

## Concentration and derivatization in silicone rubber

### traps for mass spectrometric and gas chromatographic

## analysis of air and water pollutants

 $\mathbf{B}\mathbf{Y}$ 

# MARIA JOSÉ FERNANDES-WHALEY

Submitted in partial fulfilment of the requirements for the degree

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in the Faculty of Natural and Agricultural Science

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### SUMMARY

Estrogens, alkylphenols and bisphenol-A, enter the environment through waste water systems and waste disposal of manufactured products e.g. detergents, paints, polycarbonates and flame-retardants. These analytes disrupt the endocrine function of living organisms affecting their reproductive health and those of future generations. Gas phase low molecular- mass aldehydes and amines are typically eye, nose, and throat irritants. Formaldehyde is classified as a probable human carcinogen. Given their negative impact on human health it is urgent to monitor pollutants at extremely low levels in both air and water. The aqueous pollutants are often concentrated using solid phase extraction cartridges or liquid-liquid extraction followed by derivatization. Methods that can most effectively and selectively pre-concentrate aldehydes and amines involve *in situ* 



derivatization. Unfortunately, the derivatizing reagents as well as their associated solvents or adsorbents, are responsible for problems encountered with these methods.

Polydimethylsiloxane (PDMS) has emerged as the ideal concentration and reaction medium for trace analysis. However the expensive commercial devices such as SPME and SBSE both require the samples to be returned to the laboratory for concentration. Due to the open tubular nature of the PDMS multichannel trap (MCT), developed in our laboratory, it is ideally suited for on-site and on-line sampling. The MCTs have a high analyte capacity owing to the large volume of PDMS available for concentration. The derivatization reaction can be performed *in situ* providing a "one-pot concentration and reaction device". This allows for reduced risk of contamination of / or losses of the sample and a sampling method that can cater for both air and water samples.

To demonstrate the versatility of the PDMS MCT, two approaches for concentration in PDMS were investigated in this study, namely, 1) the on-line concentration and *in situ* derivatization of volatile polar analytes from air followed by REMPI-TOFMS detection, and 2) the concentration of phenolic lipophilic analytes from water requiring derivatization prior to analysis by GC/MS.

1) Analyte and derivatizing reagent were simultaneously introduced into the PDMS trap using a ypress-fit connector. The reaction occurs in situ followed by thermal desorption using a thermal modulator array alone or in conjunction with a thermal desorption unit. The aldehydes and amine derivatives were successfully detected by the REMPI-TOFMS. Reaction efficiencies were determined temperature without catalysts. Formaldehyde vielded a low at room reaction/concentration efficiency of 41 % with phenylhydrazine in PDMS, while acetaldehyde, acrolein and crotonal displayed much improved values of 92, 61 and 74 % respectively. Both propylamine and butylamine yielded 28 % reaction/concentration efficiency with benzaldehyde in the PDMS matrix. Detection limits obtained with this technique were significantly lower than the permissible exposure limits set by the Occupational Safety and Health Administration. It should be noted that the detection limits were not determined by actual measurement but by extrapolation from a larger signal.

2) Aqueous analytes were concentrated in the PDMS MCT using a gravity flow rate of ~50  $\mu$ l/min. The trap was dried and 5  $\mu$ l derivatizing reagent added. At room temperature and without the presence of a catalyst, the reaction of alkylphenols with trifluoroacetic acid anhydride in the PDMS matrix was 100% complete after 5 minutes. Bisphenol-A reacted less than 50 % to completion



during this period, but the amount of derivative formed remained constant. This study revealed that extraction efficiencies of the alkylphenols and bisphenol-A off the PDMS trap have poor batch-tobatch repeatability indicating that the PDMS matrix was not homogenous. For two different PDMS batches: *tert*-octylphenol displayed an extraction efficiency of 70 and 79%, nonylphenol displayed 84 and 43% while Bisphenol-A displayed 10 and 26% respectively. The thermally desorbed derivatives were analysed by GC/MS. Despite background contamination in the desorption unit, detection limits were at the ppt level. Detection limits were not determined by actual measurement but by extrapolation from a larger signal.

Real samples were also tested.

**Keywords**: air pollutants, water pollutants, concentration, *in situ* derivatization, polydimethylsiloxane, PDMS, multichannel traps, thermal desorption, gas chromatography, mass spectrometry, resonance enhanced time-of-flight mass spectrometry.



# Konsentrering en derivatisering in silikoonrubbervalle vir massaspektrometriese en gaschromatografiese analise van lugen waterbesoedelstowwe

DEUR

## MARIA JOSÉ FERNANDES-WHALEY

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### SAMEVATTING

Estrogene, soos alkielfenole en bisfenol-A, beland in die omgewing deur afvalwatersisteme en die wegdoening van vervaardigde produkte soos wasmiddels, verf, polikarbonate en vlamvertragers. Hierdie analiete ontwrig die endokrienfunksie van lewende organismes, en affekteer hul eie voortplantingsgesondheid sowel as dié van hul toekomstige geslagte. Gasfase laemolekulêremassa aldehiede en amiene is tipies oog-, neus- en keel-irritanse. Formaldehied is geklassifiseer as 'n waaarskynlike menslike karsinogeen. In die lig van hul negatiewe impak op menslike gesondheid is dit dringend noodsaaklik om hierdie besoedelstowwe te moniteer by uiters lae konsentrasies in beide lug en water. Besoedelstowwe in water word dikwels gekonsentreer met soliedefaseekstraksiepatrone gevolg deur derivatisering. Metodes wat aldehiede en amiene doeltreffend vooraf konsentreer, behels *in situ* derivatisering. Ongelukkig is die derivatiseringsreagense sowel as hul oplosmiddels of adsorbente verantwoordelik vir probleme met hierdie metodes.

Polidimetielsiloksaan (PDMS, silikoon) het ontluik as die ideale konsentrerings- en reaksiemedium vir spooranalise. Die duur kommersiële toestelle soos SPME (soliedefase-mikroekstraksie) en SBSE (magnetieseroerder-ekstraksie) vereis egter dat die monsters na die laboratorium gestuur moet word vir konsentrering. As gevolg van die oopbuis geaardheid van die PDMS multikanaalval



(MKV) wat in ons laboratorium ontwikkel is, is dit ideaal geskik vir ter plaatse- en aanlynmonstering. Die MKV's het 'n groot kapasiteit vir analiete as gevolg van die groot volume PDMS beskikbaar vir konsentrering. Die derivatiseringsreaksie kan binne-in die val uitgevoer word, wat 'n "eenpot konsentrerings- en reaksietoestel" tot gevolg het. Dit lei tot 'n verminderde risiko van kontaminasie en/of verliese van die monster, en 'n monsteringsmetode wat geskik is vir beide watersowel as lugmonsters.

Om die veelsydigheid van die PDMS multikanaalval te demonstreer is twee prosedures ondersoek om stowwe in PDMS te konsentreer, naamlik: 1) aanlyn konsentrering en *in situ* derivatisering van vlugtige polêre analiete uit lug, gevolg deur REMPI-TOFMS (resonansversterkte multifotonionisasie - vlugtydmassaspektrometrie) deteksie, en 2) die konsentrering van fenoliese lipofiliese analiete uit water, met derivatisering voor analise met GC-MS (gaschromatografie – massaspektrometrie).

- Analiet en derivatiseringsreagens is tegelykertyd gevoer in 'n PDMS-val met 'n Y-koppelstuk. Die reaksie vind *in situ* plaas, gevolg deur termiese desorpsie met 'n termiese modulatoropstelling alleen, of saam met 'n termiese desorpsie-eenheid. Die aldehiede en amienderivate is suksesvol aangedui met 'n REMPI-TOFMS. Reaksiedoeltreffendhede is bepaal by kamertemperatuur sonder katalisatore. Formaldehied het ondoeltreffend gereageer en gekonsentreer (41%) met fenielhidrasien in PDMS, terwyl asetaldehied, akroleïen en krotonal baie beter waardes gegee het, nl. 92%, 61% en 74% respektiewelik. Beide propielamien en butielamien het 'n doeltreffendheid van 28% gehad met bensaldehied in die PDMS-matrys. Deteksielimiete met hierdie tegniek was aansienlik laer as die toelaatbare blootstellingslimiete van die Beroepsveiligheids- en Gesondheidsadministrasie.
- 2) Waterige analiete is in die PDMS gekonsentreer met 'n swaartekragvloeitempo van ongeveer 50 μl/min. Die val is gedroog en 5 μl derivatiseringsreagens is bygevoeg. By kamertemperatuur en sonder katalis was die reaksie van alkielfenole met trifluoorasynsuuranhidried in die PDMS-matriks 100% volledig na 5 minute. Bisfenol-A het minder as 50% volledig gereageer in hierdie tydperk, maar die hoeveelheid derivaat wat gevorm het, het konstant gebly. Ekstraksie-doeltreffendhede van alkielfenole en bisfenol-A het swak herhaalbaarheid getoon tussen besendings buise, wat aandui dat die PDMS-matriks nie homogeen was nie. Vir twee verskillende klompe PDMS het *ters*-oktielfenol 'n doeltreffendheid getoon van 70% en 79%, nonielfenol 84% en 43%, en bisfenol-A 10% en 26%. Die termiesgedesorbeerde derivate is geanaliseer met GC-MS. Ten spyte van agtergrondkontaminasie in die desorbeerder was deteksielimiete by die dele-per-triljoenvlak. Regte veldmonsters is ook getoets.



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## ABBREVIATIONS

| AAA            | - | acetic acid anhydride                                |
|----------------|---|--|
| ACGIH          | - | American Conference of Governmental Industrial       |
| BEA            | _ | benzylethanolamine                                   |
| BPA            | - | Bisphenol-A  |
| BSA            | - | n-o-bis(trimethylsilyl) acetamide                    |
| BSTFA          | _ | n-o-bis (trimethylsilyl) trifluoroacetamide          |
| C              | _ | analyte concentration/ alkane carbon number          |
| CIS            | _ | cooled injection system                              |
| CI             |   | confidence level                                     |
| C C            | _ | initial analyte concentration in sample              |
|                | - | DDMS concentration                                   |
|                | - | as the way   |
| C w            | - | calbowax   |
|                | - | water concentration                                  |
| D <sub>c</sub> | - | analyte distribution ratio between 2 phases          |
|                | - | diffusion constant (m.s.) of analyte in mobile phase |
| DNBS           | - | dinitrobenzene sulphonic acid                        |
| DNFB           | - | dinitrofluorobenzene                                 |
| DNPH           | - | dintirophenylhydrazine                               |
| DNSH           | - | dansylhydrazine                                      |
| $d_p$          | - | particle diameter                                    |
| DVB            | - | divinylbenzene                                       |
| E1             | - | estrone  |
| E2             | - | 17β-estradiol  |
| E3             | - | estriol  |
| ECD            | - | electron capture detector                            |
| EDC            | - | endocrine disrupting compound                        |
| EDU            | - | Airsense® enrichment desorption unit                 |
| EE2            | - | 17α-ethinylestradiol                                 |
| EI             | - | electron impact ionization                           |
| ELISA          | - | enzyme linked immunosorbent assay                    |
| EPA            | - | U.S. Environmental Protection Agency                 |
| ESI            | - | electrospray ionization                              |
| F              | - | column flow rate                                     |
| FIA            | - | flow injection analysis                              |
| FID            | _ | flame ionization detector/ detection                 |
| GC             | - | gas chromatgraphy                                    |
| GWRC           | _ | Global Water Research Coalition                      |
| Н              | _ | nlate height   |
| НСНО           | _ | formaldehyde   |
| HFR_           | _ | hentafluorobutyrl                                    |
| HER A          | - | heptafluorobutyric acid anhydride                    |
| HEBCI          | - | heptafluorobutanovl chloride                         |
|                | - | hydroxymethylpiperidine                              |
|                | - | high performence liquid chromate grapher             |
|                | - | nigh performance inquid chromatography               |
| $n_r$          | - | reduced plate neight                                 |
| HSSE           | - | neadspace sorptive extraction                        |
|                | - | ion trap detector                                    |
| K              | - | capacity factor                                      |

#### Abbreviations



| Κ                 | - | equilibrium distribution coefficient                            |
|-------------------|---|---|
| Ka                | - | acid in water dissociation constant                             |
| K <sub>fg</sub>   | - | SPME fibre/ gas distribution constant                           |
| K <sub>fh</sub>   | - | SPME fibre/ headspace distribution constant                     |
| K <sub>fs</sub>   | - | distribution coefficient between the SPME fibre and sample      |
| K                 | - | octanol-water partitioning coefficient                          |
| L                 | _ | column/ trap length   |
| LASER             | - | Light Amplification by Stimulated Emission of Radiation         |
| LC                | _ | liquid chromatography   |
| LLE               | - | liquid-liquid extraction  |
| LOD               | _ | limit of detection  |
|                   | _ | limit of quantitation   |
| LPME              | _ | liquid phase microextraction                                    |
| ITPRI             | _ | linear temperature programmed retention index                   |
| M                 | _ | neutral molecule  |
| M*                | _ | high-energy molecule  |
| 101               |   | total mass of analyte in the sample                             |
| $m_0$             | - | multichennel tren   |
| MC I              | - | analyte mass in DDMS  |
| m <sub>PDMS</sub> | - | analyte mass in FDMS  |
| MPI               | - |   |
| MSD MSD           | - | mass spectrometry   |
| MSD               | - | mass selective detector   |
|                   | - | n-metnyl-n-(trimetnylsilyl)-trifluoroacetamide                  |
| MIBSIFA           | - | n-( <i>tert</i> -butylaimetnylsilyi)-n-metnyltrifluoroacetamide |
| N                 | - | theoretical number of plates                                    |
| n                 | - | amount extracted  |
| NCI               | - | negative chemical ionization                                    |
| Nd: YAG           | - | neodymium-doped yttrium aluminium garnet                        |
| NIOSH             | - | National Institute for Occupational Safety and Health           |
| NP                | - | 4-nonylphenol   |
| NPD               | - | nitrogen phosphorous detector                                   |
| NSD               | - | nitrogen specific detector                                      |
| OSHA              | - | Occupational Safety and Health Administration                   |
| OTT               | - | open tubular trap   |
| $\mathbf{p}_{0}$  | - | column outlet pressure  |
| PA                | - | polyacrylate  |
| PAH               | - | polyaromatic hydrocarbon  |
| PCB               | - | polychlorinated biphenyl  |
| PDMS              | - | polydimethylsiloxane  |
| PEL               | - | permissible exposure limits                                     |
| PFBA              | - | pentafluorobenzaldehyde   |
| PFBBr             | - | pentafluorobenzylbromide  |
| PFBC1             | - | pentaflurobenzoyl chloride                                      |
| PFBHA             | - | pentafluorobenzylhydroxylamine                                  |
| PFBOH             | - | pentaflurobenzoic acid  |
| PFP-              | - | pentafluoropropionyl  |
| PFPAA             | - | pentafluoropropionic acid anhydride                             |
| PFPH              | - | pentafluorophenylhydrazine                                      |
| PTV               | - | programmed temperature vaporization                             |
| p <sub>i</sub>    | - | column inlet pressure   |
| p <sub>m</sub>    | - | flow meter pressure   |
| * ···             |   | L   |





| ppb               | - | part-per-billion   |
|-------------------|---|--|
| PPBT              | - | packed particle bed trap                                 |
| ppm               | - | part-per-million   |
| ppt               | - | part-per-trillion  |
| p <sub>w</sub>    | - | saturated water vapour pressure                          |
| REMPI             | - | resonance enhanced multiphoton ionization                |
| RI                | - | retention index  |
| RIC               | - | reconstructed ion chromatogram                           |
| SBSE              | - | stir bar sorptive extraction                             |
| SEP               | - | sample enrichment probe                                  |
| SIBA              | - | n-succinimidyl benzoate                                  |
| SIM               | - | selected ion monitoring                                  |
| SPE               | - | solid phase extraction                                   |
| SPI               | - | single photon ionization                                 |
| SPME              | - | solid phase microextraction                              |
| Тс                | - | column temperature                                       |
| Т                 | - | absolute temperature                                     |
| Т                 | - | 17β -testosterone  |
| ТСРН              | - | trichlorophenylhydrazine                                 |
| TCT - CP 4020     | - | Chrompack® thermal desorption cryotrap unit              |
| TDS               | - | thermal desorption system                                |
| TDU-CIS           | - | Gerstel® thermal desorption unit cooled injection system |
| TFA               | - | trifluoroacetic/acetate                                  |
| TFAA              | - | trifluoroacetic acid anhydride                           |
| TIC               | - | total ion chromatogram                                   |
| t <sub>m</sub>    | - | unretained compound retention time                       |
| $T_m$             | - | flow meter temperature                                   |
| ТМА               | - | thermal modulator array                                  |
| TMCS              | - | trimethylchlorosilane                                    |
| TMS               | - | trimethylsilyl   |
| TMSI              | - | n-trimethylsilylimidazole                                |
| TOFMS             | - | time-of-flight mass spectrometry                         |
| ТОР               | - | <i>tert</i> -octylphenol                                 |
| t <sub>r</sub>    | - | analyte retention time                                   |
| u                 | - | linear velocity $(m.s^{-1})$                             |
| UV                | - | ultraviolet  |
| $\mathbf{V}_{0}$  | - | void volume  |
| V <sub>b</sub>    | - | breakthrough volume                                      |
| $V_{\rm f}$       | - | volume of the fibre                                      |
| $V_L$             | - | column stationary phase volume                           |
| VOC               | - | volatile organic compound                                |
| V <sub>PDMS</sub> | - | PDMS volume  |
| Vr                | - | retention volume   |
| Vs                | - | sample volume  |
| VUV               | - | vacuum ultraviolet                                       |
| V <sub>w</sub>    | - | water volume   |
| WHO               | - | World health organization                                |
| β                 | - | phase ratio  |
| v                 | - | reduced velocity in the trap (packed column)             |
| ω                 | - | base width of analyte peak                               |



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Figure 6.40 (B) Extracted ion chromatogram of PDMS degradation peaks m/z 73, 207, 211 and 281.

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Figure 6.42 Selected Ion Mode (SIM) chromatogram of the 20 ml UP Sports Centre river water sample on PDMS MCT M3. Sample extracted at a flow rate of 50  $\mu$ l/min, dried and allowed to react with 5  $\mu$ l trifluoroacetic acid anhydride for 10 min, followed by thermal desorption. Selected ions were m/z 231, 203, 245, 316, 405, 420. *Tert*-octylphenol trifluoroacetate derivative (TOP-TFA) m/z 231, t<sub>R</sub> = 21.2 min; 4-*n*-nonylphenol trifluoroacetate derivative (NP-TFA) m/z 203, t<sub>R</sub> = 25.6 min; bisphenol-A trifluoroacetate derivative (BPA-TFA) m/z 405, t<sub>R</sub> = 27.2 min.

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#### **APPENDICES:**

Figure A2.1 Reaction efficiency graphs for the on-line derivatization of acrolein and crotonal with phenylhydrazine. The graph displays i) the amount of gas standard released over that time interval as determined by their permeation rate and ii) the amount of analyte gas trapped using *in-situ* derivatization on the SPME fibre as calculated using the internal standard and effective carbon number response for the signal obtained from the GC-FID for the derivative. A comparison of the gradients obtained from the standard and the actual amount of analyte trapped gives an approximation of the reaction/trapping efficiency for this reaction.

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#### Figure A3.1

A) GC-TOFMS chromatogram obtained for the underivatized phenols, TOP  $t_R = 17.47$ min, NP  $t_R = 20.30$  min and BPA  $t_R = 26.25$  min.

B) GC-TOFMS confirmation chromatogram for the trifluoroacetate derivatives prepared in a vial in acetone as described in section 6.2.6. TOP-TFA  $t_R = 15.99$  min, NP-TFA  $t_R =$ 19.00 min and BPA-TFA  $t_R = 20.35$  min. *Notice the absence of underivatized phenols*.

Figure A3.2 GC-TOFMS mass spectrum obtained for the TOP-TFA derivative  $t_R = 15.99$  min. M<sup>+</sup> m/z 302, base peak m/z 231.

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