT-PCR). AstV isolates were characterised by nucleotide sequence analysis of RT-PCR amplicons, generated in the 5' and 3' ends of the genome, using type-common and type-specific primer pairs.

Of a total of 35 clinical isolates, 22 AstV strains were characterised and compared to 25 environmental strains obtained from 15 surface water and wastewater samples. Cell culture amplification of selected specimens enabled the amplification of isolates present in low titres as well as the isolation of multiple AstVs serotypes from single sewage samples. All AstVs from the stool specimens and water samples were identified as HAstVs. All eight HAstV serotypes were represented in the combined study samples. Phylogenetic analyses of the nucleotide sequences of each of the HAstV isolates and comparisons with isolates from the rest of the world showed that some SA strains formed unique genetic clusters, as has been observed in other studies at other AstVs in some clinical and environmental samples were sites. identical. The existence of HAstVs in water samples highlights the potential health risk posed by these waters used for recreational and domestic purposes. This study also presents new baseline data on the molecular epidemiology of HAstVs in SA.

KEYWORDS: Human astroviruses, sewage, multitypes, reverse transcriptase-polymerase chain reaction, molecular characterisation, sequencing, cell culture amplification



# KARAKTERISERING VAN ASTROVIRUSSE VAN GESELEKTEERDE KLINIESE EN OMGEWINGS BRONNE

deur

#### SANDRAMA NADAN

PROMOTOR: PROF MB TAYLOR

MEDE-PROMOTOR: PROF DO MATSON

DEPARTEMENT : GENEESKUNDIGE VIROLOGIE

GRAAD: MAGISTER SCIENTIAE (GENEESKUNDIGE VIROLOGIE)

#### **OPSOMMING**

Astrovirusse (AstVs), geklassifiseer in die familie Astroviridae, bevat beide dierlike en menslike patogene. Daar is agt serotipes van mens astrovirusse (HAstVs) en is na rotavirusse die mees algemene patogeen van virale diarree in kinders. Met die naderende lisensiëring van 'n rotavirus entstof, moet die belangrikheid van HAstVs met betrekking tot diarree herevalueer word. Alhoewel HAstV infeksie in alle ouderdomsgroepe gerapporteer word, is kinders, bejaardes en immuungekompromitteerde individue meer vatbaar. Oordrag is deur die fekaal-orale roete en die voorkoms van AstVs in gekomtamineerde waterbronne is al beskryf. Die kliniese betekenis van omgewings AstVs tot mense is onbekend.

Die doel van hierdie studie was om molekulêre tegnieke te optimiseer en toe te pas vir die opsporing en karakterisering van AstVs in mens-stoelgang- en water-monsters. Die HAstV-serotipes wat primêr verantwoordelik was vir gastroenteritis kan dan vasgestel word. Stoelgang monsters van diere sou ook ondersoek word vir AstVs om 'n SA bron van dierlike AstV te verkry vir verder karaktiseering en vergelykende studies. Nukleotied basispaar volgorde-bepaling van kliniese en omgewings isolate kan dan



vergelyk word om vas te stel wat die verhouding van SA AstVs tot stamme in die res van die wêreld is. Hierdie data sal ook inligting verskaf tot die moontlik rol van water in menslike infeksie en die bron van fekale kontaminasie van oppervlak water.

Mens stoelgang monsters, water konsentrate en hul selkultuur ekstrakte is ondersoek vir HAstVs deur middel van ensiem immuunbepaling en tipe-algemeen tru-transkripsie polimerase ketting reaksie (TT-PKR). AstV isolate is deur nukleotied basispaar volgorde-bepaling, van die 3' en 5'-kant van die genoom, deur tipe-algemene en tipe-spesifieke voorvoerdes gekarakteriseer.

Uit 'n totaal van 35 kliniese isolate, kon 22 AstV stamme gekarakteriseer en vergelyk word met 25 omgewings stamme geïsoleer uit 15 oppervlak en rioolwater monsters. Deur middel van selkultuur vermeerdering kon isolate teenwoodig in lae titers in geselekteerde monsters geamplifiseer word. Selkultuur amplifikasie het ook die isolasie van veelsoortige AstVs van enkele rioolmonsters bevorder. Alle AstVs vanuit stoelgang en water monsters was geïdentifiseer as HAstVs. Ten minste een van elk van die agt HAstV serotipes is geïdentifiseer uit die totale aantal studie monsters. Filogenetiese analise van die nukleotied basispaar volgorde-bepaling van elk van die HAstV isolate, asook vergelyking met isolate uit die res van die wêreld, het getoon dat geselekteerde SA stamme hul eie unieke groepe vorm, soortgelyk aan patrone gerapporteer in ander studies. Sekere AstV-stamme vanaf kliniese en omgewingsbronne was identies. Die identifisering van HAstVs in water monsters beklemtoon die potensiële gesondheidsrisiko vir hierdie waterbronne vir individue wat huishoudelike ontspanningsdoeleindes gebruik. Hierdie studie verteenwoordig nuwe inligting oor die molekulêre epidemiologie van HAstVs in SA.

SLEUTELWOORDE: Mens astrovirusse, riool, veelsoortige tipes, tru-transkripsie polimerase kettingreaksie, molekulêre karakterisering, nukleotied basispaar volgorde-bepaling, selkultuur vermeerdering



# **PUBLICATIONS AND PRESENTATIONS**

#### **Publications**

Nadan S, Walter JE, Grabow WOK, Mitchell DK, Taylor MB.

Molecular characterization of astroviruses: comparison between clinical and environmental isolates from South Africa. Applied and Environmental Microbiology (submitted)

#### Presentations

Nadan S, Grabow WOK, Taylor MB. The molecular detection and characterisation of astroviruses from human stool specimens and sewage [Poster/Presentation]. Faculty Day, Faculty of Health Sciences, University of Pretoria 21-22 August 2001: Pretoria.

Taylor MB, **Nadan S**, Grabow WOK, Walter JE. Molecular epidemiology of human astroviruses from the Tshwane area (Pretoria), Gauteng [Presentation]. Joint Congress of the Infectious Diseases & Sexually Transmitted Diseases Societies of Southern Africa. 2-7 December 2001: Spier Estate, Stellenbosch, South Africa.

**Nadan S**, JE Walter, Grabow WOK, Taylor MB. The molecular detection and characterisation of astroviruses from human stool specimens and sewage [Presentation]. "Microbial Diversity" 12th Biennial Congress of the South African Society for Microbiology, Faculty of Health Sciences, University of the Free State. 2-5 April 2002: Bloemfontein.



WB van Zyl, **S Nadan**, JC Vivier, JME Venter, K Riley, EKM Tlale, LR Seautlueng, WOK Grabow, MB Taylor. The prevalence of enteric viruses in patients with gastroenteritis in the Pretoria and Kalafong Academic Hospitals, South Africa [Poster]. "Microbial Diversity" 12th Biennial Congress of the South African Society for Microbiology, Faculty of Health Sciences, University of the Free State. 2-5 April 2002: Bloemfontein.



# TABLE OF CONTENTS

Acknowledgements	Page
Dedication	ii
Summary	10
Opsomming	V
Publications and Presentations	vii
Table of Contents	ix
List of Tables	xvi
List of Figures	xviii
Abbreviations	xx
CHAPTER 1: GENERAL INTRODUCTION	N 1
CHAPTER 2: LITERATURE REVIEW	4
2.1 History	4
2.2 Classification and morphology	6
2.2.1 Morphology	6
2.2.2 Taxonomy and classification	8
2.3 Biochemical and biophysical character	stics 10
2.3.1 Buoyant density and sedimentation va	lue 10
2.3.2 Stability	11
2.3.3 Nucleic acid composition	12
2.3.4 Polypeptide composition	12
2.4 Antigenic properties and serotypes	14
2.4.1 Human astrovirus (HAstV) serotypes	14
2.4.2 Distribution of HAstV serotypes	15
2.4.3 Animal astrovirus (AstV) serotypes	16



			Page
2.5	Molecular b	piology	17
2.5.1	Genomic o	rganisation and expression	17
	2.5.1.1	Open reading frame 1a	19
	2.5.1.2	Open reading frame 1b	20
	2.5.1.3	Open reading frame 2	20
2.5.2	Genotypic	properties	21
2.6	Propagation	n of astrostviruses	23
2.6.1	Propagation	n of human astroviruses	23
2.6.2	Propagation	n of animal astroviruses	25
2.7	Viral detec	tion and characterisation	26
2.7.1	Virus/Antig	en detection and characterisation assays	26
	2.7.1.1	Electron microscopy	26
	2.7.1.2	Immune electron microscopy	27
	2.7.1.3	Immunofluorescence and immunoperoxid	ase
		assays	28
	2.7.1.4	Enzyme immunoassays	29
2.7.2	Molecular a	assays for the detection and characterisation	n
	of HAstVs		30
	2.7.2.1	Hybridisation assays	30
	2.7.2.2	Reverse transcriptase-polymerase chain	
		reaction (RT-PCR)	32
2.8	Clinical asp	pects	35
2.8.1	Clinical ma	nifestations	35
2.8.2	Pathogenes	sis	37
2.8.3	Diagnostic	assays	38
	2.8.3.1	Viral detection	38
	2.8.3.2	Serology	38
	2.8.3.3	Viral isolation	38
284	Treatment		39



			Page
2.9	Epidemio	ology	39
2.9.1	Routes o	of transmission	40
	2.9.1.1	Role of food and water in the transmis	ssion
		of HAstVs	41
2.9.2	Preventi	on and control	42
2.10	Economi	c impact	43
2.11	Occurren	nce of AstVs in South Africa	43
AIMS	OF THIS	INVESTIGATION	44
CHAP	TER 3:	MATERIALS AND METHODS	46
3.1	Study sa	ample	46
3.1.1	Human s	stool specimens	46
3.1.2	Animal s	tool specimens	47
3.1.3	Water ar	nd sewage samples	47
3.1.4	AstV iso	lates from river, dam and sewage water sa	amples 47
3.2	Viral rec	overy and concentration	48
3.2.1	Glass we	ool adsorption-elution procedure	48
3.2.2	Polyethy	lene glycol/sodium chloride concentration	
	techniqu	e	48
3.2.3	Ultrafiltr	ation	49
3.3	Cell cult	ure procedures	49
3.3.1	Cell cult	ures	49
	3.3.1.1	Human colonic carcinoma cell line	49
	3.3.1.2	Human hepatoma cell line	50
	3.3.1.3	Madin-Darby Bovine Kidney cell line	50
3.3.2	Media a	nd reagents	50
	3.3.2.1	Serum	50
	3.3.2.2	Growth media	51



			Page
	3.3.2.3	Maintenance media	51
	3.3.2.4	Cryopreservation media	51
	3.3.2.5	Starvation media	51
	3.3.2.6	Trypsin-EDTA	51
3.3.3	Subcultu	iring of cell cultures	52
3.3.4	Maintena	ance of cell cultures	52
3.3.5	Cryopres	servation of cells	52
3,3.6	Revival o	of cryopreserved cells	53
3.3.7	Infection	of cells cultures	53
	3.3.7.1	Sample preparation	53
	3.3.7.2	Infection procedure	54
3.3.8	Harvesti	ng of infected cell cultures	54
3.3.9	HAstV re	eference strains	54
3.3.10	O Assessi	ment of cell cultures for the isolation of AstVs	55
3,4	Viral det	ection	56
3.4.1	Electron	microscopy	56
3.5	Antigen	detection	56
3.5.1	Enzyme	immunoassay	56
3.6	Molecula	ar detection	57
3.6.1	RNA ext	raction	58
	3.6.1.1	Sample preparation	58
	3.6.1.2	QIAamp Viral RNA Mini Kit	59
	3.6.1.3	TRIzol® RNA extraction	59
	3.6.1.4	RNeasy Mini Kit	60
3.6.2	Oligonud	cleotide primers	61
	3.6.2.1	Type-common primers	61
	3.6.2.2	Type-specific primers	62
3.6.3	Optimisa	ition of RT-PCR	63
361	Amplification by RT-PCR		63

			Page
	3.6.4.1	Mon2/Mon67	64
	3.6.4.2	Mon348/Mon340	65
	3.6.4.3	Mon2/prBEG	65
	3.6.4.4	Type-specific RT-PCR	66
3.6.5	Detection	of RT-PCR amplicons	66
	3.6.5.1	Polyacrylamide gel electrophoresis	66
	3.6.5.2	Agarose gel electrophoresis	66
	3.6.5.3	Oligonucleotide probe hybridisation assay	66
3.7	Molecular	characterisation	67
3.7.1	PCR produ	act sequencing	68
3.7.2	Phylogene	tic analysis	68
3.8	Statistical	analysis	70
CHAP	TER 4:	RESULTS	72
4.1	Specimens	s and samples	72
4.1.1	Human sto	pol specimens	72
4.1.2	Animal sto	pol specimens	73
4.1.3	Water and	sewage samples	74
4.1.4	AstV isola	tes from river, dam and sewage samples	75
4.2	Optimisati	on of RT-PCR for the detection of AstVs	75
4.2.1	Optimisati	on of HAstV type-common RT-PCRs	75
	4.2.1.1	Mon2/Mon67 primer pair	75
	4.2.1.2	Mon348/Mon340 primer pair	75
	4.2.1.3	Mon2/prBEG primer pair	78
4.2.2	Optimisati	on of HAstV type-specific RT-PCRs	78
4.3	Assessme	nt of cell cultures for the isolation of HAstVs	79
4.4	Detection	and characterisation of AstVs	82
4.4.1	Human sto	pol specimens	82
4.4.2	Animal sto	pol specimens	88



		Page
4.4.3	Water and sewage samples	89
4.4.4	AstV isolates from river, dam and sewage sam	ples 92
4.5	Nucleotide sequence and phylogenetic analysis	of South
	African strains	94
СНА	PTER 5 : DISCUSSION	103
СНА	PTER 6: CONCLUSION	113
СНА	PTER 7 ; REFERENCES	115
APPE	NDIX A	145
A.1	Glass wool adsorption elution procedure	
A.2	PEG/NaCl concentration method	
APPE	NDIX B	147
B.1	Procedure for nucleic acid sequencing reaction	S
APPE	NDIX C	150
C.1	Summary of astrovirus detection from specimens	animal stool
APPE	NDIX D	153
D.1	ABSTRACT: Nadan S, Grabow WOK, Tayl	or MB. The
	molecular detection and characterisation of ast	roviruses from
	human stool specimens and sewage.	



- D.2 ABSTRACT: Taylor MB, Nadan S, Grabow WOK, Walter JE. Molecular epidemiology of human astroviruses from the Tshwane area (Pretoria), Gauteng.
- D.3 ABSTRACT: Nadan S, JE Walter, Grabow WOK, Taylor MB. The molecular detection and characterisation of astroviruses from human stool specimens and sewage.
  - D.4 ABSTRACT: WB van Zyl, S Nadan, JC Vivier, JME Venter, K Riley, EKM Tlale, LR Seautlueng, WOK Grabow, MB Taylor. The prevalence of enteric viruses in patients with gastroenteritis in the Pretoria and Kalafong Academic Hospitals, South Africa.

APPENDIX E 159

Nadan S, Walter JE, Grabow WOK, Mitchell DK, Taylor MB. Molecular characterization of astroviruses: comparison between clinical and environmental isolates from South Africa.



# LIST OF TABLES

		age
Table 1:	Polypeptide composition of selected astroviruses during all of replication	13
Table 2:	Sensitivity of different techniques for the detection of HAstVs	27
Table 3:	Composition of different PCR buffers tested in the RT-PCR optimisation reactions	64
Table 4:	Year-to-year detection of HAstV, HRV and HAdV-40/41 in human stool specimens from tertiary hospitals	72
Table 5:	Animal host, collection site and consistency of animal and bird stool specimens analysed	73
Table 6:	Volume and type of water and sewage samples screened for astroviruses (AstVs)	74
Table 7:	Astrovirus (AstV) positive from water and sewage samples from different geographic regions in southern Africa	76
Table 8:	Assessment of the sensitivity of cell cultures for the isolation and propagation of human astroviruses (HAstVs)	81
Table 9:	The human astrovirus (HAstV) detection results from human stool specimens and cell culture derivatives thereof	83
Table 10:	Summary of characterisation of astrovirus isolates from human stool specimens.	85
Table 11:	Summary of virus detection results on animal stool specimens where screening results were query positive	88
Table 12:	Detection and characterisation of astroviruses (Ast in sewage and water samples collected, from April 1999 to October 2000, from the Tshwane Metropolitan Area	Vs) 90



		Page
Table 13:	Detection and characterisation of astrovirus isolates from water and sewage samples referred for routine virological analysis	93
Table 14:	Summary of the characterised South African (SA) human astrovirus (HAstV) strains from clinical and sewage sources identifying the representative strain included in the phylogenetic analysis	95
Table 15:	Summary of pairwise sequence comparisons between South African (SA) astrovirus (AstV) strains and the AstV isolates in GenBank	101



# LIST OF FIGURES

	Pa	age
Figure 1:	Microbial agents associated with infant and childhood diarrhoea in developed and developing countries	2
Figure 2:	Electron micrograph of negatively stained HAstV in a faecal specimen	6
Figure 3:	Image processing of negatively stained HAstV-1 viewed along the two-fold axis of symmetry	8
Figure 4:	Genomic organisation of HAstV-1	17
Figure 5:	Phylogenetic analysis of human astrovirus nucleotid sequences	e 22
Figure 6:	Analysis of PCR products derived from RT-PCR amplification of HAstV-1 to 7 reference strains and HAstV-8 positive stool specimen with type-common primers Mon2/Mon67	77
Figure 7:	Agarose gel analysis of 289 base pair (bp) RT-PCR products amplified from HAstV-6 RNA using the buffers 1 to 12 from the Opti-Prime™ PCR Optimization Kit	77
Figure 8:	Analysis of 319 bp amplicons derived from HAstV-1 to 3 and HAstV-5 to 7 reference strains, and the HAstV-8 positive stool specimen using the optimised RT-PCR reaction mix and primers pair Mon2/prBEG	78
Figure 9:	Agarose gel analysis of RT-PCR products derived from HAstV-1 to 7 reference strains using type- specific primers in the optimised type-specific RT-PCR reaction mix	79
Figure 10:	Distribution of HAstV genotypes identified between January 1996 and October 2000 in human diarrhoeal stool specimens, taken from hospitalised patients from the Tshwane Metropolitan Area	87



Figure 12: Relationship of selected South African human astrovirus (HAstV) sequences of ORF2, to the Oxford (Ox) reference strains  Figure 13: Maximum likelihood phylogenetic tree based on a 208 nucleotide (nt) region of the 3' end of ORF2 showing the relationships between representatives of the South African environmental and clinical human astrovirus (HAstV) isolates and the		P	age
astrovirus (HAstV) sequences of ORF2, to the Oxford (Ox) reference strains  Figure 13: Maximum likelihood phylogenetic tree based on a 208 nucleotide (nt) region of the 3' end of ORF2 showing the relationships between representatives of the South African environmental and clinical human astrovirus (HAstV) isolates and the	Figure 11:	Distribution of HAstV genotypes detected between April 1999 and October 2000 in sewage samples collected from sewage works in the Tshwane	_
208 nucleotide (nt) region of the 3' end of ORF2 showing the relationships between representatives of the South African environmental and clinical human astrovirus (HAstV) isolates and the	Figure 12:	astrovirus (HAstV) sequences of ORF2, to the	97
prototypes of masty types 1 to 8	Figure 13:	208 nucleotide (nt) region of the 3' end of ORF2 showing the relationships between representatives of the South African environmental and clinical	98



## **ABBREVIATIONS**

Å : angstrom
aa : amino acid
Ab : antibody

AMPS : ammonium persulfate

AMV : avian myeloblastosis virus

Ag : antigen

ANV : avian nephritis virus

ARC : Agricultural Research Council

AstV : astrovirus

ATCC : American Type Culture Collection

BAstV : bovine astrovirus

BK : bovine kidney cell line

bp : base pair

(C) : carboxy terminal CCC : child care centre

CaCo-2 : colonic carcinoma cell line

cpe : cytopathic effect
CsCl : caesium chloride
CV : caliciviruses
DAstV : duck astrovirus

DTT : dithiothreitol

EIA : enzyme immunoassay

ELISA : enzyme-linked immunosorbent assay

EM : electron microscopy

E-MEM : Eagle's Minimum Essential Medium

EtBr : ethidium bromide
FAstV : feline astrovirus
FCS : foetal calf serum
FEA : feline embryo cells

GBEB : glycine-beef-extract buffer g/cm<sup>3</sup> : gram per cubic centimeter

g/ml : gram per millilitre

HAdV-40/41: human adenovirus 40/41

HAstV : human astrovirus

HAstV-1 human astrovirus serotype 1 HAstV-2 human astrovirus serotype 2 HAstV-3 human astrovirus serotype 3 HAstV-4 human astrovirus serotype 4 HAstV-5 human astrovirus serotype 5 HAstV-6 human astrovirus serotype 6 HAstV-7 human astrovirus serotype 7 : HAstV-8 human astrovirus serotype 8 HIV : Human Immunodeficiency Virus
HEK : human embryonic kidney cell line
HEL : human embryonic lung fibroblasts

HRP : horseradish peroxidase

HRV : human rotavirus HS : horse serum

h : hour

IEM : immune electron microscopy

IF : immunofluorescence

ISEM : immunosorbent electron microscopy

kb : kilobase kDa : kilodalton L : litres

LLCMK2 : rhesus monkey kidney cell line

MAb : monoclonal antibody

MDBK : Madin-Darby bovine kidney cell line

min : minute ml : millilitre

MW : molecular weight
(N) : amino terminal
NaCl : sodium chloride
NCR : non-coding region

NBK : neonatal bovine kidney cell line NIV : National Institute for Virology

nm : nanometre
nt : nucleotide
NV : Norwalk virus
OAstV : ovine astrovirus
ORF : open reading frame
PEG : polyethylene glycol

PAGE: polyacrylamide gel electrophoresis

PAstV : porcine astrovirus

PBK : primary bovine kidney cell line
PBS : phosphate buffered saline
PCR : polymerase chain reaction

RFLP : restriction fragment length polymorphism

p.i. ; post infection

PLC/PRF/5 : primary hepatoma cell line

RIA : radio immunoassay RNA : ribonucleic acid

rRNA : ribosomal ribonucleic acid

RT : reverse transcription

RT-PCR : reverse transcriptase-polymerase chain reaction

SA : South Africa

SDS-PAGE: sodium dodecyl sulphate-polyacrylamide gel

electrophoresis

SPIEM : solid phase immune electron microscopy

SRV : small round virus

SRSV : small round structured virus

ss : single stranded

TEMED : N,N,N',N'-tetramethyl-ethylenediamine

tRNA : transfer ribonucleic acid

TYPE-EIA: typing enzyme immunoassay

TAstV : turkey astrovirus

TAstV-1 : turkey astrovirus serotype 1
TAstV-2 : turkey astrovirus serotype 2
TMB : 3,3',5,5'-tetramethylbenzidine

UK : United Kingdom

UNICEF : United Nations International Children's Fund

US : United States of America

UTR: untranslated region

UV : ultraviolet WT : wild type