



UNIVERSITEIT VAN PRETORIA
UNIVERSITY OF PRETORIA
YUNIBESITHI YA PRETORIA

**Improved molecular diagnostics and characterization of
Theileria parva isolates from cattle and buffalo
in South Africa**

By

Kgomotso Penelope Sibeko

A thesis submitted to the Faculty of Veterinary Science, University of Pretoria, South Africa,
in fulfillment of the requirements for the degree

Philosophiae Doctor

December 2009



DEDICATION

I dedicate this thesis to the memory of my late grandmother,

Mrs Ethel Magaret Sibeko,

who has been a source of inspiration and a pillar of strength,
without whose unlimited support I would not have realized this dream;

I will be forever grateful for everything she has been to me.



DECLARATION

I hereby declare that this thesis is my own work. It is submitted in fulfillment of the degree, **Philosophiae Doctor**, in the University of Pretoria, South Africa. It has not been submitted before for any degree or examination in any other University, nor has it been prepared under the aegis or with the assistance of any other body or organization or person outside the University of Pretoria, South Africa, other than as indicated in the acknowledgements which follow.

Kgomotso P. Sibeko



ACKNOWLEDGEMENTS

I would like to extend my sincere appreciation to the following individuals, organizations and institutions:

The Lord, for the gift of life, wisdom, strength and success.

Dr Nicola Collins [University of Pretoria (UP), Department of Veterinary Tropical Diseases (DVTD)], my supervisor, for her mentorship, expert advice, support and superb supervision throughout my studies.

Dr Marinda Oosthuizen (UP), my co-supervisor, for her expert advice and support during my bereavement.

Dr Dirk Geysen (Institute of Tropical Medicine (ITM), Belgium), my co-supervisor, for the opportunity to receive training in his laboratory at the ITM and continuous support throughout my studies.

Dr Abdullah Latif (ARC-OVI), my co-supervisor, for his valuable input during the preparation of this thesis.

Prof Koos Coetzer, the head of DVTD (UP), for providing funds for this work and an opportunity to undertake my studies in the DVTD.

Dr Fred Potgieter [ARC-Onderstepoort Veterinary Institute (ARC-OVI)], for his expert advice and valuable insight and input on the project.

Prof Conrad Mathee (University of Stellenbosch), for his assistance with phylogenetic analyses.

Dr Visva Pillay (Roche Diagnostics, South Africa), for providing training on the reverse line blot assay.

Mrs Milana Troskie (DVTD-UP), Mr Jacob (Ko) De Witte (ITM) and Ms Lieve Vermeiren (ITM), for technical support.

The BioPAD BRIC and the South Africa/Belgium Bilateral Agreement grants, for funding my tuition and research project.

The ITM and the ARC-OVI, for making their facilities available for this project.

The Statistics Department (UP), for assistance with the real-time PCR data analysis.

Tshepo Matjila, Raksha Bhoora and Darshana Morar, for their encouragement, support and great friendship.

The staff and postgraduate students of the DVTD, for making my PhD journey pleasant through their support both academically and socially.

Pastor Dr Paul Mbedzi, my mentor, for his encouragement and support, and teaching me that perseverance is the mother of success.

Otsile Sibeko, my beloved son, for his love, understanding and patience throughout my study period.

The late Mrs Ethel Sibeko and Mrs Mirriam Tilodi, my grandmothers, for being a pillar of strength and believing in me.

My family, for their encouragement, immeasurable support and taking pride in my achievements.



TABLE OF CONTENTS

DEDICATION	ii
DECLARATION	iii
ACKNOWLEDGEMENTS	iv
TABLE OF CONTENTS	vi
LIST OF FIGURES.....	xii
LIST OF TABLES.....	xvi
THESIS SUMMARY	xvii
CHAPTER 1 General Introduction.....	1
1.1 Background	1
1.2 Thesis rationale	2
1.3 Thesis objectives	3
1.4 References.....	4
CHAPTER 2 Literature Review	5
2.1 Introduction	6
2.2 The parasite: <i>Theileria parva</i>	6
2.2.1 The life cycle of <i>T. parva</i>	7
2.2.1.1 In the mammalian host	7
2.2.1.2 In the vector tick	7
2.3 Disease syndromes caused by <i>T. parva</i> infections	9
2.3.1 East Coast fever (ECF)	9
2.3.2 Corridor disease.....	9

2.3.3	January disease (Zimbabwean theileriosis)	10
2.4	Epidemiology of theileriosis in southern Africa.....	11
2.4.1	Introduction and eradication of cattle theileriosis, East Coast fever, in southern Africa	11
2.4.2	Emergence of other theilerial disease syndromes.....	11
2.4.3	Transformation of buffalo-derived <i>T. parva</i> into cattle-derived <i>T. parva</i> ...	12
2.4.4	Carrier state of <i>T. parva</i>	12
2.5	Treatment and control of theileriosis.....	13
2.5.1	Tick control	13
2.5.2	Chemotherapy.....	14
2.5.3	Immunization.....	15
2.6	Detection of <i>T. parva</i> infections	16
2.6.1	Conventional methods	16
2.6.2	Serological methods	17
2.6.2.1	Indirect immunofluorescent antibody test	17
2.6.2.2	Enzyme-linked immunosorbent assay (ELISA)	17
2.6.3	Molecular techniques.....	18
2.6.3.1	Conventional PCR assays.....	18
2.6.3.2	PCR-based hybridization assays.....	18
2.6.3.3	PCR-based RFLP assays	19
2.6.3.4	Real-time PCR assays.....	19
2.7	Characterization of <i>T. parva</i> stocks.....	20
2.7.1	Monoclonal antibody screening assays	21
2.7.2	Molecular characterization	22
2.8	Aim.....	23
2.9	Thesis overview	23
2.10	References.....	25



CHAPTER 3	Development and evaluation of a real-time PCR test for detection of <i>Theileria parva</i> infections in Cape buffalo (<i>Syncerus caffer</i>) and cattle	38
3.1	Abstract	39
3.2	Introduction	39
3.3	Materials and methods	42
3.3.1	Sample collection	42
3.3.2	DNA extraction	43
3.3.3	Design of primers and hybridization probes	43
3.3.4	Optimized real-time PCR conditions	44
3.3.5	Specificity of the real-time PCR assay	47
3.3.6	Sensitivity of the real-time PCR assay	49
3.3.7	Comparison of the real-time PCR assay with other molecular tests	49
3.3.8	Proficiency testing	50
3.4	Results	51
3.4.1	Specific detection of <i>T. parva</i> using the real-time PCR assay with hybridization probes	51
3.4.2	Analytical sensitivity	56
3.4.3	Comparison of molecular tests	57
3.4.4	Proficiency testing	59
3.5	Discussion	59
3.6	Summary	63
3.7	References	64
CHAPTER 4	Four p67 alleles identified in South African <i>Theileria parva</i> field samples	69
4.1	Abstract	70
4.2	Introduction	71
4.3	Materials and methods	72

4.3.1	Sample collection	72
4.3.2	DNA isolation.....	73
4.3.3	PCR amplification of the p67 gene from <i>T. parva</i>	73
4.3.4	Cloning and sequencing of p67 amplicons.....	75
4.3.5	Sequence analysis	76
4.3.5.1	Sequence editing.....	76
4.3.5.2	Sequence alignment.....	76
4.3.6	Phylogenetic analysis	77
4.4	Results.....	78
4.4.1	Amplicon analysis by agarose gel electrophoresis	78
4.4.2	Sequence analysis	79
4.4.3	Phylogenetic analysis	82
4.5	Discussion	85
4.6	Conclusion	89
4.7	References.....	90
CHAPTER 5	Characterization of <i>Theileria parva</i> parasites occurring in buffalo (<i>Syncerus caffer</i>) in South Africa: In search of cattle-type p104 alleles	95
5.1	Abstract	96
5.2	Introduction	97
5.3	Materials and methods.....	98
5.3.1	Sample collection	98
5.3.2	DNA isolation.....	98
5.3.3	Analysis of the p104 gene from <i>T. parva</i> samples using PCR-RFLP	99
5.3.4	PCR-RFLP profile analysis	101
5.3.5	Cloning and sequencing of p104 PCR products.....	101
5.3.6	Sequence analysis	101
5.4	Results.....	102

5.4.1	p104 PCR-RFLP profile analysis	102
5.4.2	p104 gene sequence analysis	107
5.5	Discussion	110
5.6	Summary	113
5.7	References.....	115
CHAPTER 6	Analysis of the gene encoding the <i>Theileria parva</i> polymorphic immunodominant molecule (PIM) reveals evidence of the presence of cattle-type alleles in South Africa.....	119
6.1	Abstract	120
6.2	Introduction	120
6.3	Materials and methods.....	122
6.3.1	Sample collection	122
6.3.2	DNA isolation and selection of <i>T. parva</i> -positive samples.....	122
6.3.3	Amplification of the PIM gene from <i>T. parva</i> samples.....	124
6.3.4	Analysis of the PIM gene from <i>T. parva</i> samples using PCR-RFLP	124
6.3.5	Cloning and sequencing of PIM PCR products.....	124
6.3.6	Sequence analysis	125
6.4	Results.....	125
6.4.1	PIM PCR-RFLP profile analysis	125
6.4.2	PIM gene sequence analysis	137
6.5	Discussion	145
6.6	Summary	149
6.7	References.....	150
CHAPTER 7	General Discussion and Conclusion.....	154
7.1	Improvement of molecular diagnosis of <i>T. parva</i> infections.....	155
7.2	Molecular characterization of South African <i>T. parva</i> parasites	157



7.2.1	Evidence of cattle-type p67, p104 and PIM alleles in <i>T. parva</i> parasite populations in South Africa.....	157
7.2.1.1	Identification of cattle-type alleles from cattle <i>T. parva</i> samples	157
7.2.1.2	Identification of cattle-type alleles from buffalo <i>T. parva</i> samples	158
7.2.2	Extensive genetic diversity among South African <i>T. parva</i> parasites	159
7.3	Conclusion	161
7.4	References.....	163
	LIST OF PUBLICATIONS	165

LIST OF FIGURES

- Figure 2.1** The life cycle of *Theileria parva* [from International Laboratory for Research on Animal Diseases (ILRAD) 1983. Annual report 1982. Nairobi, ILRAD].....8
- Figure 3.1** CLUSTAL X (1.81) multiple sequence alignment of the V4 variable region of published *Theileria* 18S rRNA gene sequences. Accession numbers for each sequence are provided. Amplification primers are highlighted in yellow. *T. parva*-specific hybridization probes are coloured in red, and hybridization probes designed to detect the presence of any *Theileria* species are in blue. Differences between the *T. parva* sequence and the four most closely related species (*T. annulata*, *T. lestoquardi*, *Theileria* sp. (buffalo), and *T. taurotragi*) are highlighted in cyan. This figure continues on page 49.....45
- Figure 3.2** Detection of *T. parva* positive control DNA using the real-time PCR assay with *Theileria* genus-specific forward and reverse primers together with *T. parva*-specific hybridization probes. (a): Melting curve analysis showing *T. parva*-specific melting peak at $63 \pm 0.62^\circ\text{C}$ and no fluorescence in the negative controls. (b): Discrimination between *T. parva*, *T. annulata* and *T. taurotragi* using melting curve analysis, with the melting peak for *T. parva* at $63 \pm 0.62^\circ\text{C}$, for *T. annulata* at $48 \pm 0.09^\circ\text{C}$ and for *T. taurotragi* at $45 \pm 0.19^\circ\text{C}$53
- Figure 3.3** Specific detection of *T. parva* DNA. Amplicons were generated with the *T. parva*-specific primer together with the *Theileria* genus-specific reverse primer and detected with the *T. parva*-specific hybridization probe set. The *T. parva*-specific melting peak at $63 \pm 0.62^\circ\text{C}$ was only observed in the *T. parva* positive control DNA samples, with no indication of amplification from any of the other *Theileria* species tested.....54

- Figure 3.4** Discrimination between *T. parva* (blue lines), and *Theileria* sp. (buffalo) (green lines) using melting curve analysis. Amplicons were generated using the *T. parva*-specific forward primer and the *Theileria* genus-specific reverse primer, and detected with the *T. parva*-specific hybridization probe set. Amplification curves showing increase in fluorescence at (a) 640 and (b) 705 nm in *T. parva* and *Theileria* sp. (buffalo) DNA samples. (c): Melting curve analysis at 640 nm, showing the *T. parva*-specific melting peak at $63 \pm 0.62^\circ\text{C}$ only for *T. parva* DNA.55
- Figure 3.5** The sensitivity and 95% confidence intervals for a 10-fold dilution series from 10^0 (3870 parasites/reaction) to 10^{-6} (0.00387 parasites/reaction) prepared from blood from a naturally infected buffalo (KNP102) with a parasitaemia of 0.009%.56
- Figure 3.6** (a): Comparison of the sensitivity of the real-time PCR assay to that of other molecular assays in detecting *T. parva* from 309 field samples. One microlitre (~15 ng) of input DNA was used in the real-time PCR assay (LC), 2 μl (~30 ng) in the conventional PCR and probing test (PP), 2.5 μl (~37.5 ng) in the RLB, and 5 μl (~75 ng) in the *cox III* assay. (b): Improved ability of the real-time PCR assay to detect *T. parva* in 105 field samples when the input DNA was increased from 1 μl (~15 ng) to 2.5 μl (~ 37.5 ng)58
- Figure 3.7** Comparison of results obtained from the DVTD and OVI laboratories when the real-time PCR assay was used to detect *T. parva*.59
- Figure 4.1** Amplicon profiles obtained from amplification of the central region of the p67 gene from buffalo-derived *T. parva* isolates collected from different geographical areas in South Africa. Lanes: M=1 kb plus DNA marker (Fermentas Life Sciences), 1= Wel 23/04, 2= Mab 43, 3=Mar 1, 4= Itha 3, 5= Itha 8, 6= HIP 5, 7 = KNP W8, 8= KNP 102, 9 = negative control. See Table 4.1 for geographical origin of isolates.79
- Figure 4.2** Alignment of the inferred amino acid acid sequences of a portion of the ~600 bp variable region of the p67 gene amplified from representative *T. parva* strains. The alignment was generated using the multiple sequence alignment program Mafft version 6 (Katoh *et al.*, 2002) (<http://align.bmr.kyushu-u.ac.jp/mafft/software/>).81

Figure 4.3 Phylogenetic relationship of *T. parva* strains as revealed by p67 gene sequences. The phylogenetic tree was calculated by maximum parsimony analysis using TBR swapping in PAUP*4.0b10 (Swofford, 2003) and the tree where unique insertions are weighted 10:1 is shown. Nodal support was assessed with 1000 bootstrap replicates and indicated above for 1:1; 1:3; 1:10 weighting of indels while values below represent bootstrap support for the nucleotide analyses only (see text for details).84

Figure 5.1 p104 gene *AluI* RFLP profiles showing the *T. parva* Muguga RFLP profile and identical profiles obtained from cattle samples from a farm near Ladysmith. 103

Figure 5.2 A simplified similarity dendrogram for p104 PCR-RFLP profiles cluster analysis based on the Dice Coefficient analysis produced using BioNumerics v5.1. 104

Figure 5.3 A detailed similarity dendrogram for p104 PCR-RFLP profiles cluster analysis based on the Dice Coefficient analysis produced using BioNumerics v5.1 showing actual profiles used to create the dendrogram. Figure 5.3 continues on page 110. 105

Figure 5.4 Phylogenetic relationship of *T. parva* strains as revealed by p104 amino acid sequence analysis. The phylogenetic tree was calculated by Bayesian analysis; the first 10,000 trees were discarded as burnin and the majority-rule consensus tree was generated. 109

Figure 6.1 Representative PIM gene *BcII* PCR-RFLP profiles showing (a) heterogeneous profiles obtained from buffalo *T. parva* samples from KNP, (b), (c) and (d) homogeneous profiles obtained from buffalo samples from Hluhluwe-iMfolozi, Mabalingwe and Ithala, respectively. 127

Figure 6.2 PIM gene *BcII* PCR-RFLP profiles obtained from (a) cattle *T. parva* samples from Ladysmith and (b) clones produced from cattle sample, Lad 10. 128

Figure 6.3 A simplified similarity dendrogram generated by BioNumerics v5.1 cluster analysis of PCR-RFLP profiles from cloned PIM amplicons using the Dice Coefficient analysis. 129

- Figure 6.4** A detailed similarity dendrogram generated by BioNumerics v5.1 cluster analysis of PCR-RFLP profiles from cloned PIM amplicons using the Dice Coefficient analysis showing actual profiles used to produce the dendrogram. Figure 6.4 continues on pages 136 to 139..... 130
- Figure 6.5** Characteristic ‘signatures’, indicated by brackets (I), were identified in *T. parva* PIM RFLP profiles obtained from cloned PCR products from samples collected from buffalo from Kruger National Park and Hluhluwe-iMfolozi Game Park. (a): PIM RFLP profiles with specific band ‘signatures’ obtained from clones from sample KNP W8 from Kruger National Park, characterized by five (clones 1, 2, 3, 6, 7 and 11) and six (clones 13 and 14) small DNA fragments of sizes ranging from ~ 50 to 130 bp. (b): PIM RFLP profiles obtained from clones from sample HIP 5 from Hluhluwe-iMfolozi with the ‘four band signature’ consisting of ~ 50, 80, 100 and 120 bp DNA fragments..... 136
- Figure 6.6** Multiple sequence alignment of the inferred PIM amino acid sequences obtained from buffalo and cattle *T. parva* samples collected from different geographical areas in South Africa (Table 6.1). PIM sequences were aligned with previously published *T. parva* PIM sequences, Muguga (accession number: L06323), Marikebuni (accession number: L41148) and 7104 (accession number: L41833). The sequence alignment was constructed manually because of the extreme polymorphism in the central region of the PIM gene. Amino acid motifs characteristic of cattle-type PIM sequence are shown in solid-line blocks including the tetrapeptide repeat characteristic of the central variable region of cattle-derived PIM sequences at positions 428-447. All amino acid motifs characteristic of buffalo-type PIM sequence are shown in broken-line blocks. Figure 6.6 continues on pages 145 and 146. 139
- Figure 6.7** Multiple sequence alignment of representative amino acid sequences of ‘mixed’-type *T. parva* PIM sequences, showing subtypes M-I and M-II. Regions typical of buffalo-type and cattle-type PIM sequences are indicated in broken-line and solid-line blocks, respectively. This alignment excludes the region between positions 101 and 300, which has buffalo-type PIM sequences in all subtypes..... 144



LIST OF TABLES

Table 3.1	Origin and number of samples used for the evaluation of the <i>T. parva</i> real-time PCR test.....	43
Table 3.2	Specificity of the <i>T. parva</i> real-time PCR test using the <i>T. parva</i> -specific forward primer, the <i>Theileria</i> genus-specific reverse primer and both probe sets	48
Table 4.1	Source of samples used for characterization of <i>T. parva</i> parasites and results obtained from PCR amplification of the p67 gene. Table 4.1 continues in page 79.....	74
Table 4.2	p67 reference sequences used for analysis of data obtained in this study.....	77
Table 4.3	Number and type of p67 sequences obtained from 21 selected <i>T. parva</i> samples collected from both cattle and buffalo	82
Table 5.1	Geographic origin and source of blood samples (n=100) used for characterization of <i>T. parva</i> parasites.....	100
Table 5.2	Comparison of p104 PCR-RFLP cluster analysis and phylogenetic analysis of p104 amino acid sequences from 19 selected <i>T. parva</i> samples.....	110
Table 6.1	Geographical origin and source of blood samples (n=109) used for characterization of <i>T. parva</i> parasites.....	123
Table 6.2	Number of different types of PIM sequences obtained from clones produced from 27 selected <i>T. parva</i> samples	138

THESIS SUMMARY

The aim of this study was to improve the official diagnostic test package in South Africa for detection of *Theileria parva* infections in cattle and Cape buffalo (*Syncerus caffer*) and to investigate the presence of cattle-type *T. parva* parasites in buffalo and cattle in South Africa. To improve diagnosis of *T. parva* infections, a *T. parva*-specific real-time polymerase chain reaction (PCR) assay based on hybridization probe technology was developed. Oligonucleotide primers and hybridization probes used in the assay were designed based on the 18S ribosomal RNA (rRNA) gene. The primers amplify *T. parva* and *Theileria* sp. (buffalo) DNA but the hybridization probes specifically detect *T. parva* amplicons. Because of the high sequence similarity between the *T. parva* and *Theileria* sp. (buffalo) 18S rRNA genes, amplification of *Theileria* sp. (buffalo) DNA could not be avoided; no other bovine blood pathogens tested were amplified by these primers. The real-time PCR assay demonstrated superior sensitivity compared to other molecular tests used in detection of *T. parva* infections, reliably detecting the parasite in carrier animals with a piroplasm parasitaemia as low as $8.79 \times 10^{-4}\%$ with minute template DNA input. The assay requires less time to perform with a low risk of contamination because of the closed-tube system that does not require handling of amplicons for post-PCR analysis.

The presence of cattle-type *T. parva* parasites in buffalo and cattle was investigated using restriction fragment length polymorphism (RFLP) profiles of PCR products and sequences of the parasite genes which code for the antigenic proteins p67, p104, and the polymorphic immunodominant molecule (PIM). Cattle-type p67, p104 and PIM alleles were identified from three *T. parva* samples obtained from cattle from a farm near Ladysmith in the KwaZulu-Natal Province. These cattle-type alleles were identical to those previously identified from a cattle-derived *T. parva* stock, *T. parva* Muguga, a parasite stock that causes East Coast fever (ECF) in Kenya; however, ECF was not diagnosed in animals in this farm. Cattle-type alleles identical to those previously reported were not identified from *T. parva* buffalo samples, but variants of p67 allele 1 as well as p104 allele 1, both previously obtained from *T. parva* Muguga, were identified. It is not known if parasites that possess these variants can cause disease, and the risk of their adapting to cattle as in the case of ECF and January disease needs to be evaluated. Furthermore, these findings suggest that cattle-like alleles may not be exclusively associated with cattle-derived *T. parva* parasites. Most of the p67, p104 and PIM gene sequences obtained in this study were not identical to known sequences; furthermore, novel alleles were identified, demonstrating extensive genetic diversity in the

South African *T. parva* parasite population in buffalo. The significance of the parasites that possess ‘novel’ alleles in the epidemiology of theileriosis in South Africa still needs to be determined. The identification of variants and novel alleles reveals that p67, p104 and PIM gene PCR-RFLP profiles are more complex than previously thought and the classification of buffalo- and cattle-derived *T. parva* parasites in South Africa based on p67, p104 and PIM gene profiles would not be possible. Identification of more reliable markers that can be directly associated with the theilerial disease syndromes remains a challenge.