

APPENDIX A

SUBJECT DATA COLLECTION SHEET

Study: Low dose erythromycin in improving the outcome of HIV-infected children

DATA COLLECTION SHEET: Visit 1

1. PATIENT INFORMATION

Name

Hospital number

Study number

Date of admission

DD/MM/YY

Gender

male

female

Age

Months

Date of birth

DD/MM/YY

2. PREVIOUS MEDICAL HISTORY

a. HIV			
	Patient previously tested	Y	N
	If Yes, previous result	positive	negative
	Currently: 1. Consent for ELISA	Y	N
	2. CD4 count	%	total
	3. Viral load		
	Antiretroviral treatment	Y	N
	Specify: 1. Drugs	1.	
	2.		
	3.		
	4.		



	2. Start date ARV		
b. Previous admissions	Previously admitted to hospital	Y	N
c. Treatment	Did the patient receive antibiotics? If yes specify:	Y	N
c. Anthropometric Findings			
Length		cm	
Weight		kg	

d. Examination

General	Temperature on admission		°C
	Generalized lymphadenopathy (0.5 cm present in at least 2 sites, bilateral lymph nodes counting as one site)	Y	N
	Clinically pale	Y	N
	Oedema	Y	N
	Jaundice	Y	N
	Hepatomegaly	Y	N
	Splenomegaly	Y	N
	Oral thrush	Y	N
	Parotomegaly	Y	N
	Eczema	Y	N
	HIV encephalopathy	Y	N



	Neurodevelopmentally normal	Y	N	
Respiratory system	Respiratory rate	/ min		
	Heart rate	/ min		
	Peripheral saturation	without oxygen	%	
		with oxygen	%	
	Recession		Y N	
	intercostal	Subcostal	suprasternal	
	Flaring of alae nasi		Y N	
	Clinically cyanosed		Y N	
	Grunting		Y N	
	AUSCULTATORY FINDINGS			
	Focal abnormality		Y N	
	Diffuse abnormality		Y N	
	Clear chest		Y N	
	Hyperinflation		Y N	
	Crepitations		Y N	
	Bronchial breathing		Y N	
	Wheezing		Y N	
	Comments:			



OTHER SYSTEMS	
CVS	
GIT	
CNS	
ENT	

STUDY VISIT COMPLETION FORMS

Study: Low dose erythromycin in improving the outcome of HIV-infected children with bronchiectasis

Pt #: _____ Init: _____

Visit	CD4 count	Viral load	Sputum MCS	Cytokine assays (blood/ sputum)	Sputum resp virus/TB	Liver function test
Visit 1	X	X	X	X	X	X
Visit 2			X			
Visit 3			X			
Visit 4			X			
Visit 5			X			
Visit 6			X			
Visit 7			X			
Visit 8			X			
Visit 9			X			
Visit 10			X			
Visit 11			X			
Visit 12	X	X	X	X	X	X

STUDY VISIT COMPLETION FORMS

Study: Low dose erythromycin in improving the outcome of HIV-infected children with bronchiectasis

Pt #: _____ Init: _____

	CXR	PET CT	Lung function test	Nitric oxide	Sweat test	Clinical exam
Visit 1	X	X	X	X	X	X
Visit 2			X			X
Visit 3			X			X
Visit 4			X			X
Visit 5			X			X
Visit 6			X			X
Visit 7			X			X
Visit 8			X			X
Visit 9			X			X
Visit 10			X			X
Visit 11			X			X
Visit 12	X	X	X	X		X



APPENDIX B

ETHICAL APPROVAL

The Research Ethics Committee, Faculty Health Sciences, University of Pretoria complies with ICH-GCP guidelines and has US Federalwide Assurance. FWA 00002567, Approved dd 22 May 2002 and Expires 24 Jan 2009.
RB 0000 2235 IORG0001762 Approved dd Jan 2006 and Expires 21 Nov 2008.



Date: 3/06/2008

PROTOCOL NO.	100/2008~A
NEW TITLE	Low dose erythromycin in improving outcome of HIV-positive children with bronchiectasis
STUDY DE.G.R.EE	PhD
SPONSORS POSTAL ADDRESS	Level D3 New Steve Biko Academic Hospital, Malherbe Street, Capital park.
MEETING DATE OF THIS STUDY	28/05/2008

This Protocol and Informed Consent and all the attachments have been considered by the Faculty of Health Sciences Research Ethics Committee, University of Pretoria on 28/05/2008 and found to be acceptable.

Advocate AG Nienaber	(female) BA (Hons) (Wits); LLB; LLM (UP); Dipl.Datometrics (UNISA)
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*Dr L Schoeman	(female) BP harm, BA Hons (Psy), PhD
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DR R SOMMERS; MBChB; MMed (Int); MPhar.Med.
SECRETARIAT of the Faculty of Health Sciences Research Ethics
Committee, University of Pretoria, Pretoria Academic Hospital

* Members attending the meeting.

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The Research Ethics Committee, Faculty Health Sciences, University of Pretoria complies with ICH-GCP guidelines and has US Federal wide Assurance.

- * FWA 00002567, Approved dd 22 May 2002 and Expires 24 Jan 2009.
- * IRB 0000 2235 IORG0001762 Approved dd Jan 2006 and Expires 13 Aug 2011.

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UNIVERSITY OF PRETORIA
YUNIBESITHI YA PRETORIA

Faculty of Health Sciences Research Ethics Committee

Fakulteit van Gesondheidswetenskappe Navorsingsetiekkomitee

Date: 20/11/2008

PROTOCOL NO.	100/2008~B
Informed Consent Document	(Give reason for ■■■)
PROTOCOL TITLE	Chronic inflammatory lung disease in human immunodeficiency virus (HIV) infected children. Epidemiological considerations, aetiological determinants and the efficacy of low dose erythromycin in bronchiectasis.
INVESTIGATOR	Principal Investigator: Refiloe Masekela
SUPERVISOR	R.J Green
DEPARTMENT	Dept: Paediatrics Phone: 012 354 5271 Fax: 012 354 5275 E-Mail: Refiloe.masekela@up.ac.za Cell: 079 489 0936
MEETING DATE OF THIS STUDY	19/11/2008

This **Amendment** has been considered by the Faculty of Health Sciences Research Ethics Committee, University of Pretoria on 19/11/2008 and found to be acceptable

** Members attended & Feedback at the meeting .*

- *Dr A Nienaber (female) BA (Hons) (Wits); LLB; LLM (UP); Dipl.Datometrics (UNISA)
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- *Dr L Schoeman (female) BP harm, BA Hons (PSy), PhD
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- *Prof TJP Swart BChD, MSc (Odont), MChD (Oral Path), **PGCHE**
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SECRETARIAT of the Faculty of Health Sciences Research Ethics Committee, University of Pretoria, Pretoria Academic Hospital

APPENDIX C

PATIENT INFORMATION LEAFLET, CONSENT FORM AND ASSENT FORM

A. CONSENT FORM AND PATIENT INFORMATION SHEET

Study title: Low dose erythromycin therapy in improving outcome of HIV-positive children with bronchiectasis

Dear Parent / Guardian

Your child _____ is currently suffering from a problem of the chest and lungs that has come about because of HIV infection. This letter serves to request your permission to enrol your child in a study to investigate treatment options for children with this condition.

What is the purpose of the study?

This condition is usually treated with antibiotics when your child gets sick. We would like to test the effect of using a small quantity of a common antibiotic known as erythromycin on the disease process. Erythromycin is a medicine that reduces inflammation, and is often used in a number of other conditions and is generally tolerated well. It is possible that erythromycin may reduce the chances of your child getting sick and may have a good effect on improving his general condition and prevention infections in the lung. Your child may or may not get the antibiotic as we would like to see if will have any effect in the improvement of his/her lung status.

What is the duration of the study?

The duration of this study will be 12 months. Your child will need to give one tablet every evening for this whole time.

Procedures to be followed

Participating in this study would imply that your child would be treated with usual antibiotics in a standard way and the erythromycin as well. Your child's condition and response to the treatment will be monitored monthly at the usual clinic you attend.

Simultaneously, a small volume of the blood, urine and sputum, that are routinely collected, will also be tested for specific cytokine responses to infection.

Cytokines are specific substances released by the fighting cells of the body during stress situations like infection. Some of these cytokines are pro-inflammatory, or causing inflammation, while others are anti-inflammatory, or regulating this immune response by opposing the inflammatory response. These two groups of agents are usually working in a balanced way, and should something like specific chest infections impair this balance, damage to the cells may occur. We will be following your child up very closely at monthly intervals. We will then collect the normal sputum samples and monitoring his/her progress as well as looking for complications that the medication can cause. After 12 months we will repeat the sputum, urine and blood testing for the cytokine levels as well as the chest x-rays and CT scans. This information will guide us in better understanding of the lung damage caused by chest infections in your child.

What will be done at each visit will be as follows:

Visit 1:

- Sputum samples
- Blood tests
- Lung function tests
- Nitric oxide measurements
- Chest x-ray
- PET CT scan
- Sweat test
- Clinical examination

Visit 2-11

- Lung function test
- Sputum sample
- Clinical examination

Visit 12 (study ends)

- Lung function test
- Nitric oxide measurement
- Chest x-ray
- PET CT chest

- Sputum sample
- Blood sample
- Clinical examination

Risks and discomfort involved

It is important to note that no additional discomfort will be caused to the usual blood tests and investigations performed on a child with this condition.

We do not expect side effects from short-term erythromycin use and the risk is very small. Erythromycin used for a long time may cause some nausea, vomiting or diarrhoea. It is also possible that this antibiotic may make the other bugs in your child's lung resistant (stop responding) to some of the antibiotics we may need to use for pneumonia. In rare cases erythromycin can cause an allergic reaction and it may also cause damage to the liver which results in swelling of the liver and abdominal pains.

Drug interactions

Erythromycin can also interact with other medicines your child may be taking for example midazolam (Dormicum) by decreasing the level in the blood. Erythromycin can also increase the level of the following drugs in the blood: ebastine, carbamazepine (Tegretol), ciclosporin, ergotamine and warfarin. Should your child be taking any of these drugs the doctor will monitor the levels of these drugs closely and may not enrol your child in the study.

As mentioned previously, erythromycin is used in a number of other conditions and generally tolerated well. Should your child's condition deteriorate or an adverse (bad) reaction happen with the medication you are to contact Dr Masekela immediately at any time of day or night on the number 079 489 0936/ 012 354 5271. The medication will be stopped in case of a severe reaction to the medication; that is an allergic reaction or evidence of liver damage from the medication.

HIV testing

This letter then further serves to ask your permission to do a HIV test on your child if it was not done before. A specific consent form in the ward will also be used. It is important that the doctor who presents this form to you explain the following to you:

- The reasons we want to test your child
- That HIV is virus or bug that attacks the fighting cells of your body and make the body weak so that it can't fight infections as well as before
- How HIV is transmitted: through sexual contact, blood transfusions or dirty needles e.g. drug users or from mother to child. Transmission from mother to child can happen either during the pregnancy, the birth process or breastfeeding
- The stages of HIV in an adult and how it differs in children
- Currently there is no cure for HIV. We can however treat the infections the child gets because the body is weak. The doctor should also explain to you how anti-retroviral drugs can improve the quality of life and where they are available
- If your child's test is positive, the probability that you are also positive is high, and you should yourself be tested as well. Advice regarding future pregnancies and the availability of any treatment should also be given.

It is important to know that the results may only be given to you and that post-test counselling will also be done. You may decline the HIV test and the treatment of your child will not be influenced by that decision.

Has the study received ethical approval

The study protocol was submitted to the Faculty of Health Sciences Research Ethics Committee, University of Pretoria, and that committee has granted written approval.

The study has been structured in accordance with the Declaration of Helsinki, which deals with the recommendations guiding doctors in biomedical research involving humans.

Confidentiality

All information obtained during the course of this trial is strictly confidential, and personal information will remain confidential at all times. Data that may be reported

in scientific journals will not include any information, which identifies your child as a patient in this study.

Source of additional information

During your child’s stay in hospital she will be under the care of Dr Masekela on 012 354 5271/079 489 0936, If you have any questions, which were not fully explained by the doctor obtaining consent, please do not hesitate to ask him/her.

This letter serves to request your permission to enrol your child in this study group. You may refuse such permission and your child’s management will not be affected in any way.

INFORMED CONSENT

I, _____

the parent/guardian of

_____ (*Name of patient*)

_____ (*Hospital Number*)

Have been informed about and understand the nature, benefits, risks and purpose of the trial, and all my questions have been answered. I hereby give permission that my child may participate in the above study.

I further have been fully informed regarding HIV and all my questions have been answered. I therefore give / do not give consent to perform an HIV test on my child

Signature of the parent/guardian

Date

I hereby confirm that the above parent / guardian have been fully informed about the nature, conduct and risks of the above trial.

Name of the investigator



Signature of the investigator

Date

Name of witness

Signature of witness

Date

B. ASSENT FORM

Name of study: Low dose erythromycin in improving outcome of HIV-positive children with bronchiectasis

I understand that I have been asked to participate in a study about my chest problem (bronchiectasis). I understand that I will use erythromycin (the new medicine), in addition to my normal medicines that I take for my chest.

I will be asked to have a check up of my chest by the doctor and I will be asked to blow in the two machines to check the size and the swelling of my lungs.

I will be asked to come for visits to the doctor regularly in order that the doctor can check how my chest is doing and a photograph will be taken of my chest to see how far my chest problem is. I will have to ask questions about my illness with every visit. I will also give my mucus to the doctor to check with every visit. I will be blowing into the machine with every visit so that the doctor can check my lungs. I will also have blood taken from me at the beginning of the study and after 12 visits (one year). A new photo of my chest will also be done after one year.

I understand that the medicine that the doctor wants to give me can make me feel sick. I can vomit or my stomach can work a lot from taking the medicine. This medicine may also give me a rash or bad reaction. The doctor will check me to see if it's very bad in which case she will decide to stop it if I get too sick.

I understand that I do not have to participate. If I do participate, I can quit at any time. I also understand that I do not have to answer any questions I don't want to answer or do anything I don't want to do.

My parents, teachers or anyone else will not know what I have said or done in the study. No one but the researchers will know.

This study is being done by Dr Refiloe Masekela of Pretoria Academic hospital. Her phone number is 012-354 5271 or 079 4890936.

If I have any questions or concerns about the study, I can call and ask her about them. When I sign my name, this means that I agree to participate in the study and



that all of my questions have been answered. I have also been given a copy of this form.

Name: _____ Signature _____

Name of Witness _____

Witness signature _____ Date _____

APPENDIX D

BHALLA SCORE

Category	0	1	2	3
Severity of bronchiectasis	Absent	Mild (luminal diameter slightly greater than accompanying vessel)	Moderate (lumen 2-3 times the diameter of vessel)	Severe (lumen >3 times diameter of vessel)
Peribronchial thickening	Absent	Mild (wall thickness equal to diameter of adjacent vessel)	Moderate (wall thickness greater than and up to twice the diameter of adjacent vessel)	Severe (wall thickness >2 times the diameter of adjacent vessel)
Extent of bronchiectasis*	Absent	1-5	6-9	>9
Extent of mucous plugging*	Absent	1-5	6-9	>9
Sacculations or abscesses*	Absent	1-5	6-9	>9
Generations of bronchial divisions involved (bronchiectasis/plugging)	Absent	Up to 4 th generation	Up to the 5 th generation	Up to 6 th generation and distal
No of bullae	Absent	Unilateral (not >4)	Bilateral (not >4)	>4
Emphysema*	Absent	1-5	>5	
Collapse/consolidation	Absent	Subsegmental	Segmental/lobar	

No of bronchopulmonary segments affected: for the calculation of the CT score is subtracted from 25 [108]

REFERENCES

1. UNAIDS report on the global AIDS epidemic 2010.
https://www.unaids.org/globalreport/Global_report.htm. Accessed 17/04/2012.
2. Van Rie A, Beyers N, Gie RP, et al. Childhood tuberculosis in an urban population in South Africa: burden and risk factors. *Arch Dis Child* 1999;80:433-437.
3. Lazarus JV, Olsen M, Ditiu L, et al. Tuberculosis-HIV co-infection: policy and epidemiology in 25 countries in WHO European region. *HIV Med* 2008;9:406-414.
4. Callahan CW, Redding GJ. Bronchiectasis in children. Orphan disease or persistent problem? *Pediatr Pulmonol* 2002;33:492-496.
5. Keistinen T, Säynäjäkangas O, Tuuponen T, et al. Bronchiectasis: an orphan disease with a poorly-understood prognosis. *Eur Respir J* 1997;10:2784-2787.
6. Kapur N, Karadag B. Differences and similarities in non-cystic fibrosis bronchiectasis between developing and affluent countries. *Paediatr Respir Rev* 2011;12:91-96.
7. Twiss J, Metcalfe R, Edwards E, et al. New Zealand national incidence of bronchiectasis “too high” for a developed country. *Arch Dis Child* 2005;90:737-740.
8. Chang AB, Grimwood K, Mulholland EK, et al. Bronchiectasis in indigenous children in remote Australian communities. *Med J Aust* 2002;177:200-204.
9. Karadag B, Karakoc F, Ersu R, et al. Non-cystic fibrosis bronchiectasis in children: a persisting problem in developing countries. *Respiration* 2005;72:233-238.
10. Bouyahia O, Essadem L, Matoussi N, et al. Etiology and outcome of bronchiectasis in children: a study of 41 patients. *Tunis Med* 2008;86:996-999.
11. Singleton R, Morris A, Redding G, et al. Bronchiectasis in Alaska Native children: causes and clinical courses. *Pediatr Pulmonol* 2000;29:182-189.
12. Kim HY, Kwon JW, Seo J, et al. Bronchiectasis in children: 10-year experience at a single institution. *Allergy Asthma Immunol Res* 2011;3:39-45.
13. Li AM, Sonnappa S, Lex C, et al. Non-CF bronchiectasis: does knowing the aetiology lead to changes in management? *Eur Respir J* 2005;26:8-14.

14. O'Donnell AE, Barker AF, Olowite JS, et al. Treatment of idiopathic bronchiectasis with aerosolized recombinant human DNase I. rhDNase Study Group. *Chest* 1998;113:1329-1334.
15. Berman DM, Mafut D, Kajokic B, et al. Risk factors for the development of bronchiectasis in HIV-infected children. *Pediatr Pulmonol* 2007;42:871-875.
16. Sheikh S, Madiraju K, Steiner P, et al. Bronchiectasis in pediatric AIDS. *Chest* 1997;112:1202-1207.
17. Holmes A, Trotman-Dickenson B, Edwards A, et al. Bronchiectasis in HIV disease. *QMJ* 1992;85:875-882.
18. Jeena PM, Coovadia HM, Thula SA, et al. Persistent and chronic lung disease in HIV-infected and un-infected African children. *AIDS* 1998;12:1183-1193.
19. Zar HJ. Chronic lung disease in human immunodeficiency virus (HIV) infected children. *Pediatr Pulmonol* 2008;43:1-10.
20. Equi A, Balfour-Lynn IM, Bush A, et al. Long term azithromycin in children with cystic fibrosis: a randomised, placebo-controlled crossover trial. *Lancet* 2002;360:978-984.
21. McCormack J. Effect of long term treatment with azithromycin on disease parameters in cystic fibrosis. *Thorax* 2002;57:212-216.
22. Saiman L, Marshall BC, Mayer-Hamblett N, et al. Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA* 2003;290:1749-1756.
23. Clement A, Tamalet A, Leroux E, et al. Long term effects of azithromycin in patients with cystic fibrosis. *Thorax* 2006;61:895-902.
24. Anwar GA, Bourke SC, Afolabi G, et al. Effects of long-term low-dose azithromycin in patients with non-CF bronchiectasis. *Respir Med* 2008;102:1494-1496.
25. Khair OA, Devalia JL, Abdelaziz MM, et al. Effect of erythromycin on *Haemophilus influenzae* endotoxin-induced release of IL-6, IL-8, and sICAM-1 in cultured human bronchial epithelial cells. *Eur Respir J* 1995;8:1451-1457.
26. Gorrini M, Lupi A, Viglio S, et al. Inhibition of human neutrophil elastase by erythromycin and flurythromycin, two macrolide antibiotics. *Am J Respir Cell Mol Biol* 2001;25:492-499.

27. Takizama H, Desaki M, Ohtoshi T, et al. Erythromycin modulates IL-8 expression in normal and inflamed bronchial epithelial cells. *Am J Respir Crit Care Med* 1997;156:266-271
28. Yalçın E, Kiper N, Özçelik U, et al. Effects of clarithromycin on inflammatory parameters and clinical conditions in children with bronchiectasis. *J Clin Pharm Ther* 2006;31:49-55.
29. Serisier DJ, Martin ML. Long-term, low-dose erythromycin in bronchiectasis subjects with frequent infective exacerbations. *Respir Med* 2011;105:946-949.
30. Tsang KW, Ho PI, Chan KN, et al. A pilot study of low-dose erythromycin in bronchiectasis. *Eur Respir J* 1999;13:361-364.
31. Koh YY, Lee MH, Sun YH, et al. Effect of roxithromycin on airway responsiveness in children with bronchiectasis: a double-blind, placebo-controlled study. *Eur Respir J* 1997;10:994-999.
32. Palardini M, Frank I, Pandrea I, et al. Mucosal immune dysfunction in AIDS pathogenesis. *AIDS Rev* 2008;10:36-46.
33. Haase AT. Population biology of HIV-1 infection: viral and CD4⁺ T cell demographics and dynamics in lymphatic tissues. *Annu Rev Immunol* 1999;17:625-656.
34. Beck JM, Rosen MJ, Peavy HH. Pulmonary complications of HIV infection. Report of the fourth NHLBI workshop. *Am J Respir Crit Care Med* 2001;164:2120-2126.
35. Kanki P, Travers K, Hernandez-Avilla M, et al. Slower heterosexual spread of HIV-2 compared with HIV-1. *Lancet* 1994;343:943-946.
36. Marlink R, Kanki P, Thior I, et al. Reduced rate of disease development with HIV-2 compared to HIV-1. *Science* 1994;265:1587-1590.
37. Taylor BS, Sobieszczyk ME, McCutchan FE, et al. The challenge of HIV-1 subtype diversity. *N Engl J Med* 2008;358:1590-1602.
38. Kanki PJ, Hamel DJ, Sankale J-L, et al. Human immunodeficiency virus type-1 subtypes differences in disease progression. *J infect Dis* 1999;179:68-73.
39. Peeters M. The genetic variability of HIV-1 and its implications. *Transfus Clin Biol* 2001;8:222-225.
40. Hu DJ, Dondero TJ, Rayfield MA, et al. The emerging genetic diversity of HIV. *JAMA* 1996;275:210-216.

41. Chalmet K, Staelens D, Blot S, et al. Epidemiological study of phylogenetic transmission clusters in a local HIV-1 epidemic reveals distinct differences between subtype B and non-B infections. *BMC Infect Dis* 2010;10:262.
42. Renjifo B, Gilbert P, Chaplin B, et al. Preferential in-utero transmission of HIV-1 subtype C as compared to HIV-1 subtype A or D. *AIDS* 2004;18:1629-36.
43. Renjifo B, Fawzi W, Mwakagile D, et al. Differences in perinatal transmission among human immunodeficiency virus type 1 genotypes. *J Hum Virol* 2001;4:16-25.
44. Lindegren ML, Steinberg S, Byers RH. Epidemiology of HIV/AIDS in children. *Pediatr Clin North Am* 2000;47:1-20.
45. Klugman KP. Emerging infectious diseases-South Africa. *Emerg Infect Dis* 1998;4:517-520.
46. Mortality and causes of death in South Africa 2005: findings from death notification. <http://www.statssa.gov.za>. Accessed 27/04/2008.
47. South Africa HIV and AIDS statistics. <http://www.avert.org/safricastats/>. Accessed 15/08/2010.
48. Centers for Disease Control and Prevention: *HIV/AIDS Surveillance Report* 1999 11:1-24.
49. Luziriaga K, Sullivan JL. Viral and immunopathogenesis of vertical HIV-1 transmission. *Pediatr Clin North Am* 2000;47:65-78.
50. Gray L, Newell ML, Thorne C, et al. Fluctuations in symptoms of human immunodeficiency virus-infected children: The first 10 years of life. *Pediatrics* 2001;108:116-122.
51. Connor EM, Sperling RS, Gelber R, et al. Reduction of maternal-infant transmission of human immunodeficiency virus type 1 with zidovudine treatment. *N Engl J Med* 1994;331:1173-1180.
52. Centre for Disease Control and Prevention. Epidemiology of HIV/AIDS-United States 1981-2005. *MMWR* 2006;55:589-592.
53. Dorenbaum A, Cunningham CK, Gleber RD, et al. Two-dose intrapartum/newborn nevirapine and standard anti-retroviral therapy to reduce perinatal HIV-1 transmission: a randomized trial. *JAMA* 2002;288:189-198.
54. Lallemand M, Jourdan G, Le Coeur S, et al. Single-dose perinatal nevirapine plus standard zidovudine to prevent mother-to-child transmission of HIV-1 in Thailand. *N Engl J Med* 2004;351:217-228.

55. Mandelbrot L, Landreau-Mascaro A, Rekacewicz C, et al. Lamivudine-zidovudine combination for prevention of maternal infant transmission of HIV-1. *JAMA* 2001;283:2083-2093.
56. The European Mode of Delivery Collaboration. Elective caesarean section versus vaginal delivery in prevention of vertical HIV-1 transmission: a randomised clinical trial. *Lancet* 1999;353:1035-1039.
57. Centre for Disease Control and Prevention. Success in implementing PHS guidelines to reduce perinatal transmission of HIV-1993, 1995 and 1996. *MMWR* 1998;47:68-91.
58. Ebrahim S, Daponte A, Guidozi F. The impact of free antenatal care on perinatal mortality. *Int J Gynaecol Obstetr* 2000;71:205-207.
59. Myer L, Harrisson A. Why do women seek antenatal care late? Perspectives from rural South Africa. *Br Med J* 2003;48:268-272.
60. Berg CJ. Prenatal care in developing countries: The World Health Organization Technical Working Group on Antenatal Care. *J Am Med Womens Assoc* 1995;50:182-186.
61. Read JS. The mode of delivery and the risk of vertical transmission of human immunodeficiency virus type-1. *N Engl J Med* 1999;340:977-987.
62. Wilfert CM, Fowler MG. Balancing maternal and infant benefits and the consequences of breast-feeding in the developing world during the era of HIV infection. *J Infect Dis* 2007;195:165-167.
63. Dunn DT, Newell ML, Ades AE, et al. Risk of human immunodeficiency virus type-1 through breastfeeding. *Lancet* 1992;340;585-88.
64. AIDS epidemic update: special report on HIV/AIDS: December 2006. Geneva: Joint United Nations Programme on HIV/AIDS (UNAIDS) and World Health Organization, 2006.
65. Kuhn L, Aldrovandi GM, Sinkala M, et al. Extended antiretroviral prophylaxis to reduce breast-milk HIV-1 transmission. *N Engl J Med* 2008;359:130-141.
66. Iliff PJ, Piwoz EG, Tavengwa NV, et al. Early exclusive breastfeeding reduces the risk of postnatal HIV-1 transmission and increases HIV-free survival. *AIDS* 2005;19:699-708.
67. Coovadia HM, Rollins NC, Bland RM, et al. Mother-to-child transmission of HIV-1 infection during exclusive breastfeeding in the first 6 months of life: an intervention cohort study. *Lancet* 2007;369:1107-1116.

68. Kumwenda NI, Hoover DR, Mofenson LM, et al. Extended anti-retroviral prophylaxis to reduce breast-milk HIV-1 transmission. *N Engl J Med* online 10.1056.
69. Stewart R, Loveday M. Public HAART project in South Africa. Progress to November 2004. http://www.hst.org.za/uploads/files/haart_progress1104.pdf. Accessed 17/06/2008.
70. Dual therapy to start for PMTCT to start early next year communiqué 1 December 2007. <http://www.doh.gov.za/docs/pr/2007/index.html>. Accessed 19/06/2010.
71. Mortality and causes of death in South Africa 2005: findings from death notification. <http://www.statssa.com/Publications/P03093>. Accessed 27/04/2008.
72. Williams BG, Gouws E, Boschi-Pinto C, et al. Estimates of world-wide distribution of child deaths from acute respiratory infections. *Lancet Infect Dis* 2002;2:25-32.
73. Mulholland K. Magnitude of the problem of childhood pneumonia. *Lancet* 1999;354:590-592.
74. Ikeogu MO, Wolf B, Mathe S. Pulmonary manifestations in HIV seropositivity and malnutrition in Zimbabwe. *Arch Dis Child* 1997;76:124-128.
75. Lucas SB, Peacock CS, Hounnou A, et al. Disease in children infected with HIV in Abidjan, Cote d'Ivoire. *BMJ* 1996;312:335-338.
76. Vetter KM, Djomand G, Zadi F, et al. Clinical spectrum of human immunodeficiency virus disease in children in a West African city. *Pediatr Infect Dis J* 1996;15:438-442.
77. Graham SM. HIV and respiratory infections in children. *Curr Opin Pulm Med* 2003;9:215-220.
78. Madhi SA, Petersen K, Madhi A, et al. Increased disease burden and antibiotic resistance of bacterial causing severe community-acquired pneumonia lower respiratory tract infections in human immunodeficiency type 1-infected children. *Clin Infect Dis* 2000;31:170-176.
79. Zar HJ, Hanslo D, Tannenbauem E, et al. Aetiology and outcome of pneumonia in human immunodeficiency virus-infected children hospitalized in South Africa. *Acta Paediatr* 2001;90:119-125.
80. Zampoli M, Morrow B, Hsiao NY, et al. Prevalence and outcome of

- cytomegalovirus-associated pneumonia in immunodeficiency virus infection. *Pediatr Infect Dis J* 2011;30:413-417.
81. King JC Jr. Community respiratory viruses in individuals with human immunodeficiency virus infection. *Am J Med* 1996;102:19-24.
 82. McIntosh K. Respiratory viral infections. In: Pizzo PA, Wilfert CM, eds. *Pediatric AIDS. The Challenge of HIV Infection in Infants, Children and Adolescents*, 2nd ed. Baltimore: Williams and Wilkins, 1994:365-376.
 83. King JC, Burke AR, Clemens JD, et al. Respiratory syncytial virus illnesses in human immunodeficiency virus-infected and non-infected children. *Pediatr Infect Dis J* 1993;12:733-739.
 84. Chandwani S, Borkowsky W, Krasinski K, et al. Respiratory syncytial virus infection in human immunodeficiency virus-infected children. *J Pediatr* 1990;117:251-254.
 85. Jaspan HB, Huang LC, Cotton MF, et al. Bacterial disease and antimicrobial susceptibility patterns in HIV-infected hospitalized children: a retrospective cohort study. *PLoS One* 2008;3:e3260.
 86. Punpanich E, Groome M, Muhe L, et al. Systematic review on the etiology and antibiotic treatment of pneumonia in human immunodeficiency virus-infected children. *Pediatr Infect Dis J* 2011;30:192-202.
 87. Wolff AJ, O'Donnell AE. Pulmonary manifestations of HIV infection in the era of highly active antiretroviral therapy. *Chest* 2001;120:1888-1893.
 88. Gingo MR, George MP, Kessinger CJ, et al. Pulmonary function abnormalities in HIV-infected patients during the current antiretroviral therapy era. *Am J Respir Crit Care Med* 2010;182:790-796.
 89. Van Rie A, Beyers N, Gie RP, et al. Childhood tuberculosis in an urban population in South Africa: burden and risk factors. *Arch Dis Child* 1999;80:433-437.
 90. Hesseling AC, Cotton MF, Jennings T, et al. High incidence of tuberculosis among HIV infected infants-South African population-based study. *Clin Infect Dis* 2009;48:108-114.
 91. Lazarus JV, Olsen M, Ditiu L, et al. Tuberculosis-HIV co-infection: policy and epidemiology in 25 countries in WHO European region. *HIV Med* 2008;9:406-414.

92. Coovadia HM, Jeena P, Wilkinson D. Childhood human immunodeficiency virus and tuberculosis co-infections: reconciling conflicting data. *Int J Tuberc Lung Dis* 1998;2:844-851.
93. Marais BJ, Gie RP, Hesselning AC, et al. A refined symptom-based approach to diagnose pulmonary tuberculosis in children. *Pediatrics* 2006;118:e1350-e1359.
94. Hesselning AC, Cotton MF, Fordham von Reyn C, et al. Consensus statement on the revised World Health Organization recommendations for BCG vaccination in HIV-infected infants. *Int J Tuberc Lung Dis* 2008;12:1376-1379.
95. Karpelowsky JS, Alexander AG, Peek SD, et al. Surgical complications of bacilli Calmette-Guérin (BCG) infection in HIV-infected children: time for a change in policy. *S Afr Med J* 2008;98:801-804.
96. Zar HJ, Cotton MF, Strauss S, et al. Effect of isoniazid prophylaxis on mortality and incidence of TB in children with HIV: randomised controlled trial. *BMJ* 2007; 334:1-7.
97. Madhi SA, Nachman S, Violari A, et al. Primary isoniazid prophylaxis against tuberculosis in HIV-exposed children. *N Engl J Med* 2011;365:21-31.
98. Zwi KJ, Pettifor JM, Soderlund N. Paediatric hospital admissions at a South African urban regional hospital: the impact of HIV 1992-1997. *Ann Trop Paediatr* 1999;19:135-142.
99. Nachmann S, Gona P, Dnakner W, et al. The rate of serious bacterial infections among HIV-infected children with immune reconstitution who have discontinued opportunistic infection prophylaxis. *Pediatrics* 2005;115:e488-e494.
100. Laennec RTH. A treatise on the disease of the chest. Forbes J, trans New York: Library of the New York Academy of Medicine, Hafner publishing, 1962;78.
101. Reid LM. Reduction in bronchial subdivision in bronchiectasis. *Thorax* 1950;5:233-247.
102. Whitwell D. A study of the pathology and pathogenesis of bronchiectasis. *Thorax* 1952;7:213-239.
103. Chang AB, Redding GJ. Bronchiectasis. In: Chernick V, Boat TF, Wilmott RW, Bush A, editors. *Kendig's disorders of the respiratory tract in children*. 7th ed. Philadelphia: Saunders Elsevier; 2006. p 460-477.

104. Helbich TH, Heinz-Peer G, Fleischmann D, et al. Evolution of CT findings in patients with cystic fibrosis. *Am J Roentgenol* 1999;173:81-88.
105. Loeve M, Hop WC, de Bruijne M, et al. Chest computed tomography scores are predictive of survival in patients with cystic fibrosis awaiting lung transplantation. *Am J Respir Crit Care Med* 2012;185:1096-1103.
106. Brody AS, Klein JS, Molina PL, et al. High-resolution computed tomography in young patients with cystic fibrosis: distribution of abnormalities and correlation with pulmonary function. *J Pediatr* 2004;145:32-38.
107. Bhalla M, Turcios N, Aponte V, et al. Cystic fibrosis: scoring system with thin-section CT. *Radiology* 1991;179:783-788.
108. Montella S, Maglione M, Bruzzese D, et al. Magnetic resonance imaging is an accurate and reliable method to evaluate non-cystic fibrosis paediatric lung disease. *Respirology* 2012;17:87-91.
109. Goeminne P, Dupont L. Non-cystic fibrosis bronchiectasis: diagnosis and management in 21st century. *Postgrad Med J* 2010;86:493-501.
110. Säynäjäkangas O, Keistinen T, Tuuponen T, et al. Bronchiectasis in Finland: trends in hospital treatment. *Respir Med* 1997;91:395-398.
111. Zhang L, Irion K, da Silva Porto N, et al. High-resolution computed tomography in paediatric patients with postinfectious bronchiolitis obliterans. *J Thorac Imaging* 1999;14:85-89.
112. Dherani M, Pope D, Mascarenhas M, et al. Indoor air pollution from unprocessed solid fuel use and pneumonia risk in children aged under five years: a systematic review and meta-analysis. *Bull World Health Organ* 2008;86:390-398.
113. Perez-Padilla R, Schilman A, Riojas-Rodriguez H. Respiratory health effects of indoor air pollution. *Int J Tuberc Lung Dis* 2010;14:1079-1086.
114. Ng'ang'a LW, Odhiambo JA, Mungai MW, et al. Prevalence of exercise induced bronchospasm in Kenyan school children: an urban-rural comparison. *Thorax* 1998;53:919-926.
115. Volkmer RE, Ruffin RE, Wigg NR, et al. Prevalence of respiratory symptoms in South Australian preschool children II. Factors associated with indoor air quality. *J Paediatr Child Health* 1995;31:112-120.
116. World Health Organization. Ten Facts on the tobacco epidemic and global tobacco control.

http://www.who.int/features/factfiles/toacco_epidemic/tobacco_epidemic_facts/en/index1.html. Accessed 27/05/2009.

117. Stewart DW, Jones GN, Minor KS. Smoking, depression, and gender in low-income African Americans with HIV/AIDS. *Behav Med* 2011;37:77-80.
118. Tesoriero JM, Gieryic SM, Carrascal A, et al. Smoking among HIV positive New Yorkers: prevalence, frequency, and opportunities for cessation. *AIDS Behav* 2010;14:824-835.
119. Chan-Yeung M, Domich-Ward H. Respiratory health effects of exposure to environmental tobacco smoke. *Respirology* 2003;8:131-138.
120. Savitski AN, Mesaros C, Blair IA, et al. Secondhand smoke inhibits both Cl⁻ and K⁺ conductances in normal human bronchial epithelial cells. *Respir Res* 2009;10:120.
121. Feldman JG, Minkoff H, Schneider MF, et al. Association of cigarette smoking with HIV prognosis among women in the HAART era: a report from the women's interagency HIV study. *Am J Public Health* 2006;96:1060-1065.
122. Stokes DC. Pulmonary infections in the immunocompromised paediatric host. In: Ed Chernick V. Boat TF, Wilmott RW, Bush A, editors. *Kendig's disorders of the respiratory tract in children*. Philadelphia: Saunders Elsevier; 2006. p 453-462.
123. Cole PJ. Inflammation: a two-edged sword- the model of bronchiectasis. *Eur J Respir Dis Suppl* 1986;147:6-15.
124. Tsang KW, Chan K, Ho P, et al. Sputum elastase in steady state bronchiectasis. *Chest* 2000;117:420-426.
125. Richmann-Eisenstat JBY, Jorens PG, Hebert CA, et al. Interleukin 8: an important chemoattractant in sputum of patients with chronic inflammatory airways diseases. *Am J Physiol* 1993;264:L413-L418.
126. Aldallal N, McNaughton EE, Manzel LJ, et al. Inflammatory response in airway epithelial cells isolated from patients with cystic fibrosis. *Am J Respir Crit Care Med* 2002;166:1248-1256.
127. Rubin BK. Mucus structure and properties in cystic fibrosis. *Pediatr Respir Reviews* 2007;8:4-7.
128. Zheng L, Lam WK, Tipoe GL, et al. Over expression of matrix metalloproteinases-8 and -9 in bronchiectasis airways in vivo. *Eur Respir J* 2002;20:170-176.

129. Kapur N, Masters IB, Chang AB. Exacerbations in noncystic fibrosis bronchiectasis: Clinical features and investigations. *Respir Med* 2009;103:1681-1697.
130. Bilton D, Canny G, Conway S, et al. Pulmonary exacerbation: Towards a definition for use in clinical trials. Report from the EuroCare CF Working Group on outcome parameters in clinical trials. *J Cyst Fibros* 2011;10:S79-S81.
131. Wedzicha JA, Donaldson GC. Exacerbations of chronic obstructive pulmonary disease. *Respir Care* 2003;48:1204-1213.
132. Fuschillo S, De Filice A, Balzano G. Mucosal inflammation in idiopathic bronchiectasis: cellular and molecular mechanisms. *Eur Respir J* 2008;31:396-406.
133. Hladik F, Sakchalathorn P, Ballwever L, et al. Initial events in establishing vaginal entry and infection by human immunodeficiency virus type-1. *Immunity* 2007;26:145-147.
134. Cohen MS, Shaw GM, McMichael AJ, et al. Acute HIV-1 infection. *N Engl J Med* 2011;364:1943-1954.
135. Feghali CA, Wright TM. Cytokines in acute and chronic inflammation. *Front Biosci* 1997;2:d12-d26.
136. Watanabe D, Uehira T, Yonemoto H, et al. Sustained high levels of serum interferon- γ during HIV-1 infection: a specific trend different from the other cytokines. *Viral Immunol* 2012;23:619-625.
137. Bacot BK, Paul ME, Navarro M, et al. Objective measures of allergic disease in children with human immunodeficiency virus infection. *J Allergy Clin Immunol* 1997;100:707-711.
138. Clerici M, Shearer GM. The Th1-Th2 switch is a critical step in the etiology of HIV infection. *Immunol Today* 1993;14:107-111.
139. Empson M, Bishop AG, Nightingale B, et al. Atopy, anergic status, and cytokine expression in HIV-infected subjects. *J Allergy Clin Immunol* 1999;103:833-842.
140. Liu Z, Liu Q, Pesce J, et al. Requirements for the development of IL-4 producing T cells during intestinal nematode infections: what it takes to make a Th2 cell in vivo. *Immunol Rev* 2004;201:57-74.

141. Patella V, Florio G, Petraroli A, et al. HIV-1 gp120 cytokines induces IL-4 and IL-13 release from human Fc epsilon R+ cells through interaction with VH3 region of IgE. *J Immunol* 2000;164:589-595.
142. Proesmans M, Els C, Vermeulen F, et al. Change in IgG and evolution of lung function in children with cystic fibrosis. *J Cystic Fibros* 2011;10:128-131.
143. Garside JP, Kerrin DP, Brownlee KG, et al. Immunoglobulin and IgG subclass levels in a regional pediatric cystic fibrosis clinic. *Pediatr Pulmonol* 2005;39:135-140.
144. de Paulis A, De Palma R, Di Giola I, et al. Tat protein is an HIV-1 encoded beta-chemokine homolog that promotes migration and up-regulates CCR3 expression on human Fc epsilon R+ cells. *J Immunol* 2000;165:7171-7179.
145. Keating SM, Golub ET, Nowicke M, et al. The effect of HIV infection and HAART on inflammatory biomarkers in a population-based cohort of women. *AIDS* 2011;25:1823-1832.
146. Fahey JL, Taylor MB, Manna B, et al. Prognostic significance of plasma markers of immune activation, HIV viral load and CD4 T-cell measurements. *AIDS* 1998;12:1581-1590.
147. Lee N, Wong CK, Chan PK, et al. Hypercytokinemia and hyperactivation of phospho-p38 mitogen-activated protein kinase in severe human influenza A virus infection. *Clin Infect Dis* 2007;45:723-731.
148. Eller J, Lapa e Silva JR, Poulter LW, et al. Cells and cytokines in chronic bronchial infection. *Ann NY Acad Sci* 1994;725:331-345.
149. Loikides S, Bouros D, Papatheodorou G, et al. Exhaled H₂O₂ in steady-state bronchiectasis: relationship with cellular composition in induced sputum, spirometry, and extent of severity of disease. *Chest* 2002;121:81-87.
150. Janeway CA Jr, Medzhitov R. Innate immune recognition. *Annu Rev Immunol* 2002;20:197-216.
151. Simpson JL, Grissell TV, Douwes J, et al. Innate immune activation in neutrophilic asthma and bronchiectasis. *Thorax* 2007;62:211-218.
152. Mikami M, Llewellyn-Jones CG, Bayley D, et al. The chemotactic activity of sputum from patients with bronchiectasis. *Am J Respir Crit Care Med* 1998;157:723-728.

153. Angrill J, Augusti C, de Celis R, et al. Bronchial inflammation and colonization in patients with clinically stable bronchiectasis. *Am J Respir Crit Care Med* 2001;164:1628-1632.
154. Griese M, Kappler M, Gaggar A, et al. Inhibition of airway proteases in cystic fibrosis lung disease. *Eur Respir J* 2008;32:783-795.
155. Birrer P, McElvaney NG, Ruderberg A, et al. Protease-antiprotease imbalance in the lungs of children with cystic fibrosis. *Am J Respir Crit Care Med* 1994;150:207-213.
156. Sagel SD, Kapsner RK, Osberg I. Induced sputum matrix metalloproteinase-9 correlates with lung function and airway inflammation in children with cystic fibrosis. *Pediatr Pulmonol* 2005;39:224-232.
157. Colombo C, Costantini D, Rocchi A, et al. Cytokine levels in sputum of cystic fibrosis patients before and after antibiotic therapy. *Pediatr Pulmonol* 2005;40:15-21.
158. Hill SL, Burnett D, Hewerson KA, et al. The response of patients with purulent bronchiectasis to antibiotics for four months. *Q J Med* 1988;66:163-173.
159. Stockley RA, Hill SL, Morrison HM. Effect of antibiotic treatments on sputum elastase in bronchiectatic outpatients in a stable clinical state. *Thorax* 1984;39:414-419.
160. Ip M, Shum D, Lauder I, et al. Effect of antibiotics on sputum inflammatory contents in acute exacerbations of bronchiectasis. *Respir Med* 1993;87:449-454.
161. Bouchon A, Dietrich J, Colonna M. Cutting edge: inflammatory responses can be triggered by TREM-1, a novel receptor expressed on neutrophils and monocytes. *J Immunol* 2000;164:4991-4995.
162. Gingras MC, Lapillonne H, Margolin JF. TREM-1, MDL, and DAP12 expression is associated with a mature stage of myeloid development. *Mol Immunol* 2002;38:817-824.
163. Bouchon A, Facchetti F, Weigand MA, et al. TREM-1 amplifies inflammation and is a crucial mediator of septic shock. *Nature* 2001;410:1103-1107.
164. Bleharski JR, Kiessler V, Buonsanti C, et al. A role for triggering receptor expressed on myeloid cells-1 in host defense during the early-induced and adaptive phase of the immune response. *J Immunol* 2003;170:3812-3818.

165. Richeldi L, Mariani M, Lose M, et al. Triggering receptor expressed on myeloid cells: role in the diagnosis of lung infections. *Eur Respir J* 2004;24:247-250.
166. Gibot S, Cravoisy A, Levy B, et al. Soluble triggering receptor expressed on myeloid cells and the diagnosis of pneumonia. *N Engl J Med* 2004;350:451-458.
167. Barraud D, Gibot S. Triggering receptor expressed on myeloid cell 1. *Crit Care Clin* 2011;27:265-279.
168. Rohde G, Rasdak MP, Borg I, et al. Levels of soluble triggering receptor expressed on myeloid cells 1 in infectious exacerbations of chronic obstructive pulmonary disease. *Respiration* 2012;83:133-139.
169. Tintinger GR, van der Merwe JJ, Fickl H, et al. Soluble triggering receptor expressed on myeloid cells in sputum of patients with community-acquired pneumonia or pulmonary tuberculosis: a pilot study. *Eur J Clin Microbiol Infect Dis* 2012;31:73-76.
170. Shu CC, Lee LN, Lee CH, et al. Use of soluble triggering receptor expressed on myeloid cells-1 in non-tuberculous mycobacterial lung disease. *Int J Tuberc Lung Dis* 2011;15:1415-1420.
171. del Fresno C, Gómez-Piña V, Lores V, et al. Monocytes from cystic fibrosis patients are locked in an LPS tolerance state: down-regulation of sTREM as putative underlying mechanism. *PLoS One* 2008;3:e2667.
172. Feldman C. Bronchiectasis: new approaches to diagnosis and management. *Clin Chest Med* 2011;32:535-546.
173. Elkins MR, Jones A, van der Schans C. Positive expiratory pressure physiotherapy for airway clearance in people with cystic fibrosis. *Cochrane Database Syst Rev* 2006; ID CD003147.
174. Murray MP, Pentland JL, Hill AT. A randomized crossover trial of chest physiotherapy in non-cystic fibrosis bronchiectasis. *Eur Respir J* 2009;34:1086-1092.
175. Bush A, Payne S, Pike G, et al. Mucus properties in children with primary ciliary dyskinesia: comparison with cystic fibrosis. *Chest* 2006;129:118-123.
176. Kellett F, Redfern J, Niven RM. Evaluation of nebulised hypertonic saline (7%) as an adjunct to physiotherapy in patients with stable bronchiectasis. *Respir Med* 2005;99:27-31.

177. Daviskas E, Anderson SD, Young IH. Effect of mannitol and repetitive coughing on the sputum properties in bronchiectasis. *Respir Med* 2010;104:371-377.
178. Wills P, Greenstone M. Inhaled hyperosmolar agents for bronchiectasis. *Cochrane Database Syst Rev* 2006; ID CD002996.
179. Rubin BK. Aerosolized antibiotics for non-cystic fibrosis bronchiectasis. *J Aerosol Med Pulm Drug Deliv* 2008;21:71-76.
180. Orriols R, Roig J, Ferrer J, et al. Inhaled antibiotic therapy in non cystic fibrosis patients with bronchiectasis and chronic bronchial infection by *Pseudomonas aeruginosa*. *Respir Med* 1999;93:476-480.
181. Scheinberg P, Shore E. A pilot study of the safety and efficacy of tobramycin solution for inhalation inpatients with severe bronchiectasis. *Chest* 2005;127:1420-1426.
182. Tsang KW, Tan KC, Ho PL, et al. Inhaled fluticasone in bronchiectasis: a 12 month study. *Thorax* 2005;60:239-243.
183. Martinez-Garcia MA, Perpina-Tordera M, Roman-Sanchez P, et al. Inhaled steroids improve quality of life in patients with steady-state bronchiectasis. *Respir Med* 2006;100:1623-1632.
184. Kapur N, Bell S, Kolbe J, et al. Inhaled steroids for bronchiectasis. *Cochrane Database Syst Rev* 2009;1:CD000996.
185. Foisy MM, Yakiwchuk EM, Singh AE. Adrenal suppression and Cushing's syndrome secondary to an interaction between ritonavir and fluticasone: a review of the literature. *HIV Med* 2008;9:389-396.
186. Togami K, Chono S, Morimoto K. Distribution characteristics of clarithromycin and azithromycin, macrolide antimicrobial agents used for treatment of respiratory infections, in lung epithelial lining fluid and alveolar macrophages. *Biopharm Drug Dispos* 2011;32:389-397.
187. Yasuda H, Ajiki Y, Koga T, et al. Interaction between biofilms formed by *Pseudomonas aeruginosa* and clarithromycin. *Antimicrob Agents Chemother* 1993;37:1749-1755.
188. Shinkai M, Henke MO, Rubin BK. Macrolide antibiotics as immunomodulatory medications: proposed mechanisms of action. *Pharmacol Ther* 2008;117:393-405.

189. Siracusa A, Brugnami G, Fiordi T, et al. Troleandomycin in the treatment of difficult asthma. *J Allergy Clin Immunol* 1993;92:677-682.
190. Nagai H, Shishido H, Vonedo R, et al. Long term low-dose administration of erythromycin to patients with diffuse panbronchiolitis. *Respiration* 1991;58:145-149.
191. Trenadiel J, Zalcman G, Gerber F, et al. Diffuse panbronchiolitis: efficacy of low-dose erythromycin. *Respir Med* 1993;87:229-230.
192. Hoiby N. Diffuse panbronchiolitis and cystic fibrosis: East meets West. *Thorax* 1994;49:531-532.
193. Kudoh S, Azuma A, Yamamoto M, et al. Improvement of survival in patients with diffuse panbronchiolitis treated with low-dose erythromycin. *Am J Respir Crit Care Med* 1998;157:1892-1898.
194. Davies G, Wilson R. Prophylactic antibiotic treatment of bronchiectasis with azithromycin. *Thorax* 2004;59:540-541.
195. Coeman M, van Durme Y, Bauters F, et al. Neomacrolides in the treatment of patients with severe asthma and/or bronchiectasis: a retrospective observational study. *Thorax* 2011;5:377-386.
196. McCormack J. Effect of long term treatment with azithromycin on disease parameters in cystic fibrosis. *Thorax* 2002;57:212-216.
197. Saiman L, Anstead M, Mayer-Hamblett N, et al. Effect of azithromycin on pulmonary function in patients with cystic fibrosis uninfected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA* 2010;303:1707-1715.
198. Cymbala AA, Edmonds LC, Bauer MA, et al. The disease-modifying effects of twice-weekly oral azithromycin in patients with bronchiectasis. *Treat Respir Med* 2005;4:117-122.
199. Kanoh S, Rubin BK. Mechanisms of action and clinical application of macrolides as immunomodulatory medications. *Clin Microbiol Rev* 2010;23:590-615.
200. Phaff SJ, Tiddens HAWM, Verbrugh HA, et al. Macrolide resistance of *Staphylococcus aureus* and Haemophilus species associated with long-term azithromycin use in cystic fibrosis. *J Antimicrob Chemother* 2006;57:741-746.

201. Tramper-Stranders GA, Wolfs TF, Flear A, et al. Maintenance azithromycin with cystic fibrosis: long-term outcomes related to macrolide resistance and pulmonary function. *Pediatr Infect Dis J* 2007;26:8-12.
202. Olivier KC, Weber DJ, Wallace RJ, et al. Nontuberculous mycobacteria. I: Multicentre prevalence study in cystic fibrosis. *Am J Respir Crit Care Med* 2003;167:828-834.
203. Pasteur MC, Bilton D, Hill AT, et al. British thoracic society guideline for non-CF bronchiectasis. *Thorax* 2010;65:1-58.
204. Suri R, Marshall LJ, Wallis C, et al. Safety and use of sputum induction in children with cystic fibrosis. *Pediatr Pulmonol* 2003;35:309-313.
205. Jones T, Price P. Development and experimental medicine application of PET in oncology: a historical perspective. *Lancet Oncol* 2012;13:e116-e125.
206. Castell F, Cook GJR. Quantitative techniques in ¹⁸F-FDG-PET scanning in oncology. *Br J Cancer* 2008;98:1597-1601.
207. Smith TAD. The rate-limiting step for tumour ¹⁸F-fluoro-2-deoxy-D-glucose (FDG) incorporation. *Nucl Med Biol* 2001;28:1-4.
208. Warburg O. On respiratory impairment in cancer cells. *Science* 1956;124:269-270.
209. Endo K, Oriuchi N, Higuchi T, et al. PET and PET/CT using ¹⁸F-FDG in the diagnosis and management of cancer patients. *Int J Clin Oncol* 2006;11:286-296.
210. Deichen JT, Prante O, Gack M, et al. Uptake of [18F] flourodeoxyglucose in human monocyte-macrophages in vitro. *Eur J Nucl Med Mol Imaging* 2003;30:267-273.
211. Shreve PD, Anzal Y, Wahl RL. Pitfalls in oncologic diagnosis with FDG PET imaging: physiologic and benign variants. *Radiographics* 1999;19:61-77.
212. Matsui T, Nakata N, Nagai S, et al. Inflammatory cytokines and hypoxia contribute to 18F-FDG uptake by cell involved in pannus formation in rheumatoid arthritis. *J Nucl Med* 2009;50:920-926.
213. Scharko A, Perlman S, Hinds P, et al. Whole body positron emission tomography imaging of simian immunodeficiency virus-infected rhesus macaques. *Proc Natl Acad Sci USA* 1996;93:6423-6430.
214. Wallace M, Pyzalski R, Horejsh D, et al. Whole body positron emission tomography imaging of activated lymphoid tissues during acute simian-human

- immunodeficiency virus 89.6DP infection in rhesus macaques. *Virology* 2000; 274:255-261.
215. Sathekge M, Maes A, D'Asseler Y, et al. Tuberculous lymphadenitis: FDG PET and CT findings in responsive and nonresponsive disease. *Eur J Nucl Med Mol Imaging* 2012;39:1184-1190.
216. Sathekge M, Maes A, Kgomo M, et al. Use of 18F-FDG PET to predict response to first-line tuberculostatics in HIV-associated tuberculosis. *J Nucl Med* 2011;52:880-885.
217. Sato H, Hiyama T, Kaito K, et al. Usefulness of F-18FDG PET/CT in the assessment of disseminated Mycobacterium avium complex infection. *Ann Nucl Med* 2009;23:757-762.
218. Jones HA, Sriskandan S, Peters AM, et al. Dissociation of neutrophil emigration and metabolic activity in lobar pneumonia and bronchiectasis. *Eur Respir J* 1997;10:795-803.
219. Labiris NR, Nahmias C, Freitag AP, et al. Uptake of 18 fluorodeoxyglucose in the cystic fibrosis lung: a measure of lung inflammation? *Eur Respir J* 2003;21:848-854.
220. Jones HA, Marino PS, Shakur BH, et al. In vivo assessment of lung inflammatory cell activity in patients with COPD and asthma. *Eur Respir J* 2003;21:567-573.
221. Klein M, Cohen-Cymbarknoh M, Armoni S, et al. 18F-fluorodeoxyglucose PET/CT imaging of lungs in patients with cystic fibrosis. *Chest* 2009;136:1220-1228.
222. Chen DL, Ferkol TW, Mintun MA, et al. Quantifying pulmonary inflammation in cystic fibrosis with positron emission tomography. *Am J Respir Crit Care Med* 2006;173:1363-1369.
223. McGrath EE, McCabe J, Anderson PB. Guidelines on the diagnosis and treatment of pulmonary non-tuberculous mycobacteria infection. *Int J Clin Pract* 2008;62:1947-1955.
224. Bonard D, Messou E, Seyler C, et al. High incidence of atypical mycobacteriosis in African HIV-infected adults with low CD4 counts: a 6 year cohort study in Cote d'Ivoire. *AIDS* 2004;24:1961-1964.
225. www.who.int/childgrowth/standards/en/. Accessed 10/6/2011.
226. Rosenfeld M, Emerson J, Williams-Warren J, et al. Defining a pulmonary

- exacerbation in cystic fibrosis. *J Pediatr* 2001;139:359-365.
227. Verweel G, van Rossum AM, Hartwig NG, et al. Treatment with highly active antiretroviral therapy in human immunodeficiency type-1 virus infected children is associated with a sustained effect on growth. *Pediatrics* 2002;109:E25.
228. Shikuma CM, Zackin R, Sattler F, et al. Changes in weight and lean body mass during highly active antiretroviral therapy. *Clin Infect Dis* 2004;39:1223-1230.
229. Steinkamp G, Wiedemann B. Relationship between nutritional status and lung function in cystic fibrosis: cross sectional and longitudinal analyses from the German CF quality assurance (CFQA) project. *Thorax* 2002;57:596-601.
230. Von Gottberg A, de Gouveia L, Madhi SA, et al. Impact of conjugate *Haemophilus influenzae* type b (Hib) vaccine introduction in South Africa. *Bull World Health Organ* 2006;84:811-818.
231. Madhi SA, Petersen K, Khoosal M, et al. Reduced effectiveness of *Haemophilus influenzae* type b conjugate vaccine in children with a high prevalence of human immunodeficiency virus type 1 infection. *Pediatr Infect Dis J* 2002;21:315-321.
232. McNally LM, Jeena PM, Gajee A, et al. Lack of association between the nasopharyngeal carriage of *Streptococcus pneumoniae* and *Staphylococcus aureus* in HIV-1 infected South African children. *J Infect Dis* 2006;194:385-390.
233. Theart AC, Marais BJ, Gie RP, et al. Criteria used for the diagnosis of childhood tuberculosis at primary health care level in a high-burden, urban setting. *Int J Tuberc Lung Dis* 2005;9:1210-1214.
234. Munro KA, Reed PW, Joyce H, et al. Do New Zealand children with non-cystic fibrosis bronchiectasis show disease progression? *Pediatr Pulmonol* 2011;46:131-138.
235. Haidopoulou K, Calder A, Jones A, et al. Bronchiectasis secondary to primary immunodeficiency in children: longitudinal changes in structure and function. *Pediatr Pulmonol* 2009;44:669-675.
236. Pohling J, Zipperlen K, Hollett NA, et al. Human immunodeficiency virus type 1-specific CD8⁺ T cell subset abnormalities in chronic infection persist through

- effective antiretroviral therapy. *BMC Infect Dis* 2010;10:129 doi;10.1186/1471-2334-10-129.
237. Zar HJ, Latief Z, Hughes J, et al. Serum immunoglobulin E levels in human immunodeficiency virus-infected children with pneumonia. *Pediatr Allergy Immunol* 2002;13:328-333.
238. Mazengara LR, Nathoo KJ, Rusakaniko S, et al. Serum IgG subclasses levels in paediatric patients with pneumonia. *Cent Afr J Med* 2001;47:142-145.
239. Green RJ, Becker PJ, Labuschagne D, et al. Disease progression unrelated to passive environmental tobacco smoke exposure in HIV-infected children. *Int J Collaborative Res Int Med Public Health* 2012;4:130-135.
240. Kabali C, Cheng DM, Brooks C, et al. Recent cigarette smoking and HIV disease progression: no evidence of an association. *AIDS Care* 2011;10:1-10.
241. Kodgule R, Salvi S. Exposure to biomass smoke as a cause of disease in women and children. *Curr Opin Allergy Clin Immunol* 2012;12:82-90.
242. Murray EL, Brondi L, Kleinbaum D, et al. Cooking fuel type, household ventilation, and the risk of acute lower respiratory tract infections in urban Bangladeshi children: a longitudinal study. *Indoor Air* 2012;22:132-139.
243. Rehfuess EA, Tzala L, Best N, et al. Solid fuel use and cooking practices as a major risk factor for ALRI mortality among African children. *J Epidemiol Community Health* 2009;63:887-892.
244. Interim WHO clinical staging of HIV/AIDS and HIV/AIDS case definition for surveillance.
<http://www.who.int/hiv/pub/guidelines/clinicalstaging.pdf> Accessed 20/05/2008.
245. Centers for Disease Control and Prevention (CDC) 1994 revised classification system for human immunodeficiency virus infection in children less than 13 years of age. *MMWR Recomm Rep* 1994;43:1-10.
246. Bonfield TL, Panushka JR, Konstan MW, et al. Inflammatory cytokines in cystic fibrosis lungs. *Am J Respir Crit Care Med* 1995;152:2111-2118.
247. Saiman L. Microbiology of early CF lung disease. *Paediatr Resp Rev* 2004;5:S367-S369.
248. Bartling TR, Drumm ML. Oxidative stress causes IL-8 promoter hyperacetylation in cystic fibrosis airway cell models. *Am J Respir Cell Mol Biol* 2009;40:58-65.

249. Cozzi-Lepri A, French MA, Baxter J, et al. Resumption of HIV replication is associated with monocyte/macrophage derived cytokine and chemokine changes: results from a large international clinical trial. *AIDS* 2011;25:1207-1217.
250. Norris PJ, Pappalardo BL, Custer B, et al. Elevations in IL-10, TNF-alpha, and INF-gamma from the earliest point of HIV type 1 infection. *AIDS Res Hum Retroviruses* 2006;22:757-762.
251. Stacey AR, Norris PF, Qin L, et al. Induction of a striking systemic cytokine cascade prior to peak viremia in acute human immunodeficiency virus type 1 infection, in contrast to more modest and delayed responses in acute hepatitis B and C virus infections. *J Virol* 2009;83:3719-3733.
252. Thobakgale CF, Streeck H, Mkhwanazi N, et al. Short communication: CD8 (+) T cell polyfunctionality profiles in progressive and nonprogressive pediatric HIV type 1 infection. *AIDS Res Hum Retroviruses* 2011;27:1005-1012.
253. Shebl FM, Yu K, Landgren O, et al. Increased levels of circulating cytokines in HIV-related immunosuppression. *AIDS Res Hum Retroviruses* 2012;28:809-815.
254. Vigano A, Principi N, Crupi L, et al. Elevation of IgE in HIV-infected children and its correlation with progression of disease. *J Allergy Clin Immunol* 1995;95:627-632.
255. Gingo MR, Wenzel SE, Steele C, et al. Asthma diagnosis and airway bronchodilator response in HIV-infected patients. *J Allergy Clin Immunol* 2012;129:708-714.
256. Bowser CS, Kaye J, Joks RO, et al. IgE and atopy in perinatally HIV-infected children. *Pediatr Allergy Immunol* 2007;18:298-303.
257. ISAAC Steering Committee. Worldwide variation in prevalence of symptoms of asthma, allergic rhinoconjunctivitis and atopic eczema. ISAAC. *Lancet* 1998;351:1225-1232.
258. Knynyk JA, Parsons JP, Para MF, et al. HIV and asthma, is there an association. *Respir Med* 2012;106:493-499.
259. Rudikoff D. The relationship between HIV infection and atopic dermatitis. *Curr Allergy and Asthma Rep* 2002;2:275-281.
260. Strachan DP, Sibbald B, Weiland SK, et al. Worldwide variations in prevalence of symptoms of allergic rhinoconjunctivitis in children: International

- Study of Asthma and Allergies in Childhood (ISAAC). *Pediatr Allergy Immunol* 1997;8:161-176.
261. Garcia-Rodriguez JF, Corominas M, Fernandez-Vilarich P, et al. Rhinosinusitis and atopy in patients infected with HIV. *Laryngoscope* 1999; 109:939-944.
 262. Zanzinger K, Schellack C, Nausch N, et al. Regulation of triggering receptor expressed on myeloid cells 1 expression on mouse inflammatory monocytes. *Immunology* 2009;128:185-195.
 263. Gan WQ, Man SFP, Senthilselvan A, et al. Association between chronic obstructive pulmonary disease and systemic inflammation: a systematic review and a meta-analysis. *Thorax* 2004;59:574-580.
 264. Rasdak MP, Taube C, Haselmayer P, et al. Soluble triggering receptor expressed on myeloid cells 1 is released in patients with stable chronic obstructive pulmonary disease. *Clin Dev Immunol* 2007;52040.
 265. Lin CH, Yao M, Hsu SC, et al. Soluble triggering receptor expressed on myeloid cells-1 as an infection marker for patients with neutropenic fever. *Crit Care Med* 2011;39:993-999.
 266. Alavi A, Gupta J, Alberini M, et al. Positron emission tomography in non-malignant thoracic disorders. *Semin Nucl Med* 2002;32:293-321.
 267. Paik J, Lee K, Choe S, et al. Augmented ¹⁸F-FDG uptake in activated monocytes occurs during the priming process and involves tyrosine kinases and protein-kinase C. *J Nucl Med* 2004;45:124-128.
 268. Santamaria F, Montella S, Pifferi M, et al. A descriptive study of non-cystic fibrosis bronchiectasis in a pediatric population of central and southern Italy. *Respiration* 2009;77:160-165.
 269. Guran T, Ersu R, Karadag B, et al. Association between inflammatory markers in induced sputum and clinical characteristics in children with non-cystic fibrosis bronchiectasis. *Pediatr Pulmonol* 2007;42:362-369.
 270. Dogru D, Nik-Ain A, Kiper N, et al. Bronchiectasis: the consequence of late diagnosis in chronic respiratory symptoms. *J Trop Pediatr* 2005;51:362-365.
 271. Umeda Y, Demura Y, Ishizaki T, et al. Dual-time-point ¹⁸F-FDG PET imaging for diagnosis of disease type and disease activity in patients with idiopathic interstitial pneumonia. *Eur J Nucl Med Mol Imaging* 2009;36:1121-1130.

272. Win T, Screatton NJ, Porter J, et al. Novel positron emission tomography/computed tomography of diffuse parenchymal lung disease combining a labelled somatostatin receptor analogue and 2-deoxy-2 [¹⁸F] fluoro-d-glucose. *Mol Imaging* 2012;11:91-98.
273. Gingo MR, Gorge MP, Kessinger CJ, et al. Pulmonary function abnormalities in HIV-infected patients during the current antiretroviral therapy era. *Am J Respir Crit Care Med* 2010;182:790-796.
274. Guibot A, Tubiana R, Breton G, et al. Immune and virological benefits of 10 years of permanent viral control with antiretroviral therapy. *AIDS* 2010;24:617-619.
275. Kaufmann GR, Furrer H, Ledergerber B, et al. Characteristics, determinants, and clinical relevance of CD4 T cell recovery to < 500 cells/microL in HIV type-1 infected individuals receiving potent antiretroviral therapy. *Clin Infect Dis* 2005;41:361-372.
276. Fowler SJ, French J, Screatton NJ, et al. Nontuberculous mycobacteria in bronchiectasis: prevalence and patient characteristics. *Eur Respir J* 2006;28:1204-1210.
277. Vanini V, Petruccioli E, Gioia C, et al. IP-10 is an additional marker for tuberculosis (TB) detection in HIV-infected persons in a low-TB endemic country. *J Infect* 2012;65:49-59.
278. Lane BR, King SR, Bock PJ, et al. The C-X-C chemokine IP -10 stimulates HIV-1 replication. *Virology* 2003;307:122-134.
279. Stylianou E, Aukrust P, Bendtzen K, et al. Interferon and interferon (IFN)-inducible protein-10 during highly active anti-retroviral therapy (HAART)-possible immunosuppressive role of IFN-alpha in HIV infection. *Clin Exp Immunol* 2002;130:279-285.
280. Bruce MC, Poncz L, Klinger JD, et al. Biochemical and pathologic evidence for proteolytic destruction of lung connective tissue in cystic fibrosis. *Am Rev Respir Dis* 1985;132:529-535.
281. Suter S, Schaad UB, Tegner H, et al. Levels of free granulocyte elastase in bronchial secretions from patients with cystic fibrosis: effect of antimicrobial treatment against *Pseudomonas aeruginosa*. *J Infect Dis* 1986;153:902-909.
282. Meyer KC, Lewandoski JR, Zimmerman JJ, et al. Human neutrophil elastase and elastase/alpha1-antiprotease complex in cystic fibrosis. Comparison with

- interstitial lung disease and evaluation of the effect of intravenously administered antibiotic therapy. *Am Rev Respir Dis* 1991;144:580-585.
283. Gaggar A, Li Y, Weathington N, et al. Matrix metalloprotease-9 dysregulation in lower airway secretions of cystic fibrosis patients. *Am J Physiol Lung Cell Mol Physiol* 2007;293:L96-L104.
284. Downey DG, Brockbank S, Martin SL, et al. The effect of treatment of cystic fibrosis pulmonary exacerbations on airways and systemic inflammation. *Pediatr Pulmonol* 2007;42:729-738.
285. Kapur N, Grimwood K, Masters IB, et al. Lower airway microbiology and cellularity in children with newly diagnosed non-CF bronchiectasis. *Pediatr Pulmonol* 2012;47:300-307.
286. Roberts L, Passmore JA, Williamson C, et al. Plasma cytokine levels during HIV-1 infection predict HIV disease progression. *AIDS* 2010;24:819-831.
287. Doucet-Populaire F, Buriánková K, Weiser J, et al. Natural and acquired macrolide resistance in mycobacteria. *Curr Drug Targets Infect Disord* 2002;2:355-370.

TERMINOLOGY AND ABBREVIATIONS

ABPA	Allergic bronchopulmonary aspergillosis
AIDS	Acquired immunodeficiency syndrome
CAP	Community acquired pneumonia
CCR3	CC chemokine receptor-3
CCR5	CC chemokine receptor-5
CDC	Centre for Disease Control
CF	Cystic fibrosis
CFTR	Cystic fibrosis transmembrane regulator
COPD	Chronic obstructive pulmonary disease
CRF	Circulating recombinant forms
CXR	Chest x ray
D _{LCO}	Pulmonary diffusion capacity for carbon monoxide
FEV ₁	Forced expiratory volume in one second
FEF ₂₅₋₇₅	Forced expiratory flow over 25-75% of expiration
F _{ce} R1	Human F epsilon R positive cells
FVC	Forced vital capacity
G-CSF	Granulocyte colony stimulating factor
GM-CSF	Granulocyte macrophage colony stimulating factor
HAART	Highly active anti-retroviral therapy
<i>H. influenzae</i>	<i>Haemophilus influenzae</i>
HIV	Human immunodeficiency virus
<i>H. parainfluenzae</i>	<i>Haemophilus parainfluenzae</i>

HRCT	High resolution chest tomography
ICAM-1	Intracellular adhesion molecule-1
Ig	Immunoglobulin
IL	Interleukin
INF- γ	Interferon gamma
IP-10	Interferon gamma inducible protein-10
IPT	Isoniazid prophylaxis treatment
LRTI	Lower respiratory tract infection
MCP-1	Monocyte chemotactic protein-1
MIP-1	Macrophage inflammatory protein-1
MMP	Metalloproteinase
MRSA	Methicillin resistant staphylococcus aureus
NE	Neutrophil elastase
NF κ β	Nuclear factor kappa-beta
NTM	Non-tuberculous mycobacteria
PA	Pseudomonas aeruginosa
PAMPs	Pathogen associated molecular patterns
PcP	Pneumocystis jirovecii pneumonia
PET	Positron emission tomography
PMTCT	Prevention of mother to child transmission
PRP	Pattern recognition proteins
RAST	Radio Allergo Sorbent Test
rhDNAse	Recombinant DNAse

RSV	Respiratory syncytial virus
S. aureus	Staphylococcus aureus
sTNFR1	Soluble tumour necrosis factor receptor-1
sTREM	Soluble triggering receptor expressed on myeloid cells
Tat	HIV-trans-activating protein
TB	Tuberculosis
Th1	T helper-1
Th2	T helper-2
TLR	Toll-like receptors
TNF- α	Tumour necrosis factor alpha
URF	Unique recombinant forms
VCAM-1	Vascular cell adhesion molecule -1
WHO	World Health Organisation