

**Selective induction of apoptosis by 7-
methyljuglone, its derivatives and isolated
compounds from *Foeniculum vulgare* Mill.
on human cancer cells**

by

Brigitte Binneman

Submitted in partial fulfilment of the requirements for the degree of

Masters of Science (Plant Science)

University of Pretoria

Faculty of Natural and Agricultural Sciences

Department of Plant Science

Supervisor: Prof Namrita Lall

September 2008

Table of contents

List of Figures.....	i
List of Tables.....	vii
List of Abbreviations.....	x
Abstract.....	xiii

Chapter 1 Introduction

1.1 Background.....	1
1.2 The role of natural products in western medicine.....	2
1.3 Global use of plants as medicine.....	6
1.4 Medicinal plant use in Africa.....	6
1.5 Plants as a source of anti-cancer agents.....	8
1.6 Plant derived anticancer agents in clinical development.....	16
1.7 Targeting natural products.....	20
1.8 Rationale for studying anticancer botanicals.....	21
1.9 Problem statement.....	22
1.10 The aim of the study.....	22
1.11 References.....	24

Chapter 2 Cell death and cancer

2.1 Cell death.....	30
2.2.1 Apoptosis.....	33
2.2.2 Necrosis.....	37
2.2.3 Oncosis.....	39
2.2.4 Autophagy.....	40
2.1 Cancer.....	40
2.1.1 What is cancer?.....	40
2.1.2 Types of cancer.....	42



2.1.3	Benign and malignant tumours.....	43
2.1.4	Cancer stages.....	43
2.1.5	Cancer globally.....	45
2.1.6	South African cancer statistics.....	47
2.1.7	Unproven methods for cancer treatment.....	56
2.1.8	Cancer prevention.....	57
2.1.9	Cancer treatment.....	59
1.	Surgery.....	59
2.	Radiation.....	59
3.	Chemotherapy.....	60
4.	Hormones.....	61
2.3	References.....	62

Chapter 3 Anticancer activity of traditionally used plant extracts

3.1	Introduction	67
3.2	Asteraceae.....	68
3.2.1	Ethnobotanical use of <i>Artemisia</i>	68
3.2.2	<i>Artemisia afra</i>	69
3.2.3	Phytochemicals in the <i>Artemisia</i> genus.....	69
3.3	Apiaceae	70
3.3.1	Ethnobotanical use of <i>Centella</i>	70
3.3.2	<i>Centella asiatica</i>	70
3.3.3	Phytochemicals in the <i>Centella</i> genus.....	71
3.4	Ebenaceae.....	72
3.4.1	Ethnobotanical use of <i>Euclea</i>	72
3.4.2	<i>Euclea natalensis</i>	73
3.4.3	Phytochemicals in the <i>Euclea</i> genus.....	75
3.5	Euphorbiaceae.....	76
3.5.1	Ethnobotanical use of <i>Euphorbia</i>	76
3.5.2	<i>Euphorbia ingens</i>	76
3.5.3	Phytochemicals in the <i>Euphorbia</i> genus.....	77
3.6	Apiaceae.....	78
3.6.1	Ethnobotanical use of <i>Foeniculum</i>	78



3.6.2	<i>Foeniculum vulgare</i>	79
3.6.3	Phytochemicals in the <i>Foeniculum</i> genus.....	80
3.7	Hypoxidaceae	83
3.7.1	Ethnobotanical use of <i>Hypoxis</i>	83
3.7.2	<i>Hypoxis hemerocallidea</i>	84
3.7.3	Phytochemicals in the <i>Hypoxis</i> genus.....	84
3.8	Alliaceae.....	85
3.8.1	Ethnobotanical use of <i>Tulbaghia</i>	85
3.8.2	<i>Tulbaghia violacea</i>	85
3.8.3	Phytochemicals in the <i>Tulbaghia</i> genus.....	86
3.9	Positive controls used for cytotoxicity.....	87
3.9.1	Doxorubicin a quinonoid anticancer drug.....	87
3.9.2	Zearalenone a phytoestrogen.....	88
3.10	XTT assay.....	89
3.2	Materials and Methods.....	90
3.2.1	Collection of plant material.....	90
3.2.2	Extraction of plant materials.....	91
3.2.3	Cell lines.....	92
3.2.3	Cytotoxicity assay.....	93
3.3.4	Statistical analysis with GraphPad Prism4.....	93
3.3	Results.....	96
3.4	Discussion.....	96
3.5	References.....	100

Chapter 4 Cytotoxicity of 7-methyljuglone and its derivatives on cancer cells and selected few on non cancerous cells

4.1	Introduction.....	108
4.1.1	Quinonoids.....	109
4.1.2	Bioactivity, cytotoxicity of 7-methyljuglone.....	111
4.2	Materials and Methods.....	112
4.2.1	Synthesis of 7-methyljuglone and its derivatives.....	112
4.2.2	Culture of cancer cells.....	114



4.2.3	Cytotoxicity in peripheral blood mononuclear cells (PBMCs)	114
4.3	Results	115
4.3.1	Cytotoxicity on four human cancer cell lines	115
4.3.2	Cytotoxicity on U937 cells	119
4.3.3	Cytotoxicity on PBMCs	119
4.4	Discussion	121
4.4.1	Cytotoxicity on four human cancer cell lines	121
4.4.2	Cytotoxicity on U937 cells	122
4.4.3	Cytotoxicity on PBMCs	123
4.5	References	124

Chapter 5 Isolation of the bioactive compounds of *Foeniculum vulgare*

5.1	Introduction	128
5.1.1	Chromatography	128
5.1.4	Steps for isolation	129
5.2	Materials and Methods	130
5.2.1	Collection of plant material	130
5.2.2	Isolation of bioactive compounds	130
5.2.2.1	Preparation of extract	130
5.2.2.2	Column chromatography	130
5.2.2.3	Identification of the isolated compounds	132
5.2.2.4	Culture of cancer cells	134
5.2.2.5	Cytotoxicity of various fractions from <i>F. vulgare</i>	134
5.2.2.6	Cytotoxicity of the compounds Isolated <i>F. vulgare</i>	134
5.2.2.7	Cytotoxicity of the compounds Isolated <i>F. vulgare</i> on U937 cells	135
5.2.2.8	Cytotoxicity of the compounds Isolated <i>F. vulgare</i> on PBMCs	136
5.3	Discussion	136
5.3.1	Cytotoxicity of the first fractions from <i>F. vulgare</i> (Bioassay guided isolation)	136
5.3.2	Cytotoxicity of the compounds isolated from <i>F. vulgare</i>	137
5.3.3	Cytotoxicity of the compounds Isolated <i>F. vulgare</i> on U937 cells	137
5.3.4	Cytotoxicity of the compounds Isolated <i>F. vulgare</i> on PBMCs	137

5.4	References.....	139
-----	-----------------	-----

Chapter 6 Mechanistic studies of potent anticancer compounds

6.1	Different methods to detect different types of cell death with the focus on apoptosis.....	140
6.1.1	Cell cycle analysis.....	140
6.1.2	Annexin V-FITC/PI.....	143
6.1.3	Caspase 3 and 7 activity.....	145
6.1.4	Acridine orange and ethidium bromide nuclear staining.....	151
6.1.5	DNA fragmentation.....	151
6.2	Materials and Methods	154
6.2.1	Cell cycle analysis U937 cells.....	154
6.2.2	Cell cycle analysis MCF-7 cells.....	155
6.2.3	Annexin V-FITC/PI U937 cells.....	155
6.2.4	Annexin V-FITC/PI MCF-7 cells.....	155
6.2.5	Caspase 3/7 activity U937 cells.....	156
6.2.6	Acridine orange and ethidium bromide nuclear staining U937 and THP-1 cells.....	157
6.2.7	DNA fragmentation U937 cells.....	157
6.3	Results.....	158
6.3.1	Cell cycle analysis.....	158
6.3.1.1	Cell cycle analysis U937 cells.....	158
6.3.1.2	Cell cycle analysis MCF-7 cells.....	161
6.3.2	Annexin V-FITC/PI.....	163
6.3.2.1	Annexin V-FITC/PI staining U937 cells.....	163
6.3.2.2	Annexin V-FITC/PI staining MCF-7 cells.....	168
6.3.3	Caspase 3 and 7.....	166
6.3.3.1	Caspase 3 and 7 activity after 24 hours.....	166
6.3.4	Acridine orange and ethidium bromide nuclear staining U937 and THP-1 cells.....	167
6.3.5	DNA fragmentation.....	170
6.5	Discussion.....	170
6.5.1	Cell cycle analysis.....	170
6.5.1.1	Cell cycle analysis U937 cells.....	171
6.5.1.2	Cell cycle analysis MCF-7 cells.....	172
6.5.2	Annexin V-FITC.....	174

6.5.2.1	Annexin V-FITC/PI staining U937 cells.....	174
6.5.2.2	Annexin V-FITC/PI staining MCF-7 cells.....	175
6.5.3	Caspase 3/7 activity after 24 hours.....	175
6.5.4	Acridine orange and ethidium bromide nuclear staining U937 and THP1 cells.....	175
6.5.5	DNA fragmentation.....	176
6.6	References.....	177

Chapter 7 Discussion and conclusion

7.1	Discussion and conclusion.....	181
7.2	Recommendations for future work.....	186

Chapter 8 Acknowledgments

Acknowledgments		187
-----------------	--	-----

Chapter 9 Appendices

Appendix A.....		189
9.1 ¹ H-NMR and ¹³ C-NMR of isolated compounds from <i>Foeniculum vulgare</i>		189
Appendix B.....		194
9.2 Publications and conference presentations resulting from this thesis.....		194
9.2.1 Publications.....		194
9.2.2 Conference presentations.....		194
National.....		194
International.....		195

List of figures

Chapter 1

Figure 1.1 (a) Natural alkaloids 'Vinblastine' and 'Vincristine' isolated from (b) *Catharanthus roseus*. 11

Figure 1.2 Semi-synthetic derivatives of epipodophyllotoxin, an isomer of podophyllotoxin (a) etoposide and (b) teniposide which are clinically active (c) epipodophyllotoxin (d) podophyllotoxin. 12

Figure 1.3 (a) Taxol isolated from (b) *Taxus brevifolia*. 14

Figure 1.4 A natural alkaloid 'camptothecin' from *Camptotheca acuminata*. 15

Figure 1.5 (a) 'Homoharringtonine' isolated from the Chinese tree, *Cephalotaxus harringtonia* var. *drupacea*. 16

Figure 1.6 (a) A novel synthetic flavonoid 'flavopiridol' (b) One of the water-soluble analogs of the combretastatins, 'combretastatin' phosphate (CA4). (c) 'Roscovitine' derived from olomucine. 18

Chapter 2

Figure 2.1 The upper row represents disturbances in growth, differentiation, and tissue integrity that lead to the phenotypes that characterize the different stages of cancer, shown in the lower row. 41

Figure 2.2 New cancer cases for males of all ages in South Africa. 49

Figure 2.3 Cancer deaths for males of all ages in South Africa.	49
Figure 2.4 The 5-year prevalent cases for males 15 years and older in South Africa.	50
Figure 2.5 New cancer cases for females of all ages in South Africa.	51
Figure 2.6 Cancer deaths for females of all ages in South Africa.	51
Figure 2.7 The 5-year prevalent cases for females 15 years and older in South Africa.	52
Figure 2.8 Schematic diagram showing the range of efficacy of chemopreventative agents.	61
 Chapter 3	
Figure 3.1 (a) <i>A. afra</i> (b) The distribution of <i>A. afra</i> in South Africa.	69
Figure 3.2 (a) <i>C. asiatica</i> round or kidney-shaped leaves. (b) The distribution of <i>C. asiatica</i> in South Africa.	71
Figure 3.3 <i>Euclea natalensis</i> : (a) Tree (b) Fruit.	74
Figure 3.4 The distribution of the subspecies of <i>E. natalensis</i> in South Africa.	74
Figure 3.5 (a) The yellowish-green inflorescence of <i>E. ingens</i> . (b) The distribution of <i>E. ingens</i> in Southern Africa.	77
Figure 3.6 (a) The small yellow flowers and leaves are numerous needle-shaped giving <i>F. vulgare</i> a feathery appearance. (b) The distribution of <i>F. vulgare</i> in South Africa.	80
Figure 3.7 (a) The star shaped flowers and long strap like leaves of	

H. hemerocallidea. (b) The distribution of *H. hemerocallidea* is extensively in the grassland areas of South Africa. 84

Figure 3.8 (a) The purple flowers occur in groups at the tip of slender stalks of *T. violacea*. (b) The distribution of *T. violacea* is predominantly in the Eastern Cape and southern KwaZulu-Natal. 86

Figure 3.9 Quinonoid doxorubicin. 88

Figure 3.10 Zearalenone a non-steroidal estrogenic mycotoxin and phytoestrogen. 89

Figure 3.11 Metabolization of XTT to water soluble formazan salt by viable cells. 90

Figure 3.12 Schematic representation of the preparation of the cells and 96-well plates for the experiment. 94

Figure 3.13 Schematic representation of the preparation of extracts/compounds for addition to the 96-well plates which contain the cells. 95

Figure 3.14 Schematic representation of the XTT assay. 96

Chapter 4

Figure 4.1 (A) Compounds 1-15 (B) 19 (C) 16-18. 113

Figure 4.2 (a) A tube before use. (b) After blood collection. (c) After centrifugation. 115

Figure 4.3 Dose response of 8-Bromo-5-hydroxy-7-methyl-1, 4-naphthoquinone on MCF-7 cell viability. 116

Figure 4.4 Dose response curve fit of 8-Bromo-5-hydroxy-7-methyl-

1, 4-naphthoquinone on MCF-7 cells.	116
Figure 4.5 Percentage inhibition of selected compounds on U937 cells. Bars and error bars indicate mean \pm SD of quadruplicates U937.	119
Figure 4.6 Percentage inhibition of selected compounds on peripheral blood mononuclear cells. Bars and error bars indicate mean \pm SD of quadruplicates.	120
Chapter 5	
Figure 5.1 Silica gel column of the <i>F. vulgare</i> ethanol extract.	131
Figure 5.2 TLC plates (hexane: ethyl acetate (6:4) as eluent) after treatment with Vanillin in sulphuric acid (H ₂ SO ₄).	132
Figure 5.3 Syringin the first isolated compound.	133
Figure 5.4 Second isolated compound 4-methoxycinnamyl alcohol.	133
Figure 5.5 Percentage inhibition of selected compounds on U937 cells. Bars and error bars indicate mean \pm SD of quadruplicates.	135
Figure 5.6 Percentage inhibition of selected compounds on peripheral blood mononuclear cells. Bars and error bars indicate mean \pm SD of quadruplicates.	136
Chapter 6	
Figure 6.1 The different stages of the cell cycle G1 (cell grows), S (replication of DNA), G2-(cell prepare to divide) and M (cell division).	142

Figure 6.2 Biological basis of annexin V-FITC binding assay.	144
Figure 6.3 The three sequential stages of the apoptosis cascade.	149
Figure 6.4 Induction of the initiator caspases and activation of the execution caspases which finally leads to apoptotic death as a result of the very complex cascade of events	150
Figure 6.5 Agarose gel electrophoresis of DNA extracted from cultures of P-815 cells. Ethidium bromide stain photographed in ultraviolet light. Lane 1: DRlgest III molecular weight markers; lane 2: control culture; lane 3: culture showing extensive apoptosis induced by heating; lane 4: culture showing massive necrosis 72 hours after repeated freezing and thawing.	153
Figure 6.6 DNA content histograms of U937 cell cycle analysis (a) Control after 24 hours (b) Control after 48 hours (c) Compound 5 (b) after 24 hours (d) Compound 5 after 48 hours (e) Cisplatin (c) after 24 hours (f) Cisplatin after 48 hours (d) (Keyes: C=sub-G1 peak; D=G ₀ /G ₁ peak; E=S peak; F=G ₂ /M peak).	160
Figure 6.7 U937 annexin V-FITC/PI stained (a) after 24 hours on exposure to control (b) after 48 hours on exposure to control (c) after 24 hours on exposure to cisplatin (d) after 48 hours on exposure to cisplatin (e) after 24 hours on exposure to compound 5 (f) after 24 hours on exposure to compound 5.	164
Figure 6.8 MCF-7 annexin V-FITC/PI staining of cells treated with compound 5 (a) after 24 hours (b) after 48 hours.	165
Figure 6.9 Percentage caspase 3/7 expression after a 24 hour treatment in U937 cells (544:620).	167

Figure 6.10 (a) Control THP-1 cells viable (b) Control cells with DMSO (c) nuclear fragmentation (d) orange nuclei (e) nuclear fragmentation (f) blebbing (g) dumbbell (h) blebbing (i) nuclear fragmentation. 168

Figure 6.11 First lane loading dye, second lane 4-methoxycinnamyl alcohol with characteristic necrotic smear and third lane control cells without treatment. 170

List of tables

Chapter 1

- Table 1.1** The world's 25 best selling pharmaceuticals in 1991. 3
- Table 1.2** Drugs derived from natural products launched in Europe, Japan and the United States 2001-2005. 5
- Table 1.3** Summary of anticancer agents derived from natural products. 19

Chapter 2

- Table 2.1** A comparison of apoptosis with necrosis, modified from Fang (2006). 31
- Table 2.2** Cancer terminology. 42
- Table 2.3** Statistics for all cancer, males in South Africa. 52
- Table 2.4** Statistics for all cancers, females in South Africa. 54

Chapter 3

- Table 3.1** Plant samples collected for the present study: 91
- Table 3.2** Summary of the cytotoxicity results towards the cancer cell lines as well as Vero cells. 69

Chapter 4

- Table 4.1** List of naphthoquinones studied for anticancer activity

modified from (Mahapatra, *et al*, 2007). 113

Table 4.2 Summary of all the IC₅₀ results on all four the human cancer cell lines of all the derivatives of 7-methyljuglone and the positive controls. 117

Table 4.3 Cytotoxic activity of 8-Fluoro-5-hydroxy-7-methyl-1, 4-naphthoquinone, 5-Hydroxy-7-methyl-1,4-naphthoquinone (7-MJ) and 2,5-dihydroxy-7-methyl-1,4-naphthoquinone on peripheral blood mononuclear cells. 121

Chapter 5

Table 5.1 The IC₅₀ values of various fractions from column on the HeLa cell line. 134

Table 5.2 The IC₅₀ values of the compounds tested on the selected human cancer cell lines. 135

Chapter 6

Table 6.1 Optical properties for the fluorescent probes. 145

Table 6.2 Results of cell cycle analysis using nuclear PI staining. U937 cells were exposed to the compounds at the indicated concentrations for 24 hours and 48 hours before cell cycle analysis was performed. 160

Table 6.3 Results of cell cycle analysis using nuclear PI staining. U937 cells were exposed to 4-methoxycinnamyl alcohol at the indicated concentration (10 µg/m) for 48 hours before cell cycle analysis was performed. 161

Table 6.4 Results of cell cycle analysis using nuclear PI staining.

MCF-7 Cells were exposed to the compounds at the indicated concentrations for 24 and 48 hours before cell cycle analysis was performed.	162
Table 6.5 Summary of U937 annexin V-FITC/PI staining after 24 and 48 hours.	164
Table 6.6 Annexin V-FITC/PI results for MCF-7 cells at 24 and 48 hours.	166
Table 6.7 Summary of caspase 3 and 7 results.	177
Table 6.8 Summary of the morphological changes in U937 and THP-1 cells due to exposure to the naphthoquinone derivatives as determined by fluorescence microscopy of nuclear stains, acridine orange and ethidium bromide.	169

List of abbreviations

Acquired Immunodeficiency Syndrome	AIDS
Adenosine diphosphate	ADP
Adenosine triphosphate	ATP
Afrikaans	Afr.
Basal cell carcinoma	BCC
Base pairs	bp
Before Christ	B.C.
Benign prostatic hypertrophy	BPH
Calcium	Ca ²⁺
Carbon dioxide	CO ₂
Chloroform	CHCl ₃
Chronic myelogenous leukaemia	CML
Colon carcinoma	COLO-205
Dalton's lymphoma ascetic	DLA
Death effector domains	DEDS
Deoxyribonucleic acid	DNA
Dysplastic oral keratinocyte	DOK
Ehrlich ascites carcinoma	EAC
English	Eng.
Fas ligand	FasL
Fifty percent inhibitory concentration	IC ₅₀
Gas chromatography	GC
Gas chromatography-mass spectrometry	GC-MS
Glutathione (oxidized form)	GSSG
Glutathione (reduced form)	GSH

Hematologic remission	CHR
High-performance liquid chromatography	HPLC
Homoharringtonine	HHT
Human colon carcinoma	Caco-2
Human gastric adenocarcinoma	MK-1
Human immunodeficiency virus	HIV
Human papillomavirus	HPV
Human prostate cancer	LNCaP
Human uterine carcinoma	HeLa
H.G.W.J. Schwelckerdt Herbarium	PRU
Interleukin converting enzyme	ICE
Intraperitoneal	i.p.
Kilo-base pair	kb
Kilograms	kg
Lymphocytic leukaemia	P-388
Mammary gland epithelial, pleural effusion adenocarcinoma	MCF-7
Methanol	MeOH
Murine melanoma	B16F10
Micro mol	μ M
Multi Drug Resistance	MDR
Minimal inhibitory concentration	MIC
Murine lymphocytic leukaemia	P-388
Murine melanoma	B16F10
Naphthoquinones	NQ
National Cancer Institute	NCI
Metastases	M
Nicotinamide adenine dinucleotide phosphate	NADPH

Mitochondrial permeability transition pore	MPTP
Node involvement	N
Non-small cell lung cancer	NSCLC
Oesophagus carcinoma	SNO
Peripheral blood mononuclear cells	PBMCs
Prostate epithelial carcinoma	DU-145
Prostate Specific Antigen	PSA
Squamous cell carcinoma	SCC
Sodium 3'-[1-(phenyl amino-carbonyl)-3,4-tetrazolium]-bis-[4-methoxy-6-nitro) benzene sulfonic acid hydrate	XTT
Squamous oesophageal carcinoma	WHCO3
Surveillance Epidemiology and End Results	SEER
Tuberculosis	TB
Tumour	T
Tumour necrosis factor	TNF
Tumour node metastases	TNM
TUNEL	TdT-mediated dUTP nick end labeling
United States	US
United States of America	USA
United States Department of Agriculture	USDA
Vero cells	African green monkey kidney cells
World Health Organization	WHO
Zearalenone	ZEA

Abstract

A naphthoquinone, 7-methyljuglone and some of its 5-hydroxy, 5-acetoxy-, 5-alkoxy- and 1,2,4,5-tetra-*O*-acetate derivatives were tested for their activity in four human cancer cell lines: breast adenocarcinoma, cervical epithelial carcinoma, oesophageal carcinoma and prostate epithelial carcinoma. Compound 2,5-dihydroxy-7-methyl-1,4-naphthoquinone was found to be the most effective one (exhibited a fifty percent inhibitory concentration (IC_{50}) in the range of 5.3 to 14.7 μ M), while the parent compound 7-methyljuglone was less active than several of these derivatives. The IC_{50} values of 5-hydroxy-6-methyl-1,4-naphthoquinone were found to be between 19.1 and 15.4 μ M on the four cell lines. However this compound showed toxicity on peripheral blood mononuclear cells. Six derivatives were selected for mechanistic studies. Considering the findings from cell cycle analysis, caspase 3/7 activation and annexinV-FITC dual labelling, 5-hydroxy-6-methyl-1,4-naphthoquinone was found to have antitumour effect by inducing apoptosis. Two derivatives namely, '8-fluoro-5-hydroxy-7-methyl-1,4-naphthoquinone' and '2,5-dihydroxy-7-methyl-1,4-naphthoquinone' were found to be not toxic on peripheral blood mononuclear cells suggesting their action is specific for tumour cells. Compound 2,5-dihydroxy-7-methyl-1,4-naphthoquinone was found to induce apoptosis through caspase 3/7 activation. In view of the enhanced potencies associated with these derivatives, these analogues may hold considerable therapeutic potential for the treatment of leukaemia cancers.

The ethanol extracts of seven plant species (ethnobotanically selected) were also tested for their cytotoxicity, assayed by the XTT assay, against four human cancer cell lines at concentrations ranging from 0.78 to 100 μ g/ml. Of all the ethanol extracts, *Foeniculum vulgare* was found to have the best activity on HeLa cells, which exhibited an IC_{50} value of 19.97 ± 0.048 μ g/ml. Therefore, it was selected for isolation of the bioactive principles. The extract of *Foeniculum vulgare* was fractionated using column chromatography with hexane and ethyl acetate at different ratios as eluent. Two known compounds, '4-methoxycinnamyl alcohol' and 'syringin' were isolated. The IC_{50} values of '4-methoxycinnamyl alcohol' and 'syringin' were found to be 7.82 ± 0.28 μ g/ml and 10.26 ± 0.18 μ g/ml respectively on HeLa cells. Both compounds were tested for their cytotoxicity against U937 cells and also on

peripheral blood mononuclear cells. At the concentrations of 10 and 100 $\mu\text{g/ml}$ '4-methoxycinnamyl alcohol' showed similar cell proliferation as that of the positive control 'cisplatin'. 'Syringin' however, had much lower cytotoxicity on the U937 cells than '4-methoxycinnamyl alcohol'. IC_{50} was found to be $91.14 \pm 0.63 \mu\text{g/ml}$. Both 'syringin' and '4-methoxycinnamyl alcohol' were not cytotoxic at concentrations of 1 and 10 $\mu\text{g/ml}$ on the PBMCs as compared to cisplatin. '4-Methoxycinnamyl alcohol' was selected based on its activity on the cancer cells, for further investigation with regard to its mechanism of action. On gel electrophoresis it did not show a typical ladder pattern, instead a characteristic smear resulted which indicated necrosis.

Two best derivatives of 7-methyljuglone ('8-fluoro-5-hydroxy-7-methyl-1,4-naphthoquinone' and '2,5-dihydroxy-7-methyl-1,4-naphthoquinone') and the ethanol extract of *F. vulgare* warrant further investigation to be considered for their potential as anticancer agents.