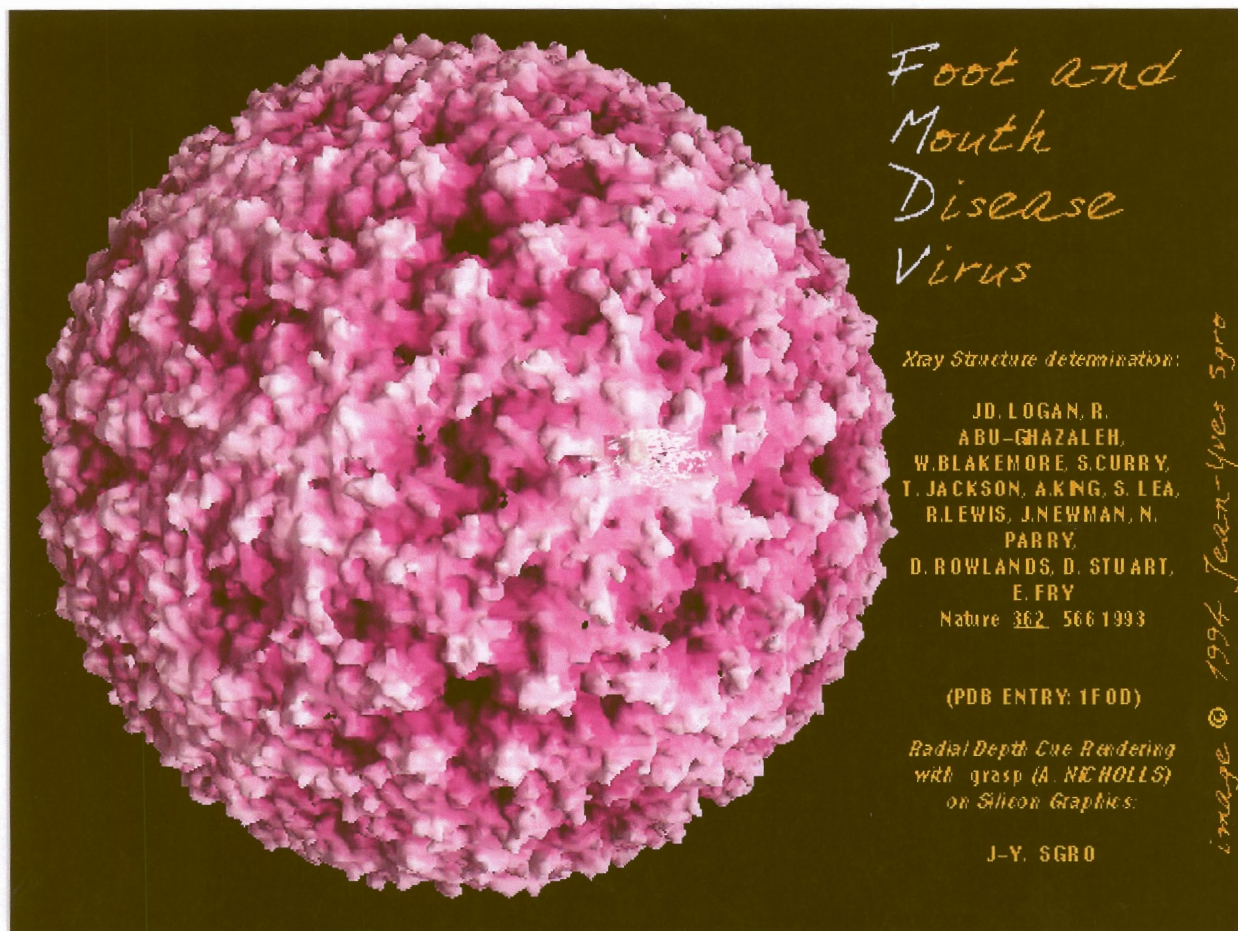


**DEVELOPMENT OF RECOMBINANT VACCINES AGAINST
FOOT-AND-MOUTH DISEASE**

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“In the temple of science are many mansions ... and various indeed are they that dwell therein and the motives that led them there.

“Many take to science out of a joyful sense of superior intellectual power; science is their own special sport to which they look for vivid experience and the satisfaction of ambition;
many others are to be found in the temple who have offered the products of their brains on this altar for purely utilitarian purposes.

Were an angel of the Lord to come and drive all the people belonging to these two categories out of the temple, it would be noticeably emptier but there would still be some men of both present and past times left inside ... If the types we have just expelled were the only types there were, the temple would never have existed any more than one can have a wood consisting of nothing but creepers ...

... and those who have found favor with the angel ... are somewhat odd, uncommunicative, solitary fellows, really less like each other than the hosts of the rejected.

“What has brought them to the temple ... no single answer will cover ... escape from everyday life, with its painful crudity and hopeless dreariness, from the fetters of one's shifting desires. A finely tempered nature longs to escape from this noisy cramped surroundings into the silence of the high mountains where the eye ranges freely through the pure still air and fondly traces out the restful contours apparently built for eternity.”

This is an excerpt from a speech given in 1918 by a young German scientist called Albert Einstein.

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ABBREVIATIONS

BEA – bromoethylamine hydrobromide
BGH – bovine growth hormone
BHK – baby hamster kidney
CMV – cytomegalovirus
CTE – C-terminal extention
DEAED – diethylaminoethyl dextran
EIF – eukaryotic initiation factor
ELISA – enzyme linked immunosorbent assay
FLC – full-length clone
FMDV – foot-and-mouth disease virus
IB-RS-2 – Instituto Biologico Rim Suino
IRES – internal ribosome entry site
KNP – Kruger National Park
MEGA – molecular evolutionary genetics analysis
MOI – multiplicity of infection
NCR – non coding region
NTP – nucleotide phosphate
OIE – Office des Epizooties
ORF – open reading frame
PBS – phosphate buffered saline
PCR – polymerase chain reaction
PK – pig kidney
PTB – polypyrimidine tract-binding
RPMI – Roswell Park Memorial Institute
RT-PCR – reverse-transcribed polymerase chain reaction
SAT – South African Territories
TCID – tissue culture infectious dose
UTR – untranslated region
VNT – virus neutralization test



VP – viral protein

VPg – viral genome-linked protein

SUMMARY

The South African Territories (SAT) types of foot-and-mouth disease virus (FMDV) show marked genomic and antigenic variation throughout sub-Saharan Africa. This variation is to a large extent geographically linked and requires therefore the use of custom-made vaccines. Adaptation of field isolates as vaccine strains is cumbersome, time consuming and expensive. A possible means of circumventing the adaptation process is to construct recombinant or chimeric FMD viruses, followed by the production of conventional, inactivated vaccine utilizing these viruses. The advantage of such a strategy would be the ability to manipulate the antigenicity of these viruses by substituting the antigenic coding regions (i.e. structural proteins) of a full-length cDNA clone of a suitable strain.

Towards this objective the structural-protein-coding region (P1) of a SAT 2 vaccine strain, ZIM/7/83/2, was determined and compared with two other known SAT 2 P1 regions. Five hypervariable regions were identified of which four are situated in VP1. The cleavage sites for proteolytic processing and especially the regions adjacent to these sites, differ between types A and SAT 2. The genetic heterogeneity of two FMDV proteinases, the Leader and 3C, of representatives of six different serotypes, was subsequently investigated. The results revealed these genomic regions of the SAT viruses originating from southern Africa to be distinct from types A, O and C. Interestingly, it was also seen that the Leader and 3C proteinases of the SAT types are less variable than their European counterparts. These results were in contrast to that obtained for the structural proteins, which showed the SAT 2 P1 region to be at least 2-3 times more variable than that of types A, O and C. Despite the observed differences in the proteinases, a three-dimensional structural model for the Lb form of the ZIM/7/83/2 Leader proteinase predicted the three-dimensional fold of the enzyme to be conserved.

A chimeric cDNA clone between types A and SAT 2 was constructed by inserting the external capsid-coding region of ZIM/7/83/2 into the genetic backbone of the A₁₂

cDNA clone. The subsequent evaluation of the resulting recombinant FMD virus indicated the virus to be immunogenically identical to the wild type ZIM/7/83/2. However, the recombinant virus was found to be a slower antigen producer and less stable than the wild type SAT 2. These characteristics make the recombinant FMD virus constructed in this study unsuitable for conventional vaccine production. Alternative means, such as the use of a SAT 2 cDNA clone, should be investigated.